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Supplementary Materials for

The risk variant rs11836367 contributes to breast cancer onset and metastasis by attenuating Wnt signaling via regulating NTN4 expression

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Tables S1 to S9

Fig. S1.





(A) Flowchart of studying GWAS breast cancer susceptible SNP rs17356907 in enhancer region.
(B) Luciferase reporter assays after transient transfection of MCF-7 breast cancer cells with different constructs. DNA fragments containing different alleles of the GWAS-SNP rs17356907 and its LD-SNPs in enhancer or cis-regulatory element (CRE) were cloned into *NTN4*-promoter-

driven luciferase constructs. Luciferase signals were normalized to renilla signals. (C) Schematic illustrating the sgRNAs designed to target different loci in CRISPR-activation (CRISPRa) and CRISPR-interference (CRISPRi) assays. AC-sgNC and KB-sgNC contain non-targeting control guide RNA. Negative control AC-sgSD and KB-sgSD contain a guide RNA targeting a locus, which is the same distance from the NTN4 promoter as enhancer. Positive control AC-sgPro and KB-sgPro target the NTN4 promoter. AC-sgEnh-1, AC-sgEnh-2, KB-sgEnh-1 and KB-sgEnh-2 target enhancer. AC-sgCRE and KB-sgCRE target CRE. (D and E) sgRNAs illustrated in (C) were used for CRISPRa in MCF-7 cells (D) or for CRISPRi in MCF10A cells (E), and NTN4 mRNA expression was measured by qRT-PCR. (F) Luciferase reporter assays after transient transfection of MCF10A and MCF-7 cells with different constructs. DNA fragments containing different alleles of the GWAS-SNP rs17356907 and its LD-SNPs in enhancer and PRE1 were cloned into NTN4-promoter-driven luciferase constructs. Luciferase signals were normalized to renilla signals. (G) The luciferase reporter assay was performed after transient transfection of MCF-7 cells with NTN4-promoter-driven luciferase vectors containing different haplotypes of 4 SNPs (rs10859915, rs17356907, rs11836367 and rs61938093). Luciferase signals were normalized to renilla signals. (H) Gel image showing PCR amplification of the rs11836367 region in wild type (WT) and 151 bp deletion clones (Δ 151) in MCF10A and MCF7 cell lines. M: DNA Marker. (I) Bar plot showing allele-specific T-sgRNA and C-sgRNA preferentially introduced the T and C alleles of rs11836367 respectively when coupled with Cas9-NG. Data are represented as mean \pm SEM. * $P \le 0.05$, ** $P \le 0.01$, *** $P \le 0.001$. P values in (B, D, E, F and G) were determined by two-way ANOVA followed by Dunnett's multiple comparisons test. P values in (I) were determined Student's t-test. Data are representative of three independent experiments.

Fig. S2.



Fig. S2. rs11836367 allele specifically influences GATA3 binding affinity. Related to Fig. 2.

(A) Predicted binding affinity of GATA2 and GATA3 with different allele of rs11836367 through PERFECTPS-APE software. (B) ChIP-qPCR confirmed the capture-efficiency of several sgRNAs used for *in-situ* Capture assay. sgEnh1-1/2/3/4/5 target enhancer containing rs11836367, and non-targeting sgGal4 is a negative control. sgEnh1-3 and sgEnh1-5 have higher capture efficiency for DNA containing the rs11836367 locus. (C) Overexpression of GATA3 in 293T nuclear extracts validated by western blotting. (D) RNA-seq of T47D (GSE99479) showed that overexpression of GATA3 in T-47D increased *NTN4* expression. (E) qRT-PCR analysis of the *GATA3* and *NTN4* expression in the control or *GATA3* knockdown cells line MCF7-dCas9-KRAB by CRISPRi. (F) The correlation analysis of *NTN4* expression with *GATA3* expression in normal breast tissues and breast cancer tissues from TCGA using GEPIA. Data are represented as mean \pm SEM. * $P \le 0.05$, ** $P \le 0.01$, *** $P \le 0.001$. P value in figures (B, D and E) were calculated using the Student's t-test. Data in figures (B and E) are representative of two to three independent experiments.



Fig. S3. Loss of *Ntn4* in *PyMT* mice affected mammary tumor progression, metastasis and cancer stem cells. Related to Fig. 3.

(A and B) Expressions of Ntn4 mRNA and NTN4 protein in the mammary gland were examined by qRT-PCR (A) and Immunohistochemical staining (B) respectively. n = 3 for both groups. As indicated by the arrow, *Ntn4* is mainly expressed by myoepithelial cells in wild-type (WT) mammary glands, but is absent in Ntn4^{-/-} mice. Scale bar, 20 µm. (C) The expression of Ntn4 is higher in myoepithelial cells (MEP) than in luminal epithelial cells (LEP) based on microarray data GSE7513. (D) Quantification of Ki-67, ER, PR and HER2 in different stages of tumors of 13-week-old PvMT; $Ntn4^{wt}$ and PvMT; $Ntn4^{-/-}$ mice. n = 3-6 for both groups. (E) Representative images of IHC staining (upper) and quantification (lower) of Ki-67, ER, PR and HER2 in the late-stage tumors of PyMT; $Ntn4^{wt}$ and PyMT; $Ntn4^{-/-}$ mice. n = 3-6 for both groups. Scale bar, 20 μ m. (F) The size of metastatic foci in the lung sections was measured in PvMT; Ntn4^{wt}, PyMT; $Ntn4^{+/-}$ and PyMT; $Ntn4^{-/-}$ at the endpoint. n > 25 for each group. (G) Representative images of IHC staining for Ki-67 (left) and quantification of Ki-67 labeling index (right) in the lung sections of PyMT; $Ntn4^{wt}$ and PyMT; $Ntn4^{-/-}$ late-stage mice. n = 3 for both groups. Scale bar, 50 µm. (H) qRT-PCR examination of Pymt and Krt8 expression in blood samples was used as a measure of circulating cancer cells in PyMT; $Ntn4^{wt}$, PyMT; $Ntn4^{+/-}$ and PyMT; $Ntn4^{-/-}$ mice. n = 8 for each group. (I) Kaplan-Meier analysis of overall survival in PyMT; $Ntn4^{wt}$ (n = 16), PvMT; $Ntn4^{+/-}$ (n = 14) and PvMT; $Ntn4^{-/-}$ (n = 18) mice. Mice were euthanized when the largest tumor reached 2 cm in diameter. (J) CD24⁺CD90⁺ cells in *PyMT;Ntn4^{wt}* (n=3) and *PyMT;Ntn4^{-/-}* tumors (n=3) were quantified by flow cytometry. (K) Limiting dilution assay demonstrated the tumor-initiating frequencies of primary tumor cells isolated from PyMT;Ntn4^{wt} and PyMT;Ntn4^{-/-} mice. (L) Pictures of tumors in three groups transplanted with 50, 500, 5000 of PvMT;Ntn4^{wt} and *PvMT;Ntn4*^{-/-} tumor cells. Mice transplanted with 5000 cells were sacrificed after 4 weeks of transplantation, and the mice transplanted with 50 and 500 cells were sacrificed after 8 weeks of transplantation. Arrowheads point to fat pads (no tumors). Scale bar, 1 cm. (M) Tumor weight transplanted with 5000 tumor cells of PyMT;Ntn4^{wt} and PyMT;Ntn4^{-/-} after 4 weeks of transplantation. Data are represented as mean \pm SEM. * $P \le 0.05$, ** $P \le 0.01$, *** $P \le 0.001$. P values in figure (A, C, D, E, G, J and M) were calculated using Student's t-test. Ptrend values in figures (F and H) were determined by one-way ANOVA and P values between two groups were determined by one-way ANOVA followed by Dunnett's multiple comparisons test. Data in figure (I) was first calculated by comparing three groups for log-rank P value and P_{trend} value, and P values between two groups were further calculated using log-rank test adjusted with Bonferroni correction.



Fig. S4. The effects of *NTN4* overexpression and silencing on breast cancer cells' biological behaviors. Related to Fig. 4.

(A) Pan-cancer expression of *NTN4* in different tumor tissues and corresponding normal tissues analyzed by UALCAN. Boxes represent the median and interquartile range; whiskers extend to the minimum value and maximum value. (B)

Boxplot showing relative expression of NTN4 in normal breast tissues (Normal), luminal breast cancer (Luminal), HER2-positive breast cancer (HER2), and triple-negative breast cancer (TNBC) tissues in the UALCAN database. Each box represents the median and interquartile range. (C) Kaplan Meier plotter reveals that NTN4 expression is positively correlated with the overall survival (OS), the distant metastasis-free survival (DMFS), post-progression survival (PPS) and relapse-free survival (RFS) of breast cancer patients, which is based on NTN4 mRNA level from the database of the online platform KMplot. Auto select best cutoff was chosen in the analysis. HR, hazard ratio. (D and E) gRT-PCR (D) and western blotting (E) detected NTN4 expression in a panel of benign human breast epithelial cell line (MCF10A) and breast cancer cell lines (other cell lines). (F) The expression of NTN4 is downregulated in human breast cancer stem cells (CD24⁻CD44⁺) based on expression data GSE7513. (G) Cellular proliferation was analyzed by CCK8 in NTN4 inducible overexpression cell lines SKBR3-dCas9-VP64-MPH with 50 ng/mL Dox. (H-J) Representative images of the cellular migration (H), invasion (I) and sphere formation (J) in the indicated NTN4 inducible overexpression cell lines SKBR3-dCas9-VP64-MPH and MDA-MB-231-dCas9-VP64-MPH as described in Fig. 5c. Scale bar, 50 µm. (K-M) Representative images of the cellular migration (K), invasion (L) and sphere formation (M) analyses in NTN4 knockdown SUM159-dCas9-KRAB cell line as described in Fig. 5h. Scale bar, 50 μ m. Data are represented as mean \pm SEM. * $P \le 0.05$, ** $P \le 0.01$, *** $P \le 0.001$. P values in figure (A, B, F and G) were calculated using Student's t-test. P values in figure (C) were calculated using logrank test. BLCA, Bladder urothelial carcinoma. BRCA, Breast invasive carcinoma. CESC, Cervical squamous cell carcinoma and endocervical adenocarcinoma. CHOL, Cholangio carcinoma. COAD, Colon adenocarcinoma. ESCA, Esophageal carcinoma. GBM, Glioblastoma multiforme. HNSC, Head and Neck squamous cell carcinoma. KICH, Kidney chromophobe. KIRC, Kidney renal clear cell carcinoma. KIRP, Kidney renal papillary cell carcinoma. LIHC, Liver hepatocellular carcinoma. LUAD, Lung adenocarcinoma. LUSC, Lung squamous cell carcinoma. PAAD, Pancreatic adenocarcinoma. PRAD, Prostate adenocarcinoma. PCPG, Pheochromocytoma and paraganglioma. READ, Rectum adenocarcinoma. SARC, Sarcoma. SKCM, Skin cutaneous melanoma. THCA, Thyroid carcinoma. THYM, Thymoma. STAD, Stomach adenocarcinoma. UCEC, Uterine corpus endometrial carcinoma.





Fig. S5. *NTN4* inhibits the expressions of Wnt/β-catenin signaling target genes. Related to Fig. 5.

(A) Top ten results of PANTHER analysis showing frequencies of differential expression genes from RNA-seq result to pathways indicated. (B) GSEA analysis of RNA-seq data for two WNT

target gene signatures, which were NUSSE_WNT_TARGETS (Blue curve: NES = -1.71, FDR q-val = 0.003) and FEVR_CTNNB1_TARGETS (Red curve: NES = -1.45, FDR q-val = 0.027). **(C and D)** qRT-PCR analysis of the Wnt/ β -catenin target genes *CCND1, CD44, MYC, MMP7* and *POU5F1* in the *NTN4*-inducible overexpression cell lines SKBR3-dCas9-VP64-MPH and MDA-MB-231-dCas9-VP64-MPH (A) and in the *NTN4* knockdown cells line SUM-159-dCas9-KRAB (B). **(E)** Western blotting analysis of protein levels of p-LRP6, LRP6, p-GSK3 β , GSK3 β and β -catenin in total cell lysates and nuclear fractions prepared from *NTN4*-knockdown cells SUM-159-dCas9-KRAB. **(F)** Representative images of IHC staining (left) and quantification (right) of AXIN2 and CCND1 in *PyMT;Ntn4^{wt}* and *PyMT;Ntn4^{-/-}* mammary tumors. n = 3 in both groups. Scale bar, 20 µm. Data are represented as mean ± SEM. * $P \le 0.05$, ** $P \le 0.01$, *** $P \le 0.001$. *P* values in figures (C, D and F) were calculated using Student's t-test. Data in figures (C and D) are representative of three independent experiments.



Fig. S6. *NTN4* inhibits breast cancer cell proliferation and metastasis by antagonizing Wnt/ β -catenin signaling through its NTN4_{LA} domain. Related to Fig. 6.

(A) Western blot detection of HA-tagged full-length or truncated NTN4 protein in cell lysates and supernatants of 293T cells, which were transiently transfected with full-length or truncated *NTN4* constructs. (**B-D**) Cellular proliferation (B), migration (C) and cancer stem cell features (D) were analyzed in the MDA-MB-231 cell line treated with different condition mediums (CM). Scale bar, 50 μ m. Data are represented as mean \pm SEM. *p \leq 0.05, **p \leq 0.01, ***p \leq 0.001. *P* values in figures (B-D) were calculated using the Student's t-test. Data are representative of three to six independent experiments.

Supplementary Tables:

Table S1. Summary of LD-SNPs.

Table S2. Gene Expression Omnibus (GEO) identifiers for ChIP-seq, DHSs and ATAC-seq datasets.

Table S3. Summary of LD SNP with enhancer modification in BCAC, GTEX and RegulomeDB database.

Table S4. Characteristics of the breast cancer patients and cancer-free control.

Table S5. Genotypes and allele frequencies of the SNP in enhancer and their associations with the risk of breast cancer.

Table S6. Association of haplotypes and diplotypes of SNPs in enhancer with breast cancer risk.

Table S7. Primer sequences.

Table S8. List of reagents and antibodies.

Table S9. WNT target gene sets used for GSEA.