## **Supplementary Figures**



**Supplementary figure 1. Visualization of Cre recombinase activities in the pancreas.** a) Scheme of tamoxifen injection. b) RFP staining showing positive cells (arrow) in the pancreas. Scale bar: 50µm. c) Histology of pancreata 3 weeks after the starting of the first tamoxifen injection. Scale bar: 100µm.



Supplementary figure 2. Long-term tamoxifen diet induces spontaneous pancreatitis in NEMO<sup>ACol1a2</sup> animals. a) H&E staining of pancreas. Scale bar: 1 mm. b) Amylase activities measured from serum (n=6). c) H&E staining of different organs. Scale bar: 100μm. d) Periportal inflammation in the liver of NEMO<sup>ΔCol1a2</sup> mice. Scale bar: 100μm. The n number represents biological replicates.



Supplementary figure 3. Characterization of animals after caerulein and tamoxifen injections. a) Measurement of the pancreas weight as a ratio of the body weight from the salineand caerulein-injected animals ( $n\geq3$ ). b) Ratio of pancreas weight to the body weight from animals at 3 weeks (n=8). c) Amylase and lipase activities measured from serum ( $n\geq4$ ). d) Cleaved caspase 3 staining indicating the presence of apoptotic cells. Arrow: apoptotic acinar cells; arrow head: apoptotic non-parenchemal cells. Scale bar: 100µm. e) Quantification of the number of acinar cells per field on pancreas sections (3 week) (n=4). f) Low magnification of pancreas histology. Scale bar: 200µm. g) Co-staining of PDGFR $\beta$  and cleaved caspase 3 to identify apopotic PSCs. Arrow: apoptotic PSCs. Scale bar: 25 µm. Whiskers: Min to Max. T-test (two-tailed): \*\*\*p<0.001. All n numbers represent biological replicates.



Supplementary figure 4. FACS analyses on the infiltrating cells from pancreata and immune cells in the peripheral circulation. Cells were isolated from pancreata (a) using gradient centrifugation or from blood (b) of NEMO<sup>WT</sup> and NEMO<sup> $\Delta$ Col1a2</sup> animals .



Supplementary figure 5. Eosinophils are recruited to the pancreas of NEMO<sup> $\Delta$ Col1a2</sup> mice after long-term tamoxifen diet. a) Siglec-F staining showing eosinophil infiltration in the pancreas. DAPI: nuclei. Scale bar: 50µm. b-c) Expression of the chemokine CCL24 and eosinophil markers (CCR3, Siglec-F and PRG2) from pancreatic tissue assessed by qPCR (n=6). Whiskers: Min to Max. Mann-Whitney (two-tailed): #p<0.05; ##p<0.01. T-test (two-tailed): \*p<0.05. The n number represents biological replicates.



Supplementary figure 6. Analysis of the RFP reporter mouse lines. RFP stainings on the NEMO<sup>WT</sup>, NEMO<sup> $\Delta$ Col1a<sup>2</sup></sup>, NEMO<sup>WT, RFP and NEMO<sup> $\Delta$ Col1a<sup>2</sup>,RFP</sup> pancreata 3 weeks after caerulein and tamoxifen injections. Scale bar: 50µm.</sup>

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Supplementary figure 7. Comparison of the gene expression profiles between NEMO<sup> $\Delta Col1a^2$ </sup> mice and human AIP samples. DNA microarray performed on the pancreatic samples of NEMO<sup>WT</sup> and NEMO<sup> $\Delta Col1a^2$ </sup> animals collected at the 3-week time point. a) GSEA was performed on the microarray results of the NEMO<sup> $\Delta Col1a^2$ </sup> group with the NEMO<sup>WT</sup> group as a reference (n=3). b) Analysis of microarray data from AIP patients (n=4) using GSEA with AICP patients (n=5) as a reference. NES: Net enrichment score. FDR: False discovery rate.

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Supplementary figure 8. PSC<sup> $\Delta NEMO$ </sup> cells show upregulation of ECM components. Expressions of genes involved in ECM modifications in PSCs analyzed by qPCR (n $\geq$ 3). Whiskers: Min to Max. T-test (two-tailed): \*p<0.05; \*\*p<0.01. The n number represents biological replicates.







Chan LK et al., Commun Biol 2022.











## Supplementary Tables

Primary antibody	Dilutions	Company	
rabbit anti-α-amylase	1:400	Sigma (A8273)	
goat anti-CK19	1:100	Santa Cruz (sc-33111)	
rat anti-CD45	1:100	BD Pharmingen (#550539)	
mouse anti-αSMA	1:100	Millipore (CBL171)	
rabbit anti-RFP	1:100	Rockland (600-401-379)	
rat anti-B220	1:100	BioLegend (103247)	
rat anti-CD3	1:100	BioLegend (100201)	
rat anti-CD4	1:100	BD Pharmingen (550278)	
rat anti-CD8	1:100	BD Pharmingen (550281)	
rabbit anti-vimentin	1:100	abcam (ab7783)	
rat anti-SiglecF	1:100	Miltenyi Biotec (130-102-167)	
goat anti-nestin	1:100	Santa Cruz (sc-21248)	

Supplementary table 1. List of primary antibodies for histological staining.

Antibody	Dilutions	Company	
CD11c (BV510)	1:100	BioLegend (117337)	
CD11b (APC)	1:100	BioLegend (101212)	
CD45 (APC/Cy7)	1:400	BD Pharmingen (557659)	
Ly6G (BV421)	1:100	BioLegend (127627)	
CCR3 (PerCP)	1:100	R&D System (FAB729C-025)	
SiglecF (PE-Vio770)	1:25	MACS (130-102-167)	
CD4 (FITC)	1:100	MACS (120-001-955)	
CD4 (PE)	1:100	BioLegend (100407)	
IL17A (APC)	1:50	MACS (130-103-027)	
F4/80 (AF448)	1:50	Bio-Rad (MCA497A448T)	
CD3ε (PE/Cy7)	1:100	BioLegend (100320)	

Supplementary table 2. List of antibodies for flow cytometry.

Primary antibodies	Dilutions	Company	
NEMO	1:1000	BD Transduction Laboratories (#611306)	
GAPDH	1:1000	Santa Cruz (sc-25778)	
TOM20	1:1000	Santa Cruz (sc-11415)	
p-JNK	1:1000	Cell Signaling (9255S)	
JNK	1:1000	Santa Cruz (sc-7345)	
p-p38	1:1000	Cell Signaling (9211S)	
p38	1:1000	Cell Signaling (9212)	
p-ERK	1:1000	Cell Signaling (4377)	
ERK2	1:1000	Santa Cruz (sc-154)	

p-STAT3	1:1000	Cell Signaling (9145S)
STAT3	1:1000	Cell Signaling (4904S)

Supplementary table 3. List of antibodies for western blot.

Gene	Primer Sequence	Gene	Primer Sequence
Col1a1	F 5' catgttcagctttgtggacct 3'	Icam1	F 5' cccacgctacctctgctc 3'
	R 5' gcagctgacttcagggatgt 3'		R 5' gatggatacctgagcatcacc 3'
Col1a2	F 5' agggcctgatggaaacaag 3'	Nfkbia	F 5' cagaacaacctgcagcagac 3'
	R 5' cccctctctctggaage 3'		R 5' tttcagaagtgcctcagcaat 3'
Col3a1	F 5' tcccctggaatctgtgaatc 3'	Ccl6	F 5' tetttateettgtggetgtee 3'
	R 5' tgagtcgaattggggagaat 3'		R 5' tggagggttatagcgacgat 3'
Vim	F 5' gtaccggagacaggtgcagt 3'	Ccl8	F 5' tcagcccagagaagctgac 3'
	R 5' ttetettecateteacgeate 3'		R 5' gggggatcttcagctttagtaca 3'
Cxcr4	F 5' tggaaccgatcagtgtgagt 3'	Ccl9	F 5' ggctgctcaggtttcactgt 3'
	R 5' gggcaggaagatcctattga 3'		R 5' agaacgatagggcggtaggt 3'
Fn1	F 5' gatgccgatcagaagtttgg 3'	Hprt	F 5' ggagcggtagcacctcct 3'
	R 5' ggttgtgcagatctcctcgt 3'		R 5' cctggttcatcatcgctaatc 3'
Tgfb1	F 5' tggagcaacatgtggaactc 3'	Ccl11	F 5' agageteeaagegettet 3'
	R 5' gtcagcagccggttacca 3'		R 5' gcaggaagttgggatgga 3'
Rpl13	F 5' cctgctgctctcaaggttgt 3'	Ccl12	F 5' ccatcagtcctcaggtattgg 3'
	R 5' ggtacttccacccgacctc 3'		R 5' cttccggacgtgaatcttct 3'
Il6	F 5' gctaccaaactggatataatcagga 3'	Ccl24	F 5' gtgcctgacctccagaacat 3'
	R 5' ccaggtagctatggtactccagaa 3'		R 5' gaggggatggtcacagaatc 3'
Prg2	F 5' ggagcgtctgctcttcatct 3'	Ccr2	F 5' acctgtaaatgccatgcaagt 3'
	R 5' cccctggaggacactcttct 3'		R 5' tgtcttccatttcctttgatttg 3'
Ifng	F 5' atctggaggaactggcaaaa3'	Ccr3	F 5' gaatcaaagagctggggtca 3'
	R 5' ttcaagacttcaaagagtctgagg 3'		R 5' caggaggccgatgatgaa 3'
Il12a	F 5' ccaggtgtcttagccagtcc 3'	Siglecf	F 5' gaacttacctggcactggtgtt 3'
	R 5' gcagtgcaggaataatgtttca 3'		R 5' gctccactctgcaggacttt 3'
Il4	F 5' cctgctcttctttctcgaatgt 3'	Ikbkg	F 5' ctggcttcaagtcacccaag 3'
	R 5' cacatecateteegtgeat 3'		R 5' gtgctaaagcaccaccagt 3'
<i>II5</i>	F 5' acattgaccgccaaaaagag 3'	Cpa1	F 5' agcagtetteggeaatgag 3'
	R 5' atccaggaactgcctcgtc 3'		R 5' cagaagtccaacttcaagtgctc 3'
<i>II17</i>	F 5' ggacagcccttctttgtctg 3'		
	R 5' tgctttttatatttcattacgtggtt 3'		

**Supplementary table 4.** List of primers for qPCR.