## Science Advances

## Supplementary Materials for

## Glucose-sensing glucagon-like peptide-1 receptor neurons in the dorsomedial hypothalamus regulate glucose metabolism

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Figs. S1 to S6



**Figure S1**| **A**, ICV Exn4 induced increased c-Fos expression in the GLP-1R neurons of paraventricular nucleus of hypothalamus (PVN) in the *GLP-1R-Cre* mice; **B**, Quantification of c-Fos positive cells ration (relative to total GLP-1R neurons), \*\*\*p<0.001 (t-test); **C**, ICV Exn4 induced increased c-Fos expression in the GLP-1R neurons of arcuate nucleus (ARC) in the *GLP-1R-Cre* mice; **D**, Quantification of c-Fos positive cells ration (relative to total GLP - 1R neurons), \*\*p<0.01 (t-test); **E**, AAV-DIO-synaptophysin-mGFP mediated GFP expression in the NTS of *Gcg-Cre* mice. **F**, To verify the injection site for retro-AAV-DIO-GFP in the DMH of *Gcg-Cre* mice, we mixed with AAV-mCherry during injection, which was showed by the representative brain slice. n numbers are indicated in each panel.



**Figure S2**| **A**, Experimental paradigm for retro-AAV viral injection in the PVN and DMH respectively of *Gcg-Cre* mice. **B**, Cre-dependent GFP expression in NTS demarks GLP-1 neurons which project to PVN (25%, mCherry), DMH (10%, GFP) and both PVN and DMH (65%, yellow). **C**, Cre-dependent hM3Dq expression in NTS of *Gcg-Cre* mice, which expressed c-fos after CNO injection. **D**, The representative image showed c-fos expression after chemogenetically activation of NTS GLP-1 neurons of *Gcg-Cre* mice. **E**, Cre-dependent retro-hM3Dq expression in NTS of *Gcg-Cre* mice, which expressed c-fos after CNO injection. **D**, The representative image showed c-fos expression in NTS of *Gcg-Cre* mice, which expressed c-fos after CNO injection. **D**, The representative image showed c-fos expression after chemogenetically activation of NTS GLP-1 neurons of *Gcg-Cre* mice. **E**, Cre-dependent retro-hM3Dq expression in NTS of *Gcg-Cre* mice, which expressed c-fos after CNO injection. **F**, GLP-1 concentration is showed in hM3Dq group than control group when chemogenetically activation of NTS GLP-1 neurons of Gcg-Cre mice (p>0.05, t-test).

**Fig. S3.** 



Figure S3|A, Experimental paradigm for viral injection of AAV-Cre in GLP1R<sup>t/f</sup> mice; B, Body weight gain increased 5 weeks after depletion of DMH GLP-1R expression. \*\*p<0.01, \*\*\*p<0.001; C, Epididymal fat pad is heavier than control group, \*\* p<0.01 (ttest); D, AAV-Cre-mCherry virus expression pattern in the DMH in GLP1R<sup>t/f</sup> mice; E, DMH brain slices were harvest freshly and mRNA was extracted immediately after. By using the GLP-1R specific primers, we did the quantitative PCR to analyze the mRNA expression. The representative gel bands showed GLP-1R expression are lower in AAV-Cre group (right two lanes) when compare to control group (left two lanes). F, Q-PCR results confirmed the knockout of GLP-1R expression in DMH , \*\*\* p<0.001 (t-test); G&H, We further confirmed the GLP-1R protein expression in DMH by western-blot. \*\* p<0.01 (t-test); I, GLP-1R expression was not changed in the PVN and ARC after depletion GLP-1R in the DMH. J, Energy expenditure analysis indicated that GLP-1R depletion mice showed lower energy expenditure in the dark period; K, In the postprandial glucose experiment setup, 1hr food intake amount was measured. There's no difference between control group and GLP-1R depletion group. While the baseline glucose level (t=-60min) was higher in GLP-1R depletion group when compare to control. \*p<0.05 (t-test) (L); Both 1hr food intake (t-test) (M) and baseline blood glucose level (t=-60min) (t-test) (N) were not different before Exn9 injection when investigating the GLP-1R antagonist-Exn9 effect on postprandial glucose (t-test).





**Figure S4**| **A**, Experimental paradigm for viral injection in *GLP-1R-Cre* mice. **B**, AAV-DIO-mCherry expression in the DMH of *GLP-1R-Cre* mice. **C**, C-Fos expression in the GLP-1R neurons within the DMH after CNO injection in *GLP-1R-Cre* mice. **D**, Baseline glucose levels before CNOinjection (fasting condition) were not different (p>0.05, t-test); **E**, Representative trace for miniature-EPSC before and after Exn4 treatment inDMH GLP-1R neurons of *GLP-1R-Cre* mice. **F**, Cumulative amplitude probability plots of mEPSCs in typical DMH GLP-1R neurons before and after Exn4 application. Inset in f: pooled data. n=9/3, cells/animals; p>0.05, paired t-test; **G**, Cumulative frequency probability plots of mEPSCs in typical DMH GLP-1R neurons before and after Exn4 application. Inset in g: pooled data. n=9/3, cells/animals; p>0.05, paired t-test. **H**, Representative trace for miniature-IPSC before and after Exn4 treatment in DMH GLP-1R neurons of GLP-1R-Cre mice. **I**, Cumulative amplitude probability plots of mLPSCs in typical DMH GLP-1R neurons before and after Exn4 application. Inset in g: pooled data. n=9/3, cells/animals; p>0.05, paired t-test. **H**, Representative trace for miniature-IPSC before and after Exn4 treatment in DMH GLP-1R neurons of GLP-1R-Cre mice. **I**, Cumulative amplitude probability plots of mLPSCs in typical DMH GLP-1R neurons before and after Exn4 application. Inset in i: pooled data. n=10/3, cells/animals; p>0.05, paired t-test. The data presenting style for panel E-J was adapted from our previous publication as described in main Figure2.



**Figure S5**[ **A**, Experimental paradigm for recording GLP-1R positive neurons within DMH by injecting AAV-DIOmCherry into *GLP-1R-Cre* mice; **B**, Action potential firing rate is not significantly changed when comparing vehicle group with Exn4 group. *p*>0.05 (paired t-test); **C&E**, Representative trace of voltage-step recording to isolate Na<sup>+</sup> and K<sup>+</sup> currents before and after treatment of GLP-1R agonist-Exn4; Exn4 did not alter voltage dependent sodium channel activity(**D**) but did change potassium channel activity (**F**) \**p*<0.05,\*\**p*<0.01 (repeat measurement, *posthoc* pair-t-test); **G**, Representative trace and stimulation protocol for measurement of *I*<sub>A</sub> before and after Exn4 treatment; **H**, I-V curve shows no significant shift after Exn4 treatment (repeat measurement, group effect *p*>0.05, group x voltage effect *p*>0.05);

Fig. S6.



**Figure S6| A**, Experimental paradigm for investigation of in vivo Ca<sup>2+</sup> image in the DMH of *GLP-1-R-Cre* mice. **B**, The expression of GCaMP6 and location for optic fiber; **C**, Experimental paradigm for fiber photometry recording in DMH during glucose uptake in *GLP-1R-Cre* mice. **D**, Blood glucose levels were significantly increased after 5min glucose uptake. \*\*\*p<0.001 (t-test) **E**, The expression of GFP after injection of AAV-DIO-synaptophysin-mGFP in DMH of *GLP-1R-Cre* mice; **F**, No GLP-1R afferents were observed in the IML region. **G**, Cre-dependent mGFP expression in the RPa originating from GLP-1R afferents. Chemogenetical stimulation of DMH GLP-1R neurons induced c-Fos expression in the NTS (**H**) and PRa (**I**) of *GLP-1R-Cre* mice. **J**, C-Fos positive cells are significantly higher in the NTS after CNO injection in the MM<sub>3</sub>Dq group when compare to control animals (AAV-mCherry), but no in the RPa (\*\*\* p<0.001, t-test). **K**, Cre-dependent ChR2 expression in the DMH of *GLP-1R-Cre* mice. **L**, Location of optic fiber in the optogenetic experiment setup. **M**, Schematic showing tracing of DMH GLP-1R+ neurons-liver projection: PRV-GFPand AAV-DIO-mCherry were injected into liver and DMH respectively in *GLP-1R-Cre* mice; **N**, Representative imageshowing GLP-1R+ neurons (red) and liver projecting neurons (green). Py:pyramidal tract; RPa: raphe pallidus nucleus; IML:intermedio-lateral nucleus; IMM: intermedio-medial nucleus