## **Supplementary Information**

# Ex vivo-expanded human CD19<sup>+</sup>TIM-1<sup>+</sup> regulatory B cells suppress immune responses *in vivo* and are dependent upon the TIM-1/STAT3 axis

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a) Summarised data of expansion factor of expBreg when expanded at a 1:1 CD154<sup>+</sup> CHO cell-CD19<sup>+</sup> B cell ratio with different cytokine combinations are presented. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (b) Representative FACS plots and summarised data of % CD19<sup>+</sup>7AAD<sup>+</sup>B cells at day-3 and day-7 of B cell expansion are shown. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (c) Human CD19<sup>+</sup> B cell expansion could be maintained in culture for at least 14 days when CD154<sup>+</sup> CHO cells were replaced at day-3 and day-7 within the ex vivo culture system. Data from experiments performed with cells from different healthy donors (n = 3) are presented. (d) Summarised data of % CD19<sup>+</sup>IL-10<sup>+</sup> B cells at day-14 of B cell expansion are shown, when CD154<sup>+</sup> CHO cells were replaced either at day-3 only, or at day-3 and day-7 within the ex vivo culture system. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (e) Summarised data of IL-10 production by human CD19<sup>+</sup> B cells at day-14 of B cell expansion are shown, when CD154<sup>+</sup> CHO cells were replaced either at day-3 only, or at day-3 and day-7 within the ex vivo culture system. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (f) Representative FACS plots and summarised data of % CD19<sup>+7</sup>AAD<sup>+</sup>B cells at dav-14 of B cell expansion are shown, when CD154<sup>+</sup> CHO cells were replaced either at day-3 only, or at day-3 and day-7 within the *ex vivo* culture system. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. Error bars in each panel represent Mean +/- SD (a, b, d, e, f). One-way ANOVA with Tukey's multiple comparisons test (a) and 2-tailed paired t-test (b, c, d, e, f) were used. Source data are provided as a Source Data file.



(a) Gating strategy & representative FACS plots of live CD4<sup>+</sup>CFSE<sup>+</sup> T cells when autologous CD4<sup>+</sup>CFSE<sup>+</sup> T cells were cultured with anti-CD3/CD28 beads for 5 days +/- non-expanded CD19<sup>+</sup> B cells or expBreg at 1:1 ratio of B:T cells. Live/dead exclusion was performed using 7-AAD. (b) Representative histograms of live CD4<sup>+</sup>VPD<sup>+</sup>T cells and summarised data of suppressive potency of expBreg when expanded at a 1:1 CD154<sup>+</sup> CHO cell-CD19<sup>+</sup> B cell ratio with different cytokine combinations, are presented. Autologous CD4<sup>+</sup>VPD<sup>+</sup> T cells were cultured with anti-CD3/CD28 beads for 5 days +/- expBreg. % Inhibition of CD4<sup>+</sup> T cell proliferation is an expression of Division Index of live CD4<sup>+</sup>CFSE<sup>+</sup>T cells at day 5 relative to that of the Stimulated CD4<sup>+</sup>T cell control. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (c) Summarised data of % inhibition of CD4+ T cell proliferation are presented, when autologous CD4<sup>+</sup>VPD<sup>+</sup> T cells were cultured with anti-CD3/CD28 beads for 5 days +/-nonexpanded CD19<sup>+</sup> B cells (nCD19<sup>+</sup> B cells), expBreg or CD19<sup>+</sup> B cells that had been stimulated with platebound agonistic CD40 mAb (at [1ug/ml], [5ug/ml] or [10ug/ml] concentrations) for 7 days. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (d) Summarised data of % inhibition of CD4<sup>+</sup> T cell proliferation by expBreg when in the presence of blocking CD154 mAb, blocking CD40 mAb, or both. Blocking mAbs were added at day-0 of the 5-day suppression assay. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (e) Summarised data of % Inhibition of CD4<sup>+</sup> T cell proliferation by expBreg and expression of CD154 on CD4<sup>+</sup> T cells when measured daily in the 5-day suppression assay. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (f) Summarised data of % inhibition of CD4<sup>+</sup> T cell proliferation by expBreg when in the presence of TGF<sup>β</sup> inhibitor SB 431542 [µM/L] or DMSO. SB 431542 or DMSO were added at day-0 of the 5day suppression assay. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (g) Summarised data of IL-35 production by nCD19<sup>+</sup> B cells or expBreg. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. Error bars in each panel represent Mean +/- SD (b, c, e, f, g). One-way ANOVA with Tukey's multiple comparisons test (b, c, d) and 2-tailed paired t-test (f, g) were used. Source data are provided as a Source Data file.



CD19-APC Cy7

(a) Representative FACS plots of live CD19<sup>+</sup> B cells and summarised data demonstrating expression of cell surface markers by expBreg. Gating is on FMO controls. % Expression is compared between expBreg and autologous non-expanded CD19<sup>+</sup> B cells (nCD19<sup>+</sup> B cells). Data from experiments performed with cells from different healthy donors (n = 5) are presented. Each dot is an individual response. (b) Representative FACS plots of live CD19<sup>+</sup> B cells demonstrating expression of cell surface markers by non-expanded CD19<sup>+</sup> B cells (nCD19<sup>+</sup> B cells). Gating is on FMO controls. Data from experiments performed with cells from different healthy donors (n = 5) are presented. Each dot is an individual response. (b) Representative FACS (nCD19<sup>+</sup> B cells). Gating is on FMO controls. Data from experiments performed with cells from different healthy donors (n = 5) are presented. Each dot is an individual response. Error bars in (a) represent Mean +/- SD. 2-tailed paired t-tests (a) were used. Source data are provided as a Source Data file.

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TIM-1+ expBreg TIM-1expBreg





(a) Representative histograms and summarised data of cell surface markers expressed by IL-10<sup>+</sup> and IL-10<sup>-</sup> CD19<sup>+</sup> expBreg are presented. Gating is on FMO controls. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (b) FACS-sorting strategy to isolate expBreg based on the expression of TIM-1 (c) Representative FACS plots and summarised data of IL-10 expression by FACS-sorted TIM-1<sup>+</sup> and TIM-1<sup>-</sup> expBreg. Gating is on fluorescence minus one controls (FMO). (d) FACS-sorting strategy to isolate expBreg based on the expression of CD25. Error bars in (a, c) represent Mean +/- SD. 2-tailed paired t-tests (a, c) were used. Source data are provided as a Source Data file.



(a) FACS-sorting strategy to isolate expBreg based on the expression of CD154 or TIM-1. Gating is on fluorescence minus one controls (FMO). (b) Summarised data of % inhibition of CD4<sup>+</sup> T cell proliferation by FACS-sorted CD73<sup>-</sup>CD25<sup>+</sup>CD71<sup>+</sup> and CD73<sup>-</sup>CD25<sup>-</sup>CD71<sup>+</sup> expBreg are presented. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (c) Summarised data of % inhibition of CD4<sup>+</sup> T cell proliferation by FACS-sorted CD73<sup>-</sup>CD25<sup>+</sup>CD71<sup>+</sup> and CD73<sup>-</sup> CD25<sup>-</sup>CD71<sup>+</sup> expBreg which had been further FACS-sorted based on TIM-1 and CD154 expression, are presented. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (d) Summarised data of % CD4<sup>+</sup>TNFa<sup>+</sup> and CD4<sup>+</sup>IFNy<sup>+</sup> T cells at day-3 of suppression assay when autologous CD4<sup>+</sup> T cells were cultured with anti-CD3/CD28 beads +/- expBreg when in the presence of blocking CD80 mAb, blocking CD86 mAb, or both. Blocking mAbs was added at day-0 of suppression assay. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (e) Summarised data of % inhibition of CD4<sup>+</sup> T cell proliferation by expBreg when in the presence of blocking CD80 mAb, blocking CD86 mAb, or both. Blocking mAbs were added at day-0 of suppression assay. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (f) Summarised data of % inhibition of CD4<sup>+</sup> T cell proliferation by expBreg when in the presence of blocking PD-1 mAb. Blocking mAb was added at day-0 of suppression assay. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (g) Representative FACS plots of live CD19<sup>+</sup> expBreg and summarised data are presented which demonstrate protein expression of CD73, CD25 and CD71 after electroporation with Cas9 alone +/- TIM-1 or CD154 multiguide RNAs to generate TIM-1 or CD154 knockouts (KO). (h) Summarised data of IL-10 production by Cas9-expBreg, TIM-1 KO or CD154 KO expBreg are presented. Data from experiments performed with cells from different healthy donor (n = 4) are presented. Each dot is an individual response. (i) Summarised data of mean fluorescence intensity (MFI) of cell surface markers expressed by expBreg which had been pre-incubated with IgG isotype control (Isotype-expBreg), anti-CD154 mAb (CD154-blocked-expBreg) or anti-TIM-1 mAb (TIM-1-blocked-expBreg) for the last 48 hours of expansion, are presented. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. Error bars in (b, c, d, e, f, g, h, i) represent Mean +/- SD. Oneway ANOVA with Tukey's multiple comparisons test (d, e, g, h, i) and 2-tailed paired t-test (b, c, f) were used. Source data are provided as a Source Data file.



(a) Representative histograms of live CD19<sup>+</sup> expBreg are shown to demonstrate extracellular surface expression of IL-6 receptor (IL-6R). Gating is on fluorescence minus one controls (FMO). (b) Representative histograms of % STAT3 phosphorylation (pSTAT3) are presented when expBreg or an immortalised PANC-1 cell line were incubated with media or IL-6 ([0.01µg/ml], [1ug/ml] or [10ug/ml]) for 10 minutes and stained for pSTAT3. The PANC-1 cell line served as a positive control for IL-6-induced STAT3 phosphorylation. (c) Representative histograms of live CD4<sup>+</sup>CFSE<sup>+</sup> T cells are presented when autologous CD4<sup>+</sup>CFSE<sup>+</sup> T cells and anti-CD3/CD28 beads were co-cultured with expBreg +/- IL-21. expBreg were either pre-incubated with IL-21 [0.05µg/ml] for the last 48hrs of the 7-day expansion co-culture (IL-21-stimulated expBreg) prior to addition to the suppression assay at day-0, or IL-21 [0.05µg/ml] was directly added at day-0 of the suppression assay +/expBreg (expBreg + hIL-21). (d) Summarised data of % inhibition of CD4<sup>+</sup> T cell proliferation are presented when autologous CD4<sup>+</sup>VPD<sup>+</sup> T cells and anti-CD3/CD28 beads were co-cultured alone or with expBreg. expBreg had been pre-incubated with IL-6 ([0.01µg/ml], [1ug/ml] or [10ug/ml]) for the last 48hrs of the 7-day expansion co-culture (IL-6-stimulated expBreg). Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (e) Summarised data are presented which demonstrate % expression of CD40, CD154 and TIM-1 by expBreg and IL-21-stimulated expBreg. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (f) Summarised data of IL-10 production by expBreg and IL-21-stimulated expBreg. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (g) Summarised data of expansion factor of expBreg +/- IL-21 for the last 48hrs of the 7day expansion co-culture. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (h) Summarised data of number of CD19<sup>+</sup> B cells at day-5 of suppression assay when expBreg, IL-21-stimulated expBreg or expBreg + IL-21 had been added at day-0 of suppression assay. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Error bars in (d, e, f, g, h) represent Mean +/- SD. One-way ANOVA with Tukey's multiple comparisons test (d, h) and 2-tailed paired t-test (e, f, g) were used. Source data are provided as a Source Data file.



+ IL-2 [100ug/ml] + IL-2 [1000ug/ml]

(a) Representative histograms of live CD4<sup>+</sup>CFSE<sup>+</sup> T cells are presented when autologous CD4<sup>+</sup>CFSE<sup>+</sup> T cells and anti-CD3/CD28 beads were co-cultured with expBreg +/- STAT3 inhibitor +/- IL-21 [0.05µg/ml]. expBreg were incubated for 2 hours with the STAT3 inhibitor WP 1066 [10µM] +/- hIL-21 [0.05µg/ml], or DMSO +/hIL-21 [0.05µg/ml], washed and suppressive function analysed. (b) Summarised data are presented which demonstrate expression of CD40 and CD154 in the presence of STAT3 inhibition or DMSO. Data from experiments performed with cells from different healthy donors (n = 5) are presented. Each dot is an individual response. (c) Summarised data of IL-10 production by expBreg are presented when in the presence of STAT3 inhibition or DMSO. Data from experiments performed with cells from different healthy donors (n = 4) are presented. Each dot is an individual response. (d) Representative histograms of CD19<sup>+</sup>B cells and summarised data of % STAT5 phosphorylation (pSTAT5) are presented when Isotype-expBreg, TIM-blocked-expBreg, Cas9-expBreg or TIM-1 KO expBreg were incubated with media, IL-2 [10<sup>4</sup> U/ml] or IL-4 [1µg/ml] for 10 minutes and stained for pSTAT5. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. (e) Summarised data of % inhibition of CD4<sup>+</sup> T cell proliferation are presented when autologous CD4<sup>+</sup>CFSE<sup>+</sup> T cells and anti-CD3/CD28 beads were co-cultured with expBreg +/- IL-2. IL-2 was added at day-0 of the suppression assay, at different concentrations ([10ug/ml], [100ug/ml], 1000ug/ml]). Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. Error bars in (b, c, d, e) represent Mean +/- SD. One-way ANOVA with Tukey's multiple comparisons test (d, e) and 2-tailed paired t-test (b, c) were used. Source data are provided as a Source Data file.



(a) Humanised mouse model of human skin transplantation experimental set-up. (b) Gating strategy and representative FACS plots of live huCD45<sup>+</sup> cells in human skin allograft are presented. Gating is on fluorescence minus one controls (FMO). (c) Summarised data of the number of huCD45<sup>+</sup>CD8<sup>+</sup> T cells in spleen of mice receiving PBMC +/- nCD19<sup>+</sup> B cells or expBreg are presented. n=6 per group per experiment, one of three independent experiments is shown. Each dot is an individual mouse response. Each experiment used a different HLA-mismatched human donor pair. (d) Representative FACS plots of live huCD45<sup>+</sup>CD20<sup>+</sup>TIM-1<sup>+</sup> B cells in human skin allograft are presented. Gating is on FMO controls. (e) Summarised data of serum levels of human IgM and IgG in peripheral blood of mice receiving PBMC +/- nCD19<sup>+</sup> B cells or expBreg are presented. n=6 per group per experiment, one of three independent experiments is shown. Each dot is an individual mouse response. Each experiment used a different HLA-mismatched human donor pair. (f) Representative FACS plots and summarised data of the number of live huCD45<sup>+</sup>CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>lo</sup> putative Treg in human skin allograft of mice receiving PBMC +/- nCD19<sup>+</sup> B cells or expBreg are presented. n=6 per group per experiment, one of three independent experiments is shown. Each dot is an individual mouse response. Each experiment used a different HLA-mismatched human donor pair. (g) Representative FACS plots and summarised data of live CD4<sup>+</sup> T cells demonstrating % of putative Treg when expBreg and anti CD3/CD28 beads are co-cultured with CD4+ T cells or CD4<sup>+</sup>CD25<sup>-</sup> T cells *in vitro* for 5 days, when compared to controls. Data from experiments performed with cells from different healthy donors (n = 3) are presented. Each dot is an individual response. Error bars in (c, e, f) represent Mean +/- SEM. Error bars in (g) represent Mean +/- SD. One-way ANOVA with Tukey's multiple comparisons test (c, e, f, g) were used. Source data are provided as a Source Data file.



(a) Representative FACS plots of live CD19<sup>+</sup>CD73<sup>-</sup> B cells to demonstrate gating strategy for CD19<sup>+</sup>CD73<sup>-</sup> CD25<sup>+</sup>CD71<sup>+</sup> B Cells from human peripheral blood. Gating is on fluorescence minus one controls (FMO). (b) Representative FACS plots and summarised data of % IL-10 expression by CD24<sup>hi</sup>CD38<sup>hi</sup>, CD24<sup>hi</sup>CD27<sup>+</sup> and CD73<sup>-</sup>CD25<sup>+</sup>CD71<sup>+</sup> Breg subsets from human peripheral blood. Gating is on FMO controls. Data from experiments performed with cells from different healthy donors (n = 5) are presented. Each dot is an individual response. (c) Representative FACS plots of % TIM-1<sup>+</sup> B cells in whole CD19<sup>+</sup> B cells, CD73<sup>-</sup>CD25<sup>+</sup>CD71<sup>+</sup>, CD24<sup>hi</sup>CD38<sup>hi</sup> and CD24<sup>hi</sup>CD27<sup>+</sup> B cell subsets in peripheral blood of healthy human donors. Gating is on FMO controls. (d) Representative FACS plots of % CD154<sup>+</sup> B cells in whole CD19<sup>+</sup> B cells, CD73<sup>-</sup> CD25<sup>+</sup>CD71<sup>+</sup>, CD24<sup>hi</sup>CD38<sup>hi</sup> and CD24<sup>hi</sup>CD27<sup>+</sup> B cell subsets in peripheral blood of healthy human donors. Gating is on FMO controls. (e) Representative FACS plots of FACS-sorted CD73<sup>-</sup>CD25<sup>-</sup>CD71<sup>-</sup>, CD73<sup>-</sup> CD25<sup>+</sup>CD71<sup>+</sup> enriched, TIM-1<sup>+</sup> and TIM-1<sup>-</sup> B cells from human peripheral blood. (f) Summarised data of % TIM-1<sup>+</sup> and CD154<sup>+</sup> B cells within CD19<sup>+</sup> B cells in peripheral blood of healthy human donors (n=10) and agematched patients with SCC (n=8). Each dot is an individual response. (g) Summarised data of % IL-10 expression within CD73<sup>-</sup>CD25<sup>+</sup>CD71<sup>+</sup> B cells in peripheral blood of healthy human donors (n=10) and agematched patients with SCC (n=8). Each dot is an individual response. Error bars in (b, f, g) represent Median + interquartile range. One way ANOVA with Dunn's multiple comparisons test (b) and 2-tailed Mann Whitney test (f, g) were used. Source data are provided as a Source Data file.



(a) Hematoxylin and eosin (H&E) stained histological sections demonstrating invasive squamous cell carcinomas in an additional four patients (hashed line delineates tumour boundary; higher magnification image demonstrating representative histological features in top or bottom right-hand corners). (b) Double immunohistochemistry staining for pSTAT3 (red) and CD20 (brown) demonstrates the peritumoral accumulation of CD20<sup>+</sup> B cells (arrows), predominantly arranged as clusters, both within the tumours themselves and in proximity to their invasive margins (hashed line). (c) Higher magnification images of the annotated B cell clusters shown in (b) (hashed boxes), demonstrating double positive cells (arrows). (d) On the adjacent section, a single cell double stained for PAX5 (a master regulator of B cell development; red) and TIM 1 (brown) is identified in each cluster (arrows; higher magnification images of these cells in top right-hand corners). Each case in (a, b, c, d) represents 1 patient (n=4 patients).

## Supplementary Table 1

Antibody (Human)	Fluorochrome	Concentration	Supplier	Catalogue Number
CD19	APC-Cy7	1:100	BD Pharmingen	557791
CD25	PE-Cy7	1:100	BD Pharmingen	557741
CD24	PE	1:100	BD Pharmingen	555428
CD38	FITC	1:100	BD Pharmingen	555459
CD127	PE	1:100	BD Pharmingen	557938
IgD	PE-Cy7	1:100	BD Pharmingen	561314
pSTAT3	Alexa Fluor 647	1:6	BD Pharmingen	557815
pSTAT5	Alexa Fluor 488	1:6	BD Pharmingen	562077
CD20	PE-Cy7	1:100	eBioscience	25-0209-42
CD4	PE-Texas Red	1:100	eBioscience	61-0049-42
CD8	FITC	1:100	eBioscience	11-0088-42
γCR	PE	1:100	eBioscience	12-1329-41
<b>CD27</b>	eFluor 450	1:100	eBioscience	48-0279-42
CD138	eFluor 450	1:100	eBioscience	17-1389-41
CD1d	APC	1:100	eBioscience	17-0016-42
CD5	FITC	1:100	eBioscience	11-0058-42
CD21	eFluor 450	1:100	eBioscience	9048-0219-025
CD71	APC	1:100	eBioscience	17-0719-41
<b>CD73</b>	eFluor 450	1:100	eBioscience	48-0739-41
LAP	PE	1:100	eBioscience	12-9829-41
IL-10	eFluor 660	1:4	eBioscience	50-7108-41
TNFa	FITC	1:4	eBioscience	11-7349-41
ΙΓΝγ	PE	1:4	eBioscience	12-7319-41
CD154	PE	1:100	eBioscience	12-1548-42
IgM	PE	1:100	eBioscience	12-9998-42
CD40	APC	1:100	eBioscience	17-0409-41
CD80	FITC	1:100	eBioscience	11-0809-42
CD86	PE	1:100	eBioscience	12-0869-42
FAS-L	PE	1:100	eBioscience	12-9919-41
PD-L1	PE-Cy7	1:100	eBioscience	25-5983-41
PD-L2	APC	1:100	eBioscience	17-5888-41
TIM-1	PE	1:100	Biolegend	353903
IL-10R	APC	1:100	Biolegend	308811
CD122	APC	1:100	Biolegend	339007
IL-6Ra	PE	1:100	Biolegend	352803
CD154	FITC	1:100	Biolegend	310804
CD10	PE-Texas Red	1:100	Beckman Coulter	41116015
CD45	APC	1:100	Invitrogen	MHCD45054
CD154	APC	1:100	R&D Systems	FAB617A
CD25	PE	1:100	R&D Systems	FAB1020P-025
CD154	Unconjugated	[10µg/ml]	K&D Systems	MAB61/
CD40	Unconjugated	[10µg/ml]	R&D Systems	MAB6322
IL-10	Unconjugated	[10µg/ml]	R&D Systems	MAB217
IL-10Rα	Unconjugated	[10µg/ml]	K&D Systems	MAB2/4
CD122	Unconjugated	[10µg/ml]	R&D Systems	MAB224
CD25	Unconjugated	[10µg/ml]	R&D Systems	MAB223

Antibody (Human)	Fluorochrome	Concentration	Supplier	Catalogue Number
CD80	Unconjugated	[10µg/ml]	R&D Systems	MAB140
CD86	Unconjugated	[10µg/ml]	R&D Systems	MAB141
FASL	Unconjugated	[10µg/ml]	R&D Systems	MAB126
PD-1	Unconjugated	[10µg/ml]	R&D Systems	AF1086
IL-6	Unconjugated	[10µg/ml]	R&D Systems	MAB2061
IL-6R	Unconjugated	[10µg/ml]	R&D Systems	MAB227R
IgG <sub>1</sub> <sub>k</sub>	Unconjugated	[10µg/ml]	R&D Systems	MAB002
IgG <sub>2ak</sub>	Unconjugated	[10µg/ml]	R&D Systems	MAB004
CD25	Unconjugated	[10µg/ml]	R&D Systems	MAB1020
TIM-1	Unconjugated	[10µg/ml]	Biolegend	353902
CD20	Unconjugated	1:100	Leica Biosystems	PA0200
pSTAT3	Unconjugated	1:100	Abcam	ab76315
TIM-1	Unconjugated	1:100	Abcam	ab47635
PAX5	Unconjugated	1:500	Abcam	ab109443