

Supporting Information

FastCAT accelerates absolute quantification of proteins by using multiple short non-purified chimeric standards

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Chimeric protein (CP) standard

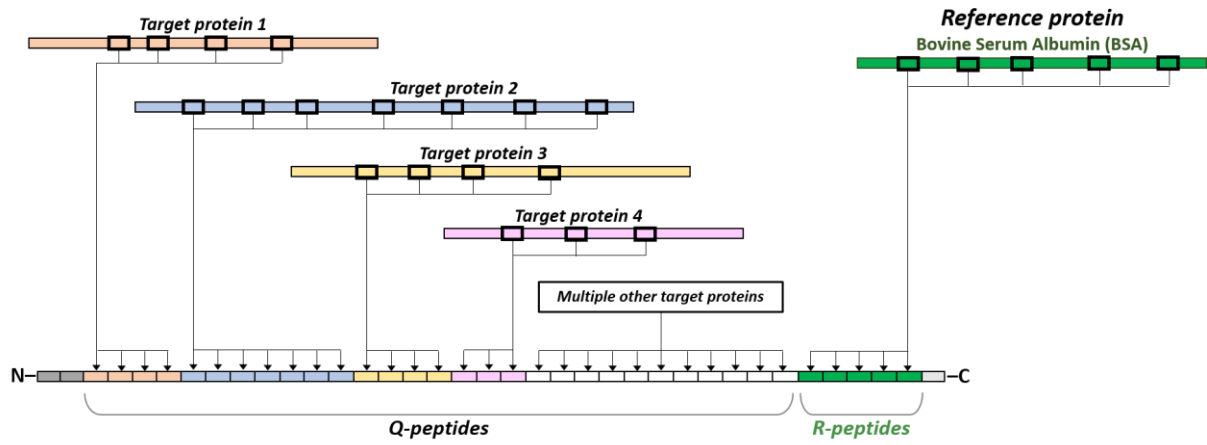


Figure S1. Scheme of chimeric protein (CP) standards used in this work. A typical CP comprises quantotypic Q-peptides selected from the sequences of target proteins and reference R-peptides selected from bovine serum albumin (BSA). They were concatenated *in-silico* into a chimeric sequence, whose N- and C-termini contain additional tag sequences (in gray) for R- and Q- peptides protection and (optionally) for the affinity purification.

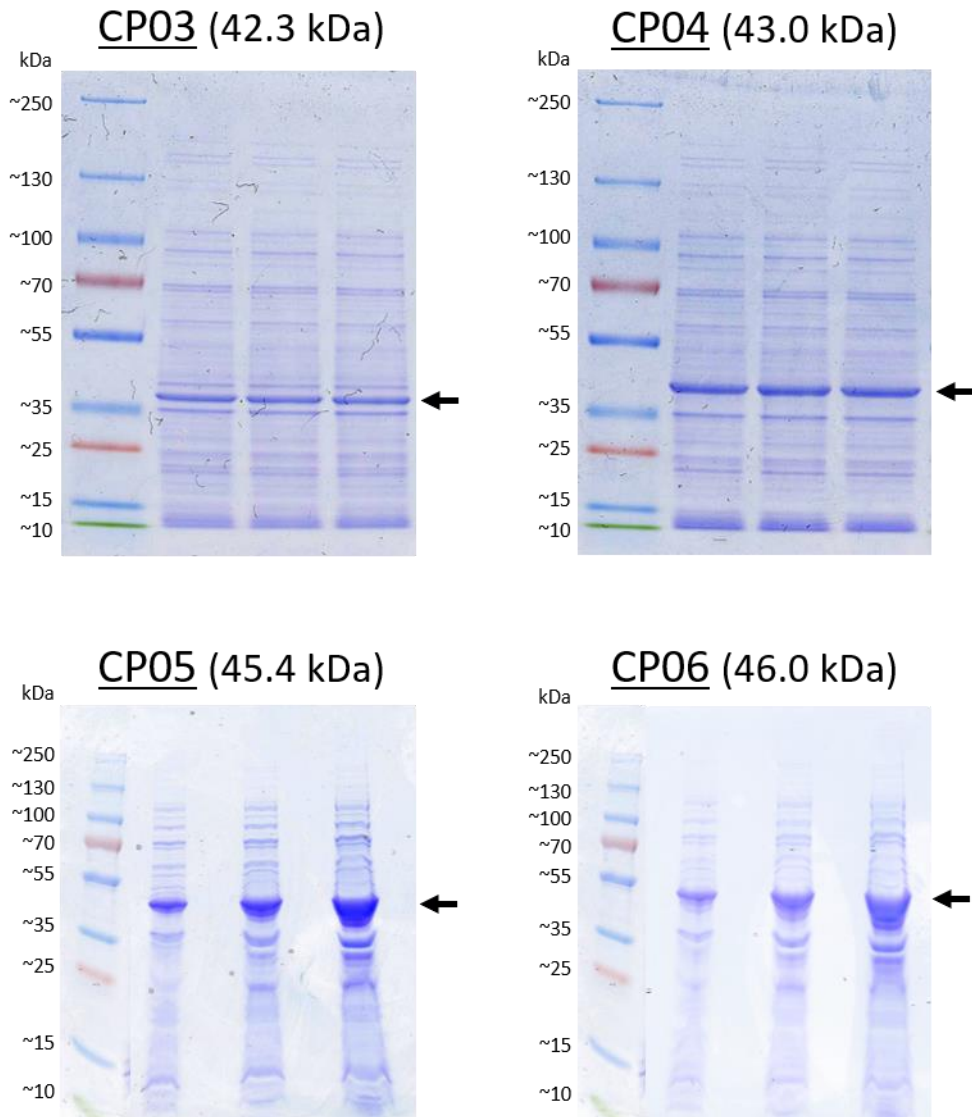


Figure S2. Images of Coomassie stained SDS-PAGE gels of four short (<50 kDa) CPs used in this study. Gel bands corresponding to the full-length CPs are designated with arrows.

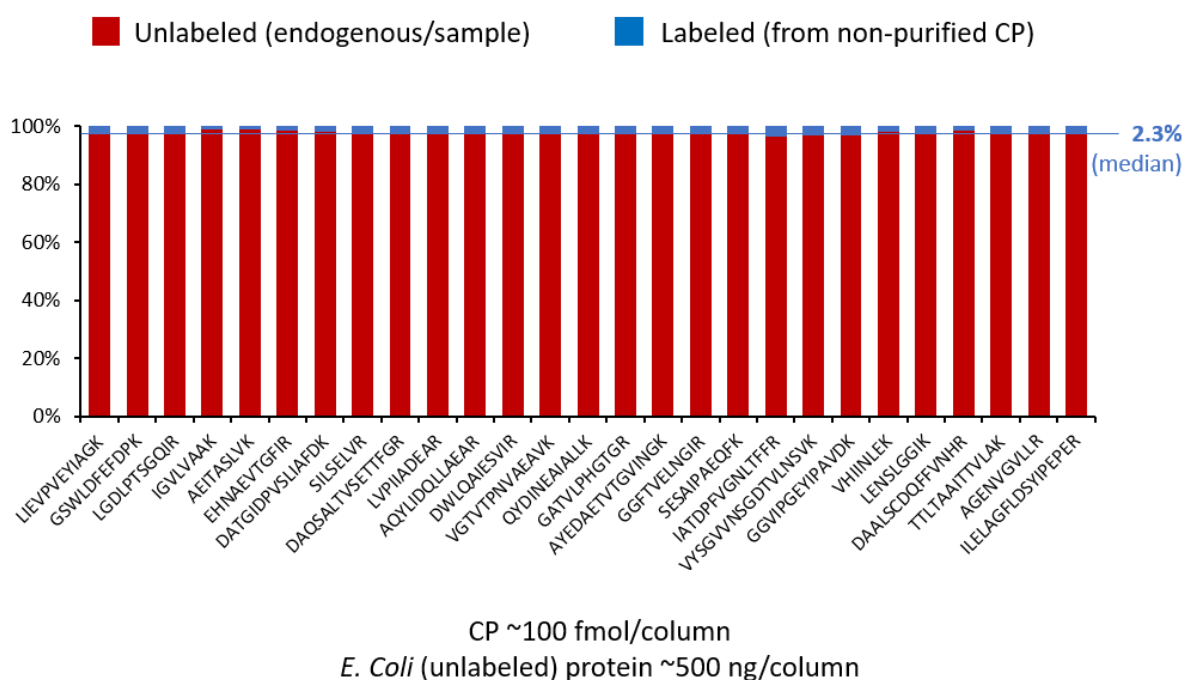


Figure S3. Contribution of extra background from the expression host (*E. coli*) introduced by spiking a unpurified CP standard (labeled, blue bars) compared to unlabeled *E. coli* proteins (unlabeled/endogenous, red bars). The protein amounts are typical for FastCAT experiments with CP at approximately 100 fmol and total protein of 500 ng loaded on column. The contribution of background (labeled) is ~2 % of the total protein. Nine abundant *E. coli* proteins (mainly ribosomal proteins and elongation factors) were considered. Their quantification was based on the abundance of at least three peptides having consistent L/H ratios (see Table S1). These peptides were detected as predominantly doubly charged ions, did not have missed cleavage sites and cysteine and methionine residues. Moreover, CP labeling efficiency exceeded 99 % and therefore all unlabeled peptides originated from background *E. coli* proteins.

A**CP05**

MGSAWSHPQFEKGGGSGGGSGGSAWSHPQFEKLEVLFFQGPAAAKVFADYEEYVKDFYELEPHKV
 AAAPGDVDRGLAGVENVTELKHLVDEPQNLIKLVQETLEFAKSELEEQLTPVAEETRLGPLVEQGR
 AATVGSLAGQPLQERQWAGLVEKLAQYQAGARNILTSNNIDVKIPTTFENGRNPNLPPETVDSLKV
 LNQELRWYIEIKAEFVEVTKITLLSALVETRNLAVSQVVHKAVLDVFEEGTEASAATAVKADLSGITG
 ARHVEDVPAFQALGSLNDLQFFRYSITYTGLSKEIPAWVVPFDPAAQITKQKWEAEPVYVQRNYP
 LELEDKNYLNNGEAGAPGKGEAGAPGEEDIQGPQKQWQQGDLQDTKLGEYGFQNALIVRDAFIGT
 LYEYSRSGSGHHHHHH

Apolipoprotein E (APOE) – Apolipoprotein D (APOD) – Alpha-1-antichymotrypsin (AACT) – Zinc-alpha-2-glycoprotein (ZAG) – Secretogranin-1 (SCG1)
 BSA (R-)peptides: native and scrambled

B**CP06**

MGSAWSHPQFEKGGGSGGGSGGSAWSHPQFEKLEVLFFQGPAAAKVFADYEEYVKDFYELEPHKV
 AAAPGDVDRGLAGVENVTELKHLVEEPNQLIKLVNELTEFAKVAAGAFQGLRGQTLAVAKTLDLG
 ENQLETLPDILLRENQLEVLVSWLHGLKFLPTNAIKILGQQVVPYATKTLSSVGGWNFGSQRVTIDS
 SYDIAKGNQWVGYDDQESVKADFVETVKFIPLIPIPERASFPVYKTDGAAPNVAPSDVGGGGGR
 VQVTSQEYSARNSEPDQEGELFQGVDPRTLGEALAPLSKLADLASDLLLQYLLQGGARVGEEDDEE
 AAEAEAEAEAEAFQLTFPLRTPAAETLSQLGQTLQSLKALSGNVIAWAESHIEIYGGATKLGDYGFN
 NALIVRDAFLGSFLYEYSRSGSGHHHHHH

Leucine-rich alpha-2-glycoprotein (A2GL) – Chitinase-3-like protein (CH3L1) – Contactin-1 (CNTN1) – Neurosecretory protein VGF (VGF) – Neuronal pentraxin-1 (NPTX1)
 BSA (R-)peptides: native and scrambled

Figure S4. Amino acid sequences of the CP05 and CP06 constructs used for the quantification of 10 neurological protein markers in CSF samples.

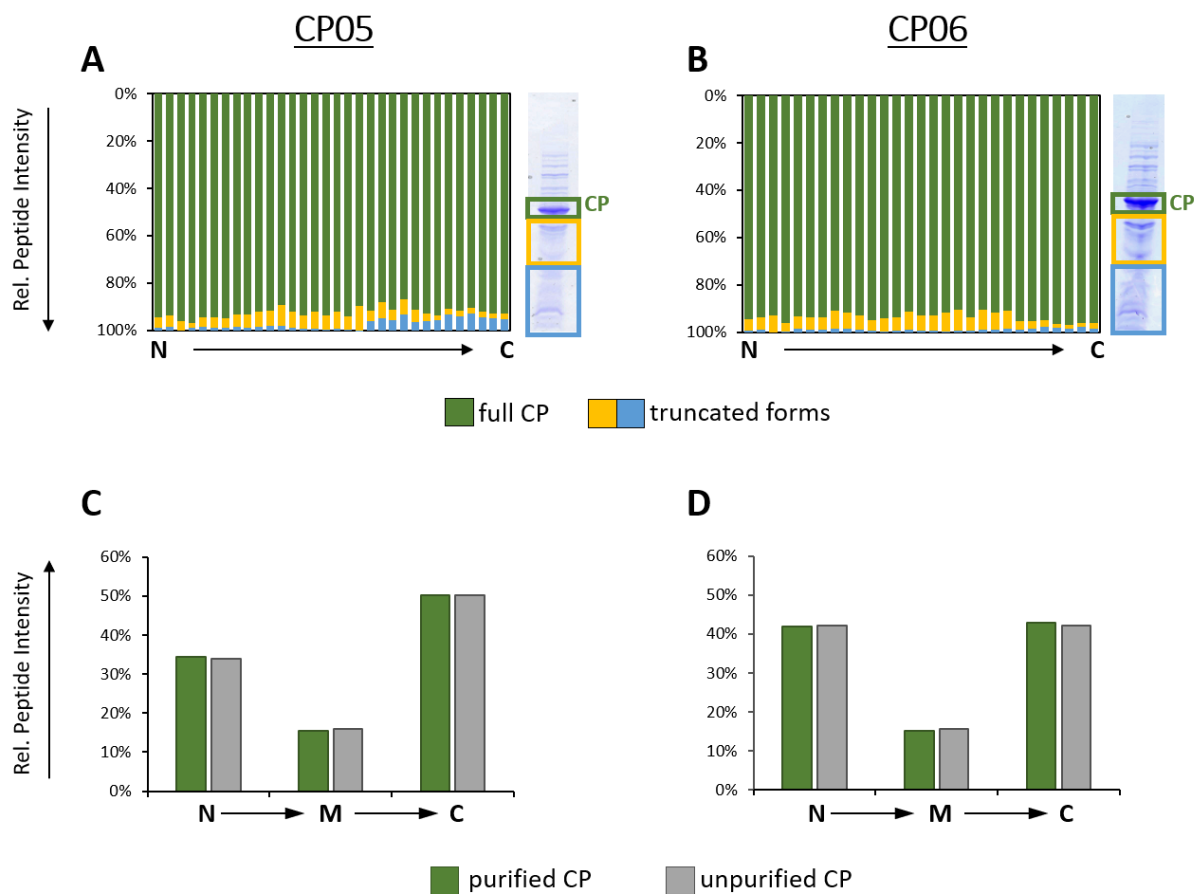
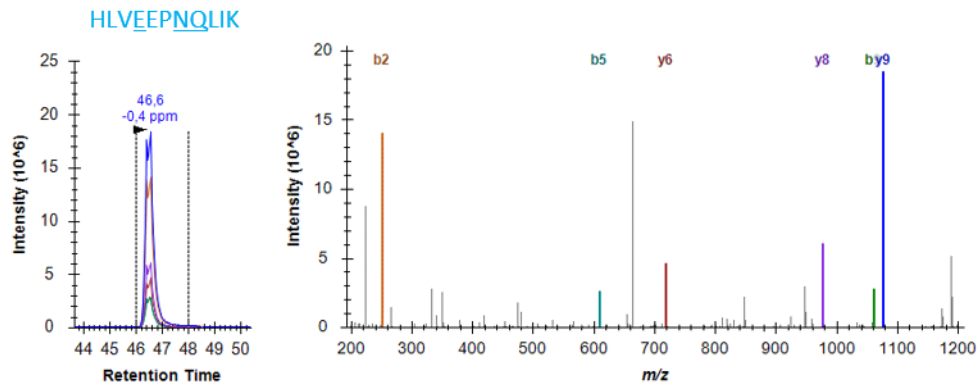
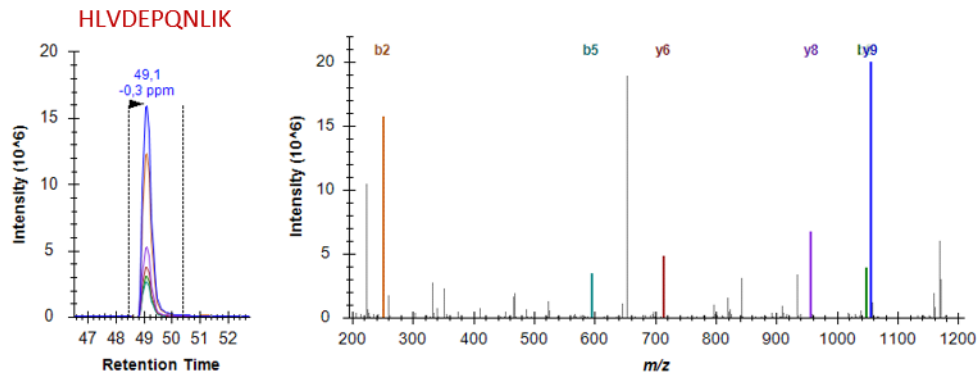
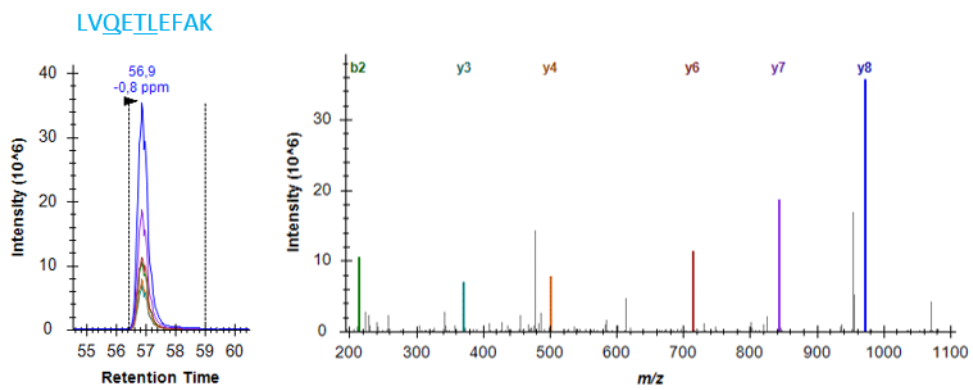
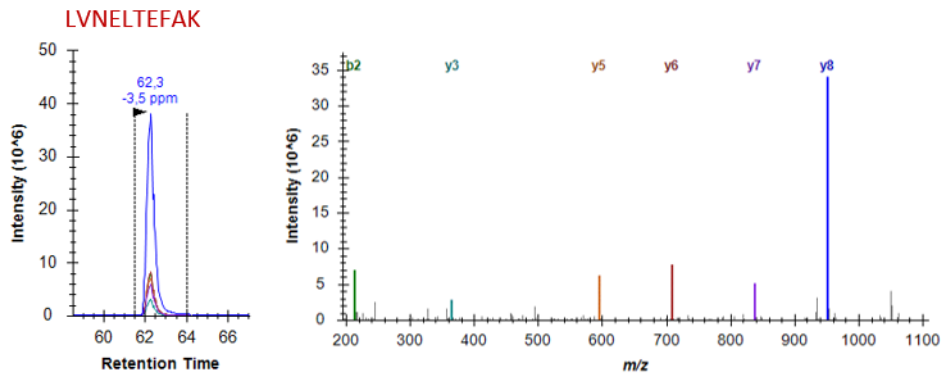
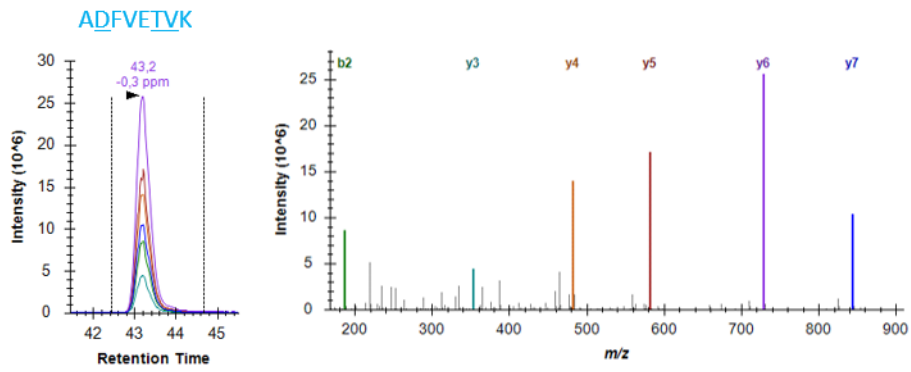
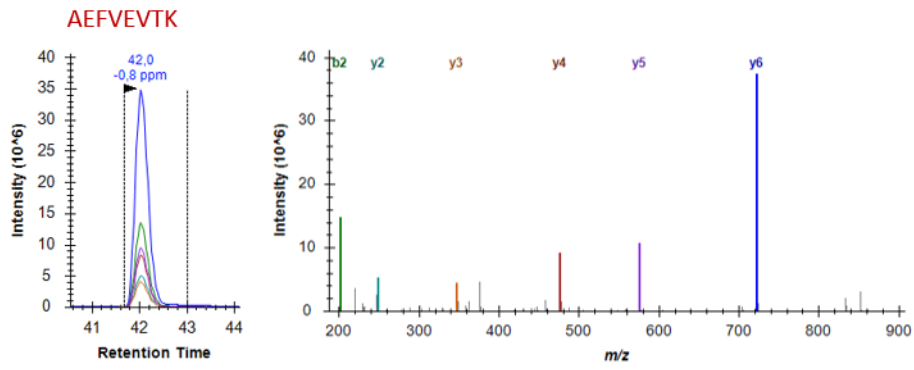
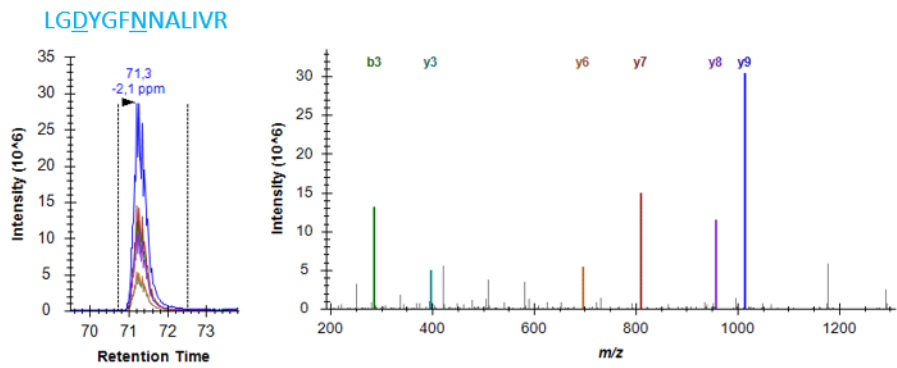
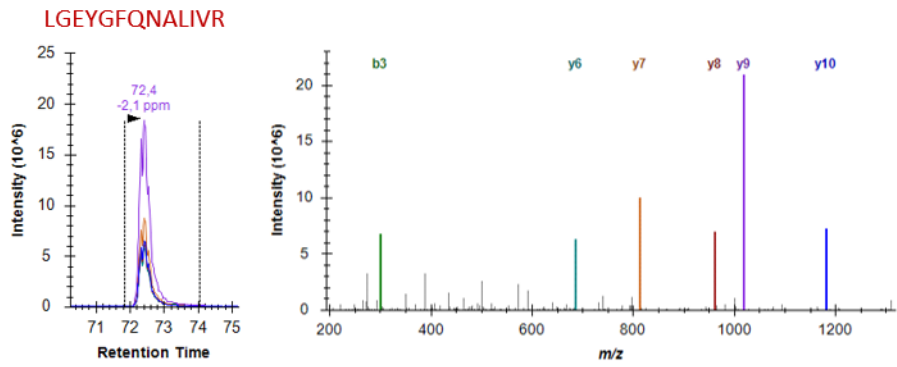


Figure S5. Truncation patterns for CP05 (A and C) and CP06 (B and D) standards.

A**B**

C**D**

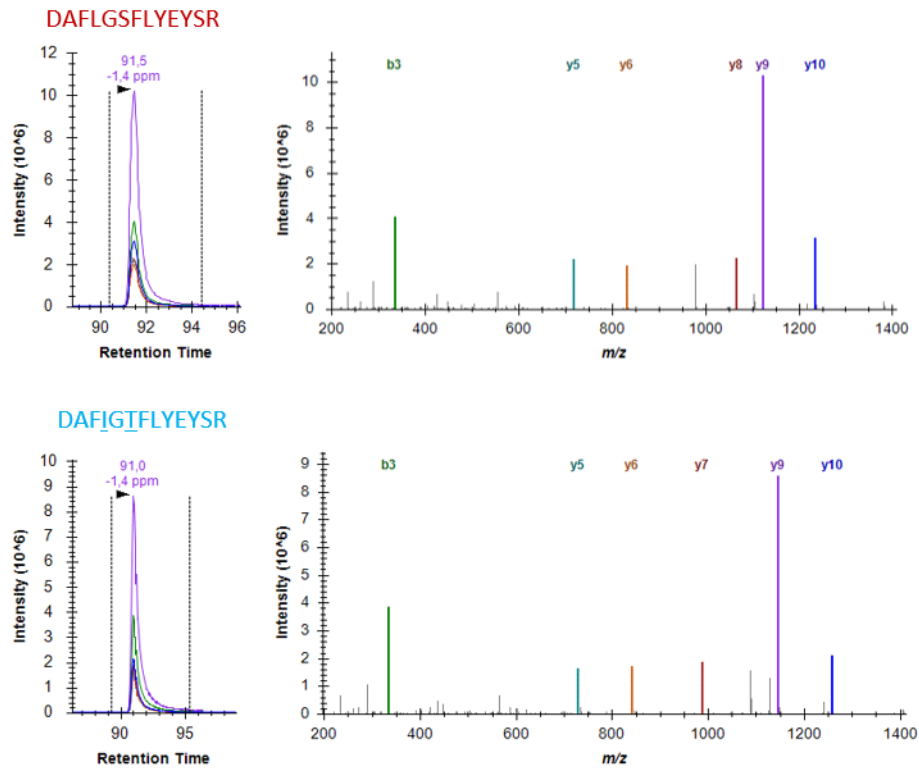
E

Figure S6. Tandem mass spectra of R-peptides in native (BSA, unlabeled) and scrambled forms (CP05/06, labeled) together with their chromatographic peaks: HLVDEPQNLIK (nat) and HLVEEPNQLIK (scr) (A), LVNELTEFAK (nat) and LVQETLEFAK (scr) (B), AEFVEVTK (nat) and ADFVETVK (scr) (C), LGEYGFQNALIVR (nat) and LGDYGFNNALIVR (scr) (D), DAFLGSFLYEYSR (nat) and DAFIGTFLYEYSR (scr) (E).

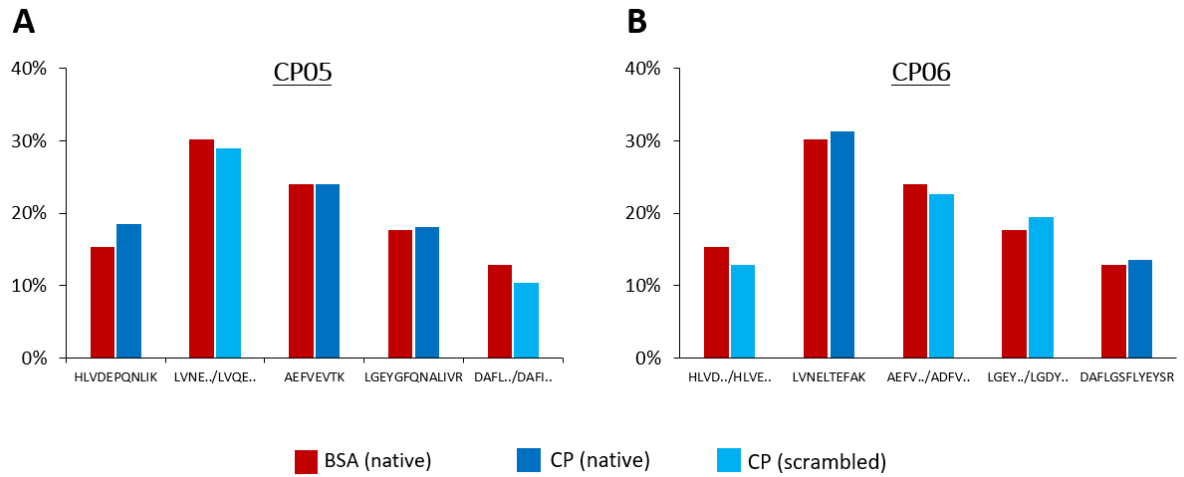


Figure S7. Matching relative peak areas of R-peptides for BSA (native) and both CP05 (**A**) and CP06 (**B**) obtained using Top6(y,b) fragment ions. Differences were <5 % for all pairs.

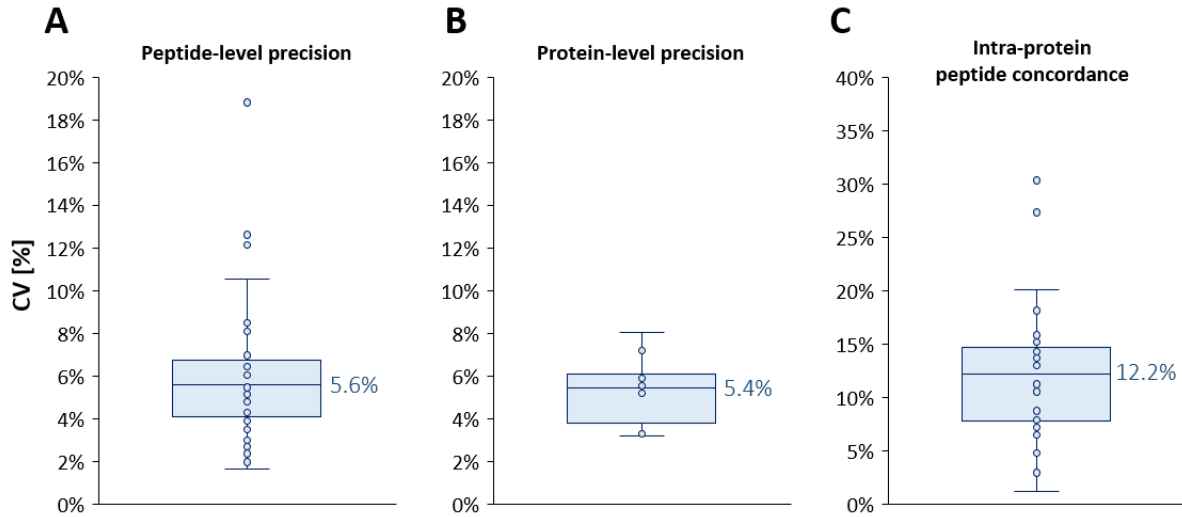


Figure S8. Precision of the FastCAT quantification estimated from the PRM analysis of the pooled CSF sample. Panels A and B show peptide-level and protein-level precision, respectively. The graphs are created with data acquired for all investigated proteins (10 target proteins and two CPs) based on three sample preparation replicates and two LC injections each. Panel C shows intra-protein peptide concordance obtained from the same data set. Median CV values are provided at the plots. Each box plot displays the median (line), the 25th and 75th percentiles (box), and the 5th and 95th percentiles (whiskers).

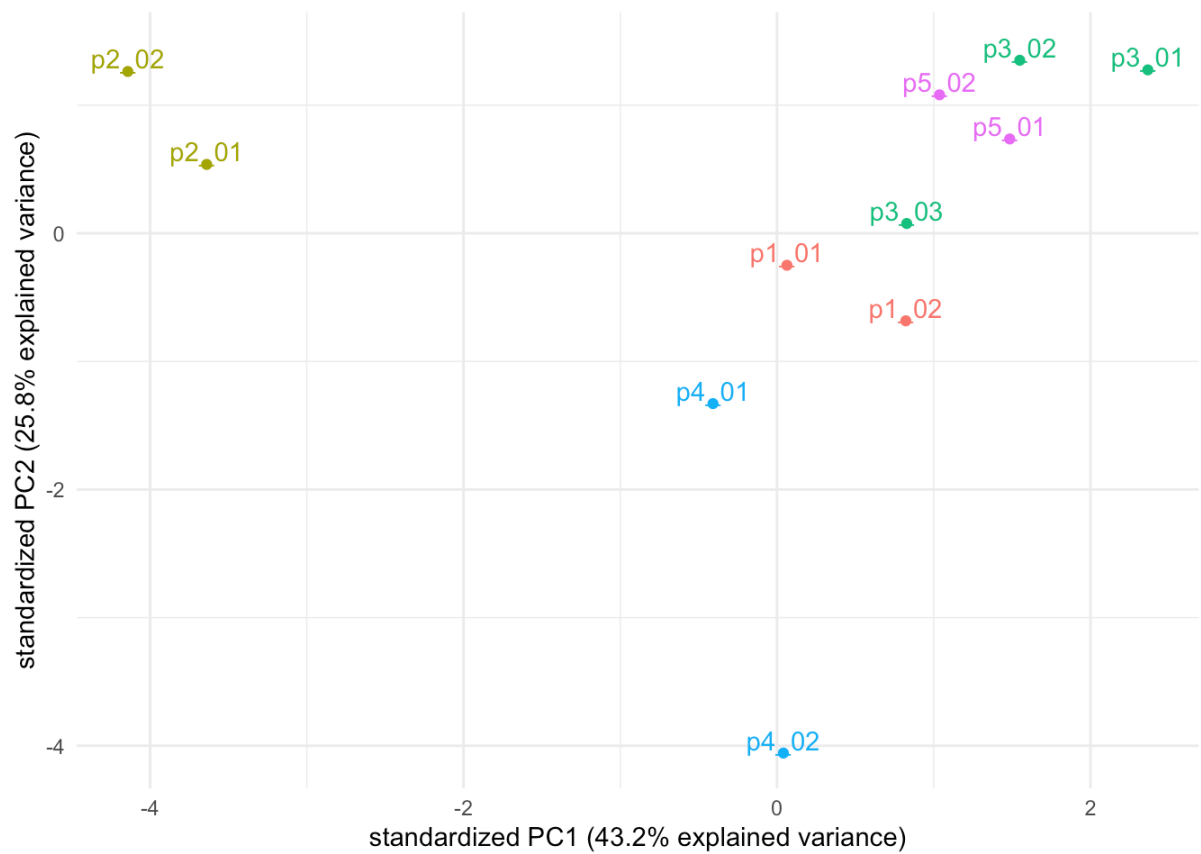


Figure S9. Principal component analysis (PCA) plot for all protein quantities (from Table S4) in patients 1-5 (p1-p5) with two (or three for p3) collections of samples. In this way, p2_01 and p2_02 stand for the first and second CSF collection from the patient p2, respectively.

Table S1. List of representative *E. coli* proteins (selected from most abundant proteins) used for estimating the amount of extra protein background introduced by spiking an unpurified CP standard. For each protein we provide UniProt accession number; sequences of peptides used for the quantification (three peptides per each protein); peptides retention times and charge states.

Protein	UniProt Accession	Peptide	RT	Charge state
DNA-directed RNA polymerase	P0A8V2	LIEVPVEYIAGK	76.7	2
		GSWLDFFEDPK	87.1	2
		LGDLPTSGQIR	57.5	2
Elongation factor Ts	P0A6P1	IGVLVAAK	53.2	2
		AEITASLVK	55.2	2
		EHNAEVTGFIR	53.7	3
50S ribosomal protein L4	P60723	DATGIDPVSLIAFDK	88.9	2
		SILSELVR	76.7	2
		DAQSALTVSETTFGR	69.9	2
Pyruvate dehydrogenase E1	P0AFG8	LVPIIADEAR	64.4	2
		AQYLIDQLLAEAR	89.5	2
		DWLQAIESVIR	105.7	2
50S ribosomal protein L1	P0A7L0	VGTVTPNVAEAVK	56.0	2
		QYDINEAIALLK	85.9	2
		GATVLPHTGTR	41.1	2
30S ribosomal protein S1	P0AG67	AYEDAETVTGVINGK	64.3	2
		GGFTVELNGIR	70.9	2
		SESAIPAEQFK	55.2	2
Elongation factor G	P0A6M8	IATDPFVGNLTFFR	89.9	2
		VYSGVVNSGDTVLNSVK	67.9	2
		GGVIPGEYIPAVDK	70.7	2
30S ribosomal protein S2	P0A7V0	VHIINLEK	51.9	2
		LENSLGGIK	51.7	2
		DAALSCDQFFVNHR	66.1	3
Elongation factor Tu1	P0CE47	TTLTAAITTVLAK	82.1	2
		AGENVGVLLR	61.4	2
		ILELAGFLDSYIPEPER	98.4	2

Table S2. Relative error of the CP05/06 quantification based on the scrambled R-peptides and using either MS1-derived (DDA) peptides peak areas or different MS2 fragment ions (PRM).

Native form	Scrambled form in CP	Δ RT [min]	CP with scrambled form	MS1-based (DDA)			MS2-based (PRM)						
				CP [fmol] by nat	CP [fmol] by scr	% error	CP [fmol] by nat	CP [fmol] by scr					
							Top1	% error	Top3(y)	% error	Top6(y,b)	% error	
HLVDEPQNLIK	HLVEE <u>P</u> NQLIK	2.5	CP06	275	290	5.4%	294	241	17.9%	241	17.7%	236	19.7%
LVNELTEFAK	LVQ <u>E</u> TLEFAK	5.4	CP05	270	295	9.0%	280	180	35.8%	245	12.5%	250	10.9%
AEFVEVTK	A <u>D</u> FV <u>E</u> TVK	1.2	CP06	275	274	0.4%	294	185	36.9%	268	8.7%	265	9.7%
LGEYGFQNALIVR	LG <u>D</u> YGF <u>N</u> NALIVR	1.1	CP06	275	312	13.4%	294	329	12.1%	341	16.4%	310	5.4%
DAFLGSFLYEYSR	DAF <u>I</u> G <u>T</u> FLYEYSR	0.5	CP05	270	188	31.0%	280	214	23.9%	203	27.4%	211	24.7%

Table S3. Protein concentrations (ng/mL) in 11 CSF samples from 5 patients diagnosed with multiple sclerosis. CSF was collected twice for four patients and three times for the patient #3. Patient age during the 1st collection is given in years (y), while the time difference (Δt) to the 2nd (and 3rd for patient 3) puncture is given in months (m).

Protein	Concentration in CSF [ng/mL]										
	Patient 1		Patient 2		Patient 3			Patient 4		Patient 5	
	time 1	time 2	time 1	time 2	time 1	time 2	time 3	time 1	time 2	time 1	time 2
	age, 43 y	Δt , 40 m	age, 61y	Δt , 17 m	age, 36	Δt , 10 m	Δt , 16 m	age, 30 y	Δt , 32 m	age, 30 y	Δt , 26 m
APOE	1489.28	1604.15	1106.19	922.07	1926.15	1807.81	1772.67	1854.96	2075.59	1996.34	1811.10
APOD	1197.68	1307.78	1185.15	1203.23	1284.59	1281.67	1297.31	1083.58	1060.41	1246.55	1327.29
AACT	759.84	684.61	765.58	723.96	580.30	564.96	770.17	810.35	1127.28	540.90	588.94
ZAG	100.05	113.18	102.01	96.32	85.98	85.89	87.58	102.12	108.05	85.19	85.76
SCG1	636.56	684.09	451.77	388.55	942.32	885.69	869.22	788.86	862.85	852.69	850.08
LRG1	67.98	71.42	77.14	80.84	50.02	47.78	114.29	38.45	92.91	27.13	33.82
CH3L1	51.77	35.09	35.16	33.37	53.82	49.85	44.63	38.90	27.48	27.90	25.74
CNTN1	238.90	256.07	159.70	148.28	229.68	230.81	229.77	241.20	268.62	257.04	246.15
VGF	139.34	163.29	62.64	60.65	175.94	133.49	141.99	73.35	98.14	110.97	99.89
NPTX1	75.67	86.02	42.50	37.18	77.01	72.52	71.98	64.28	81.85	70.87	68.53

Table S4. Chimeric protein (CP) standards used in this study together with their molecular weights, labeling efficiencies as well as amino acid sequences.

Chimeric Protein	Molecular weight	Labeling efficiency	Amino acid sequence
CP01	264.8 kDa	>99 %	<p>MGSAWSHPQFEKGGGSGGGSGGSAWSHPQFEKLEVLVFGPAAAKVFADYEEYVKDFYELEPHKVA AAFPGD VDRGLAGVENVTELKGSTLSAQQGSQFKNEPVSYLELINGRNIGDSYVA AKLDNGYNHLIEVVRLVLGIPTYG RFSPLVASNERL TEAEGSSLYIGGRSTDAEEDPQVIK FADQDNDLVNLRLSNFVDTTVAWLNRYRFFVSVTRVQL TDLNRDVFIPDVFNNYKILAIFFPGPSQYINVPYLVKVIWPYTYLYFRGTFFVSHKGHEISVFSLTHKAPLGLHF HASWLKIINNPEATQRAVYWVEHVSRSHYHVG SALAKAEQNGYGVTVHYEELSSAKGSAYAHAENTLRSEGD LTQITPPSALGVQVRAEVEAVQIIAESLKAANQLTDRPTIINVAISPSSDRGHGAGGADVLTYKDLNQGNSYVQD KSLVFDNHDNQRSEYTG LGAITEFRVVEFLDHLIDLGVAGFRIGALDTSRLIELL VRLADLLLQEERVNGEVPD VGAFIRITPDVVGAIVQEATTYFRATQLSEQLGSELTPKLLAADYADGVSQPRILSWNAVNL YGLKASEQPGL TAIHTAFLRLLPAQYEDGISAPRGIA TNDVGI AKDTFSFLTGSGRAINEGGFQSLPQKFG LNEGSEPPQAYGIGLKI TTHYTLNPRAILVYLVEKITPGWEENWAGALDVKINPQHSIPTLV DNGFTIWESRALGLEFNKINLDPYALYVIV EKTNFDDSA LFYASGESLKVLTGGDAGSNRLGSDVQPPGRVFLQ LLENQRNLSQTFGNIWRVFDLPDFL TWR GSTVQFAARDGAISHVVF KYFNAYLSDRFSEPYDSTLSDVIK NVANLEATKDADAIVQQT LAKGTQAEIVVAD VTKVGDVTEVAEVAFLASSKSVLLTLRIDALEGVYKALNLD FDKAITFPLFWENKLTLYGIDGSPPVRELPY YEEANGSRAVGV ELNKAPADPEAFKAIQVYLVEKLLNLQ AGEHLKPEFLKINPQHTIPTLV DNGFALWESRGL VVSGTRSLSDVSLTGRGNSYLILPPRISGSPSNHVTVVRNAA FSGDSYVSHRSVVNVGLGQALRSTQEEVDHIRS AVHDEVEFLKVST AIDAVISVGRVALQIQDVATSSRLEEISLR LLELHDNRLFNDFVLR LAVLDLSHNRA SLSG IQSHAFKTFDFGNPIHTLRDFGVELEDLQITRSTISSTTTV RTSTSSLTGNPRTSVATVAGGAVVGATKLLVPGS SSTTTTSSLRLLPDTTDEQLLTSALEEKSTGNIFAAKVF AVNSAGRIFADNVYGRGVAEDFAPSFVKH DGGSPIT GYIEKHYPNPAVRVVGSEADTGRDAPTTESYLRSTVPFATSESNRLVGGNDLDTPLIITRIAELEGLGSRIQYLE DSTRIGEAISSIRFITQIVDVTKAEDAGVYLVVARTYYDFG FVALDIKVVQVSVSAHALLWDLNDGKDVLS VAFSADNRLWDLAAGKLNNDLIARFQEALAGLSKLA AHDALGGA AKSAGSGVSTTAIEKIVQVQIDDVGKSEL DVFSDWLQVARFSQSDFGLDQGETLLRESLLEITIYHQG AAYQEAPVDAEVA VTPKISAFGLFTYSVFSIIAGS LKVFANPVQLEFYGFVPWSIIQRQVGASSGSSGAVSKIV QIQVGP AHLNVKVQDNSVLVEGNHEERVLP AIEA FQKFSEATLDEIIRFHGVALAFNALDSKENAILTDIWNITP FKVIVDILLK TILVDLQV GKLAELHAASVVAKEAG LEIELAPKAGFAGDDAPRSYELPDGQVITIGNERVAPEEHPV LLTEAPLNPKGYSFTTTAEREITSLAPSTIKEITA LAPSTIKQEYDESGPGIVHRQEYDESGPSIVHRSIELLEK LQASGDFPTTKEQNSPIYISRATVA AFAASEGHAPR YQVIVKSSDAQSQATASEAESKALSASSLIALSSRD PVLTA FQLSWELKDDYGITEDDII EVRATAWVIVHRVEL ELWLKVVLPDLAVLRALGIEEQSGGAARVDEYGFFLYWK YYLSDLARIEQADYLPTEQDILRVPTTGILEYPFD LDGIVFRTHIYPWFQNSSVILFLNKAGIAVEGDIKDAFGI VSYAVKDTALASTLIASQDARDFLSPGELELEV TLDKVFGQLATTYRLQYAPLNRISLEVTDRLLQPAPGTIEKFYPEDVAEELIQDITLRIGFPWSEIRTTHTAGFL ANDRLFYLQVKA AIELNRGDNILLTKVPPEVPPSKLVDFI INRSLVGVQKQIDVETVADRYIQA TDLSEASRNAE YDQFLEDIQKATAWVIVHRALSASSLIALSSRH L VDEPQNLIK YL YEIARQTALVELLKLGEYGFQNALIVRDAF LGSFLYEYSRLVNELTEFAKSGSGHHHHHH</p>

CP02	79.3 kDa	>99 %	MGSAWSHPQFEKGGGSGGGSGGSAWSHPQFEKLEVLFGQPAAAKVFADYEEYVKDFYELEPHKVA AAFPGD VDRGLAGVENVTELKLG DYGFNNALIVRDAFIGTFLYEYSRAAAAYVLQETPVVNALVDENEIVYRSDGEVEIA SEKVIDTAYEIIKGIILIGEGIGNAEEQAAEFLKVQNDSDIVFFDYR VLEATLAQDFSKLVTWYDNEFGYTNRYA GEDAAAGAAETL FVAKAEIEAVQIIAETLKVVEFLEHILDLGVAGFRIVQVNLDDV GKVG D VTEVAEAAVFLA TSKAGLAIEGDIKANEILSDIWNITPFKGAEDFAPTFVKIADLEGIYKALQLD FEYKALVFETWQGP LEVRWVAI DGEQY GEGSSRLIDDNVANALKVVDHAYEAVVIGAGGAGLRYLYDVARFHGATSINLVGDLDVTNPKGTVA HDGDYLVIAKTVEADAAHGSVTRTIEADAAHGSVTRVVELITYIATKYAVFDTGSRVTENVLAFIYKQLLFSAG AELNKLDLGT VVSPVSGPKLGANTLLELVIFGRAFGGNTQDFGRVVFQFLEASAGSKLNADTSLFILASKGQETST QPIATIFAWSRAGQSHLGLPIFGSAVEAKGVEPSHAISGARALIANGTGPYFYLPKNTVIASGGYGRAAAAQIN Y IRSGNVVPGYHGAVLRVALLGAGAGIGNPLGLLLKVPQVILAVGLPARHLVEEPNQLIKLVNELTEFAKSGSHH HHHH
CP03	42.3 kDa	>99 %	MGSAWSHPQFEKGGGSGGGSGGSAWSHPQFEKLEVLFGQPAAAKVFADYEEYVKDFYELEPHKVA AAFPGD VDRGLAGVENVTELKIGEEYISDL DQLRVIFLENYRLLSYVDDEAFIRVLDLERTLSDYNIQKES TLHLVLRITIT LEVEPSDTIENVKEGIPPDQQRIGDYAGIKVLGIDGGEGKEALDFFARVVGLSTLPEIYEKANELLINVKGVIFYE SHGKGNPTVEVELTTEKTFAEALRAADALLKTAGIQIVADDLTVTNPKNVNDVIAPAFVKVNQIGTLESIKA VDDFLISLDGTANKHLVDEPQNLIKLYE IARQTALVELLKLGEYGFQNALIVRDAFLGSFLYEYSRLVNELTEF AKGSGHHHHHH
CP04	43.0 kDa	>99 %	MGSAWSHPQFEKGGGSGGGSGGSAWSHPQFEKLEVLFGQPAAAKVFADYEEYVKDFYELEPHKVA AAFPGD VDRGLAGVENVTELKHLVDEPQNLIKLVNELTEFAKIGEEYISDL DQLRVIFLENYRLLSYVDDEAFIRVLDLE RTLSDYNIQKES TLHLVLRITITLEVEPSDTIENVKEGIPPDQQRILY E IARLVTDLTKAEFVEVTKIGDYAGIKVL GIDGGEGKEALDFFARVVGLSTLPEIYEKANELLINVKGVIFYESHGKGNPTVEVELTTEKTFAEALRAADALL KTAGIQIVADDLTVTNPKNVNDVIAPAFVKVNQIGTLESIKA VDDFLISLDGTANKLGEYGFQNALIVRDAFLG SFLYEYSRSGSHHHHHH
CP05	45.4 kDa	>99 %	MGSAWSHPQFEKGGGSGGGSGGSAWSHPQFEKLEVLFGQPAAAKVFADYEEYVKDFYELEPHKVA AAFPGD VDRGLAGVENVTELKHLVDEPQNLIKLVQETLEFAKSELEEQLTPVAEETRLGPLVEQGRAATV GSLAGQPLQ ERQWAGLVEKLAVYQAGARNILTSNNIDVKIPTTFENGRNPNLPETVDSLKVLNQELRWYEIEKAEFVEVTKI TLLSALVETRNLAVSQVVHKAVLDVFEEGTEASAATAVKADLSGITGARHVEDVPAFQALGSLNDLQFFRYSL TYIYTGLSKEIPAWVPFDPAAQITKQKWEAEPVYVQRNYPSELELDKNYLN YGEEGAPGKGEAGAPGEEDIQGP TKWQQQGDLDQDTKLGEYGFQNALIVRDAFIGTFLYEYSRSGSHHHHHH
CP06	46.0 kDa	>99 %	MGSAWSHPQFEKGGGSGGGSGGSAWSHPQFEKLEVLFGQPAAAKVFADYEEYVKDFYELEPHKVA AAFPGD VDRGLAGVENVTELKHLVEEPNQLIKLVNELTEFAKVAAGAFQGLRGQTLLAVAKTLDLGENQLETLPDILLR ENQLEVLEVSWLHGLKFPLTNAIKILGQQVPYATKTLSSVGGWNFGSQRVTIDSSYDIAGKNQWVG YDDQESV KADFVETVKFIPLIPERASFPFVYKTDGAAPNVAPSDVGGGGGRVQVTSQEYSARNSEPD EGELFQGVDPRT HLGEALAPLSKLADLASDLLLQYLLQGGARVGEED EEAEEAEAEAEAEAEERFQLTFPLRTPAAETLSLQGQTLQS LKALSGNVIAWAESHIEIYGGATKLG DYGFNNALIVRDAFLGSFLYEYSRSGSHHHHHH