

Supplementary File S5. *NtNRAMP3* promoter analysis.

A. *NtNRAMP3* promoter sequence

Sequence of *NtNRAMP3* promoter (*promNtNRAMP3* + 5'UTR + START codon) located in the scaffold AWOK01S026429 (position 12116...13800):

AGAATCTGCGAGCATCTCAAAGGAATCTATGGAATGCTCGGGAAGAAGCGAATACCACGTCAAGGCTCCCCTCGT
GAGTGTCTCCTCGAATTTCTTCAGCAAAACAGATTCAATCTCGTGGGAGCTAAAATCGTTCCCCTCACCACCG
TTGCATAGGTGATAATATGTTTATGAGAGTCCGAAGTCCCTCAAGATTGTGTGTATTGTTAACACCATTATTTGC
CATTTTCTGTAATTTTTGCTAAGAACATATAATCAAAACATGTTAGTAAAAAAGACAAGGATCAACTTAATTACA
CGATTTTCTAGGCCCCACGATGGACGCCAAACTATTTGCCCGTAAAACGATACAGTTGAATTTATACGTACTTTC
TAGACAAGTGAATTAATTCGATCCTGAAATAATAAAAAGAATTGAATAAATATGCAATATTTAGCCTTAATATGAA
AATGAAATAGCAGAAACCGTAGTTTCGGGAGCAGGGCTTCCGGGCACAACATCGATGAAATCAAAAAGGCGAGAGG
ATAAAGTTGTATAAAGCTTTTAATATATTGTAGCGTAAAGTATGCTAGAAAATGCGTCCCTTTTACAATGATAACA
GAGCTCATTATTTATAGTTGCATCTAGGGAATAAGGTCCTAGGATTATGTCTTCTTTAATATGAGGGCTATTAA
AGAATGTGTAACGGCAGGCATGAATGATAAATTTTTGTAAACGGCCAATGACTAAATACTGTAGAATATTTCTTC
ATTAAATGCTATAGTGACAGGCATTTATTGCATATTTATGATCATTATTCTTTTTCGGTAATAAATGAAACAGTT
GTCTTCGGATTTAGCTATCCTCTGCCTTCGGCTTCACGTGTAACCTCTTATAACAATTACTTAACATACCATATT
TTACCCATACATATAGATTTATAAAAAATATAAAACACAGAAAAACTTTACTGCATTTGCACCAAAAAGGTAAT
GGAAAGGATCAAAGTATTTAATTTTTTTGTGTTAATTTGGAATATCCGTTATTTATTTTTCAAAGAAAAAATCAA
ATATTTCAAATTACAGTAAAATAAATATAAAACCTTCAAATAAATTATCGTCCAAGAGATAGGAAAAAGTAAAA
ACAATTTAGTCTTAGTTTCACAATGTCAGACTCTCCCTGTTCTTCCCCTCATCACTAAATAAGAAAAAGTCAATC
GATCTCTTATGCTTCTATCTCAAATTAAGAAAAATTTCAATTATATCTATTTTAAGCGGACTTATTTTCATTCAAA
TAACCAGAAAAAAGATTTTCCAAAACCTAGGGGTTGTATCCTTTACCGCTTACTAAACTAAGAAAGACATTGATAAA
TCCAAGATTATAAATATAACATCGATAAATAAACATAATTCCAATTATAGTTGCTATACTTTTGGCTTTACATTG
TATTTTATTTTTTAGCTAAATTCATACTCATTTCAAAGTGAAAAAAGAAAAACAAAATAAAAGGAGAAAGGAA
ATTGGGGTGCAAATAGAAAGCGGTAGCGTAATGGTAGTTTTCAAATAATCATTGCTCTTATCGTCTGAAAAAGAAA
ACCAGAAAAAGATCCAAAGCCTCCACTATAAAACAACATCACTTCGCACAAACCGAACTCACCTGTACCTTTTAA
CGCTAATATCTCTTTACAGATTTCTCTTCTAAATG

AGAATC – primers used for *promNtNRAMP3* amplification (sequence of the forward primer and complementary sequence of the reverse primer)

ATG – START codon (not amplified with *NtNRAMP3* promoter sequence)

Sequence length: 1684 bp

B. *In situ* analysis of the *NtNRAMP3* promoter

In situ analysis of the *NtNRAMP3* promoter sequence was performed by PlantCARE software (<http://bioinformatics.psb.ugent.be/webtools/plantcare/html/>) to identify the *cis*-acting elements within its region. For the analysis, a 1684 bp sequence of the *NtNRAMP3* promoter (counting from the START codon upstream) was used.

Table 1. Cis-acting regulatory elements present in *NtNRAMP3* promoter sequence.

<i>cis</i> -acting regulatory element	Sequence	Position in <i>NtNRAMP3</i> promoter sequence	Function	Reference
METAL RESPONSIVE SEQUENCES				
MRE1	TGC(A/G)C(T/G/C/A)C	959	sequence of the Metal Response Element 1	1
IDE2	CA(A/C)G(C/T)(A/C)A	56	sequence of the Iron Deficiency Response Element 2	2
GENERAL REGULATORY ELEMENTS				
TATA-box	ATTATA	1394	core promoter element around -30 of transcription start site	3
CAAT-box	(C)C(A)AAT	multi-position	common <i>cis</i> -acting element in promoter and enhancer regions from <i>Pisum sativum</i> or <i>Arabidopsis thaliana</i>	4
PHYTOHORMONE RESPONSIVE SEQUENCES				
ABRE	ACGTG	861	<i>cis</i> -acting element involved in the abscisic acid responsiveness from <i>Arabidopsis thaliana</i>	5
CGTCA-motif	CGTCA	58	<i>cis</i> -acting regulatory element involved in the MeJA-responsiveness from <i>Hordeum vulgare</i>	6
LIGHT RESPONSIVE SEQUENCES				
BOX4	ATTAAT	387	part of a conserved DNA module involved in light responsiveness from <i>Petroselinum crispum</i>	7, 8
G-box	CACGTC	56	<i>cis</i> -acting regulatory element involved in light responsiveness from <i>Zea mays</i>	9
GATA-motif	GATAGGG	1108	part of a light responsive element from <i>Arabidopsis thaliana</i>	10
TCCC-motif	TCTCCCT	1157	part of a light responsive element from <i>Spinacia oleracea</i>	11
chs-CMA1a	TTACTTAA	882	part of a light responsive element from <i>Daucus carota</i>	12
STRESS RESPONSIVE SEQUENCES				
ARE	AAACCA	1574	<i>cis</i> -acting regulatory element essential for the anaerobic induction from <i>Zea mays</i>	13

promNtNRAMP3

IDE2

+ AGAATCTGCG AGCATCTCAA AGGAATCTAT GGAATGCTCG GGAAGAAGCG AATAC **CACGT CA**AGGCTCCC 70

+ CTCGTGAGTG TCTCCTCGAA TTTCTTCAGC AAAACAGATT CAATCTCGTG GGGAGCTAAA ATCGTTCCCC 140

+ TTCACCACCG TTGCATAGGT GATAATATGT TTATGAGAGT CCGAAGTCCC TCAAGATTGT GTGTATTGTT 210

+ AACACCATTA TTTGCCATTT TCTGTAATTT TTGCTAAGAA CATATAATCA AAACATGTTA GTAAAAAAGA 280

+ CAAGGATCAA CTTAATTACA CGATTTTCTA GGCCCCACGA TGGACGCCAA ACTATTTGCC CGTAAAACGA 350

+ TACAGTTGAA TTATACGTA CTTTCTAGAC AAGTGA**ATTA** **AT**TCGATCCT GAAATAATAA AAGAATTGAA 420

+ TAAATATGCA ATATTTAGCC TTAATATGAA AATGAAATAG CAGAAACCGT AGTTTCGGGA GCAGGGCTTC 490

+ CGGGCACAAAC ATCGATGAAA TCAAAAGGCG AGAGGATAAA GTTGATAAAA GCTTTTAATA TATTGTAGCG 560

+ TAAGTATGCT AGAAAATGCG TCCCTTTTAC AATGATAACA GAGCTCATT TTTATAGTTG CATCTAGGGA 630

+ ATAAGGTCCT AGGATTATGT CCTTCTTTAA TATGAGGGCT ATTAAAGAAT GTGTAACGGC AGGCATGAAT 700

+ GATAAATTTT TTGTAACGGC **CAAT**GTACTA AATACTGTAG AATATTCTTC ATTAAATGCT ATAGTGACAG 770

+ GCATTTATTG CATATTTATG ATCATTATTC TTTTTCGGTA ATAAATGAAA CAGTTGTCTT CGGATTTAGC 840

+ TATCCTCTGC CTTCGGCTTC **ACGTG**TAACT CTCTTATACA **ATTACTTAAC** ATACCATATT TTACCCTATA 910

MRE1

+ CATATAGATT CATAAAAAAT ATAAAACACA GAAAACTTT ACTGCATT **TG CACC**AAAAAG GTAATGGGAA 980

+ GGATCAAAGT ATTTAATTTT TTGTGTTAAT TTGGAATATC CGTTATTTAT TTTTCAAAGA AAAAATT**CAA** 1050

+ **AT**ATTCAAAA TTACAGTAAA ATAAATATAA AACCTTCAA ATAATTATCG TCCAAGAGAT **AGG**AAAAAGT 1120

+ GAAAAACAAT TTAGTCTTAG TTTACAATG TCAGACTCTC **CCTGTTCTTC** CCCTCATCAC TAAATAAGAA 1190

+ AAAGTCAATC GATCTTTAT GCTTCTATCT **CAAA**TAAGA AAAATTTCAA TTATATCTAT TTTAAGCGGA 1260

+ CTTATTTTCA **TCAA**TAACC AGAAAAAAGA TTTTCAAAA CTAGGGGTTG TATCCTTTAC CGCTTACTAA 1330

+ ACTAAGAAGA CATTGATAAA TCCAAGATTA TAAATATAAC ATCGATAAAT AAACATAATT **CCAATTATAG** 1400

+ TTGCTATACT TTTGGCTTTA CATTGATTTT TATTTTTTAG CTAAATTCAT ACTCATTCA AAGTGAAAA 1470

+ AAGGAAAAAC AAAATAAAG GAGAAAGGAA ATTGGGGTGC **AAAT**AGAAAG CGGTAGCGTA ATGGTAGTTT 1540

+ TCAAAAATCA TTCGTCTTAT CGTCTGAAAA GAAAAACCAG AAAAAGATCC AAAGCCTCCA CTATAAACA 1610

+ ACATCACTTC GCACAAACCG AACTCACCTG TACCTTTTAA CCGCTAATAT CCTCTTTACA GATTTCTCTT 1680

+ CTA

References:

1. Li, Y., et al., *Spatial-temporal analysis of zinc homeostasis reveals the response mechanisms to acute zinc deficiency in Sorghum bicolor*. New Phytologist, 2013. 200(4): p. 1102-1115.
2. Ogo, Y., Kobayashi, T., Itai, R.N., Nakanishi, H., Kakei, Y., Takahashi, M., Toki, S., Mori, S., Nishizawa, N.K. 2008. *A Novel NAC Transcription Factor, IDEF2, That Recognizes the Iron Deficiency-responsive Element 2 Regulates the Genes Involved in Iron Homeostasis in Plants*. J. Biol. Chem. 283, 13407-13417
3. Basehoar, A.D., Zanton, S., Pugh, B. 2004. *Identification and Distinct Regulation of Yeast TATA Box-Containing Genes*. Cell, 116(5):699-709.
4. Frangeul, L., Glaser, P., Rusniok, C., Buchrieser, C., Duchaud, E., Dehoux, P., Kunst, F. 2004. *CAAT-Box, contigs-Assembly and Annotation Tool-Box for genome sequencing projects*. Bioinformatics, 20(5):790-797
5. Shen, Q., Ho, T.H. 1995. *Functional dissection of an abscisic acid (ABA)-inducible gene reveals two independent ABA-responsive complexes each containing a G-box and a novel cis-acting element*. The Plant Cell. 7, 295-307.
6. Fink, J.S., et al., *The CGTCA sequence motif is essential for biological activity of the vasoactive intestinal peptide gene cAMP-regulated enhancer*. Proceedings of the National Academy of Sciences, 1988. 85(18): p. 6662-6666.
7. Hiratsuka, K., Chua, N.H. 1997. *Light regulated transcription in higher plants*. J. Plant. Res. 110(1), 131-139.
8. Weisshaar, B., Block, A., Armstrong, G.A., Herrmann, A., Schulze-Lefert, P., Hahlbrock, K. 1991. *Regulatory elements required for light-mediated expression of the Petroselinum crispum chalcone synthase gene*. Symp. Soc. Exp. Biol. 45,191-210
9. Sibénil, Y., P. Doireau, and P. Gantet, *Plant bZIP G-box binding factors. Modular structure and activation mechanisms*. European Journal of Biochemistry, 2001. 268(22): p. 5655-5666.
10. Chawla, R. and D.A. DeMason, *Molecular expression of PsPIN1, a putative auxin efflux carrier gene from pea (Pisum sativum L.)*. Plant growth regulation, 2004. 44(1): p. 1-14.
11. Lafyatis, R., et al., *Sequence specific protein binding to and activation of the TGF- β 3 promoter through a repeated TCCC motif*. Nucleic acids research, 1991. 19(23): p. 6419-6425.
12. Zhu, M., et al., *Characterization of the promoter of Artemisia annua amorpho-4, 11-diene synthase (ADS) gene using homologous and heterologous expression as well as deletion analysis*. Plant molecular biology reporter, 2014. 32(2): p. 406-418.
13. Paul, A.-L. and R.J. Ferl, *In vivo footprinting reveals unique cis-elements and different modes of hypoxic induction in maize Adh1 and Adh2*. The Plant Cell, 1991. 3(2): p. 159-168.