1	Supplementary materials
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3	Gut microbiota differs between treatment outcomes early after fecal microbiota
4	transplantation against recurrent Clostridioides difficile infection
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17 Supplementary figures and tables





20 Figure S1. Study design of the clinical trial. Patients were randomly allocated to three 21 treatment arms, namely FMTv (FMT preceded by vancomycin), fidaxomicin, and vancomycin. 22 FMTv group was used to assess the effect of FMT on the gut microbiota and is highlighted 23 with a box in dashed lines. The vancomycin and fidaxomicin groups were used to compare the 24 outcome-associated changes induced by FMT to those induced by antibiotic treatment. Fecal samples were collected before any treatments (FMTv or antibiotics) (W0), and 1 (W1), 8 (W8), 25 26 and 26 weeks (W26) after treatments. Blue and orange colors refer to responders and non-27 responders at W8, respectively. Outcomes were determined based on combined clinical 28 resolution and a negative PCR test. The width of blue and orange colors is proportional to the 29 number of patients within each outcome.





35 and in donors. The four most abundant phyla are colored as indicated, and the less abundant

36 phyla are merged as "Others".



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Figure S3. Beta diversity of samples and taxa estimated with Bray-Curtis distance and visualized with nonmetric multidimensional scaling (NMDS) ordination. Bray-Curtis distance was estimated at the family level. Dots are samples colored differently according to their outcomes with ellipses indicating 50% confidence regions for clusters. Arrows point to the families directing the effect and the arrow lengths indicate the strength of the association. Comparison of beta diversity between outcomes was performed with permutational multivariate analysis of variance (PERMANOVA) with 10,000 iterations.





50 Figure S4. Beta diversity of functional potential estimated with PICRUSt2 based on Bray-

51 Curtis distance and visualized with NMDS ordination. The analysis was performed with

- 52 permutational multivariate analysis of variance (PERMANOVA) with 10,000 iterations.
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56 Figure S5. Correlations between the relative abundances of families and MetaCyc 57 pathways. Blue color indicates positive Spearman's rank-order correlation coefficient and red 58 color indicates negative correlations. The abundance of pathway labelled "Higher in responders" 59 is more abundant in responders and the one labelled "Lower in responders" is less abundant in 60 responders. The stars for each correlation refers to the significance level: *, 0.01 < P < 0.05; **, 0.001 < *P* < 0.01; ***, *P* < 0.001. 61



Figure S6. Tree plots of discriminant taxa as compared to pre-FMT. Comparisons were 65 66 performed between post-FMT and pre-FMT samples within each outcome. The smallest nodes 67 represent the genus level and the node in the center represents the kingdom bacteria. Node sizes 68 are proportional to the mean relative abundance at the given phylogenetic level. Nodes with labels have significantly different relative abundances between time points, where the blue 69 70 color indicates the taxa have increased over time, red color indicates decreased abundances, 71 and grey color indicates no significant changes in abundances.





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75 Figure S7. Importance and receiver operating characteristic (ROC) curve obtained by random forest modelling. (a) The top 30 most important genera are shown and their 76 77 importance scores are summed (the black vertical line) based on the family they belonged to in 78 descending order. Colors specify higher or lower abundances of corresponding families 79 between outcomes. The variable 'importance' is obtained from the random forest model built 80 based on the samples at week 1, via 10-fold cross validation with 100 iterations. (b) The ROC 81 curve describing the ability of the model to differentiate outcomes when applied at different 82 levels of sensitivity (true positive rate) and specificity (true negative rate). Area under the ROC curve (AUC) is used to measure the discriminatory power of the model (AUC 0.79). 83



Figure S8. Relative abundances of the top 30 most important genera at differentiating
outcomes identified by random forest modelling. The X axis lists the genera in a descending
order from left to right according to importance. Colors refer to outcomes.



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Figure S9. Correlation between the abundance ratios (*Escherichia* to *Blautia*) determined

by qPCR and 16S. The X and Y axis is the abundance ratio determined by qPCR and 16S

respectively, and log10 transformed for visualization. The strength of correlation is calculated

using the non-parametric Spearman's rank-order correlation coefficient.



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101 Figure S10. Prediction of outcomes based on the abundance ratio between Escherichia 102 and Blautia at week 1. (a) The distribution of abundance ratios between genus Escherichia 103 and Blautia for each outcome determined by qPCR. The optimal threshold of abundance ratios that best differentiated outcomes was obtained by the Youden's J statistic and calculated to be 104 105 5.6, where a larger value of abundance ratio than 5.6 is indicative of treatment failure. A prediction accuracy of 91.3% was obtained based on the chosen threshold. (b) The ROC curve 106 107 describing the ability of the model to differentiate outcomes when applied at different levels of 108 sensitivity (true positive rate) and specificity (true negative rate). Area under the ROC curve 109 (AUC) is used to measure the discriminatory power of the model (AUC 0.922). 110

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114 Figure S11. Observed richness for FMTv, vancomycin, and fidaxomicin treatments over

115 time, stratified by outcomes. Boxplot is used to show the distribution of richness. The median

- 116 richness is compared with the Wilcoxon rank sum test.
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121 Figure S12. Rarefaction curve for observed richness and Shannon diversity for samples 122 from the FMTv treatment. Donors (blue) are higher in alpha diversity than patients (red). 123 Although the richness did not reach saturation before 50,000 reads, Shannon diversity was 124 close to maximum after 2,000 reads. The black vertical lines mark the sequencing depth of 125 10,000 reads, which is the depth used for rarefaction before calculating alpha diversity. Dots represent the means of alpha diversity in corresponding sample types. Error bars show the 126 127 standard deviation of alpha diversity obtained from 100 random rarefactions at the 128 corresponding sequencing depth. 129

130 Table S1. Comparison of Bray-Curtis distance between time points or donors within each

131 outcome for FMTv treatment.

Time	Outcome	P adj	
0-1	Responders	0.002	
0-1	Non-responders	0.612	
0-8	Responders	0.002	
0-8	Non-responders	0.077	
0-26	Responders	0.002	
0-26	Non-responders	0.117	
1-8	Responders	0.961	
1-8	Non-responders	0.147	
1-26	Responders	0.889	
1-26	Non-responders	0.153	
8-26	Responders	0.961	
8-26	Non-responders	0.961	
0-Donor	Responders	0.258	
0-Donor	Non-responders	0.258	
1-Donor	Responders	0.385	
1-Donor	Non-responders	0.258	
8-Donor	Responders	0.258	
8-Donor	Non-responders	1.000	
26-Donor	Responders	0.258	
26-Donor	Non-responders	0.385	

135The comparison was performed between time points or between time and donor for each136outcome. Permutational multivariate analysis of variance (PERMANOVA, 10,000137permutations) was used to test the comparison. P values adjusted with the Benjamini-Hochberg138method are shown as P_{adj} . Significant P_{adj} values are in bold.

140 Table S2. Comparison of Bray-Curtis distances between time points within each outcome

141 for vancomycin and fidaxomicin treatments.

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Time	Outcome	Treatment	P adj
0-1	Responders	Vancomycin	1.000
0-1	Non-responders	Vancomycin	0.441
0-8	Responders	Vancomycin	1.000
0-8	Non-responders	Vancomycin	0.541
0-26	Non-responders	Vancomycin	0.590
1-8	Responders	Vancomycin	1.000
1-8	Non-responders	Vancomycin	0.441
1-26	Non-responders	Vancomycin	0.674
8-26	Non-responders	Vancomycin	1.000
0-1	Responders	Fidaxomicin	1.000
0-1	Non-responders	Fidaxomicin	1.000
0-8	Responders	Fidaxomicin	0.749
0-8	Non-responders	Fidaxomicin	0.590
0-26	Responders	Fidaxomicin	0.541
0-26	Non-responders	Fidaxomicin	0.441
1-8	Responders	Fidaxomicin	1.000
1-8	Non-responders	Fidaxomicin	0.928
1-26	Responders	Fidaxomicin	0.830
1-26	Non-responders	Fidaxomicin	0.593
8-26	Responders	Fidaxomicin	1.000
8-26	Non-responders	Fidaxomicin	1.000

143 The comparison was performed between time points within each outcome. Permutational

144 multivariate analysis of variance (PERMANOVA, 10,000 permutations) was used to test the

145 comparison. *P* values adjusted with the Benjamini-Hochberg method are shown as P_{adj} .