

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

- | | |
|-----|-----------|
| n/a | Confirmed |
|-----|-----------|
- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
 - A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
 - The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
 - A description of all covariates tested
 - A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
 - A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
 - For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
 - For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
 - For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
 - Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection	Illumina, HiSeqXten (Chromium WGS), Omni-C
Data analysis	alienness, version 2, link http://alienness.sophia.inra.fr/cgi/index.cgi assembly-stats, version 1.0.1, link https://github.com/rjchallis/assembly-stats AUGUSTUS, version 3.3, link http://bioinf.uni-greifswald.de/augustus/binaries/old/augustus-3.3.tar.gz blast+, version 2.2.31+, link https://ftp.ncbi.nlm.nih.gov/blast/executables/blast+/2.2.31/ BUSCO, version 5, link https://gitlab.com/ezlab/busco bwa, version 2.4.2, link https://github.com/lh3/bwa CAFE5, version 5.0.0, link https://github.com/hahnlab/CAFE5/releases/tag/v5.0 clusterProfiler, version 3.16.1, link https://bioconductor.org/packages/release/bioc/html/clusterProfiler.html cutadapt, version 1.10, link https://github.com/marcelm/cutadapt/tree/v1.10 DSK, version 2.1.0, link http://gatb-tools.gforge.inria.fr/versions/bin/dsk-2.1.0-Linux.tar.gz eggno-mapper, version 2.1.7, link https://github.com/eggno-mapper/eggno-mapper/releases/tag/2.1.7 FastQC, version 0.11.8, link https://www.bioinformatics.babraham.ac.uk/projects/fastqc/fastqc_v0.11.8.zip funannotate, version 1.7.4, link https://funannotate.readthedocs.io/en/latest/ genomertools, version 1.5.8, link http://genomertools.org/pub/genomertools-1.5.8.tar.gz gggenes, version 0.4.1, link https://cran.r-project.org/web/packages/gggenes/index.html ggplot2, version 3.3.5, link https://cran.r-project.org/web/packages/ggplot2/index.html ggtree, version 3.2.1, link https://bioconductor.org/packages/release/bioc/html/ggtree.html InterProScan, version 5.26-65.0, link https://github.com/ebi-pf-team/interproscan/wiki/Interproscan5_26_65_ReleaseNotes iqtree, version 2.2.0, link http://www.iqtree.org KAT, version 2.1.1, link https://github.com/TGAC/KAT Kraken2, version 2.0, link https://github.com/DerrickWood/kraken

MAFFT , version 7.453, link <https://mafft.cbrc.jp/alignment/software/source.html>
 MEGA, version 7, link <https://www.megasoftware.net/>
 minion, version 15-065, link <http://wwwdev.ebi.ac.uk/enright-dev/kraken/reaper/src/reaper-15-065.tgz>
 mirDeep2, version 2_0_0_8, link <https://github.com/rajewsky-lab/mirdeep2>
 MITE_Hunter, version 11-2011, link http://target.iplantcollaborative.org/mite_hunter/MITE%20Hunter-11-2011.zip
 MUSCLE, version 3.8.31, link <https://www.drive5.com/muscle/downloads.htm>
 OrthoFinder, version 2.5.2, link <https://github.com/davidemms/OrthoFinder/releases/tag/2.5.2>
 OrthoMCL, version 2.0.9, <http://orthomcl.org/common/downloads/software/v2.0/orthomclSoftware-v2.0.9.tar.gz>
 PAML, version 3.1, link <http://abacus.gene.ucl.ac.uk/software/paml.html>
 PASA, version 2.1.0, link <https://github.com/PASAPipeline/PASAPipeline>
 pathview, version 1.34.0, link <https://bioconductor.org/packages/release/bioc/html/pathview.html>
 Prokka, version 1.12-beta, link <https://github.com/tseemann/prokka>
 ProtTest, version 3.4, link <https://github.com/ddarriba/prottest3>
 R, version 3.6.3, link <https://cran.r-project.org/src/base/R-3/>
 R, version 4.1.0, link <https://cran.r-project.org/>
 r8s, version 1.8.1, link <https://sourceforge.net/projects/r8s/files/r8s1.81.tar.gz>
 RaxML, version 8.2.4, link <https://github.com/stamatak/standard-RAxML>
 RECON, version 1.08, link <http://www.repeatmasker.org/RepeatModeler/RECON-1.08.tar.gz>
 RepeatModeler, version 1.0.11, link <http://www.repeatmasker.org/RMDownload.html>
 Supernova, version 2.1.1, link <https://github.com/major/supernova/archive/v2.1.0.tar.gz>
 SyMAP, version 4.2, link <http://www.agcol.arizona.edu/software/symap/index.html>
 TransDecoder, version 5.3.0, link <https://github.com/TransDecoder/TransDecoder>
 TransposonPSI, version 08222010, link https://sourceforge.net/projects/transposonpsi/files/TransposonPSI_08222010.tgz/download
 trimAl, version 1.4, link <https://github.com/scapella/trimal>
 Trinity, version 2.9.1, link <https://github.com/trinityrnaseq/trinityrnaseq>
 Custom codes, link <https://github.com/xieyichun50/Myriapod-genomes>

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

The final assemblies were submitted to NCBI Assembly under accession numbers WWPM000000000 (Glomeris maerens) [<https://www.ncbi.nlm.nih.gov/nucleotide/WWPM000000000>], JAAFCF000000000 (Helicorhormorpha holstii) [<https://www.ncbi.nlm.nih.gov/nucleotide/JAAFCF000000000>], WWPL000000000 (Anaulaciulus tonginus) [<https://www.ncbi.nlm.nih.gov/nucleotide/WWPL000000000>], JAAIVG000000000 (Niponia nodulosa) [<https://www.ncbi.nlm.nih.gov/nucleotide/JAAIVG000000000>], JAAACE000000000 (Trigoniulus corallinus) [<https://www.ncbi.nlm.nih.gov/nucleotide/JAAACE000000000>], JAHWFP000000000 (Lithobius niger) [<https://www.ncbi.nlm.nih.gov/nucleotide/JAHWFP000000000>], JAHWFO000000000 (Rhysida immarginata) [<https://www.ncbi.nlm.nih.gov/nucleotide/JAHWFO000000000>], and JAFIDM000000000 (Thereuonema tuberculata) [<https://www.ncbi.nlm.nih.gov/nucleotide/JAFIDM000000000>] in NCBI. The raw reads generated in this study were deposited to the NCBI database under the BioProject accessions PRJNA598061 (Glomeris maerens) [<https://www.ncbi.nlm.nih.gov/bioproject/PRJNA598061>], PRJNA564202 (Helicorhormorpha holstii) [<https://www.ncbi.nlm.nih.gov/bioproject/PRJNA564202>], PRJNA598060 (Anaulaciulus tonginus) [<https://www.ncbi.nlm.nih.gov/bioproject/PRJNA598060>], PRJNA606398 (Niponia nodulosa) [<https://www.ncbi.nlm.nih.gov/bioproject/PRJNA606398>], PRJNA564195 (Trigoniulus corallinus) [<https://www.ncbi.nlm.nih.gov/bioproject/PRJNA564195>], PRJNA738717 (Lithobius niger) [<https://www.ncbi.nlm.nih.gov/bioproject/PRJNA738717>], PRJNA701115 (Rhysida immarginata) [<https://www.ncbi.nlm.nih.gov/bioproject/PRJNA701115>], and PRJNA699399 (Thereuonema tuberculata) [<https://www.ncbi.nlm.nih.gov/bioproject/PRJNA699399>]. The genome annotation files were deposited in the Figshare (<https://doi.org/10.6084/m9.figshare.15088722>). The scripts for carrying out analyses of this study were deposited in github (<https://github.com/xieyichun50/Myriapod-genomes>). The databases are available for download from the following websites: eggNOG http://eggnog5.embl.de/download/eggnog_5.0/, GO <http://geneontology.org/>, KEGG <https://www.genome.jp/kegg/pathway.html>, and KOG <https://www.hsls.pitt.edu/obrc/index.php?page=URL1144075392>. Source data are provided with this paper.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

To avoid sequence polymorphism among individuals of the same species, only one individual of each species was chosen and harvested for genome sequencing of *Lithobius niger*, *Rhysida immarginata*, *Thereuonema tuberculata*, *Anaulaciulus tonginus*, *Glomeris maerens* and *Niponia nodulosa*.

For transcriptome sequencing, one individual was chosen and harvested in *Lithobius niger*, *Rhysida immarginata*, *Anaulaciulus tonginus*,

Glomeris maerens and Niponia nodulosa. For the transcriptomes of Thereuonema tuberculata, various number of individuals (5 eggs, 5 stadium I, 5 stadium II, 3 stadium III, 1 stadium IV, 1 stadium V, 1 stadium VI and 1 adult) were chosen and pooled for transcriptome sequencing.

Data exclusions No data were excluded from analysis.

Replication Genomic and the transcriptomic sequencing were not replicated.

Randomization Randomization is not applicable to this study.

Blinding Blinding was not relevant for this study

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

- | | |
|-------------------------------------|---|
| n/a | Included in the study |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Antibodies |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Eukaryotic cell lines |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology and archaeology |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Animals and other organisms |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Human research participants |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Clinical data |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Dual use research of concern |

Methods

- | | |
|-------------------------------------|---|
| n/a | Included in the study |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Flow cytometry |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals This study did not involve laboratory animals.

Wild animals Adult *R. immarginata*, *L. niger* and *A. tonginus* (10 for each sex) were collected in an agricultural garden of New Asia College, The Chinese University of Hong Kong. Mixed sexes and different developmental stages of 30 adult *T. tuberculata*, 20 adult *G. maerens* and 15 adult *N. nodulosa* were purchased from an exotic pet shop in Hong Kong. Live animals were collected, and each species was contained in a sealed and air-ventilated plastic box during transportation to the laboratory. The remaining animals after the experiment were kept alive in culture under laboratory conditions.

Field-collected samples Alive adult *R. immarginata*, *L. niger* and *A. tonginus* (10 for each sex) were collected in an agricultural garden of New Asia College, The Chinese University of Hong Kong. The animals were all caught by hand-picking in a soil ground and under flowerpots during the daytime in the summer afternoon. The remaining animals after experiments were kept alive in culture under laboratory conditions.

Ethics oversight The study did not require an ethical approval.

Note that full information on the approval of the study protocol must also be provided in the manuscript.