

## Supporting Information

# Interactions of an Anti-sickling drug with Hemoglobin in Red Blood Cells from a Patient with Sickle Cell Anemia

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## Experimental procedures

### *Proteins and chemicals*

The GBT440 incubated blood samples were centrifuged at 1000 g for 10 minutes. The supernatant was discarded, and the RBC was washed with 3-fold volumes of PBS by centrifuging at 1000 rpm for 10 minutes in order to remove excess GBT400. The washing step proceeded 3 times. Then 50  $\mu$ L of RBC was added 200  $\mu$ L of water and then well mixed and incubated at room for 20 minutes to lyse the RBC. The 250  $\mu$ L of Hb lysate solution was added 25  $\mu$ L of condensed PBS ( $\times 10$ , Fisher Sci), and then centrifuged at 13000 g for 20 minutes to remove the debris. For the NaBH<sub>3</sub>CN stabilized Hb samples, the Hb lysates (4mM) were incubated with NaBH<sub>3</sub>CN (50 mM) at room temperature for 20 minutes before any analyses. Hb concentration was measured based on the absorbencies at  $\lambda = 541, 576, 630, \text{ and } 700 \text{ nm}$  using recently published extinction coefficients <sup>1</sup>. All the Hb concentrations in this study are reported based on heme concentration.

### *Reverse phase high performance liquid chromatograph (RP-HPLC)*

RP-HPLC was performed with a Zorbax 300 SB C3 column (4.6x250 mm) coupled to a Waters HPLC system consisting of a Waters 626 pumps, 2487 dual-wavelength detector and a 600s controller installed with Empower 2 (Waters Corp, Milford, MA) <sup>2</sup>. 20  $\mu$ g of Hb in 25  $\mu$ L of water was loaded on C3 column equilibrated with 35% acetonitrile containing 0.1% TFA. Globin chains were eluted with a gradient of 35-50% acetonitrile within 100 min at a flow rate of 1 mL/min. The eluent was monitored at 280nm for globin chains and at 405 nm for the heme components.

### *LC/MS/MS Analysis*

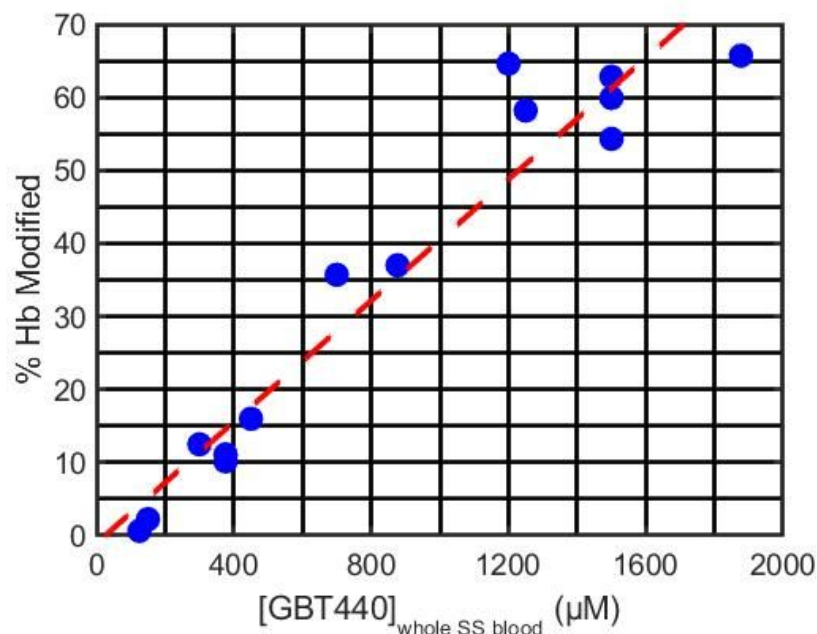
Hb samples from patient hemolysate (with and without GBT440) were digested with trypsin, desalted and analyzed by mass spectrometry using our previously described method<sup>3</sup>. Briefly, tryptic peptides were analyzed by reverse phase liquid chromatography mass spectrometry (RP LC/MS/MS) using an Easy nLC II Proxeon nanoflow HPLC system coupled online to a Q-Exactive Orbitrap mass spectrometer (Thermo Scientific). Data were acquired using a top10 method (for 60 minutes), dynamically choosing the most abundant precursors (scanned at 400-2000 m/z) from the survey scans for HCD fragmentation.

Mass spectrometry was used to confirm the presence of GBT440 attached peptides for each patient hemolysate same and by searching data against the Swiss-Prot Human database (release 2014\_03, containing 542782 sequence entries) using the Mascot (version 2.4) search engine (Matrix Sciences, London, UK) as described previously<sup>3</sup>. Variable modifications for all MS/MS data including the following masses: methionine oxidation (+16) and GBT440 binding to N-terminal peptides and lysine residues (+337). Mascot output files were analyzed using the software Scaffold 4.2.0 (Proteome Software Inc.). Hb peptide identifications were accepted if they could be established at greater than 99.9% probability and contained at least 2 identified peptides. Probabilities were assigned by the Protein Prophet algorithm<sup>4</sup>.

### ***Intact Mass Analysis***

Subunits of intact HbS from patient hemolysate (incubated with GBT440 (1;0.3 and 1:1 ratios) were isolated using RP-HPLC prior to LC-MS analysis. Samples were analyzed using a Q-Exactive as described above with the instrument configured to acquire data in Full-MS mode. Isotopically resolved charge state envelopes (multiply charged ions of intact monomers) representing mass spectra of intact Hb subunits were deconvoluted using the protein deconvolution software Xtract (Thermo Scientific). The deconvoluted mass spectrum yielded a measured molecular mass that was within 0.025 Da (3 ppm) of the calculated value.

**Figure S1**



**Figure S1.** Percent hemoglobin modified for whole SS blood samples treated with varying concentrations of GBT440. Blood was drawn from four SS patients with hematocrits between 21% and 28%. The percent hemoglobin modified for each sample was determined by the linear combination analysis in Figure 1 of the main text. Briefly, the best linear least squares fit was found between the oxygen affinity curve of each sample treated with GBT440 and an untreated sample and a 1:50 erythrocyte suspension treated with 200  $\mu$ M GBT440, a concentration found by Li *et al* to modify all hemoglobin, from the same donor. From a linear regression analysis, the following relationship was found with an  $R^2$  of 0.93: **% Hb Modified =  $0.0415 \times [\text{GBT440}]_{\text{whole SS blood}} - 1.0391$** . From this, it was concluded that GBT440 shifts the hemoglobin quaternary equilibrium to the R conformation in SS erythrocytes in a linearly dose dependent manner throughout the range of modification desirable for therapeutic intervention.

References:

- [1] Meng, F., and Alayash, A. I. (2017) Determination of extinction coefficients of human hemoglobin in various redox states, *Analytical Biochemistry* 521, 11-19.
- [2] Kassa, T., Jana, S., Strader, M. B., Meng, F., Jia, Y., Wilson, M. T., and Alayash, A. I. (2015) Sick Cell Hemoglobin in the Ferryl State Promotes  $\beta$ Cys-93 Oxidation and Mitochondrial Dysfunction in Epithelial Lung Cells (E10), *Journal of Biological Chemistry* 290, 27939-27958.
- [3] Strader, M. B., Hicks, W. A., Kassa, T., Singleton, E., Soman, J., Olson, J. S., Weiss, M. J., Mollan, T. L., Wilson, M. T., and Alayash, A. I. (2014) Post-translational Transformation of Methionine to Aspartate Is Catalyzed by Heme Iron and Driven by Peroxide: A NOVEL SUBUNIT-SPECIFIC MECHANISM IN HEMOGLOBIN, *The Journal of Biological Chemistry* 289, 22342-22357.
- [4] Keller A, N. A., Kolker E, Aebersold R. (2002) Empirical statistical model to estimate the accuracy of peptide identifications made by MS/MS and database search., *Anal Chem.* 74, 10.