

Figure S1. Preprocessing of the scRNA-seq Datasets for the Integrated Analyses, Related to Figure 1.

(A) scRNA-seq data of the *KIC* PDA by Dominguez et al. were reprocessed. Identities of the cell clusters were matched to the previously reported cluster names based on the expression of genes mentioned in Dominguez et al. including clusters 0, 1, 2, 3, 4, 8, proliferating fibroblasts (Prolif.FB), normal mesothelial cells (Meso), fEMT tumor cells (fEMT), pEMT tumor cells (pEMT), myeloid cells, acinar cells and endothelial cells (EC).

(B) The heatmap displaying the top marker genes for each cell cluster of the reprocessed data from Dominguez et al.

(C) scRNA-seq data of the *KPC* PDA by Elyada et al. were reprocessed. Identities of the cell clusters were matched to the previously reported cluster names based on the expression of genes mentioned in Elyada et al. including iCAFs, apCAFs and myCAFs.

(D) The heatmap displaying the top marker genes for each cell cluster of the reprocessed data from Elyada et al.

(E) Three scRNA-seq datasets of fibroblasts in PDA (Hosein et al., Dominguez et al. and Elyada et al.) were integrated. Graph-based clustering of cells with UMAP was performed with the integrated data and 11 clusters of fibroblasts were identified (Figure 1G). The top marker genes for each cell cluster of the integrated data were shown as the heatmap.

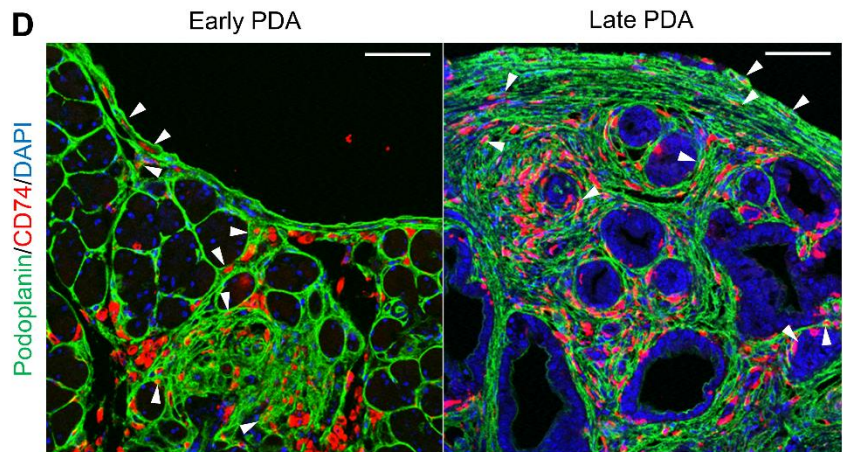
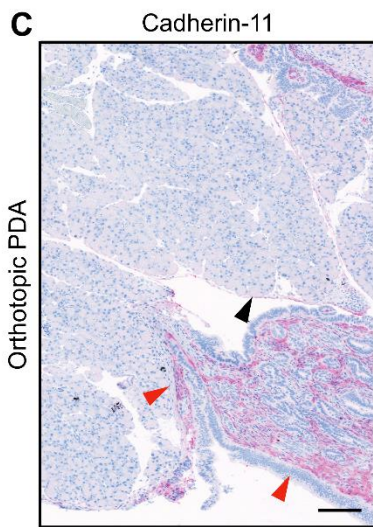
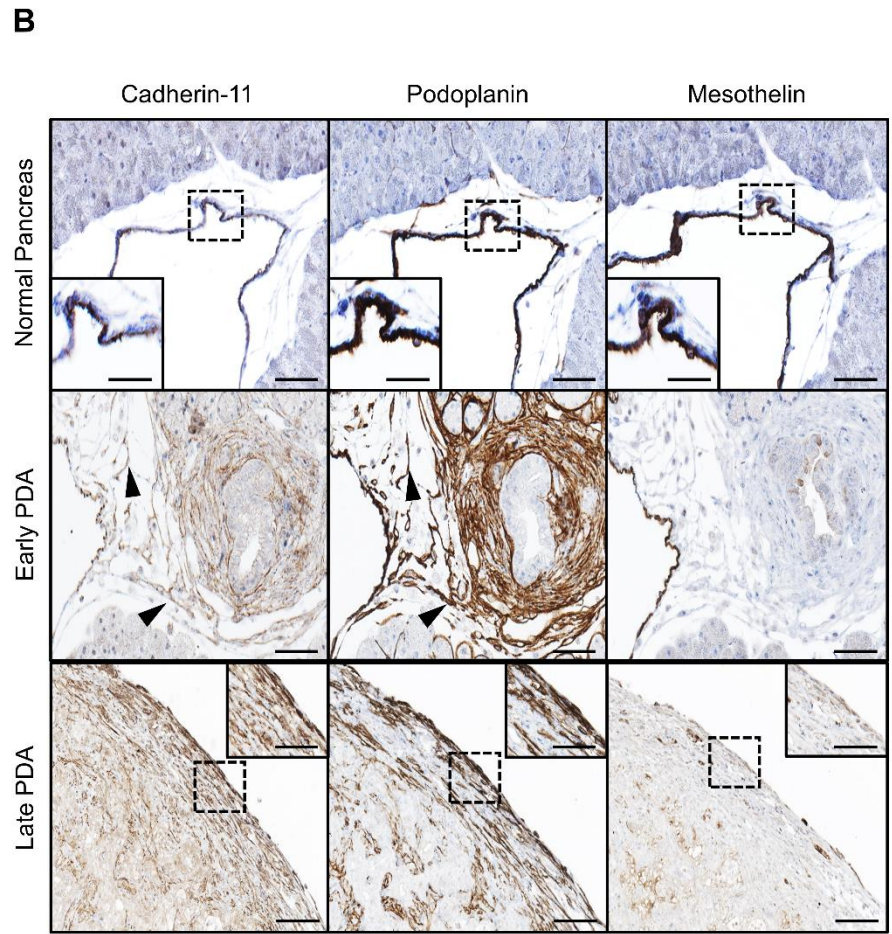
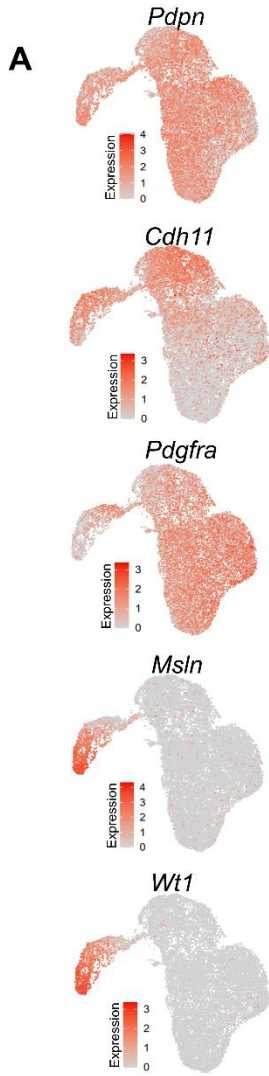


Figure S2. Mesothelial Cells Form a Lining of Normal Pancreas and Contribute to the Peripheral CAF Formation in PDA, Related to Figure 2.

(A) UMAP plots of *Pdpr*, *Cdh11*, *Pdgfra*, *Msln* and *Wt1* from the integrated fibroblast data. Color keys indicate log normalized expression.

(B) IHC staining of mesothelium in normal pancreata, early *KPfc* tumors (40-day-old) and late *KPfc* tumors (60-day-old) by cadherin-11, podoplanin and mesothelin. Arrows, expanding mesothelial cells. Original scale bar 50 μ m. Inset scale bar 12.5 μ m.

(C) The primary mouse PDA cell line BMFA3 derived from *KPfc* tumor was orthotopically injected into the pancreata of C57BL/6 mice. Mouse pancreata were harvested one week after the implantation and subjected to IHC staining for cadherin-11. The cadherin-11⁺ mesothelium lining the uninvolved normal pancreas tissues (black arrow). The cadherin-11⁺ mesothelium adjacent to the tumor lesions (red arrows). Scale bar 200 μ m.

(D) Multiple IHC staining of podoplanin (green), CD74 (red) and DAPI (blue) in early *KPfc* tumors (40-day-old) and late *KPfc* tumors (60-day-old). Arrows, podoplanin⁺CD74⁺ mesothelial cells or apCAFs. Scale bar 50 μ m.

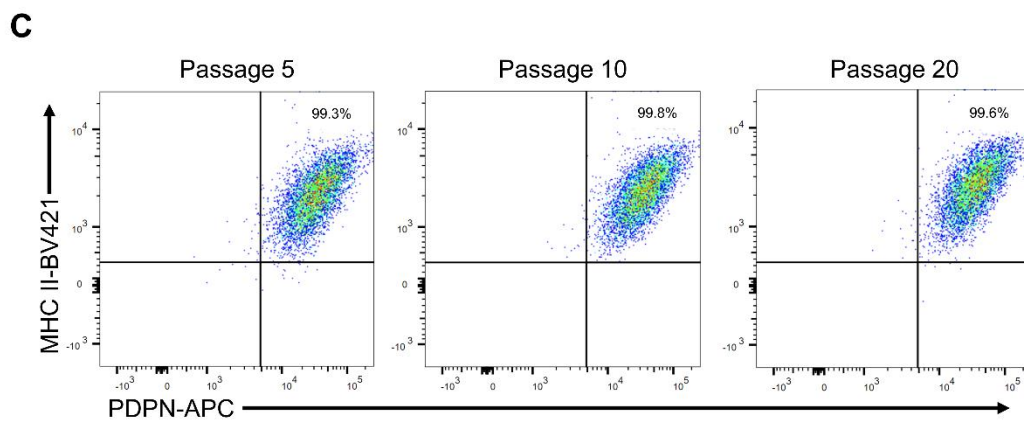
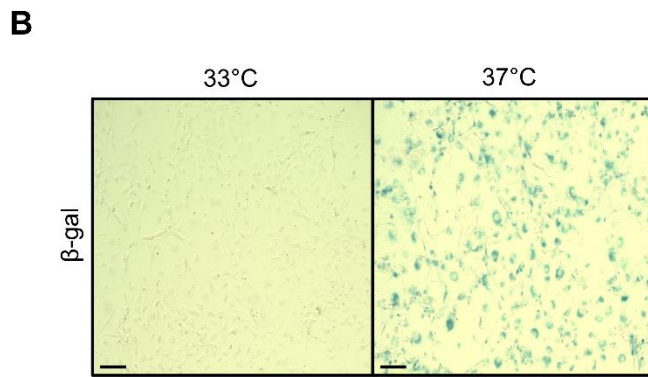
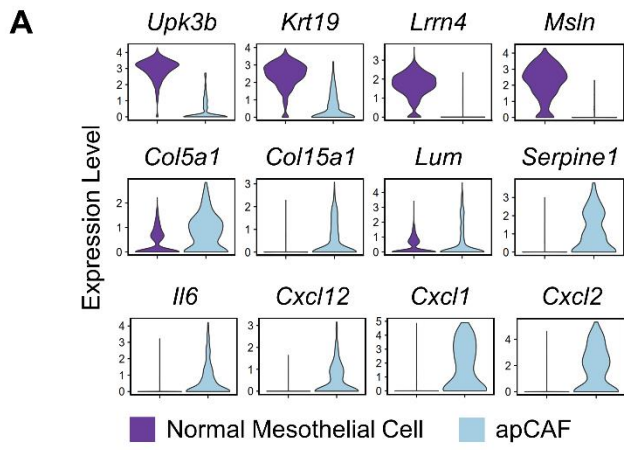


Figure S3. Gene Signature of Normal Mesothelial Cells and apCAFs and Characterization of PanMeso Cells, Related to Figures 2 and 4.

(A) Violin plots showing the mesothelial genes (*Upk3b*, *Krt19*, *Lrrn4*, *Msln*) and fibroblastic genes (*Col5a1*, *Col15a1*, *Lum*, *Serpine1*, *Il6*, *Cxcl12*, *Cxcl1*, *Cxcl2*) in normal mesothelial cells and apCAFs from the integrated data (Figure 1G). The width of the violin plots represents frequency of cells in each region. Values of Y axis indicate log normalized expression of genes.

(B) PanMeso cells were cultured at 33 °C or 37 °C for seven days and subjected to staining for β -Galactosidase to examine cell senescence. Scale bar 50 μ m.

(C) PanMeso cells were cultured at 33 °C for 5, 10 and 20 passages. Cells from different passages were subjected to flow cytometry analysis for podoplanin and MHC II.

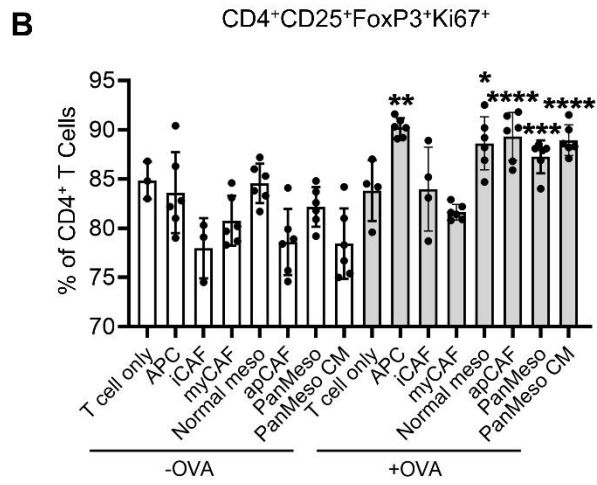
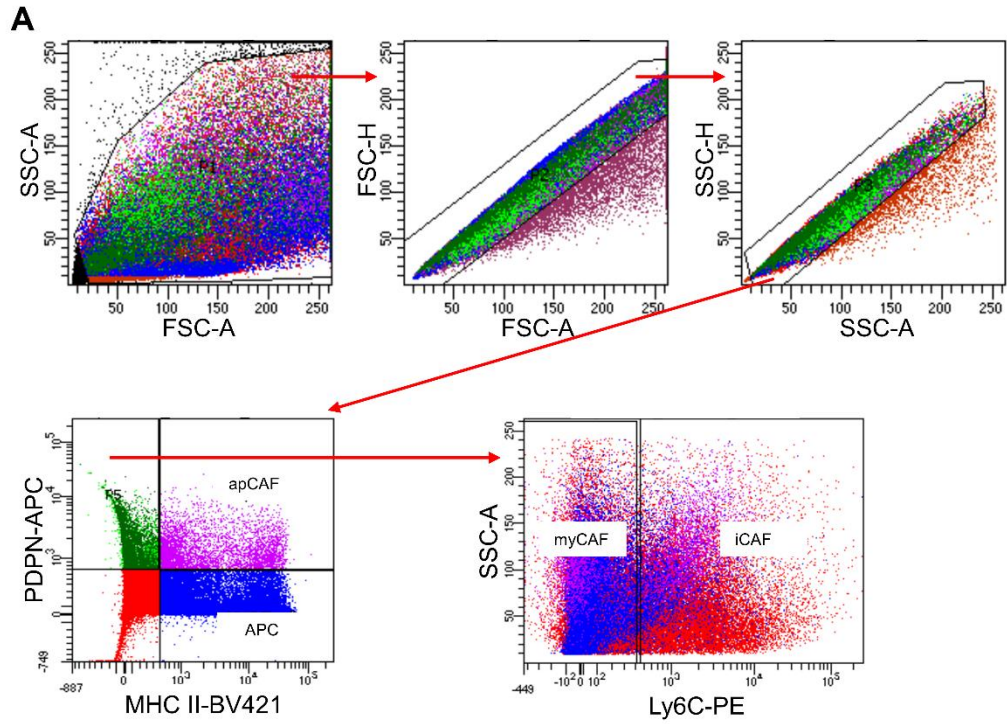


Figure S4. apCAFs Induce Treg Expansion, Related to Figure 5.

(A) Late stage *KPfc* tumors were digested into cell suspension. APCs (podoplanin⁻MHC II⁺), apCAFs (podoplanin⁺MHC II⁺), iCAFs (podoplanin⁺MHC II⁻Ly6C⁺), myCAFs (podoplanin⁺MHC II⁻Ly6C⁻) from *KPfc* tumors were sorted by FACS.

(B) CD4⁺ T cells after being co-cultured with APCs/iCAFs/myCAFs/normal mesothelial cells/apCAFs were subjected to flow cytometry for the analysis of CD25, FoxP3 and Ki67. CD4⁺CD25⁺FoxP3⁺Ki67⁺ T cells were quantified, n=3-6, data shown as mean ± SD, statistical analysis, t-test, *P<0.05, **P<0.01, ***P<0.001, ****P<0.0001 vs without OVA control.

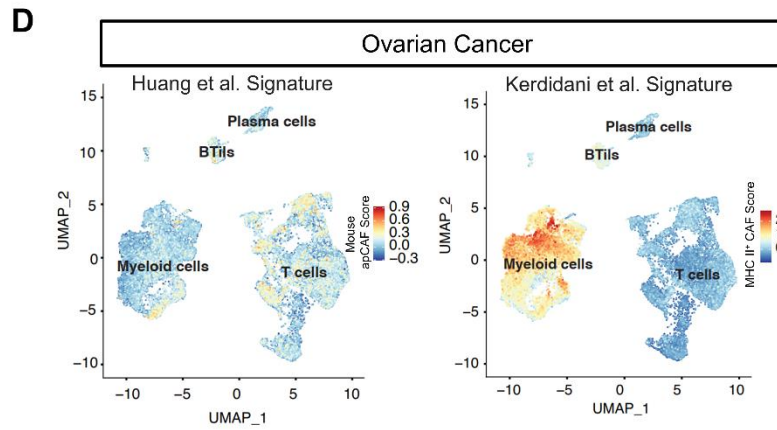
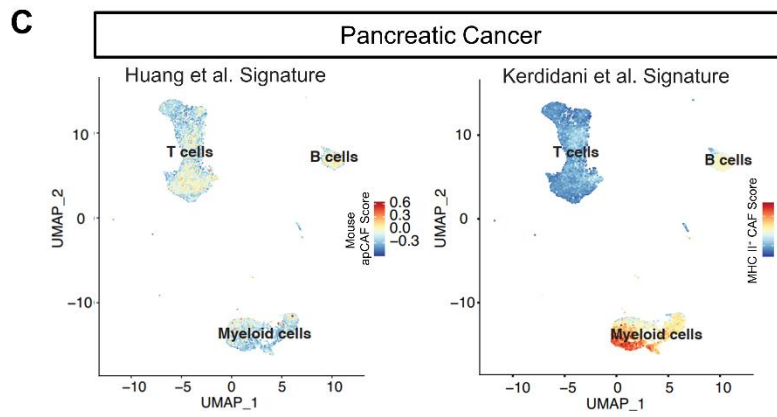
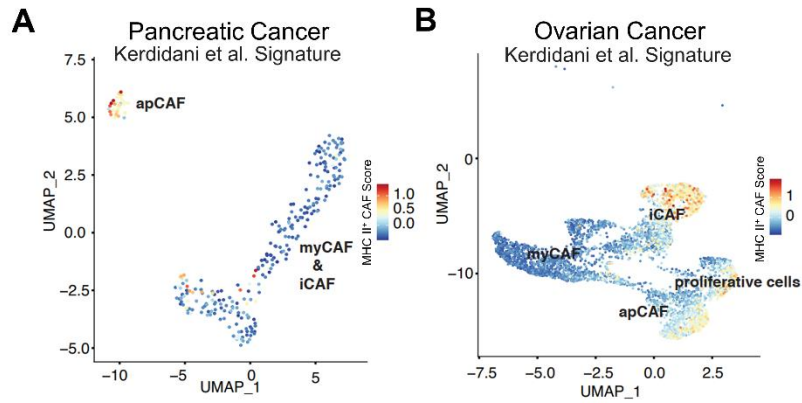


Figure S5. Scoring of apCAF Signatures in Immune Compartments of Human Cancer, Related to Figure 6.

(A-B) The MHC II⁺ CAF gene signature generated from Keridani et al. was scored in the fibroblast compartments of two human cancer scRNA-seq datasets including PDA (A) and ovarian cancer (B).

(C-D) The immune cell subpopulations from the human PDA and ovarian cancer scRNA-seq datasets were first identified based on the cluster annotations mentioned in the original publications. We then scored our mouse apCAF signature and Keridani et al MHC II⁺ CAF signature in the immune cell compartments of the human PDA (C) and ovarian cancer (D) datasets.

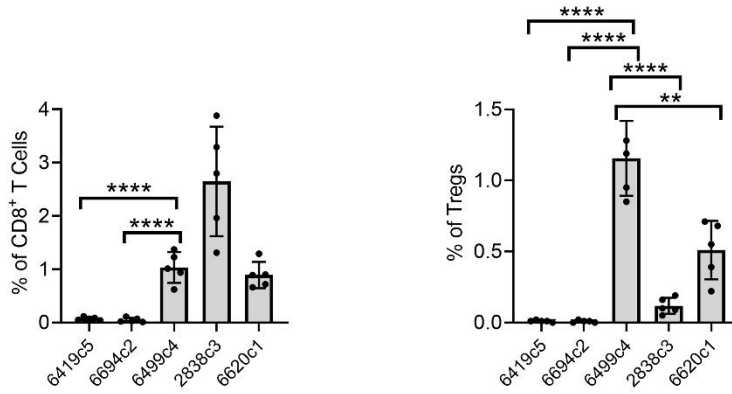
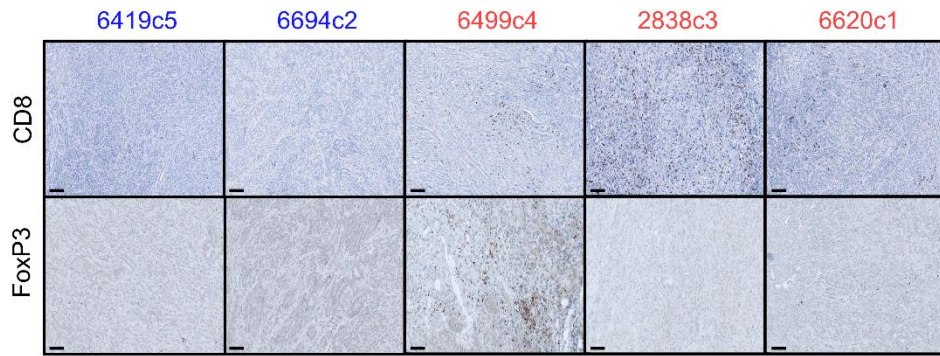


Figure S6. T Cell Characterization of Tumors Derived from the *KPC* Clones, Related to Figure 8.

T cell-inflamed (red) and non-T-cell-inflamed (blue) *KPC* cells were injected orthotopically. Tumors were harvested three weeks after injection and subjected to IHC staining for CD8 and FoxP3. Scale bar 50 μm . CD8⁺ T cells and FoxP3⁺ Tregs were quantified, n=5, data shown as mean \pm SD, statistical analysis, t-test, **P<0.01, ****P<0.0001.

Table S1. qRT-PCR primer sequences, Related to STAR Methods

Mouse Gene Name	Primer Sequence
<i>Msln</i>	Forward Primer: CTTGGGTGGATACCACGTCTG Reverse Primer: CTTCTGTCTTACAGCCATAGCC
<i>Upk3b</i>	Forward Primer: AGACCTGATTGCCTACGTGC Reverse Primer: GGTGTCCTTAGTTGAGACATGCT
<i>Ezr</i>	Forward Primer: CAATCAACGTCCGGGTGAC Reverse Primer: GCCAATCGTCTTTACCACCTGA
<i>Nkain4</i>	Forward Primer: CTCTGGAACGGCAAGTCTTTG Reverse Primer: GTGGCCGGTATTGAATGGTG
<i>Pdpm</i>	Forward Primer: ACCGTGCCAGTGTGTTCTG Reverse Primer: AGCACCTGTGGTTGTTATTTTGT
<i>Cdh11</i>	Forward Primer: CTGGGTCTGGAACCAATTCTTT Reverse Primer: GCCTGAGCCATCAGTGTGTA
<i>Cd74</i>	Forward Primer: AGTGCGACGAGAACGGTAAC Reverse Primer: CGTTGGGGAACACACACCA
<i>H2-Ab1</i>	Forward Primer: AGCCCCATCACTGTGGAGT Reverse Primer: GATGCCGCTCAACATCTTGC
<i>Cdh2</i>	Forward Primer: AGCGCAGTCTTACCGAAGG Reverse Primer: TCGCTGCTTTCATACTGAACTTT
<i>Vim</i>	Forward Primer: CGTCCACACGCACCTACAG Reverse Primer: GGGGGATGAGGAATAGAGGCT
<i>Snai1</i>	Forward Primer: CACACGCTGCCTTGTGTCT Reverse Primer: GGTCAGCAAAGCACGGTT
<i>Snai2</i>	Forward Primer: TGGTCAAGAAACATTTCAACGCC Reverse Primer: GGTGAGGATCTCTGGTTTTGGTA
<i>Zeb1</i>	Forward Primer: GCTGGCAAGACAACGTGAAAG Reverse Primer: GCCTCAGGATAAATGACGGC
<i>Il6</i>	Forward Primer: TAGTCCTTCCTACCCCAATTTCC Reverse Primer: TTGGTCCTTAGCCACTCCTTC
<i>Cxcl1</i>	Forward Primer: CTGGGATTCACCTCAAGAACATC Reverse Primer: CAGGGTCAAGGCAAGCCTC
<i>Pdgfrb</i>	Forward Primer: TTCCAGGAGTGATACCAGCTT Reverse Primer: AGGGGGCGTGATGACTAGG
<i>Tagln2</i>	Forward Primer: CCTGGCCGTGAGAACTTCC Reverse Primer: GTCCGTGGTGTTAATGCCATAG
<i>Col1a1</i>	Forward Primer: GTCCTCTTAGGGGCCACT Reverse Primer: CCACGTCTCACCATTGGGG
<i>Col12a1</i>	Forward Primer: AAGTTGACCCACCTTCCGAC Reverse Primer: GGTCCACTGTTATTCTGTAACCC
<i>Tgfb1</i>	Forward Primer: CTCCCGTGGCTTCTAGTGC Reverse Primer: GCCTTAGTTTTGACAGGATCTG
<i>Il1a</i>	Forward Primer: CGAAGACTACAGTTCTGCCATT Reverse Primer: GACGTTTCAGAGGTTCTCAGAG
<i>Cxcl12</i>	Forward Primer: TGCATCAGTGACGGTAAACCA Reverse Primer: TTCTTCAGCCGTGCAACAATC
<i>Col1a2</i>	Forward Primer: GTAACTTCGTGCCTAGCAACA Reverse Primer: CCTTTGTCAGAATACTGAGCAGC