

OMTN, Volume 28

Supplemental information

SREBP1 regulates *Lgals3*

activation in response to cholesterol loading

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Supplemental Figures

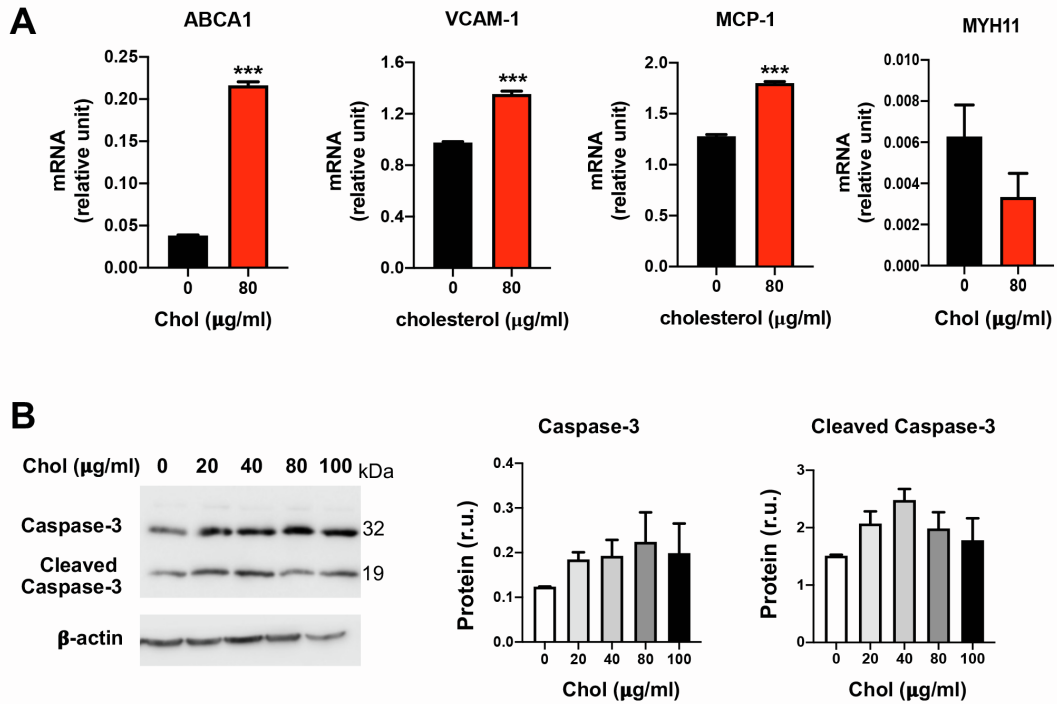


Figure S1. Cholesterol loading stimulates inflammatory marker gene expression without causing apoptosis

MOVAS cell culture, cholesterol feeding, and data quantification were performed as described for Figure 1. qRT-PCR data are presented as mean ± SD, n = 3 replicates. Western blot data are presented as mean ± SEM, n = 3 independent repeat experiments. Statistics: ANOVA followed by Tukey test; **P<0.01, ***P<0.001, compared to the basal condition (the 1st bar in each plot); non-significant comparisons are not labeled.

- A. Upregulation of inflammatory marker mRNAs (qRT-PCR) due to cholesterol feeding (80 μg/ml).
- B. Full length and cleaved caspase-3 protein (Western blot).

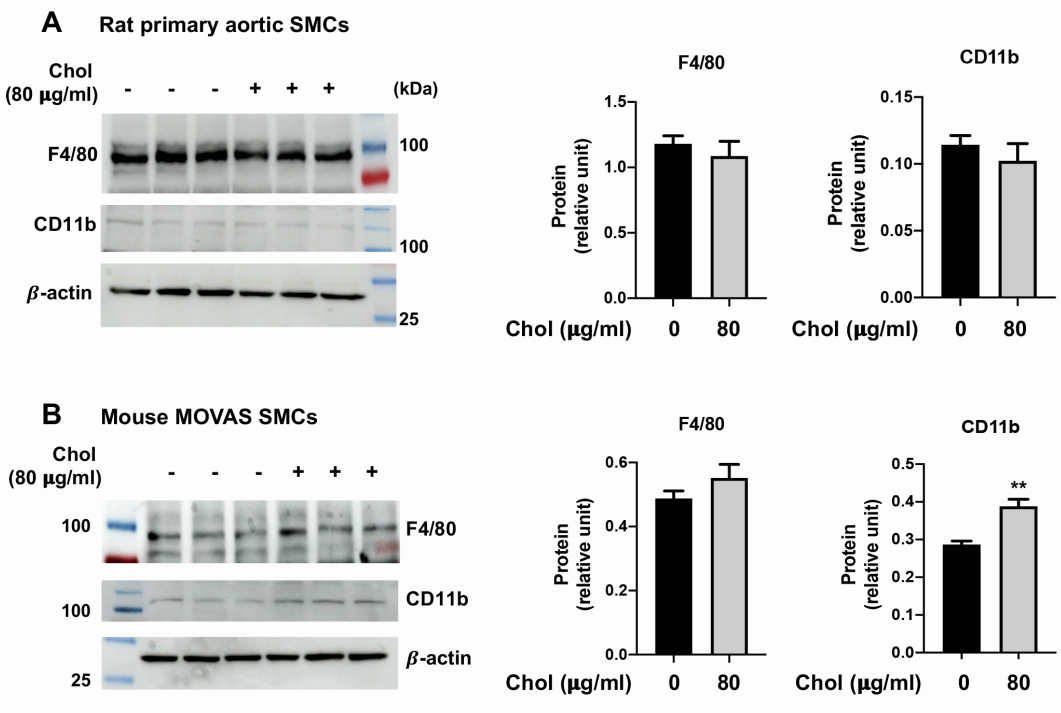


Figure S2. Effect of cholesterol loading on macrophage markers

Rat primary SMCs (A) and the MOVAS mouse SMC line (B) were used. Cell culture, cholesterol loading, and data quantification were performed as described for Figure 1. Data quantification for Western blots: Mean ± SD, n = 3 replicates. Statistics: Student's t-test. **P<0.01.

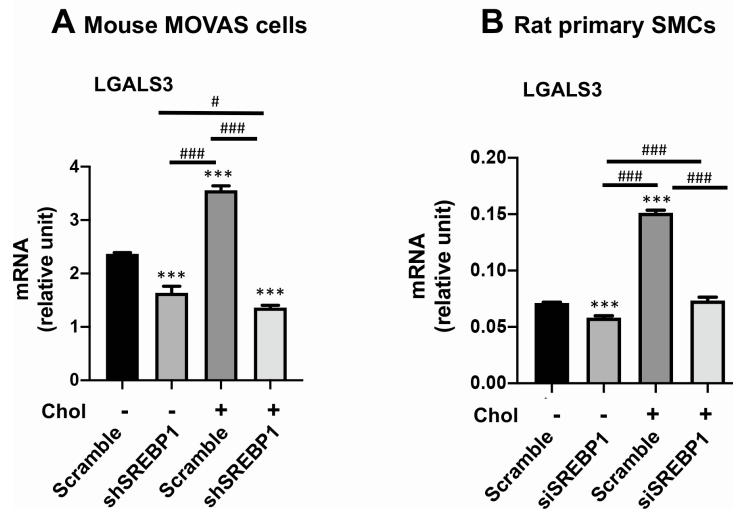


Figure S3. Effect of SREBP1 silencing on LGALS3 mRNA levels.

Both the MOVAS mouse SMC line (A) and rat primary SMCs (B) were used. Cell culture, cholesterol loading, and data quantification were performed as described for Figure 1.

Data quantification: Mean \pm SD, n = 3 replicates. Statistics: one-way ANOVA followed by Tukey test; ###P<0.001 (between paired bars); ***P<0.001, compared to the basal condition (the 1st bar in each plot); non-significant comparisons are not labeled.

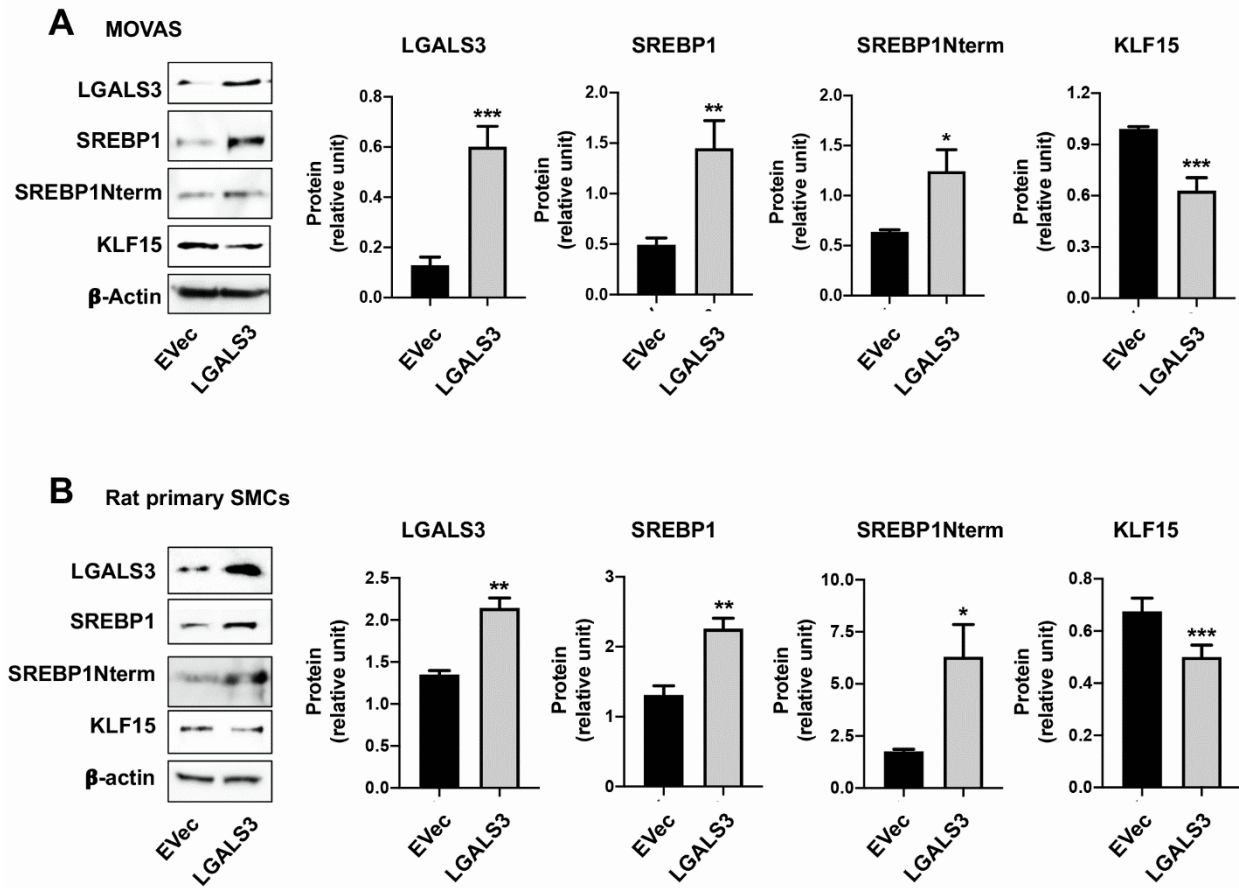
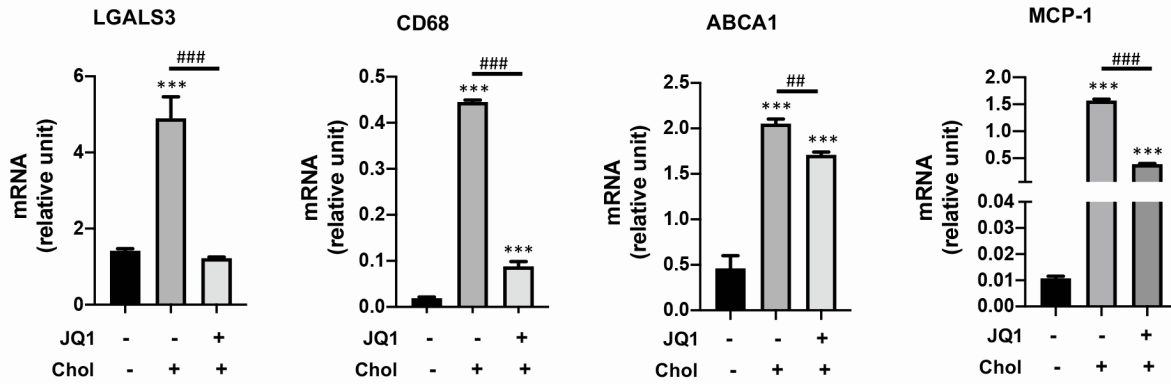


Figure S4. LGALS3 gain-of-function upregulates the full length and N-terminal half molecules of the SREBP1 protein

MOVAS cells (A) or rat primary aortic SMCs (B) were used. Experiments were performed as in Figure 5B. Data are presented as mean \pm SEM, $n = 3$ independent repeat experiments. Statistics: ANOVA followed by Tukey test; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, compared to EVec (empty vector control).

A Mouse MOVAS cells



B Rat primary SMCs

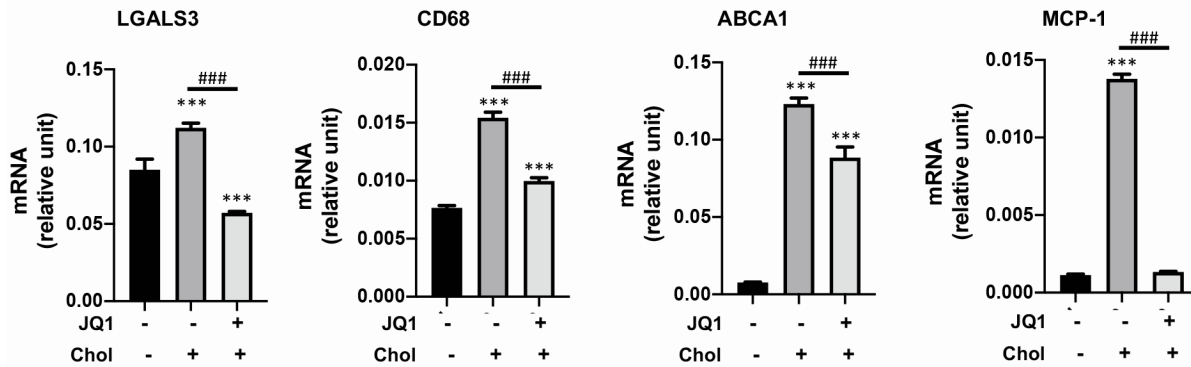


Figure S5. Inhibitory effect of the BETs inhibitor JQ1 on LGALS3 mRNA levels

Both the MOVAS mouse SMC line (A) and rat primary SMCs (B) were used. Cell culture, cholesterol loading, and data quantification were performed as described in detail for Figure 1. Data quantification: Mean \pm SD, n = 3 replicates. Statistics: one-way ANOVA followed by Tukey test; ##P<0.01, ###P<0.001 (between paired bars); ***P<0.001, compared to the basal condition (the 1st bar in each plot); non-significant comparisons are not labeled.

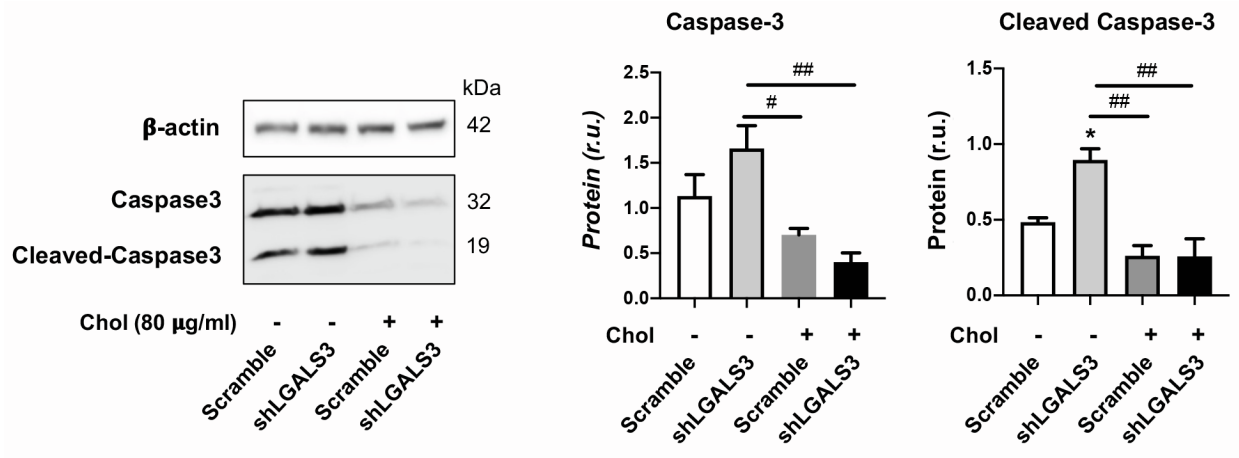


Figure S6. Effect of LGALS3 loss-of-function on cleaved caspase-3

Experiments were performed as in Figure 5A. Data are presented as mean \pm SEM, n = 3 independent repeat experiments. Statistics: ANOVA followed by Tukey test; #P<0.05, ##P<0.01 (between paired bars); *P<0.05, compared to the basal condition (the 1st bar in the plot); non-significant comparisons are not labeled; r.u., relative unit.

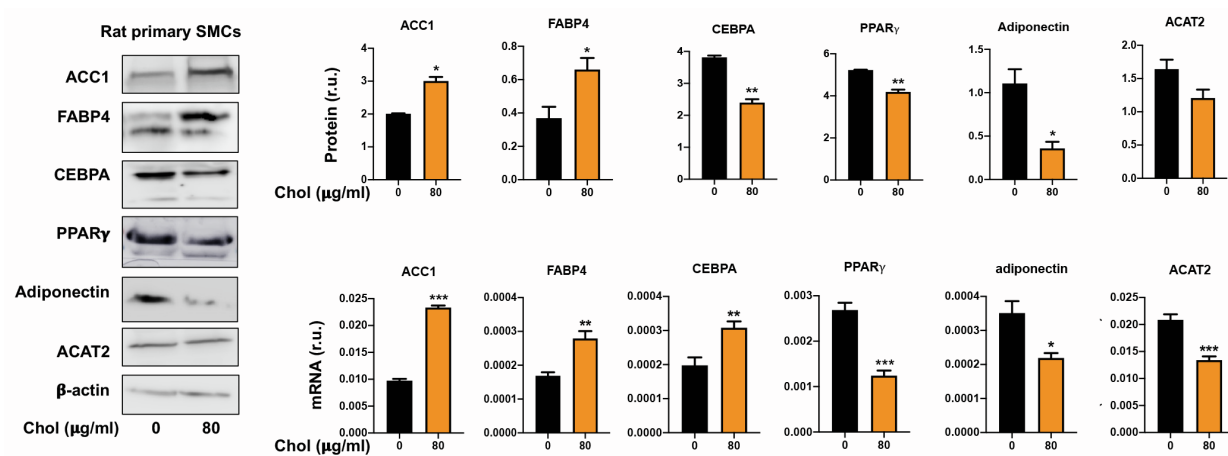


Figure S7. Chol loading alters the levels of proteins involved in lipid storage

Experiments were performed as in Figure 7D but in rat primary SMCs instead. Western blot data are presented as mean \pm SEM, n = 3 independent repeat experiments. qRT-PCR data are presented as mean \pm SD, n = 3 replicates. Statistics: ANOVA followed by Tukey test; *P<0.05, **P<0.01, ***P<0.001, compared to the control without cholesterol loading.

Supplemental Tables

Table S1. Kits and reagents

Product	Manufacturer	Catalog Number
JetPRIME Transfection reagent	Polyplus transfection	114-07
Lipofectamine RNAiMAX Transfection Reagent	Invitrogen	3778150
PageRuler™ PLUS Prestained Protein Ladder	Thermo Scientific	26620
Clarity Western ECL Substrate	Bio-Rad	170-5060
QIAGEN Plasmid MIDIprep Kit	QIAGEN	12143
PureLink™ Quick Plasmid Miniprep Kit	Invitrogen	K210011
PureLink™ Quick Gel Extraction and PCR Purification Combo Kit	Invitrogen	K220001
NE-PER™ Nuclear and Cytoplasmic Extraction Reagents	Invitrogen	78835
LightShift™ Chemiluminescent EMSA Kit	Invitrogen	20148
Pierce™ Protein A/G Magnetic Agarose Beads	Invitrogen	78610
UltraPure™ Salmon Sperm DNA Solution	Invitrogen	15632011
Mouse IgG	Invitrogen	10400C
PerfeCTa SYBR® Green SuperMix	Quantabio	95054
V5-TAG MAGNETIC BEADS	Fisher Scientific	NC0777490
Pierce™ Anti-DYKDDDDK Magnetic Agarose	Thermofisher	A36797

Table S2. shRNAs and siRNAs for gene silencing

Name	Sequence (5'-3')
Rat <i>Srebf1</i> siRNA (ID: s134945)	Sense: CCUGCGAAGUGCUCACAAATT
	Antisense: UUUGUGAGCACUUCGCAGGGT
Rat <i>Lgals3</i> siRNA (ID: s136243)	Sense: GCAAACCAUUCAAAAUACATT
	Antisense: UGUUUUUUGAAUGGUUUUGCCG
Human <i>Srebf1</i> siRNA (ID: 5140)	Sense: GGCAAAGCUGAAUAAAUCUTT Antisense: AGAUUUUAUUCAGCUUUGCCTC
Human <i>Lgals3</i> siRNA (ID: s8149)	Sense: CGGUGAAGCCCAAUGCAAATT Antisense: UUUGCAUUGGGCUUCACCGTG
Rat <i>Klf15</i> shRNA#1	CCGGCTACCCTGGAGGAGATTGAAGCTCGAGCTTCAATCTCCTC CAGGGTAGTTTTTG
	AATTCAAAACCTACCCTGGAGGAGATTGAAGCTCGAGCTTCAAT CTCCTCCAGGGTAG
Rat <i>Klf15</i> shRNA#2	CCGGCCAAACCTATTGGCTCAGGATCTCGAGATCCTGAGCCAAT AGGTTTGGTTTTTG
	AATTCAAAACCAAACCTATTGGCTCAGGATCTCGAGATCCTGA GCCAATAGGTTTTGG
Rat KLF15 shRNA#3	CCGGCCAGGGCAGCATCTTGGATTTCTCGAGAAATCCAAGATGC TGCCCTGGTTTTTG
	AATTCAAAACCAAGGGCAGCATCTTGGATTTCTCGAGAAATCCAA GATGCTGCCCTGG
Mouse <i>Srebf1</i> shRNA#1	CCGGGCGGCTGTTGTCTACCATAAGCTCGAGCTTATGGTAGACA ACAGCCGCTTTTTG
	AATTCAAAAGCGGCTGTTGTCTACCATAAGCTCGAGCTTATGGT AGACAACAGCCGC
Mouse <i>Srebf1</i> shRNA#2	CCGGGCTGCTATGAGGAGGGTATTCTCGAGAATACCCTCCTCA TAGCAGGCTTTTTG
	AATTCAAAAGCCTGCTATGAGGAGGGTATTCTCGAGAATACCC TCCTCATAGCAGGC
Mouse <i>Srebf1</i> shRNA#3	CCGGCCTGCACTTCTTGACACGTTTCTCGAGAAACGTGTCAAGA AGTGCAGGTTTTTG
	AATTCAAAACCTGCACTTCTTGACACGTTTCTCGAGAAACGTGT CAAGAAGTGCAGG
Mouse <i>Srebf1</i> shRNA#4	CCGGGCTGAATAAATCTGCTGTCTTCTCGAGAAGACAGCAGATT TATTCAGCTTTTTG
	AATTCAAAAGCTGAATAAATCTGCTGTCTTCTCGAGAAGACAGC AGATTTATTCAGC
Mouse <i>Lgals3</i> shRNA#1	CCGGGGAGCTTATCCTGGCTCAACTCTCGAGAGTTGAGCCAGG ATAAGCTCCTTTTTG
	AATTCAAAAGGAGCTTATCCTGGCTCAACTCTCGAGAGTTGAG CCAGGATAAGCTCC
Mouse <i>Lgals3</i> shRNA#2	CCGGCCGCATGCTGATCACAATCATCTCGAGATGATTGTGATCA GCATGCGGTTTTTG

	AATTCAAAAACCGCATGCTGATCACAATCATCTCGAGATGATTGT GATCAGCATGCGG
Mouse <i>Lgals3</i> shRNA#3	CCGGGCAGTACAACCATCGGATGAACTCGAGTTCATCCGATGGT TGTACTGCTTTTTG
	AATTCAAAAAGCAGTACAACCATCGGATGAACTCGAGTTCATCC GATGGTTGTACGCC
Mouse <i>Klf15</i> shRNA#1	CCGGACCGAAATGCTCAGTGGGTTACTCGAGTAACCCACTGAG CATTTCGGTTTTTTG
	AATTCAAAAACCGAAATGCTCAGTGGGTTACTCGAGTAACCCA CTGAGCATTTCGGT
Mouse <i>Klf15</i> shRNA#2	CCGGGCGATCTCACTCGGGTGTGAACTCGAGTTCACACCCGAG TGAGATCGCTTTTTG
	AATTCAAAAAGCGATCTCACTCGGGTGTGAACTCGAGTTCACAC CCGAGTGAGATCGC
Mouse <i>Klf15</i> shRNA#3	CCGGCTACCCTGGAGGAGATTGAAGCTCGAGCTTCAATCTCCTC CAGGGTAGTTTTTG
	AATTCAAAAACTACCCTGGAGGAGATTGAAGCTCGAGCTTCAAT CTCCTCCAGGGTAG
Mouse <i>Klf15</i> shRNA#4	CCGGGCGATCTCACTCGGGTGTGAACTCGAGTTCACACCCGAG TGAGATCGCTTTTTG
	AATTCAAAAAGCGATCTCACTCGGGTGTGAACTCGAGTTCACAC CCGAGTGAGATCGC
Mouse <i>Klf15</i> shRNA#5	CCGGACCGAAATGCTCAGTGGGTTACTCGAGTAACCCACTGAG CATTTCGGTTTTTTG
	AATTCAAAAACCGAAATGCTCAGTGGGTTACTCGAGTAACCCA CTGAGCATTTCGGT
Mouse <i>Klf15</i> shRNA#6	CCGGCCCTCAAAGTTTGTGCGAATTCTCGAGAATTCGCACAAAC TTTGAGGGTTTTTG
	AATTCAAAAACCTCAAAGTTTGTGCGAATTCTCGAGAATTCGCA CAAACCTTGAGGG
Mouse <i>Brd4</i> shRNA#1	CCGGGCCATCTACACTACGAGAGTTCTCGAGAACTCTCGTAGTG TAGATGGCTTTTTG
	AATTCAAAAAGCCATCTACACTACGAGAGTTCTCGAGAACTCTCG TAGTGTAGATGGC
Mouse <i>Brd4</i> shRNA#2	CCGGGATGTGTTTGAAATGCGCTTTCTCGAGAAAGCGCATTTC AACACATCTTTTTG
	AATTCAAAAAGATGTGTTTGAAATGCGCTTTCTCGAGAAAGCGCA TTTCAAACACATC
Mouse <i>Brd4</i> shRNA#3	CCGGGCAGCAGCAGCAGCAGCAACCCTCGAGGGTTGCTGCTGC TGCTGCTGCTTTTTG
	AATTCAAAAAGCAGCAGCAGCAGCAGCAACCCTCGAGGGTTGCT GCTGCTGCTGCTGC
Mouse <i>Brd4</i> shRNA#4	CCGGCCCAGCTCCTCTGACAGTGAAGCTCGAGCTTCACTGTCA GAGGAGCTGGATTTTTG
	AATTCAAAAACCCAGCTCCTCTGACAGTGAAGCTCGAGCTTCAC TGTCAGAGGAGCTGGA
Mouse <i>Brd3</i> shRNA#1	CCGGAGCTGAACCTGCCTGATTATCCTCGAGGATAATCAGGCAG GTTGAGCTTTTTTG

	AATTCAAAAAGCTGAACCTGCCTGATTATCCTCGAGGATAATCA GGCAGGTTTCAGCT
Mouse <i>Brd3</i> shRNA#2	CCGGCCACAGATGATATAGTGCTAACTCGAGTTAGCACTATATC ATCTGTGGTTTTTG
	AATTCAAAAACACAGATGATATAGTGCTAACTCGAGTTAGCACT ATATCATCTGTGG
Mouse <i>Brd2</i> shRNA#1	CCGGCAGCCCAAGAAATCTAAGAACTCGAGTTTCTTAGATTTCT TGGGCTGTTTTTG
	AATTCAAAAACAGCCCAAGAAATCTAAGAACTCGAGTTTCTTAG ATTTCTTGGGCTG
Mouse <i>Brd2</i> shRNA#2	CCGGCCTCAGAATGTATGCAGGATTCTCGAGAATCCTGCATACA TTCTGAGGTTTTTG
	AATTCAAAAACCTCAGAATGTATGCAGGATTCTCGAGAATCCTGC ATACATTCTGAGG
Scrambled- sequence Control shRNA	CCGGTAGCGACTAAACACATCAATCCTCGAGGATTGATGTGTTT AGTCGCTATTTTTG
	AATTCAAAAATAGCGACTAAACACATCAATCCTCGAGGATTGATG TGTTTAGTCGCTA

Table S3. Primers for cloning to overexpress a gene

Name	Sequence (5'-3')	Application
Mouse <i>Srebf1</i> cDNA	Forward: GTGGAATTCTATGGACTACAAGGATGAC GATGAC	Lentiviral vector (inserted into pLenti-puro)
	Reverse: CTCTAGACTGCTGGAAGTGACGGTGGTTCCGCC	
Mouse <i>Klf15</i> cDNA	Forward: CCGCTAGCGATGGATTACAAGGATGACGACG	Lentiviral vector (inserted into pLenti-puro)
	Reverse: CGACCGGTAGGTTGATGGCGGCTACTGCGC	
Mouse <i>Lgals3</i> cDNA	Forward: GTGGATCCAATGGCAGACAGCTTTTCGCTT	Lentiviral vector (inserted into pLenti-puro)
	Reverse: CACTCGAGCGGATCATGGCGTGGTTAGCGC	
Rat <i>Lgals3</i> cDNA	Forward: GTGGATCCAATGGCAGACGGCTTCTCACTT	Lentiviral vector (inserted into pLenti-puro)
	Reverse: CGAATTCCAGATCATGGCGTGGGAAGCGCT	

Table S4. Primers for qRT-PCR

Mouse GAPDH	Forward: AAGGTCGGTGTGAACGGATTT
	Reverse: CTTTGTCAACAAGAGAAGGCAGC
Mouse ACTA2	Forward: ACTCTGTCTGGATCGGTGGC
	Reverse: TTCGTTCGTATTCCCTGTTTGCT
Mouse VCAM1	Forward: TGGAAATGTGCCCGAAAC
	Reverse: GCCTGGCGGATGGTGTAC
Mouse ABCA1	Forward: CCGTCTTTCCAGGACAGTAT
	Reverse: CAGGGTGGCTCTTCTCATC
Mouse CD68	Forward: ACCGTGACCAGTCCCTCTT
	Reverse: AAGGCGATGAGCACCAGGGTGAGGA
Mouse LGALS3	Forward: CTGGAGGCTATCCTGCTGC
	Reverse: AACCAATCCTGTTTGCGTTGG
Mouse KLF15	Forward: CCGAAATGCTCAGTGGGTAC
	Reverse: GAGTCAGGGCTGGCACAAGA
Mouse SREBP1 C-terminal domain (primer 1)	Forward: CCAGGTGACCCGGCTATTC
	Reverse: CCAAGGGCATCTGAGAACTCC
Mouse SREBP1 N-terminal domain (primer 2)	Forward: CGGCACCCGCTGCTTTA
	Reverse: TGGCACTGGCTCCTCTTTGA

Table S5. Antibodies for Western blot (WB) or ChIP or IP

Antigen	Manufacturer	Catalog Number	Dilution Ratio	Application
BRD2	Proteintech	22236-1-AP	1:1000	WB, ChIP
Beta-actin	Proteintech	60008-1-Ig	1:10000	WB
FLAG tag	Sigma Aldich	F3165	1:1000	WB
Alpha-SMA	Proteintech	55135-1-AP	1:5000	WB
LGALS3	Cell Signaling Technology	87985	1:1000	WB
CD68	Santa Cruz	sc-17832	1:250	WB
KLF15	Santa Cruz	sc-271675	1:1000	WB, ChIP
SREBP1(2A4) (for full-length and N-term)	Santa Cruz	sc-13551	1:1000	WB, ChIP
MRTF-A/MKL1	Cell Signaling Technology	14760	1:1000	WB
Histone-3	Cell Signaling Technology	14269	1:1000	WB
GAPDH	Cell Signaling Technology	5174	1:1000	WB
Caspas3/P17/P19	Proteintech	19677-1-AP	1:1000	WB
ACC1	Proteintech	21923-1-AP	1:1000	WB
FABP4	abcam	ab23693	1:1000	WB
CEBPA	Proteintech	18311-1-AP	1:1000	WB
PPAR gamma	Proteintech	16643-1-AP	1:1000	WB
Adiponectin	Cell Signaling Technology	2789s	1:1000	WB
ACAT2	Proteintech	14755-1-AP	1:1000	WB

Table S6. Primers for ChIP-qPCR

ChIP: Anti-SREBP ^N term or anti-KLF15, S1 site in <i>Lgals3</i> promoter	Forward: TGGGTGCGGTCGGCTAGGGCCC
	Reverse: CCCCACGGACCACGGAGCTTC
ChIP: Anti-SREBP ^{N1} term or anti-KLF15, S2 site in <i>Lgals3</i> promoter	Forward: GTCCTCCCTTCCCTCACAC
	Reverse: CACCCAGACTCTCAGACTCACCC
ChIP: Anti-SREBP ^{N1} term or Anti-KLF15, S3 site in <i>Lgals3</i> promoter	Forward: CAGGGATCAAAGTTAGGCGTC
	Reverse: GTCGCTGTGCCCTTGCTTAC
ChIP: Anti-BRD2 or anti-H3K27ac, Primer #1 for <i>Srebf1</i> promoter	Forward: CTGCCACCCAAGTGCTGG
	Reverse: CTGAAGGGCCAGTGGGCTC
ChIP: Anti-BRD2 or anti-H3K27ac, Primer #2 for <i>Srebf1</i> promoter	Forward: GAACCTCTCCTCCCTCCC
	Reverse: CTGTAACAGAGGTCCTGAG
ChIP: Anti-BRD2 and anti-H3K27ac, Primer S1 for <i>Lgals3</i> promoter	Forward: TGGGTGCGGTCGGCTAGGGCCC
	Reverse: CCCCACGGACCACGGAGCTTC
ChIP: Anti-BRD2 and anti-H3K27ac, Primer S2 for <i>Lgals3</i> promoter	Forward: GTCCTCCCTTCCCTCACAC
	Reverse: CACCCAGACTCTCAGACTCACCC

Table S7. Oligo sequences for EMSA

Name	Sequence (5'-3')	Biotin label
S1 WT F (SREBP1)	CTGTGGGGAAGCTCCGTGGTCCGTGGGGCGCGGTCCAG CCAGGCGCCTGC	5' labeling
S1 WT R (SREBP1)	GCAGGCGCCTGGCTGGACCGCGCCCCACGGACCACGGA GCTTCCCCACAG	
S1-mu1 F (SREBP1)	CTGTGGGGAAGCTCCGTGGTCCGTGTGGTGCGGTCCAG CCAGGCGCCTGC	5' labeling
S1-mu1 R (SREBP1)	GCAGGCGCCTGGCTGGACCGCACCACACGGACCACGGA GCTTCCCCACAG	
S1-mu2 F (SREBP1)	CTGTGGGGAAGCTCCGTGGTCCCTTGTGGTGCGGTCCAGC CAGGCGCCTGC	5' labeling
S1-mu2 R (SREBP1)	GCAGGCGCCTGGCTGGACCGCACCACAAGGACCACGGA GCTTCCCCACAG	
S1-mu3 F (SREBP1)	CTGTGGGGAAGCTCCGTGGTCCCTTGTGTTGCGGTCCAGC CAGGCGCCTGC	5' labeling
S1-mu3 R (SREBP1)	GCAGGCGCCTGGCTGGACCGCAACACAAGGACCACGGA GCTTCCCCACAG	
S2 WT F (KLF15)	GTAAGCCCTAGGCATAGAGTGGGGTGCATAAGTGTTTG GTAGATATTAG	5' labeling
S2 WT R (KLF15)	CTAATATCTACCAAACACTTATGCACCCCACTCTATGCCTA GGGCTTTAC	
S3- F	CATCCCCCTGCCCGGGTGTGGGGAGAAGTCAGCAGAAT GGGGGGC	
S3- R	GCCCCCATTCTGCTGACTTCTCCCCACACCCGGGGCAG GGGAATG	
S2-mu1 F (KLF15)	GTAAGCCCTAGGCATAGAGTGTGTTGCATAAGTGTTTGG TAGATATTAG	5' labeling
S2-mu1 R (KLF15)	CTAATATCTACCAAACACTTATGCAACACACTCTATGCCTA GGGCTTTAC	
S2-mu2 F (KLF15)	GTAAGCCCTAGGCATAGATTGTGTTGCATAAGTGTTTGG TAGATATTAG	5' labeling
S2-mu2 R (KLF15)	CTAATATCTACCAAACACTTATGCAACACAATCTATGCCTA GGGCTTTAC	
S2-mu3 F (KLF15)	GTAAGCCCTAGGCATAGATTGTGTTTCATAAGTGTTTGG TAGATATTAG	5' labeling
S2-mu3 R (KLF15)	CTAATATCTACCAAACACTTATGAAACACAATCTATGCCTA GGGCTTTAC	

Note: The oligo sequences are within the respective S1 and S2 Lgals3 promoter regions.