

Supporting Information for

ORIGINAL ARTICLE

**Tetrahedral DNA nanostructures synergize with MnO<sub>2</sub> to enhance antitumor immunity *via* promoting STING activation and M1 polarization**

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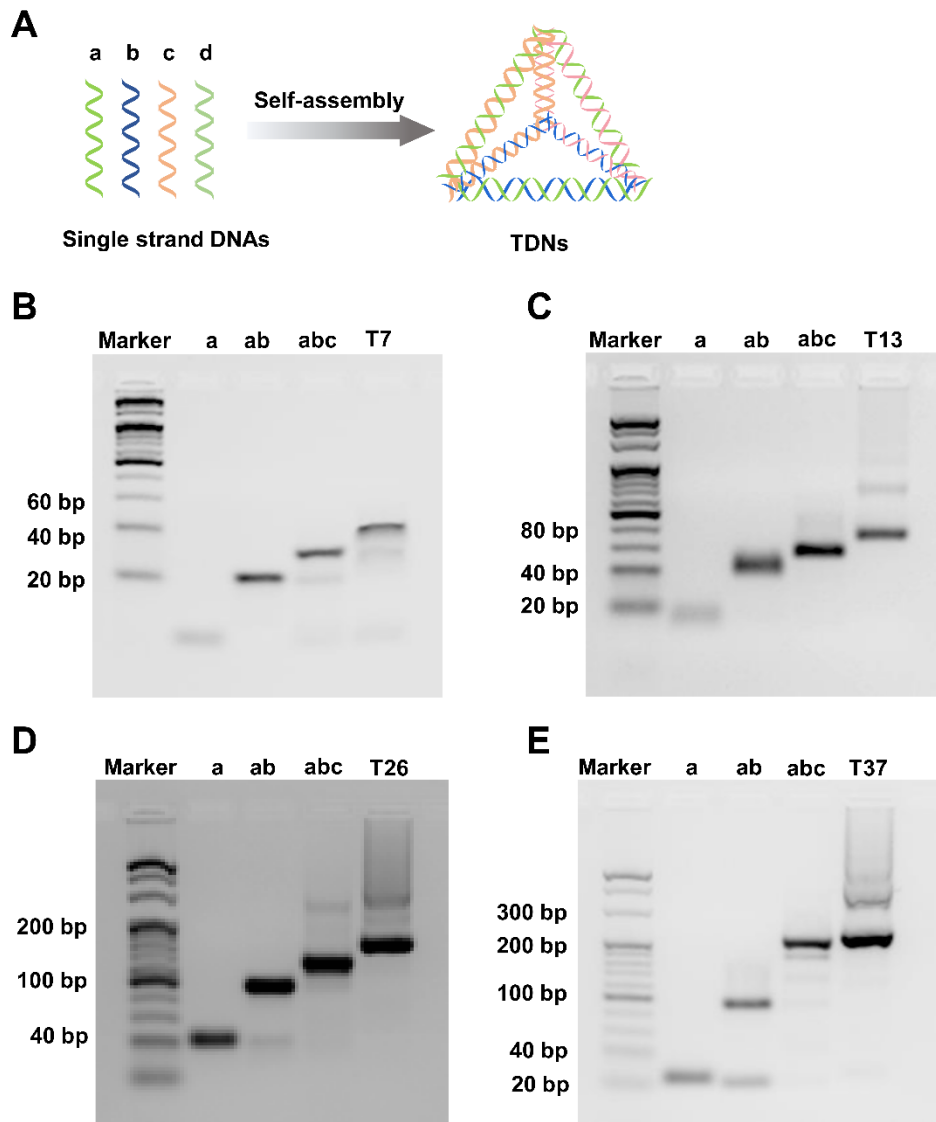
Received 23 August 2021; received in revised form 8 October 2021; accepted 18 November 2021

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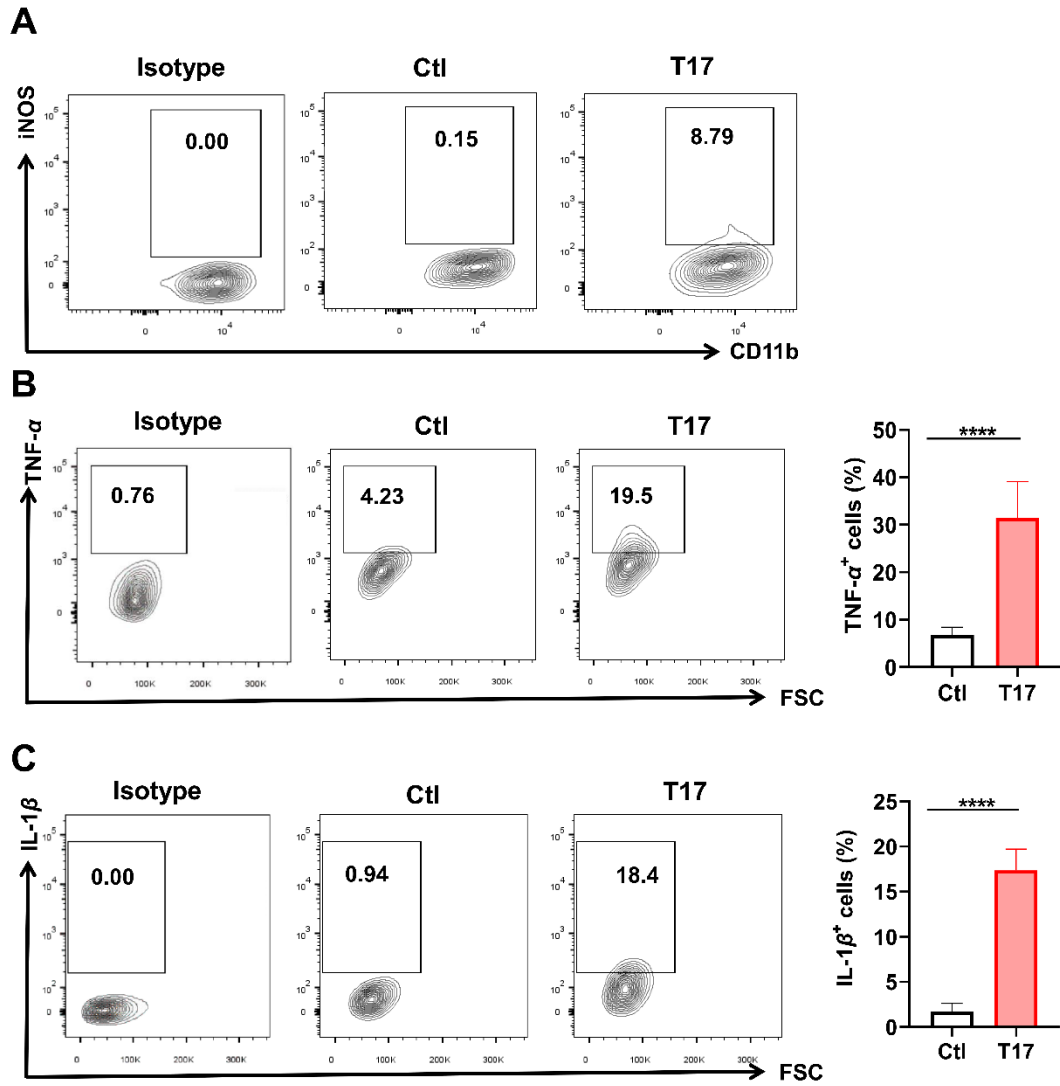
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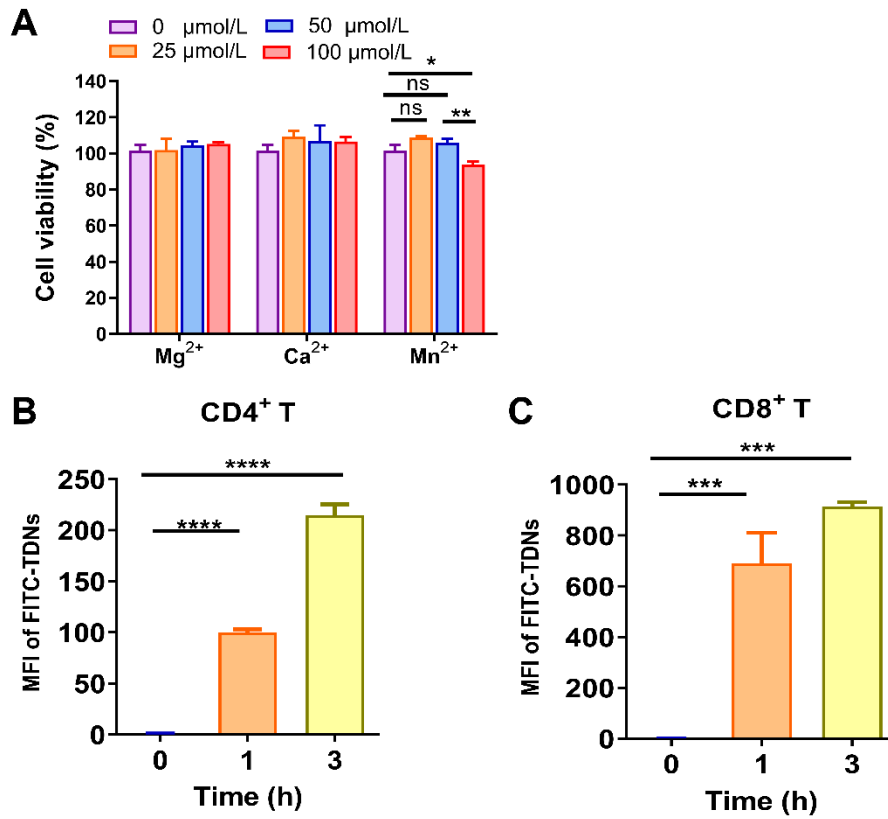
**Supporting Figures S1–S6**



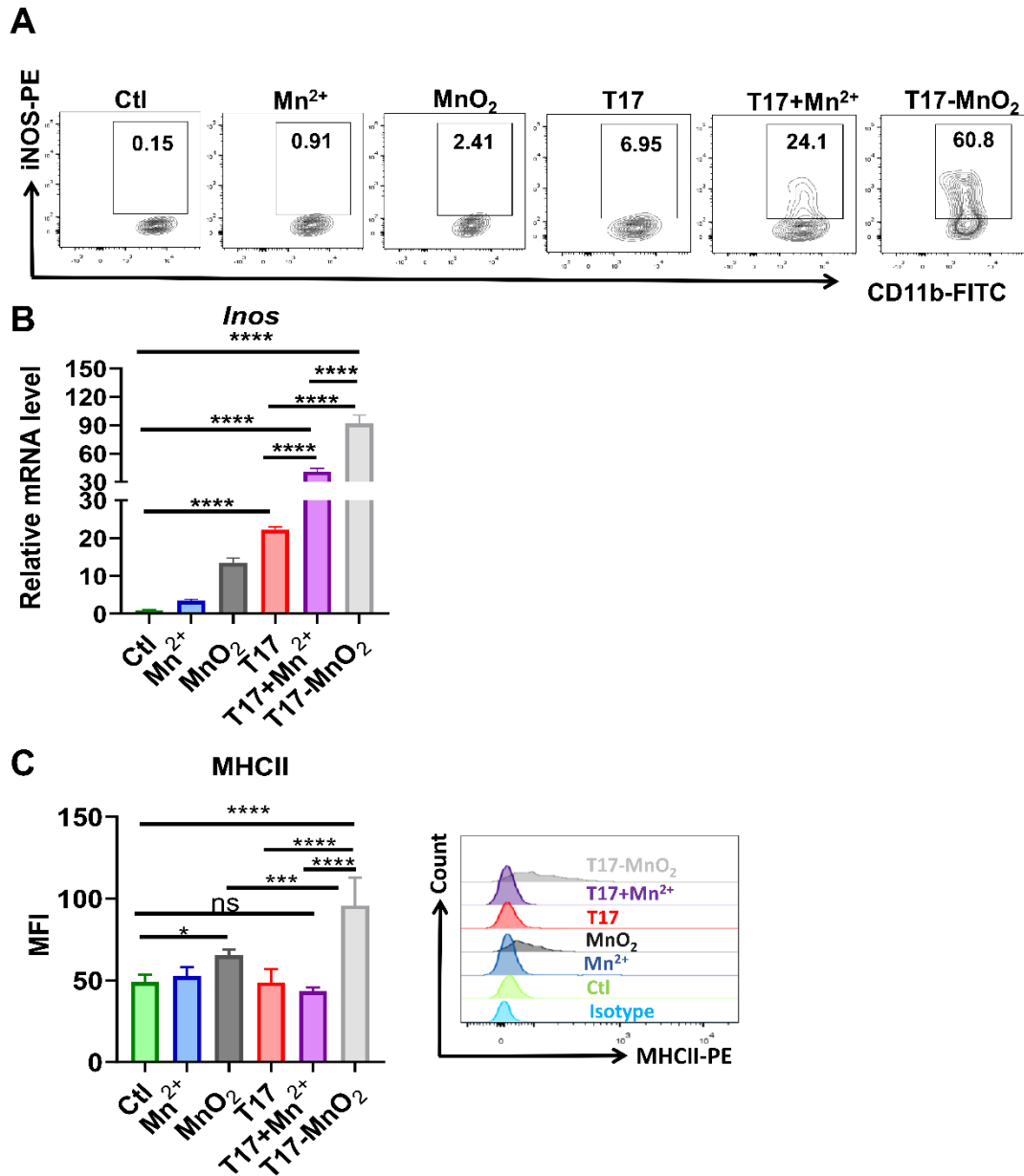
**Figure S1** Synthesis of TDNs with different sizes. (A–E) The synthesis of TDNs with sizes of 7, 13, 26 and 37 bp was analyzed by agarose gel electrophoresis.



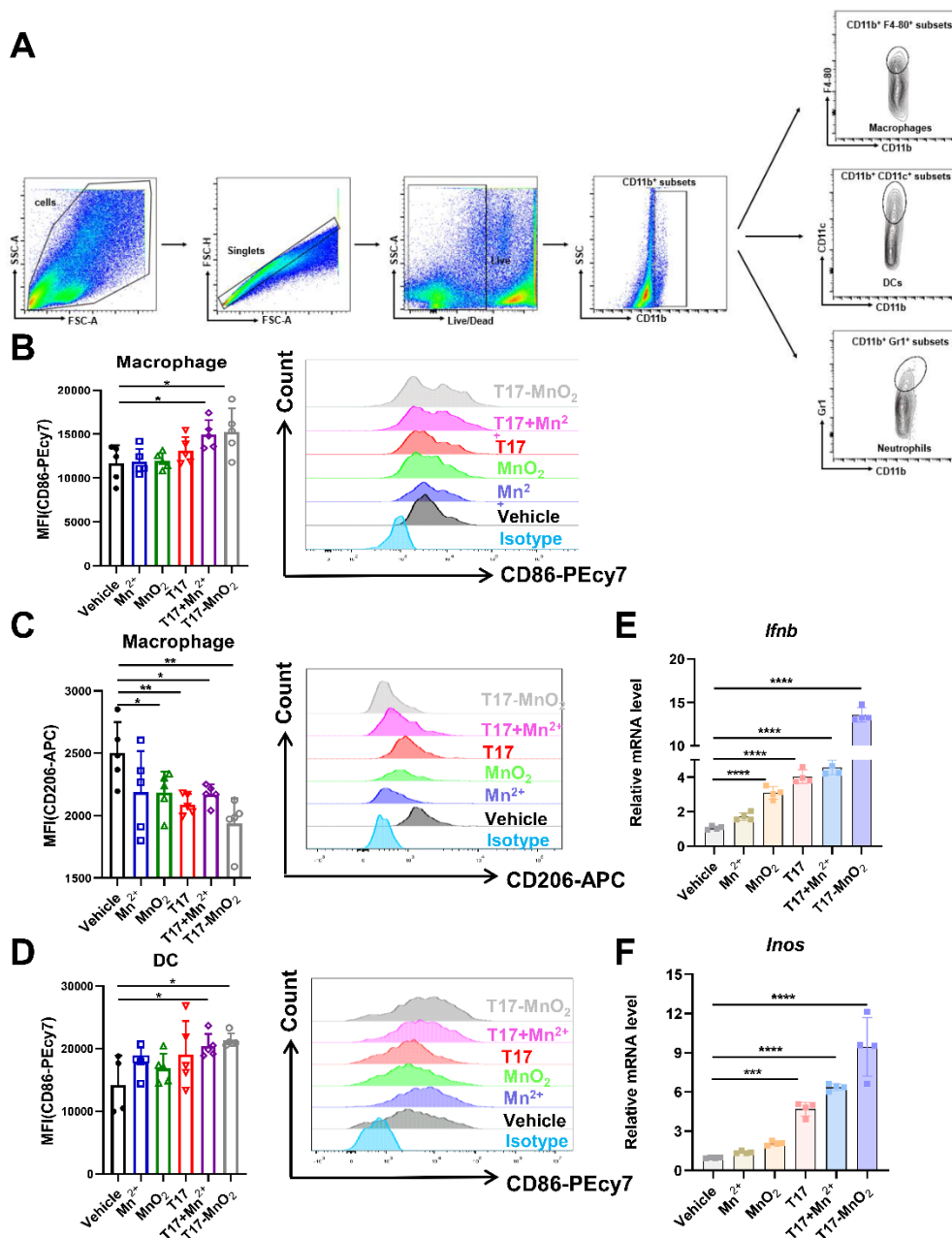
**Figure S2** Inflammatory cytokines expression in macrophages after T17 treatment. After treatment of T17 (200 nmol/L), the expression of (A) INOS, (B) TNF- $\alpha$ , (C) IL-1 $\beta$  in BMDMs was detected by flow cytometry, and the representative figures were shown. Data are presented as mean  $\pm$ SD ( $n=3$ ). \*\*\*\*  $P<0.0001$ .



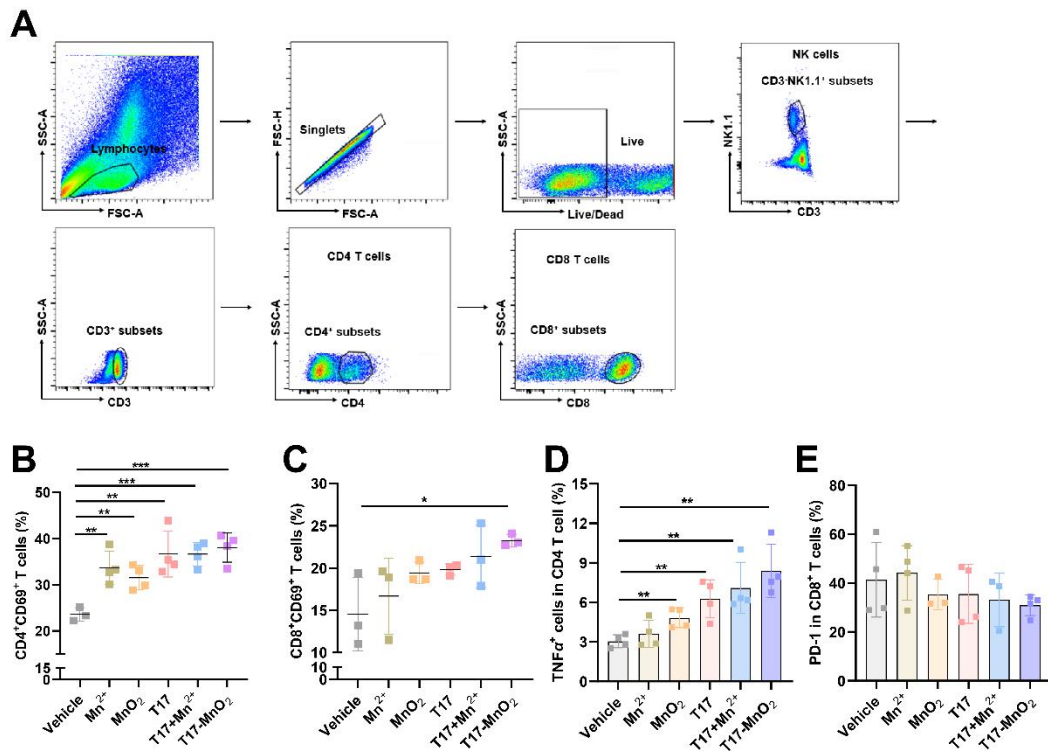
**Figure S3** The cytotoxicity of divalent metal cations in macrophages and cellular uptake of T17 by T cells. (A) After treatment with different divalent metal cations including  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$  and  $\text{Mn}^{2+}$  for 24 h, cell viability of BMDMs was detected by CCK-8 assay. Cellular uptake of T17-FITC (200 nmol/L) by (B)  $\text{CD4}^{+}$  T cells and (C)  $\text{CD8}^{+}$  T cells were detected by flow cytometry. Data are presented as mean  $\pm$ SD ( $n=3$ ). \*  $P<0.05$ , \*\*  $P<0.01$ , \*\*\*  $P<0.001$ , \*\*\*\*  $P<0.0001$ , ns, not significant.



**Figure S4** T17-MnO<sub>2</sub> promoted the M1 polarization and the antigen-presentation of BMDMs. After treatment of MnCl<sub>2</sub> (50  $\mu$ mol/L), MnO<sub>2</sub> (50  $\mu$ mol/L), T17 (200 nmol/L), T17+MnCl<sub>2</sub>, and T17-MnO<sub>2</sub> for 24 h, the protein level (A) and mRNA level (B) of INOS was detected by flow cytometry and real-time PCR, and the protein level of MHCII (C) was detected by flow cytometry. Data are presented as mean  $\pm$ SD ( $n=3$ ). \*  $P<0.05$ , \*\*\*  $P<0.001$ , \*\*\*\*  $P<0.0001$ , ns, not significant.



**Figure S5** T17-MnO<sub>2</sub> promotes M1 polarization *in vivo*. Hepa1-6 growth and mice treated with vehicle control (0.9% NaCl), Mn<sup>2+</sup> (50 mmol/L), MnO<sub>2</sub> (2 mmol/L), T17 (3 μmol/L), T17 plus Mn<sup>2+</sup> or T17-MnO<sub>2</sub>. Mice were sacrificed and tumor-infiltrating leukocytes were analyzed by flow cytometry at Day 20. (A) Representative gating strategies of myeloid cells. (B) Representative flow cytometry histogram figure (left) and quantification (right) of CD86 expression on macrophages, (C) CD206 expression on macrophages and (D) CD86 expression on DCs. The mRNA level of *Ifnb* (E) and *Inos* (F) was detected by real-time PCR. Data are presented as mean ± SD (*n*=4). \**P*<0.05, \*\**P*<0.01, \*\*\**P*<0.001, \*\*\*\**P*<0.0001.



**Figure S6** T17-MnO<sub>2</sub> stimulates CD4<sup>+</sup> and CD8<sup>+</sup> T cell activation. Mice were inoculated with Hepa1-6 cells on Day 0. On Day 8, 14, and 17 mice were treated with either vehicle control (0.9% NaCl), Mn<sup>2+</sup> (50 mmol/L), MnO<sub>2</sub> (2 mmol/L), T17 (3 μmol/L), T17 plus Mn<sup>2+</sup> or T17-MnO<sub>2</sub> by i.t. administration. On Day 20, mice were sacrificed and tumor infiltrated leukocytes were analyzed by flow cytometry. (A) Representative gating strategies of NK cells, CD4<sup>+</sup> T cells and CD8<sup>+</sup> T cells. (B, C) Quantification of tumor-infiltrating CD4<sup>+</sup>CD69<sup>+</sup> T cells and CD8<sup>+</sup>CD69<sup>+</sup> T cells. (D) TNF-α<sup>+</sup> in CD4<sup>+</sup> T cells. (E) Quantification of PD-1<sup>+</sup> cells in tumor-infiltrating CD8<sup>+</sup> T cells. Data are presented as mean ±SD (n=4). \* P<0.05, \*\* P<0.01 \*\*\* P<0.001.