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Supplemental information

**DIPA-CRISPR is a simple and accessible method
for insect gene editing**

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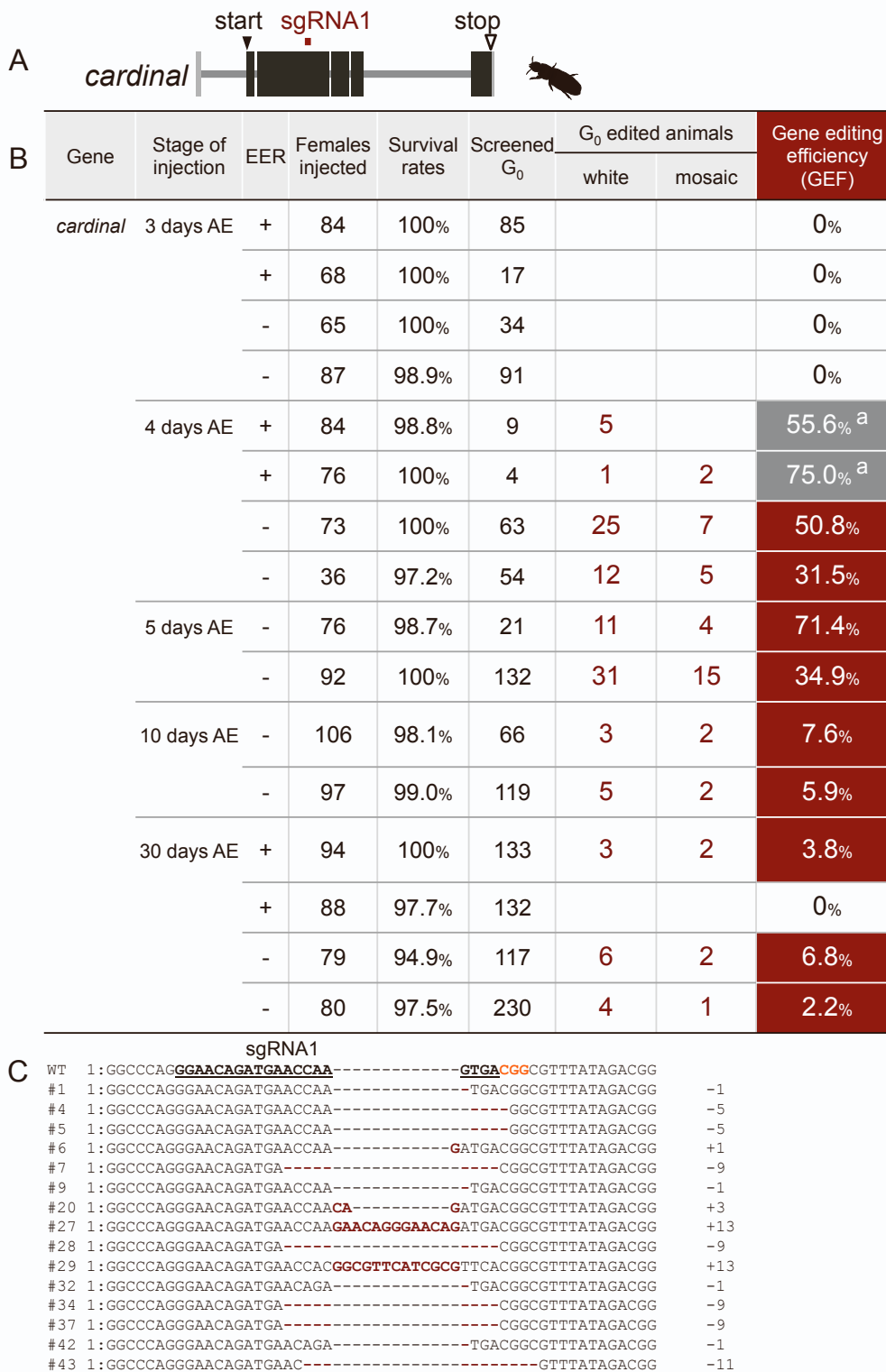


Figure S2. Disruption of *Tribolium castaneum cardinal* by DIPA-CRISPR. Related to Figures 3 and 4.

(A) The CRISPR target site of *T. castaneum cardinal* (XP_008200769). The sgRNA1 targeting the exon 3 of *cardinal* (Shirai and Daimon, 2020) was used.

(B) The detailed results of DIPA-CRISPR in *T. castaneum*. Cas9 ribonucleoprotein (RNP) solution containing 3.3 µg/µL Cas9 (IDT, Cat#1081059) and 1.3 µg/µL sgRNA were injected into adult females of selected days (i.e., 3, 4, 5, 10 or 30 days) after adult emergence (AE). The injected females were pooled, and the results are from the eggs laid during the first 24 h (females at 4, 5, 10 or 30 days AE) or 48 h (females at 3 days AE), as

they were too young to lay eggs during the first 24 h) after injection. The results from two independent experiments are shown. EER, presence (+) or absence (-) of an endosomal escape reagent saponin (100 ng/ μ L) in injection solution. a: the gene-editing efficiency (GEF) values are very high, but these values may not be reliable as they are calculated based on very small numbers of G_0 insects hatched and/or survived.

(C) The DNA sequences of *cardinal* mutant alleles in G_0 edited insects. Hemizygous G_0 males with white eyes were randomly chosen and subjected to direct Sanger sequencing of genomic PCR products. Each row represents each G_0 mutant, and the length of indel is shown on the right.

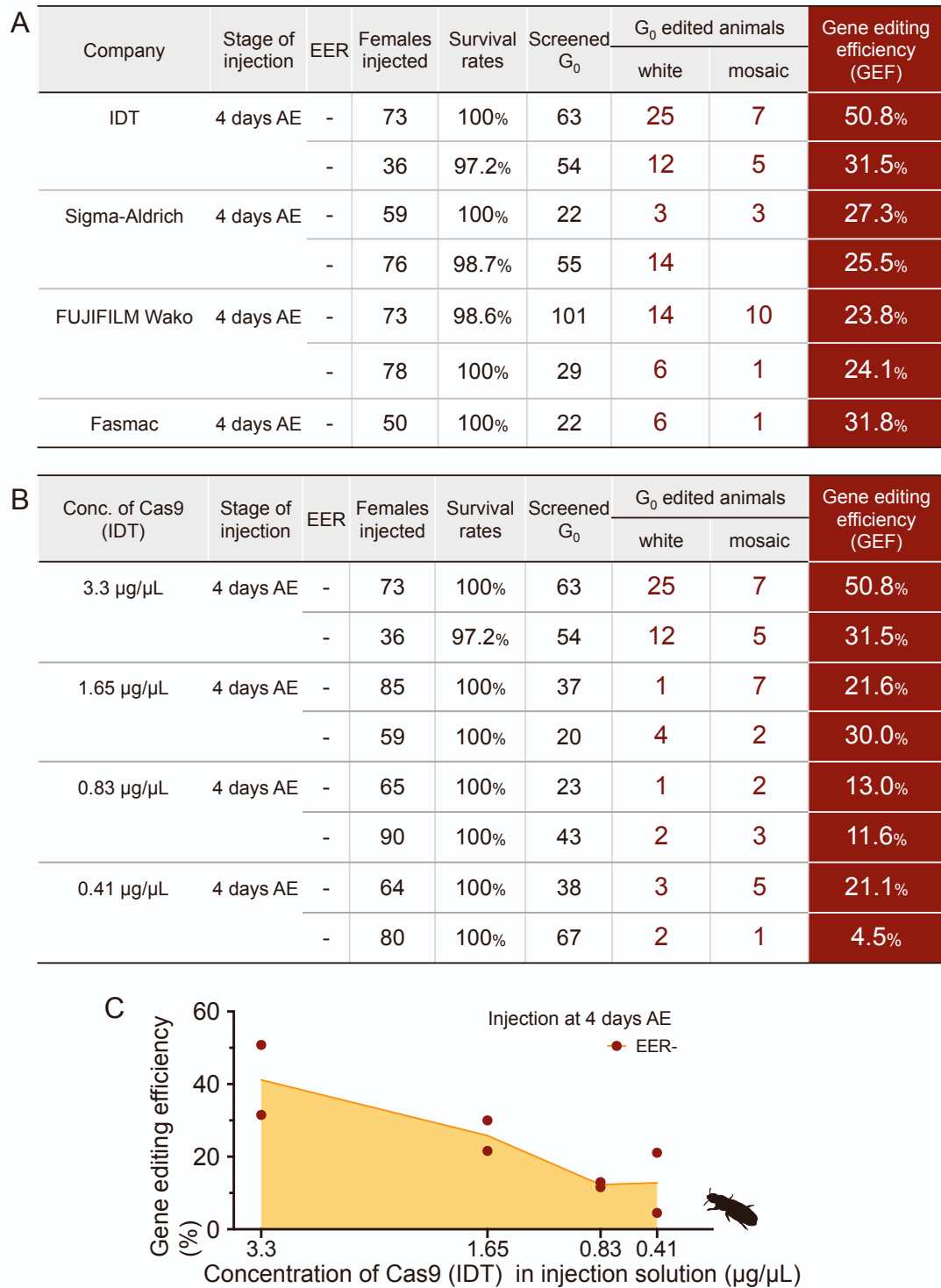


Figure S3. Performance of different Cas9 products and different doses of Cas9 in *Tribolium castaneum*. Related to Figures 3 and 4.

(A) Comparisons of Cas9 products from four vendors. Cas9 RNP solution containing 3.3 µg/µL Cas9 and 1.3 µg/µL sgRNA were injected into adult females of 4 days after adult eclosion (AE).

(B) A dilution series of Cas9 RNPs (diluted by water, the molar ratio of Cas9 and sgRNA was fixed to be 1 : 2) were injected into females.

(C) Gene editing efficiency shown in (B) was plotted against the concentration of Cas9 in injection solution. Each point represents the result of each replication.

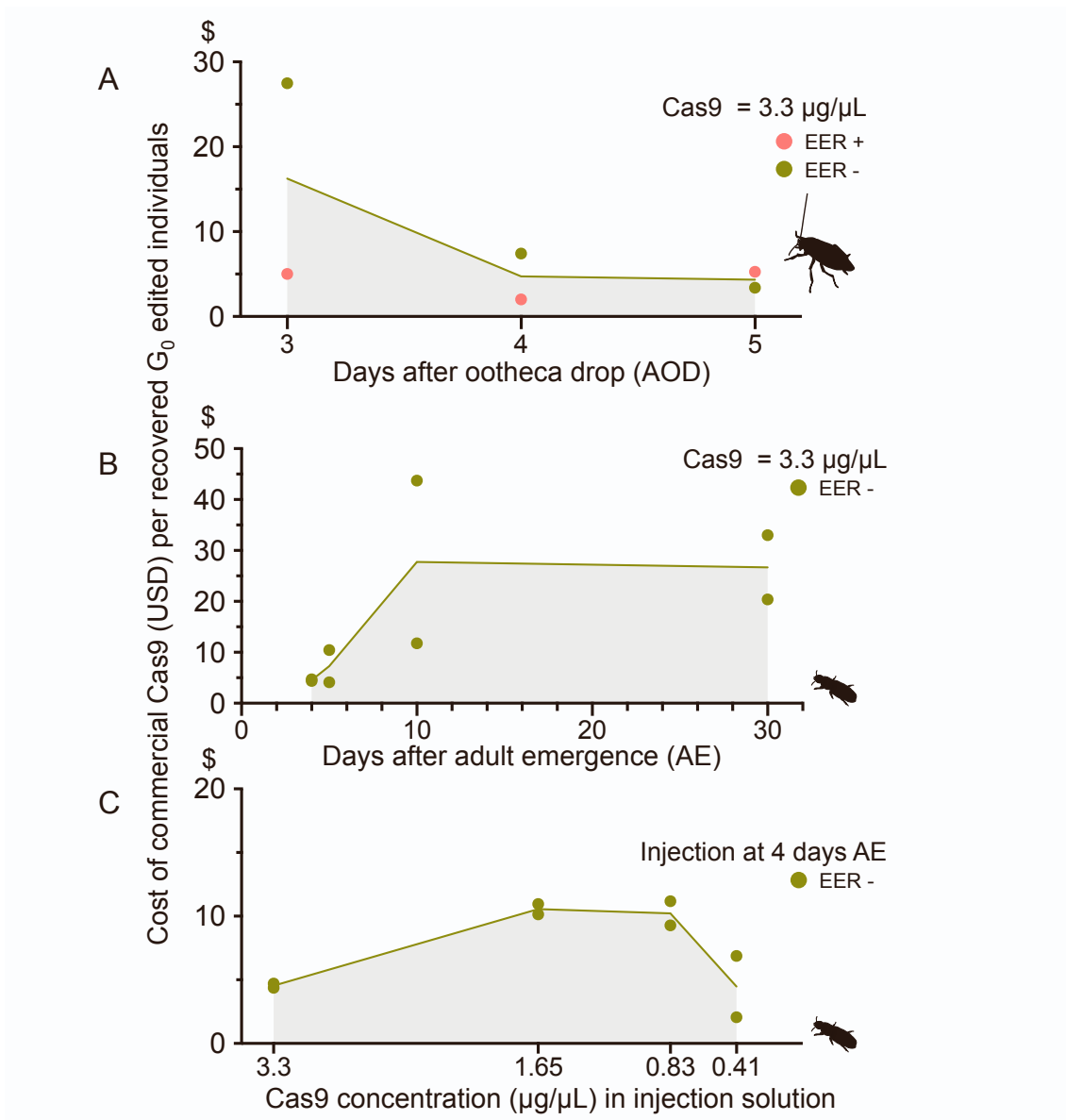


Figure S4. Cost of commercial Cas9 in DIPA-CRISPR. Related to Figures 1 and 3.

(A and B) Cost of commercial Cas9 (IDT, cat#10000735, 1.25 USD/µg) per recovered G_0 edited individuals was plotted against the time of injection for *B. germanica* (A) and *T. castaneum* (B). The results show that the cost can be significantly reduced by injecting at an appropriate timing in both species. Cost was calculated as follows: $1.25 \text{ (USD/}\mu\text{g)} \times \text{amount } (\mu\text{g)} \text{ of Cas9 injected per adult} \times \text{total number of adults injected} / \text{total number of } G_0 \text{ edited individuals recovered}$. Note that this calculation does not include the cost for sgRNA synthesis, as it greatly varies depending on the method (i.e., *in vitro* transcription or chemical synthesis).

(C) Relationship of the cost of Cas9 and the concentration of Cas9 in injection solution in *T. castaneum*. The results shown in Figure S3B were used for calculation.

Supplementary Table 1. DIPA-CRISPR experiments in *Drosophila melanogaster*. Related to Figure 4.

Gene	sgRNA	Stage of injection	EER	Females injected	Survival Rates	Screened G ₀	G ₀ edited animals		Gene editing efficiency (GEF)	
							white	mosaic		
<i>white</i>	chemical synthesis	2 h AE	-	59	81.4%	212			0%	
		2 h AE	-	29	96.6%	101			0%	
		10 h AE	-	31	87.1%	2			0%	
		10 h AE	-	26	96.2%	33			0%	
		10 h AE	-	80	96.3%	270			0%	
		24 h AE	-	61	85.2%	481			0%	
		24 h AE	-	71	91.5%	426			0%	
		2 d AE	-	47	53.2%	324			0%	
		2 d AE	-	58	77.6%	437			0%	
		4 d AE	-	53	60.4%	329			0%	
		4 d AE	-	63	76.2%	421			0%	
		7 d AE	-	62	66.1%	841			0%	
		7 d AE	-	65	80.0%	573			0%	
		Random	Chl 2 mM	113	93.1%	891			0%	
		in vitro transcription	Random	-	71	91.5%	340			0%
			Random	Chl 0.5 mM	74	95.9%	417			0%
			Random	Chl 2 mM	70	88.6%	662			0%

Cas9 ribonucleoprotein (RNP) solution containing 3.3 µg/µL Cas9 (IDT, Cat#1081059) and 1.3 µg/µL sgRNA (a mixture of sgRNA1 and sgRNA2) were injected into adult females of selected hours or days after adult emergence (AE), or females that were randomly chosen from vials (Random). sgRNAs were purchased (chemical synthesis) or synthesized by in vitro transcription. The injected females were pooled, and the results are from the eggs laid during the first 48 h after injection. EER, presence or absence (-) of an endosomal escape reagent chloroquine (Chl) in injection solution.