

# Supporting Information

for

## **Unconventional Site-Selectivity in Palladium-Catalyzed Cross-Couplings of Dichloroheteroarenes under Ligand- Controlled and Ligand-Free Systems**

*Jacob P. Norman, Nathaniel G. Larson, Emily D. Entz, Sharon R. Neufeldt\**

*Department of Chemistry and Biochemistry, Montana State University  
Bozeman, MT 59717, USA*

\*Correspondence: [sharon.neufeldt@montana.edu](mailto:sharon.neufeldt@montana.edu)

## Table of Contents

<b>I. Experimental Details</b>	
<b>A. General Materials and Methods</b>	S3
<b>B. Synthesis and Characterization of Reagents</b>	
1. Chlorinated Heteroarenes	S4
2. Grignard Reagents	S8
<b>C. Suzuki Cross-Couplings with Pd/IPr</b>	
1. General Procedures	S9
2. Optimization (Table 1)	S10
3. Isolation and Characterization of Cross-Coupled Products (Table 1, Scheme 2)	S12
4. Discussion about Mass Balance	S20
5. Scope Limitations of the Suzuki Coupling	S23
6. Comparison of Results with IPr (C4-Selective) vs. dppf (C2-Selective)	S30
<b>D. Kumada Cross-Couplings with Pd/IPr</b>	
1. General Procedures	S30
2. Optimization	S31
3. Isolation and Characterization of Kumada Products from Scheme 3	S32
4. Discussion about Decomposition of Putative Products <b>S31a</b> and <b>S33a</b>	S33
<b>E. Negishi Cross-Couplings with Pd/IPr</b>	
1. General Procedures	S37
2. Optimization	S38
3. Isolation and Characterization of Negishi Products from Scheme 3	S39
<b>F. Multistep Syntheses using C4-Selective Cross-Coupling with Pd/IPr (Scheme 4)</b>	S40
<b>G. Ligand-Free Suzuki Cross-Couplings</b>	
1. Ligand-Free Control Reaction Based on Previously-Reported Conditions (Scheme 5b)	S42
2. Coupling of <b>1</b> under Fairlamb's Recently Reported Pd/PPh <sub>3</sub> Conditions	S43
3. General Procedures for the Ligand-Free 'Jeffery' Conditions Reported Herein	S43
4. Optimization of the Ligand-Free Conditions (Table 2)	S44
5. Isolation and Characterization of Cross-Coupled Products (Scheme 6)	S45
6. Discussion about Mass Balance	S48
7. Comparison of Ligand-Free Results (C4-Selective) vs. C2-Selective Conditions	S50
<b>II. References</b>	S51
<b>III. NMR Spectra</b>	S52

## I. Experimental Details

### A. General Materials and Methods

NMR spectra were recorded at 298 K on a Bruker DRX 500 MHz (500.233 MHz for  $^1\text{H}$ , 125.795 MHz for  $^{13}\text{C}$ , 470.639 MHz for  $^{19}\text{F}$ ), a Bruker Avance III 600 MHz (600.130 MHz for  $^1\text{H}$  or 150.903 MHz for  $^{13}\text{C}$ ) spectrometer, or a Bruker Ascend 400 MHz (400.130 MHz for  $^1\text{H}$  NMR, 100.613 for  $^{13}\text{C}$ ).  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts are reported in parts per million (ppm) relative to TMS, with the residual solvent peak used as an internal reference [ $^1\text{H}$  NMR:  $\text{CHCl}_3$  (7.26 ppm),  $\text{C}_6\text{D}_5\text{H}$  (7.16 ppm),  $\text{DMSO}-d_5$  (2.50 ppm);  $^{13}\text{C}$  NMR:  $\text{CDCl}_3$  (77.16 ppm),  $^{13}\text{C}$  NMR:  $\text{C}_6\text{D}_6$  (128.06 ppm),  $\text{DMSO}-d_6$  (39.52 ppm)]. Multiplicities are reported as follows: singlet (s), broad singlet (br s) doublet (d), doublet of doublets (dd), doublet of doublets of doublets (ddd), triplet (t), quartet (q), and multiplet (m). GC data were collected using a Shimadzu GC-2010 Plus with a flame ionization detector equipped with a SH-Rxi-5ms capillary column (15 m x 0.25 mm ID x 0.25  $\mu\text{m}$  df). GCMS data were collected with a Shimadzu GC-2030 paired with a Shimadzu GCMS-QP2020 NX and equipped with a SH-Rxi-5ms capillary column (30 m x 0.25 mm ID x 0.25  $\mu\text{m}$  df). LC-MS analyses were performed on either an Agilent 6538 Q-TOF MS or a Bruker micro-TOF MS, both coupled to an Agilent 1290 Infinity UHPLC system. A 50 mm long Eclipse Plus C18 column (Agilent Technologies, Santa Barbara, CA; i.d. 2.1 mm, 1.8  $\mu\text{m}$  particle size) was used for separation. A 6-minute gradient was used at a flow rate of 0.6 mL/min: 0-1 min 95% buffer A (100%  $\text{H}_2\text{O}$  with 0.1% formic acid), followed by a gradient from 1-4 minutes of 5-95% buffer B (100% acetonitrile with 0.1% formic acid), 1 minute 95% buffer B, and returning to 95% buffer A for 1 minute. The mass spectrometers were operated in positive-ion mode with electrospray ionization. All reactions that require heating were conducted in aluminum single-size vial reaction blocks (Chemglass) fitted to an IKA stirring hot plate equipped with a temperature probe.

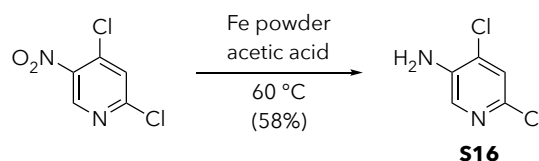
Unless otherwise noted below, all commercially-obtained chemicals were used as received. PEPPSI-SIPr and PEPPSI-IPr were obtained from Sigma Aldrich.  $(\eta^3\text{-1-}^t\text{Bu-indenyl})_2(\mu\text{-Cl})_2\text{Pd}_2$ ,  $(\eta^3\text{-1-}^t\text{Bu-indenyl})\text{Pd}(\text{IPr})(\text{Cl})$ , and  $(\eta^3\text{-1-}^t\text{Bu-indenyl})\text{Pd}(\text{IPent})(\text{Cl})$  were obtained from Umicore. Other  $(\eta^3\text{-1-}^t\text{Bu-indenyl})\text{Pd}(\text{NHC})(\text{Cl})$  precatalysts were prepared from the corresponding *N*-heterocyclic carbene ligands IMes, SIMes, SIMix, or SIPr (each obtained from Strem Chemicals or Sigma Aldrich) according to a literature procedure.<sup>1</sup> Unless otherwise noted, dichloroheteroarenes and arylboronic acid starting materials were obtained from Oakwood Chemical. 4,5-Dichloro-3(2H)-pyridazinone (**S45**)<sup>2</sup> and 3,4,5-trichloropyridazine (**S46**)<sup>3</sup> were prepared according to literature procedures. Potassium fluoride, 1,4-dioxane, benzyl bromide, iodine, potassium *tert*-butoxide, tetrabutylammonium hexafluorophosphate, and palladium (II) chloride were obtained from Acros Organics. Potassium carbonate, methylmagnesium bromide, triphenylphosphine, tri-*tert*-butylphosphine, 4-vinylphenylboronic acid, 2,4-dichloro-3-cyanopyridine, and 2,4-dichloro-5-methylpyridine were obtained from Alfa Aesar. 2,4-dichloro-5-nitropyridine and 3,3'-bromomethyloxetane were obtained from Combi-Blocks. 2,4-Dichloro-1,8-naphthyridine and 3-amino-2,4-dichloropyridine were obtained from Millipore Sigma. 3-Amino-2,4-dichloropyridine was obtained from Synthonix. Sodium carbonate, THF, toluene, and methanol were obtained from Fisher Scientific. Magnesium turnings, *tert*-butyl bromide, cyclopentyl bromide, benzothiophene (thianaphthene), 2-bromo-5-methylpyridine, lithium chloride, tetrabutylammonium bromide and chloride, tris(dibenzylideneacetone)dipalladium(O), diphenylphosphinoferrrocene, tri-*o*-tolylphosphine, and tricyclohexylphosphine were obtained from Oakwood

chemical. Diisobutylaluminum hydride (DIBAL), magnesium bromide diethyl etherate, propylene carbonate, potassium bromide, and *n*-butyllithium were obtained from Sigma-Aldrich. Palladium (II) acetate, trimethyl phosphine, Q-Phos, and CataCXium A were obtained from Strem Chemical. 2,4-Dichloroquinoline, zinc dichloride, and Pd(PPh<sub>3</sub>)<sub>4</sub> were obtained from TCI Chemicals. Benzene was obtained from Beantown Chemical. For the purposes of Kumada or Negishi cross-couplings or for preparation of Grignard reagents, THF was purified on a JC Meyer solvent dispensing system and stored under N<sub>2</sub> prior to use. For the purpose of Suzuki-Miyaura cross-couplings, THF was used as received from Fisher Scientific. For the purpose of Suzuki-Miyaura cross-couplings under ligand-free reaction conditions, DMF was purified on a JC Meyer solvent dispensing system and stored under N<sub>2</sub> prior to use. 1,4-Dioxane required for Pd/dppf-mediated Suzuki-Miyaura cross-couplings was used as received from Acros Organics but kept under N<sub>2</sub> prior to and during use. Grignard reagents were titrated according to a literature procedure.<sup>4</sup>

Deuterated solvents (CDCl<sub>3</sub>, C<sub>6</sub>D<sub>6</sub>, DMSO-*d*<sub>6</sub>) were obtained from Cambridge Isotopes and stored over molecular sieves. Manual flash column chromatography was performed on SiliCycle silica gel 60 (40-63 μm particle size) and thin layer chromatography was performed on SiliCycle TLC plates pre-coated with extra hard silica gel 60 F<sub>254</sub>. Automated flash column chromatography was performed with a Biotage Selekt equipped with Biotage Sfar silica flash cartridges (20 μm particle size; 50 Å pore width) for normal phase separations, or Silica C18 cartridges (30 μm particle size; 100 Å pore width) for reversed phase separations.

## B. Synthesis and Characterization of Reagents

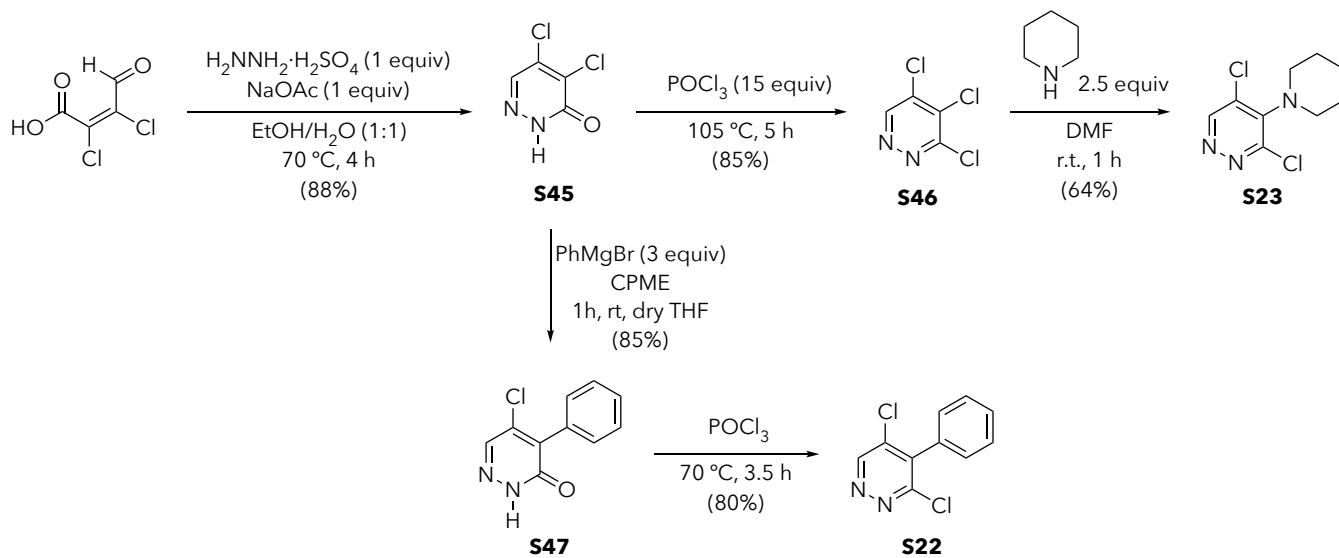
### 1. Chlorinated Heteroarenes



**3-Amino-4,6-dichloropyridine (S16):** Glacial acetic acid (14.2 mL) was heated to 60 °C in a round-bottom flask equipped with a Vigreux reflux condenser and stir bar. Iron powder (4.175 g, 74.8 mmol, 3.5 equiv) and 2,4-dichloro-5-nitropyridine (4.122 g, 21.36 mmol, 1.0 equiv) were added to the flask consecutively while stirring. The mixture was allowed to stir at 60 °C for 2.5 h. Partway through the reaction time, the mixture thickened such that stirring was encumbered, so an additional 8 mL glacial acetic acid was added to the flask. The resulting suspension was vacuum-filtered through celite and the filter cake was washed thoroughly with ethyl acetate. Solvent was removed from the filtrate under reduced pressure, and the resulting solids were taken up in ethyl acetate. The organic solution was washed with saturated aqueous sodium bicarbonate, deionized water (3x), and brine. The organic layer was dried over magnesium sulfate, filtered, and concentrated under reduced pressure. The resulting white solid was dried under vacuum overnight, affording **S16** as a very pale pink solid (2.030 g, 58% yield). <sup>1</sup>H NMR

(500 MHz, CDCl<sub>3</sub>, δ): 7.88 (s, 1H), 7.21 (s, 1H), 4.16 (br s, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, CDCl<sub>3</sub>, δ): 139.7, 139.3, 136.2, 129.7, 123.9

**Scheme S1.** Dichloropyridazines Synthetic Map



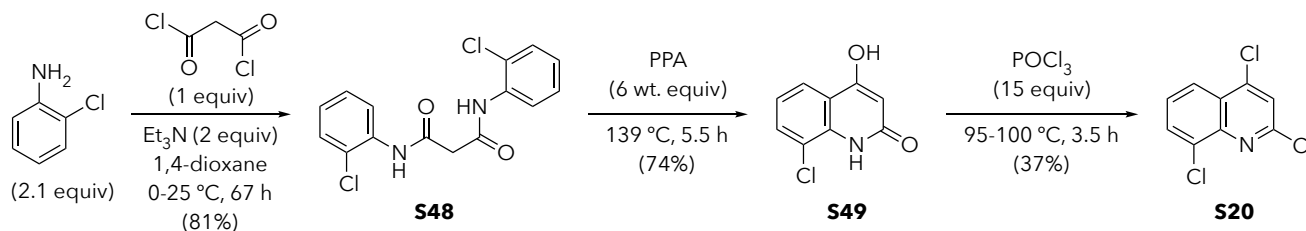
**5-Chloro-4-phenyl-3(2H)-pyridazinone (S47):** Compound **S47** was prepared according to a procedure adapted from the literature.<sup>5</sup> 4,5-Dichloro-3(2H)-pyridazinone (**S45**)<sup>2</sup> (1.2 g, 7.27 mmol, 1.0 equiv) was added to an oven-dried 50 mL Schlenk flask equipped with a stir bar. The flask was fitted with a rubber septum, the septum was secured with copper wire, and the flask was evacuated and backfilled with N<sub>2</sub> (3x). Dry THF (12 mL) was transferred into the flask by cannula. The flask was cooled in an ice bath, and a solution of phenylmagnesium bromide (1.6 M in CPME, 13.6 mL, 21.82 mmol, 3.0 equiv) was added dropwise over 10 min by cannula. The flask was removed from the ice bath and the reaction was stirred at room temperature for 1 h. The flask was returned to the ice bath, opened to air, and quenched by the slow addition of aqueous NH<sub>4</sub>Cl (12 mL; 100 g/L). The resulting mixture was partitioned between deionized water and ethyl acetate. The aqueous layer was extracted with ethyl acetate (2x). The combined organic fractions were washed with brine, dried over magnesium sulfate, filtered, and concentrated under reduced pressure. The resulting solids were triturated in hexanes, filtered, and dried under vacuum to afford **S47** as a white solid (1.277 g, 85% yield). <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>, δ): 13.40 (br s, 1H), 8.03 (s, 1H), 7.47-7.40 (multiple overlapping signals, 5H). Spectral data are consistent with the literature report.<sup>5</sup>

**3,5-Dichloro-4-phenylpyridazine (S22):** Phosphorous(V) oxychloride (3.4 mL) and **S47** (1.266 g, 6.127 mmol, 1.0 equiv) were combined in an oven-dried round-bottom flask equipped with a stir bar. The flask was then fitted with a Vigreux reflux condenser and sealed with a rubber septum fastened with copper wire. An outgassing line, which was assembled with PVC tubing, a needle adapter, and a needle, was routed from the condenser through a mineral oil bubbler, which was in turn routed through an aqueous KOH trap. An in-gas needle was introduced to the condenser, and the system was purged with N<sub>2</sub>. The in-gas line was closed off such that hydrogen chloride evolution could be monitored visually. The reaction mixture was heated to 70 °C and stirred for 3.5 h. Stirring was

continued while the reaction cooled to ambient temperature. The reaction was quenched by slow, dropwise addition of aqueous NaOH (5 mol % in water) while stirring until bubbling ceased. The mixture was diluted with water and filtered affording a brown solid. The wet solid was taken up in benzene, and then concentrated under reduced pressure; this process was repeated once. The resulting solids were dried under vacuum overnight to afford **S22** as a brown solid (1.109 g, 80% yield).  $^1\text{H NMR}$  (500 MHz, DMSO- $d_6$ ,  $\delta$ ): 9.54 (s, 1H), 7.58-7.51 (multiple overlapping signals, 3H), 7.46-7.43 (m, 2H);  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz, DMSO- $d_6$ ,  $\delta$ ): 155.6, 151.6, 138.8, 137.9, 131.7, 129.7, 128.71, 128.68. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{10}\text{H}_6\text{Cl}_2\text{N}_2$  223.9908; Found 223.9896.

**3,5-Dichloro-4-(1-piperidiny)-pyridazine (S23)**: 3,4,5-Trichloropyridazine (**S46**)<sup>3</sup> (550.3 mg, 3.0 mmol, 1.0 equiv) was combined with DMF (9.38 mL) and a stir bar in a 5-dram vial. The vial was fitted with a septum and placed under an atmosphere of  $\text{N}_2$  gas. Piperidine (744  $\mu\text{L}$ , 7.5 mmol, 2.5 equiv) was added dropwise through the septum via a 100  $\mu\text{L}$  syringe to the stirring solution, resulting in rapid evolution of HCl gas and increased turbidity. The reaction was stirred at ambient temperature for 1 h. DMF was removed under vacuum, and the residue was purified by flash column chromatography on silica gel in 10% acetone in hexanes ( $R_f = 0.22$ ). The combined fractions were dried under vacuum to afford **S23** as a white solid with minor brown discoloration (445 mg, 64% yield).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.68 (s, 1H), 3.36-3.34 (m, 4H), 1.77-1.64 (multiple overlapping signals, 6H);  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 155.5, 148.0, 143.1, 123.9, 50.6, 25.6, 23.7. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_9\text{H}_{11}\text{Cl}_2\text{N}_3$  231.0330; Found 231.0326.

**Scheme S2. Synthesis of 2,4,8-Trichloroquinoline (S20)**

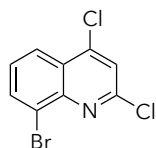


**$N^1,N^3$ -Bis(2-chlorophenyl)-propanediamide (S48)**: Compound **S48** was prepared according to a modified literature procedure.<sup>6</sup> Malonyl dichloride was purified by simple vacuum distillation prior to use; triethylamine was distilled from calcium hydride, sparged with  $\text{N}_2$ , and stored over 4 Å molecular sieves prior to use. 2-Chloroaniline (2.679 g, 21 mmol, 2.1 equiv) was weighed into a 100-mL round bottom flask, followed by dry 1,4-dioxane (24 mL) and a stir bar. The flask was fitted with a rubber septum and the septum was secured with copper wire. An ingas needle and smaller diameter outgassing needle were introduced through the septum, and the contents of the flask were sparged with  $\text{N}_2$  for 15 min. Dry triethylamine (2 equiv, 20 mmol, 2.8 mL) was added through the septum via 1-mL syringe, and the sealed flask was cooled in an ice bath. Under  $\text{N}_2$ , a solution of malonyl dichloride (973  $\mu\text{L}$ , 10 mmol, 1 equiv) in dry 1,4-dioxane (28.5 mL) was added dropwise to the cooled stirring solution through the septum via a 10-mL syringe over 20 min. The solution became bright yellow and evolution of HCl gas was observed. The

flask was removed from the ice bath and stirred at ambient temperature for 67 h. The flask was opened to air and the bright yellow mixture was poured into a 500-mL beaker. While stirring, aqueous HCl (2 M, 36 mL) was added slowly to the mixture. The mixture was stirred for an additional 30 min, resulting in precipitation of yellow solids. The solids were collected by vacuum filtration, washed with water, and dried under vacuum, affording **S48** as a yellow powder (2.6132 g, 81% yield). A minor dioxane impurity (3 mol%, 0.8% by mass) was detected by <sup>1</sup>H-NMR analysis. <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>, δ): 10.04 (s, 2H), 7.90 (dd, *J* = 8.3, 1.3 Hz, 2H), 7.51 (dd, *J* = 8.0, 1.3 Hz, 2H), 7.43 (ddd, *J* = 8.7, 8.0, 1.3, 2H), 7.19 (ddd, *J* = 8.7, 8.0, 1.3, 2H), 3.75 (s, 2H). Spectral data are consistent with the literature.<sup>6</sup>

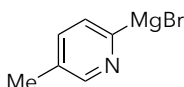
**8-Chloro-4-hydroxy-2(1H)-quinolinone (S49):** *N*<sup>1</sup>, *N*<sup>3</sup>-Bis(2-chlorophenyl)-propanediamide (**S48**) (1.661 g, 5.14 mmol, 1.0 equiv) and polyphosphoric acid (9.965 g, 6.0 wt. equiv) were measured into a round-bottom flask equipped with stir bar. The flask was fitted with a water-cooled coil-type jacketed condenser fitted with a rubber septum secured by copper wire. The system was placed under an atmosphere of N<sub>2</sub> gas, and the mixture was heated to 139 °C for 5.5 h. The system was allowed to cool to handling temperature and the mixture was poured into ice water, inducing precipitation and hardening of the tar-like mixture. The transfer was completed with alternating washes of ethyl acetate and water. The resulting slurry was stirred overnight to break up the tar, resulting in a suspension of precipitated fine solids. The suspension was decanted from residual tar and the precipitate was collected by filtration, washed with water, and dried under vacuum to afford **S49** as a pale brown powder (737 mg, 74% yield). <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>, δ): 11.61 (br s, 1H), 10.37 (br s, 1H), 7.78 (dd, *J* = 7.9, 1.3 Hz, 1H), 7.66 (dd, *J* = 7.9, 1.3 Hz, 1H), 7.17 (t, *J* = 7.9 Hz, 1H), 5.81 (s, 1H). Spectral data are consistent with the literature.<sup>7</sup>

**2,4,8-Trichloroquinoline (S20):** 8-Chloro-4-hydroxy-2(1H)-quinolinone (**S49**) (737.4 mg, 3.77 mmol, 1.0 equiv) and phosphorous(V) oxychloride (POCl<sub>3</sub>) (5.3 mL, 56.6 mmol, 15.0 equiv) were combined in a round-bottom flask equipped with a stir bar. The flask was fitted with a water-cooled coil-type jacketed condenser fitted with a rubber septum secured by copper wire. The system was placed under an atmosphere of N<sub>2</sub> and heated at 95-100 °C for 3.5 h. Evolving hydrogen chloride gas was routed through a mineral oil bubbler and then through an aqueous KOH trap using PVC tubing. Excess POCl<sub>3</sub> was removed by short-path vacuum distillation. Residual POCl<sub>3</sub> was quenched by dropwise addition of aqueous NaOH (5 mol %) then diluted with deionized water, inducing precipitation. The dark brown precipitate was filtered, washed with water, triturated in hexanes by sonication, and vacuum filtered through celite. The filtrate was concentrated under reduced pressure, but excessive residual water was observed by <sup>1</sup>H NMR. The solids were redissolved in chloroform, and the solution was washed with brine (2x), passed through a plug of magnesium sulfate, and concentrated under reduced pressure. The resulting solids were dried under vacuum overnight to afford **S20** as a pale yellow solid (324 mg, 37% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, δ): 8.13 (dd, *J* = 8.5, 1.3 Hz, 1H), 7.90 (dd, *J* = 7.6, 1.3 Hz, 1H), 7.58 (s, 1H), 7.57 (dd, *J* = 8.5, 7.6 Hz, 1H). <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, CDCl<sub>3</sub>, δ): 151.0, 144.9, 144.8, 133.4, 131.8, 127.8, 126.7, 123.4, 123.2. HRMS (TOF MS ESI+) *m/z*: [M]<sup>+</sup> Calcd for C<sub>9</sub>H<sub>4</sub>Cl<sub>3</sub>N 230.9409; Found 230.9471.

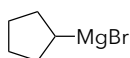


**8-Bromo-2,4-dichloroquinoline (28):** Compound **28** was prepared according to a modified literature procedure.<sup>8</sup> 2-Bromoaniline (230  $\mu$ L, 1 mmol, 1.0 equiv), malonic acid (208 mg, 2 mmol, 2 equiv) and phosphorous(V) oxychloride ( $\text{POCl}_3$ ) (2.0 mL) were combined in a 10 mL oven-dried Schlenk flask equipped with a stir bar. The flask was fitted with a water-cooled coil-type jacketed condenser equipped with a rubber septum secured by copper wire. The system was placed under an atmosphere of  $\text{N}_2$  and heated at 100  $^\circ\text{C}$  for 6 h. Evolving hydrogen chloride gas was routed through a mineral oil bubbler and then through an aqueous KOH trap using PVC tubing. The reaction mixture was allowed to cool to room temperature and was subsequently poured onto crushed ice with stirring. The resulting mixture was extracted with excess dichloromethane and washed with sat.  $\text{NaHCO}_3$  (aq) until the aqueous layer was slightly basic (pH 8). The layers were separated, and the organic layer was dried over anhydrous  $\text{MgSO}_4$ , filtered, and concentrated under reduced pressure. The crude residue was purified by flash column chromatography ( $R_f = 0.58$  in 98:2 hexanes:EtOAc).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$   $\delta$ ): 8.18 (dd,  $J = 8.4, 1.2$  Hz, 1H), 8.12 (dd,  $J = 7.6, 1.2$  Hz, 1H), 7.58 (s, 1H), 7.50 (dd,  $J = 8.4, 7.6$  Hz, 1H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$   $\delta$ ): 150.9, 145.5, 144.7, 135.3, 128.1, 126.5, 124.13, 124.05, 123.0. HRMS (ESI<sup>+</sup>)  $m/z$ :  $[\text{M}]^+$ : Calcd for  $\text{C}_9\text{H}_4\text{Cl}_2\text{BrN}$  274.8904; Found 274.8904.

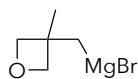
## 2. Grignard Reagents



**5-Methyl-2-pyridylmagnesium bromide (S50):** Compound **S50** was prepared according to a modified literature procedure.<sup>9</sup> To an oven-dried 25 mL Schlenk flask equipped with a stir bar, under  $\text{N}_2$ , was added freshly crushed magnesium turnings (80.2 mg, 3.3 mmol, 1.1 equiv), dry degassed THF (6.0 mL), dry lithium chloride (127.2 mg, 3 mmol, 1.0 equiv), and diisobutylaluminum hydride (30  $\mu$ L of a 1.0  $M$  solution in THF, 0.03 mmol, 0.01 equiv). The reaction mixture was stirred for 5 min, followed by the dropwise addition of a solution of 2-bromo-5-methylpyridine (516.1 mg, 3.0 mmol, 1.0 equiv) in dry degassed THF (3.0 mL). The reaction was stirred at 23  $^\circ\text{C}$  for 1 h, resulting in a dark red solution that was used without further purification.

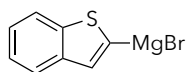


**Cyclopentylmagnesium bromide (S51):** To an oven-dried 25 mL Schlenk flask equipped with a stir bar, under  $\text{N}_2$ , was added freshly crushed magnesium turnings (53.5 mg, 2.2 mmol, 1.1 equiv) and dry degassed THF (6.7 mL). Cyclopentyl bromide (475  $\mu$ L, 4 mmol, 1.0 equiv) was then added dropwise to the reaction mixture. The reaction mixture was heated to reflux for 3 h, resulting in a light brown solution that was used without further purification.

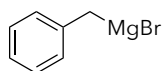


**3,3'-Methyloxetanylmagnesium bromide (S52):** To an oven-dried 25 mL Schlenk flask equipped with a stir bar, under  $\text{N}_2$ , was added freshly crushed magnesium turnings (106.9 mg, 4.4 mmol, 1.1 equiv), dry degassed THF (6.0 mL), and diisobutylaluminum hydride (40  $\mu$ L of a 1.0  $M$  solution in THF, 0.04 mmol, 0.01 equiv). The reaction mixture was stirred for 5 min, followed by the dropwise addition of a solution of 3-bromomethyl-3-methyloxetane (462  $\mu$ L, 4.0 mmol, 1.0 equiv) in dry degassed THF (7 mL). The reaction mixture was heated to reflux for 2.5 h resulting in a colorless solution containing a white precipitate. This mixture was used without further purification.





**2-Benzothiophenylmagnesium bromide (S53):** Compound **S53** was prepared according to a modified literature procedure.<sup>7</sup> To an oven-dried 25 mL Schlenk flask ("Flask A") equipped with a stir bar, under N<sub>2</sub>, was added benzothiophene (1.073 g, 8 mmol, 1 equiv) and dry degassed THF (13 mL). The resulting solution was cooled to -84 °C. *n*-BuLi (3.2 mL of a 2.5 M solution in hexanes, 8 mmol, 1.0 equiv) was added dropwise, and the mixture was stirred for 2 h while warming to 0 °C. In a separate 50 mL Schlenk flask ("Flask B") equipped with a stir bar, under nitrogen, was added magnesium bromide ethyl etherate (2.066 g, 3.0 mmol, 1.0 equiv) and dry degassed THF (14 mL). The cold contents of Flask A were added to the slurry in Flask B. The reaction mixture was stirred at 23 °C until all solids were dissolved, resulting in a rusty colored solution that was used without further purification.



**Benzylmagnesium bromide (S54):** To an oven-dried 25 mL Schlenk flask equipped with a stir bar, under N<sub>2</sub>, was added freshly crushed magnesium turnings (106.9 mg, 4.4 mmol, 1.1 equiv), dry degassed THF (6.0 mL), and diisobutylaluminum hydride (40 μL of 1.0 M solution in THF, 0.04 mmol, 0.01 equiv). The resulting mixture was stirred for 5 min, followed by the dropwise addition of benzyl bromide (475 μL, 4 mmol, 1.0 equiv) in dry degassed THF (7 mL). The reaction mixture was heated to reflux for 3 h, resulting in a clear solution containing a small quantity of white crystals. This mixture was used without further purification.

## C. Suzuki Cross-Couplings with Pd/IPr

### 1. General Procedures

**GC-Scale Reactions.** The specified solids required in the Suzuki-Miyaura reactions were added to a 1-dram reaction vial in order of increasing mass: palladium catalyst (palladium source and free ligand) or precatalyst, the specified dihalopyridine substrate if solid (0.08 mmol, 1 equiv), arylboronic acid (0.08 mmol, 1.0 equiv), potassium carbonate or cesium carbonate, and then a stir bar. Liquid reagents were pre-measured by syringe and added in quick succession: benzene, THF, or 1,4-dioxane (0.32 mL, 0.25 M) via 1-mL syringe, followed by N<sub>2</sub>-sparged deionized water via 50-μL syringe. Note: dihalopyridine substrates were added last if liquid, via microliter syringe. A septum cap equipped with an N<sub>2</sub>-ingass and outgassing needle was fastened to the 1-dram reaction vial and the headspace was sparged for 30-45 seconds. With continuous sparging, the vial was unscrewed from the septum cap and lowered while the cap was replaced with a PTFE-lined cap. The reaction was stirred vigorously at the specified temperature for the specified duration.

**Scaled-Up Reactions for Product Isolation.** Precatalyst ( $\eta^3$ -1-*t*Bu-indenyl)Pd(IPr)Cl (8.4 mg, 0.012 mmol, 3.0 mol%) was weighed into a 1-dram vial followed by solid substrate, if applicable (2,4-dichloropyridine, quinoline, or 3,5-dichloropyridazine, 0.4 mmol, 1 equiv), arylboronic acid (0.4 mmol, 1.0 equiv), potassium carbonate, and a stir bar. Liquid reagents were pre-measured by syringe and added in quick succession: benzene or THF (1.6 mL, 0.25 M) via a 1-mL syringe followed by N<sub>2</sub>-sparged deionized water via a 100-μL or 250-μL syringe. In cases where the substrate was a liquid, the substrate was added last via microliter syringe (0.4 mmol, 1 equiv). A septum cap equipped with an N<sub>2</sub>-ingass and outgassing needle was fastened to the 1-dram reaction vial and the mixture was

sparged for 30-45 seconds. With continuous sparging, the vial was unscrewed from the septum cap and lowered while the cap was replaced with a PTFE-lined cap. The reaction was stirred vigorously at the specified temperature for the specified duration. The cap was removed, the reaction mixture was diluted in ethyl acetate then filtered through a plug of celite. The reaction outcome was assessed by GC and TLC (isomeric ratios were determined by GC). The mixture was purified by flash column chromatography or crystallization under the specified conditions, and the product was dried under vacuum.

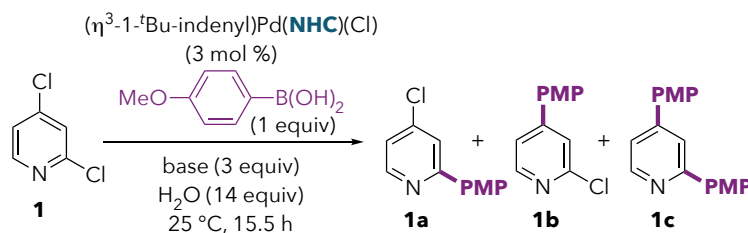
**Conditions A:** potassium carbonate (221.1 mg, 1.6 mmol, 4.0 equiv), deionized water (250  $\mu$ L, 13.8 mmol, 34.6 equiv), THF, 60  $^{\circ}$ C.

**Conditions B:** potassium carbonate (165.8 mg, 1.2 mmol, 3.0 equiv), deionized water (100  $\mu$ L, 5.53 mmol, 13.8 equiv), THF, 25  $^{\circ}$ C.

**Conditions C:** potassium carbonate (165.8 mg, 1.2 mmol, 3.0 equiv), deionized water (100  $\mu$ L, 5.53 mmol, 13.8 equiv), benzene, 25  $^{\circ}$ C.

## 2. Optimization (Table 1)

**Table S1.** Optimization of the C4-Selective Suzuki-Miyaura Coupling with Pd/IPr<sup>a</sup>



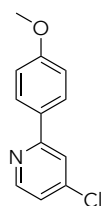
entry	trial	NHC	base	solvent	additive (equiv)	1a (%)	1b (%)	1c (%)	1a : 1b
1	1	SIPr	KF	THF	--	4	35	4	1 : 8.8
2	2	SIPr	KF	THF	--	5	44	5	1 : 8.8
3	Average	SIPr	KF	THF	--	5	39	5	1 : 7.8
4	1	SIPr	K <sub>2</sub> CO <sub>3</sub>	THF	--	7.4	67.7	9.1	1 : 9.1
5	2	SIPr	K <sub>2</sub> CO <sub>3</sub>	THF	--	6.2	63.0	10.3	1 : 10.1
6	Average	SIPr	K <sub>2</sub> CO <sub>3</sub>	THF	--	6.8	65.4	9.7	1 : 9.6
7	1	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	--	6	69	8	1 : 11.5
8	2	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	--	7	68	7	1 : 9.7
9	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	--	7	69	8	1 : 9.9
10	1	IPr	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	8	69	6.0	1 : 8.6
11	2	IPr	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	8	70	6.4	1 : 8.8
12	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	8	70	6.2	1 : 8.8
13	1	IPr	K <sub>2</sub> CO <sub>3</sub>	PhCH <sub>3</sub>	--	8	75	4	1 : 9.4
14	2	IPr	K <sub>2</sub> CO <sub>3</sub>	PhCH <sub>3</sub>	--	8	72	4	1 : 9.0
15	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	PhCH <sub>3</sub>	--	8	74	4	1 : 9.2
16	1	IPr	K <sub>2</sub> CO <sub>3</sub>	DMF	--	8	46	17	1 : 5.8
17	2	IPr	K <sub>2</sub> CO <sub>3</sub>	DMF	--	8	48	14	1 : 5.9
18	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	DMF	--	8	47	15	1 : 5.9
19	1	IPr	K <sub>2</sub> CO <sub>3</sub>	PC <sup>b</sup>	--	9	43	16	1 : 4.8
20	2	IPr	K <sub>2</sub> CO <sub>3</sub>	PC <sup>b</sup>	--	9	46	18	1 : 5.1
21	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	PC <sup>b</sup>	--	9	45	17	1 : 5.0
22 <sup>c</sup>	1	--	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	2	<1	n.d.	--
23 <sup>c</sup>	2	--	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	1	<1	n.d.	--

24 <sup>e</sup>	Average	--	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	2	<1	n.d.	--
25 <sup>d</sup>	1	--	K <sub>2</sub> CO <sub>3</sub>	THF	--	2	1	n.d.	--
26 <sup>d</sup>	2	--	K <sub>2</sub> CO <sub>3</sub>	THF	--	1	<1	n.d.	--
27 <sup>d</sup>	Average	--	K <sub>2</sub> CO <sub>3</sub>	THF	--	1	<1	n.d.	--
28	1	SIPr	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	9	61	8	1 : 6.8
29	2	SIPr	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	9	66	7	1 : 7.3
30	Average	SIPr	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	9	64	8	1 : 7.1
31	1	SIMes	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	21	30	3	1 : 1.4
32	2	SIMes	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	27	39	3	1 : 1.4
33	Average	SIMes	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	24	35	3	1 : 1.5
34	1	IMes	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	21	35	3	1 : 1.6
35	2	IMes	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	24	44	3	1 : 1.7
36	Average	IMes	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	23	40	3	1 : 1.7
37	1	SIMix	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	8	36	2	1 : 4.5
38	2	SIMix	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	8	31	2	1 : 3.9
39	Average	SIMix	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	8	34	2	1 : 4.3
40	1	IPent	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	1	31	21	1 : 31
41	2	IPent	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	1	37	24	1 : 37
42	Average	IPent	K <sub>2</sub> CO <sub>3</sub>	C <sub>6</sub> H <sub>6</sub>	--	1	34	23	1 : 34
43	1	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Br (1)	<1	3	n.d.	--
44	2	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Br (1)	1	4	n.d.	--
45	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Br (1)	<1	3	n.d.	--
46	1	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Br (3)	<1	2	n.d.	--
47	2	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Br (3)	<1	2	n.d.	--
48	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Br (3)	<1	2	n.d.	--
49	1	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Cl (3)	<1	1	n.d.	--
50	2	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Cl (3)	<1	<1	n.d.	--
51	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> Cl (3)	<1	1	n.d.	--
52	1	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> OH (3) <sup>e</sup>	1	4	n.d.	--
53	2	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> OH (3) <sup>e</sup>	1	4	n.d.	--
54	Average	IPr	K <sub>2</sub> CO <sub>3</sub>	THF	NBu <sub>4</sub> OH (3) <sup>e</sup>	1	4	n.d.	--
55	1	IPr	K <sub>2</sub> CO <sub>3</sub>	dioxane	--	6.7	77.1	7.6	1 : 12
56 <sup>f</sup>	1	IPr	K <sub>2</sub> CO <sub>3</sub>	dioxane	--	2.8	43.0	25.4	1 : 15
57	1	IPr	K <sub>2</sub> CO <sub>3</sub>	toluene	--	8.8	78.2	4.8	1 : 9
58 <sup>f</sup>	1	IPr	K <sub>2</sub> CO <sub>3</sub>	toluene	--	8.5	44.6	15.4	1 : 5

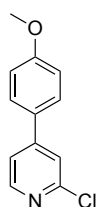
<sup>a</sup>Reactions were conducted according to the General Procedure for GC-scale reactions. GC yields calibrated against undecane as an internal standard. Calculated yield values were rounded to the nearest integer. Entries reported as <1 indicate a calibrated GC yield less than 0.5%; nd = not detected. <sup>b</sup>PC = propylene carbonate. <sup>c</sup>No NHC ligand; Pd source was (η<sup>3</sup>-1-<sup>t</sup>Bu-indenyl)<sub>2</sub>(μ-Cl)<sub>2</sub>Pd<sub>2</sub> (1.5 mol %). <sup>d</sup>No NHC ligand; Pd source was PdCl<sub>2</sub> (3 mol %). <sup>e</sup>NBu<sub>4</sub>OH was added as a solution in water (156 μL of a 40% aqueous solution, 3 equiv) added; no additional water added to the reaction. <sup>f</sup>Reaction was conducted at 100 °C.

Discussion: Unlike our optimized ligand-free conditions (*vide infra*), the addition of tetraalkylammonium salts to the Pd/IPr catalytic conditions inhibits cross-coupling (Table S1, entries 43-54). The use of high temperatures in the Pd/IPr conditions does not lead to significant improvements in C<sub>4</sub>-selectivity (compare entry 55 to 56, and entry 57 to 58), indicating that the high-temperatures used in ligand-free conditions are not solely responsible for the greatly enhanced selectivity under 'Jeffery' conditions.

### 3. Isolation and Characterization of Products (Table 1 and Scheme 2)

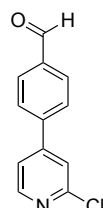


**4-Chloro-2-(4-methoxyphenyl)pyridine (1a).** Compound **1a** was prepared according to a modified literature procedure<sup>10</sup>. Pd(OAc)<sub>2</sub> (33.7 mg, 0.15 mmol, 5 mol %), 1,1'-bis(diphenylphosphino)ferrocene (109.8 mg, 0.15 mmol, 5 mol %), Cs<sub>2</sub>CO<sub>3</sub> (2.44 g, 7.5 mmol 2.5 equiv) and *p*-methoxyphenylboronic acid (455.8 mg, 3 mmol, 1 equiv) were added to an oven-dried 5-dram vial in a N<sub>2</sub>-filled glovebox. The vial was sealed with a septum and removed from the glovebox, and 2,4-dichloropyridine (**1**, 322 μL, 3 mmol, 1 equiv), deionized water (378 μL, 21 mmol, 7 equiv), and 1,4-dioxane (12 mL) were added through the septum. The reaction was allowed to stir at 70 °C for 19 h. Purification via flash column chromatography (*R<sub>f</sub>* = 0.50 in 20% ethyl acetate in hexanes) yielded **1a** as an off-white solid (504 mg, 76% yield). Spectral data are consistent with the literature.<sup>11</sup>



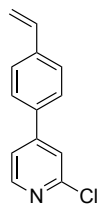
**2-Chloro-4-(4-methoxyphenyl)pyridine (1b).** Compound **1b** was prepared according to the general procedure using Conditions B and 2,4-dichloropyridine (43.2 μL, 0.4 mmol, 1.0 equiv) with stirring for 16.5 h. Three separate reactions were performed using *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv), *p*-methoxyphenylboronic acid pinacol ester (93.6 mg, 0.4 mmol, 1.0 equiv), or *p*-methoxyphenylboronic acid neopentyl glycol ester (88.0 mg, 0.4 mmol, 1.0 equiv). Purification by flash column chromatography on silica gel (*R<sub>f</sub>* = 0.29 in 10% acetone in hexanes) provided **1b** and other isolated side products as white solids with varying degrees of brown discoloration. *p*-Methoxyphenylboronic acid method: (63.3 mg, 72% yield); *p*-methoxyphenylboronic acid pinacol ester method: (61.5 mg, 70% yield); diarylated side product **1c**: 15.2 mg, 13% yield); *p*-methoxyphenylboronic acid neopentyl glycol ester method: (55.4 mg, 63% yield); diarylated side product **1c**: 10.5 mg, 9% yield). **1b**: (600 MHz, CDCl<sub>3</sub>, δ): 8.36 (d, *J* = 5.2 Hz, 1H), 7.6-7.5 (m, 2H), 7.48 (d, *J* = 1.1 Hz, 1H), 7.37 (dd, *J* = 1.6, 5.3 Hz, 1H), 7.01-6.98 (m, 2H), 3.85 (s, 3H). Spectral data are consistent with the literature.<sup>12</sup> 2,4-bis(4-methoxyphenyl)pyridine (**1c**): (600 MHz, CDCl<sub>3</sub>, δ): 8.65 (d, *J* = 5.2 Hz, 1H), 8.00 (m, 2H), 7.83 (s, 1H), 7.65 (m, 2H), 7.35 (dd, *J* = 5.2, 1.4 Hz, 1H), 7.04-7.01 (multiple peaks, 4H), 3.88 (coincidentally overlapping singlets, 6H). Spectral data are consistent with the literature.<sup>13</sup>

**Larger-Scale Preparation of 1b:** Compound **1b** was prepared according to the general procedure using Conditions B with 2,4-dichloropyridine (216 μL, 2.0 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (303.9 mg, 2.0 mmol, 1.0 equiv) with stirring for 27 h. Purification by flash column chromatography on silica gel (*R<sub>f</sub>* = 0.29 in 10% acetone in hexanes) provided **1b** as a white solid with slight yellow discoloration (265.8 mg, 61% yield). Spectral data are consistent with the literature.<sup>12</sup>

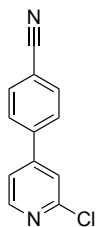


**4-(2-Chloro-4-pyridinyl)benzaldehyde (4b).** Compound **4b** was prepared according to the general procedure using Conditions B, 2,4-dichloropyridine (43.2 μL, 0.4 mmol, 1.0 equiv) and *p*-formylphenylboronic acid (60.0 mg, 0.4 mmol, 1.0 equiv) with stirring for 16 h. Purification by flash column chromatography on silica (*R<sub>f</sub>* = 0.22 in 10% acetone in hexanes) provided **4b** as a white solid (38.8 mg, 45% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, δ): 10.08 (s, 1H), 8.48 (d, *J* = 5.2 Hz, 1H), 8.01-7.99 (m, 2H), 7.78-7.76 (m, 2H), 7.57 (d, *J* = 1.5 Hz, 1H), 7.46 (dd, *J* = 5.2, 1.5 Hz, 1H). Spectral data are consistent with

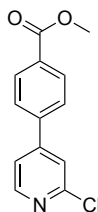
the literature.<sup>14</sup> The C2-monosubstituted isomer **S4a** was isolated as a minor product in low purity (7.5 mg, <9% yield). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, δ): 10.9 (s, 1H), 8.64 (d, *J* = 5.3 Hz, 1H), 8.17-8.15 (m, 2H), 8.01-7.99 (m, 2H), 7.18 (d, *J* = 1.8 Hz, 1H), 7.32 (dd, *J* = 1.8, 5.3 Hz, 1H). The minor product of diarylation **4c** was also isolated (8.3 mg, 7% yield). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, δ): 10.12 (s, 1H), 10.11 (s, 1H), 8.84 (d, *J* = 5.0 Hz, 1H), 8.26-8.24 (m, 2H), 8.05-8.02 (multiple peaks, 5H), 7.88-7.86 (m, 2H), 7.55 (dd, *J* = 1.6, 5.0 Hz, 1H).



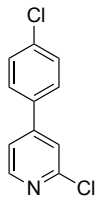
**2-Chloro-4-(4-vinylphenyl)pyridine (5b).** Compound **5b** was prepared according to the general procedure using Conditions B, 2,4-dichloropyridine (43.2 μL, 0.4 mmol, 1.0 equiv) and *p*-vinylphenylboronic acid (60.0 mg, 0.4 mmol, 1.0 equiv) with stirring for 16.5 h. Purification by flash column chromatography on silica (*R<sub>f</sub>* = 0.21 in 2.5% acetone in hexanes) provided **5b** as an off-white solid (55.7 mg, 65% yield). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, δ): 8.39 (d, *J* = 5.2 Hz, 1H), 7.56-7.55 (m, 2H), 7.51-7.49 (multiple overlapping signals, 3H), 7.39 (d, *J* = 5.2 Hz, 1H), 6.74 (dd, *J* = 17.6, 10.9 Hz, 1H), 5.83 (d, *J* = 17.6 Hz, 1H), 5.34 (d, *J* = 10.9 Hz, 1H). <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, CDCl<sub>3</sub>, δ): 152.3, 151.0, 150.1, 139.1, 135.9 (two signals are coincidentally overlapping), 127.2, 127.1, 121.7, 120.2, 115.5. HRMS (ESI Q-TOF) *m/z*: [M]<sup>+</sup> Calcd for C<sub>13</sub>H<sub>10</sub>ClN 215.0502; Found 215.04922.



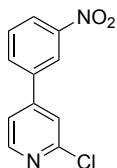
**4-(2-Chloro-4-pyridinyl)benzonitrile (6b).** Compound **6b** was prepared according to the general procedure using Conditions B, 2,4-dichloropyridine (43.2 μL, 0.4 mmol, 1.0 equiv) and *p*-cyanophenylboronic acid (58.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 16.5 h. Purification by flash column chromatography on silica (*R<sub>f</sub>* = 0.19 in 10% acetone in hexanes) provided **6b** as a white crystalline solid (60.3 mg, 70% yield); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, δ): 8.47 (d, *J* = 5.2 Hz, 1H), 7.78-7.77 (m, 2H), 7.71-7.70 (m, 2H), 7.52 (d, *J* = 1.1 Hz, 1H), 7.42 (dd, *J* = 5.2, 1.1 Hz, 1H). The spectral data are consistent with the literature.<sup>15</sup> The C2-monosubstituted isomer **S6a** was isolated as a minor product in low purity (7.8 mg, <9% yield). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, δ): 8.63 (d, *J* = 5.2 Hz, 1H), 8.11-8.10 (m, 2H), 7.78-7.77 (multiple overlapping signals, 3H), 7.33 (dd, *J* = 5.2, 1.8 Hz, 1H). The minor product of diarylation **S6c** was also isolated (14.3 mg, 13% yield). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, δ): 8.84 (d, *J* = 5.0 Hz, 1H), 8.19-8.18 (m, 2H), 7.94 (s, 1H), 7.83-7.79 (multiple overlapping signals, 6H), 7.52 (dd, *J* = 5.0, 1.3 Hz, 1H).



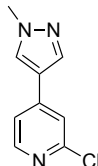
**4-(2-Chloro-4-pyridinyl)benzoic acid methyl ester (7b).** Compound **7b** was prepared according to the general procedure using Conditions B, 2,4-dichloropyridine (43.2 μL, 0.4 mmol, 1.0 equiv) and *p*-(methoxycarbonyl)phenylboronic acid (72.0 mg, 0.4 mmol, 1.0 equiv) with stirring for 16 h. Purification by flash column chromatography on silica (*R<sub>f</sub>* = 0.24 in 5% acetone in hexanes) provided **7b** as a white microcrystalline solid (59.8 mg, 60% yield). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>, δ): 8.43 (d, *J* = 4.9 Hz, 1H), 8.13-8.12 (m, 2H), 7.65-7.64 (m, 2H), 7.53 (s, 1H), 7.42 (d, *J* = 4.9 Hz, 1H), 3.93 (s, 3H). Spectral data are consistent with the literature.<sup>15</sup>



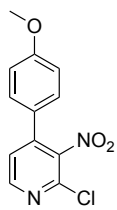
**2-Chloro-4-(4-chlorophenyl)pyridine (8b).** Compound **8b** was prepared according to the general procedure using Conditions B, 2,4-dichloropyridine (43.2  $\mu\text{L}$ , 0.4 mmol, 1.0 equiv) and *p*-chlorophenylboronic acid (62.5 mg, 0.4 mmol, 1.0 equiv) with stirring for 16 h. Purification by flash column chromatography on silica ( $R_f = 0.25$  in 5% THF in hexanes) resulted in partial separation of **8b** from the C2-monoarylated isomer. Coeluted fractions were purified by evaporative crystallization from acetone and water, providing **8b** as an off-white solid (59 mg, 66% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.43 (d,  $J = 5.2$  Hz, 1H), 7.55-7.52 (m, 2H), 7.50 (d,  $J = 1.6$  Hz, 1H), 7.47-7.45 (m, 2H), 7.38 (dd,  $J = 5.2, 1.6$  Hz, 1H). Spectral data are consistent with the literature.<sup>15</sup>



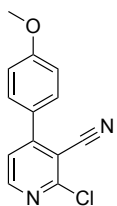
**2-Chloro-4-(3-nitrophenyl)pyridine (9b).** Compound **9b** was prepared according to the general procedure (except on the 1.0 mmol scale) using Conditions B, 2,4-dichloropyridine (108.0  $\mu\text{L}$ , 1.0 mmol, 1.0 equiv) and *m*-nitrophenylboronic acid (166.9 mg, 1.0 mmol, 1.0 equiv) with stirring for 16 h. Purification by flash column chromatography on silica ( $R_f = 0.24$  in 10% acetone in hexanes) provided **9b** as a white crystalline solid (156.7 mg, 67% yield); mp 159-161  $^\circ\text{C}$  [uncorrected, measured against benzoic acid (113-117  $^\circ\text{C}$ )].  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.52 (dd,  $J = 5.2, 0.5$  Hz, 1H), 8.48 (dd,  $J = 1.8, 1.8$  Hz, 1H), 8.33 (ddd,  $J = 8.0, 1.9, 0.9$  Hz, 1H), 7.95 (ddd,  $J = 8.0, 1.9, 0.9$  Hz, 1H), 7.71 (dd,  $J = 8.0, 8.0$  Hz, 1H), 7.60 (dd,  $J = 1.9, 0.8$  Hz, 1H), 7.48 (dd,  $J = 5.2, 1.8$  Hz, 1H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 152.9, 150.7, 149.1, 149.0, 138.8, 133.0, 130.6, 124.4, 122.3, 122.2, 120.5. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{11}\text{H}_7\text{ClN}_2\text{O}_2$  234.0196; Found 234.0193.



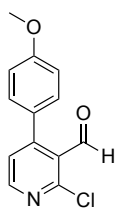
**2-Chloro-4-(1-methyl-1H-pyrazol-4-yl)pyridine (10b).** Compound **10b** was prepared according to the general procedure using Conditions B, with the modification that the reaction was heated to 75  $^\circ\text{C}$ , and with 2,4-dichloropyridine (43.2  $\mu\text{L}$ , 0.4 mmol, 1.0 equiv) and (1-methyl-1H-pyrazol-4-yl)boronic acid (75.5 mg, 0.6 mmol, 1.5 equiv) with stirring for 35 h. Products were purified via flash column chromatography using a step gradient: 0.75% methanol in  $\text{CH}_2\text{Cl}_2$  until elution of the C2-monoarylated product **S10a**, 3% methanol in  $\text{CH}_2\text{Cl}_2$  until elution of **10b**, 5% methanol in  $\text{CH}_2\text{Cl}_2$  until elution of the diarylated product **S10c** ( $R_f$  values in 0.75% methanol/ $\text{CH}_2\text{Cl}_2$ : C2-monoarylated isomer **S10a** = 0.17, C4-monoarylated isomer **10b** = 0.31, diarylated product **S10c** = 0.09). Product **10b** was isolated as an off-white solid (52 mg, 67% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.27 (d,  $J = 5.3$  Hz, 1H), 7.80 (s, 1H), 7.72 (s, 1H), 7.34 (d,  $J = 1.3$  Hz, 1H), 7.22 (dd,  $J = 5.3, 1.3$  Hz, 1H), 3.94 (s, 3H).  $^1\text{H}$  NMR (500 MHz,  $\text{DMSO}-d_6$ ,  $\delta$ ): 8.43 (s, 1H), 8.30 (dd,  $J = 5.3, 0.6$  Hz, 1H), 8.12 (d,  $J = 0.6$  Hz, 1H), 7.71 (dd,  $J = 1.5, 0.6$  Hz, 1H), 7.57 (dd,  $J = 5.3, 1.5$  Hz, 1H), 3.88 (s, 3H). Spectral data are consistent with the literature.<sup>16</sup> The C2-monoarylated isomer (**S10a**) was isolated as a minor product in low purity (10 mg, <13% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.42 (d,  $J = 5.4$  Hz, 1H), 7.93 (s, 1H), 7.92 (s, 1H), 7.44 (d,  $J = 1.9$  Hz, 1H), 7.10 (dd,  $J = 5.4, 1.9$  Hz, 1H), 3.95 (s, 3H). The minor product of diarylation (**S10c**) was also isolated (9 mg, 9% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.48 (dd,  $J = 5.2, 0.7$  Hz, 1H), 7.97 (s, 1H), 7.95 (s, 1H), 7.87 (d,  $J = 0.7$  Hz, 1H), 7.76 (s, 1H), 7.50 (dd,  $J = 1.7, 0.7$  Hz, 1H), 7.15 (dd,  $J = 5.2, 1.7$  Hz, 1H), 3.97 (s, 3H), 3.96 (s, 3H).



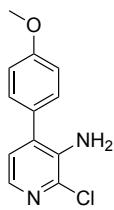
**2-Chloro-4-(4-methoxyphenyl)-3-nitropyridine (11b).** Compound **11b** was prepared according to the general procedure using Conditions A, 2,4-dichloro-3-nitro-pyridine (77.2 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 9 h. Purification by flash column chromatography on silica gel in 10-15% acetone/hexanes ( $R_f = 0.25$  in 10% acetone in hexanes) provided **11b** as a colorless crystalline solid (70.3 mg, 66% yield), with evidence of yellow impurity (1H NMR shows there is a 6% impurity comprising C2-monoarylated isomer **S11a**); mp 110-113 °C [uncorrected, measured against benzoic acid (110-113 °C)].  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.49 (d,  $J = 5.1$  Hz, 1H), 7.36-7.33 (multiple peaks, 3H), 6.99-6.97 (m, 2H), 3.85 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 161.5, 150.0, 145.1 (this small quaternary peak was selected from among impurity peaks based on comparison with an ACD labs simulated spectrum), 144.3, 142.3, 129.2, 125.0, 124.3, 115.0, 55.5. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{12}\text{H}_9\text{ClN}_2\text{O}_3$  264.0302; Found 264.0298.



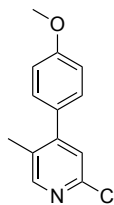
**2-Chloro-4-(4-methoxyphenyl)-3-pyridinecarbonitrile (12b).** Compound **12b** was prepared according to the general procedure using Conditions A, 2,4-dichloro-3-cyano-pyridine (69.2 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 12 h. Purification by flash column chromatography on silica gel ( $R_f = 0.21$  in 10% acetone in hexanes) provided **12b** as a colorless crystalline solid (66.4 mg, 68% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.52 (d,  $J = 5.3$  Hz, 1H), 7.60-7.57 (m, 2H), 7.36 (d,  $J = 5.3$  Hz, 1H), 7.07-7.04 (m, 2H), 3.88 (s, 3H). Spectral data are consistent with the literature.<sup>17</sup> The minor product of diarylation (**S12c**) was also isolated (4.4 mg, 3% yield;  $R_f = 0.10$  in 10% acetone in hexanes).  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.77 (d,  $J = 5.1$  Hz, 1H), 7.91-7.90 (m, 2H), 7.62-7.58 (m, 2H), 7.31 (d,  $J = 5.1$  Hz, 1H), 7.07-7.04 (multiple peaks, 4H), 3.89 (coincidentally overlapping singlets, 6H).



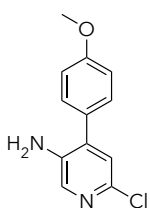
**2-Chloro-4-(4-methoxyphenyl)-3-pyridinecarboxaldehyde (13b).** Compound **13b** was prepared according to the general procedure using Conditions C, 2,4-dichloro-3-pyridinecarboxaldehyde (70.4 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 12 h. Purification by flash column chromatography on silica gel ( $R_f = 0.21$  in 10% acetone in hexanes) provided **13b** as a yellow solid (51.4 mg, 52% yield).  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 10.14 (s, 1H), 8.49 (d,  $J = 5.1$  Hz, 1H), 7.31 (d,  $J = 5.1$  Hz, 1H), 7.29-7.26 (m, 2H), 7.02-7.00 (m, 2H), 3.87 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 190.3, 161.0, 154.4, 151.5, 151.1, 130.8, 128.1, 127.6, 124.8, 114.5, 55.5. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{13}\text{H}_{10}\text{ClNO}_2$  247.0400; Found 247.0385. The C2-mononarylated isomer **S13a** was isolated as a minor product in low purity (9.9 mg, <10% yield).  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 9.97 (s, 1H), 8.65 (d,  $J = 5.3$  Hz, 1H), 7.51-7.49 (m, 2H), 7.38 (d,  $J = 5.3$  Hz, 1H), 7.03-7.02 (m, 2H), 3.88 (s, 3H).



**2-Chloro-4-(4-methoxyphenyl)-3-pyridinamine (14b).** Compound **14b** was prepared according to the general procedure using Conditions C, 3-amino-2,4-dichloro-pyridine (65.2 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 18.5 h. Purification by flash column chromatography on silica gel ( $R_f = 0.42$  in 25% ethyl acetate in hexanes) provided **14b** as a white crystalline solid (56.7 mg, 60% yield); mp 113-116 °C [uncorrected, measured against benzoic acid (111-113 °C)].  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.79 (d,  $J = 4.8$  Hz, 1H), 7.38-7.35 (m, 2H), 7.01-6.98 (m, 2H), 6.95 (d,  $J = 4.8$  Hz, 1H), 4.19 (br s, 2H), 3.85 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 159.9, 138.1, 137.6, 137.3, 135.2, 129.6, 128.7, 124.2, 114.7, 55.5. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{12}\text{H}_{11}\text{ClN}_2\text{O}$  234.0560; Found 234.0557. The C2-monoarylated isomer **S14a** was isolated as a minor product (12.2 mg, 13% yield).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.98 (d,  $J = 5.2$  Hz, 1H), 7.62-7.59 (m, 2H), 7.15 (d,  $J = 5.2$  Hz, 1H), 7.02-7.00 (m, 2H), 4.23 (br s, 2H), 3.86 (s, 3H). The minor product of diarylation **S14c** was also isolated (16.7 mg, 14% yield).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.13 (d,  $J = 4.9$  Hz, 1H), 7.68-7.66 (m, 2H), 7.46-7.44 (m, 2H), 7.03-7.00 (multiple peaks, 4H), 6.98 (d,  $J = 4.9$  Hz, 1H), 3.90 (br s, 2H), 3.864 (s, 3H), 3.859 (s, 3H).



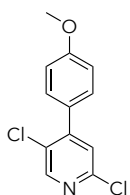
**2-Chloro-4-(4-methoxyphenyl)-5-methylpyridine (15b).** Compound **15b** was prepared according to the general procedure using Conditions B, 2,4-dichloro-5-methylpyridine (64.8 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 18.5 h. Purification by flash column chromatography on silica gel ( $R_f =$  in 5% acetone in hexanes) resulted in partial purification of the C4-monoarylated isomer. One fraction containing coeluted monoarylated isomers and diarylated product was purified further by preparatory TLC in the same eluent. Product **15b** was extracted from the silica gel with acetone and combined with remaining **15b** fractions obtained from flash column chromatography, providing an off-white solid (49.9 mg, 53% yield).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.24 (s, 1H), 7.26-7.24 (m, 2H), 7.19 (s, 1H), 7.00-6.98 (m, 2H), 3.86 (s, 3H), 2.25 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (151 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 159.9, 152.1, 150.9, 149.2, 130.3, 129.91, 129.87, 124.2, 114.2, 55.5, 17.0. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{13}\text{H}_{12}\text{ClNO}$  233.0607; Found 233.0603. The minor product of diarylation **S15c** was also isolated (21.8 mg, 18% yield).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.52 (s, 1H), 7.96-7.94 (m, 2H), 7.52 (s, 1H), 7.33-7.31 (m, 2H), 7.01-7.98 (multiple peaks, 4H), 3.87 (s, 3H), 3.86 (s, 3H), 2.30 (s, 3H).



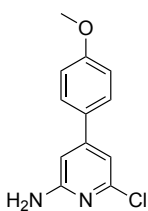
**6-Chloro-4-(4-methoxyphenyl)-3-pyridinamine (16b).** Compound **16b** was prepared according to the general procedure using Conditions C, 5-amino-2,4-dichloropyridine (78.2 mg, 0.48 mmol, 1.2 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 21 h. Purification was performed by reversed phase flash column chromatography with a 12 g C18 silica column, using an initial automated program comprising of a flow rate of 36 mL/min of water:acetonitrile (98:2 to 6:94) over 30 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 50-53% MeCN in water. After the elution, the initial program was resumed. The C4 monoarylated product **16b** coeluted with the C2 monoarylated isomer **S16a** at 52-53% MeCN. Fractions containing **16b** were partitioned between dichloromethane and saturated brine. The organic layers were combined and dried over magnesium sulfate and solvent was removed under reduced pressure affording a mixture of the product



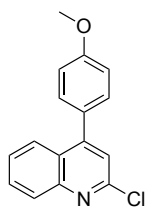
isomers. Further purification attempts were unsuccessful, and the final product **16b** was obtained as a rust colored solid in 94% purity (74.4 mg, 74% yield based on 94% purity).  $^1\text{H}$  NMR (400 MHz,  $\text{C}_6\text{D}_6$ ,  $\delta$ ): 7.55 (s, 1H), 6.55-7.00 (m, 2H), 6.90 (s, 1H), 6.66-6.71 (m, 2H), 3.28 (s, 3H), 2.88 (br s, 2H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (100 MHz,  $\text{C}_6\text{D}_6$ ,  $\delta$ ): 160.6, 141.5, 139.9, 137.5, 136.9, 130.1, 124.6, 125.3, 115.1, 55.2. HRMS (ESI<sup>+</sup>) m/z: [M]<sup>+</sup>: Calcd for  $\text{C}_{12}\text{H}_{11}\text{ClN}_2\text{O}$  234.0560; Found 234.0562.



**2,5-Dichloro-4-(4-methoxyphenyl)pyridine (17b).** Compound **17b** was prepared according to the general procedure using Conditions C, 2,4,5-trichloropyridine (73.0 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 21 h. Purification was performed by reversed phase flash column chromatography with a 12 g C18 silica column, using an initial automated program comprising a flow rate of 30 mL/min of water:methanol (98:2 to 0:100) over 40 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 65–60% MeOH in water. After the elution, the initial program was resumed. The C4 monoarylated product eluted at 67% MeOH. Fractions containing product **17b** were partitioned between ethyl acetate and saturated brine. The organic layers were combined and dried over magnesium sulfate and solvent was removed under reduced pressure affording an off-white solid (44.0 mg, 43% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{C}_6\text{D}_6$ ,  $\delta$ ): 8.16 (s, 1H), 6.94-7.02 (m, 2H), 6.87 (s, 1H), 6.66-6.71 (m, 2H), 3.26 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (150 MHz,  $\text{C}_6\text{D}_6$ ,  $\delta$ ): 160.5, 149.9, 149.51, 149.46, 130.2, 128.9, 128.0, 125.3, 113.8, 54.5 ppm. HRMS (ESI<sup>+</sup>) m/z: [M]<sup>+</sup>: Calcd for  $\text{C}_{12}\text{H}_9\text{Cl}_2\text{NO}$  253.0061; Found 253.0073.

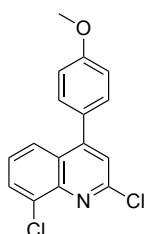


**6-Chloro-4-(4-methoxyphenyl)-2-pyridinamine (18b).** Compound **18b** was prepared according to the general procedure using Conditions C, 2-amino-4,6-dichloro-pyridine (65.2 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 25.5 h. Purification by flash column chromatography on silica gel ( $R_f = 0.22$  in 25% ethyl acetate in hexanes) provided **18b** as a colorless crystalline solid with minor brown discoloration (45.1 mg, 48% yield); mp 167-172 °C [uncorrected, measured against benzoic acid (115-118 °C)].  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.51-7.49 (m, 2H), 6.96 (m, 2H), 6.86 (d,  $J = 1.3$  Hz, 1H), 6.53 (d,  $J = 1.3$  Hz, 1H), 4.61 (br s, 2H), 3.85 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 160.8, 158.8, 152.6, 150.2, 130.0, 128.2, 114.6, 111.5, 103.8, 55.5. HRMS (ESI Q-TOF) m/z: [M]<sup>+</sup> Calcd for  $\text{C}_{12}\text{H}_{11}\text{ClN}_2\text{O}$  234.0560; Found 234.0551. The C2-monoarylated isomer **S18a** was isolated as a minor product ( $R_f = 0.30$  in 25% ethyl acetate in hexanes; 12.6 mg, 13% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.88-7.86 (m, 2H), 7.03 (d,  $J = 1.5$  Hz, 1H), 6.97-6.95 (m, 2H), 6.40 (d,  $J = 1.5$  Hz, 1H), 4.57 (br s, 2H), 3.85 (s, 3H). The column eluent was modified to 5% methanol in  $\text{CH}_2\text{Cl}_2$  to elute the diarylated product **S18c**, isolated in low purity ( $R_f = 0.02$  in 25% ethyl acetate in hexanes; 19.7 mg, <16% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.94-7.91 (m, 2H), 7.60-7.57 (m, 2H), 7.22 (d,  $J = 1.3$  Hz, 1H), 7.00-6.96 (multiple peaks, 4H), 6.58 (d,  $J = 1.3$  Hz, 1H), 4.62 (br s, 2H), 3.859 (s, 3H), 3.856 (s, 3H).



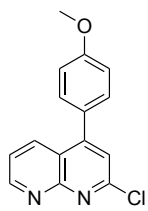
**2-Chloro-4-(4-methoxyphenyl)quinoline (19b).** Compound **19b** was prepared according to the general procedure using Conditions B on a 3.0 mmol scale using 2,4-dichloroquinoline (**19**, 594.2 mg, 3.0 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (455.9 mg, 3.0 mmol, 1.0 equiv) with stirring for 19 h. The product mixture was diluted in ethyl acetate, filtered through celite, and solvent was removed under reduced pressure (as described in the general procedure) Purification method 1:

The concentrated residue was dissolved in excess acetone in a conical flask, then water was added until the solution appeared to become saturated. Slow evaporation of the acetone/water solution from the flask induced crystallization. Yellow-tinted crystals were collected from three crops, separated from solvent, and dried in air to provide product **19b** (454 mg, 56% yield). Purification method 2: The concentrated residue was purified on a Biotage Selekt automated column using 100 grams of high-capacity silica (see *General Materials and Methods*), a linear gradient of 2-20% acetone in hexanes over 12 column volumes, at a flow rate of 120 mL/min ( $R_f = 0.60$  in 10% acetone in hexanes). GC analysis indicated coelution of the C2- and C4-monoarylated isomers (**S19a** and **19b**); however, large colorless crystals of **19b** formed selectively within the fractions, and were collected in two separate crops and dried under air to provide product **19b** (403.2 mg, 50% yield); mp 152-154 °C [uncorrected, measured against benzoic acid (113-117 °C)].  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.08 (d,  $J = 8.4$  Hz, 1H), 7.93 (d,  $J = 8.4$  Hz, 1H), 7.73 (ddd,  $J = 8.4, 7.0, 1.3$  Hz, 1H), 7.51 (ddd,  $J = 8.4, 7.0, 1.2$  Hz, 1H), 7.46-7.43 (m, 2H), 7.32 (s, 1H), 7.08-7.05 (m, 2H), 3.91 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 160.4, 151.5, 150.5, 148.6, 130.9, 130.5, 129.15, 129.12, 126.9, 126.1, 125.9, 122.1, 114.3, 55.6. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{16}\text{H}_{12}\text{ClNO}$  269.0607; Found 269.0606.

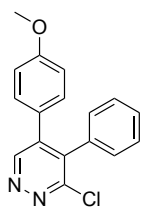


**2,8-Dichloro-4-(4-methoxyphenyl)quinoline (20b).** Compound **20b** was prepared according to the general procedure using Conditions C, 2,4,8-trichloroquinoline (102.3 mg, 0.44 mmol, 1.1 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 15.5 h. The product was purified by reversed phase flash column chromatography with a 12 g C18 silica column, using an initial automated program utilizing a flow rate of 13 mL/min of water:acetonitrile (98:2 to 5:95) over 30 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 61-70% MeCN in water. After the elution, the initial program was resumed. The C4 monoarylated product **20b** eluted cleanly at 65-67% MeCN, but the C2-monoarylated isomer **S20a** and the 2,4-diarylated product **S20c** subsequently coeluted with each other. Fractions containing **20b** were partitioned between ethyl acetate and saturated brine. The organic layers were combined and dried over magnesium sulfate, concentrated, and dried furnishing **20b** as a white solid (65.9 mg, 54% yield).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.86 (dd,  $J = 1.8, 1.4$  Hz, 1H), 7.84 (dd,  $J = 3.0, 1.4$  Hz, 1H), 7.43-7.40 (multiple peaks, 3H), 7.38 (s, 1H), 7.08-7.05 (m, 2H), 3.90 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 160.6, 152.1, 151.5, 145.0, 133.1, 130.9, 130.6, 128.9, 127.5, 126.7, 125.3, 123.2, 114.4, 55.6. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{16}\text{H}_{11}\text{Cl}_2\text{NO}$  303.0218; Found 303.0217. The minor products **S20a** and **S20c** were further purified by (normal-phase) flash column chromatography in 5-20% acetone in hexanes. The C2-monoarylated isomer **S20a** was isolated as a minor product in low purity (4.0 mg, <3% yield).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.24-8.22 (m, 2H), 8.13 (dd,  $J = 8.4, 1.3$  Hz, 1H), 8.01 (s, 1H), 7.87 (dd,  $J = 7.5, 1.3$  Hz, 1H), 7.48 (dd,  $J = 8.4, 7.5$  Hz, 1H), 7.07-7.04 (m, 2H), 3.90 (s, 3H). The minor product of 2,4-diarylation **S20c** was also isolated in low purity (7.3 mg, <5% yield).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ):

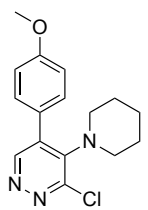
8.30-8.27 (m, 2H), 7.83-7.81 (multiple peaks, 3H), 7.49-7.46 (m, 2H), 7.33 (dd,  $J = 8.4, 7.5$  Hz, 1H), 7.09-7.07 (m, 2H), 7.07-7.04 (m, 2H), 3.92 (s, 3H), 3.89 (s, 3H).



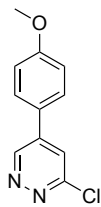
**2-Chloro-4-(4-methoxyphenyl)-1,8-naphthyridine (21b).** Compound **21b** was prepared according to the general procedure using Conditions B, with the modification that catalyst **3f** (9.8 mg, 0.012 mmol, 3.0 mol%) was used instead of **3b**, 2,4-dichloro-1,8-naphthyridine (79.6 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 12 h. Purification by flash column chromatography on silica gel ( $R_f = 0.22$  in 20% pyridine in hexanes) provided **21b** as a white solid (72.1 mg, 67% yield).  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 9.04 (dd,  $J = 4.3, 1.7$  Hz, 1H), 8.28 (dd,  $J = 8.3, 1.7$  Hz, 1H), 7.42 (dd,  $J = 8.3, 4.3$  Hz, 1H), 7.38-7.37 (multiple peaks, 3H), 7.05-7.03 (m, 2H), 3.86 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (151 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 160.7, 156.1, 154.0, 153.7, 152.3, 135.6, 130.8, 127.8, 123.0, 122.2, 120.6, 114.5, 55.5. HRMS (TOF MS ESI+)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{15}\text{H}_{11}\text{ClN}_2\text{O}$  270.0560; Found 270.0567.



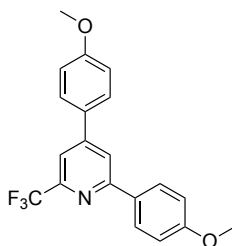
**3-Chloro-5-(4-methoxyphenyl)-4-phenylpyridazine (22b).** Compound **22b** was prepared according to the general procedure using Conditions A, 3,5-dichloro-4-phenylpyridazine (**S22**, 135.0 mg, 0.6 mmol, 1.5 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 22 h. Product **22b** was purified on an automated column using 10 grams of high-capacity silica, a linear gradient of 5-40% acetone in hexanes over 11 column volumes ( $R_f = 0.36$  in 20% acetone in hexanes). Product **22b** was isolated as a white solid (81.5 mg, 69% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 9.12 (s, 1H), 7.34-7.33 (m, 3H), 7.15-7.14 (m, 2H), 7.03-7.01 (m, 2H), 6.77-6.75 (m, 2H), 3.74 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 160.3, 157.0, 151.6, 140.5, 137.7, 133.4, 130.9, 129.7, 128.9, 128.6, 126.0, 114.3, 55.3. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{17}\text{H}_{13}\text{ClN}_2\text{O}$  296.0716; Found 296.0706. The minor product of diarylation **S22c** was also isolated in low purity (10.1 mg, <7% yield,  $R_f = 0.15$  in 20% acetone in hexanes).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 9.15 (s, 1H), 7.28-7.26 (m, 2H), 7.21-7.17 (m, 3H), 7.03-7.01 (m, 2H), 6.93-6.91 (m, 2H), 6.79-6.75 (multiple peaks, 4H), 3.782 (s, 3H), 3.778 (s, 3H).



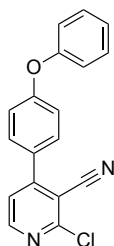
**3-Chloro-5-(4-methoxyphenyl)-4-(1-piperidinyl)pyridazine (23b).** Compound **23b** was prepared according to the general procedure using Conditions A, modified to heat at 66 °C instead of 60 °C, with 3,5-dichloro-4-(1-piperidinyl)-pyridazine (**S23**, 92.8 mg, 0.4 mmol, 1.0 equiv), and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 8 days (however, yields were comparable when the reaction was run for only 3 days). Product **23b** was purified on an automated column using 10 grams of high-capacity silica, a linear gradient of 5-40% acetone in hexanes over 11 column volumes (although **23b** eluted after 13 column volumes) at a flow rate of 40 mL/min ( $R_f = 0.21$  in 20% acetone in hexanes), providing **23b** as a red-brown solid (38.7 mg, 32% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.74 (s, 1H), 7.28-7.26 (m, 2H), 7.00-6.98 (m, 2H), 3.85 (s, 3H), 2.99-2.97 (m, 4H), 1.51-1.47 (m, 2H), 1.43-1.39 (m, 4H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 159.8, 156.5, 149.3, 143.0, 130.7, 126.4, 126.3, 114.4, 55.4, 50.2, 25.5, 23.8. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{16}\text{H}_{18}\text{ClN}_3\text{O}$  303.1138; Found 303.1129.



**3-Chloro-5-(4-methoxyphenyl)pyridazine (24b).** Prior to the reaction, 3,5-dichloropyridazine (purchased as a reddish brown solid) was purified by trituration in pentanes and filtration through a sintered funnel; the filtrate was concentrated and dried furnishing 3,5-dichloropyridazine starting material as a white solid (70% recovery). Compound **24b** was prepared according to the general procedure using Conditions A, purified 3,5-dichloropyridazine (59.6 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 6 h. Purification by flash column chromatography on silica gel ( $R_f = 0.22$  in 15% acetone in hexanes) provided **24b** as a white crystalline solid with minor red-brown discoloration (68.8 mg, 78% yield); mp 94–97 °C [uncorrected, measured against benzoic acid (113–117 °C)].  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 9.24 (d,  $J = 1.5$  Hz, 1H), 7.55–7.54 (multiple peaks, 3H), 6.97 (d,  $J = 7.4$  Hz, 2H), 3.80 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (151 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 161.9, 157.2, 148.5, 140.7, 128.6, 125.0, 123.6, 115.2, 55.5. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{11}\text{H}_9\text{ClN}_2\text{O}$  220.0403; Found 220.0403.



**2,4-Bis-(4-methoxyphenyl)-6-(trifluoromethyl)pyridine (S25c).** Compound **S34c** was prepared according to the general procedure using Conditions B, 2,4-dichloro-6-(trifluoromethyl)pyridine (86.4 mg, 0.4 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (60.8 mg, 0.4 mmol, 1.0 equiv) with stirring for 12 h. Purification by flash column chromatography on silica ( $R_f = 0.34$  in 7% acetone in hexanes) provided **S25c** as a white crystalline solid (61.8 mg, 43% yield); mp 95–98 °C [uncorrected, measured against benzoic acid (109–111 °C)].  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.09–8.06 (m, 2H), 7.96 (d,  $J = 1.3$  Hz, 1H), 7.70 (d,  $J = 1.3$  Hz, 1H), 7.67–7.64 (m, 2H), 7.06–7.00 (multiple peaks, 4H), 3.88 (s, 3H), 3.87 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 161.2, 161.1, 158.1, 150.4, 148.7 (q,  $J_{CF} = 33.9$  Hz), 130.8, 129.9, 128.7, 128.5, 122.0 (q,  $J_{CF} = 274.5$  Hz), 119.4, 115.5 (q,  $J_{CF} = 2.7$  Hz), 114.8, 114.3, 55.54, 55.49.  $^{19}\text{F}$  NMR (471 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -68.0.

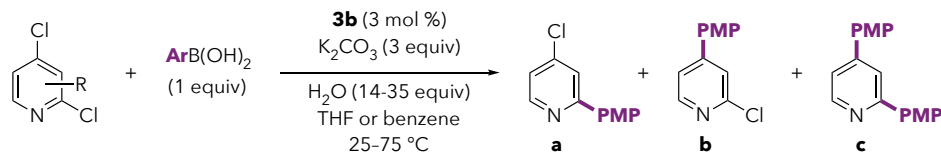


**2-Chloro-4-(4-phenoxyphenyl)-3-pyridinecarbonitrile (39b).** Compound **39b** was prepared according to the general procedure using Conditions A on a 1.0 mmol scale, 2,4-dichloro-3-cyanopyridine (173.0 mg, 1.0 mmol, 1.0 equiv) and *p*-phenoxyphenylboronic acid (214.0 mg, 1.0 mmol, 1.0 equiv) with stirring for 19 h. Purification by flash column chromatography on silica gel ( $R_f = 0.26$  in 10% acetone in hexanes) provided **39b** as a white solid (188 mg, 61% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.52 (d,  $J = 5.2$  Hz, 1H), 8.59–8.56 (m, 2H), 7.41–7.37 (multiple peaks, 3H), 7.20–7.17 (m, 1H), 7.12–7.08 (multiple peaks, 4H). Spectral data are consistent with the literature.<sup>17</sup>

#### 4. Discussion About Mass Balance

The moderate isolated yields in Scheme 2 (ligand-controlled conditions) are generally due to a combination of factors: (1) unreacted starting material, (2) formation of diarylated product, and (3) material loss during purification of the target product by column chromatography due to partial coelution of regioisomers.

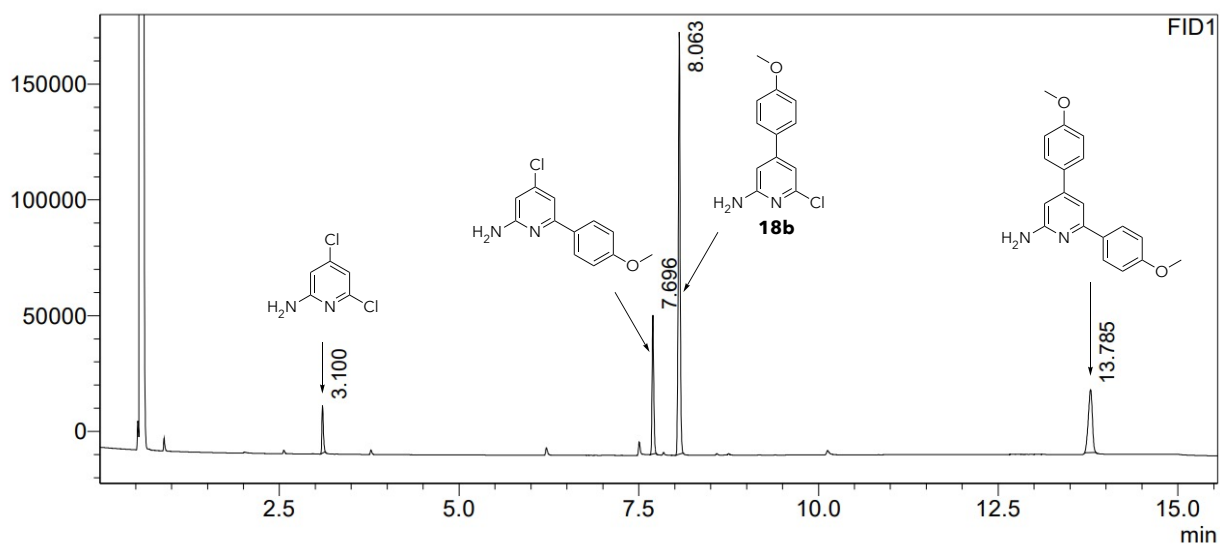
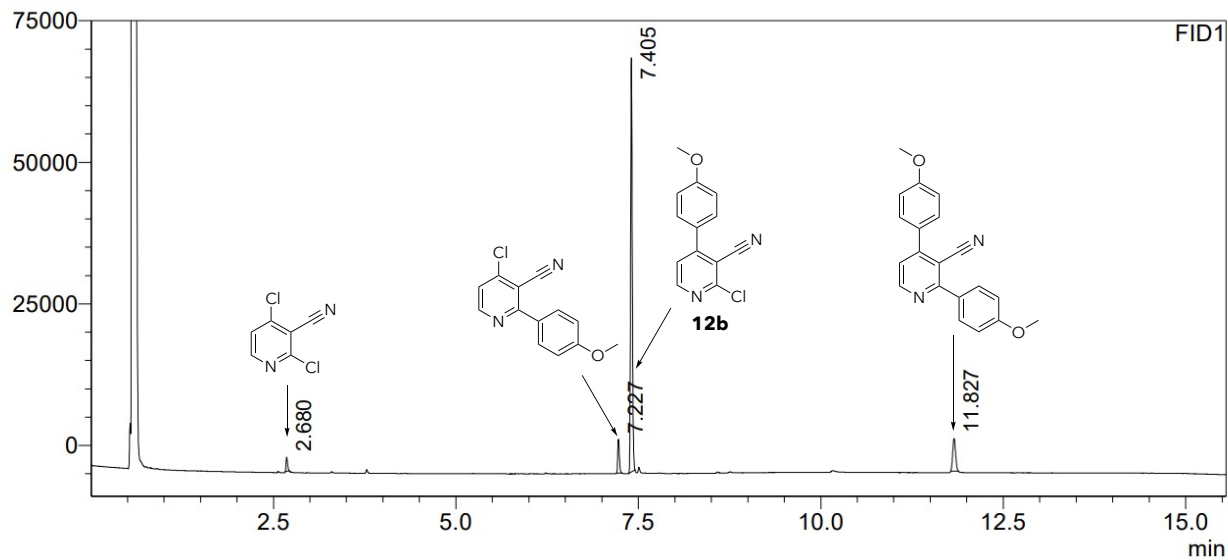
In several examples above, we report isolated yields of the minor regioisomer as well as the diarylated regioisomer. These isolated yields are summarized below. In some cases, the mass balances are above 90%. In cases with lower mass balance, there was significant material loss during isolation of products.



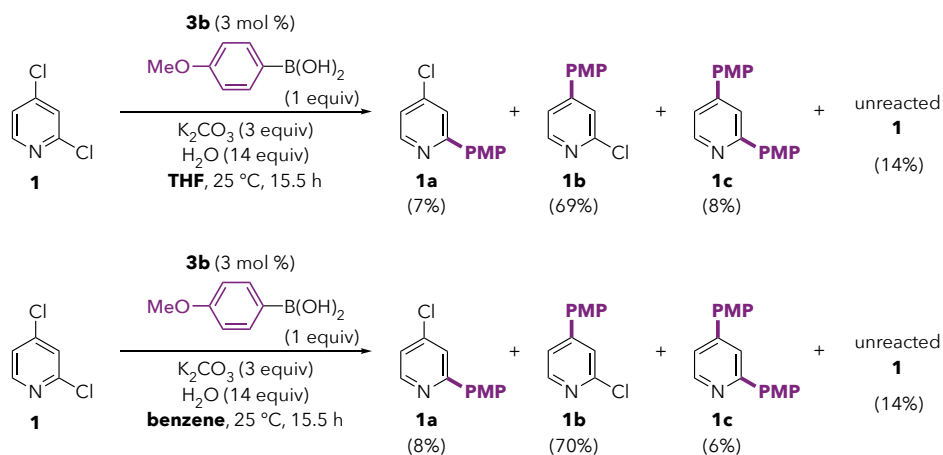
Substrate	ArB(OH) <sub>2</sub>	estimated unreacted substrate based on crude GC (uncalibrated, %) <sup>a</sup>	<b>a</b> (%)	<b>b</b> (%)	<b>c</b> (%)	Sum of substrate + <b>a</b> + <b>b</b> + <b>c</b> (%)
		6	9	71 ( <b>6b</b> )	13	99
<b>1</b>		2	13	67 ( <b>10b</b> )	9	91
		2	6 <sup>b</sup>	68 ( <b>12b</b> )	3	79
		6	13	60 ( <b>14b</b> )	14	93
		11	2 <sup>b</sup>	53 ( <b>15b</b> )	18	84
		4	13	48 ( <b>18b</b> )	16	81

<sup>a</sup>Yield calculated by extrapolation from the isolated yield of the C4 product (**b**) and the ratio of C4 product to remaining starting material signals by GC analysis of the crude reaction mixture. <sup>b</sup>Yield calculated by extrapolation from the isolated yield of the C4 product (**b**) and the ratio of C4:C2 product signals by GC analysis of the crude reaction mixture.

The GC chromatograms below illustrate the crude reaction mixtures prior to purification for two of the reactions from the table above with lower mass balance (reactions to form **12b** and **18b**). There are no significant unidentified peaks.



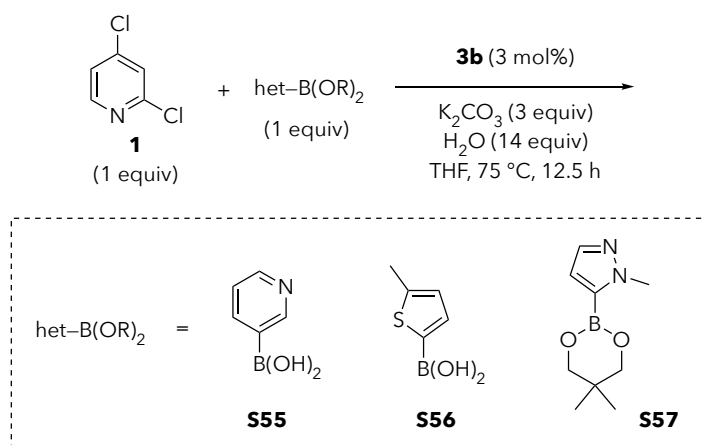
In the optimization table (Table 1), a standard was added to the reactions and calibrated GC yields were obtained. We generally see good mass balance accounted for by major and minor regioisomers, unreacted starting material, and diarylation products. Examples are illustrated below, corresponding to entries 3 and 5 of Table 1:

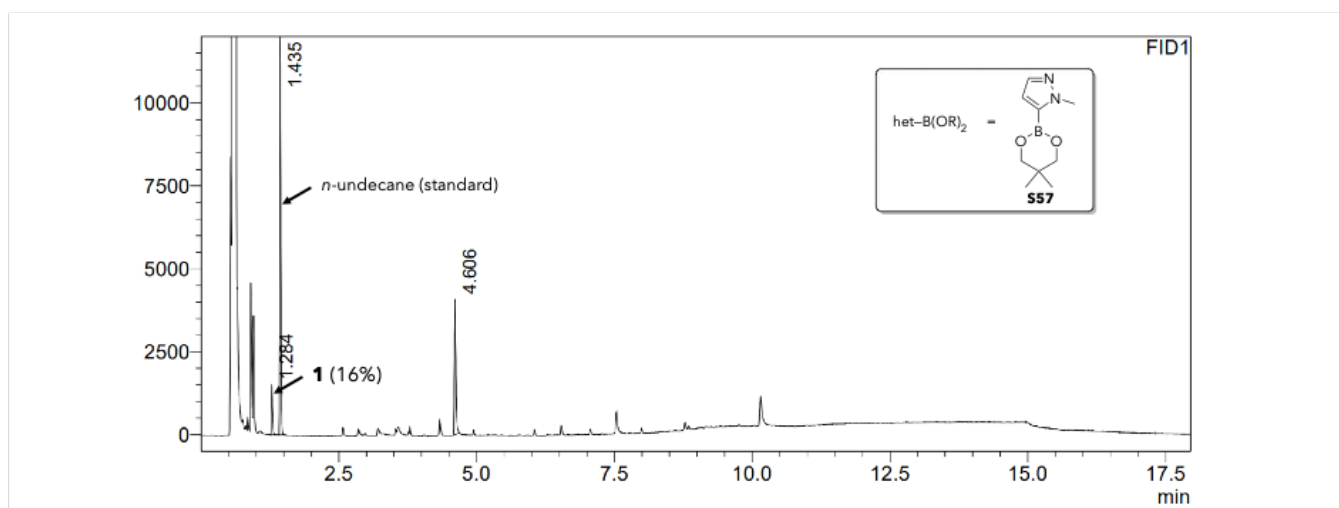
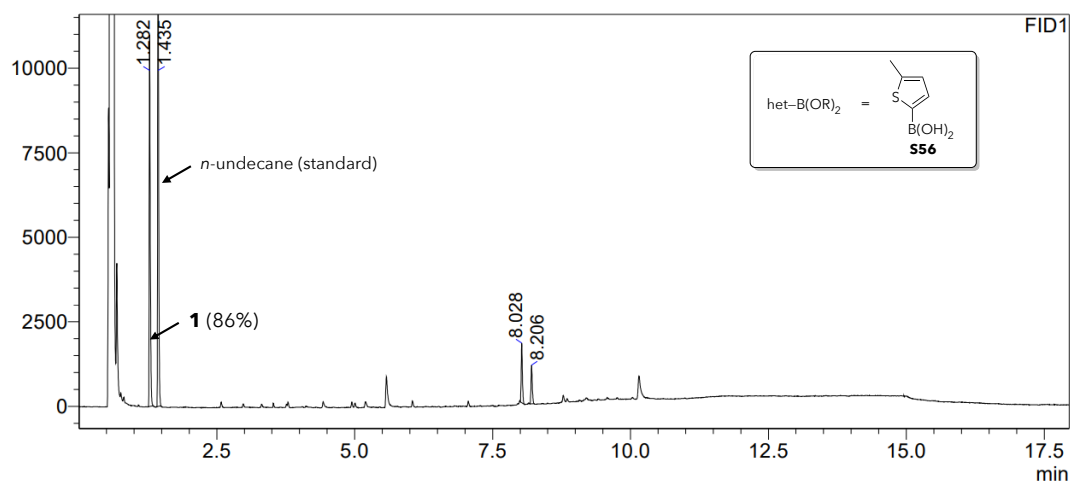
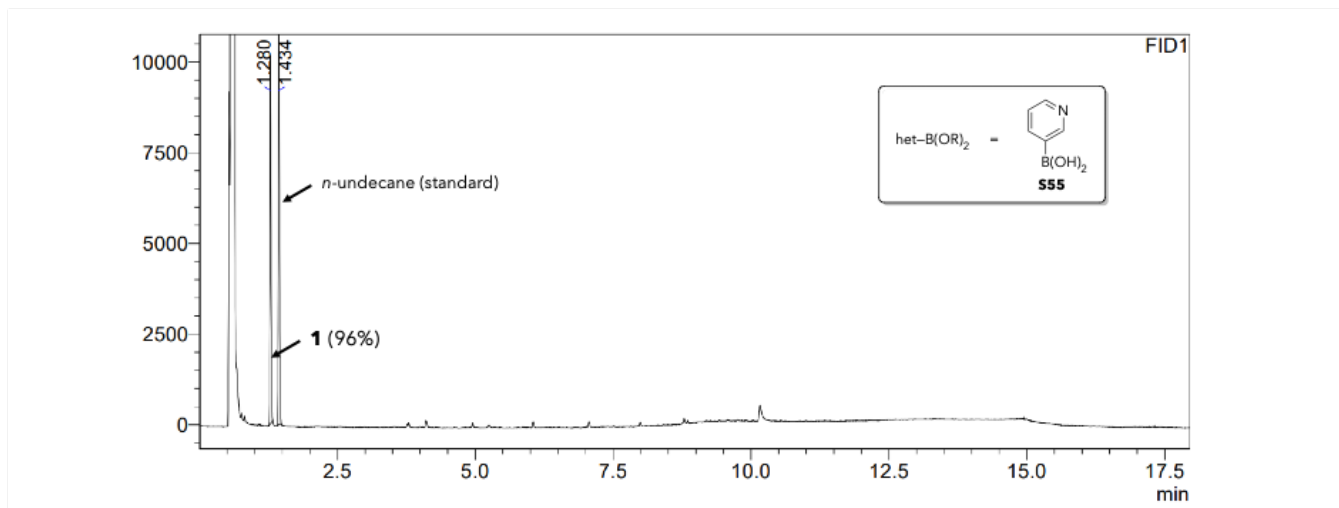


## 5. Scope Limitations of the Suzuki Coupling

**Boron Nucleophile Limitations.** Heteroaryl boronic acids and esters **S55-S57** were ineffective in the Suzuki cross-coupling with **1** under the optimized conditions using catalyst **3b** (General Procedure for GC-Scale Reactions) at room temperature or even at a higher temperature (75 °C, below). For **S55** and **S56**, most of the starting material **1** remained after the reaction time. For **S57**, most of the starting material was consumed, but only small amounts of products were detected by GC. The GC chromatograms of these crude reactions are depicted below.

**Scheme S3.** Boron Nucleophile Limitations





**Substrate Limitations: Overview.** Some substrates did not provide the desired cross-coupling selectivity. These substrates are categorized below as 2,4-dihalopyridines that lead to overarylation (**25** and **26**), brominated 2,4-



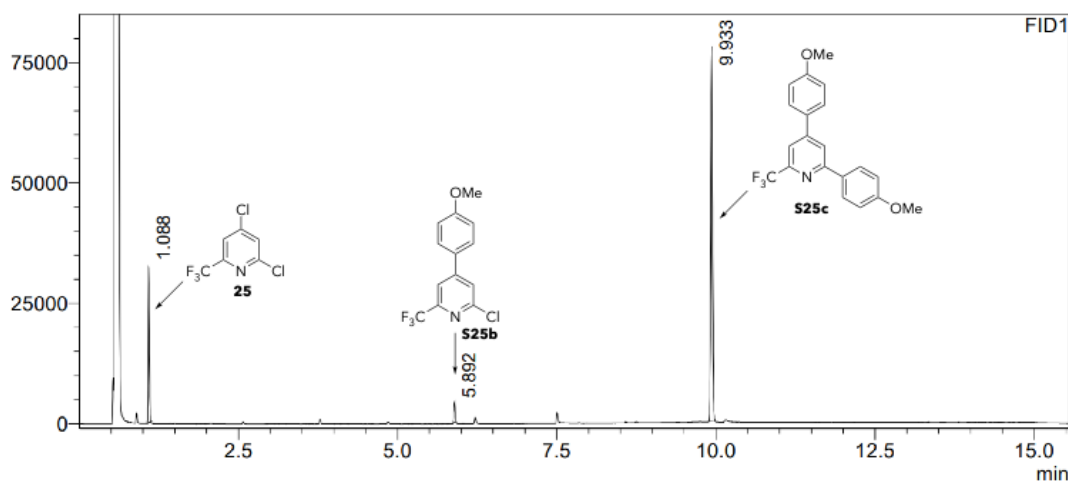
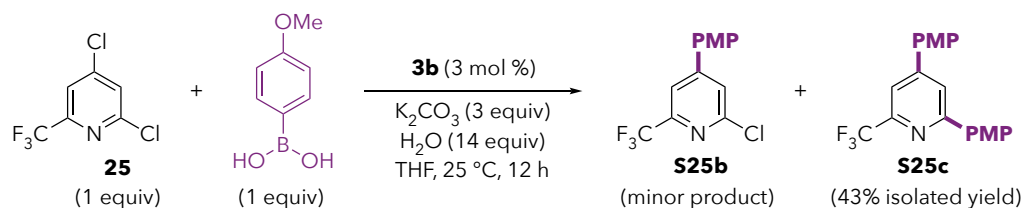
dichloropyridine or quinolines that react at bromide instead of chloride (**27** and **28**), and 2,5-dichloropyridine and pyrimidine (**29** and **30**), substrates for which the Pd/IPr catalytic system provides conventional selectivity next to nitrogen. The reactions below were conducted according to the General Procedure for GC-Scale Reactions.

**Substrates Resulting in Overarylation.** Reactions of both substrates **25** and **26** afford diarylated product **S25c** and **1c** as the major products by crude GC analysis.

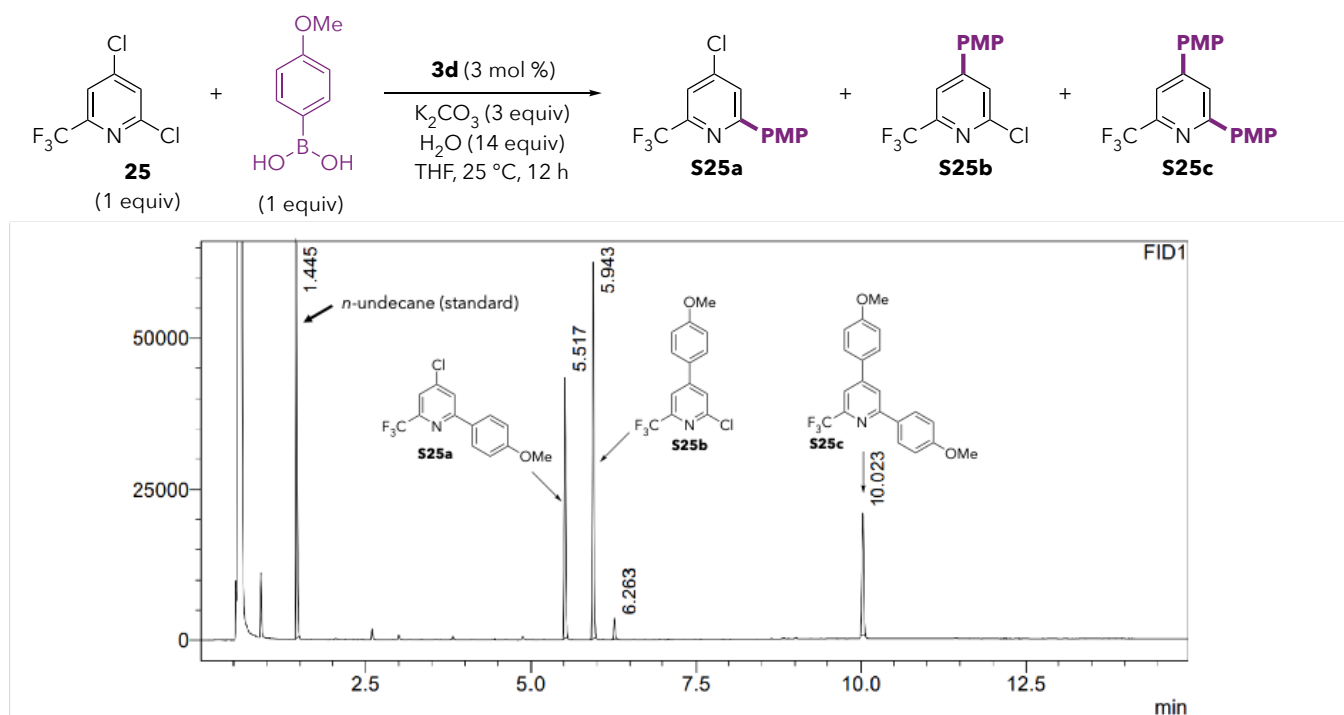
For the reaction of **25** with catalyst **3b**, a small amount of C4-monarylated product **S25b** is detected by crude GC, but the major product is **S25c** from diarylation (Scheme S4). Product **S25c** was isolated and characterized (*vide supra*). The monoarylated product was not isolated, but its identity was tentatively assigned on the basis of its mass (by GCMS analysis) and inference based on the usual selectivity exhibited by **3b** (Pd/IPr, which favors reaction at C4 with other 2,4-dichloropyridines) and **3d** (Pd/IMes, which gives nearly a 1:1 mixture of C2- and C4-arylated products with other 2,4-dichloropyridines). When **3d** (Pd/IMes) is used as the catalyst for the Suzuki coupling of **25**, much less diarylation is observed, and two monoarylated products are formed in a 1.5:1 ratio (assigned as **S25b** and **S25a**, Scheme S5). Notably, only one of these two monoarylated products is observed in the reaction using catalyst **3b**, and based on the usual selectivity with **3b** we infer that this product is **S25b**.

Dibromo substrate **26** undergoes extensive diarylation using catalyst **3b** (Scheme S6). Product **1c** has been isolated and characterized (*vide supra*), and the identities of the minor monoarylated products **S26a** and **S26b** are supported by GCMS analysis. The regiochemistry of **S26a** and **S26b** was inferred based on the usual selectivity exhibited by **3b** (Pd/IPr, which favors reaction at C4 with 2,4-dichloropyridines) and subsequently confirmed using C4-selective cross-coupling conditions recently reported by Fairlamb<sup>18</sup> (*vide infra*).

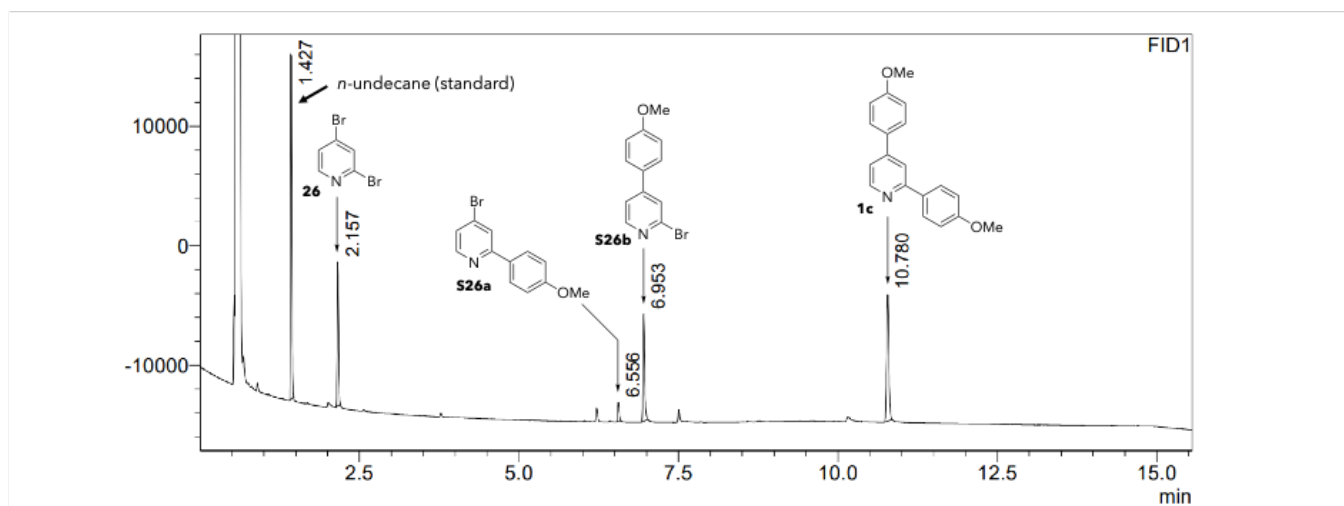
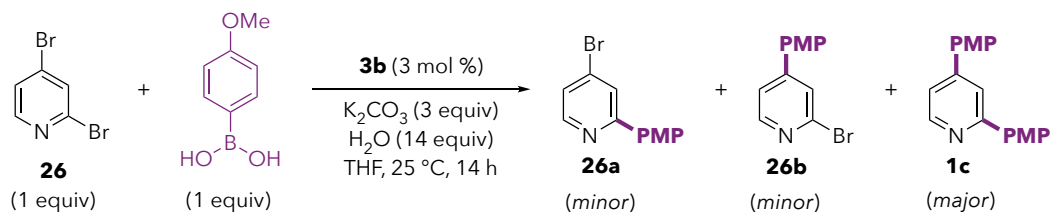
**Scheme S4.** Suzuki reaction of **25** using Pd/IPr catalyst **3b**.



**Scheme S5.** Suzuki reaction of **25** using Pd/SIMes catalyst **3d**.



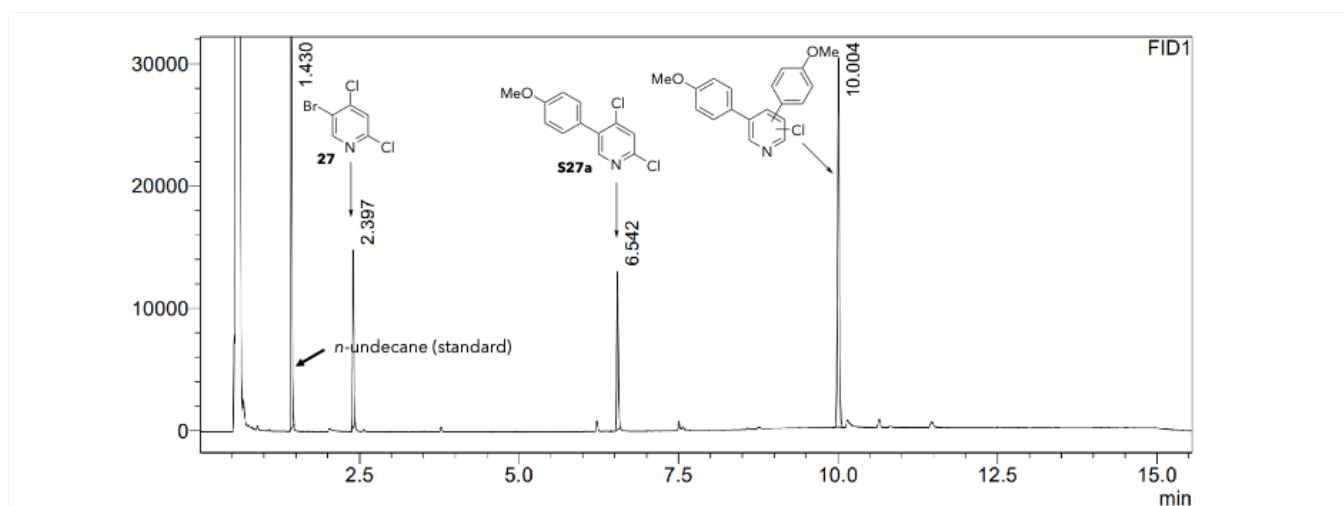
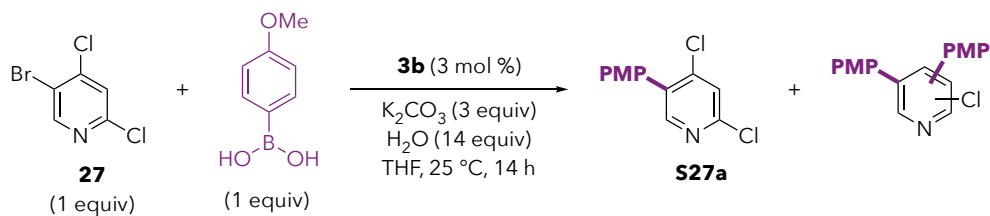
**Scheme S6.** Suzuki reaction of **26** using Pd/IPr catalyst **3b**.



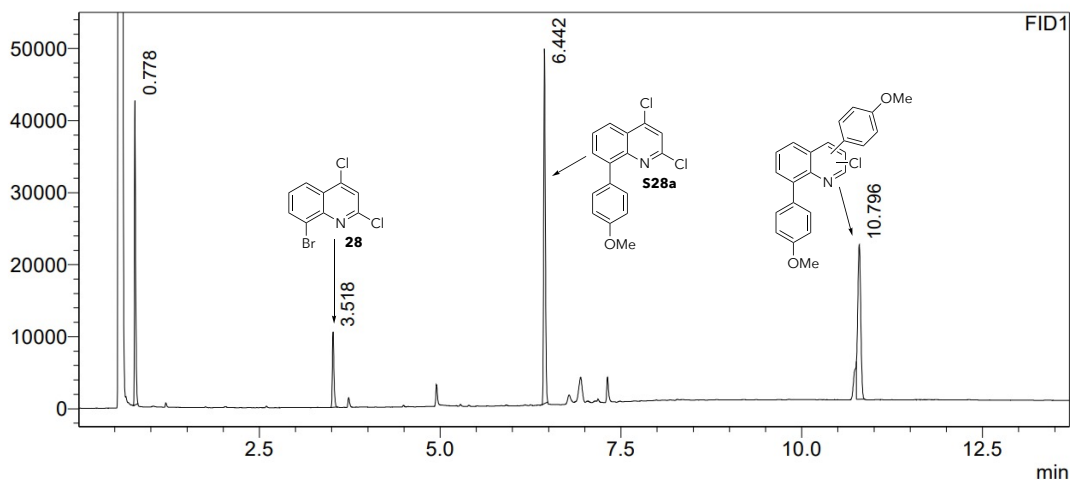
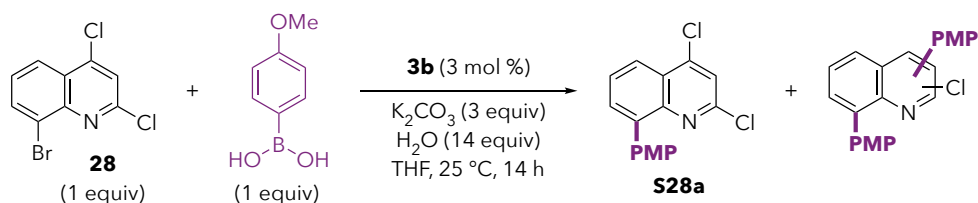
*Substrates Resulting in Reaction at Bromide.* Substrates **27** and **28** undergo extensive reaction at bromide under the optimized conditions, providing a mixture of dichloro-monoarylated products **S27a** and **S28a** and

diarylated products resulting from reaction at bromide and one of the chlorides (Schemes S7-S8). Product identities were assigned based on GCMS analysis.

**Scheme S7.** Suzuki reaction of **27** using Pd/IPr catalyst **3b**.

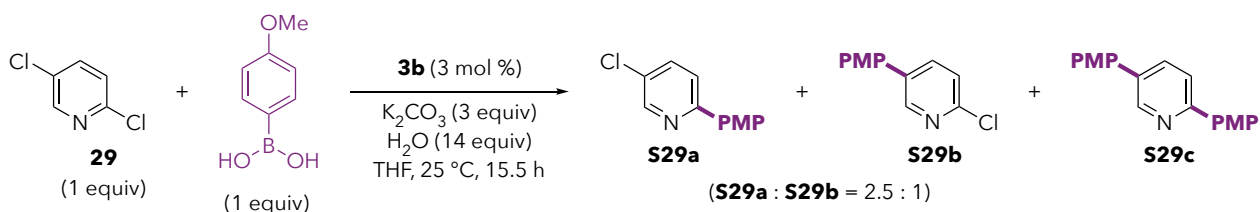


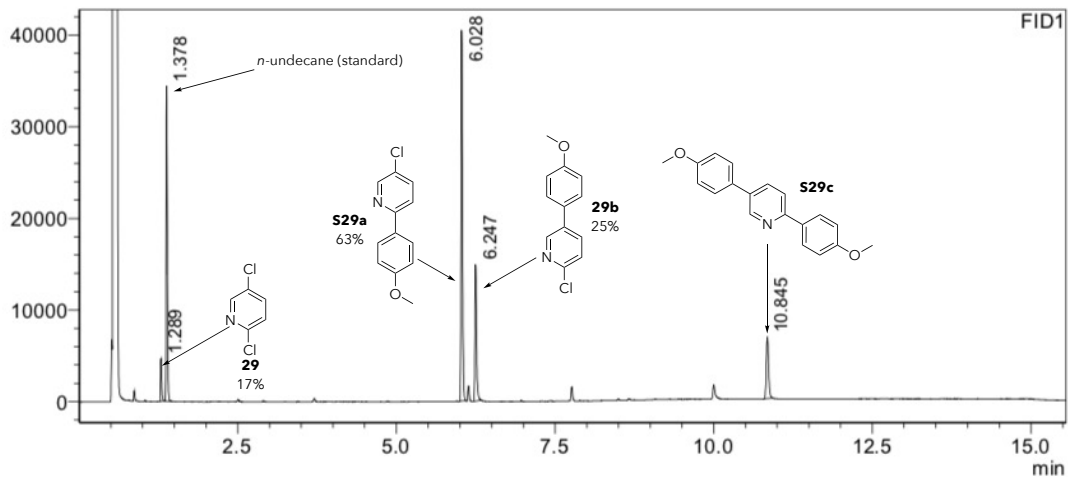
**Scheme S8.** Suzuki reaction of **28** using Pd/IPr catalyst **3b**.



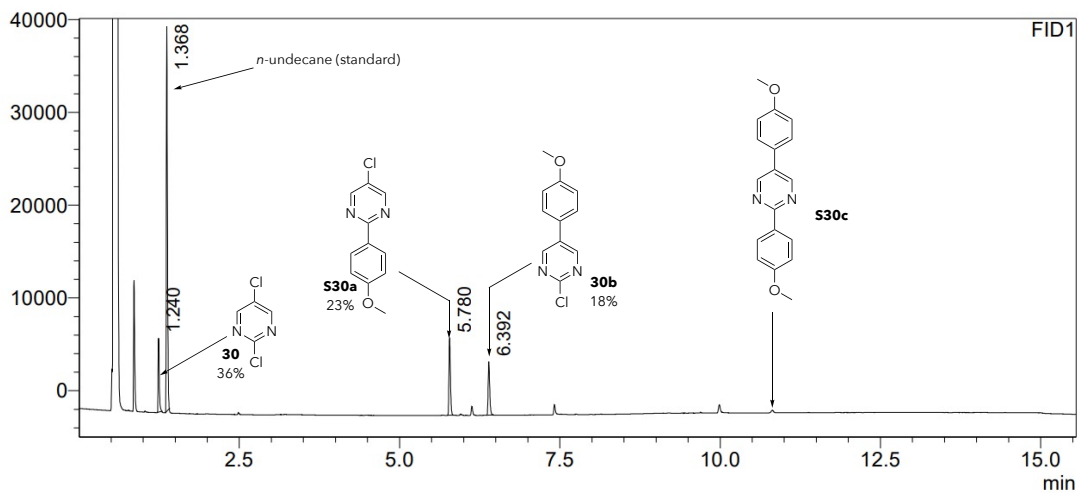
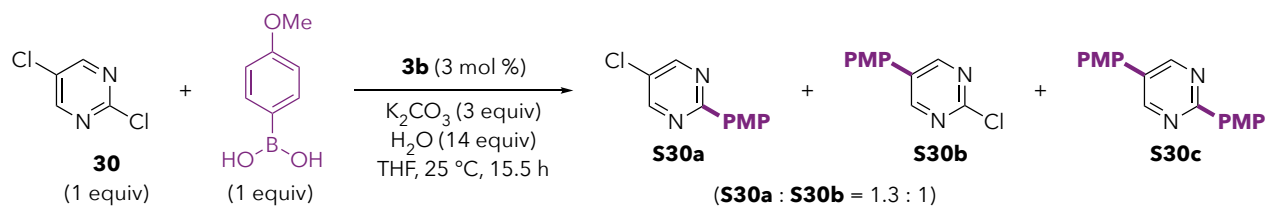
Substrates Resulting in Cross-Coupling at the Conventional Site Next to Nitrogen. 2,5-Dichloropyridine (**29**) and 2,5-dichloropyrimidine (**30**) favor reaction at C2 under the optimized system according to crude GC and GCMS analysis (ratio of **S29b** to **S29a** = 1 : 2.5, **S30b** to **S30a** = 1 : 1.3 based on the ratio of signal integrations by GC, Schemes S9-S10). As such, the use of catalyst **3b** does not enable unconventional selectivity with this substrate. The identities of the monoarylated products were assigned by a combination of GCMS analysis and comparison to the results obtained with a  $Pd(OAc)_2/dppf$  catalytic system, a system that normally favors conventional C2-selectivity in the reaction of other dichlorinated pyridines.<sup>10</sup> The identity of the diarylated products **S29c** and **S30c** were assigned based on their masses obtained by GCMS analysis.

**Scheme S9.** Suzuki reaction of **29** using Pd/IPr catalyst **3b**.





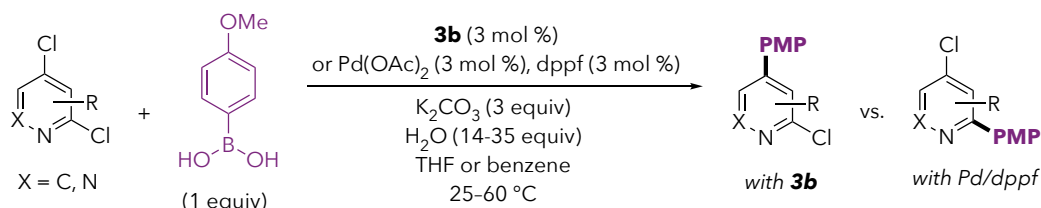
**Scheme S10.** Suzuki reaction of **30** using Pd/IPr catalyst **3b**.



## 6. Comparison of Results with IPr (C4-Selective) vs. dppf (C2-Selective)

C4-selectivity remains ligand-controlled for the substrates depicted in Scheme 2, as evidenced by the observation of different major products when using catalyst **3b** (ligand = IPr) versus using Pd(OAc)<sub>2</sub>/dppf (Table S2).

**Table S2.** Retention time (RT) of major products by GC-FID for reactions catalyzed by **3b** or by Pd(OAc)<sub>2</sub>/dppf.



entry	substrate	<b>3b</b> (min)	Pd/dppf (min)
1	<b>S11</b>	7.11	7.21
2	<b>S13</b>	7.21	7.16
3	<b>S14</b>	7.25	7.02
4	<b>S15</b>	6.52	6.68
5	<b>S16</b>	7.81	7.88
6	<b>S18</b>	8.03	7.69
7	<b>S21</b>	7.10	7.20
8	<b>S22</b>	6.79	6.48
9	<b>S23</b>	7.62	7.98
10	<b>S24</b>	7.22	6.62

## D. Kumada Cross Couplings with Pd/IPr

### 1. General Procedures

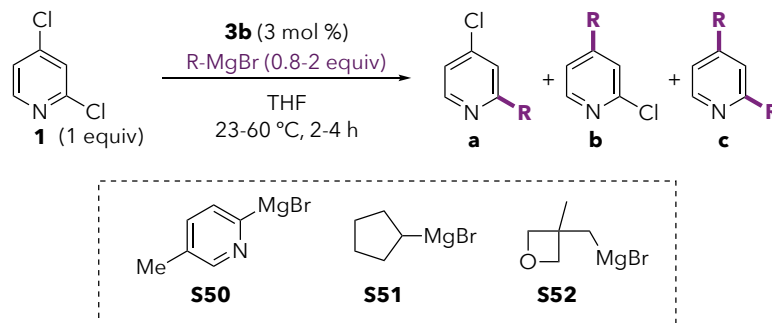
**GC-Scale Reactions.** The precatalyst **3b** (1.7 mg, 0.0024 mmol, 3.0 mol%) was added to an oven-dried 1-dram vial equipped with a stir bar. The vial was transferred to a N<sub>2</sub> atmosphere glovebox, and the Grignard reagent (0.77-2 equiv) was added as a solution in THF followed by 2,4-dichloropyridine (0.08 mmol, 1 equiv). The vial was sealed with a PTFE-lined cap and removed from the glovebox. The reaction was stirred for 2-4 hours at 23-60 °C. The reaction mixture was opened to air and quenched with EtOAc (~3.2 mL), and the internal standard undecane was added (7.5 μL, 0.036 mmol). The vial was shaken to mix the contents, and the mixture was filtered through celite prior to analysis by GC and GCMS.

**Scaled-Up Reactions for Product Isolation.** The precatalyst **3b** (8.4 mg, 0.012 mmol, 3.0 mol%) was added to an oven-dried 1- or 5-dram vial equipped with a stir bar. The vial was transferred to a N<sub>2</sub> atmosphere glovebox, and the Grignard reagent (0.77-2 equiv) as a solution in THF was added followed by the addition of 2,4-dichloropyridine (1 equiv). The vial was sealed with a PTFE-lined cap and removed from the glovebox. The reaction was stirred for 2-5 hours at 23 °C, then the vial was opened to air. The reaction mixture was diluted with EtOAc and washed with sat. aqueous NH<sub>4</sub>Cl (3 x 10 mL), sat. aqueous NaHCO<sub>3</sub> (3 x 10 mL), and brine (2 x 10 mL) in a separatory funnel. The organic layer was extracted with additional EtOAc (3 x 10 mL) and the combined organic layers were dried over anhydrous MgSO<sub>4</sub>. The crude reaction mixture was analyzed by GC, concentrated under vacuum, and purified by normal or reversed phase flash column chromatography.

## 2. Optimization

Kumada reactions were performed according to the general procedure for GC-scale reactions.

**Table S3.** Optimization of Kumada Cross-Couplings

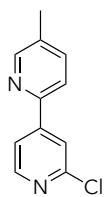


entry	T (°C)	time (h)	R-MgBr (equiv)	<b>a</b> : <b>b</b> <sup>a</sup>	<b>1</b> (%) <sup>b</sup>
1	23	2	<b>S50</b> (1)	<1 : 100	16
2 <sup>c</sup>	23	2	<b>S50</b> (1)	-- <sup>d</sup>	100
3 <sup>e</sup>	23	2	<b>S50</b> (1)	-- <sup>d</sup>	72
4	23	3	<b>S51</b> (1)	1 : 8	5
5	23	3	<b>S51</b> (0.8)	1 : 7	13
6 <sup>c</sup>	23	3	<b>S51</b> (1)	-- <sup>d</sup>	56
7	23	3	<b>S52</b> (1)	<1 : 39	30
8	23	3	<b>S52</b> (2)	<1 : 60	7
9 <sup>c</sup>	23	3	<b>S52</b> (1)	-- <sup>d</sup>	98
10 <sup>f</sup>	60	4	<b>S52</b> (1)	-- <sup>d</sup>	64

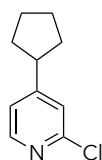
<sup>a</sup> Ratios based on the ratio of signals for **a** and **b** in the GC chromatogram. <sup>b</sup> GC yield of recovered starting material calibrated against undecane. In entry 3, the starting material was 2-bromo-4-chloropyridine instead of **1**. <sup>c</sup> No catalyst (metal free control). <sup>d</sup> "--" means no cross-coupled products **a** or **b** were observed. <sup>e</sup> 2-Bromo-4-chloropyridine was used instead of **1** in combination with Pd(OAc)<sub>2</sub> (5 mol %) and dppe (5 mol %) as the catalytic system instead of **3b**. <sup>f</sup> Pd(OAc)<sub>2</sub> (5 mol %) and dppe (5 mol %) was used as the catalytic system instead of **3b**.

**Discussion:** Catalyst-free controls using reagents **S50–S52** (Table S3, entries 2, 6, 9) show no formation of products **a** or **b**, indicating that there is no background S<sub>N</sub>Ar reactivity. Entries 1, 5, and 8 represent the optimized conditions that were used for scale-up reactions. Interestingly, products **a** and **c** were not observed in the cross couplings with **S50** and **S52**. The use of 2-bromo-4-chloropyridine as the substrate for reaction with **S50** (entry 3), a substrate that should be strongly biased toward reaction at C2, also did not lead to any detectable cross-coupled products. Reaction of the pyridyl Grignard **S50** at C2 would lead to a 2,2'-bipyridine derivative which can chelate to Pd or Mg, providing a compound that is insoluble or not volatile and therefore invisible by GC-FID. Similarly in the case of Grignard **S52**, product **a** is not obtained even when using Pd(OAc)<sub>2</sub>/dppe (entry 10), although considerably more starting material was consumed. It is possible that the C2-functionalized product **a** resulting from reaction of Grignard **S52** is prone to decomposition.

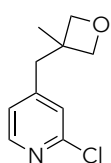
### 3. Isolation and Characterization of Kumada Products from Scheme 3



**2-Chloro-5'-methyl-4,2'-bipyridine (31b).** Product **31b** was prepared according to the general procedure using **3b** (8.4 mg, 0.012 mmol, 3 mol %), 5-methyl-2-pyridylmagnesium bromide (**S50**, 1.6 mL of a 0.25 M solution in THF, 0.40 mmol, 1.00 equiv), and **1** (43.2  $\mu$ L, 0.40 mmol, 1.00 equiv). The reaction was stirred at 23 °C for 2 h. The crude GC chromatogram indicated the presence of unreacted **1**. Purification was performed by flash column chromatography ( $R_f$ = 0.46 in 90:10 Hex:EtOAc) yielded a white solid (49.7 mg, 61% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.58-8.61 (m, 1H), 8.49 (d,  $J$  = 5.2 Hz, 1H), 7.96-7.99 (m, 1H), 7.82 (dd,  $J$ = 5.2, 1.5 Hz, 1H), 7.72 (d,  $J$  = 8.0 Hz, 1H), 7.65 (dd,  $J$  = 8.0, 1.9 Hz, 1H), 2.44 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 152.7, 151.0, 150.8, 149.7, 137.8, 134.6, 121.7, 120.7, 119.8. HRMS (ESI<sup>+</sup>)  $m/z$ :  $[\text{M}]^+$ : Calcd for  $\text{C}_{11}\text{H}_9\text{ClN}_2$  204.0454; Found 204.0455. The C2 monoarylated isomer **S31a** has been reported in previous literature, and the signals for product **31b** are not consistent with those reported for **S31a**.<sup>19</sup>



**2-Chloro-4-cyclopentylpyridine (32b).** Product **32b** was prepared according to the general procedure using **3b** (8.4 mg, 0.012 mmol, 3 mol %), cyclopentylmagnesium bromide (**S51**, 1.34 mL of a  $0.299 \pm 0.004$  M solution in THF, 0.40 mmol, 0.77 equiv), and **1** (56.2  $\mu$ L, 0.52 mmol, 1.00 equiv). The reaction was stirred at 23 °C for 4 h. The crude GC chromatogram indicated the presence of unreacted **1**. Purification was performed by reversed phase flash column chromatography with a 12 g C18 silica column, using an initial automated program comprising a flow rate of 30 mL/min of water:acetonitrile (98:2 to 0:100) over 31 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 58-69% MeCN in water over 11 column volumes. After the elution, the initial program was resumed. The C4 monoarylated product **32b** eluted at 61-64% MeCN. Fractions containing **32b** were partitioned between dichloromethane and saturated brine. The organic layers were combined and dried over magnesium sulfate and solvent was removed under reduced pressure affording **32b** as a light brown oil (50.8 mg, 62% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.24 (d,  $J$  = 5.2 Hz, 1 H), 7.17 (s, 1H), 7.06 (d,  $J$  = 5.2 Hz, 1H), 2.92-3.01 (m, 1H), 2.02-2.12 (m, 2H), 1.75-1.85 (m, 2H), 1.65-1.75 (m, 2H), 1.51-1.61 (m, 2H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 159.3, 151.7, 149.5, 123.1, 121.6, 45.1, 34.0, 25.6. HRMS (ESI<sup>+</sup>)  $m/z$ :  $[\text{M}]^+$ : Calcd for  $\text{C}_{10}\text{H}_{12}\text{ClN}$  181.0658; Found 181.0666.



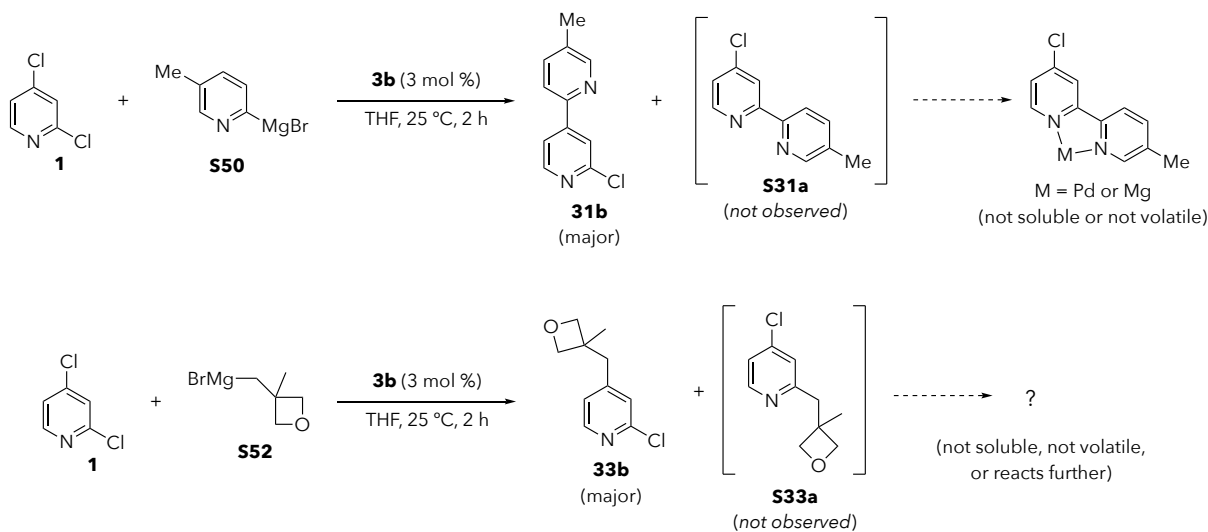
**2-Chloro-4-(3,3'-methyloxetanyl)pyridine (33b).** Product **33b** was prepared according to the general procedure using **3b** (8.4 mg, 0.012 mmol, 3 mol %), 3,3'-methyloxetanylmagnesium bromide (**S52**, 5.71 mL of a 0.14 M solution in THF, 0.80 mmol, 2.0 equiv), and **1** (43.2  $\mu$ L, 0.40 mmol, 1.00 equiv). The reaction was stirred at 23 °C for 3 h. Purification was performed by reversed phase flash column chromatography with a 12 g C18 silica column, using an initial automated program comprising a flow rate of 25 mL/min of water:acetonitrile (98:2 to 8:92) over 21 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 46-49% MeCN in water. After the elution, the initial program was resumed. The C4 monoarylated product **33b** coeluted with an aliphatic impurity at 49% MeCN. Fractions containing **33b** were partitioned between ethyl acetate and saturated brine. The organic layers were combined and dried over magnesium sulfate and solvent was removed under reduced pressure. The aliphatic impurity was removed *in vacuo* yielding an off white colored solid (46.5 mg, 59% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.30 (d,



$J = 5.0$  Hz, 1H), 7.11 (s, 1H), 6.99 (d,  $J = 5.0$  Hz), 4.59 (d,  $J = 5.8$  Hz, 2H), 4.37 (d,  $J = 5.8$  Hz, 2H), 2.96 (s, 2H), 1.28 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 151.8, 150.5, 149.6, 125.0, 123.4, 82.7, 82.1, 43.9, 39.7, 23.3 ppm. HRMS (ESI<sup>+</sup>)  $m/z$ :  $[\text{M}]^+$ : Calcd for  $\text{C}_{10}\text{H}_{12}\text{ClNO}$  197.0607; Found 197.0599.

#### 4. Discussion about Decomposition of Putative Products **S31a** and **S33a**

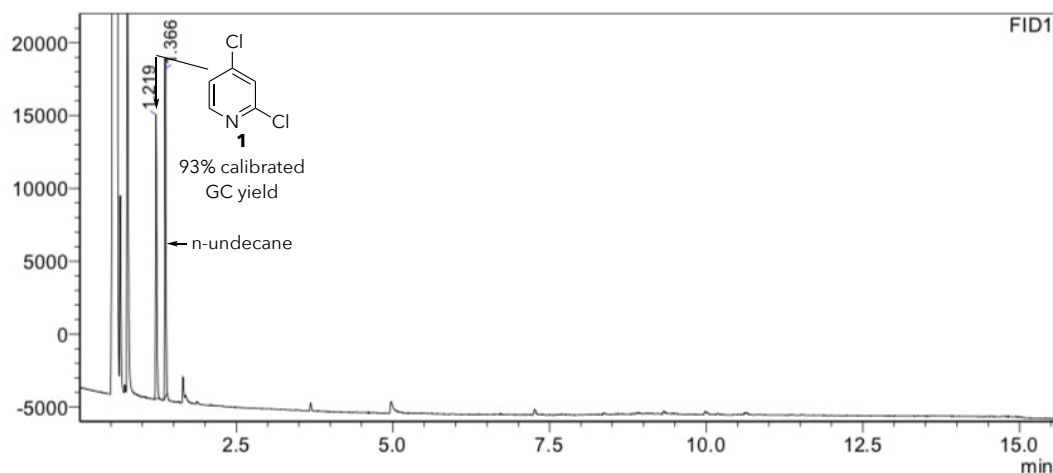
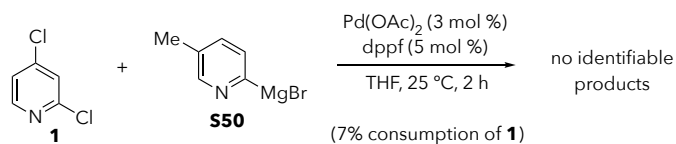
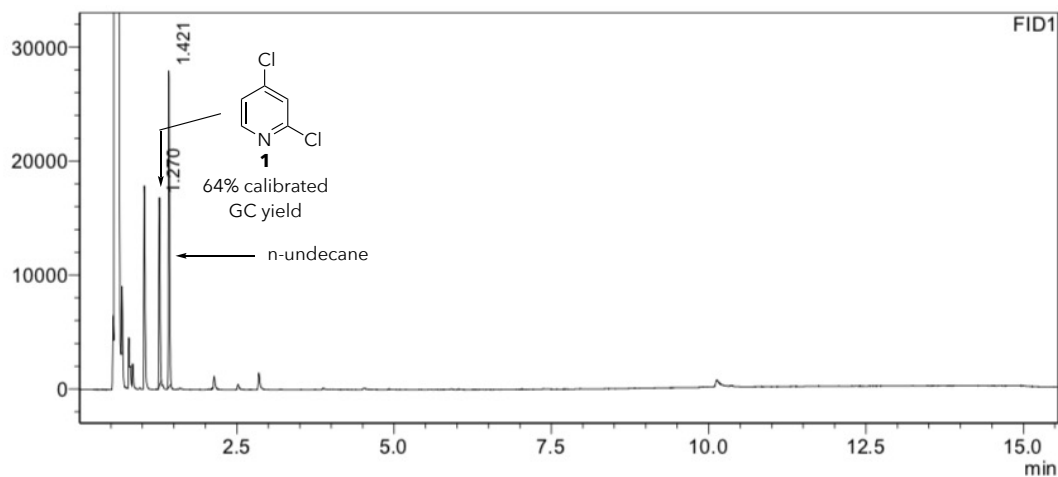
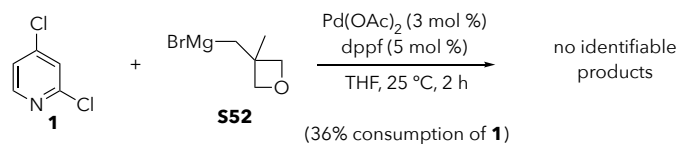
Although typical C4:C2 selectivity for Pd/IPr-catalyzed Suzuki, Kumada, and Negishi couplings of **1** is about 10:1 (range 7:1 to 13:1 for the reactions reported in the manuscript), the reactions of **1** with a pyridyl or a methyloxetanyl Grignard reagent (**S50** and **S52**) lead to the C4-functionalized products **31b** and **33b** as the only products observed. We hypothesize that this apparently high selectivity is misleading, and the selectivity in these reactions is likely similar to the other Pd/IPr-catalyzed reactions in the manuscript. It is likely that some amount of C2-functionalized products (**S31a** and **S33a**) are formed, but these products undergo further reaction in a way that makes them undetected by crude GC-FID, GC-MS, or NMR due to poor volatility or solubility. The hypothesized decompositions are represented below:



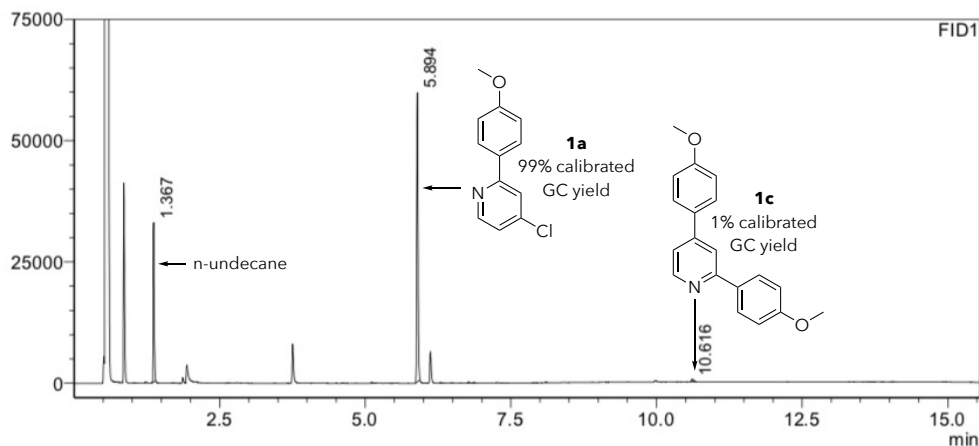
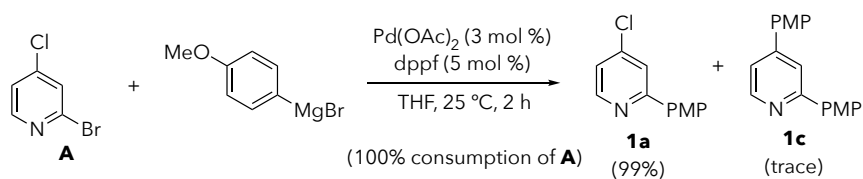
Evidence to support the hypothesis that **S31a** and **S33a** (but not other C2-functionalized products) undergo decomposition includes the following:

- (1) With the exception of the reactions with **S50** and **S52**, good mass balance is seen in the other Kumada couplings, as well as in the Suzuki and Negishi couplings (see Sections I.C.4 and I.G.6). As an example, the following reaction was performed, affording known products that have all been previously isolated and calibrated against an internal standard for GC-FID analysis. As illustrated, there is no evidence for decomposition of the C2-functionalized product **1a**. The C4:C2 product ratio is 11:1, which is comparable to other cross-couplings performed with substrate **1** using Pd/IPr. The mass balance in the crude GC chromatogram adds up to 100% (estimated error is  $\pm 3\%$ ), indicating that all material is accounted for.

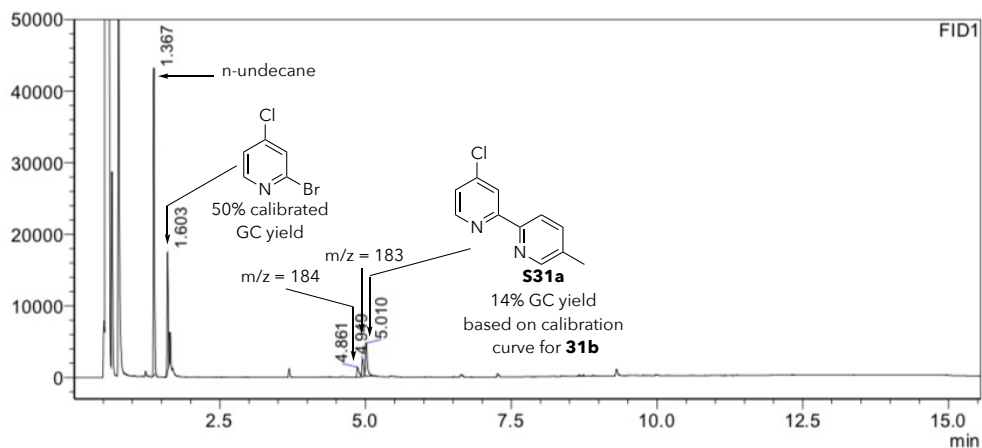
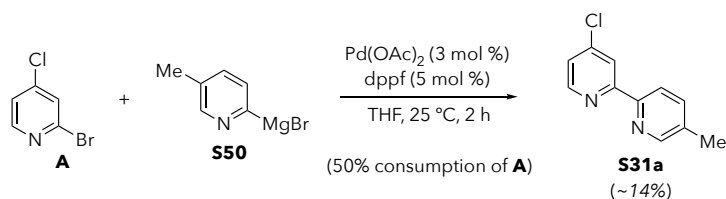


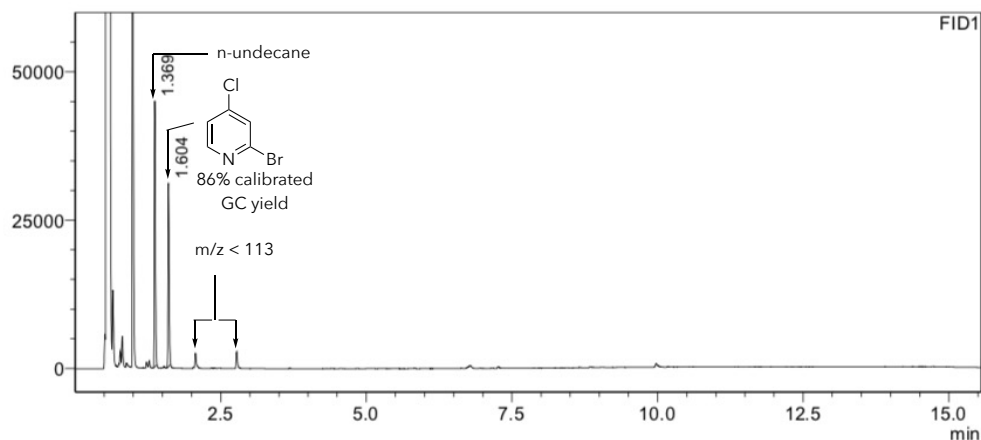
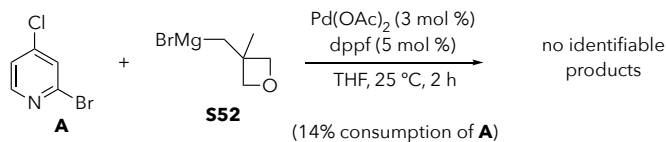


(3) The use of the Pd/dppf catalytic system for the cross-coupling of 2-bromo-4-chloropyridine would be expected to lead to improved cross-coupling at C2, because this substrate has enhanced reactivity at C2 due to the 2-Br substituent. Indeed, excellent mass balance is observed in the reaction between 2-bromo-4-chloropyridine and a simple aryl Grignard reagent:



However, the use of this system for the reactions with Grignard reagents **S50** and **S52** leads to none or only a small amount of detectable C2-functionalized products, despite partial consumption of the starting material. These results are consistent with conversion of the C2-functionalized products into materials that are not identified by crude GC-FID, GC-MS, or NMR due to poor solubility or volatility. We are continuing to explore the nature of these decomposition pathways in our lab.





## E. Negishi Cross-Couplings with Pd/*IPr*

### 1. General Procedures

**GC-Scale Reactions.** In a glovebox, Grignard reagent (0.77-1 equiv as a solution in THF) and a solution of  $\text{ZnCl}_2$  in THF (1 M, equimolar with the Grignard reagent) were combined in an oven-dried 1-dram vial ("vial A") equipped with a stir bar and stirred for at least 0.5 hours at ambient temperature. Outside of the glovebox, the precatalyst **3b** (1.7 mg, 0.0024 mmol, 3.0 mol %) was added to a separate oven-dried 1-dram vial equipped with a stir bar ("vial B"), and this vial was transferred into the glovebox. The contents of vial A, comprising the organozinc reagent (0.77-1 equiv), and 2,4-dichloropyridine (0.08 mmol, 1 equiv) were added to vial B. Vial B was sealed with a PTFE-lined cap and removed from the glovebox. Reactions were stirred for 0.75-4 hours at 23-60 °C. The reaction mixture was opened to air, diluted with ethyl acetate (~3 mL), and the internal standard undecane was added (7.5  $\mu\text{L}$ , 0.036 mmol). The vial was shaken to mix the contents, and the mixture was filtered through celite prior to analysis by GC and GCMS.

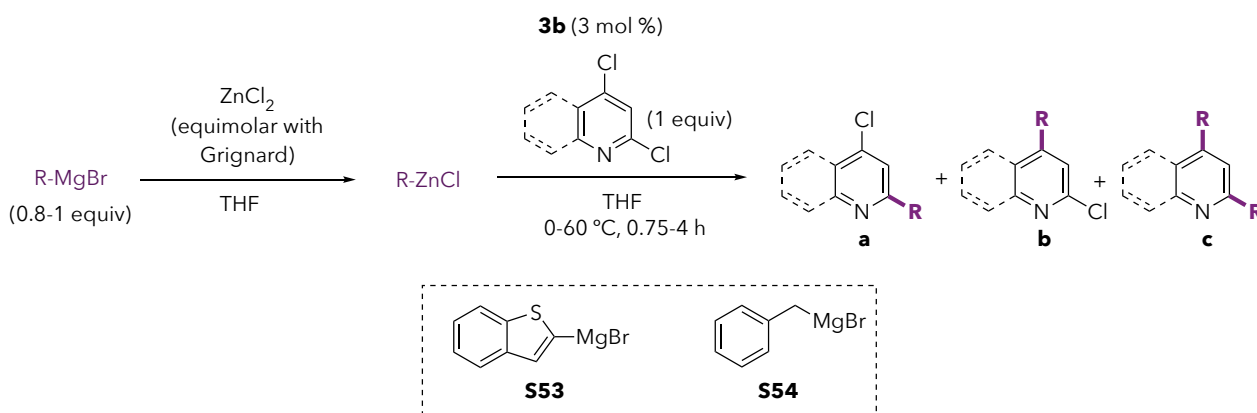
**Scaled-Up Reactions for Product Isolation.** In a glovebox, Grignard reagent (0.77-1 equiv as a solution in THF) and a solution of  $\text{ZnCl}_2$  in THF (1 M, equimolar with the Grignard reagent) were combined in an oven-dried 1- or 5-dram vial ("vial A") equipped with a stirbar. The mixture was stirred for 0.5 hours at ambient temperature. Outside of the glovebox, the precatalyst **3b** (8.4 mg, 0.012 mmol, 3.0 mol %) was added to a separate oven-dried 1- or 5-dram vial equipped with a stir bar ("vial B"), and this vial was transferred into the glovebox. The contents of vial A, comprising the organozinc reagent (0.77-1 equiv), and 2,4-dichloropyridine (0.08 mmol, 1 equiv) were added to vial B. Vial B was sealed with a PTFE-lined cap and removed from the glovebox. Reactions were stirred for 0.75-4 hours at 60 °C unless otherwise indicated. The reaction mixture was opened to air, diluted with ethyl acetate (~3 mL), and washed with sat.  $\text{NH}_4\text{Cl}$  (3 x 10 mL), sat.  $\text{NaHCO}_3$  (3 x 10 mL), and brine (2 x 10 mL) in a separatory

funnel. The organic layer was extracted with additional ethyl acetate (3 x 10 mL) and the combined organic layers were dried over anhydrous MgSO<sub>4</sub>. The crude product was analyzed by GC and purified by normal or reversed phase column chromatography.

## 2. Optimization

Negishi reactions were performed according to the general procedure for GC-scale reactions. Yield of remaining starting material (**1**) is calibrated against the undecane internal standard. Ratio of products **a** and **b** are assuming equal response factor between the isomers.

**Table S4.** Optimization of Negishi Cross Couplings.

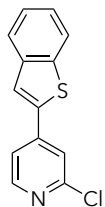


entry	T (°C)	time (h)	R-MgBr (equiv)	<b>a</b> : <b>b</b> <sup>a</sup>	<b>1</b> (%) <sup>b</sup>
1	60	0.75	<b>S59</b> (1)	1 : 68	2
2	60	0.75	<b>S59</b> (0.8)	1 : 7.7	33
3 <sup>c</sup>	60	0.75	<b>S59</b> (1)	-- <sup>d</sup>	98
4	60	3	<b>S60</b> (1)	<1 : 99	0
5	23	3	<b>S60</b> (1)	1 : 45	8
6	0	3	<b>S60</b> (1)	1 : 9	21
7 <sup>c</sup>	60	3	<b>S60</b> (1)	-- <sup>d</sup>	96
8	60	2	MeMgBr (1)	<1 : 99	51
9 <sup>e</sup>	60	4	MeMgBr (1)	1 : 16	0
10 <sup>e</sup>	60	4	MeMgBr (0.8)	1 : 4	0

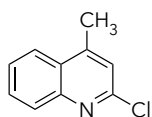
<sup>a</sup> Ratios based on the ratio of signals for **a** and **b** in the GC chromatograph. <sup>b</sup> All reactions were performed with 2,4-dichloropyridine (**1**) unless otherwise indicated. GC yield of recovered starting material calibrated against undecane. <sup>c</sup> No catalyst (metal free control). <sup>d</sup> "--" means no cross-coupled products **a** or **b** were detected. <sup>e</sup> 2,4-Dichloroquinoline (**19**) was used instead of **1**.

**Discussion:** Catalyst-free controls (Table S4, entries 3 and 7) show no reaction, indicating that there is no background S<sub>N</sub>Ar reactivity. Entries 2, 6, and 10 represent optimized conditions that were used for scale-up reactions. Although the results of entry 1 appear promising only low yields of product **B** could be isolated.

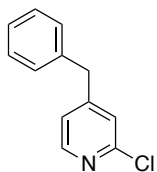
### 3. Isolation and Characterization of Negishi Products from Scheme 3



**4-Benzothiophen-2-yl-2-chloropyridine (34b).** Product **34b** was prepared according to the general procedure using **3b** (8.4 mg, 0.012 mmol, 3 mol %), benzothiophenylzinc chloride (**S53**, 2.35 mL of a  $0.170 \pm 0.001$  M solution in THF, 0.40 mmol, 0.80 equiv), and **1** (54.0  $\mu$ L, 0.50 mmol, 1.00 equiv). The reaction was stirred at 60 °C for 0.75 h. Purification was performed by reversed phase flash column chromatography with a 12 g C18 silica column, using an initial automated program comprising a flow rate of 12 mL/min of water:acetonitrile (98:2 to 5:95) over 35 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 61-70% MeCN in water. After the elution, the initial program was resumed. The C4-monoarylated product **34b** coeluted with the diarylated product at 65-67% MeCN in water. Fractions containing **34b** were partitioned between ethyl acetate and saturated brine. The organic layers were combined and dried over magnesium sulfate and concentrated under reduced pressure. Compound **34b** was further purified by normal phase flash column chromatography ( $R_f = 0.36$  in 80:20 Hex:EtOAc), affording **34b** as an off-white colored solid (68.9 mg, 70% yield). Spectral data are consistent with those previously reported.<sup>20</sup>



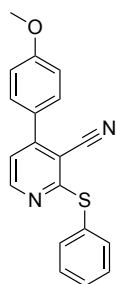
**2-Chloro-4-methylquinoline (35b).** Compound (**35b**) was prepared according to the general procedure using **3b** (8.4 mg, 0.012 mmol, 3 mol %), methylzinc chloride (1.00 mL of  $0.67 \pm 0.02$  M solution in THF, 0.40 mmol, 0.77 equiv), and 2,4-dichloroquinoline (103.0 mg, 0.52 mmol, 1.00 equiv). The reaction was stirred at 60 °C for 4 h. Purification was performed by reversed phase flash column chromatography with a 12 g C18 silica column, using an initial automated program comprising a flow rate of 30 mL/min of water:acetonitrile (98:2 to 6:94) over 35 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 56% MeCN in water. After the elution, the initial program was resumed. The C4 monoarylated product eluted at 56% MeCN. Fractions containing **35b** were partitioned between dichloromethane and saturated brine. The organic layers were combined and dried over magnesium sulfate and solvent was removed under reduced pressure affording **35b** as a white solid (50.5 mg, 71% yield). Spectral data are consistent with those previously reported.<sup>21</sup>



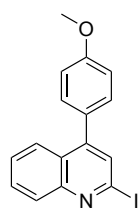
**4-Benzyl-2-chloropyridine (36b).** Compound (**36b**) was prepared according to the general procedure using **3b** (8.4 mg, 0.012 mmol, 3 mol %), benzylzinc chloride (**S54**, 4.82 mL of a 0.083 M solution in THF, 0.40 mmol, 1.00 equiv), and 2,4-dichloropyridine (43.2  $\mu$ L, 0.40 mmol, 1.00 equiv). The reaction was stirred at 0 °C for 3 h. Purification was performed by reversed phase flash column chromatography with a 12 g C18 silica column, using an initial automated program comprising a flow rate of 35 mL/min of water:acetonitrile (98:2 to 5:95) over 35 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 70-73% MeCN in water. After the elution, the initial program was resumed. The C4 monoarylated product **36b** eluted at 72-73% MeCN. Fractions containing **36b** were partitioned between dichloromethane and saturated brine. The organic layers were combined and dried over magnesium sulfate and solvent was removed under reduced pressure affording **36b** as a light brown oil (52.3 mg, 64% yield). <sup>1</sup>H NMR (500 MHz, C<sub>6</sub>D<sub>6</sub>,  $\delta$ ): 7.95 (d,  $J = 5.0$  Hz, 1H), 7.00-7.10 (multiple peaks, 3H), 6.75-6.80

(multiple peaks, 3H), 6.33 (dd,  $J = 5.0, 0.7$  Hz, 1H), 3.27 (s, 2H) ppm.  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{C}_6\text{D}_6$ ,  $\delta$ ): 153.4, 152.7, 150.2, 138.8, 129.5, 129.3, 127.3, 124.9, 123.2, 41.0 ppm. HRMS (ESI<sup>+</sup>)  $m/z$ :  $[\text{M}]^+$ : Calcd for  $\text{C}_{12}\text{H}_{11}\text{ClN}$  203.0502; Found 203.0519.

#### F. Multistep Syntheses using C4-Selective Cross-Coupling with Pd/IPr (Scheme 4)



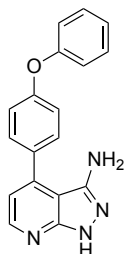
**4-(4-Methoxyphenyl)-2-(phenylthio)-3-pyridinecarbonitrile (37).** Thiophenol (40.8  $\mu\text{L}$ , 0.4 mmol, 1.0 equiv) was added dropwise to a stirring solution of 2-chloro-4-(4-methoxyphenyl)-3-pyridinecarbonitrile (**12b**, 97.9 mg, 0.4 mmol, 1.0 equiv), triethylamine (111.5  $\mu\text{L}$ , 0.8 mmol, 2.0 equiv), and DMF (0.4 mL) in a 1-dram reaction vial. The vial was sparged with  $\text{N}_2$  gas using the technique described in the “general procedure” on S9, sealed with a PTFE-lined cap, and heated to 60  $^\circ\text{C}$  for 18.3 h. After cooling, a crystalline precipitate formed. The mixture was poured into a 125 mL Erlenmeyer flask and diluted with 30 mL of deionized water, inducing precipitation of the desired product and solubilizing byproducts. The precipitate was filtered and saved, and the liquid filtrate was extracted with ethyl acetate. The organic phase was concentrated, affording mixed white and orange solids. The solids were washed with diethyl ether over a fritted funnel until all orange discoloration had passed into the filtrate. The combined organic solids were dried under vacuum furnishing **37** as a colorless crystalline solid with slight yellow discoloration (92.2 mg, 72% yield); mp 156–158  $^\circ\text{C}$  [uncorrected, measured against benzoic acid (113–117  $^\circ\text{C}$ )].  $^1\text{H}$  NMR (500 MHz,  $\text{C}_6\text{D}_6$ ,  $\delta$ ): 7.84 (d,  $J = 5.2$  Hz, 1H), 7.59–7.57 (m, 2H), 7.24–7.22 (m, 2H), 7.12–7.04 (multiple peaks, 3H), 7.00–6.67 (m, 2H), 6.29 (d,  $J = 5.2$  Hz, 1H), 3.23 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{C}_6\text{D}_6$ ,  $\delta$ ): 164.9, 161.6, 153.4, 151.3, 136.4, 130.3, 129.5, 129.4, 129.3, 119.7, 115.8, 114.6, 105.5, 54.9 (two signals are coincidentally overlapping).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 164.5, 161.4, 153.7, 151.8, 135.6, 130.1, 129.6, 129.4, 128.8, 127.9, 120.0, 115.9, 114.6, 105.1, 55.6. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{19}\text{H}_{14}\text{N}_2\text{OS}$  318.0827; Found 318.0812.



**2-Iodo-4-(4-methoxyphenyl)quinoline (38).** Reaction conditions were adapted from a literature procedure describing the di-iodination of 2,4-dichloropyridine.<sup>22</sup> 2-Chloro-4-(4-methoxyphenyl)quinoline (**19b**, 107.9 mg, 0.4 mmol, 1.0 equiv), NaI (179.9 mg, 1.2 mmol, 3.0 equiv), and acetonitrile (0.45 mL) were combined in a 1-dram vial with stir bar. Acetyl chloride (2.1 equiv, 0.84 mmol, 59.9  $\mu\text{L}$ ) was added dropwise to the stirring mixture. The vial was sparged with  $\text{N}_2$  gas using the technique described under “general procedure” on S9 sealed with a PTFE cap, and heated to reflux for 67 h. TLC analysis indicated complete consumption of quinoline starting material **19b**. The product mixture was quenched by dropwise addition of deionized water (8.5 mL) while stirring. An aqueous solution of 10%  $\text{K}_2\text{CO}_3$ /5%  $\text{NaHSO}_3$  was added to the mixture, and the mixture was shaken and extracted with  $\text{CHCl}_3$  (3x). The organic fractions were combined, dried over  $\text{Mg}_2\text{SO}_4$ , filtered, concentrated, and purified by column chromatography on silica ( $R_f = 0.46$  in 10% ethyl acetate in hexanes) to separate product **38** from polar impurities detected by TLC. Product fractions were concentrated and dried affording **38** as a white crystalline solid (119 mg, 82% yield); mp 114–117  $^\circ\text{C}$  [uncorrected, measured against benzoic acid (112–114  $^\circ\text{C}$ )].  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.06 (d,  $J = 8.4$



Hz, 1H), 7.88 (d,  $J = 8.4$  Hz, 1H), 7.67 (ddd,  $J = 8.4, 6.9, 1.3$  Hz, 1H), 7.64 (s, 1H), 7.47 (ddd,  $J = 8.4, 6.9, 1.3$  Hz, 1H), 7.40-7.37 (m, 2H), 7.04-7.01 (m, 2H), 3.87 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 160.3, 149.97, 149.50, 131.6, 130.8, 130.1, 129.3, 128.6, 127.1, 126.3, 126.2, 119.2, 114.3, 55.5. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{16}\text{H}_{12}\text{INO}$  360.9964; Found 360.9963.



**4-(4-phenoxyphenyl)-1H-Pyrazolo[3,4-b]pyridin-3-amine (40).**

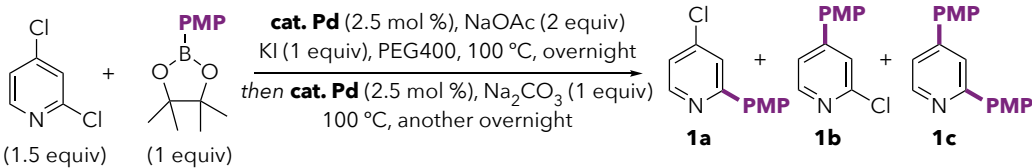
2-Chloro-4-(4-phenoxyphenyl)-3-pyridinecarbonitrile (**39b**, 122.7 mg, 0.4 mmol, 1.0 equiv), hydrazine-monohydrate (200.2 mg, 4.0 mmol, 10.0 equiv), and *i*PrOH (6.2 mL) were combined with a stir bar in a 25 mL round-bottom flask equipped with water-cooled jacketed condenser. The condenser was sealed with a rubber septum secured by copper wire, the system was sparged with  $\text{N}_2$  gas, and the reaction was heated to 90 °C with stirring for 15 h under an atmosphere of  $\text{N}_2$ . The mixture was cooled, and trace impurities were detected by TLC. Solvent was removed under reduced pressure, and the solids were columned on silica ( $R_f = 0.27$  in 4.5% methanol in  $\text{CH}_2\text{Cl}_2$ ). All rinses and transfers were completed with  $\text{CHCl}_3$ . Solvent was removed under reduced pressure, furnishing a bright yellow solid. Drying under vacuum afforded **40** (119.3 mg, 99% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 10.82 (br s, 1H), 8.50 (d,  $J = 4.8$  Hz, 1H), 7.58-7.55 (m, 2H), 7.42-7.39 (m, 2H), 7.20-7.17 (m, 1H), 7.16-7.14 (m, 2H), 7.12-7.10 (m, 2H), 6.95 (d,  $J = 4.8$  Hz, 1H), 4.05 (br s, 2H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 158.8, 156.3, 153.5, 149.4, 147.5, 145.3, 131.7, 130.4, 130.1, 124.3, 119.8, 118.6, 115.8, 104.1.

## G. Ligand-Free Suzuki Cross-Couplings

### 1. Ligand-Free Control Reaction Based on Previously-Reported Conditions (Scheme 5b)

Reactions were set up according to the literature procedure<sup>15</sup> (Table S5, entries 1-3) with the indicated modifications (entries 4-12).

**Table S5.** Analysis of the Role of Ligand in the Previously-Reported Conditions<sup>15</sup> for C4-Selective Suzuki Cross-Coupling<sup>a</sup>



entry	trial	Pd source	additional ligand (mol %)	<b>1a</b> (%)	<b>1b</b> (%)	<b>1c</b> (%)	<b>1a : 1b</b>
1	1	PEPPSI-IPr	--	<1	80	2	<1 : 80
2	2	PEPPSI-IPr	--	<1	84	2	<1 : 84
3	Average	PEPPSI-IPr	--	<1	82	2	<1 : 82
4	1	<b>3b</b>	--	<1	69	1	<1 : 69
5	2	<b>3b</b>	--	<1	81	1	<1 : 81
6	Average	<b>3b</b>	--	<1	75	1	<1 : 75
7	1	Pd(OAc) <sub>2</sub>	PPh <sub>3</sub> (5)	28	14	2	1.9 : 1
8	2	Pd(OAc) <sub>2</sub>	PPh <sub>3</sub> (5)	32	18	2	1.8 : 1
9	Average	Pd(OAc) <sub>2</sub>	PPh <sub>3</sub> (5)	30	16	2	1.9 : 1
10	1	PdCl <sub>2</sub>	--	<1	54	1	<1 : 54
11	2	PdCl <sub>2</sub>	--	<1	66	1	<1 : 66
12	Average	PdCl <sub>2</sub>	--	<1	60	1	<1 : 60

<sup>a</sup>GC yields calibrated against undecane as the internal standard.

**Discussion:** Selectivity under Yang's high-temperature conditions is significantly better than what is observed under the Pd/IPr-catalyzed room temperature conditions employed in our work. However, ligand-free control reactions using PdCl<sub>2</sub> (Table S5, entries 10-12) demonstrate that the same extremely high selectivity is observed in the absence of IPr. The selectivity under the conditions reported in reference 15 is therefore not ligand-controlled. When PPh<sub>3</sub> is used as a ligand in combination with Pd(OAc)<sub>2</sub> (entries 7-9, a catalytic system analogous to Fairlamb's work with dibromopyridine),<sup>18</sup> the reaction favors C2-arylated product **1a**, but with poor yield. In all other entries, extremely high selectivity for product **1b** is observed.

## 2. Coupling of **1** under Fairlamb's Recently Reported Pd/PPh<sub>3</sub> Conditions

The Suzuki coupling in Table S6 was set up according to conditions reported by Fairlamb et al., who observed C4-selective cross-coupling of 2,4-dibromopyridine.<sup>18</sup> Reactions were conducted on a 0.16 mmol reaction scale.

**Table S6.** Reaction of **1** under Fairlamb's Conditions

entry	solvent	Temp. (°C)	<b>1a</b> (%)	<b>1b</b> (%)	<b>1c</b> (%)	<b>1a</b> : <b>1b</b>
1	THF	40	90.6	5.2	4.9	17 : 1
2	dioxane	110	85.4	0.6	12.8	142 : 1

**Discussion:** Intriguingly, although these conditions promote C4-selective Suzuki coupling of 2,4-dibromopyridine, 2,4-dichloropyridine undergoes preferential C2-coupling.

## 3. General Procedures for the Ligand-Free 'Jeffery' Conditions Reported Herein

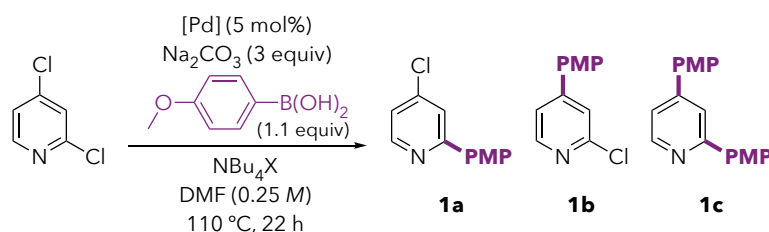
**GC-Scale Reactions.** Solids were added to a 1-dram reaction vial in order of increasing mass: palladium catalyst (typically 0.008 mmol, 5 mol %), *p*-methoxyphenylboronic acid (26.7 mg, 0.18 mmol, 1.1 equiv), sodium carbonate (50.9 mg, 0.48 mmol, 3.0 equiv), tetrabutylammonium salt (0.5 equiv), and a stir bar. Inside a nitrogen-atmosphere glovebox, liquid reagents were pre-measured by syringe and added in quick succession: DMF (0.64 mL) via 1-mL syringe, followed by 2,4-dichloropyridine (17.2  $\mu$ L, 0.16 mmol, 1.0 equiv) via 25- $\mu$ L syringe. The reaction vial was sealed under nitrogen with either a septum screwcap or a solid PTFE-lined screwcap. When indicated, N<sub>2</sub>-sparged deionized water was added through the septum cap outside the glovebox via 25- or 50- $\mu$ L syringe, and the puncture was sealed with electrical tape. The reaction was stirred vigorously at the indicated temperature for 22 h. The reaction vial was opened to air, diluted with ethyl acetate (~ 3 mL), and *n*-undecane (15  $\mu$ L, 0.44 equiv) was added as an internal standard via 25- $\mu$ L syringe. An aliquot was filtered through celite into a separate vial for GC analysis.

**Scaled-Up Reactions for Product Isolation.** PdCl<sub>2</sub> (4.4 mg, 0.025 mmol, 5.0 mol%) was weighed into a 1-dram vial followed by solid substrate if applicable (2,5-dichloropyridine, pyrimidine, or 2,4-dichloroquinoline, 0.5 mmol, 1 equiv), arylboronic acid (0.5 mmol, 1.1 equiv), sodium carbonate (159 mg, 1.5 mmol, 3.0 equiv), tetrabutylammonium bromide (806 mg, 2.5 mmol, 5.0 equiv) and a stir bar. DMF (2 mL) was added inside a nitrogen-atmosphere glovebox and sealed with either a septum screwcap or a solid PTFE-lined screwcap. Unless otherwise indicated, N<sub>2</sub>-sparged deionized water (45  $\mu$ L, 2.5 mmol, 5.0 equiv) was added through the septum cap outside the glovebox via 50- $\mu$ L syringe (*note: conversion in 0.5 mmol scale reactions was more adversely affected by dry conditions compared to 0.16 mmol scale reactions*), followed by 2,4-dichloropyridine substrate (if liquid), and the puncture was sealed with electrical tape. The reaction was stirred vigorously at 110 °C for the indicated time. After cooling to ambient temperature, the cap was removed, the reaction mixture was diluted with ethyl acetate and,

in cases where calibrated GC yields are reported, *n*-undecane (46.9  $\mu$ L, 0.44 equiv) was added as an internal standard via 50- $\mu$ L syringe. The reaction mixture was then filtered through a plug of celite. The reaction outcome was assessed by GC and TLC (isomeric ratios were determined by GC). The mixture was purified by flash column chromatography and/or reversed phase chromatography as indicated, and the product was dried under vacuum.

#### 4. Optimization of the Ligand-Free Conditions (Table 2)

**Table S7.** Optimization of Ligand-Free Suzuki Coupling

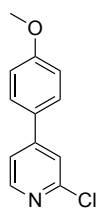


entry	trial	Pd source	NBu <sub>4</sub> X (equiv)	<b>1a</b> (%)	<b>1b</b> (%)	<b>1c</b> (%)	<b>1a : 1b</b>
1	1	PdCl <sub>2</sub>	Br (3)	0.3	82	0.4	1 : >99
2	2	PdCl <sub>2</sub>	Br (3)	0.6	80	0.4	1 : >99
3	Average	PdCl <sub>2</sub>	Br (3)	0.5	81	0.4	1 : >99
4	1	PdCl <sub>2</sub>	Br (5)	0.4	87	0.4	1 : >99
5	2	PdCl <sub>2</sub>	Br (5)	0.5	80	0.4	1 : >99
6	Average	PdCl <sub>2</sub>	Br (5)	0.5	84	0.4	1 : >99
7	1	Pd(OAc) <sub>2</sub>	Br (3)	0.7	75	0.4	1 : >99
8	2	Pd(OAc) <sub>2</sub>	Br (3)	0.6	71	0.3	1 : >99
9	Average	Pd(OAc) <sub>2</sub>	Br (3)	0.7	73	0.4	1 : >99
10	1	Pd <sub>2</sub> dba <sub>3</sub>	Br (3)	0.4	61	0.2	1 : >99
11	2	Pd <sub>2</sub> dba <sub>3</sub>	Br (3)	0.5	52	0.2	1 : >99
12	Average	Pd <sub>2</sub> dba <sub>3</sub>	Br (3)	0.5	57	0.2	1 : >99
13	1	PdCl <sub>2</sub>	Cl (3)	0.8	72	0.4	1 : 90
14	2	PdCl <sub>2</sub>	Cl (3)	1.3	72	0.6	1 : 55
15	Average	PdCl <sub>2</sub>	Cl (3)	1.1	72	0.5	1 : 65
16	1	PdCl <sub>2</sub>	Br (1)	1.2	67	0.8	1 : 56
17	2	PdCl <sub>2</sub>	Br (1)	1.2	71	0.6	1 : 59
18	Average	PdCl <sub>2</sub>	Br (1)	1.2	69	0.7	1 : 58
19	1	PdCl <sub>2</sub>	PF <sub>6</sub> (3)	2.6	44	2.4	1 : 17
20	2	PdCl <sub>2</sub>	PF <sub>6</sub> (3)	3.4	52	3.8	1 : 15
21	Average	PdCl <sub>2</sub>	PF <sub>6</sub> (3)	3.0	48	3.1	1 : 16
22 <sup>b</sup>	1	PdCl <sub>2</sub>	---	1.9	70	1.6	1 : 37
23 <sup>b</sup>	2	PdCl <sub>2</sub>	---	2.2	68	1.5	1 : 31
24 <sup>b</sup>	Average	PdCl <sub>2</sub>	---	2.1	69	1.6	1 : 33
25	1	PdCl <sub>2</sub>	---	3.3	55	3.0	1 : 17
26	2	PdCl <sub>2</sub>	---	3.7	58	3.2	1 : 16
27	Average	PdCl <sub>2</sub>	---	3.5	56	3.1	1 : 16
28	1	PdCl <sub>2</sub>	Br (1)	1.2	67	0.8	1 : 56
29	2	PdCl <sub>2</sub>	Br (1)	1.2	71	0.6	1 : 59
30	Average	PdCl <sub>2</sub>	Br (1)	1.2	69	0.7	1 : 58
31 <sup>c</sup>	1	PdCl <sub>2</sub>	Br (3)	1	80	0.6	1 : 80
32 <sup>c</sup>	2	PdCl <sub>2</sub>	Br (3)	0.8	78	0.5	1 : 98
33 <sup>c</sup>	Average	PdCl <sub>2</sub>	Br (3)	0.9	79	0.6	1 : 88
34 <sup>d</sup>	1	PdCl <sub>2</sub>	Br (3)	1.1	82	0.6	1 : 75
35 <sup>d</sup>	2	PdCl <sub>2</sub>	Br (3)	1.0	84	0.8	1 : 84
36 <sup>d</sup>	Average	PdCl <sub>2</sub>	Br (3)	1.1	83	0.7	1 : 75
37 <sup>e</sup>	1	PdCl <sub>2</sub>	Br (3)	0.8	40	0.5	1 : 50

38 <sup>e</sup>	2	PdCl <sub>2</sub>	Br (3)	0.8	49	0.4	1 : 61
39 <sup>e</sup>	Average	PdCl <sub>2</sub>	Br (3)	0.8	45	0.5	1 : 56
40 <sup>f</sup>	1	PdCl <sub>2</sub>	Br (3)	0.3	8	0.4	1 : 27
41 <sup>f</sup>	2	PdCl <sub>2</sub>	Br (3)	0.3	10	0.5	1 : 33
42 <sup>f</sup>	Average	PdCl <sub>2</sub>	Br (3)	0.3	9	0.5	1 : 29
43 <sup>c</sup>	1	PdCl <sub>2</sub>	Br (5)	0.8	85	0.4	1 : >99
44 <sup>c</sup>	2	PdCl <sub>2</sub>	Br (5)	0.7	84	0.6	1 : >99
45 <sup>c</sup>	Average	PdCl <sub>2</sub>	Br (5)	0.8	85	0.5	1 : >99

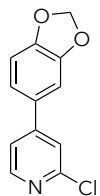
<sup>a</sup>Reactions were conducted according to the General Procedure on S37 for GC-scale reactions. GC yields calibrated against undecane as an internal standard. Calculated yield values were rounded to the nearest integer. <sup>b</sup>With KBr (3 equiv). <sup>c</sup>With H<sub>2</sub>O (5 equiv). <sup>d</sup>With H<sub>2</sub>O (10 equiv). <sup>e</sup>With 1 mol% PdCl<sub>2</sub>. <sup>f</sup>Reaction run at 25 °C.

## 5. Isolation and Characterization of Cross-Coupled Products (Scheme 6)

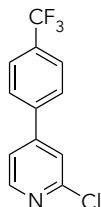


**2-Chloro-4-(4-methoxyphenyl)pyridine (1b).** Compound **1b** was prepared according to the general procedure under ligand-free conditions using 2,4-dichloropyridine (54 μL, 0.5 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (83.6 mg, 0.55 mmol, 1.1 equiv) with stirring for 30 h. Purification by flash column chromatography on silica gel in 6-20% ethyl acetate/hexanes ( $R_f = 0.42$  in 15% ethyl acetate in hexanes) provided **1b** as a white solid (82.1 mg, 75% yield). Spectral data are consistent with previously reported data and literature.<sup>12</sup>

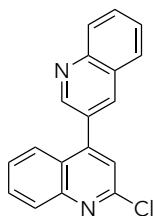
**Larger-Scale Preparation of 1b:** Compound **1b** was prepared according to the general procedure under ligand-free conditions using 2,4-dichloropyridine (270 μL, 2.5 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (417.9 mg, 2.75 mmol, 1.1 equiv) with stirring for 40 h. Purification by flash column chromatography on silica gel in 6-20% ethyl acetate/hexanes ( $R_f = 0.42$  in 15% ethyl acetate in hexanes) provided **1b** as a white solid (347 mg, 63% yield). Spectral data are consistent with previously reported data and literature.<sup>12</sup>



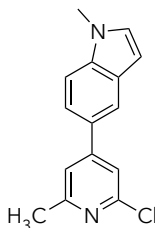
**4-(1,3-benzodioxol-5-yl)-2-chloro-pyridine (41b).** Compound **41b** was prepared according to the general procedure under ligand-free conditions using 2,4-dichloropyridine (54 μL, 0.5 mmol, 1.0 equiv) and 3,4-(methylenedioxy)benzeneboronic acid (91.3 mg, 0.55 mmol, 1.1 equiv) with stirring for 34 h. Purification by flash column chromatography on silica gel in 2-21% ethyl acetate/hexanes ( $R_f = 0.35$  in 15% ethyl acetate in hexanes) provided **41b** as a white solid (75 mg, 64% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, δ): 8.36 (d,  $J = 5.2$  Hz, 1H), 7.44 (d,  $J = 1.6$  Hz, 1H), 7.32 (dd,  $J = 5.2, 1.7$  Hz, 1H), 7.11 (dd,  $J = 8.1, 1.6$  Hz, 1H), 7.06 (d,  $J = 1.7$  Hz, 1H), 6.90 (d,  $J = 8.1$  Hz, 1H), 6.03 (s, 2H). <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, CDCl<sub>3</sub>, δ): 152.3, 151.2, 150.1, 149.2, 148.7, 131.0, 121.7, 121.4, 120.2, 109.1, 107.3, 101.8. HRMS (ESI Q-TOF)  $m/z$ : [M]<sup>+</sup> Calcd for C<sub>12</sub>H<sub>8</sub>ClNO<sub>2</sub> 233.0244; Found 233.0245.



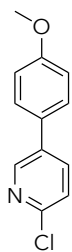
**2-Chloro-4-[4-(trifluoromethyl)phenyl]pyridine (42b).** Compound **42b** was prepared according to the general procedure under ligand-free conditions using 2,4-dichloropyridine (54  $\mu$ L, 0.5 mmol, 1.0 equiv) and *p*-(trifluoromethyl)phenylboronic acid (104.5 mg, 0.55 mmol, 1.1 equiv) with stirring for 23 h. Purification by flash column chromatography on silica gel using a linear gradient of 1-28% ethyl acetate/hexanes over 9 column volumes ( $R_f$  = 0.40 in 15% ethyl acetate in hexanes) provided **42b** as a white solid (79.1 mg, 62% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 8.46 (d,  $J$  = 5.2 Hz, 1H), 7.76 (d,  $J$  = 8.4, 2H), 7.72 (d,  $J$  = 8.4 Hz, 2H), 7.55 (d,  $J$  = 1.3 Hz, 1H), 7.43 (dd,  $J$  = 5.2, 1.3 Hz, 1H). Spectral data are consistent with literature.<sup>23</sup>



**2-Chloro-4-(3-quinolinyl)quinoline (43b).** Compound **43b** was prepared according to the general procedure under ligand-free conditions using 2,4-dichloroquinoline (99 mg, 0.5 mmol, 1.0 equiv) and 3-quinolineboronic acid (95.1 mg, 0.55 mmol, 1.1 equiv) with stirring for 27.5 h. Purification by flash column chromatography on silica gel in 20% ethyl acetate/hexanes ( $R_f$  = 0.33 in 20% ethyl acetate in hexanes) provided **43b** as a tan solid (94.1 mg, 65% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 9.05 (d,  $J$  = 1.9 Hz, 1 H), 8.32 (d,  $J$  = 1.4 Hz, 1 H), 8.24 (d,  $J$  = 8.5 Hz, 1H), 8.14 (d,  $J$  = 8.5 Hz, 1H) 7.94 (d,  $J$  = 8.1 Hz, 1H), 7.83-7.87 (multiple peaks, 3H), 7.68 (dd,  $J$  = 15.2, 7.4 Hz, 1H), 7.56 (dd,  $J$  = 15.2, 7.4 Hz, 1H), 7.47 (s, 1H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 150.5, 150.4, 148.6, 148.22, 148.15, 136.7, 131.0, 130.8, 130.0, 129.7, 129.5, 128.3, 127.8, 127.7, 127.6, 125.7, 125.5, 122.9. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{18}\text{H}_{11}\text{ClN}_2$  290.0611; Found 290.0615.



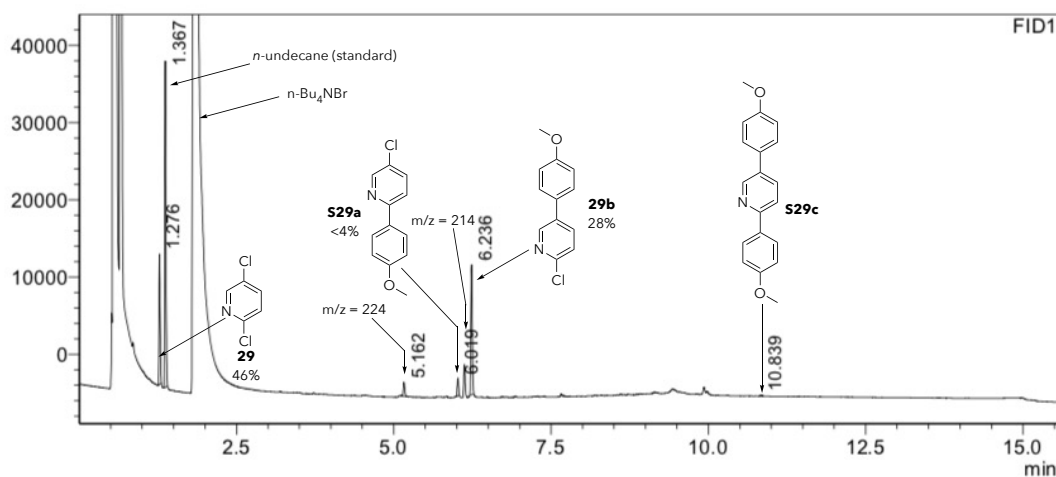
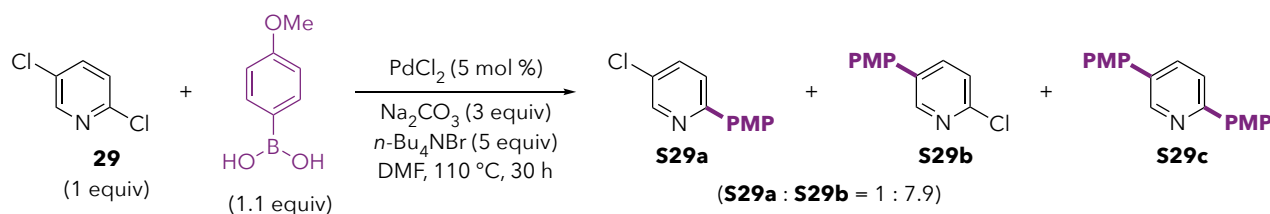
**2-Chloro-6-methyl-4-(1-methyl-1H-indol-5-yl)pyridine (44b).** Compound **44b** was prepared according to the general procedure under ligand-free conditions using 2,4-dichloro-6-methylpyridine (81 mg, 0.5 mmol, 1.0 equiv) and 1-methyl-5-indolyl-boronic acid (96.2 mg, 0.55 mmol, 1.1 equiv) with stirring for 30 h. Purification by flash column chromatography on silica gel in 5-20% ethyl acetate/hexanes ( $R_f$  = 0.45 in 20% ethyl acetate in hexanes) provided **44b** as a white solid (62.2 mg, 49% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.88 (m, 1.6 Hz, 1H), 7.45 (dd,  $J$  = 8.6, 1.6 Hz, 1H), 7.41 (d,  $J$  = 0.8 Hz, 1H), 7.38 (d,  $J$  = 8.6 Hz, 1H), 7.33 (d,  $J$  = 0.8 Hz, 1H), 7.11 (d,  $J$  = 3.1 Hz, 1H), 6.56 (dd,  $J$  = 3.1, 0.7 Hz, 1H), 3.81 (s, 3H), 2.58 (s, 3H).  $^{13}\text{C}\{^1\text{H}\}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 159.3, 153.2, 151.1, 137.4, 130.3, 129.1, 128.4, 120.7, 120.1, 119.9, 119.0, 110.0, 101.9, 33.1, 24.4. HRMS (ESI Q-TOF)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{15}\text{H}_{13}\text{ClN}_2$  256.0767; Found 256.0772.



**2-Chloro-5-(4-methoxyphenyl)pyridine (29b).** Compound **29b** was prepared according to the general procedure under ligand-free conditions, with the modification that no water was added. The reaction vial was charged with 2,5-dichloropyridine (74 mg, 0.5 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (83.6 mg, 0.55 mmol, 1.1 equiv) and stirred for 30 h. Purification by flash column chromatography on silica gel in 1-36% ethyl acetate/hexanes ( $R_f$  = 0.41 in 15% ethyl acetate in hexanes) resulted in coelution of **29b** and the diarylated product (the estimated crude yield of diarylated product is ~1% based on GC analysis). Further purification was performed using reversed phase flash column

chromatography with a 30 g C18 silica column, using an initial automated program comprising a flow rate of 35 mL/min of water:acetonitrile (98:2 to 1:99) over 20 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 98% MeCN in water. Fractions containing **29b** were partitioned between ethyl acetate and saturated brine. The organic layers were combined and dried over magnesium sulfate and dried *in vacuo* providing **29b** as a white solid (22.4 mg, 20% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, δ): 8.57 (dd, *J* = 2.6, 0.6 Hz, 1H), 7.79 (dd, *J* = 8.3, 2.6 Hz, 1H), 7.46- 7.51 (m, 2H), 7.36 (dd, *J* = 8.3, 0.6 Hz, 1H), 6.98-7.03 (m, 2H), 3.86 (s, 3H). The product of C2-arylation was also isolated (4.5 mg, 4%). Spectral data for both monoarylated products are consistent with the literature.<sup>11</sup> Side products resulting from diarylation and homocoupling of **29** (*m/z* = 224) were each detected by GCMS (Scheme S11). These side products, as well as remaining starting material, appears to account for most of the remaining mass balance.

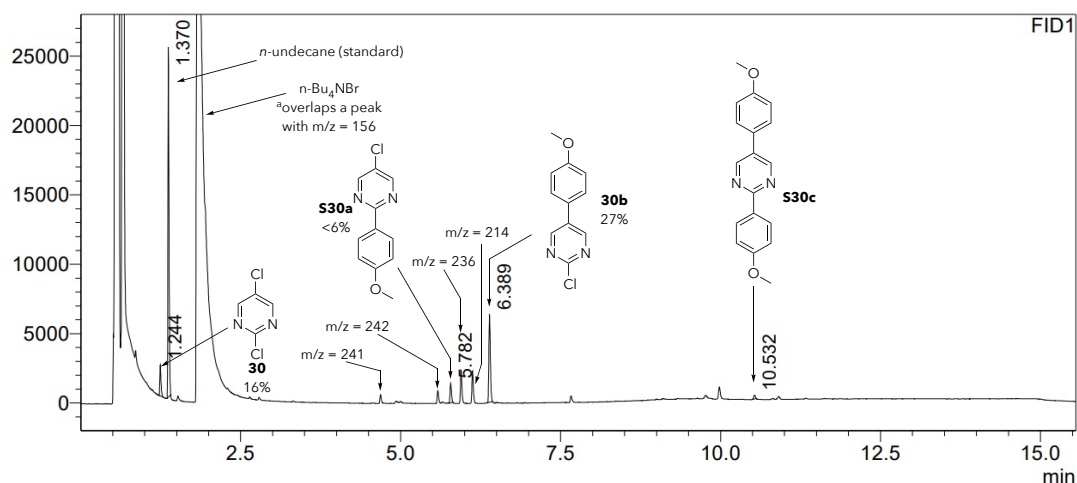
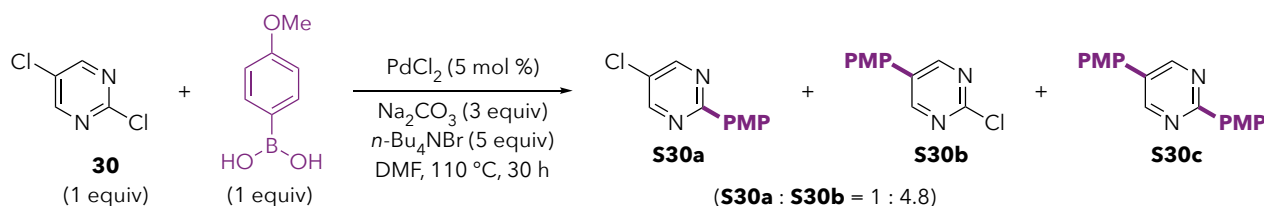
**Scheme S11.** Suzuki reaction of **29** under ligand-free conditions.



**2-Chloro-5-(4-methoxyphenyl)pyrimidine (30b).** Compound **30b** was prepared according to the general procedure under ligand-free conditions with the modification that no water was added. The reaction vial was charged with 2,5-dichloropyrimidine (74.5 mg, 0.5 mmol, 1.0 equiv) and *p*-methoxyphenylboronic acid (83.6 mg, 0.55 mmol, 1.1 equiv) and stirred for 30 h. Purification by flash column chromatography on silica gel in 1-45% ethyl acetate/hexanes (*R<sub>f</sub>* = 0.31 in 15% ethyl acetate in hexanes) resulted in coelution of the C5 isomer and a bipyrimidine side product. Fractions containing product **30b** were combined and solvent was removed under reduced pressure. Further purification was performed using reversed phase flash column chromatography with a 30 g C18 silica column, using an initial automated

program comprising a flow rate of 35 mL/min of water:acetonitrile (98:2 to 1:99) over 20 column volumes. When the product began eluting, the program was paused and reset to elute the analyte at 50-55% MeCN in water. Fractions containing **30b** were partitioned between ethyl acetate and saturated brine. The organic layers were combined and dried over magnesium sulfate and dried *in vacuo* providing **30b** as a white solid (18.5 mg, 17% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, δ): 8.78 (s, 2H), 7.46-7.53 (m, 2H), 7.00-7.08 (m, 2H), 3.87 (s, 3H). Spectral data are consistent with the literature.<sup>24</sup> The crude reaction mixture was analyzed by GCMS; in addition to remaining starting material, a small amount of C<sub>2</sub>-arylated product, and trace diarylated product, several small peaks corresponding to unidentified side products were observed (Scheme S12).

**Scheme S12.** Suzuki reaction of **30** under ligand-free conditions.

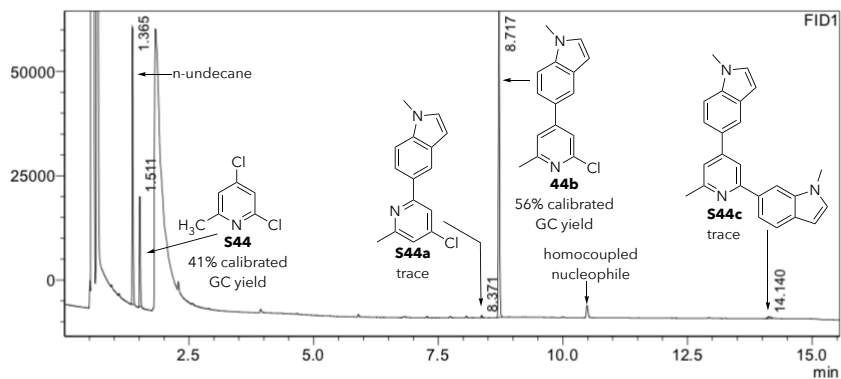
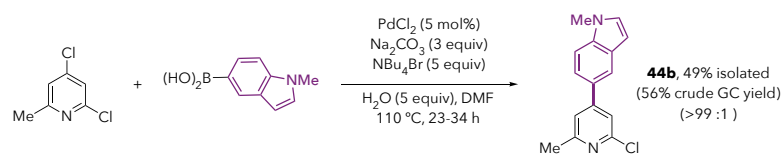
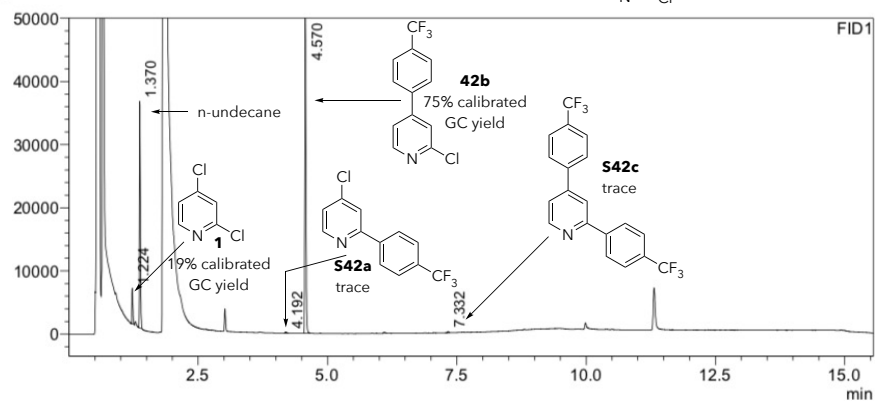
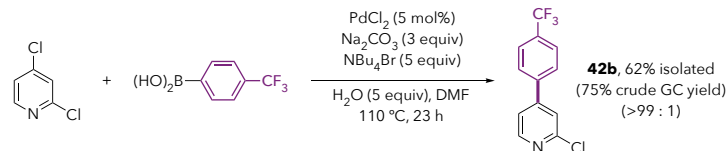
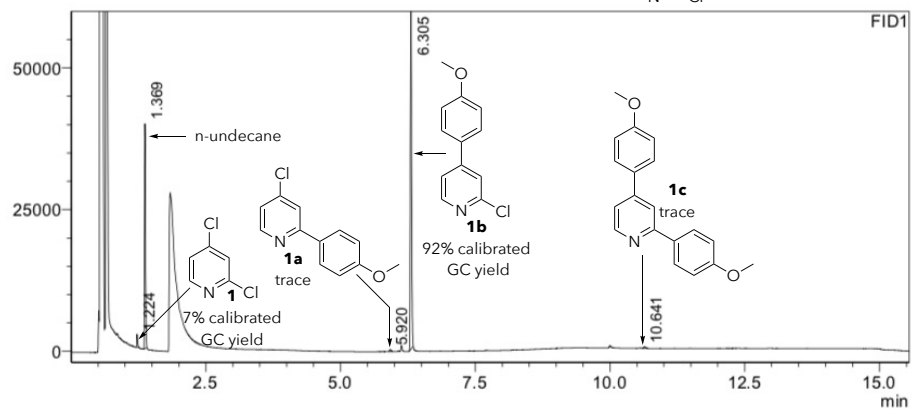
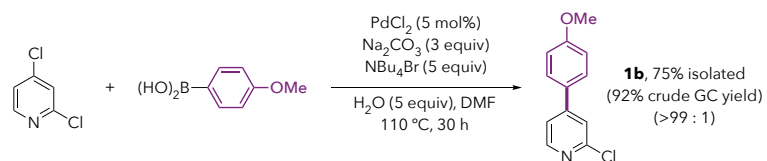


<sup>a</sup>A side product that overlaps with NBu<sub>4</sub>Br was detected in a column fraction during purification and analyzed by GCMS providing a m/z = 157. This mass is consistent with a side product resulting from an S<sub>N</sub>Ar reaction between dimethylamine and 2,5-dichloropyrimidine. Dimethylamine is a common contaminant in DMF.

## 6. Discussion about Mass Balance

Lower yields under the ligand-free conditions are near-exclusively explained by unreacted starting material as well as some material loss during product isolation, as illustrated by the three GC chromatograms below that are representative of the reactions shown in Scheme 6 (product was not isolated from the trials shown below due to the addition of internal standard, undecane, after completion of the reaction). We tentatively hypothesize that the reactions stall due to aggregation of Pd nanoparticles into unreactive Pd black, leaving a significant portion of starting material unreacted.

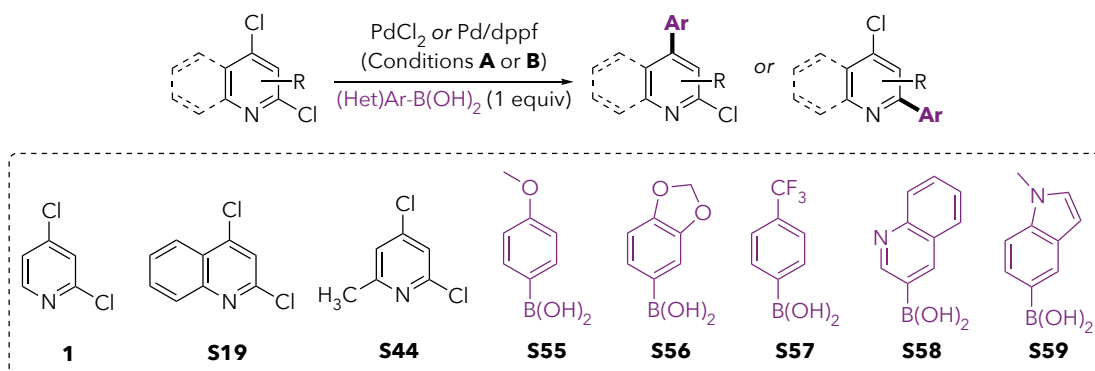




## 7. Comparison of Ligand-Free Results (C4-Selective) vs. C2-Selective Conditions

The ligand-free conditions described herein promote Suzuki-Miyaura cross-coupling of 2,4-dichloropyridines and quinoline with high C4-selectivity and promote C5-selective coupling of 2,5-dichloropyridine and pyrimidine. In contrast, Pd/dppf is a highly C2-selective catalyst for each of these substrates. Tables S8 and S9 demonstrate orthogonality between these two catalytic systems, based on the distinct GC retention times of the major products under each of these conditions.

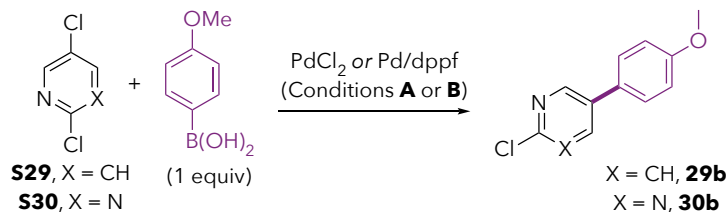
**Table S8.** Retention times (RT) of C4- and C2-arylated products by GC-FID for ligand-free reactions versus Pd(OAc)<sub>2</sub>/dppf-catalyzed reactions.<sup>a</sup>



entry	substrate	nucleophile	RT with A (min)	RT with B (min)
1	<b>1</b>	<b>S55</b>	6.31	5.91
2	<b>1</b>	<b>S56</b>	6.84	6.46
3	<b>1</b>	<b>S57</b>	4.58	4.18
4	<b>S19</b>	<b>S58</b>	10.22	10.85
5	<b>S44</b>	<b>S59</b>	8.73	8.37

<sup>a</sup>Conditions A: PdCl<sub>2</sub> (5 mol%), Na<sub>2</sub>CO<sub>3</sub> (3 equiv), N(n-Bu)<sub>4</sub>Br (5 equiv), H<sub>2</sub>O (5 equiv), DMF (0.25 M), 110 °C, 22-34h.  
Conditions B: Pd(OAc)<sub>2</sub>/dppf (5 mol%), Cs<sub>2</sub>CO<sub>3</sub> (2.5 equiv), H<sub>2</sub>O (7 equiv), 1,4-dioxane (0.25 M), 60 °C, 12 h.

**Table S9.** Retention times (RT) of C5- and C2-arylated products by GC-FID for ligand-free reactions versus Pd(OAc)<sub>2</sub>/dppf-catalyzed reactions<sup>a</sup>



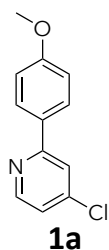
entry	substrate	nucleophile	RT with A (min)	RT with B (min)
1	<b>S29</b>	<b>S55</b>	6.24	6.02
5	<b>S30</b>	<b>S55</b>	6.39	5.78

<sup>a</sup>Conditions A: PdCl<sub>2</sub> (5 mol%), Na<sub>2</sub>CO<sub>3</sub> (3 equiv), N(n-Bu)<sub>4</sub>Br (5 equiv), H<sub>2</sub>O (5 equiv), DMF (0.25 M), 110 °C, 22-34h.  
Conditions B: Pd(OAc)<sub>2</sub>/dppf (5 mol%), Cs<sub>2</sub>CO<sub>3</sub> (2.5 equiv), H<sub>2</sub>O (7 equiv), 1,4-dioxane (0.25 M), 60 °C, 12 h.

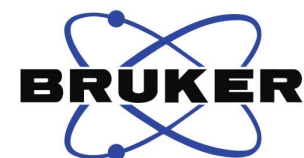
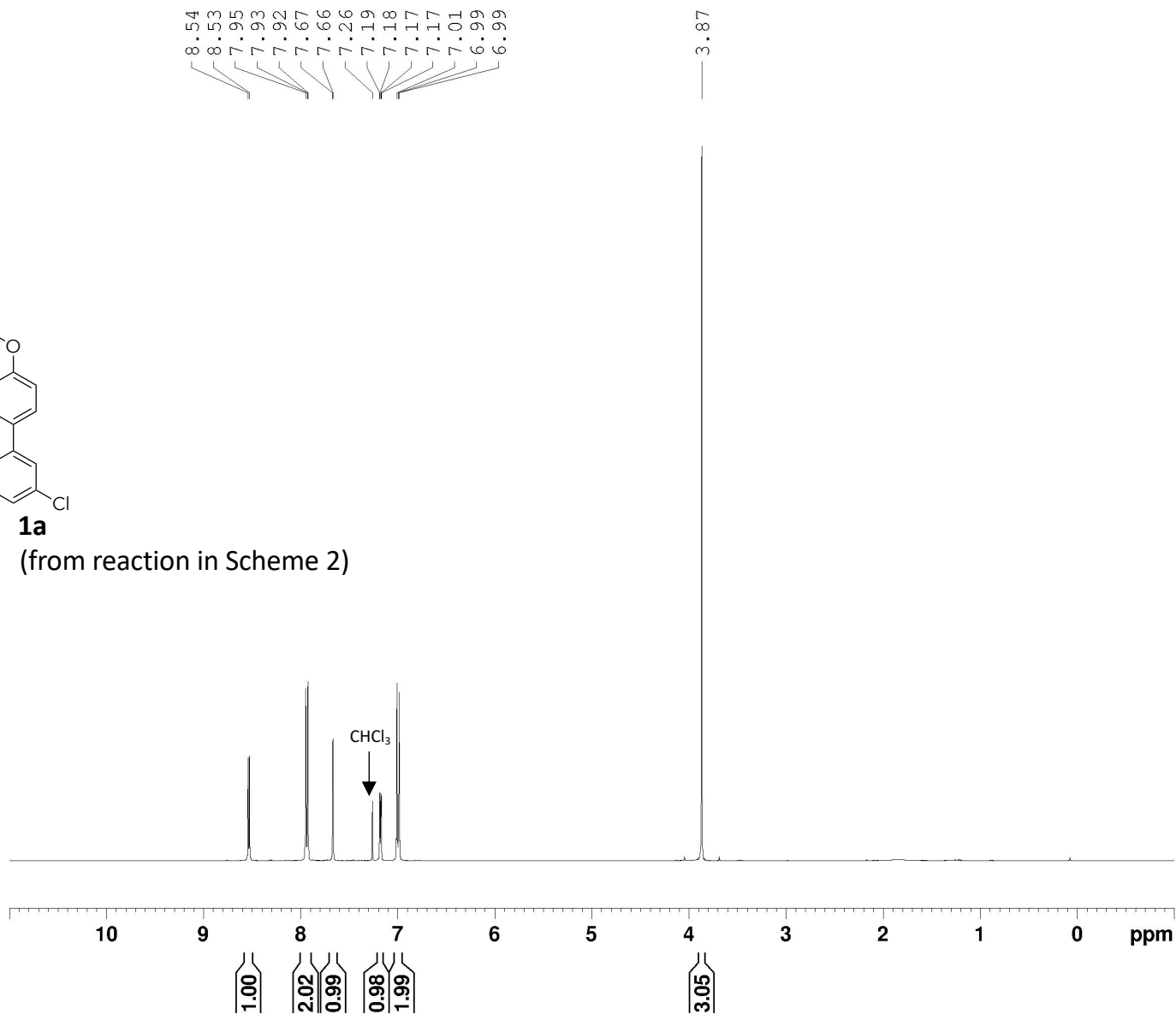
## II. References

1. Melvin, P. R.; Nova, A.; Balcells, D.; Dai, W.; Hazari, N.; Hruszkewycz, D. P.; Shah, H. P.; Tudge, M. T. *ACS Catal.* **2015**, *5*, 3680–3688.
2. Jiang, X.; Hu, F. *Synlett.* **2018**, *29*, 1207-1210.
3. Baranov, A. B.; Tsypin, V. G.; Malin, A. S.; Laskin, B. M. *Russ. J. Appl. Chem.* **2004**, *77*, 1997-2000.
4. Krasovskiy, A.; Knochel, P. *Synthesis* **2006**, *5*, 0890-0891.
5. Tran, T.-A.; Shin, Y.-J.; Kramer, B.; Choi, J.; Zou, N.; Vallar, P.; Martens, P.; Boatman, P. D.; Adams, J. W.; Ramirez, J.; Shi, Y.; Morgan, M.; Unett, D. J.; Chang, S.; Shu, H.-H.; Tung, S.-F.; Semple, G. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 1030–1035.
6. Habash, M.; Taha, M. O. *Bioorg. Med. Chem.* **2011**, *19*, 4746–4771.
7. Yahyazadeh, A.; Yousefi, H. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* **2014**, *117*, 696–701.
8. Rossiter, S.; Péron J. M.; Whitfield P. J.; Jones, K.; *Bioorganic Med. Chem Lett.* **2005**, *15*, 4806-4808.
9. Su, Z.; Yeagley, A. A.; Su, R.; Peng, L.; Melander, C. *ChemMedChem.* **2012**, *7*, 2030-2039.
10. Dai, X.; Chen, Y.; Garrell, S.; Liu, H.; Zhang, L.-K.; Palani, A.; Hughes, G.; Nargund, R. *J. Org. Chem.* **2013**, *78*, 7758–7763.
11. Reeves, E. K.; Humke, J. N.; Neufeldt, S. R. *J. Org. Chem.* **2019**, *84*, 11799–11812.
12. Thapa, S.; Gurung, S. K.; Dickie, D. A.; Giri, R.; *Angew. Chem. Int. Ed.* **2014**, *53*, 11620–11624 .
13. Sicre, C.; Alonso-Gómez, J. L.; Cid, M. M. *Tetrahedron* **2006**, *62*, 11063-11072.
14. Crouse, G. D.; Sparks, T. C.; McLead, C. L.; Demeter, D. A.; Bryan, K.; Brown, A. V.; Dent III W. H.; Cudworth, D. P.; Nugent, J. S.; Hunter R.; Samaritoni, J. G. Pesticidal Compositions. U.S. Patent US 2009/0209476 A1, August 20, **2009**.
15. Yang, M.; Chen, J.; He, C.; Hu, X.; Ding, Y.; Kuang, Y.; Liu, J. Huang, Q. *J. Org. Chem.* **2020**, *85*, 6498–6508
16. Minxing, X.; Feng, T.; Wei, H.; Jun, W.; Han, S.; Zhaoyu, L.; Chongxing, Tu.; Xinge, Z. Pyrimidinamine and pyridinamine Hedgehog signal conduction inhibitors. International Patent CN103864770 (A), June 18, **2014**. (paragraph 1443).
17. Duan, Jingwu; Jian, Bing; Lu, Zhonghui. Azaindoles as BTK Kinase Modulators and Use Thereof. International Patent WO 2011/019780 A1, February 17, **2011**. (pages 63-64).
18. Scott, N. W. J.; Ford, M. J.; Jeddi, N.; Eyles, A.; Simon, L.; Whitwood, A. C.; Tanner, T.; Willans, C. E.; Fairlamb, I. J. S. *J. Am. Chem. Soc.* **2021**, *143*, 9682-9693.
19. Louërat, F.; Gros, P.C. *Tetrahedron* **2010**, *51*, 3558-3560.
20. Baloch, M.; Roy, D.; Bensaid, S.; Guerchais, V.; Doucet, H. *Eur. J of Inorg. Chem.* **2012**, *28*, 4454-4462.
21. Levy, J. N.; Alegre-Requena, J. V.; Liu, R.; Paton, R. S.; McNally, A. *J. Am. Chem. Soc.* **2020**, *142*, 11295-11305.
22. Corcoran, R. C.; Bang, S. H. *Tetrahedron Lett.* **1990**, *31*, 6757-6758.
23. Gurung, S. K.; Thapa, S.; Vangala, A. S.; Giri, R. *Org. Lett.* **2013**, *15*, 20, 5378-5381
24. Mosquera, Á.; Riveiros, R.; Sestelo, J. P.; Sarandeses, L. A. *Org. Lett.* **2008** *10*, 17, 3745-3748

### III. NMR Spectra



(from reaction in Scheme 2)



Current Data Parameters  
 NAME NL-1-96-1H  
 EXPNO 12  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210812  
 Time 14.41 h  
 INSTRUM Avance Neo  
 PROBHD z152088\_0031 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 16  
 DS 2  
 SWH 8196.722 Hz  
 FIDRES 0.250144 Hz  
 AQ 3.9976959 sec  
 RG 101  
 DW 61.000 usec  
 DE 13.89 usec  
 TE 298.2 K  
 D1 1.00000000 sec  
 TD0 1  
 SFO1 400.1324708 MHz  
 NUC1 1H  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 24.03499985 W

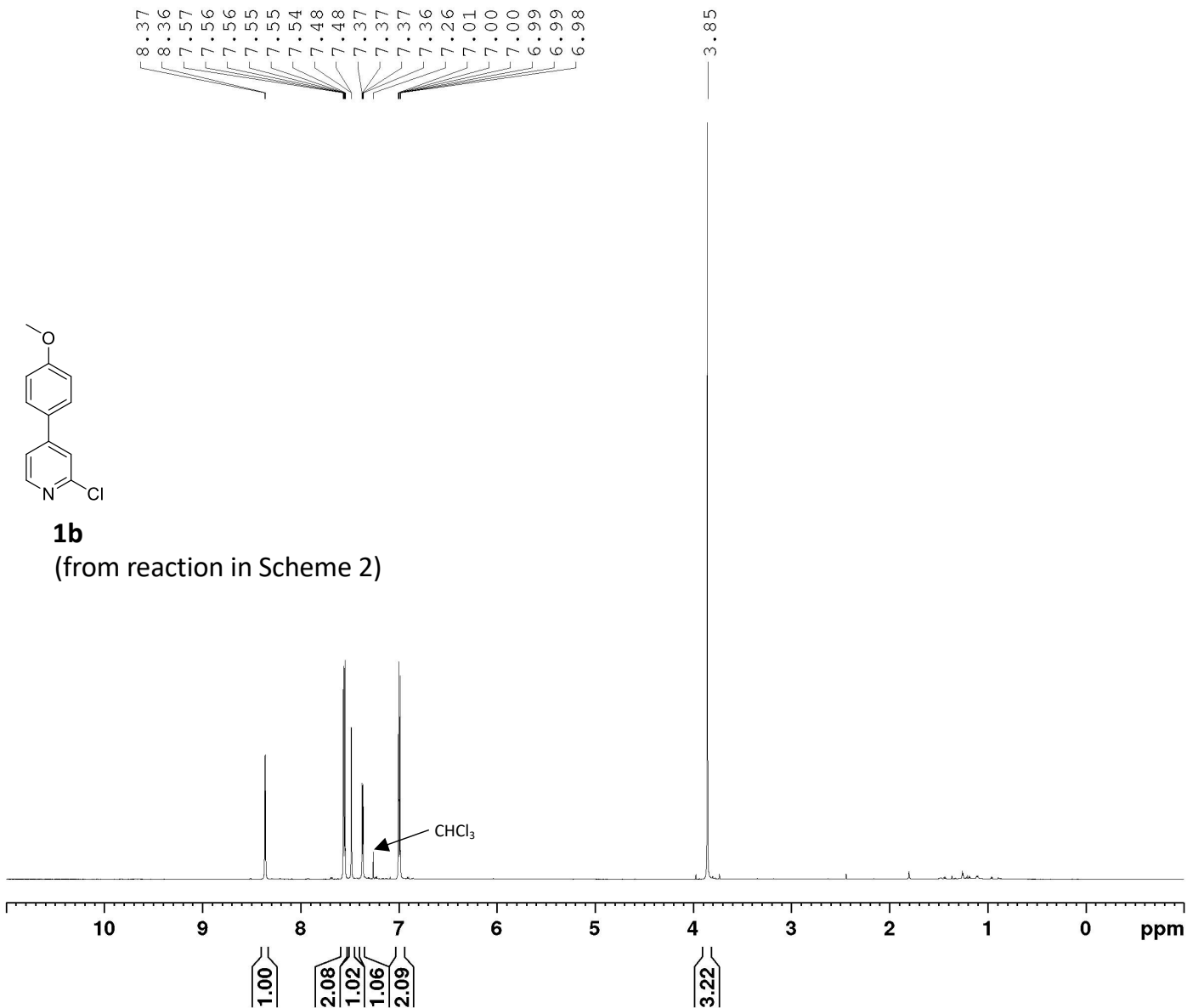
F2 - Processing parameters  
 SI 65536  
 SF 400.1300097 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

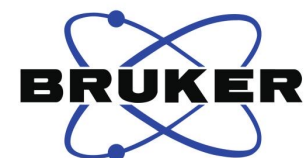


Current Data Parameters  
NAME JPN-1-174-7\_C4-mono  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200610  
Time 12.30 h  
INSTRUM spect  
PROBHD z127277\_0002 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.366798 Hz  
AQ 2.7262976 sec  
RG 4.41  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1  
SF01 600.1337060 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 5.59999990 W

F2 - Processing parameters  
SI 65536  
SF 600.1300144 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

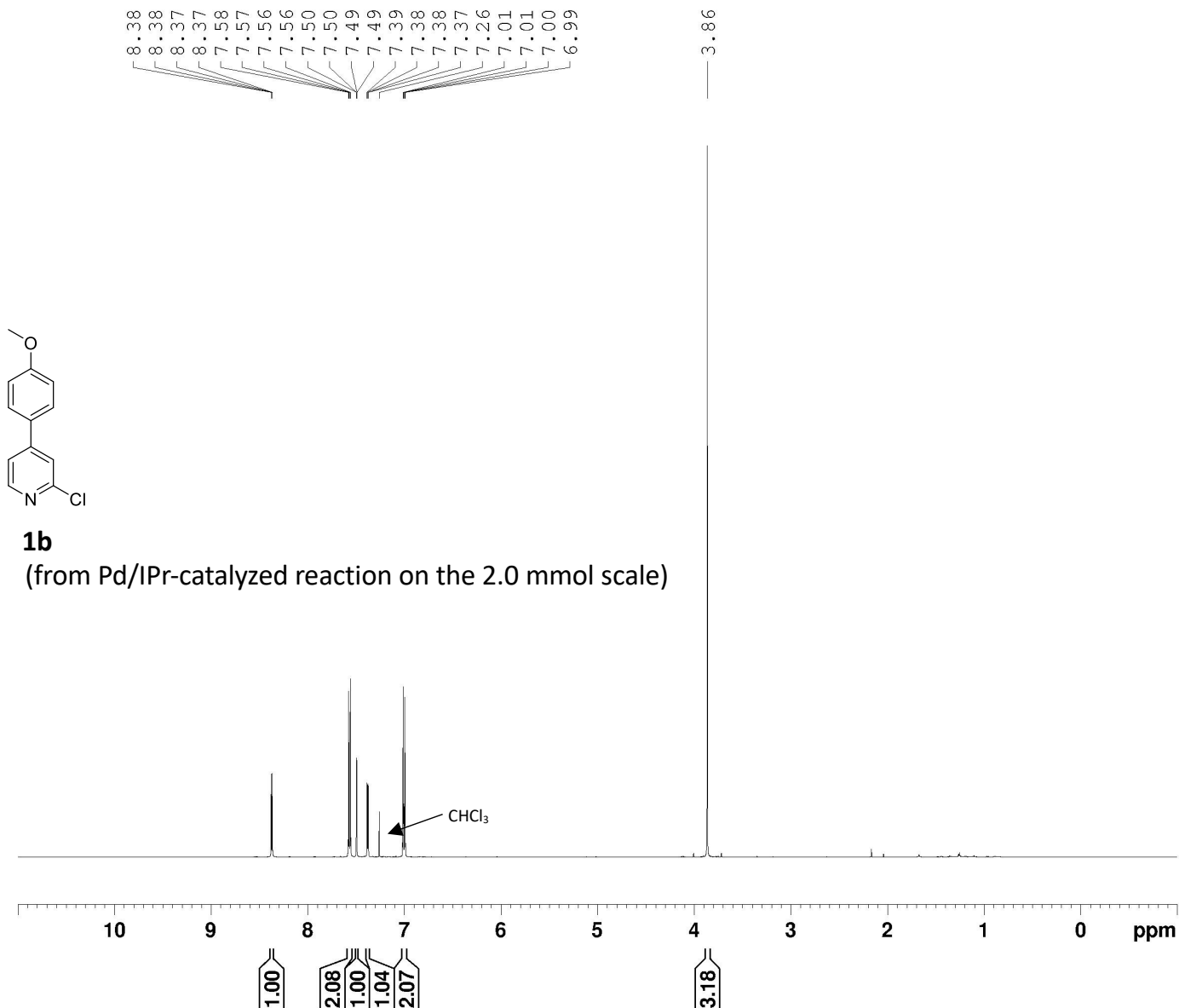




Current Data Parameters  
NAME NL-2-14-1-IPr  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20220419  
Time 17.32 h  
INSTRUM spect  
PROBHD z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 86.13  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300119 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00





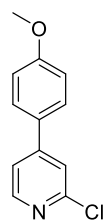
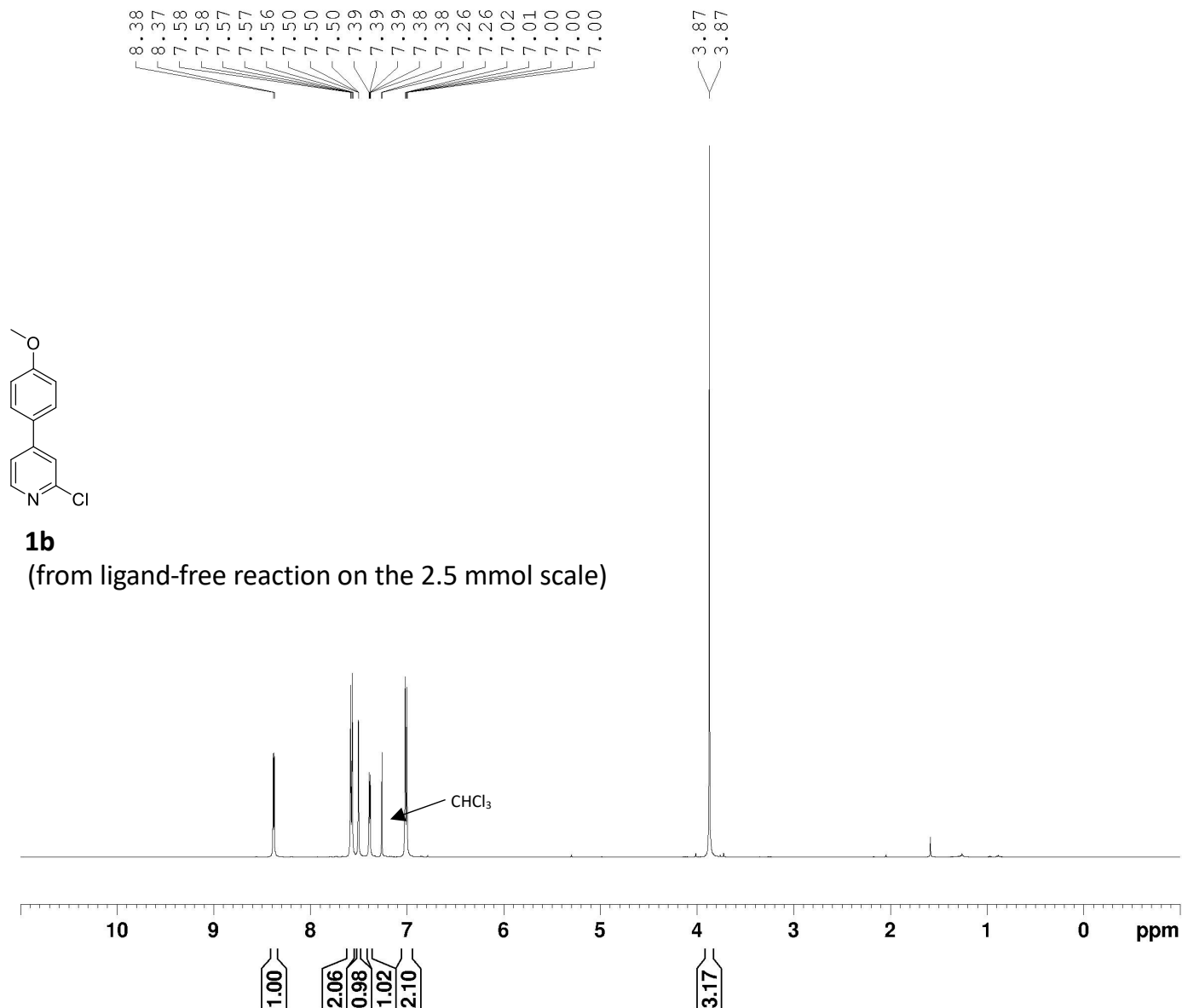
Current Data Parameters  
NAME NL-2-14-2-LF  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters

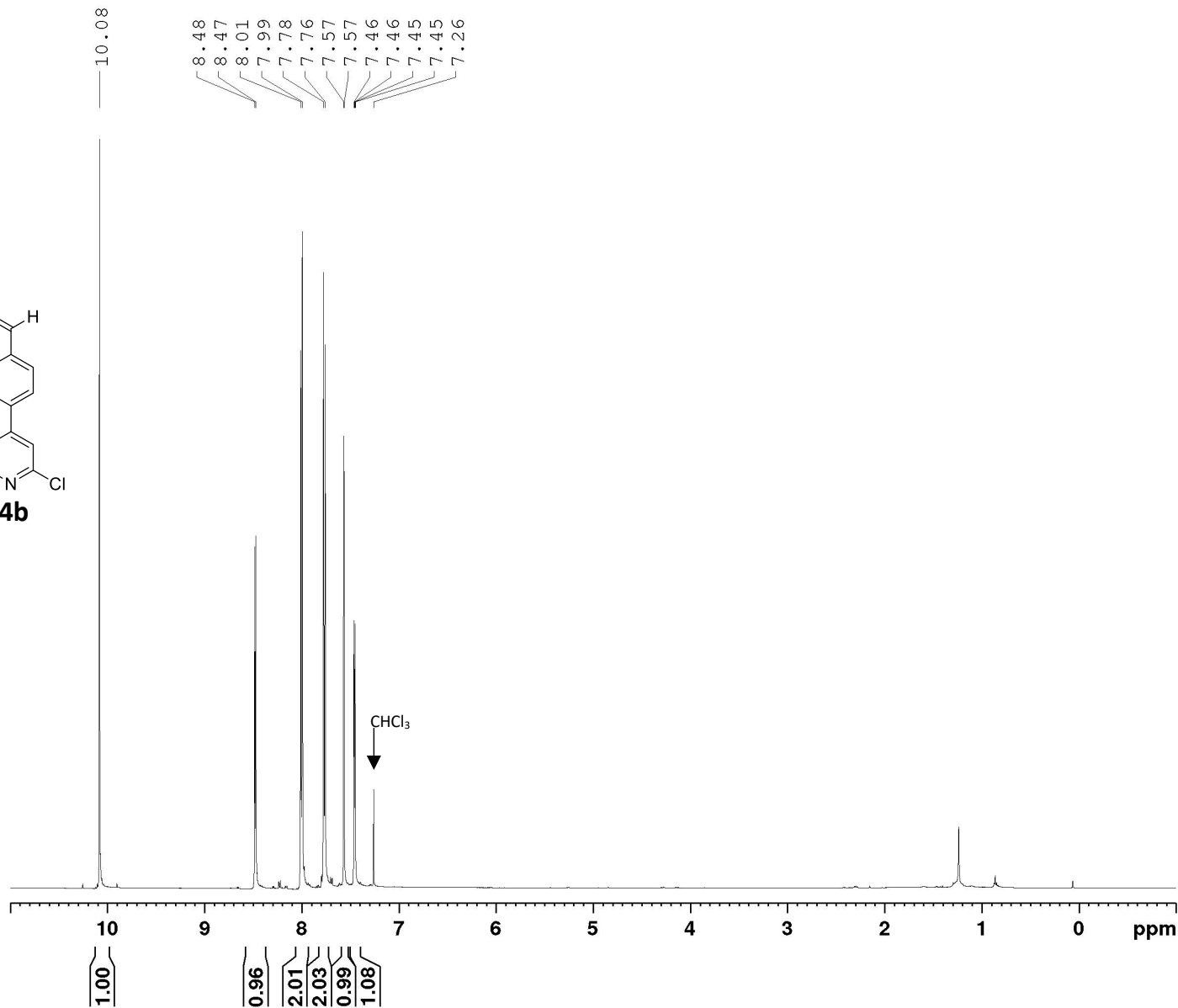
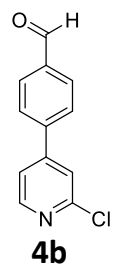
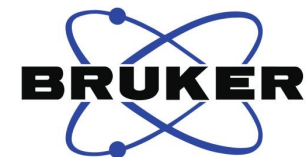
Date\_ 20220419  
Time 17.42 h  
INSTRUM spect  
PROBHD z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 151.18  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters

SI 65536  
SF 500.2300117 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



**1b**  
(from ligand-free reaction on the 2.5 mmol scale)

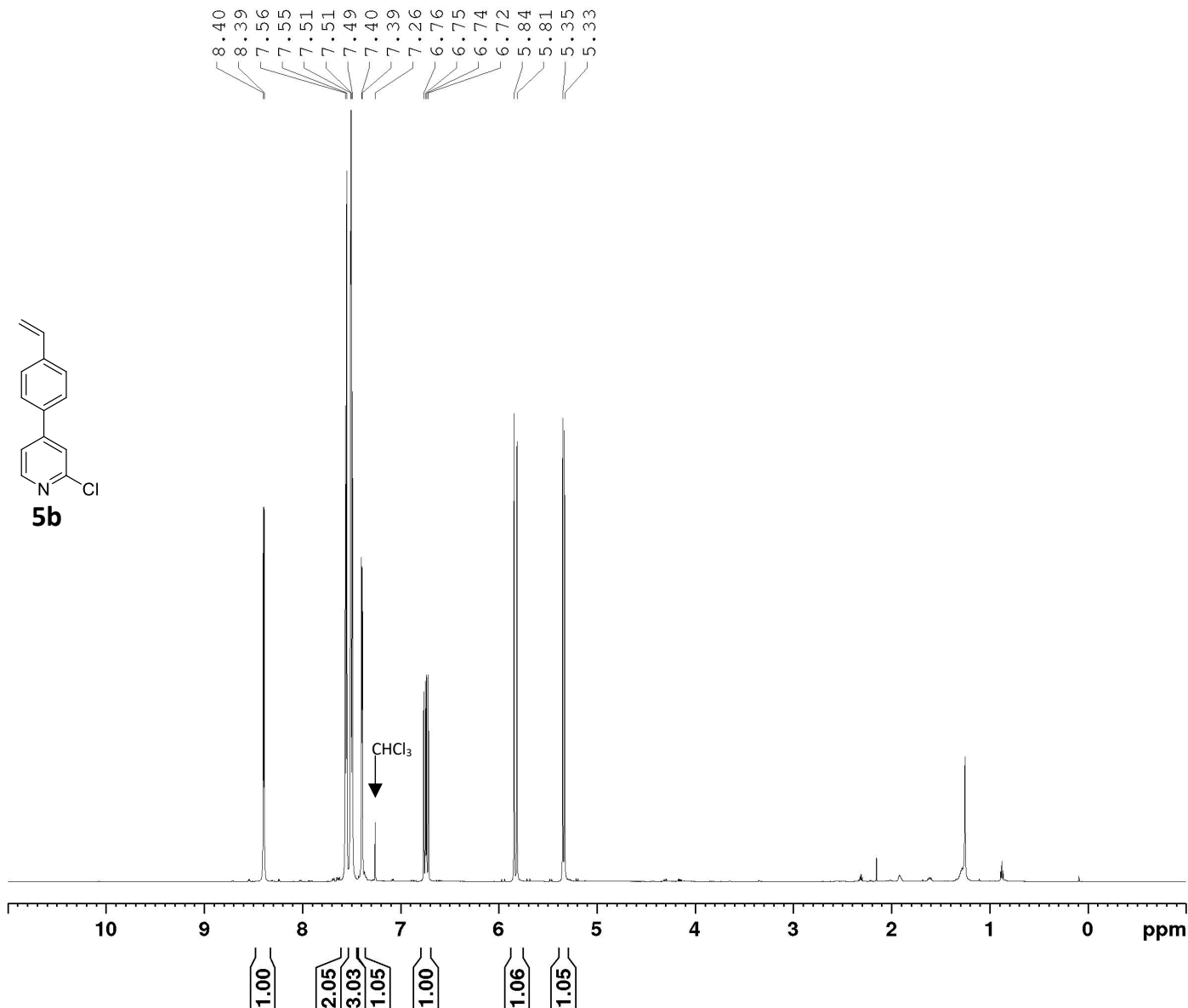
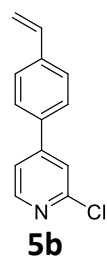


Current Data Parameters  
NAME JPN-1-174-5  
EXPNO 17  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211120  
Time 5.51 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 76.94  
DW 50.000 usec  
DE 16.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300120 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

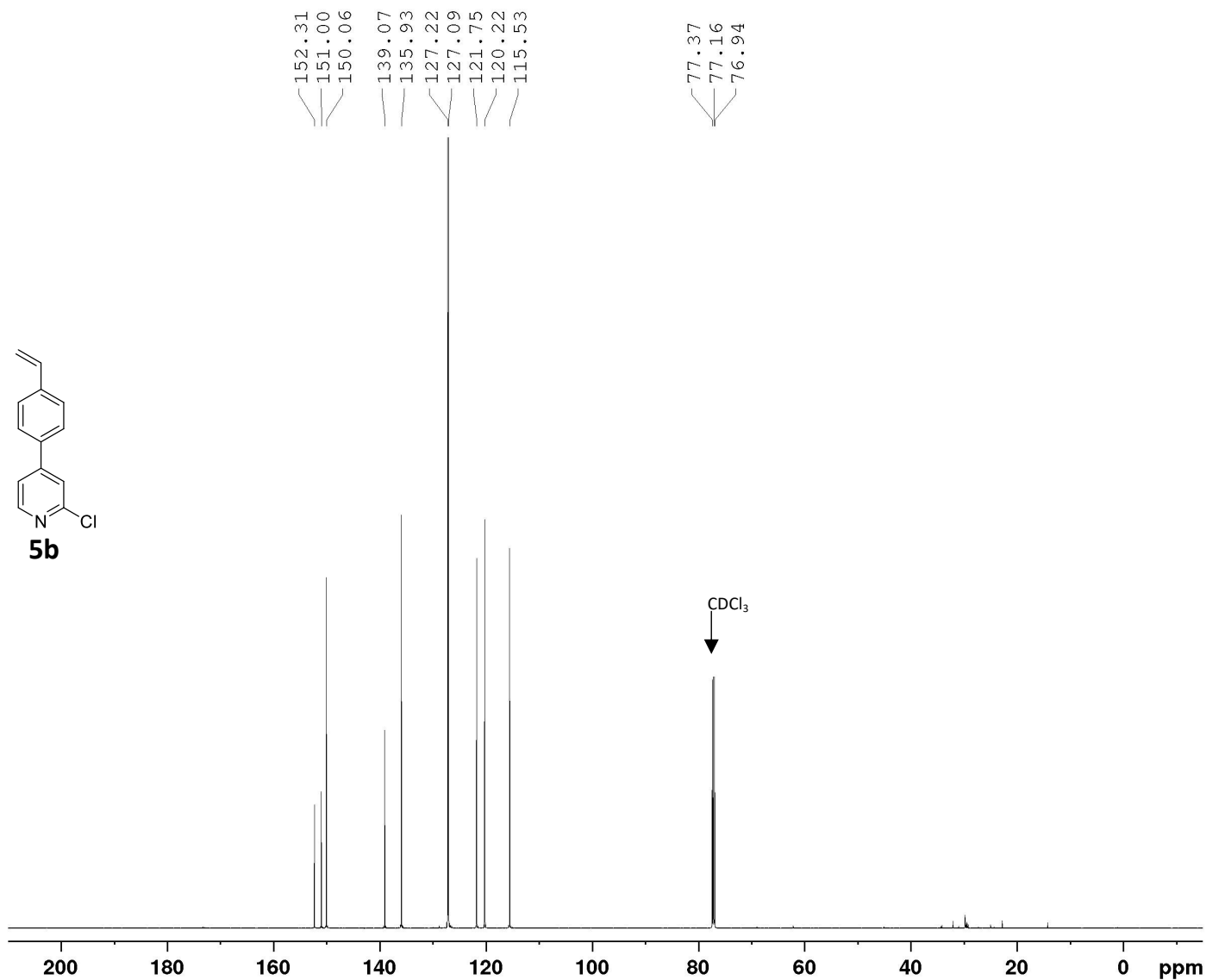
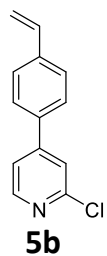




Current Data Parameters  
NAME JPN-1-171-3\_C4-mono\_dried  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200608  
Time 16.21 h  
INSTRUM spect  
PROBHD z127277\_0002 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.366798 Hz  
AQ 2.7262976 sec  
RG 4.41  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1  
SFO1 600.1337060 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 5.59999990 W

F2 - Processing parameters  
SI 65536  
SF 600.1300145 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



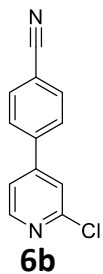
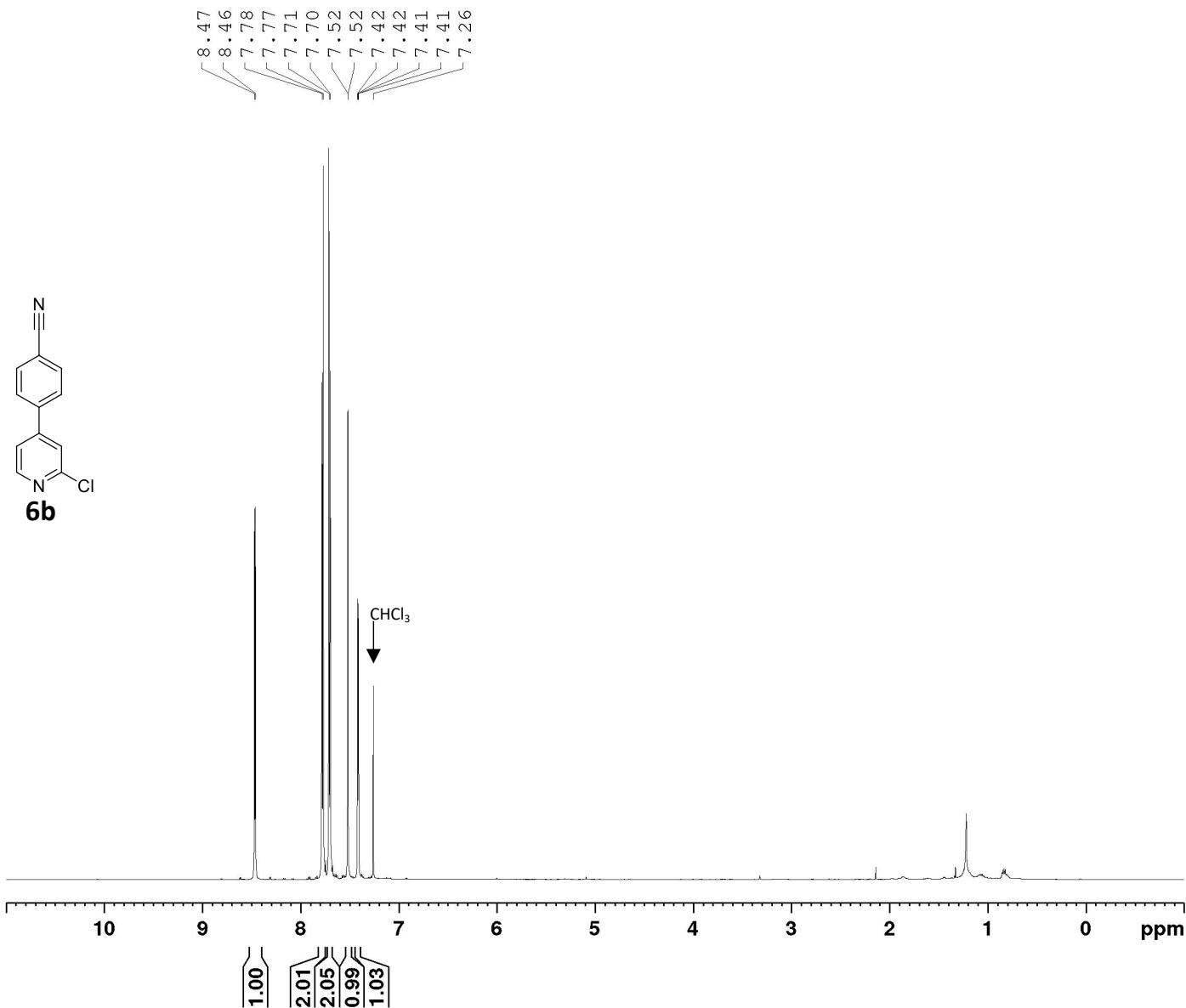
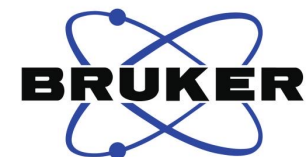
Current Data Parameters  
 NAME JPN-1-171-3\_C4-mono\_dried  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters

Date\_ 20200608  
 Time 17.13 h  
 INSTRUM spect  
 PROBHD Z127277\_0002 ( )  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 36057.691 Hz  
 FIDRES 1.100393 Hz  
 AQ 0.9087659 sec  
 RG 184.4  
 DW 13.867 usec  
 DE 18.00 usec  
 TE 300.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 150.9178981 MHz  
 NUC1 13C  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 91.00000000 W  
 SFO2 600.1324005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 70.00 usec  
 PLW2 5.59999990 W  
 PLW12 0.07314300 W  
 PLW13 0.03679000 W

F2 - Processing parameters

SI 32768  
 SF 150.9028016 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



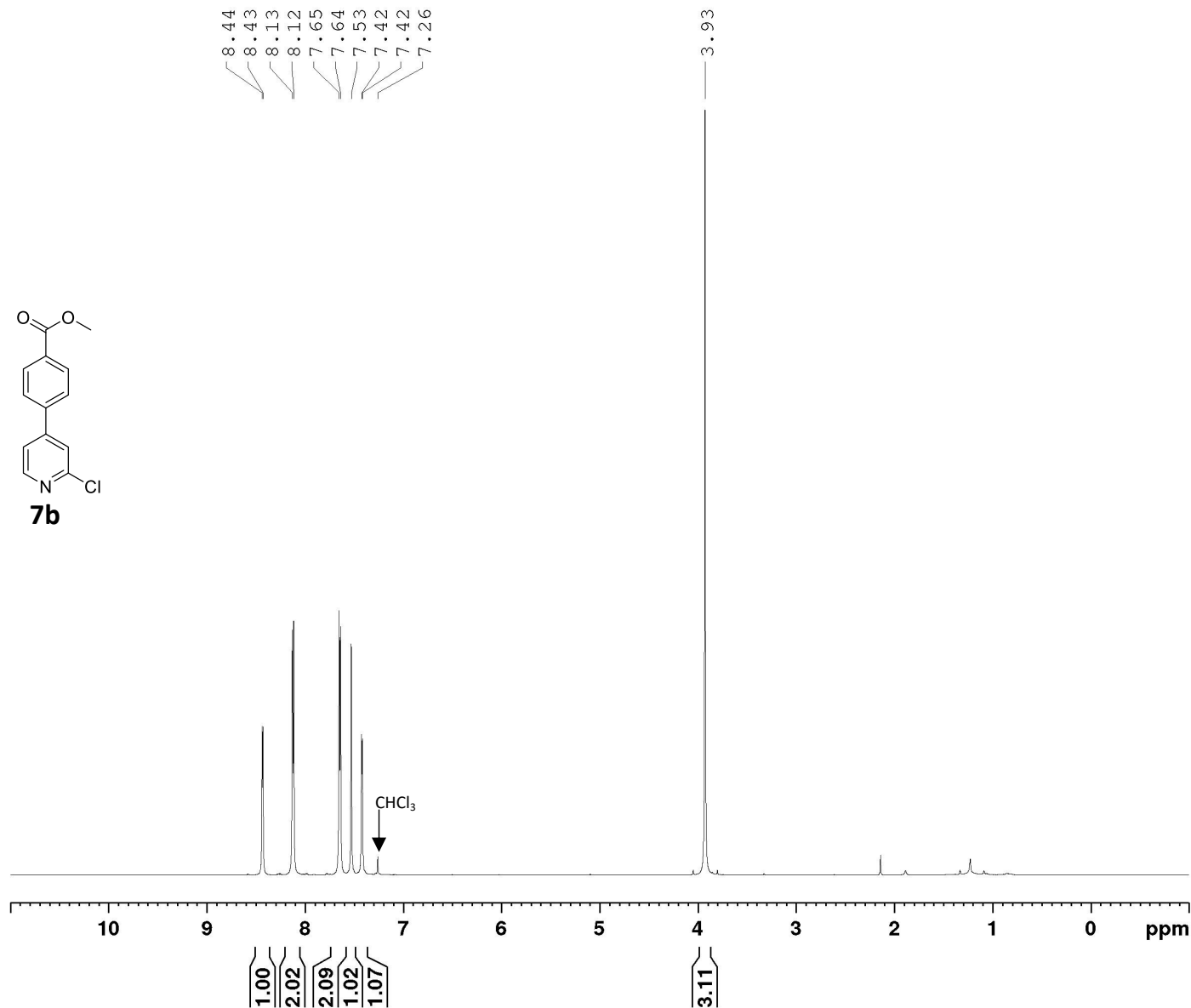
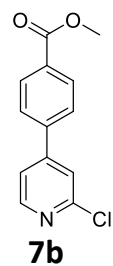
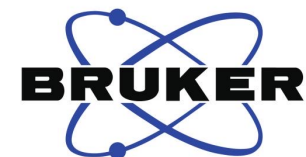
Current Data Parameters  
NAME JPN-1-171-2\_C4-mono\_dried  
EXPNO 22  
PROCNO 1

F2 - Acquisition Parameters

Date\_ 20200608  
Time 17.21 h  
INSTRUM spect  
PROBHD z127277\_0002 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.366798 Hz  
AQ 2.7262976 sec  
RG 4.41  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1  
SFO1 600.1337060 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 5.59999990 W

F2 - Processing parameters

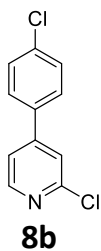
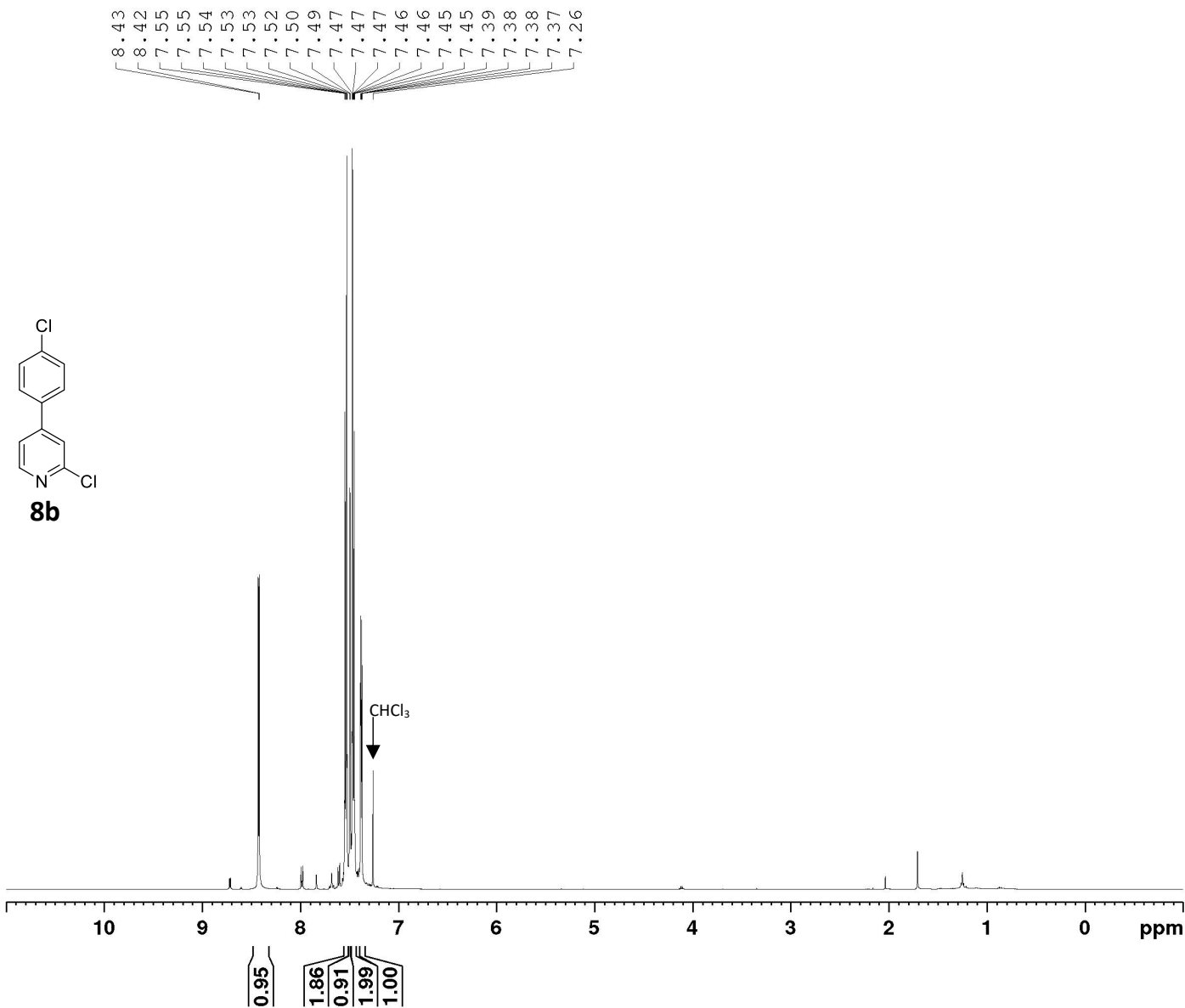
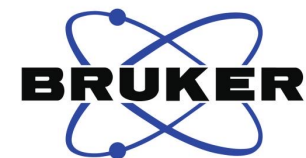
SI 65536  
SF 600.1300145 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME JPN-1-174-4\_C4-mono  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200610  
Time 12.06 h  
INSTRUM spect  
PROBHD Z127277\_0002 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.366798 Hz  
AQ 2.7262976 sec  
RG 4.41  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1  
SF01 600.1337060 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 5.59999990 W

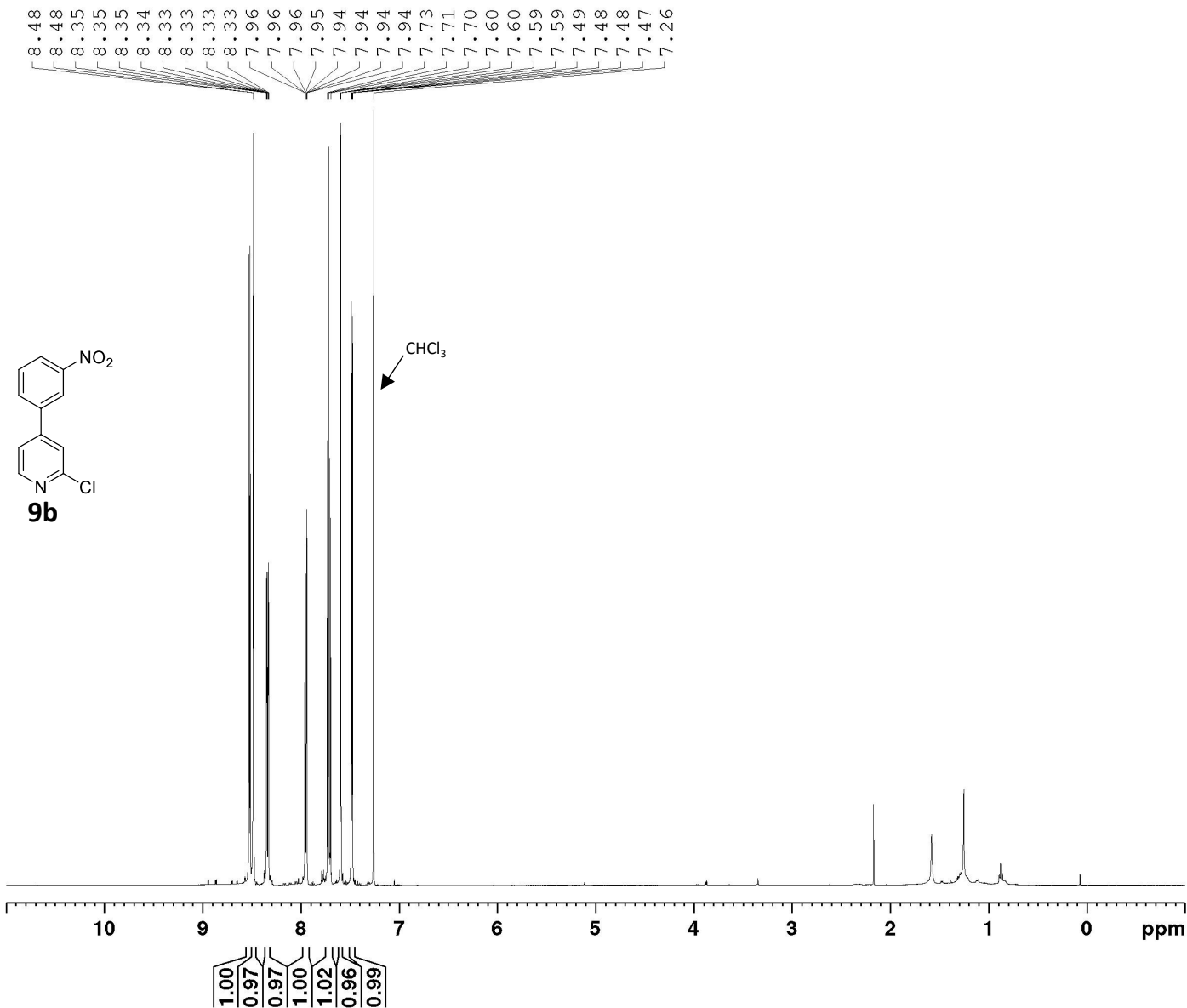
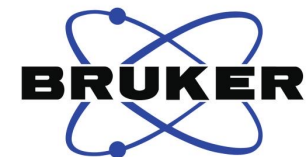
F2 - Processing parameters  
SI 65536  
SF 600.1300144 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME JPN-1-174-3a\_C4-mono  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200811  
Time 23.22 h  
INSTRUM spect  
PROBHD Z125869\_0055 (   
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 86.13  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TDO 1  
SFO1 500.2330889 MHz  
NUC1 1H  
PO 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300122 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters

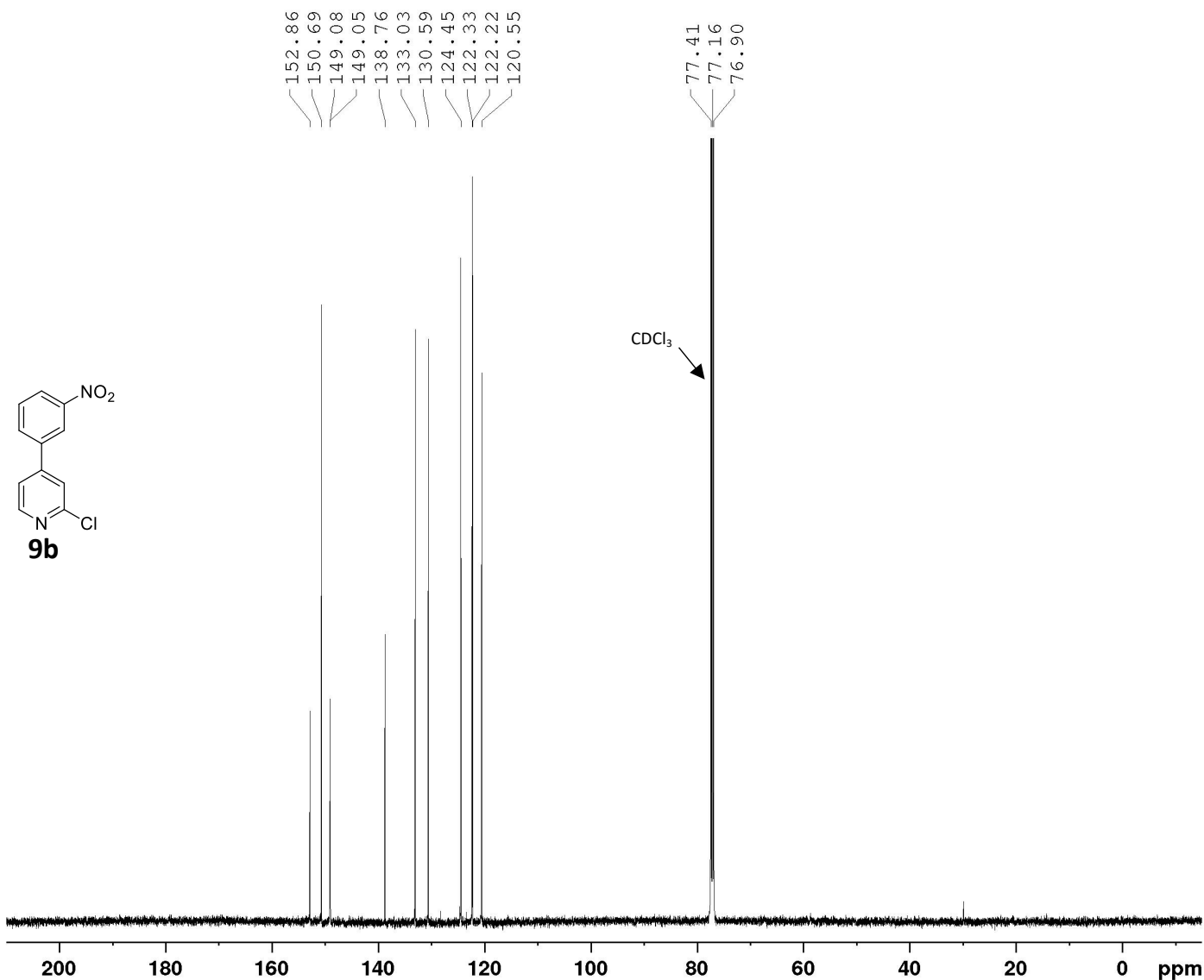
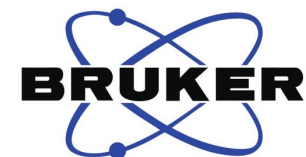
NAME JPN-1-193-1  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters

Date\_ 20211121  
Time 1.47 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 151.18  
DW 50.000 usec  
DE 16.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters

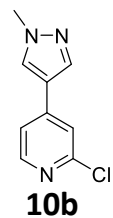
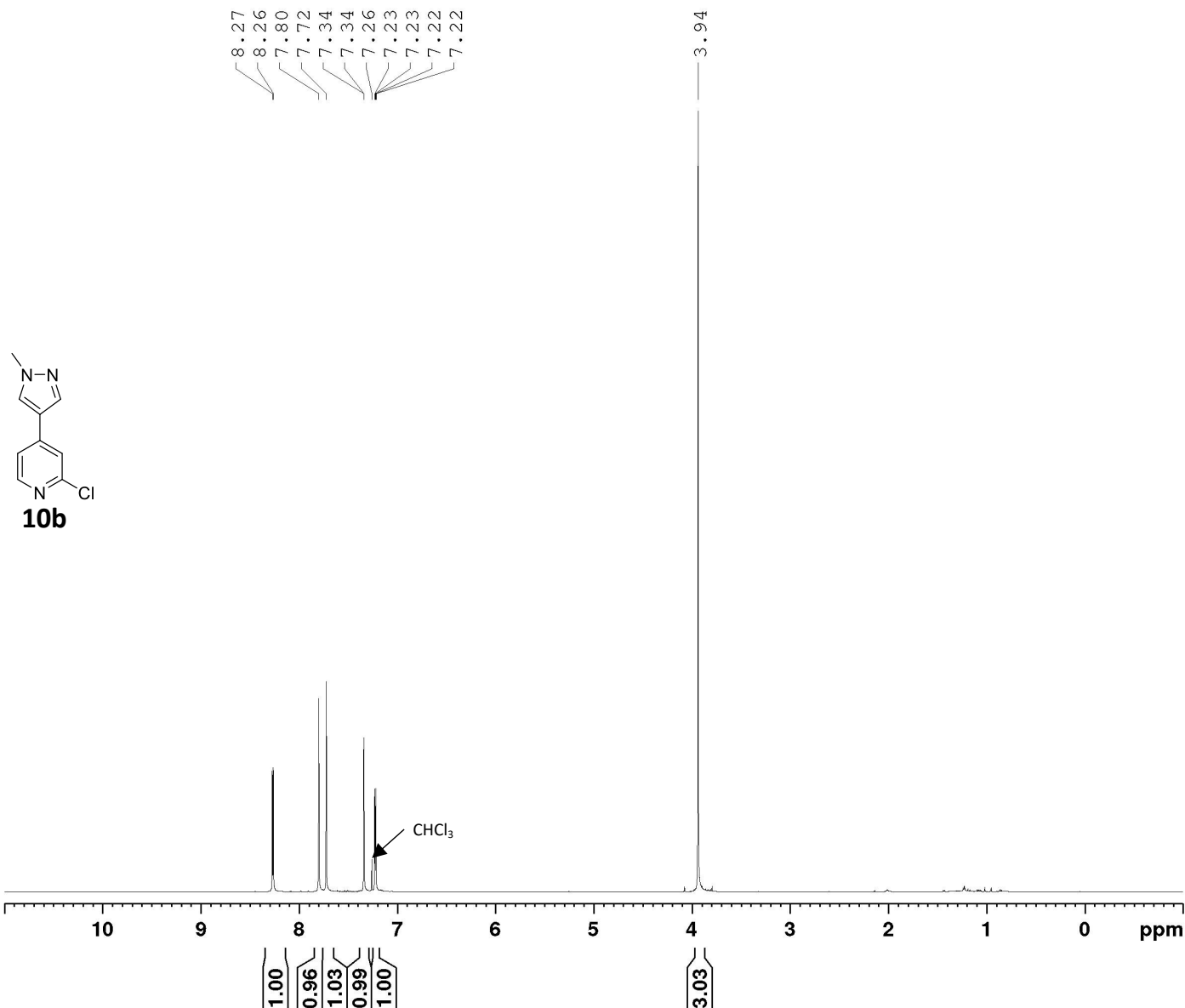
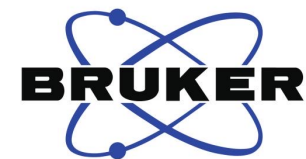
SI 65536  
SF 500.2300121 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME JPN-1-193-1  
EXPNO 11  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211121  
Time 2.43 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.908261 Hz  
AQ 1.1010048 sec  
RG 190.44  
DW 16.800 usec  
DE 18.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 125.7955118 MHz  
NUC1 13C  
P0 3.33 usec  
P1 10.00 usec  
PLW1 56.90299988 W  
SFO2 500.2320009 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 11.44699955 W  
PLW12 0.25756001 W  
PLW13 0.12955000 W

F2 - Processing parameters  
SI 32768  
SF 125.7829172 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

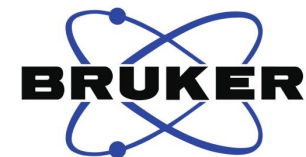


Current Data Parameters  
NAME JPN-2-11a\_C4-mono\_MeOH\_column\_cc  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200829  
Time 17.06 h  
INSTRUM spect  
PROBHD z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 37.93  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SF01 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300122 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

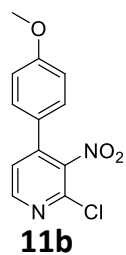
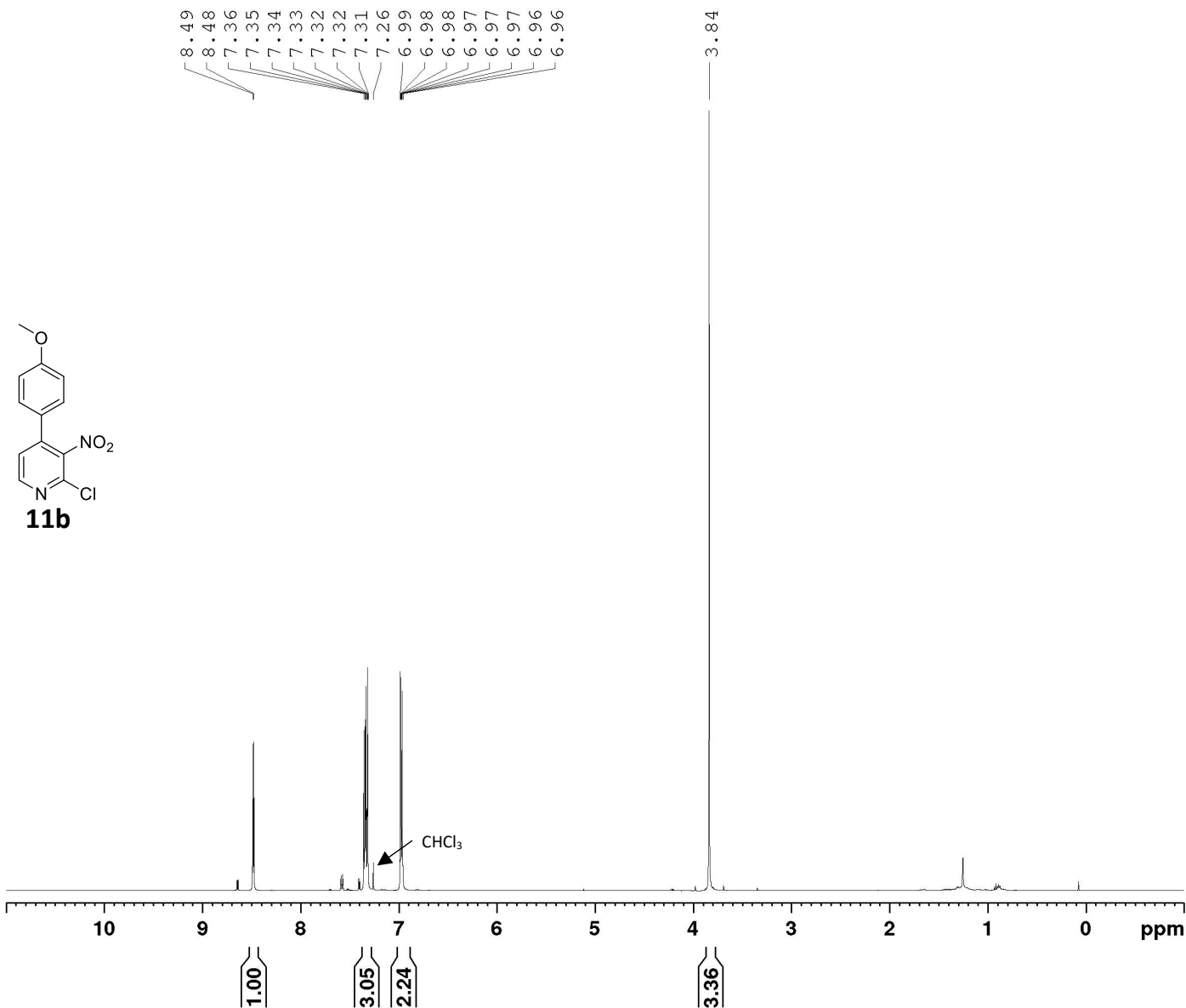


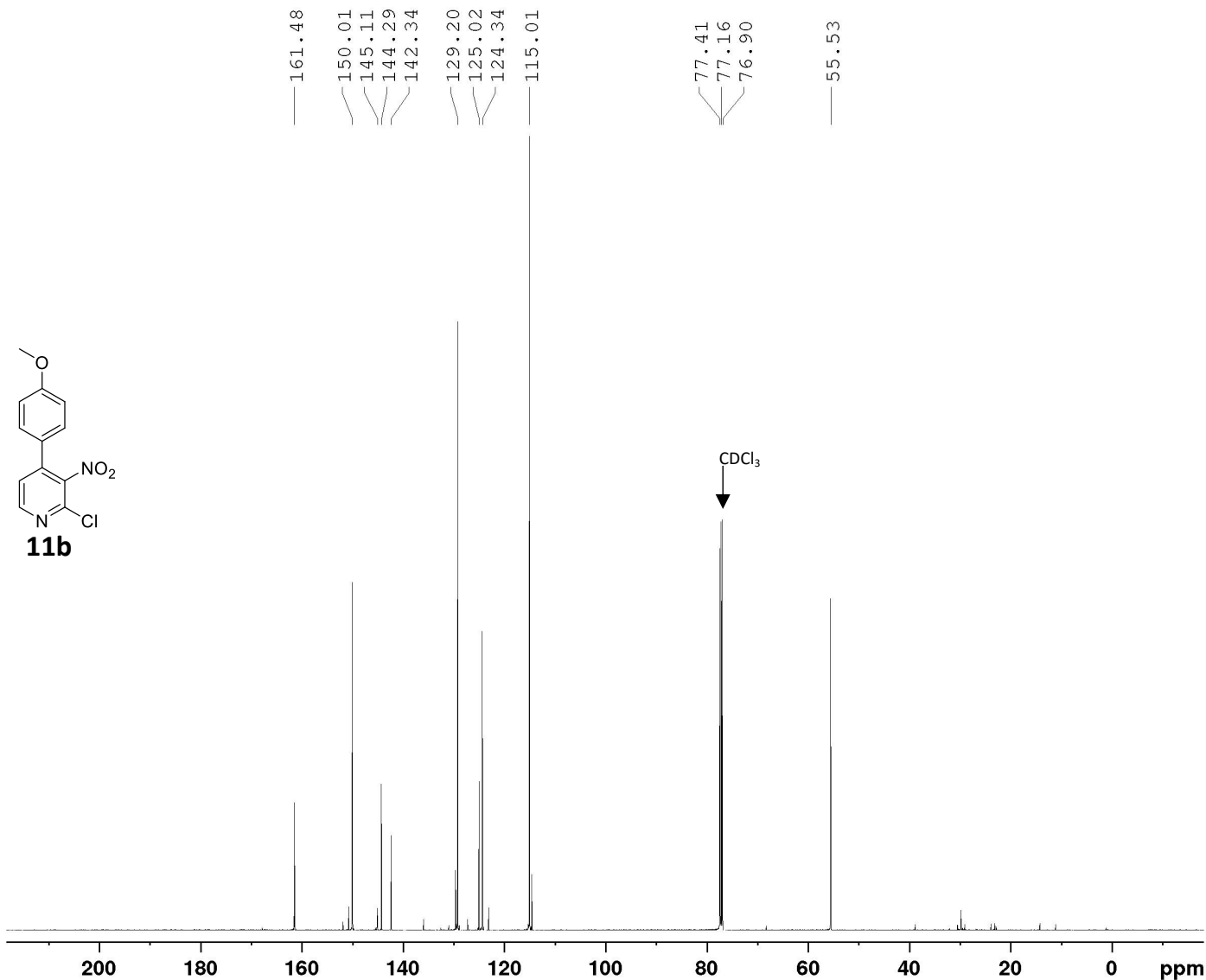
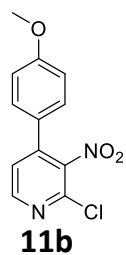


Current Data Parameters  
NAME JPN-1-163-3  
EXPNO 16  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211119  
Time 2.34 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 37.93  
DW 50.000 usec  
DE 16.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300119 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

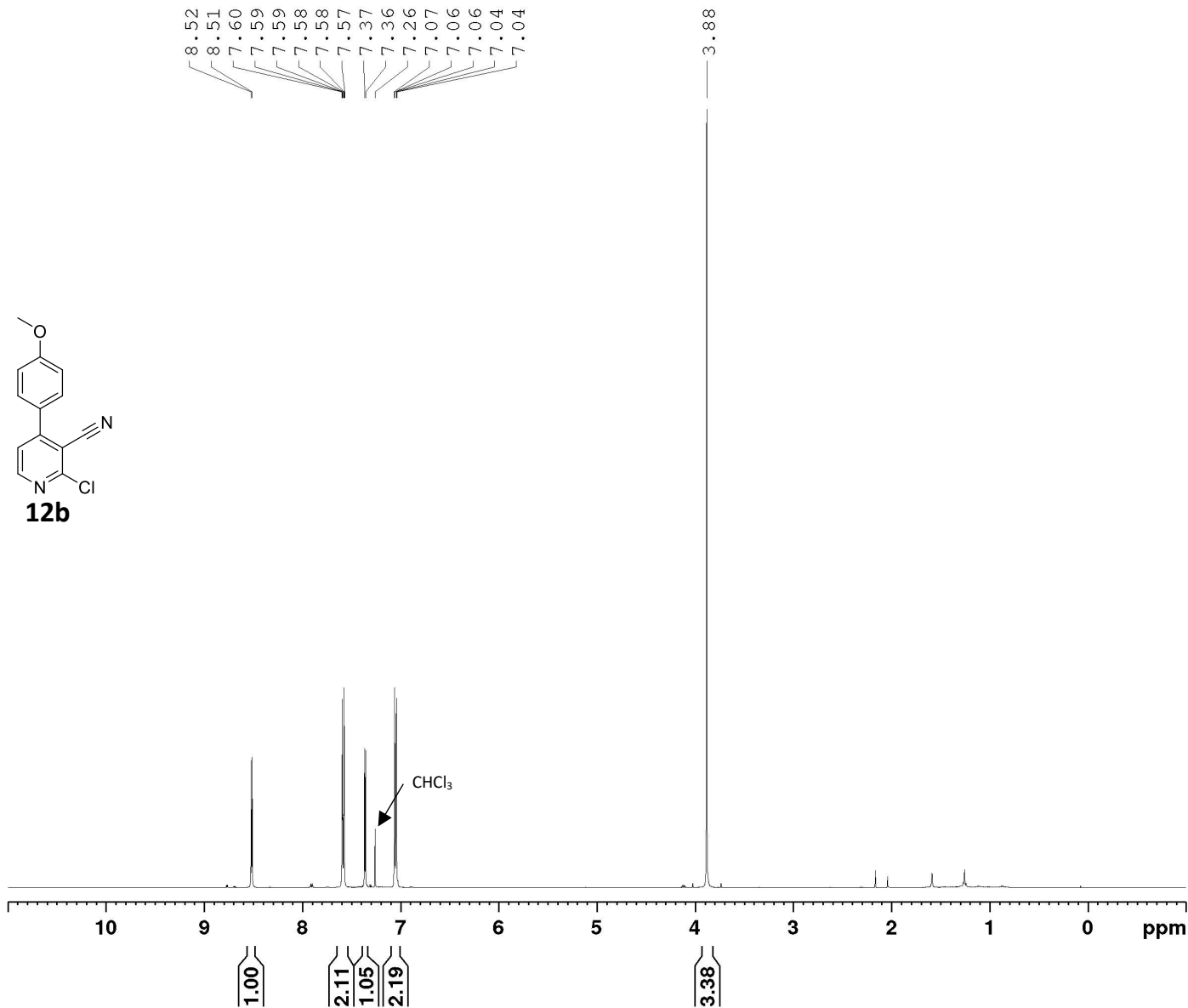
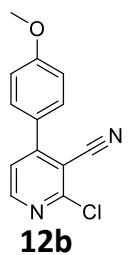




Current Data Parameters  
 NAME JPN-1-163-3  
 EXPNO 17  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211119  
 Time 3.30 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

F2 - Processing parameters  
 SI 32768  
 SF 125.7829219 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



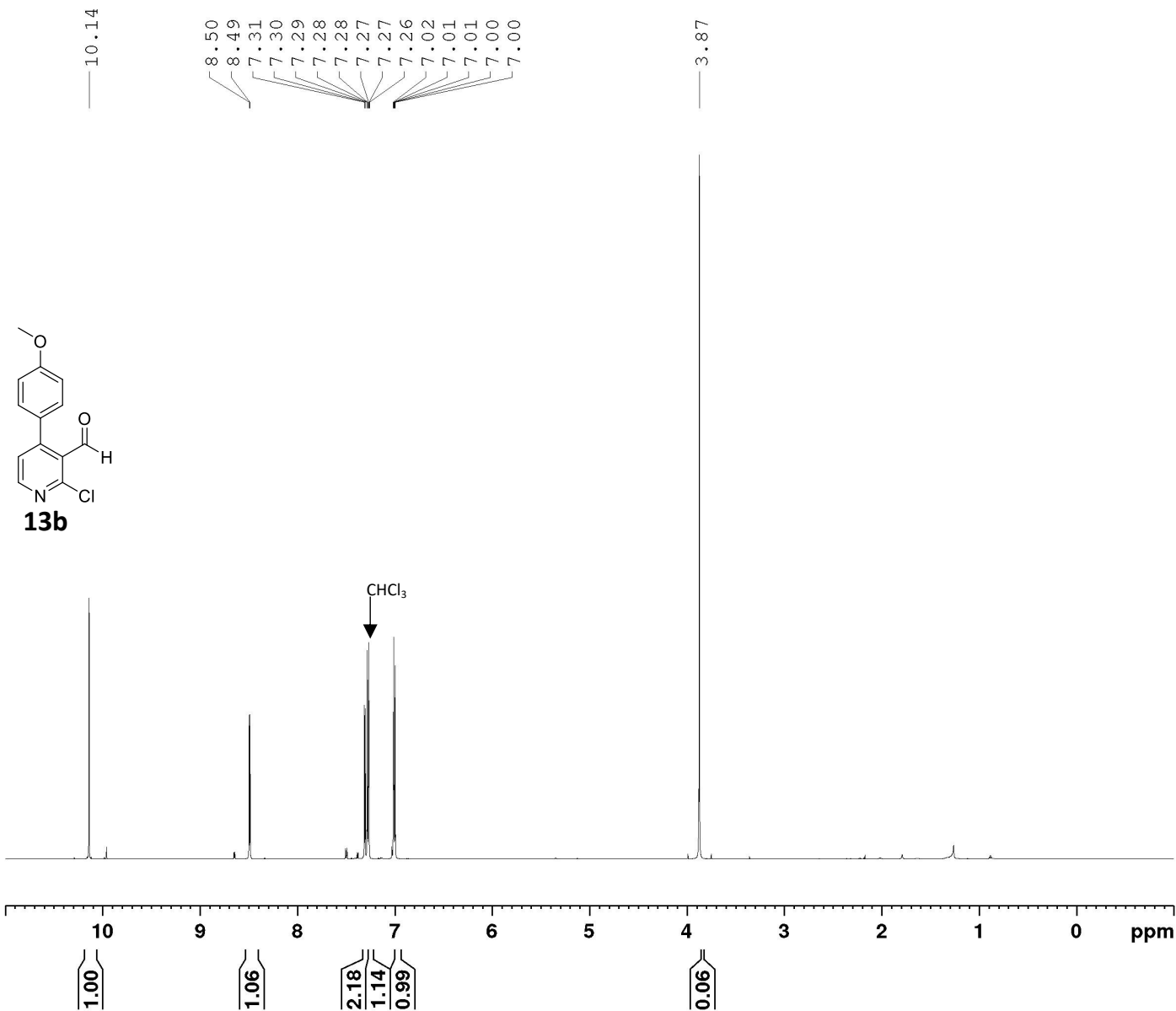
Current Data Parameters  
 NAME JPN-1-166-1\_repeat\_c4-mono  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters

Date\_ 20200601  
 Time 9.25 h  
 INSTRUM spect  
 PROBHD Z119470\_0370 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDC13  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 137.36  
 DW 50.000 usec  
 DE 6.50 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 3.13 usec  
 P1 9.38 usec  
 PLW1 20.00000000 W

F2 - Processing parameters

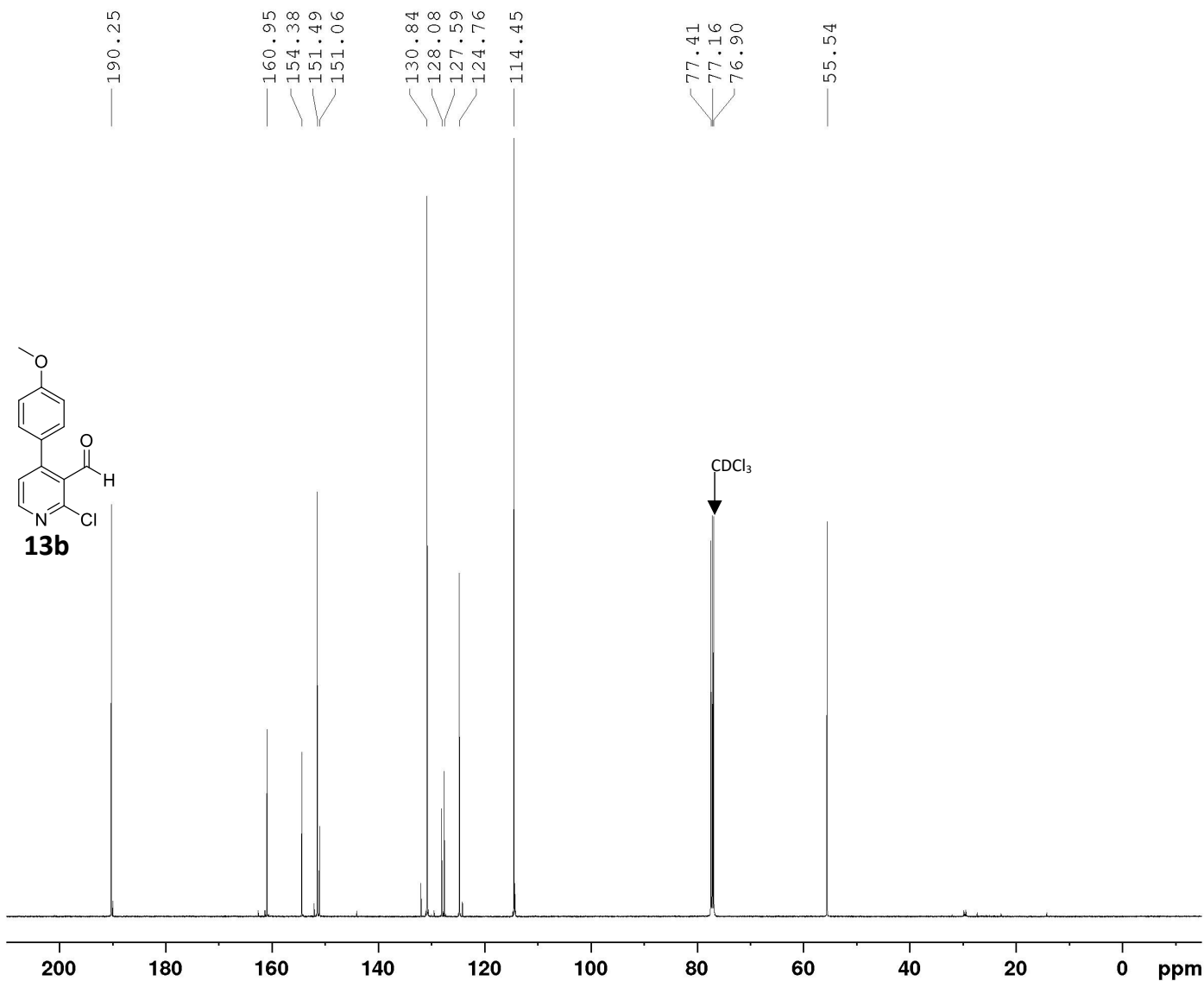
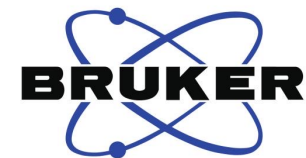
SI 32768  
 SF 500.2300128 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
NAME JPN-1-167-1\_C4-Mono  
EXPNO 24  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200608  
Time 20.22 h  
INSTRUM spect  
PROBHD Z127277\_0002 ( )  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.366798 Hz  
AQ 2.7262976 sec  
RG 4.41  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1  
SFO1 600.1337060 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 5.59999990 W

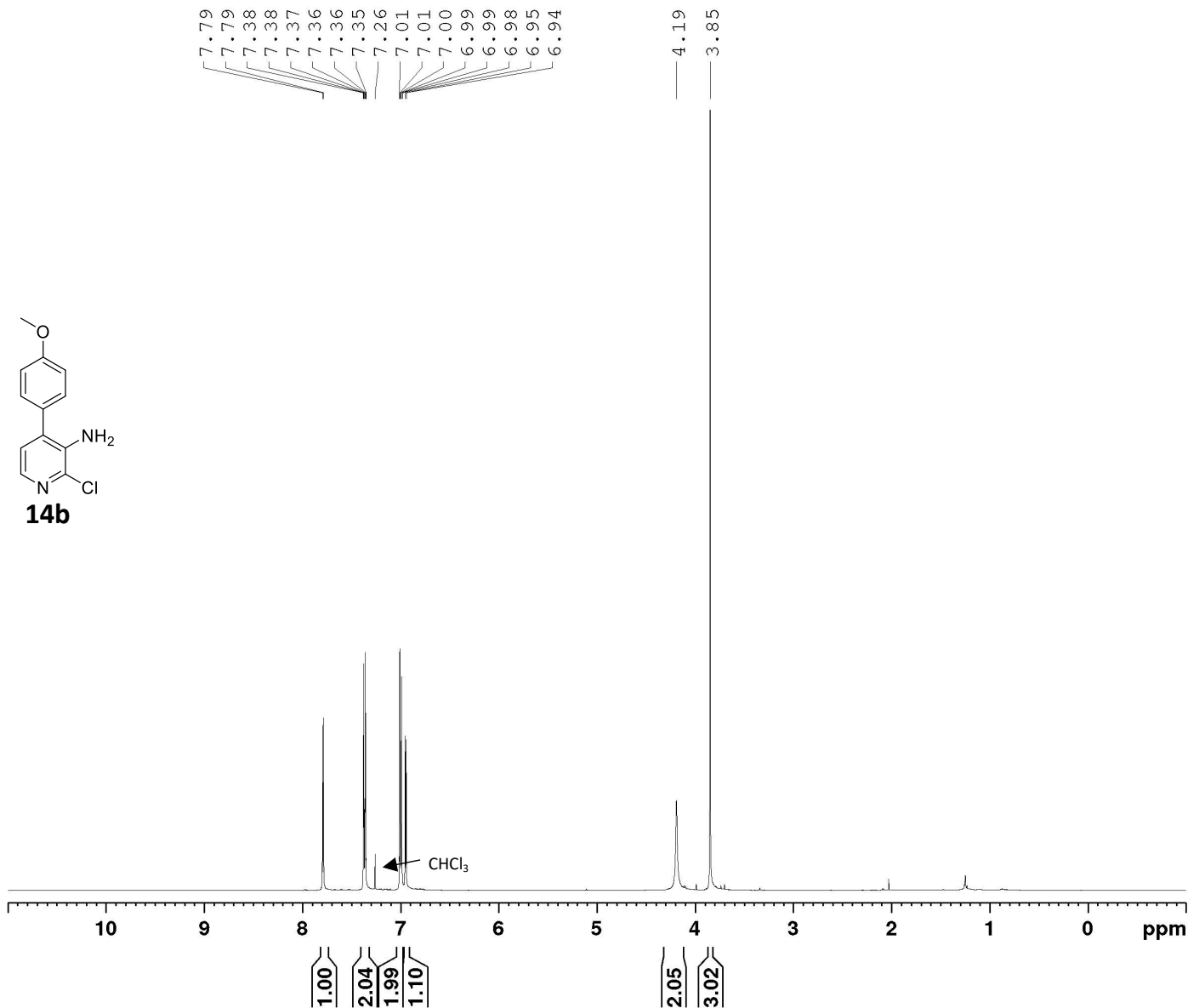
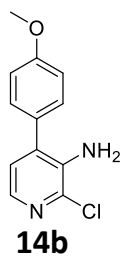
F2 - Processing parameters  
SI 65536  
SF 600.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME JPN-1-167-1\_C4-Mono  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200811  
Time 21.16 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.908261 Hz  
AQ 1.1010048 sec  
RG 190.44  
DW 16.800 usec  
DE 18.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 125.7955118 MHz  
NUC1 13C  
P0 3.33 usec  
P1 10.00 usec  
PLW1 56.90299988 W  
SFO2 500.2320009 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 11.44699955 W  
PLW12 0.25756001 W  
PLW13 0.12955000 W

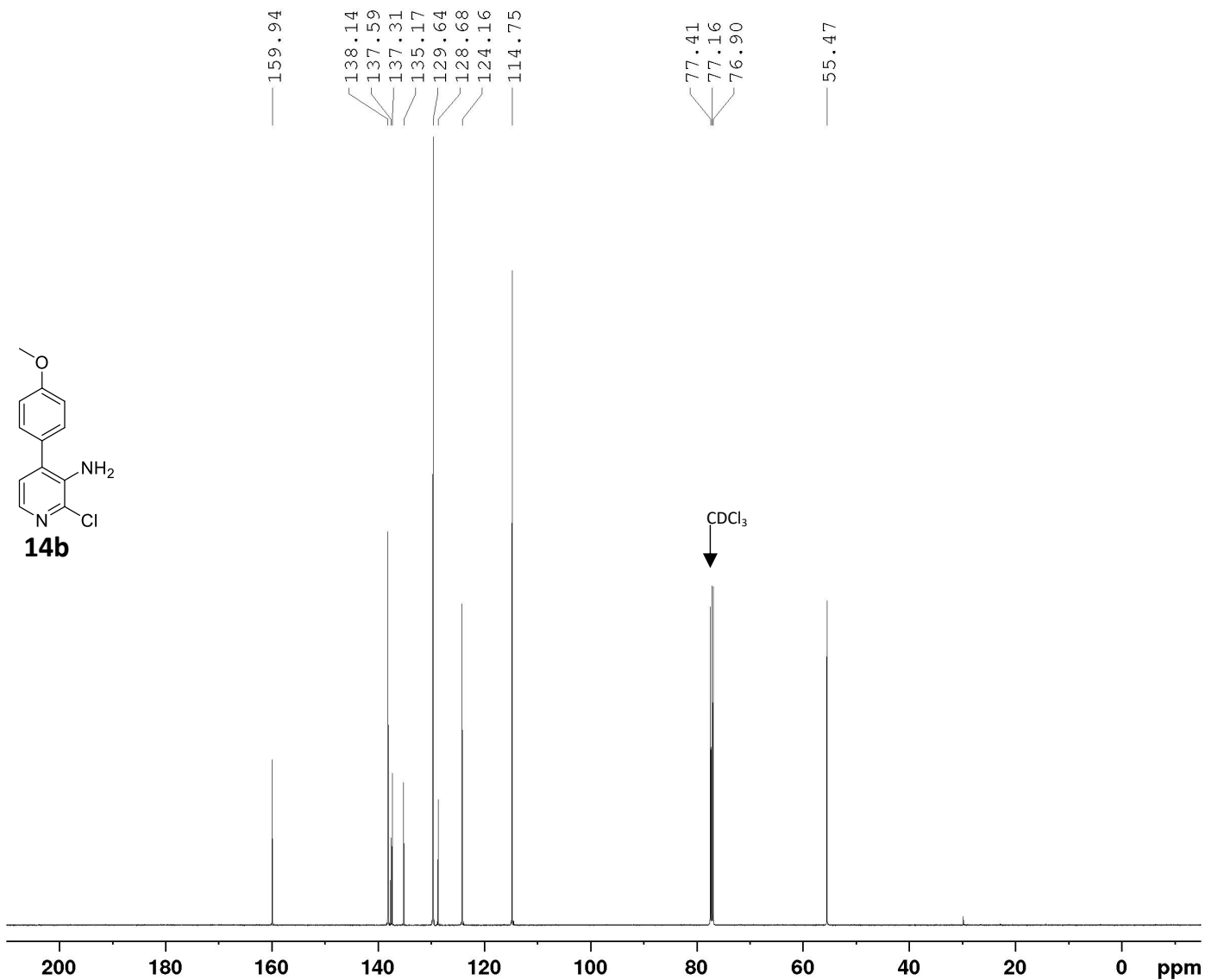
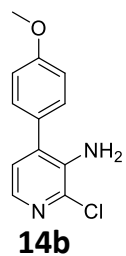
F2 - Processing parameters  
SI 32768  
SF 125.7829218 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



Current Data Parameters  
 NAME JPN-2-17-1\_C4-mono  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20200914  
 Time 10.52 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 ( )  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 30.54  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 300.0 K  
 D1 2.0000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

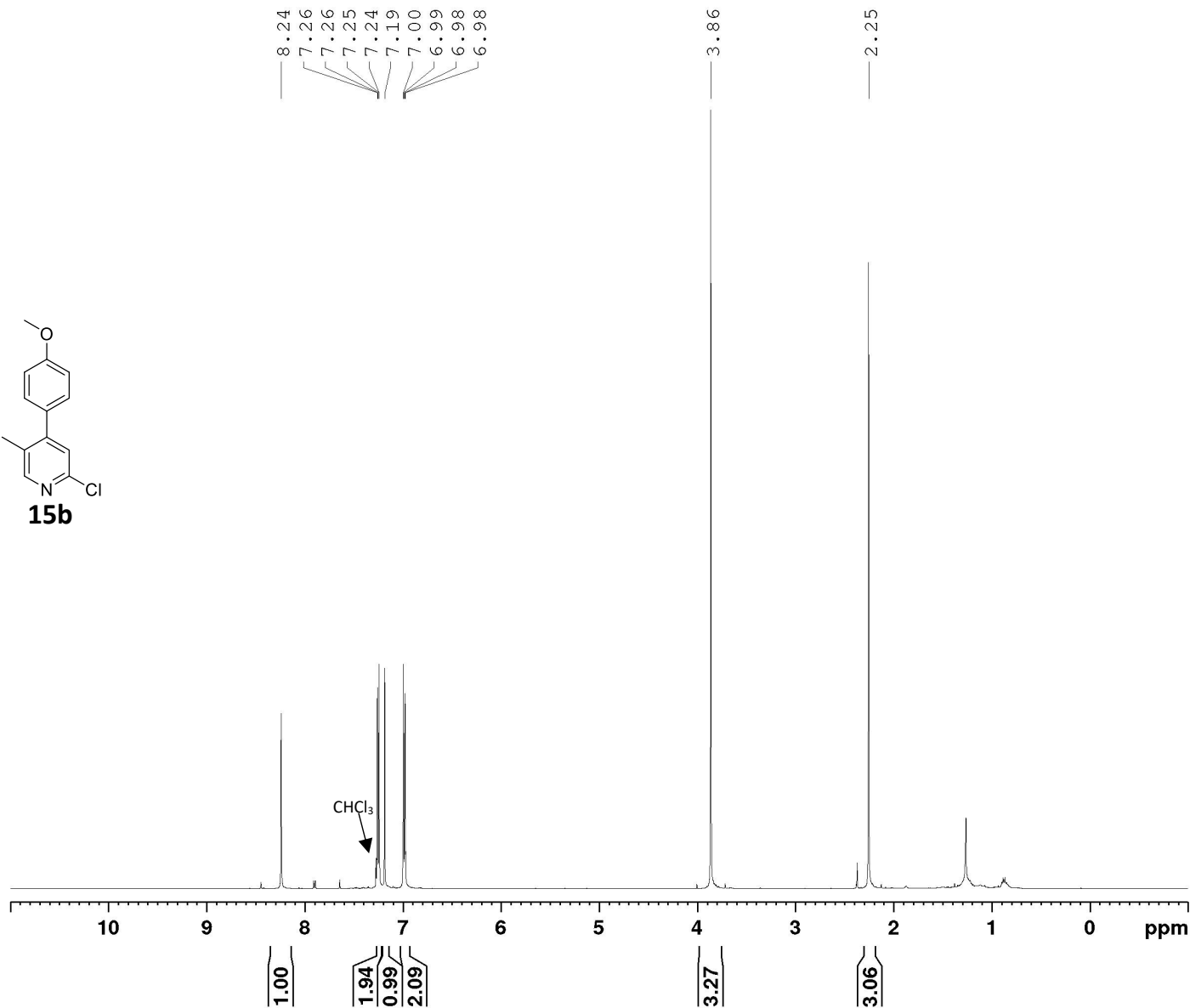
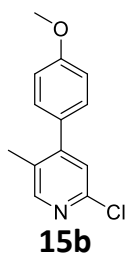
F2 - Processing parameters  
 SI 65536  
 SF 500.2300120 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
NAME JPN-2-17-1  
EXPNO 17  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211119  
Time 9.19 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.908261 Hz  
AQ 1.1010048 sec  
RG 190.44  
DW 16.800 usec  
DE 18.00 usec  
TE 298.0 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 125.7955118 MHz  
NUC1 13C  
P0 3.33 usec  
P1 10.00 usec  
PLW1 56.90299988 W  
SFO2 500.2320009 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 11.44699955 W  
PLW12 0.25756001 W  
PLW13 0.12955000 W

F2 - Processing parameters  
SI 32768  
SF 125.7829236 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

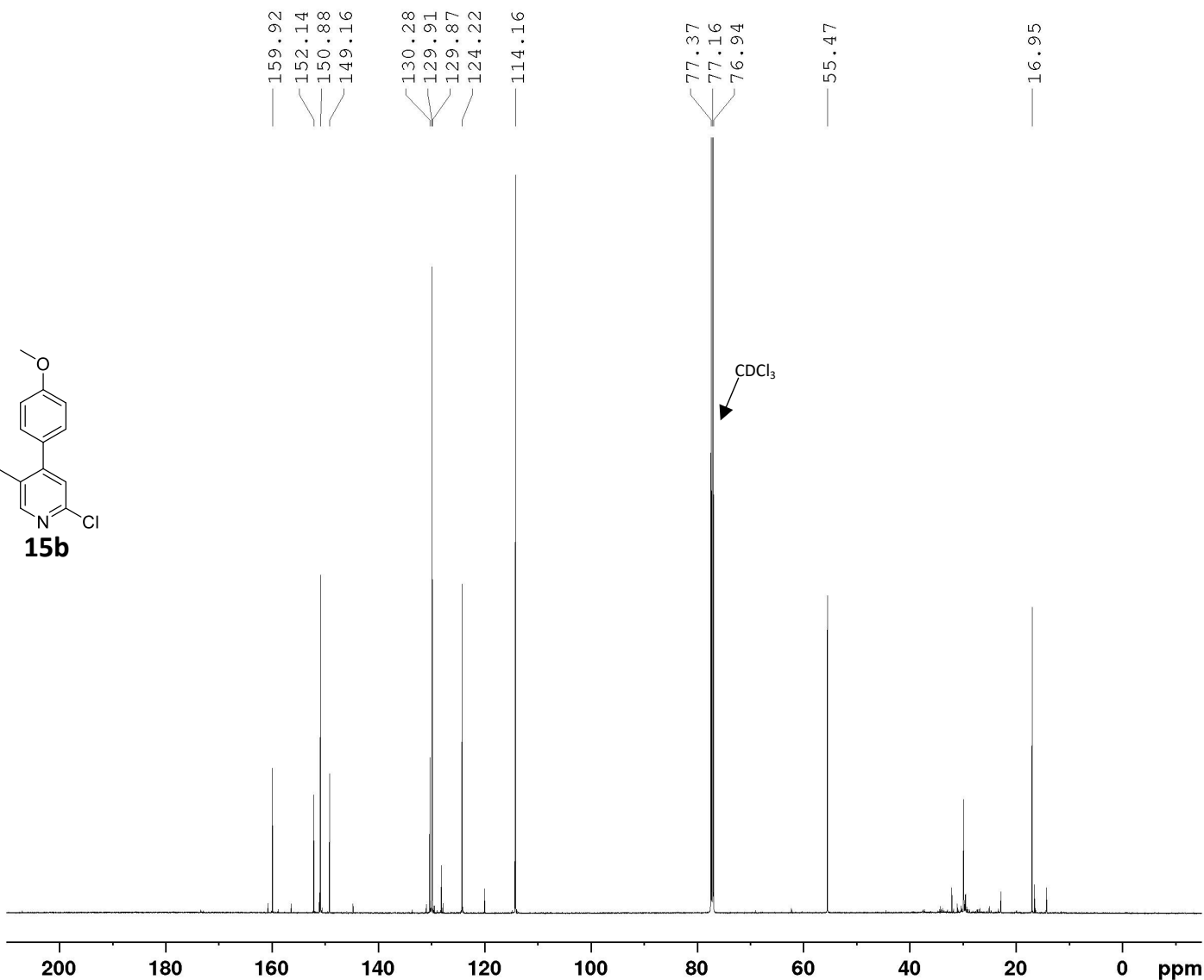
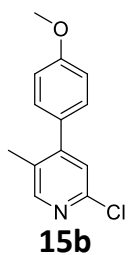


Current Data Parameters  
 NAME JPN-2-17-2\_C4-mono  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20200914  
 Time 4.30 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 25.21  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 300.0 K  
 D1 2.0000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

F2 - Processing parameters  
 SI 65536  
 SF 500.2300047 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

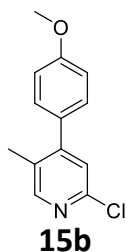




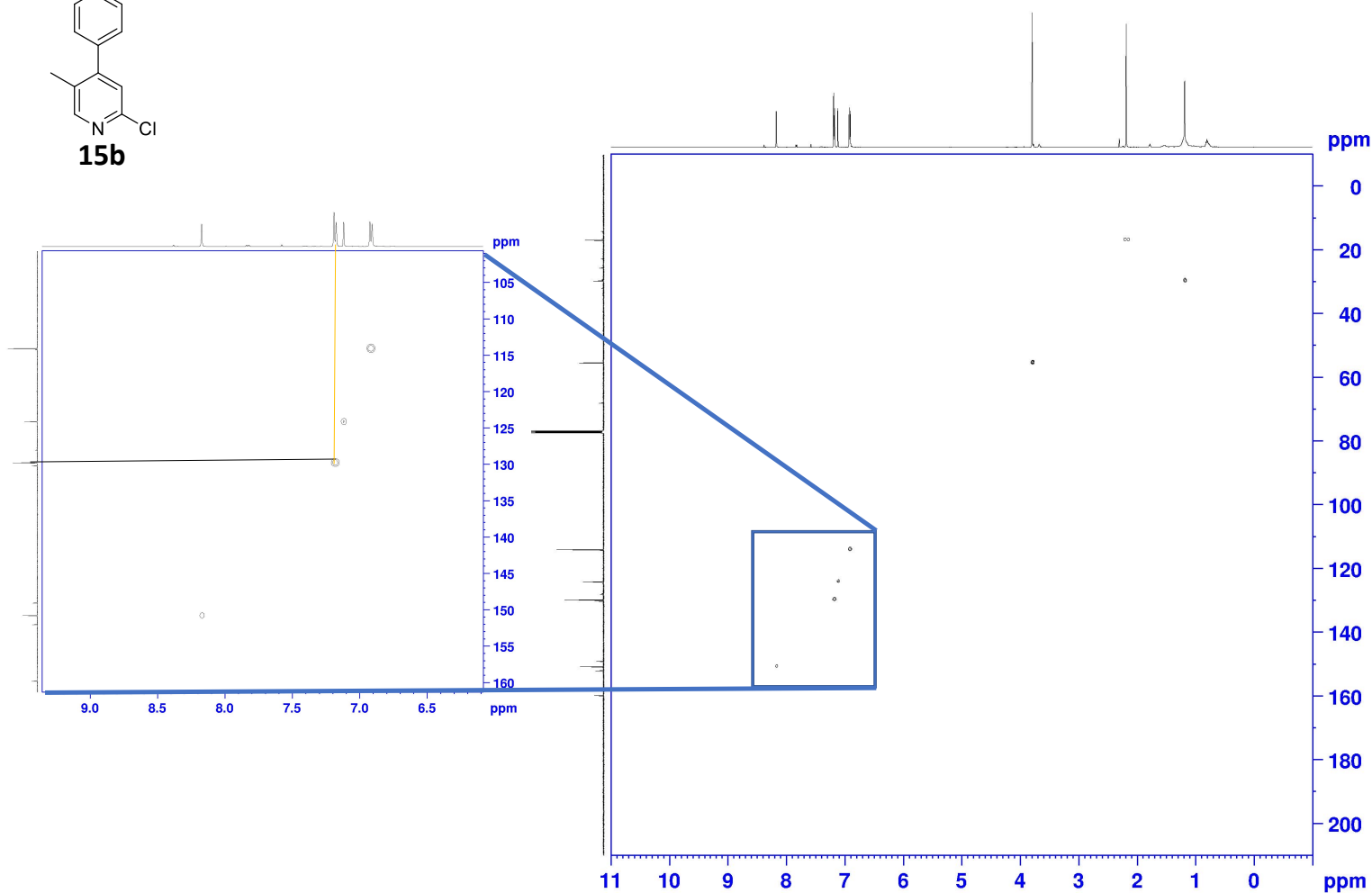
Current Data Parameters  
 NAME JPN-2-17-2  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20200519  
 Time 16.32 h  
 INSTRUM spect  
 PROBHD Z127277\_0002 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 36057.691 Hz  
 FIDRES 1.100393 Hz  
 AQ 0.9087659 sec  
 RG 184.4  
 DW 13.867 usec  
 DE 18.00 usec  
 TE 300.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 150.9178981 MHz  
 NUC1 13C  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 91.0000000 W  
 SFO2 600.1324005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 70.00 usec  
 PLW2 5.59999990 W  
 PLW12 0.07314300 W  
 PLW13 0.03679000 W

F2 - Processing parameters  
 SI 32768  
 SF 150.9027914 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



HSQC



$^{13}\text{C}$  at 130 ppm correlated to the *meta* C-H protons



```

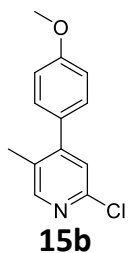
Current Data Parameters
NAME      JPN-2-1-167-4
EXPNO    12
PROCNO   1

F2 - Acquisition Parameters
Date_    20211103
Time     20.15 h
INSTRUM  spect
PROBHD   Z125869_0055 (
PULPROG  hsqcetgp
TD       1024
SOLVENT  CDCl3
NS       8
DS       16
SWH      7002.801 Hz
FIDRES   13.677346 Hz
AQ       0.0731136 sec
RG       190.44
DW       71.400 usec
DE       16.00 usec
TE       298.0 K
CONST2   145.000000
DO       0.00000300 sec
D1       1.50000000 sec
D4       0.00172414 sec
D11      0.03000000 sec
D16      0.00020000 sec
IN0      0.00001990 sec
TDev     1
ZGPGTNS
SFO1     500.2330889 MHz
NUC1     1H
F1       12.00 usec
P2       24.00 usec
PLW1     11.44699955 W
SFO2     125.7948829 MHz
NUC2     13C
CPDPRG[2] garp
P3       10.00 usec
P4       20.00 usec
PCPD2    70.00 usec
PLW2     56.90299988 W
PLW12    1.16129994 W
GPNAM[1] SMSQ10.100
GPZ1     80.00 %
GPNAM[2] SMSQ10.100
GPZ2     20.10 %
P16      1000.00 usec

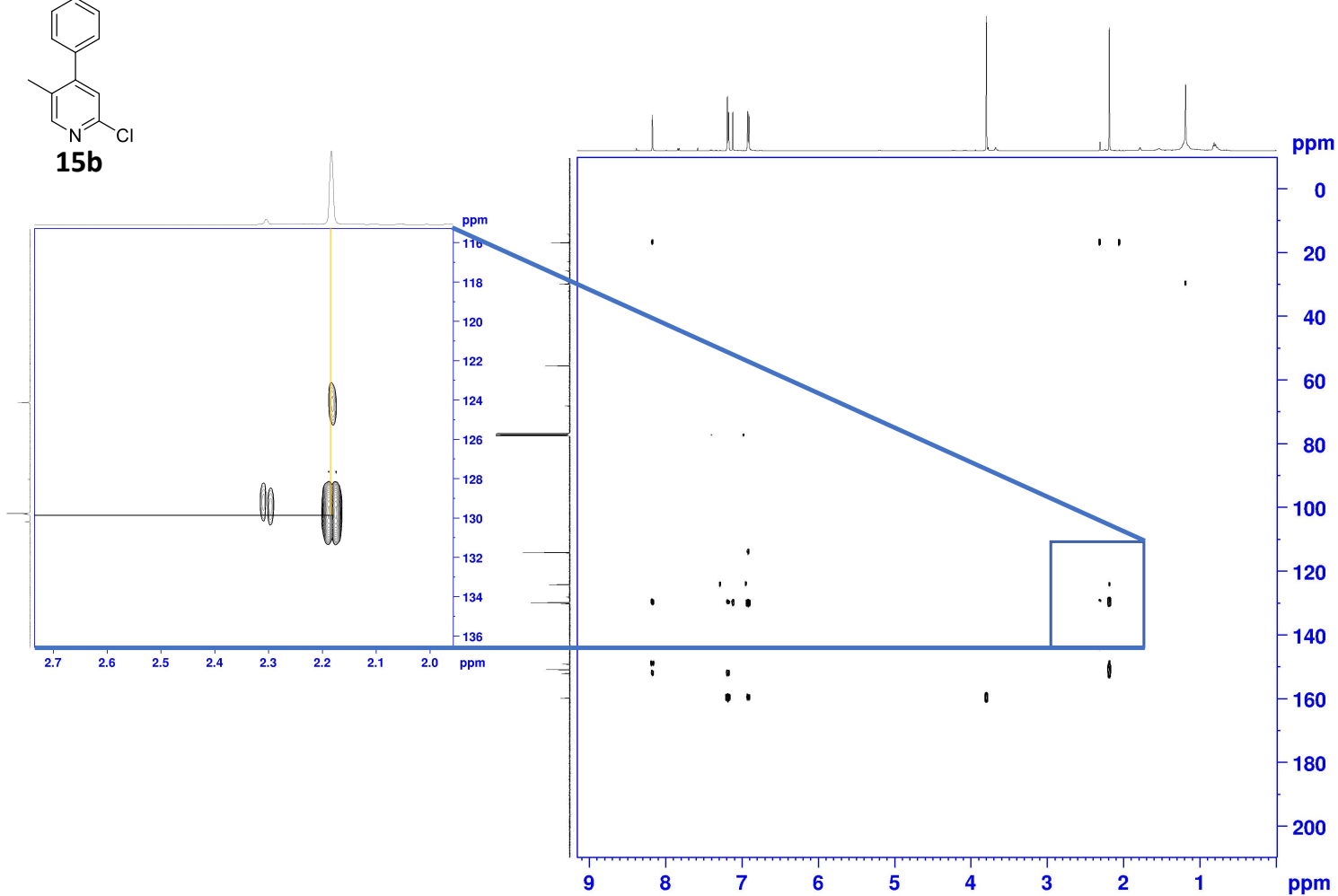
F1 - Acquisition parameters
TD       256
SFO1     125.7949 MHz
FIDRES   196.293976 Hz
SW       199.735 ppm
FnMODE   Echo-Antiecho

F2 - Processing parameters
SI       1024
SF       500.2300477 MHz
WDW      QSINE
SSB      2
LB       0 Hz
GB       0
PC       1.40

F1 - Processing parameters
SI       1024
MC2      echo-antiecho
SF       125.7829335 MHz
WDW      QSINE
SSB      2
LB       0 Hz
GB       0
  
```



### HMBC



Meta C-H protons correlate to methyl on the C5 carbon



Current Data Parameters  
 NAME JPN-2-1-167-4  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 2021103  
 Time 19.20 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (4  
 PULPROG hmcgcp1pnddf  
 TD 4096  
 SOLVENT CDCl3  
 NS 4  
 DS 16  
 SWH 4587.156 Hz  
 FIDRES 2.239822 Hz  
 AQ 0.4464640 sec  
 RG 190.44  
 DW 109.000 usec  
 DE 16.00 usec  
 TE 298.0 K  
 CNST2 145.0000000  
 CNST13 10.0000000  
 DO 0.00000300 sec  
 D1 1.5000000 sec  
 D2 0.00344828 sec  
 D6 0.05000000 sec  
 D16 0.00020000 sec  
 INO 0.00001810 sec  
 TDel 1  
 SFO1 500.2323359 MHz  
 NUC1 1H  
 P1 12.00 usec  
 P2 24.00 usec  
 PLW1 11.44699955 W  
 SFO2 125.7955118 MHz  
 NUC2 13C  
 P3 10.00 usec  
 PLW2 56.90299988 W  
 GPNAM[1] SMSQ10.100  
 GPZ1 50.00 %  
 GPNAM[2] SMSQ10.100  
 GPZ2 30.00 %  
 GPNAM[3] SMSQ10.100  
 GPZ3 40.10 %  
 P16 1000.00 usec

F1 - Acquisition parameters  
 TD 128  
 SFO1 125.7955 MHz  
 FIDRES 431.629822 Hz  
 SW 219.597 ppm  
 FRMODE QF

F2 - Processing parameters  
 SI 4096  
 SF 500.2300475 MHz  
 WDW QSINE  
 SSB 0  
 LB 0 Hz  
 GB 0  
 PC 1.40

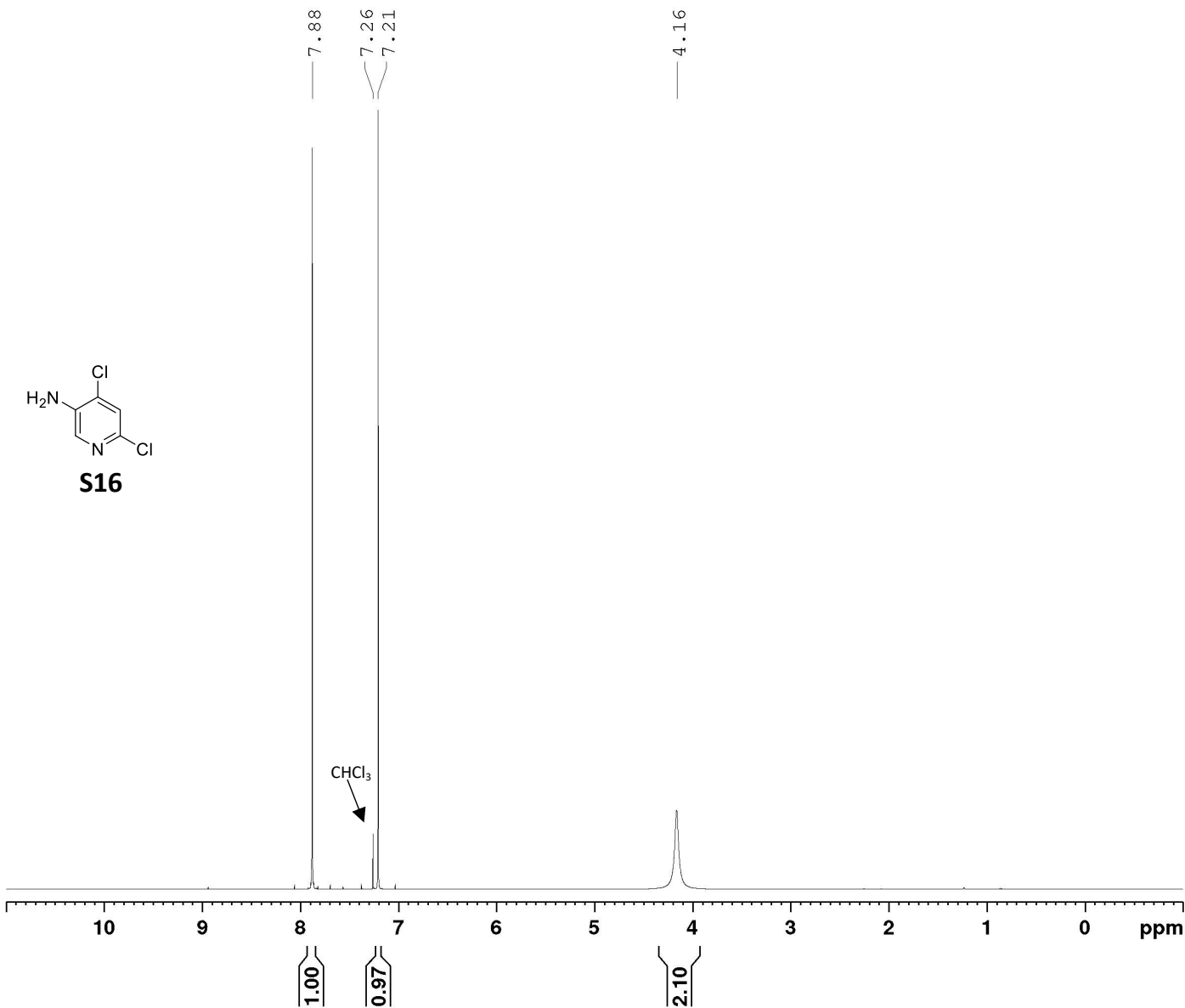
F1 - Processing parameters  
 SI 1024  
 MC2 QF  
 SF 125.7823335 MHz  
 WDW QSINE  
 SSB 0  
 LB 0 Hz  
 GB 0

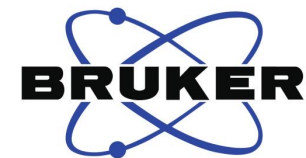


Current Data Parameters  
NAME JPN-2-54\_(5-amino-2,4-DCP)  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211127  
Time 18.56 h  
INSTRUM spect  
PROBHD Z125869\_0055 ( )  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 47.3  
DW 50.000 usec  
DE 16.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300120 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00





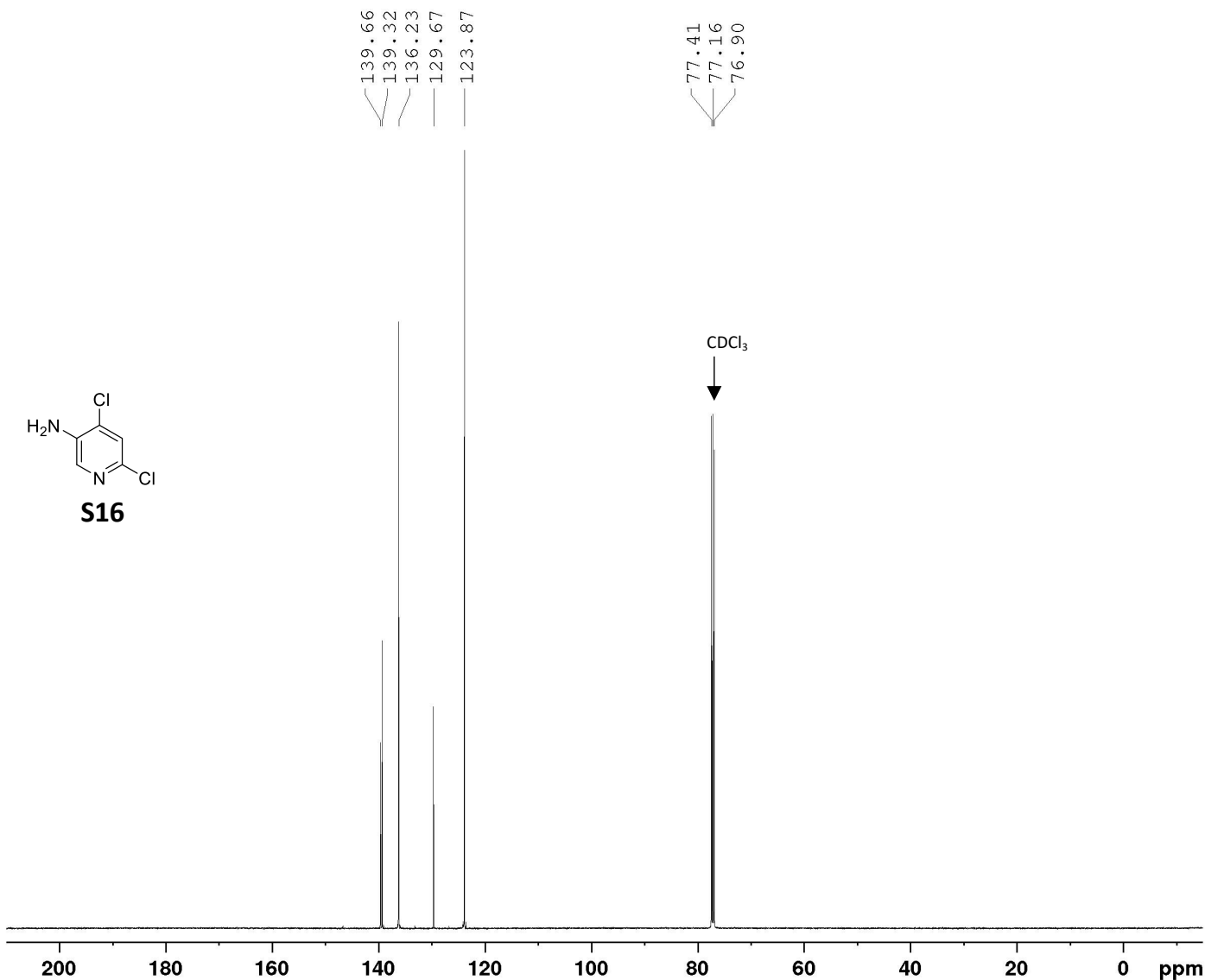
Current Data Parameters  
NAME JPN-2-54\_(5-amino-2,4-DCP)  
EXPNO 11  
PROCNO 1

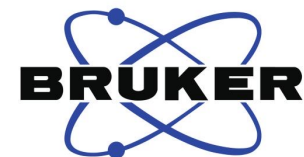
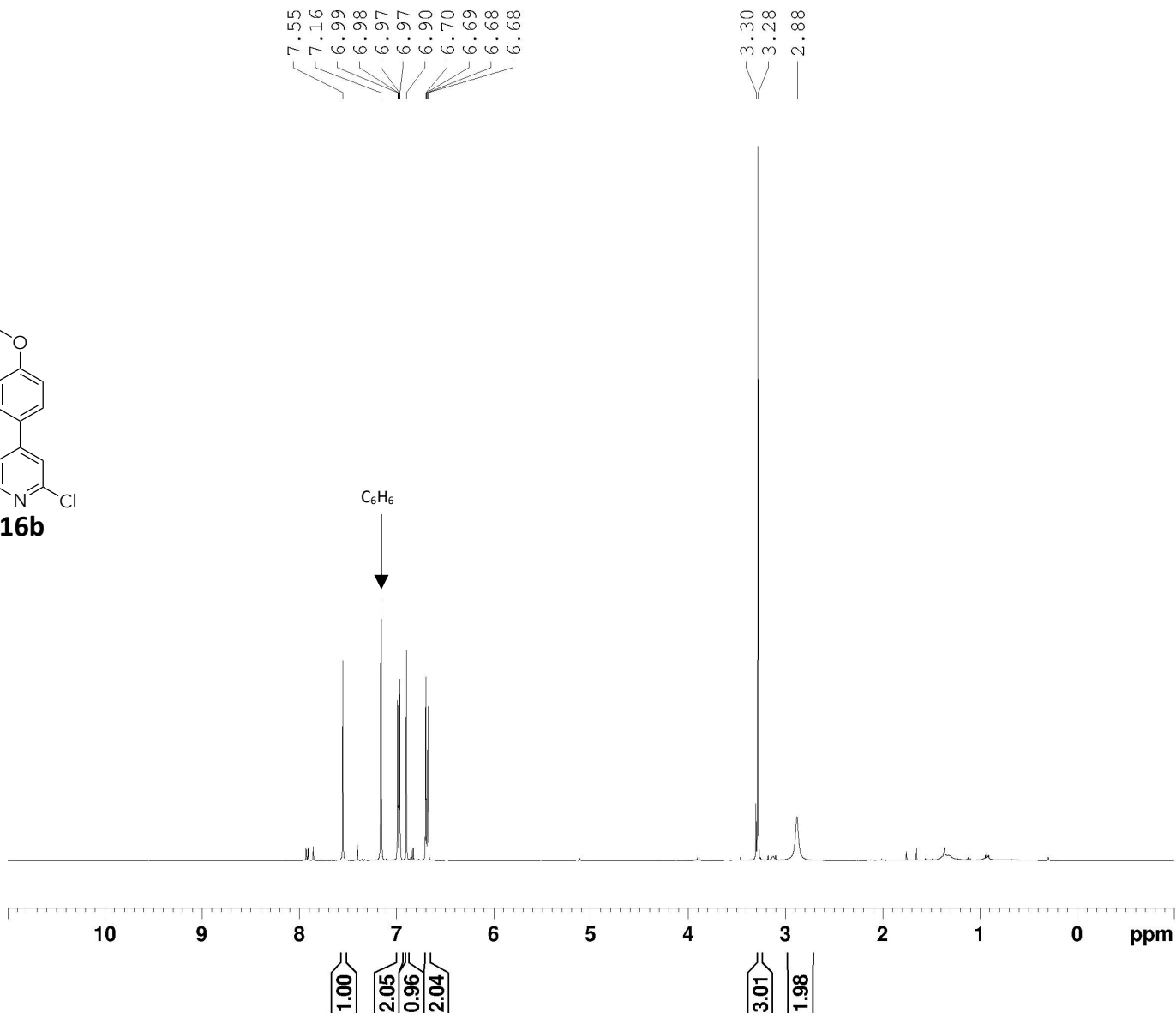
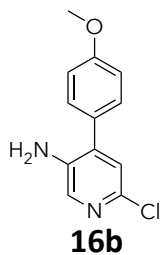
F2 - Acquisition Parameters

Date\_ 20211127  
Time 19.52 h  
INSTRUM spect  
PROBHD z125869\_0055 (zpgg30)  
PULPROG zpgg30  
TD 65536  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.908261 Hz  
AQ 1.1010048 sec  
RG 190.44  
DW 16.800 usec  
DE 18.00 usec  
TE 298.0 K  
D1 2.0000000 sec  
D11 0.0300000 sec  
TD0 1  
SFO1 125.7955118 MHz  
NUC1 13C  
P0 3.33 usec  
P1 10.00 usec  
PLW1 56.90299988 W  
SFO2 500.2320009 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 11.44699955 W  
PLW12 0.25756001 W  
PLW13 0.12955000 W

F2 - Processing parameters

SI 32768  
SF 125.7829244 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

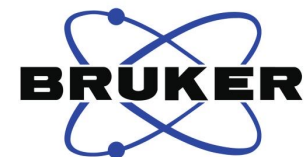
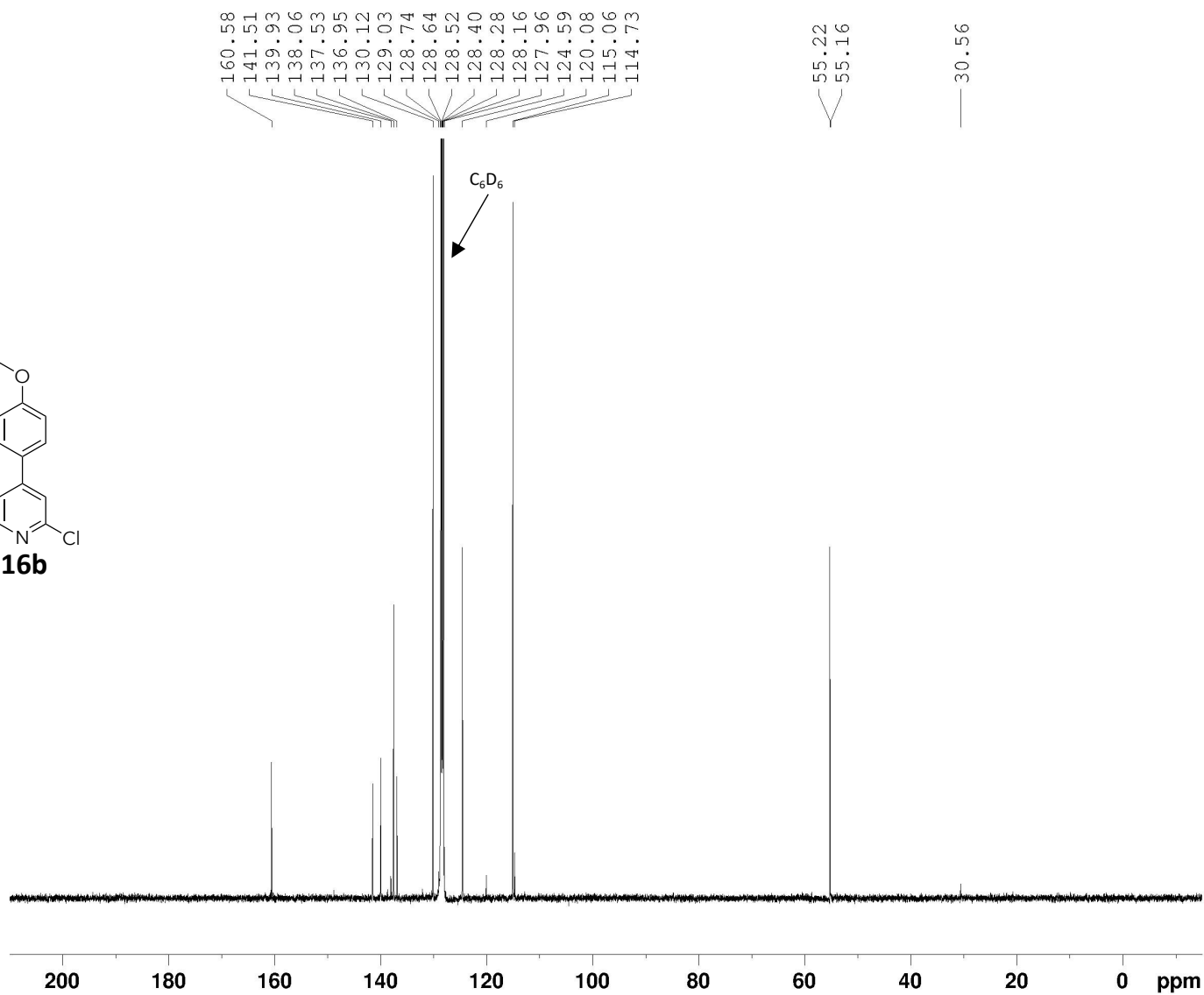
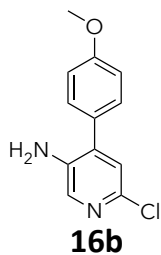




Current Data Parameters  
 NAME NL-1-94-1Hx2  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210807  
 Time 12.32 h  
 INSTRUM Avance Neo  
 PROBHD z152088\_0031 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT C6D6  
 NS 16  
 DS 2  
 SWH 8196.722 Hz  
 FIDRES 0.250144 Hz  
 AQ 3.9976959 sec  
 RG 101  
 DW 61.000 usec  
 DE 13.89 usec  
 TE 298.1 K  
 D1 1.00000000 sec  
 TD0 1  
 SFO1 400.1324708 MHz  
 NUC1 1H  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 24.03499985 W

F2 - Processing parameters  
 SI 65536  
 SF 400.1299967 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
 NAME NL-1-94-1Hx2  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210807  
 Time 19.40 h  
 INSTRUM Avance Neo  
 PROBHD z152088\_0031 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT C6D6  
 NS 1024  
 DS 4  
 SWH 23809.523 Hz  
 FIDRES 0.726609 Hz  
 AQ 1.3762560 sec  
 RG 8.125  
 DW 21.000 usec  
 DE 6.50 usec  
 TE 298.1 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 100.6228298 MHz  
 NUC1 13C  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 86.55400085 W  
 SFO2 400.1316005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz65  
 PCPD2 90.00 usec  
 PLW2 24.03499985 W  
 PLW12 0.18990999 W  
 PLW13 0.09552100 W

F2 - Processing parameters  
 SI 32768  
 SF 100.6126977 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

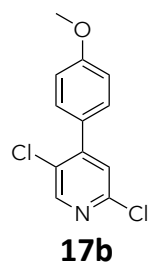
proton 300k c6d6



Current Data Parameters  
NAME NL-1-93-1n-inadequate-08-27-21  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20210827  
Time 12.31 h  
INSTRUM spect  
PROBHD Z127277\_0002 f  
PULPROG zg30  
TD 65536  
SOLVENT C6D6  
NS 16  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.366798 Hz  
AQ 2.7262976 sec  
RG 6.41  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1  
SFO1 600.1337060 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 5.59999990 W

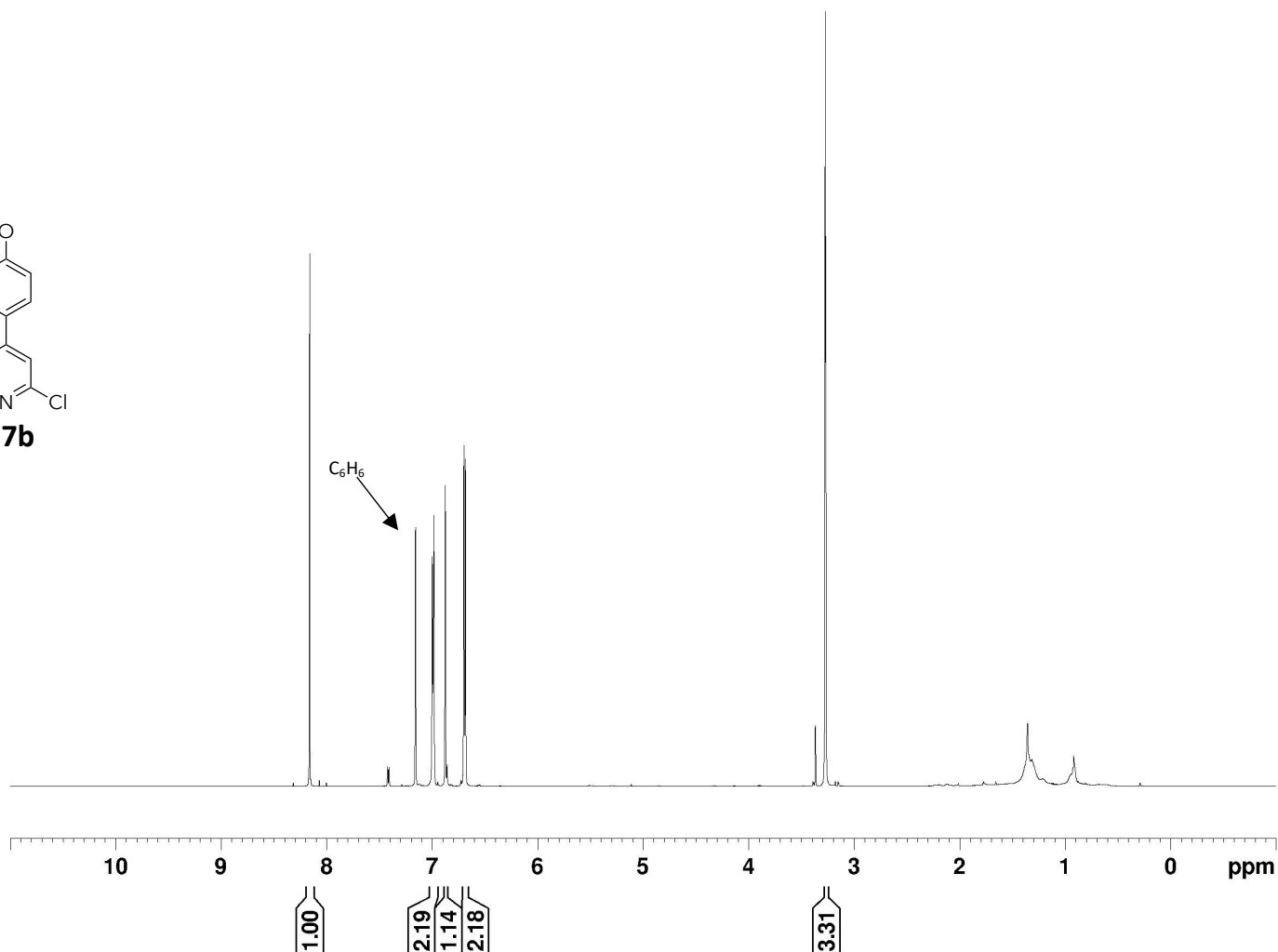
F2 - Processing parameters  
SI 65536  
SF 600.1299970 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



8.16  
7.16  
7.00  
7.00  
6.99  
6.98  
6.88  
6.87  
6.70  
6.68

3.27  
3.27

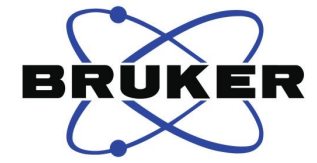
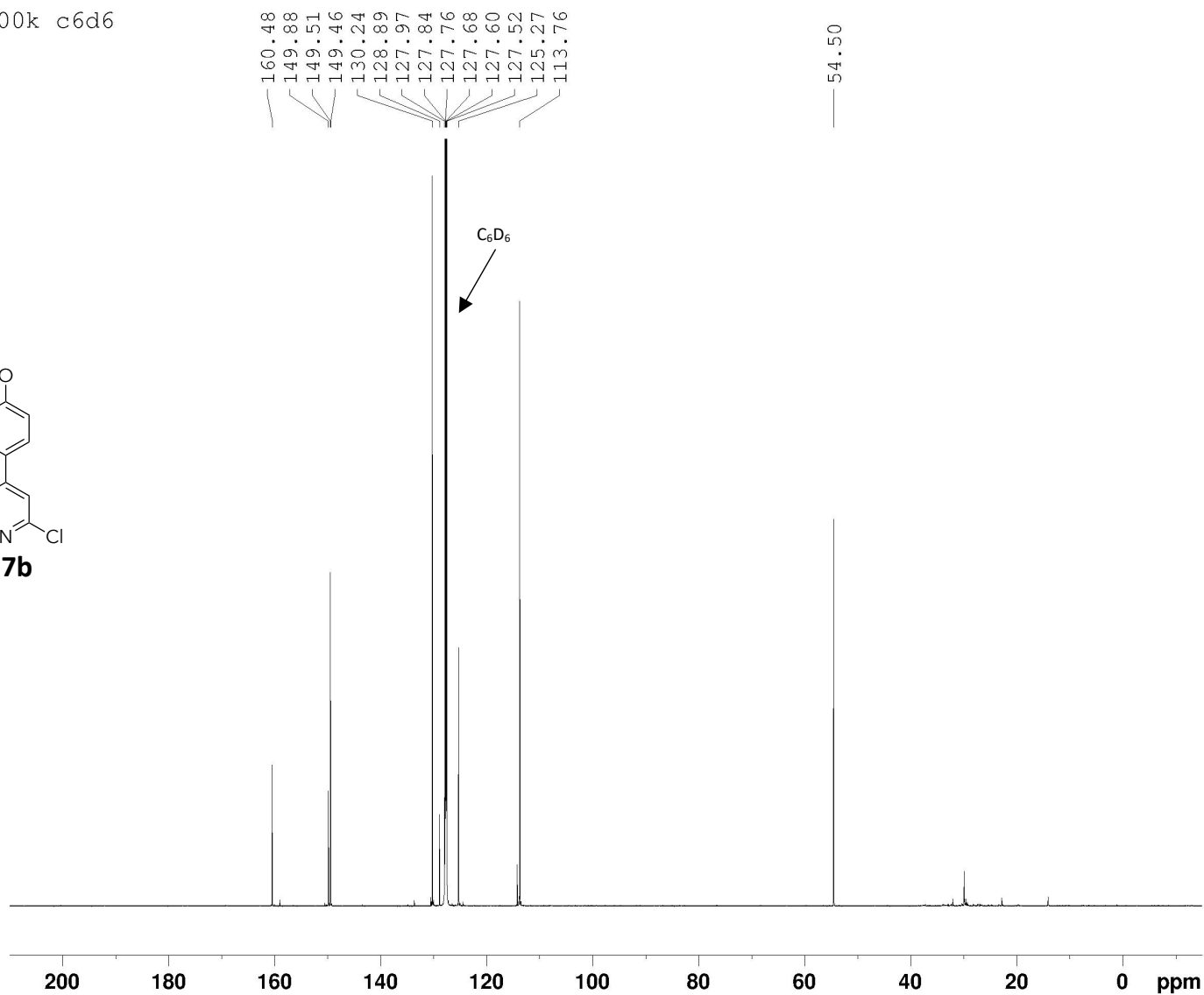
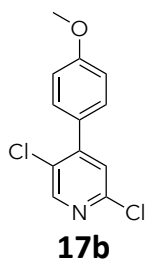
C<sub>6</sub>H<sub>6</sub>



S80



300k c6d6

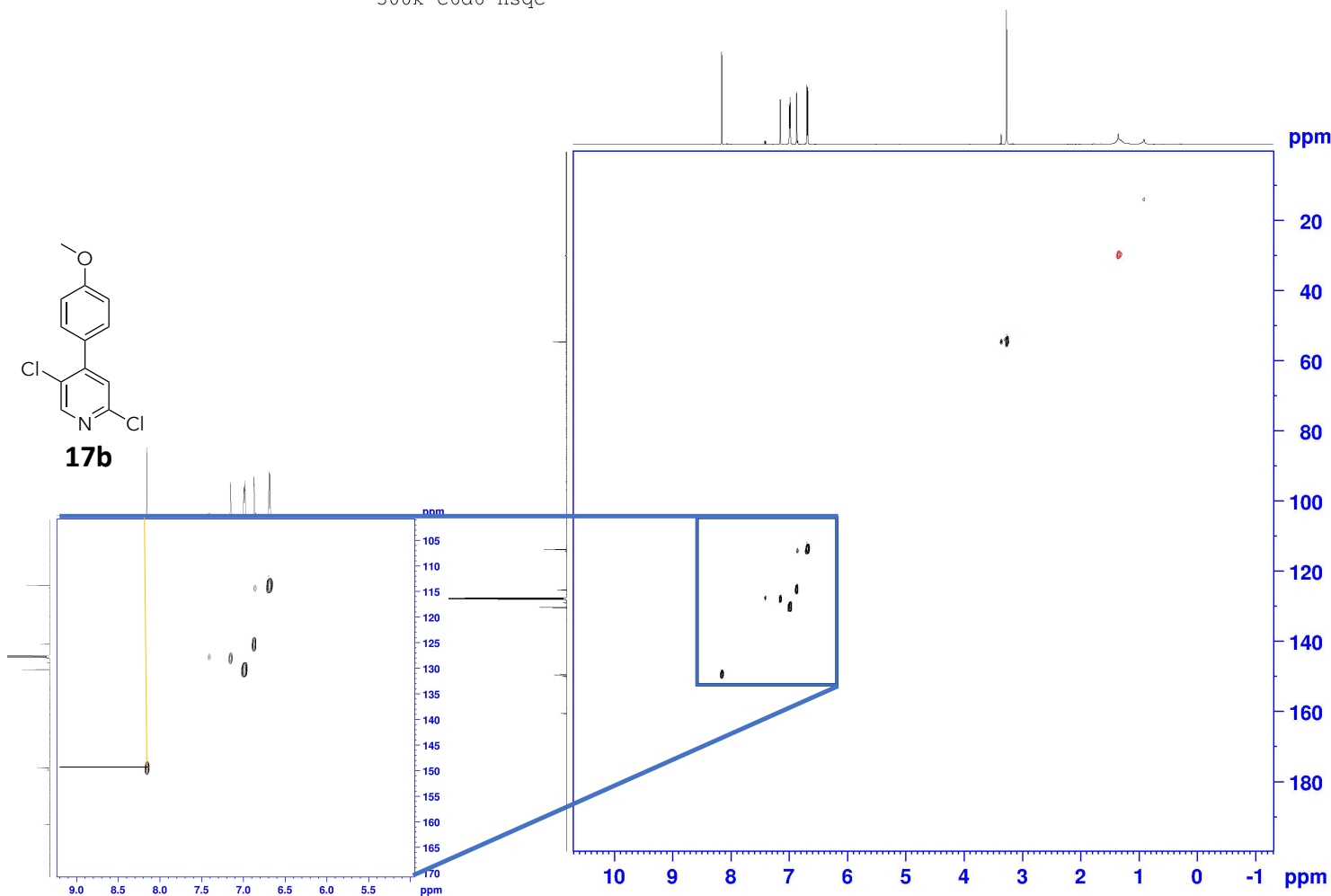
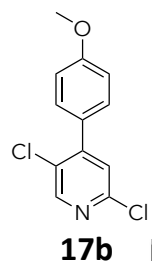


Current Data Parameters  
NAME NL-1-93-1n-inadequate-08-27-21  
EXPNO 4  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20210830  
Time 9.42 h  
INSTRUM spect  
PROBHD Z127277\_0002 (  
PULPROG zgpg30  
TD 65536  
SOLVENT C6D6  
NS 1024  
DS 4  
SWH 36057.691 Hz  
FIDRES 1.100393 Hz  
AQ 0.9087659 sec  
RG 184.4  
DW 13.867 usec  
DE 18.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 150.9178981 MHz  
NUC1 13C  
P0 4.00 usec  
P1 12.00 usec  
PLW1 91.00000000 W  
SFO2 600.1324005 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 70.00 usec  
PLW2 5.59999990 W  
PLW12 0.07314300 W  
PLW13 0.03679000 W

F2 - Processing parameters  
SI 32768  
SF 150.9028090 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

300k c6d6 hsqc



Current Data Parameters  
NAME ML-1-93-1a-15adegate-08-27-21  
EXPNO 702  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20210827  
Time 12.35 h  
INSTRUM spect  
PROBHD Z127277\_0092\_1  
PULPROG noah2\_88  
TD 1524  
SOLVENT C6D6  
NS 4  
DS 8  
SWH 7211.559 Hz  
FIDRES 14.085936 Hz  
AQ 0.0709973 sec  
RG 184.4  
DW 69.333 usec  
DE 10.00 usec  
TE 300.0 K  
CMT2 145.0000000  
CMT6 120.0000000  
CMT7 176.0000000  
CMT13 8.0000000  
D0 0.0000300 sec  
D1 1.0000000 sec  
D2 0.00344828 sec  
D4 0.0012414 sec  
D6 0.0625000 sec  
D16 0.0000000 sec  
TMO 0.0000160 sec  
L3 64  
SOL 1  
SFO1NS -CSDIT  
SFO1 600.1328256 MHz  
NUC1 1H  
P1 8.00 usec  
P2 16.00 usec  
RFW1 5.59999990 W  
SFO2 150.9199988 MHz  
NUC2 13C  
CPDPRG2 gq4r4  
P3 12.00 usec  
P14 500.00 usec  
P15 1730.00 usec  
PCPD2 60.00 usec  
PLA2 91.0000000 W  
P1W12 3.64000010 W  
SPNAM3 Ccp60\_0\_5\_20\_1  
SFOAL3 0.500  
SFOFF3 0 Hz  
SPW3 20.02099991 W  
SPNAM18 Ccp60\_xf1k\_2  
SFOAL8 0.500  
SFOFF18 0 Hz  
SFO18 5.78449990 W  
GENM(0) SMSQ10.100  
GF0 17.13 %  
GENM(1) SMSQ10.100  
GF1 39.00 %  
GENM(2) SMSQ10.100  
GF2 49.20 %  
GENM(3) SMSQ10.100  
GF3 15.00 %  
GENM(4) SMSQ10.100  
GF4 10.00 %  
GENM(5) SMSQ10.100  
GF5 5.00 %  
P16 1000.00 usec

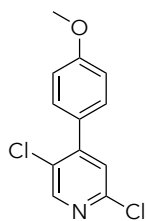
F1 - Acquisition Parameters  
TD 128  
SFO1 150.9179 MHz  
FIDRES 470.612518 Hz  
SW 199.582 ppm  
FbMxIDE Echo-Antiecho

F2 - Processing parameters  
SI 1024  
SF 600.1300000 MHz  
MSB G31NE  
SB 2  
LB 0 Hz  
GB 0  
PC 1.40

F1 - Processing parameters  
SI 1024  
MC1 echo-antiecho  
SF 150.9028085 MHz  
MSB G31NE  
SB 2  
LB 0 Hz  
GB 0

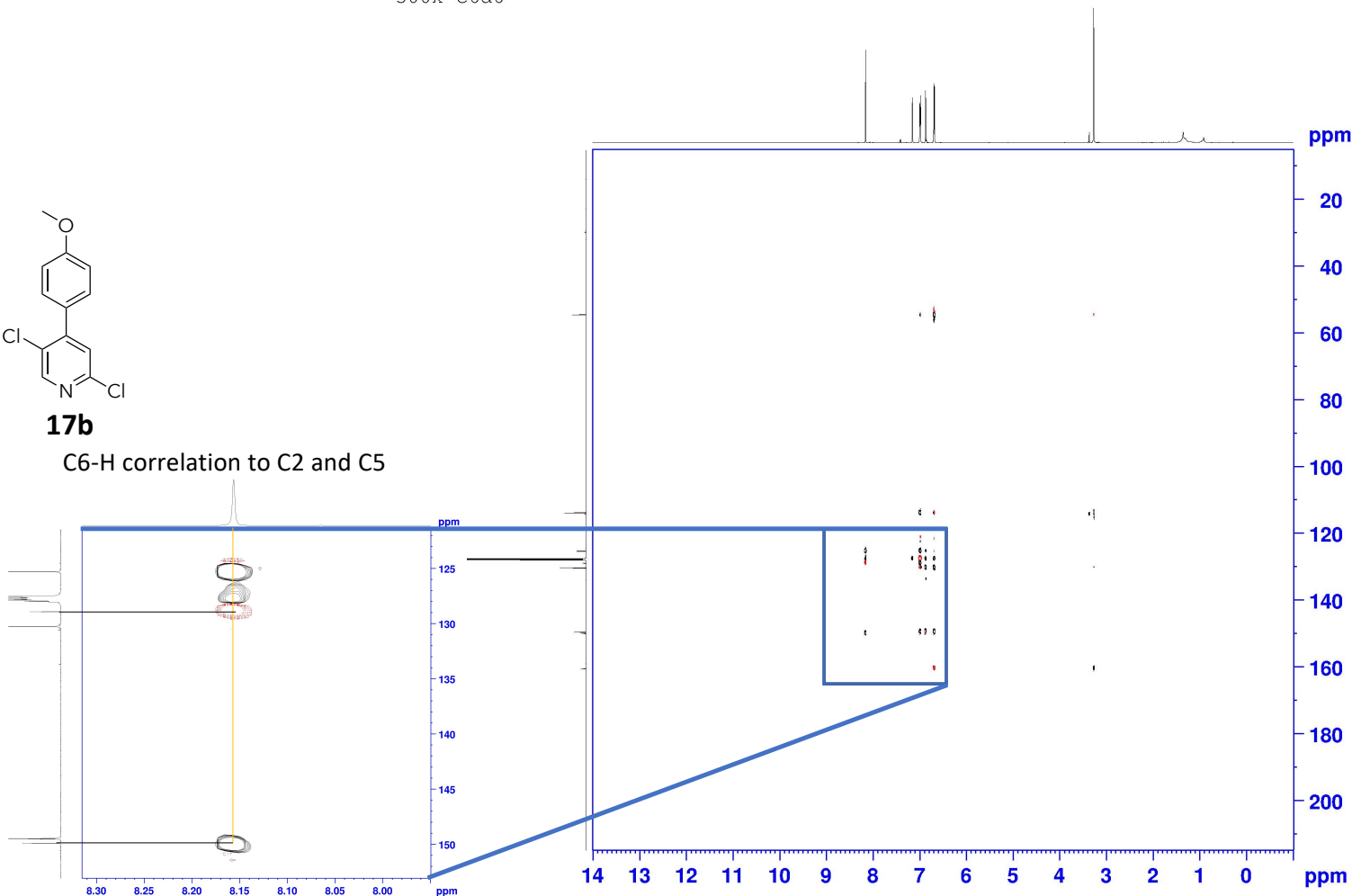
# 1,1 ADEQUATE

300k c6d6



**17b**

C6-H correlation to C2 and C5



```

Current Data Parameters
NAME: MR-1-15-17c1c1adequate-08-27-21
EXPNO: 3
PROCNO: 1

F2 - Acquisition Parameters
Date_: 20210810
Time: 11.21 h
INSTRUM: spect
PROBHD: 1127271_0602 /
PULPROG: zgpg30
TD: 2048
SOLVENT: CDCl3
NS: 256
DS: 4
SWH: 9914.413 Hz
FIDRES: 8.803147 Hz
AQ: 0.1115957 sec
RG: 194.4
DW: 55.447 usec
DE: 10.00 usec
TE: 300.2 K

CONST1 145.000000
CONST2 31.000000
CONST3 64.000000
CONST4 -1.111112
CONST5 9.1428576
CONST6 9.1428576
CONST7 11.3999996
CONST8 8.000000
CONST9 9.500000
CONST10 8.000000
CONST11 8.000000
CONST12 -0.500000
D1 0.0000000 sec
D2 2.0000000 sec
D3 0.0012414 sec
D4 0.0000000 sec
D5 0.0000000 sec
D6 0.0000000 sec
D7 0.0209253 sec
D8 0.0307015 sec
D9 0.0008207 sec
D10 0.0212982 sec
D11 0.03515625 sec
D12 0.0212425 sec
D13 0.02633975 sec
D14 0.00001580 sec
INZ0 0.00001580 sec
INZ1 0.00001580 sec
LO 1
TD0 2
YD0 0
YD1 600.1330000 MHz
NUC1 1H
P1 1H
P2 16.00 usec
P3 5.9999999 W
P4 150.9194078 MHz
NUC2 13C
CPDPRG2 bl_2step_4sp_2
P5 16.00 usec
P6 12.00 usec
P7 500.00 usec
P8 2000.00 usec
P9 1500.00 usec
P10 91.0000000 W
P11 3.4400000 W
SPRAME1 Cmp69,0.5,20,1
SPRAME2 0.5,20
SPRAME3 20.0209999 W
SPRAME4 Cmp60comp,4
SPRAME5 0.5,20
SPRAME6 20.0209999 W
SPRAME7 Cmp42,1.5,20,2
SPRAME8 0.5,20
SPRAME9 11.2119999 W
SPRAME10 Cmp42,1.5,20,2
SPRAME11 0.5,20
SPRAME12 2.8029999 W
SPRAME13 0.5,20
SPRAME14 150.900000 MHz
QZ1 10.00 W
P16 1000.00 usec

F1 - Acquisition parameters
TD 256
SFO1 150.9194 MHz
FIDRES 247.231010 Hz
SW 209.685 ppm
PROCOR Echo-Det-Loch
F2 - Processing parameters
SI 2048
SF 600.1330000 MHz
WDW EM
SSB 0
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
SF 150.900000 MHz
WDW EM
SSB 0
GB 0
PC 1.40
    
```

300k c6d6 hmbc  
2D selective HMBC



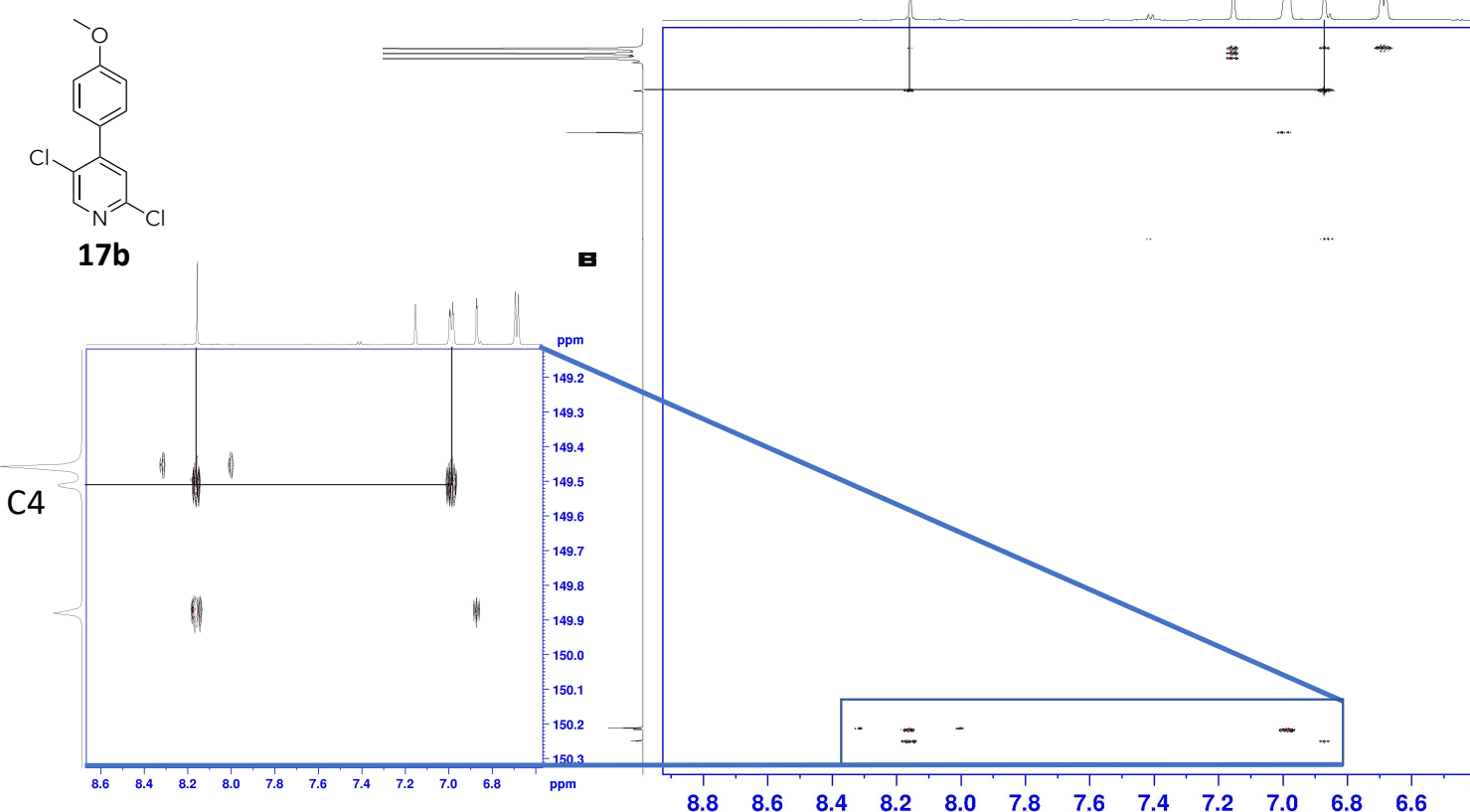
Current Data Parameters  
NAME NL-1-93-ln-inadequate-08-27-21  
EXPNO 6  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20210901  
Time 16.12 n  
INSTRUM spect  
PROBHD 2127277\_0002 (SULPROG) shmbcctgtplind  
TD 2048  
SOLVENT C6D6  
NS 32  
DS 16  
SWH 7211.539 Hz  
FIDRES 7.042818 Hz  
AQ 0.1419947 sec  
RG 184.4  
DM 69.333 usec  
DE 10.00 usec  
TE 300.0 K  
CNS16 120.000000  
CNS17 170.000000  
CNS113 8.000000  
DO 0.00000300 sec  
D1 1.30000000 sec  
D6 0.00200000 sec  
D16 0.00200000 sec  
D20 0.02000000 sec  
INO 0.00007830 sec  
INZ0 0.00003915 sec  
Dwav 1  
SFO1 600.1328206 MHz  
NUC1 13C  
P1 8.00 usec  
F2 16.00 usec  
SFO1 150.9250574 MHz  
NUC2 13C  
P3 12.00 usec  
F4 500.00 usec  
P43 770.60 usec  
SFO2 91.0000000 W  
SPNAM[3] Ccp60,0.5,20.1  
SFOAL3 0.500  
SFOFF33 0 Hz  
SPW3 20.02099991 W  
SPNAM[32] Q3\_surrog\_1  
SFOAL32 0.500  
SFOFF332 0 Hz  
SFW32 6.08045387 W  
GPNAM[1] SMSQ10.100  
GPE1 80.00 %  
GPNAM[3] SMSQ10.100  
GPE3 15.00 %  
GPNAM[4] SMSQ10.100  
GPE4 -10.00 %  
GPNAM[5] SMSQ10.100  
GPE5 -5.00 %  
P16 1000.00 usec

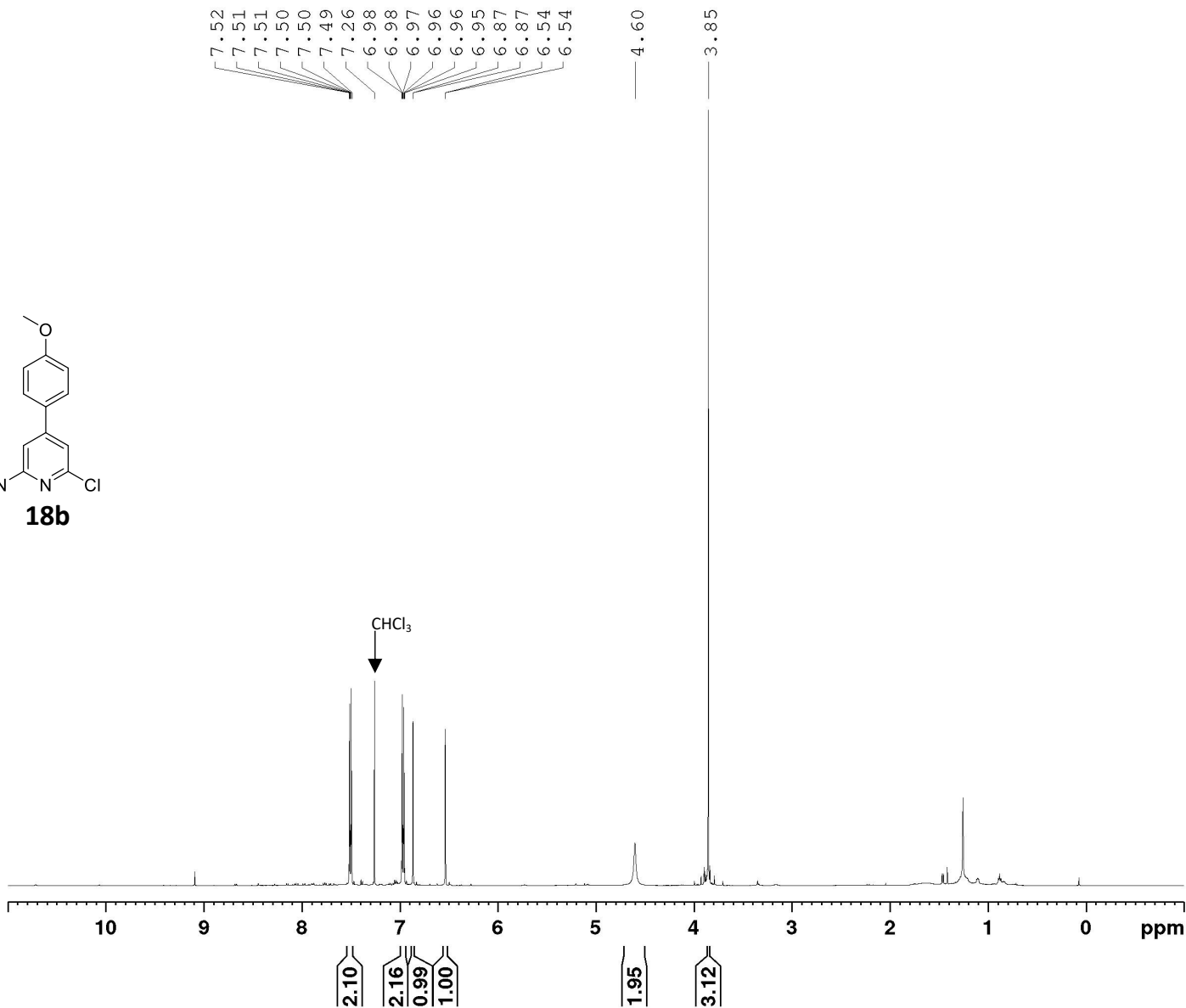
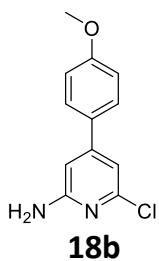
F1 - Acquisition parameters  
TD 1024  
SFO1 150.9251 MHz  
FIDRES 12.470062 Hz  
SW 42.310 ppm  
FMODE Echo-Antlecho

F2 - Processing parameters  
SI 2048  
SF 600.1300000 MHz  
WDW SINE  
SSB 4  
LB 0 Hz  
GB 0  
GC 1.40

F1 - Processing parameters  
SI 1024  
MC2 echo-antlecho  
SF 150.9028085 MHz  
WDW QSINE  
SSB 2  
LB 0 Hz  
GB 0



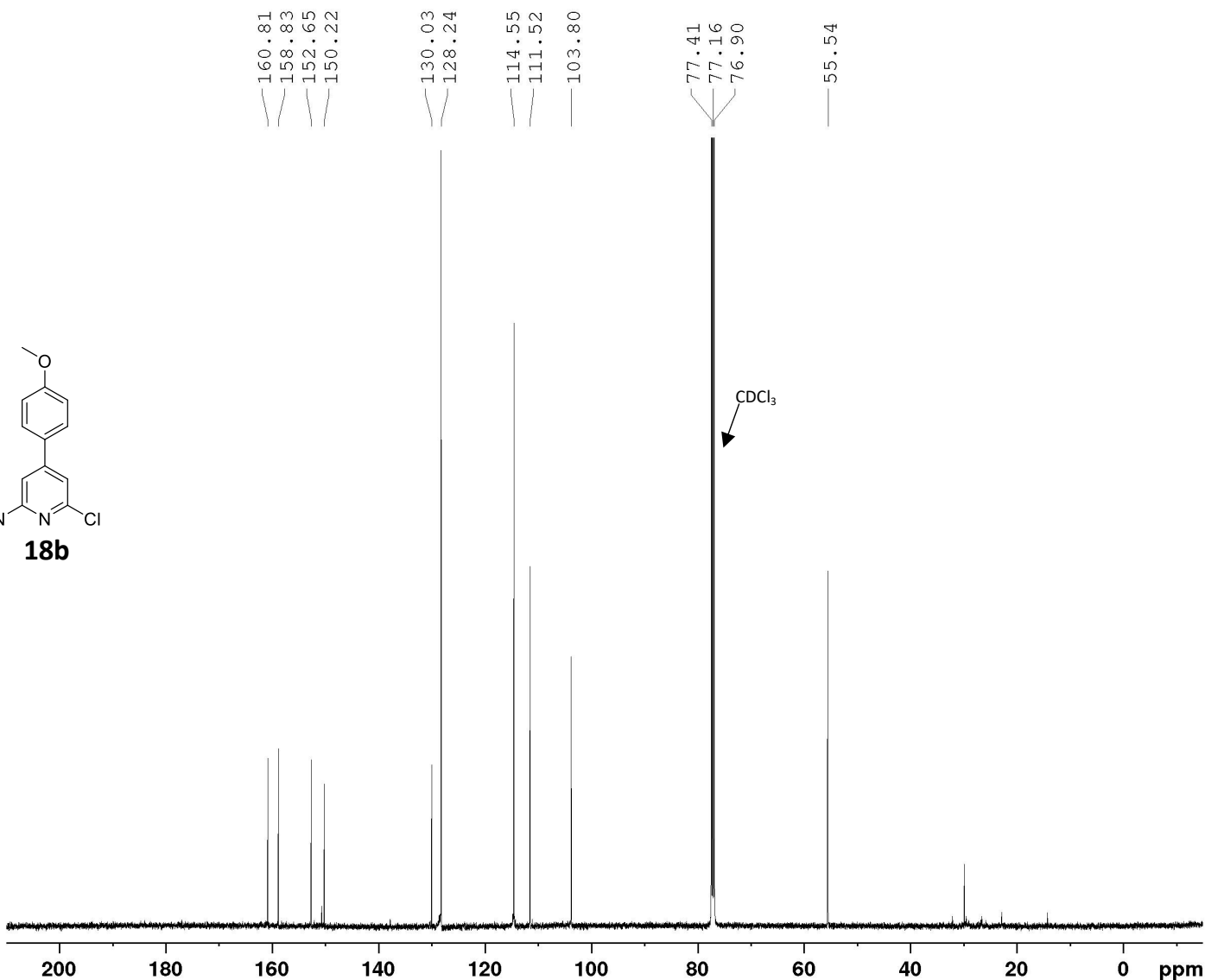
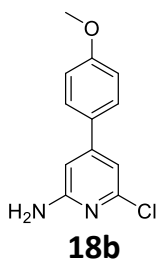
C4 reasoned by process of elimination, HMBC correlations, and chemical shift. Optimized 3-bond correlations. C(129ppm) weak correlation to C6-H, and a strong correlation to C3-H results in identification as C5. Absence of correlation from C(149.5) to C3-H and strong correlation to C6-H results in identification of C4.



Current Data Parameters  
 NAME JPN-2-16-2  
 EXPNO 16  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211120  
 Time 1.49 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 151.18  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

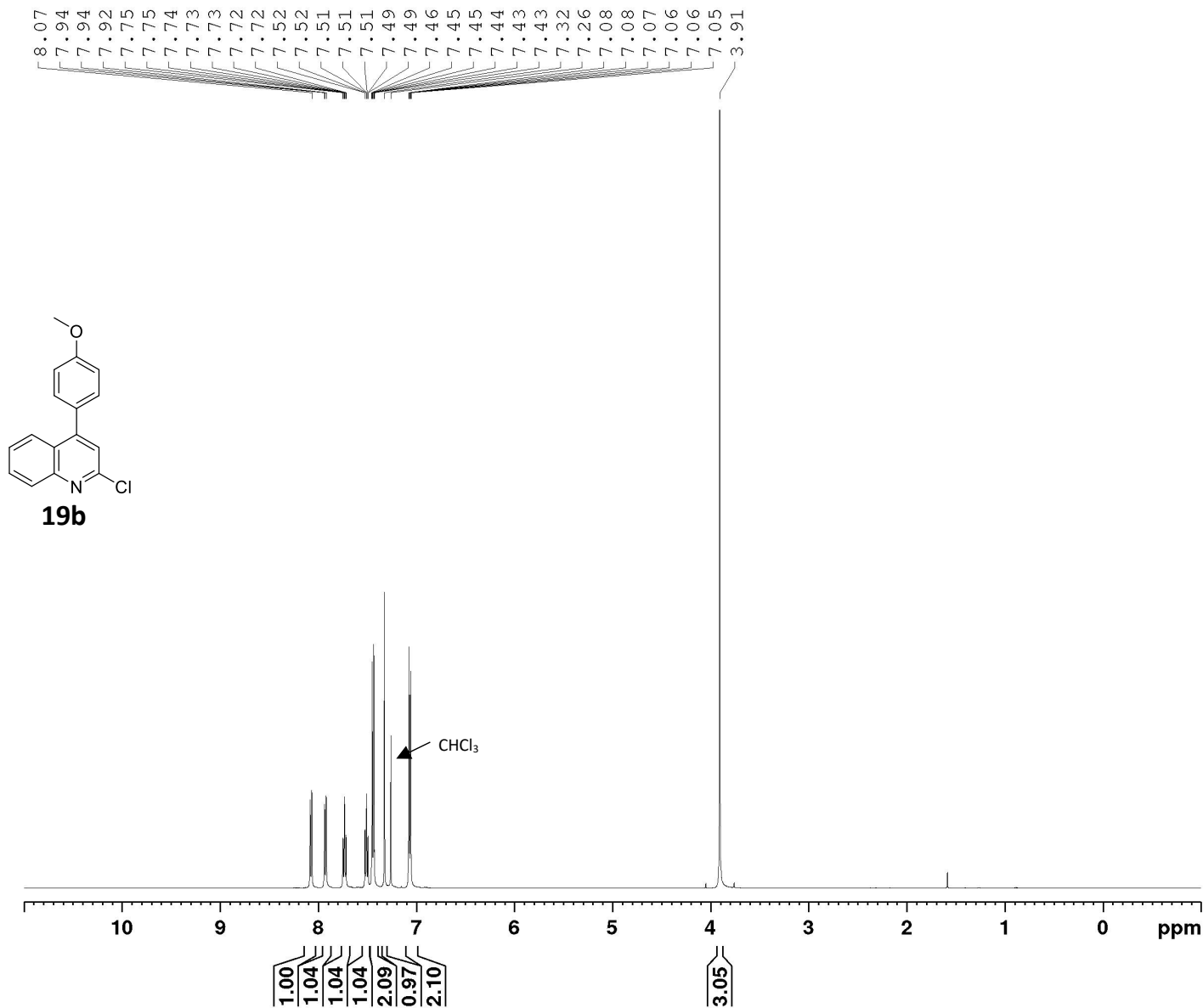
F2 - Processing parameters  
 SI 65536  
 SF 500.2300121 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
 NAME JPN-2-16-2  
 EXPNO 17  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211120  
 Time 2.45 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

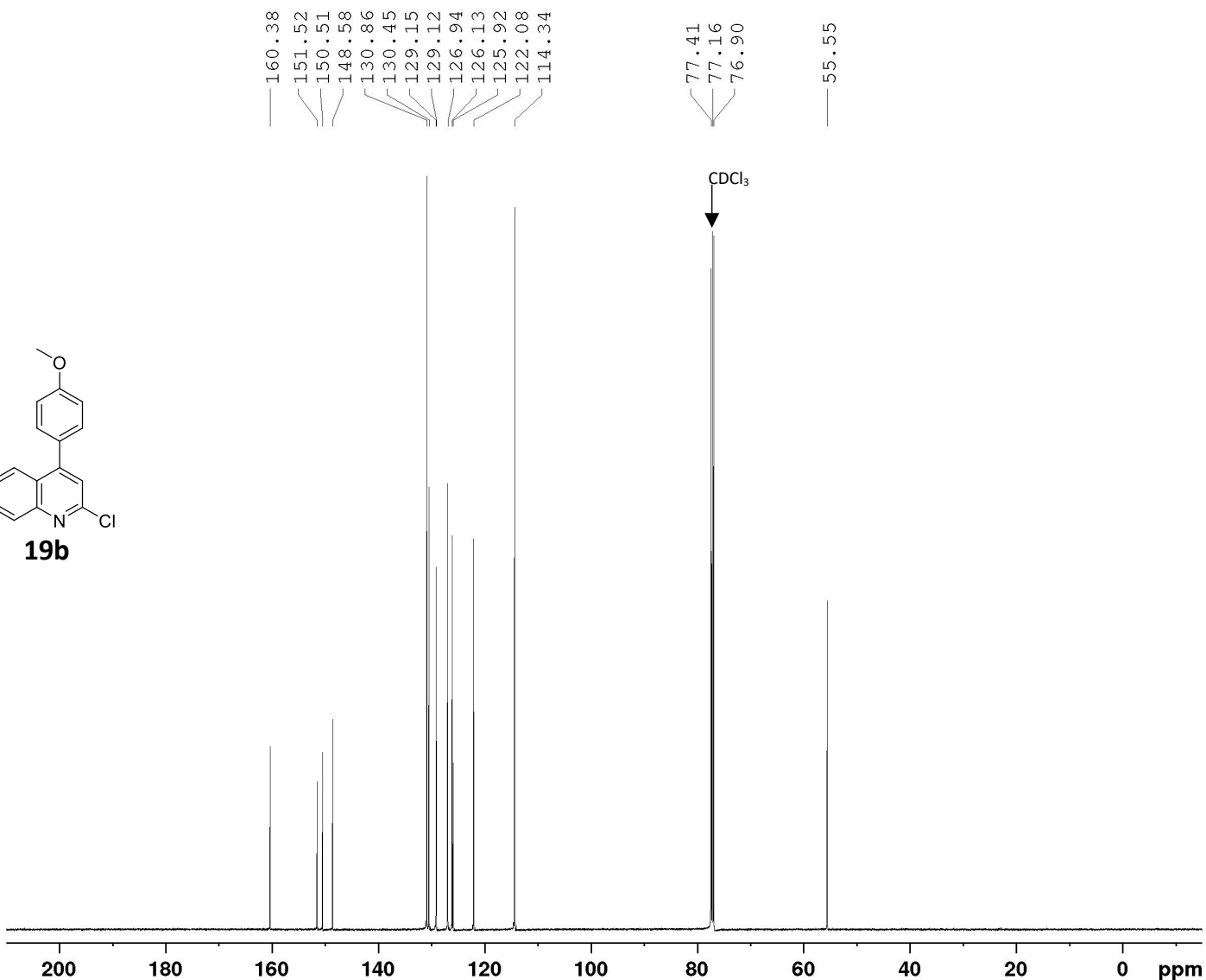
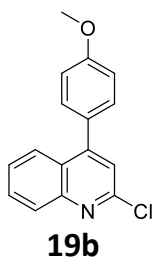
F2 - Processing parameters  
 SI 32768  
 SF 125.7829170 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
NAME JPN-2-8\_repeat\_C4-mono\_crop  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20201130  
Time 17.10 h  
INSTRUM spect  
PROBHD Z125869\_0055 (   
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 122.05  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300120 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

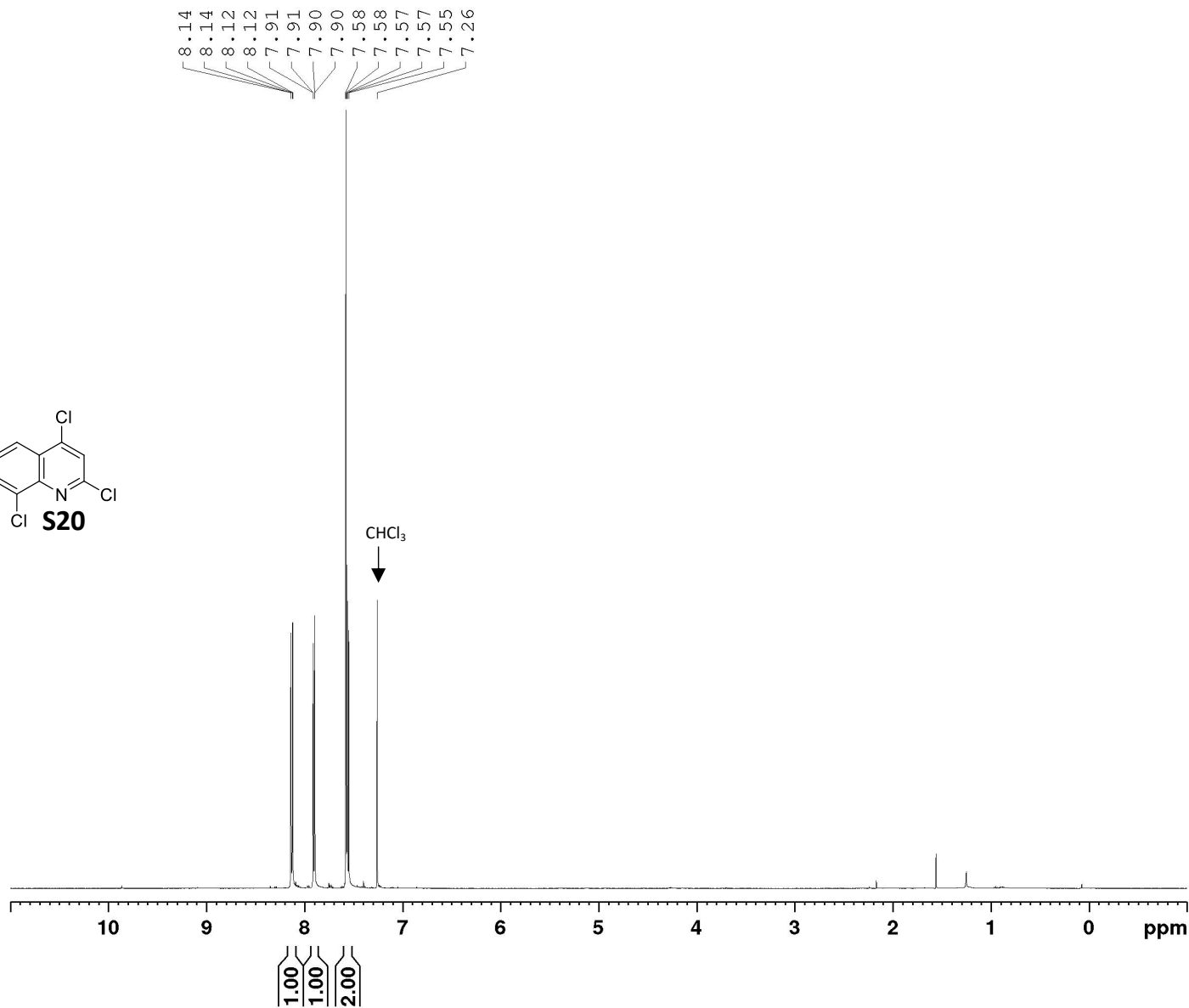
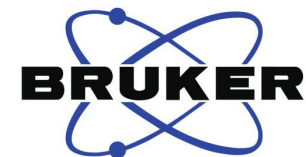


Current Data Parameters  
 NAME JPN-2-8\_repeat  
 EXPNO 17  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211119  
 Time 6.24 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

F2 - Processing parameters  
 SI 32768  
 SF 125.7829201 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

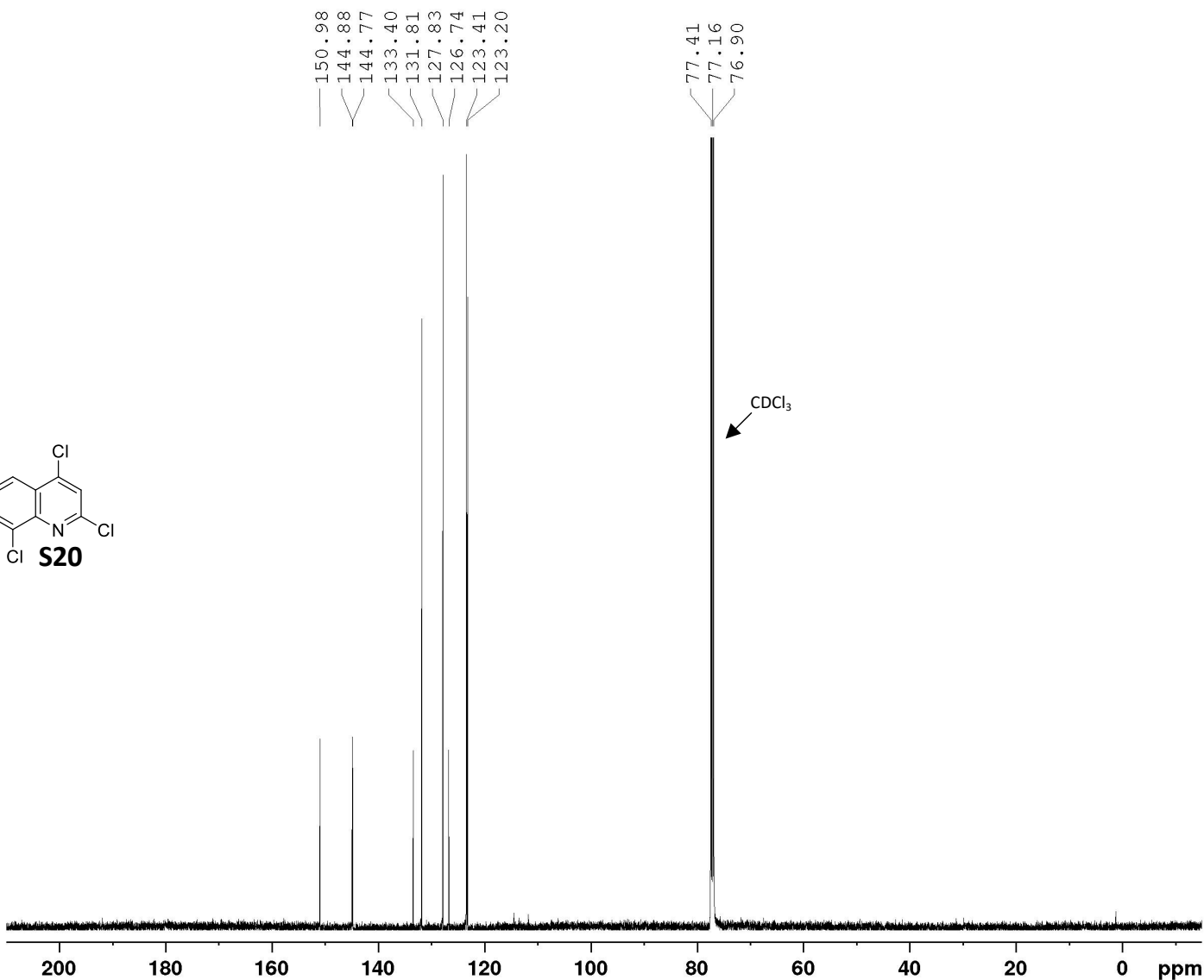




Current Data Parameters  
NAME JPN-2-42-3  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20210209  
Time 10.24 h  
INSTRUM spect  
PROBHD Z125869\_0055 (zg30)  
PULPROG zg30  
TD 65536  
SOLVENT CDCl<sub>3</sub>  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 151.18  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TDO 1  
SFO1 500.2330889 MHz  
NUC1 1H  
PO 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300121 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME JPN-2-42-3\_(2)  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211128  
 Time 21.42 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

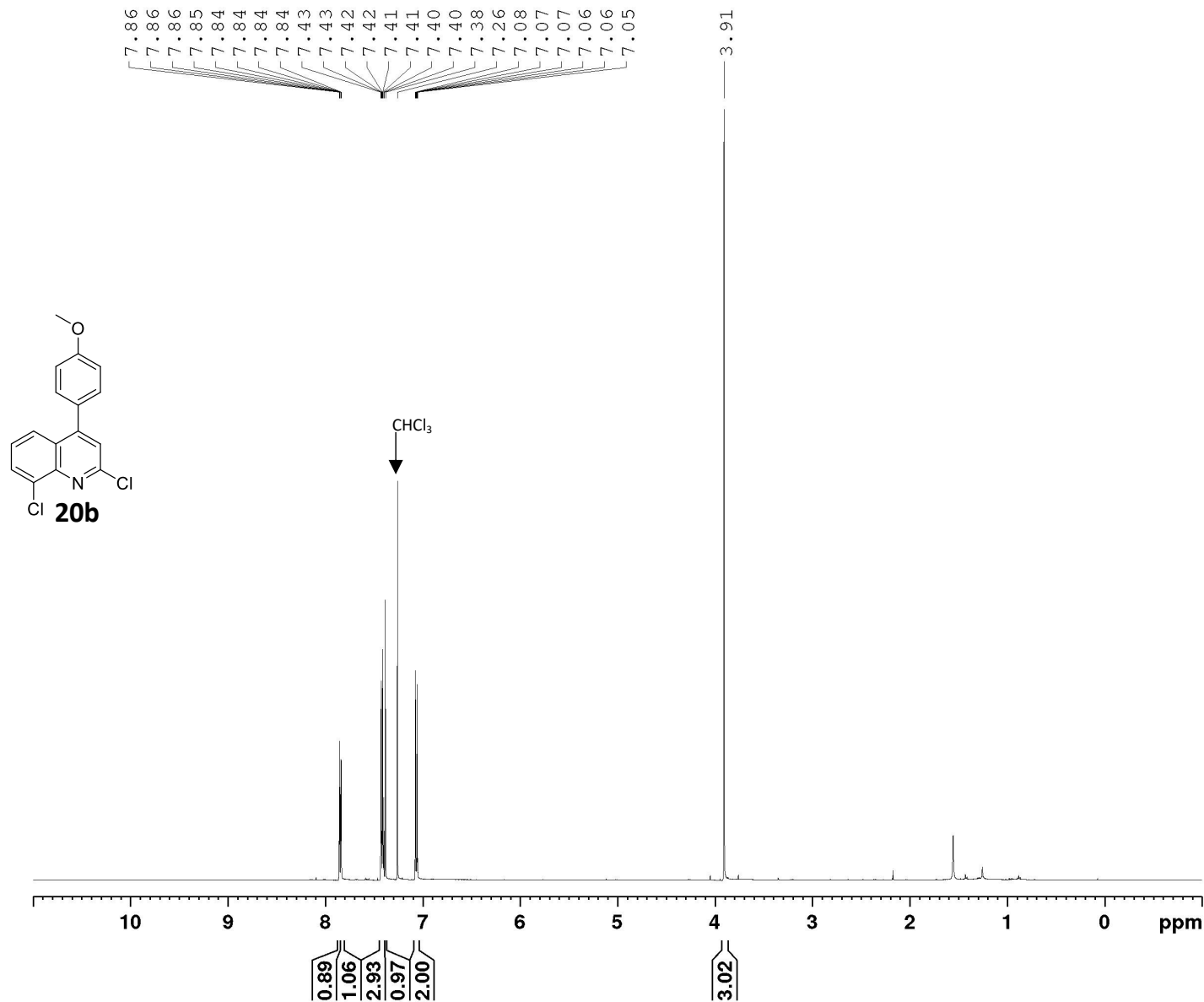
F2 - Processing parameters  
 SI 32768  
 SF 125.7829163 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

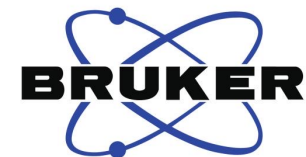
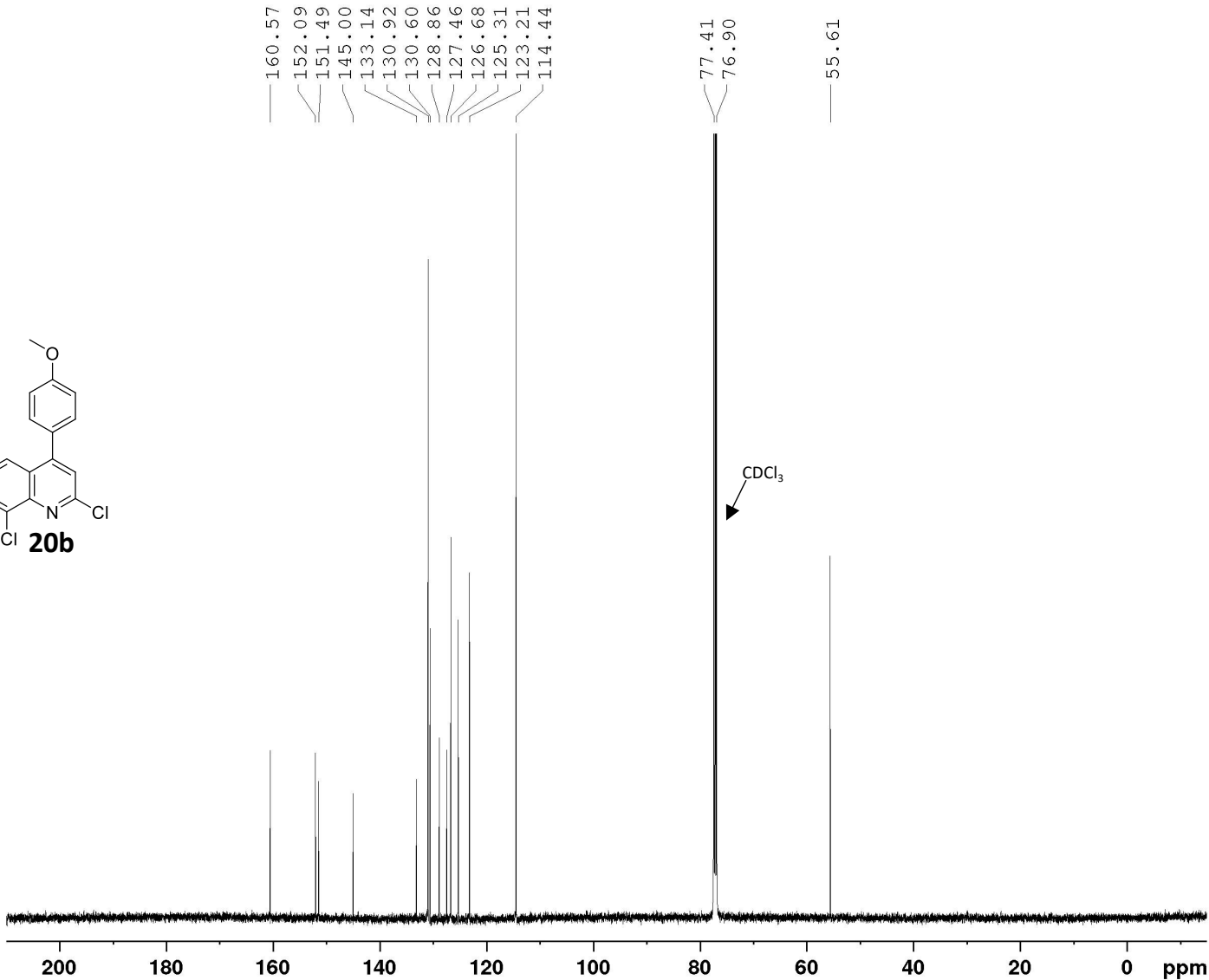
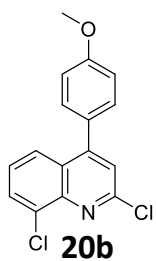


Current Data Parameters  
NAME JPN-2-47-3\_20-Nov-2021  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211120  
Time 21.51 h  
INSTRUM spect  
PROBHD z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 151.18  
DW 50.000 usec  
DE 16.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300121 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



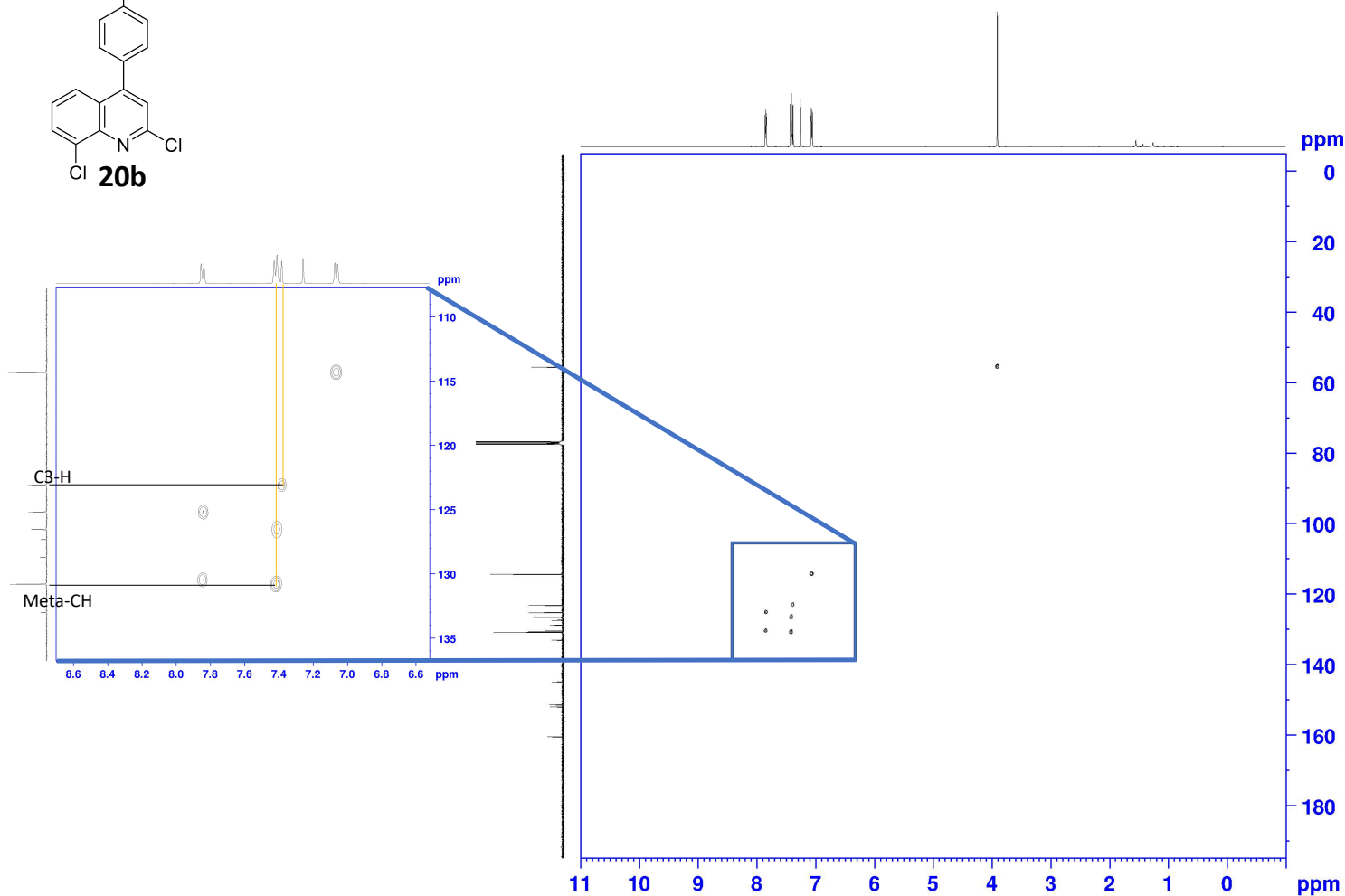
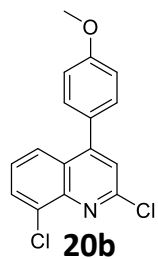


Current Data Parameters  
 NAME JPN-2-47-3\_20-Nov-2021  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211120  
 Time 22.47 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 ( )  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SF01 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SF02 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

F2 - Processing parameters  
 SI 32768  
 SF 125.7829164 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

# HSQC



```

Current Data Parameters
NAME      JPN-2-47-3
EXPNO     13
PROCNO    1

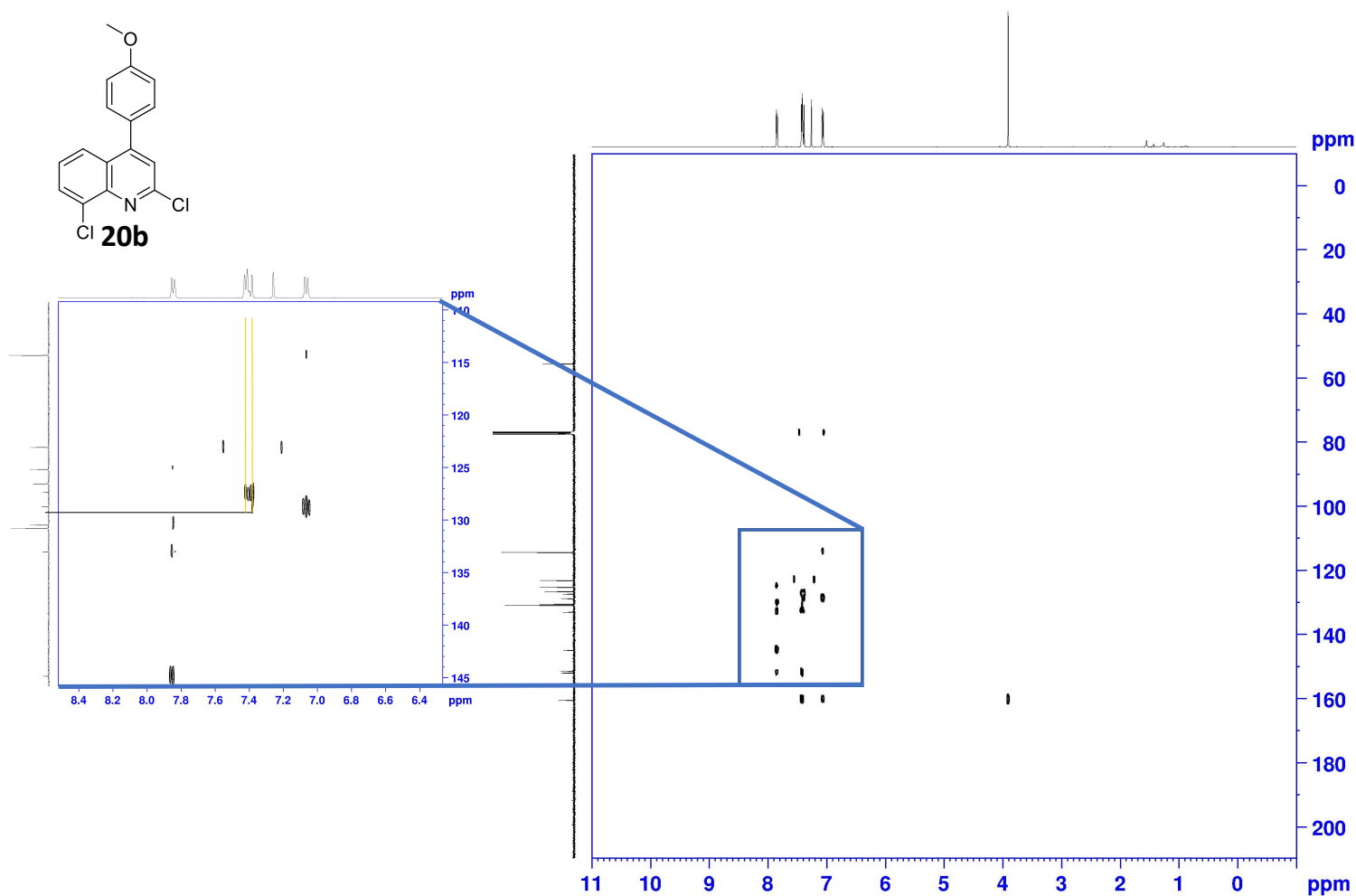
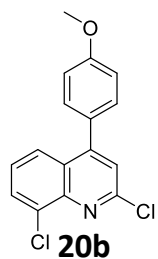
F2 - Acquisition Parameters
Date_     20211103
Time      18.57 h
INSTRUM   spect
PROBHD    2125869_0055 (
PULPROG   hsqcetgp
TD         1024
SOLVENT   CDCl3
NS         8
DS         16
SWH        7002.801 Hz
FIDRES     13.677346 Hz
AQ         0.0731136 sec
RG         190.44
DW         71.400 usec
DE         16.00 usec
TE         298.0 K
CNST2     145.000000
D0         0.0000300 sec
D1         1.5000000 sec
D4         0.00172414 sec
D11        0.0300000 sec
D16        0.0002000 sec
IND        0.00001990 sec
TDec      1
ZGOFITNS
SFO1      500.2330889 MHz
NUC1       1H
P1         12.00 usec
P2         24.00 usec
PLW1      11.44699955 W
SFO2      125.7948829 MHz
NUC2       13C
CPDPRG2   garp
P3         10.00 usec
P4         20.00 usec
PCPD2     70.00 usec
PLW2      56.90299988 W
PLW12     1.16129994 W
GPNAM(1)  SMOQ10.100
GE21      80.00 %
GPNAM(2)  SMOQ10.100
GE22      20.10 %
P16       1000.00 usec

F1 - Acquisition parameters
TD         256
SFO1      125.7949 MHz
FIDRES     196.293976 Hz
SW         199.735 ppm
FmMODE     Echo-Antiecho

F2 - Processing parameters
SI         1024
SF         500.2300122 MHz
WDW        QSINE
SSB         2
LB          0 Hz
GB          0
PC          1.40

F1 - Processing parameters
SI         1024
MC2        echo-antiecho
SF         125.7829335 MHz
WDW        QSINE
SSB         2
LB          0 Hz
GB          0
    
```

# HMBC



```

Current Data Parameters
NAME      JFN-2-47-3
EXPNO    11
PROCNO   1

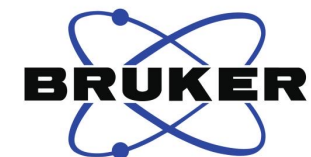
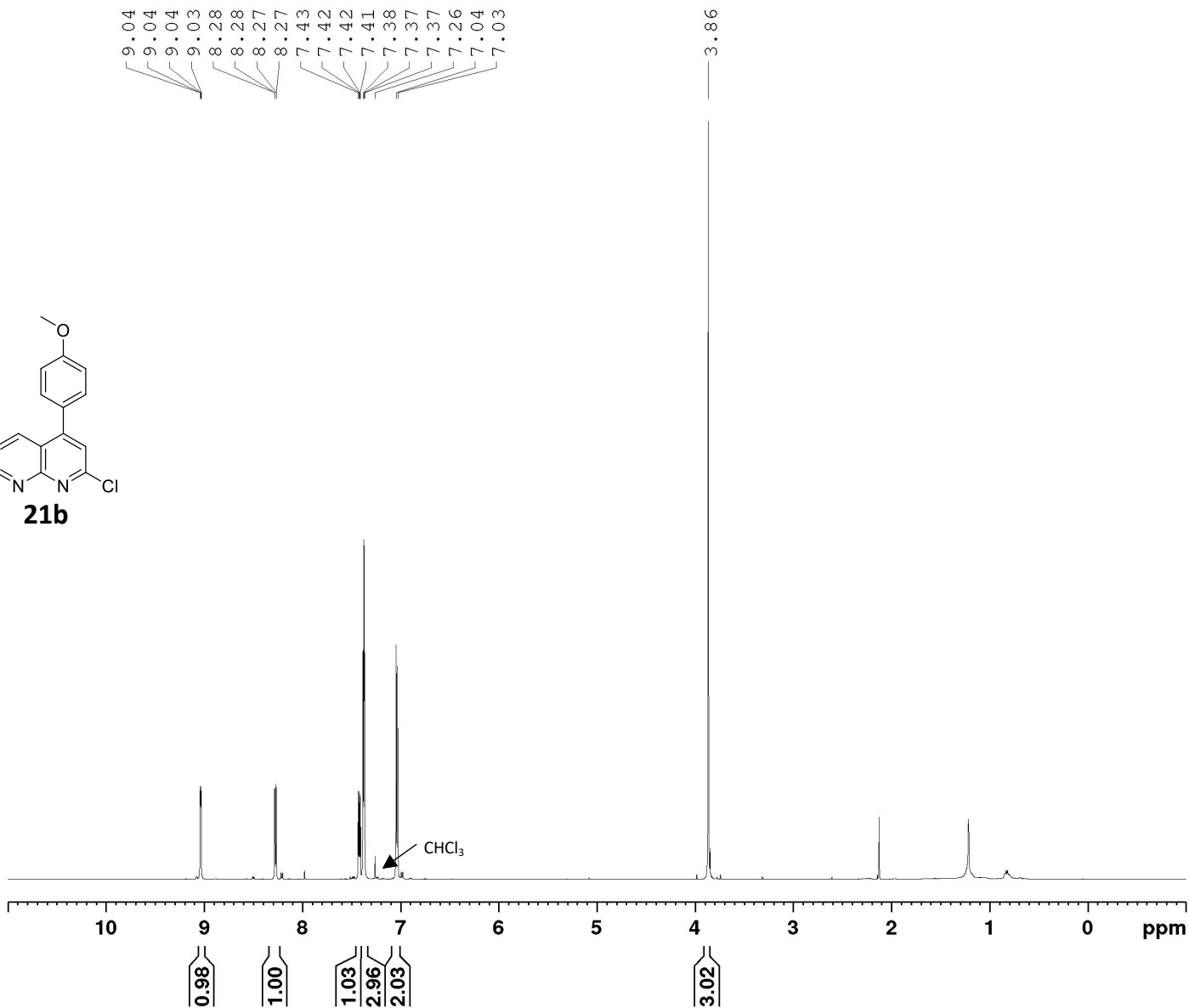
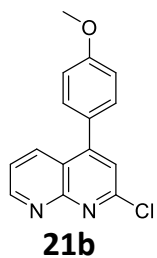
F2 - Acquisition Parameters
Date_    20211103
Time     18.01 h
INSTRUM  spect
PROBHD   Z125869_0055 (
PULPROG  hmbcgp1pndqf
TD       4096
SOLVENT  CDCl3
NS       4
DS       16
SWH      4201.681 Hz
FIDRES   2.051602 Hz
AQ       0.4874240 sec
RG       190.44
DW       119.000 usec
DE       16.00 usec
TE       298.0 K
CNST2    145.0000000
CNST13   10.0000000
D0       0.00000300 sec
D1       1.50000000 sec
D2       0.00344828 sec
D6       0.05000000 sec
D16      0.00020000 sec
IN0      0.00001810 sec
TDav     1
SFO1     500.2321971 MHz
NUC1     1H
P1       12.00 usec
P2       24.00 usec
PLW1     11.44699955 W
SFO2     125.7955118 MHz
NUC2     13C
P3       10.00 usec
PLW2     56.90299988 W
GPNAM[1] SMSQ10.100
GPZ1    30.00 %
GPNAM[2] SMSQ10.100
GPZ2    30.00 %
GPNAM[3] SMSQ10.100
GPZ3    40.10 %
P16     1000.00 usec

F1 - Acquisition parameters
TD       128
SFO1     125.7955 MHz
FIDRES   431.629822 Hz
SW       219.597 ppm
FwMODE   QF

F2 - Processing parameters
SI       4096
SF       500.2300126 MHz
WDW      QSINE
SSB      0
LB       0 Hz
GB       0
PC       1.40

F1 - Processing parameters
SI       1024
MC2      QF
SF       125.7829432 MHz
WDW      QSINE
SSB      0
LB       0 Hz
GB       0
    
```

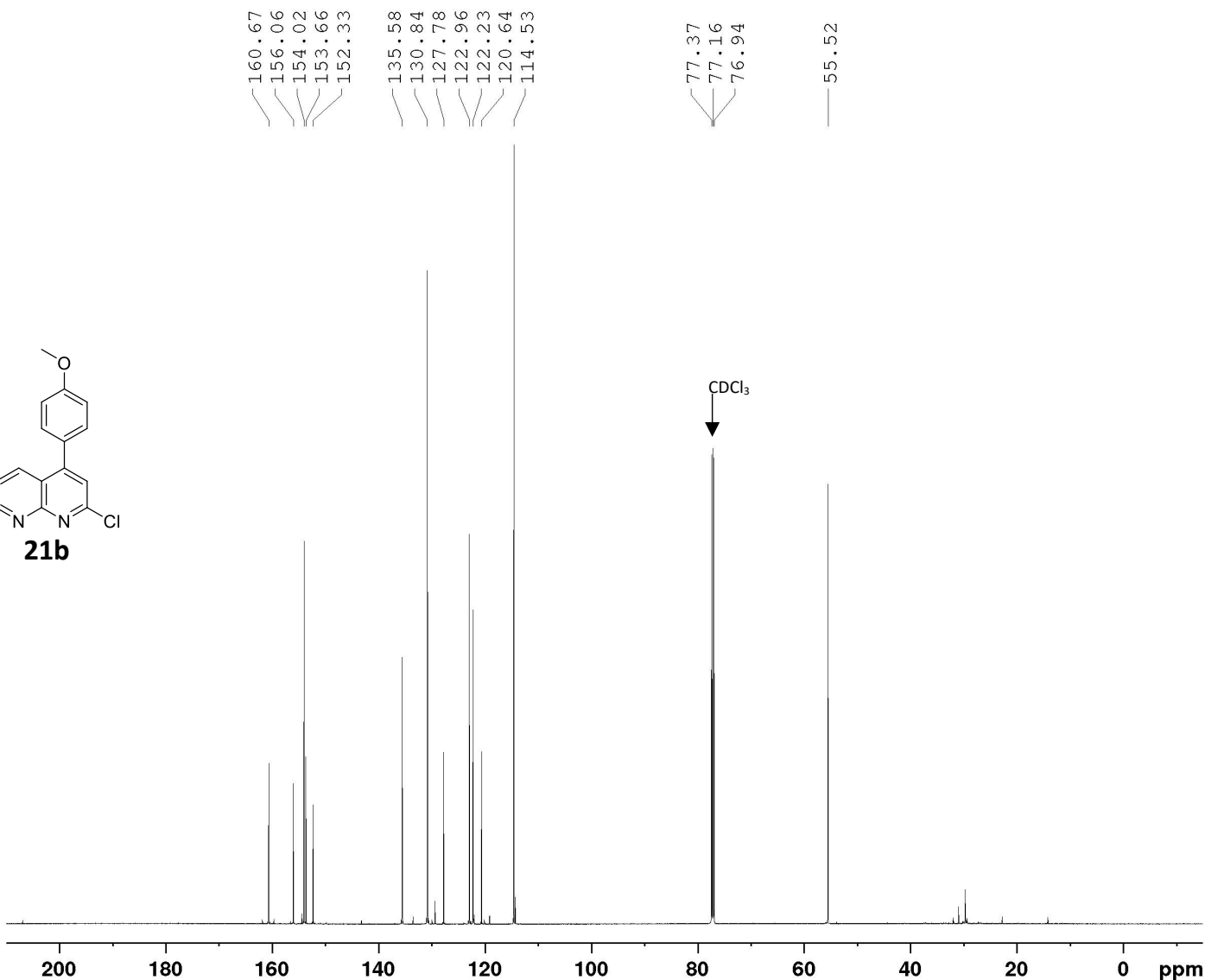
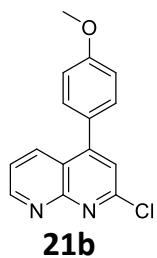
C9 correlation to *meta* C-H and C3-H



Current Data Parameters  
 NAME JPN-1-169-2\_C4-monoaryl  
 EXPNO 22  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20200527  
 Time 10.24 h  
 INSTRUM spect  
 PROBHD Z127277\_0002 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 12019.230 Hz  
 FIDRES 0.366798 Hz  
 AQ 2.7262976 sec  
 RG 4.41  
 DW 41.600 usec  
 DE 10.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TD0 1  
 SFO1 600.1337060 MHz  
 NUC1 1H  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 5.59999990 W

F2 - Processing parameters  
 SI 65536  
 SF 600.1300147 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

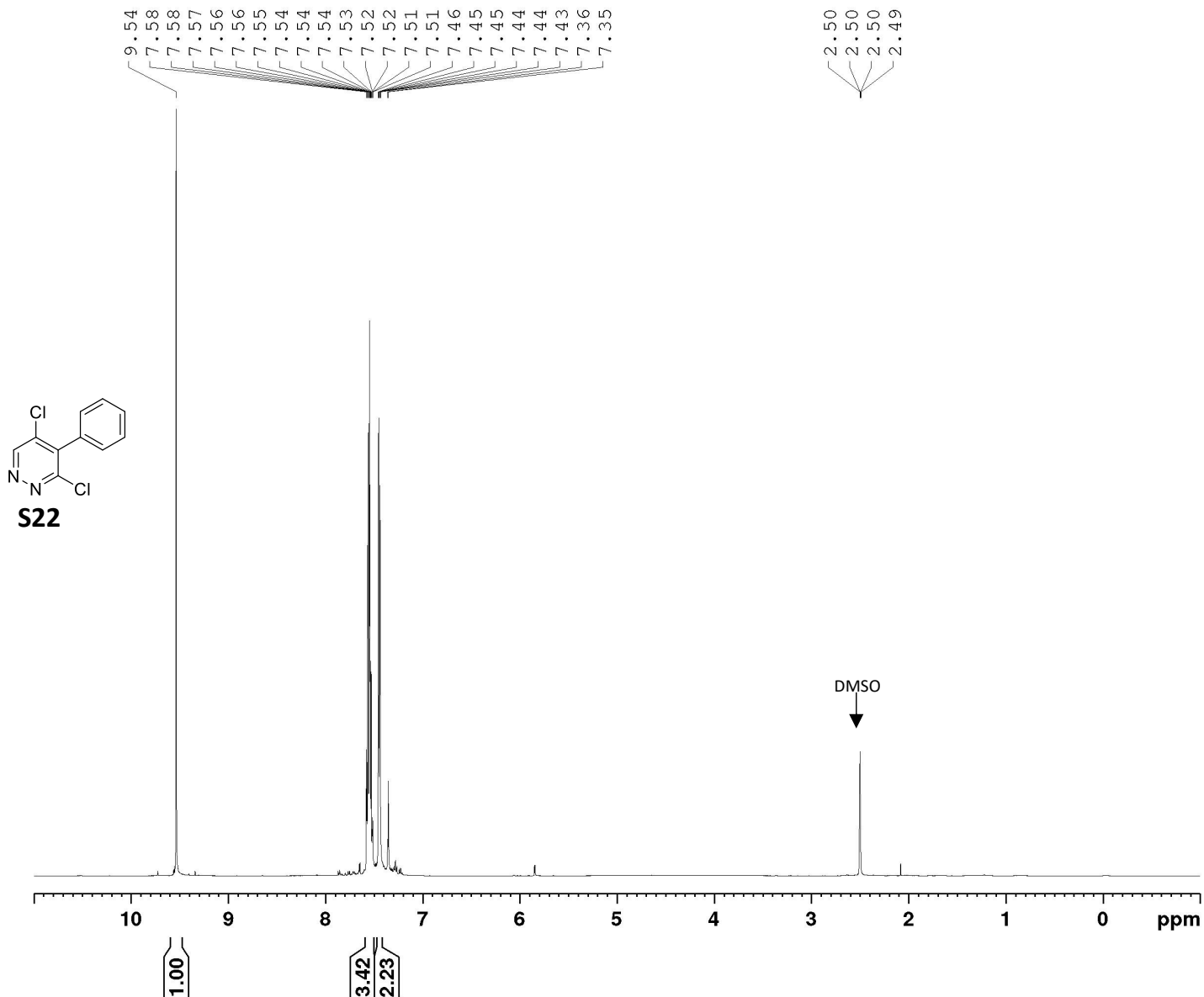


Current Data Parameters  
 NAME JPN-1-169-2\_C4-monoaryl  
 EXPNO 23  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20200527  
 Time 11.16 h  
 INSTRUM spect  
 PROBHD Z127277\_0002 (  
 PULPROG zgpg30  
 ID 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 36057.691 Hz  
 FIDRES 1.100393 Hz  
 AQ 0.9087659 sec  
 RG 184.4  
 DW 13.867 usec  
 DE 18.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 150.9178981 MHz  
 NUC1 13C  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 91.00000000 W  
 SFO2 600.1324005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 70.00 usec  
 PLW2 5.599999990 W  
 PLW12 0.07314300 W  
 PLW13 0.03679000 W

F2 - Processing parameters  
 SI 32768  
 SF 150.9028026 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 EC 1.40

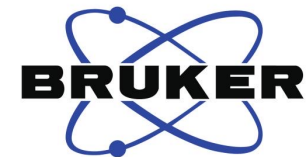
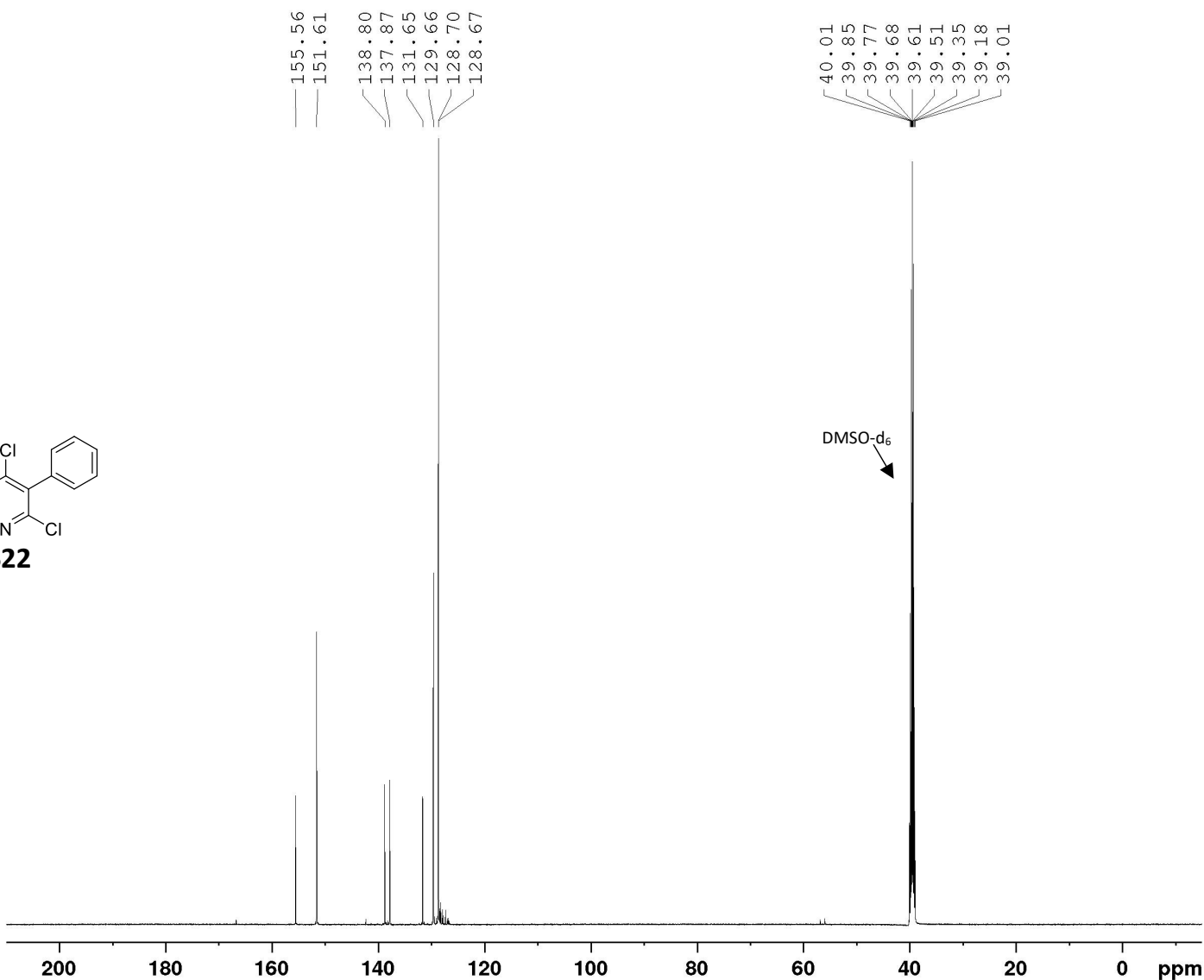
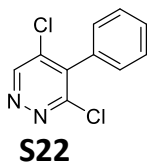




Current Data Parameters  
NAME JPN-2-22-2\_dried  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20201010  
Time 10.30 h  
INSTRUM spect  
PROBHD Z125869\_0055 ( )  
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 69.97  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TDO 1  
SFO1 500.2330889 MHz  
NUC1 1H  
PO 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

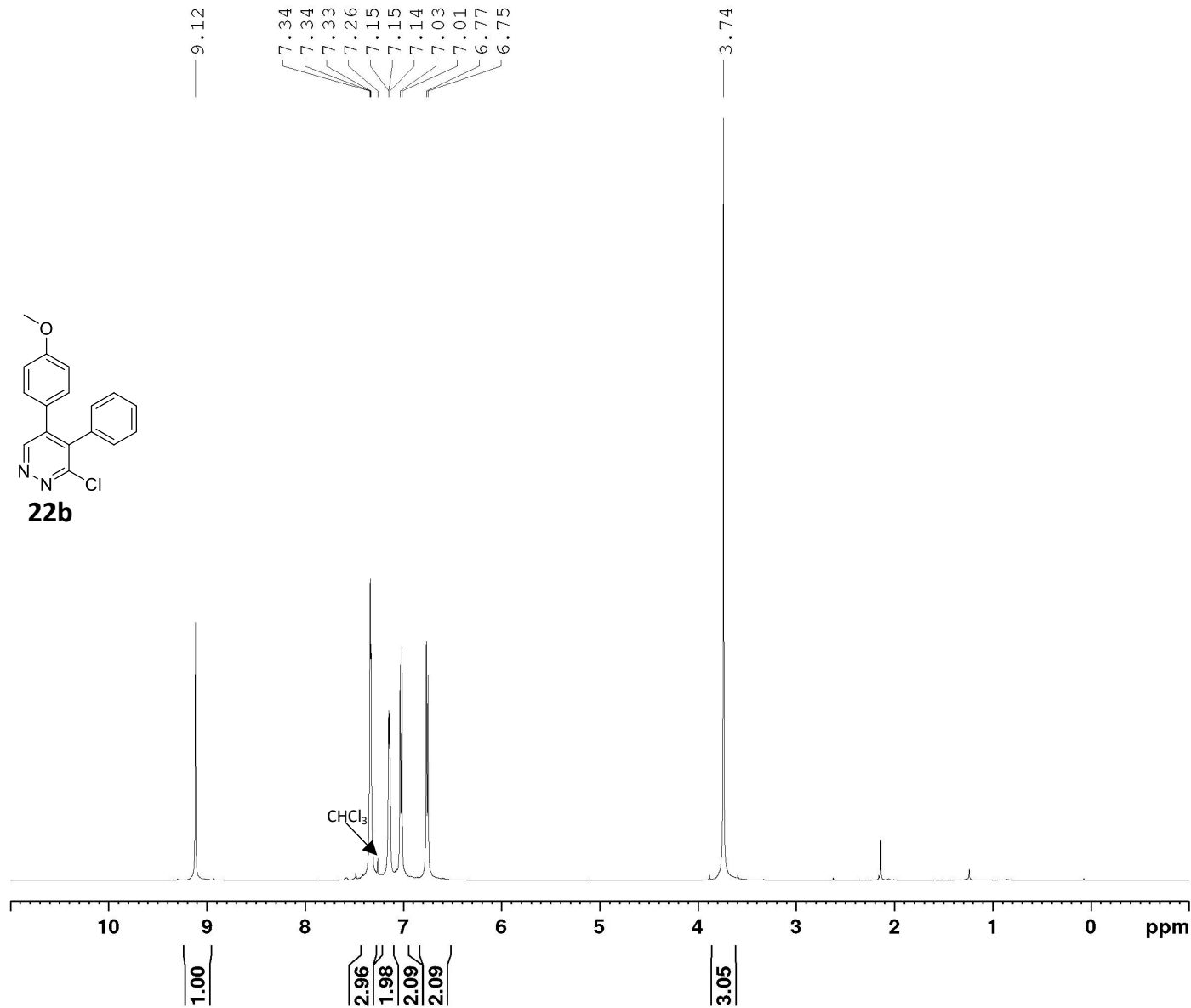
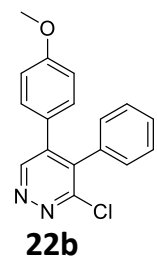
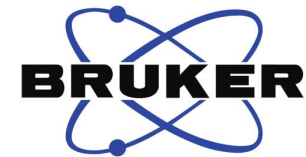
F2 - Processing parameters  
SI 65536  
SF 500.2300035 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME JPN-2-22-2\_dried  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20201010  
 Time 12.37 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT DMSO  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

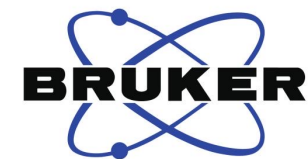
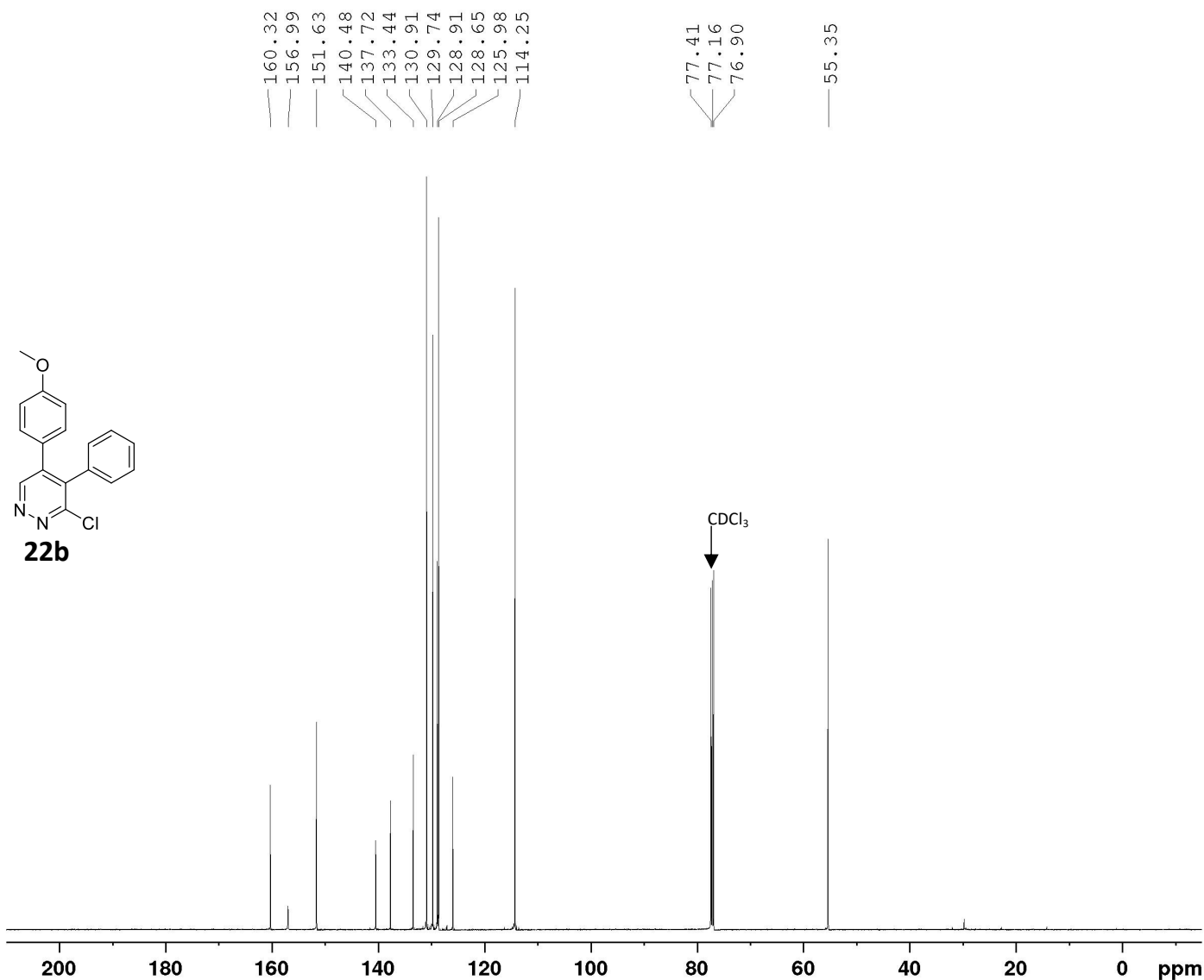
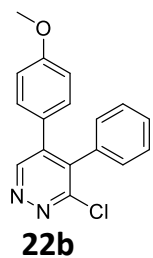
F2 - Processing parameters  
 SI 32768  
 SF 125.7829976 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
NAME JPN-2-27-1\_repeat\_C5-mono  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20201130  
Time 17.40 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 22.16  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

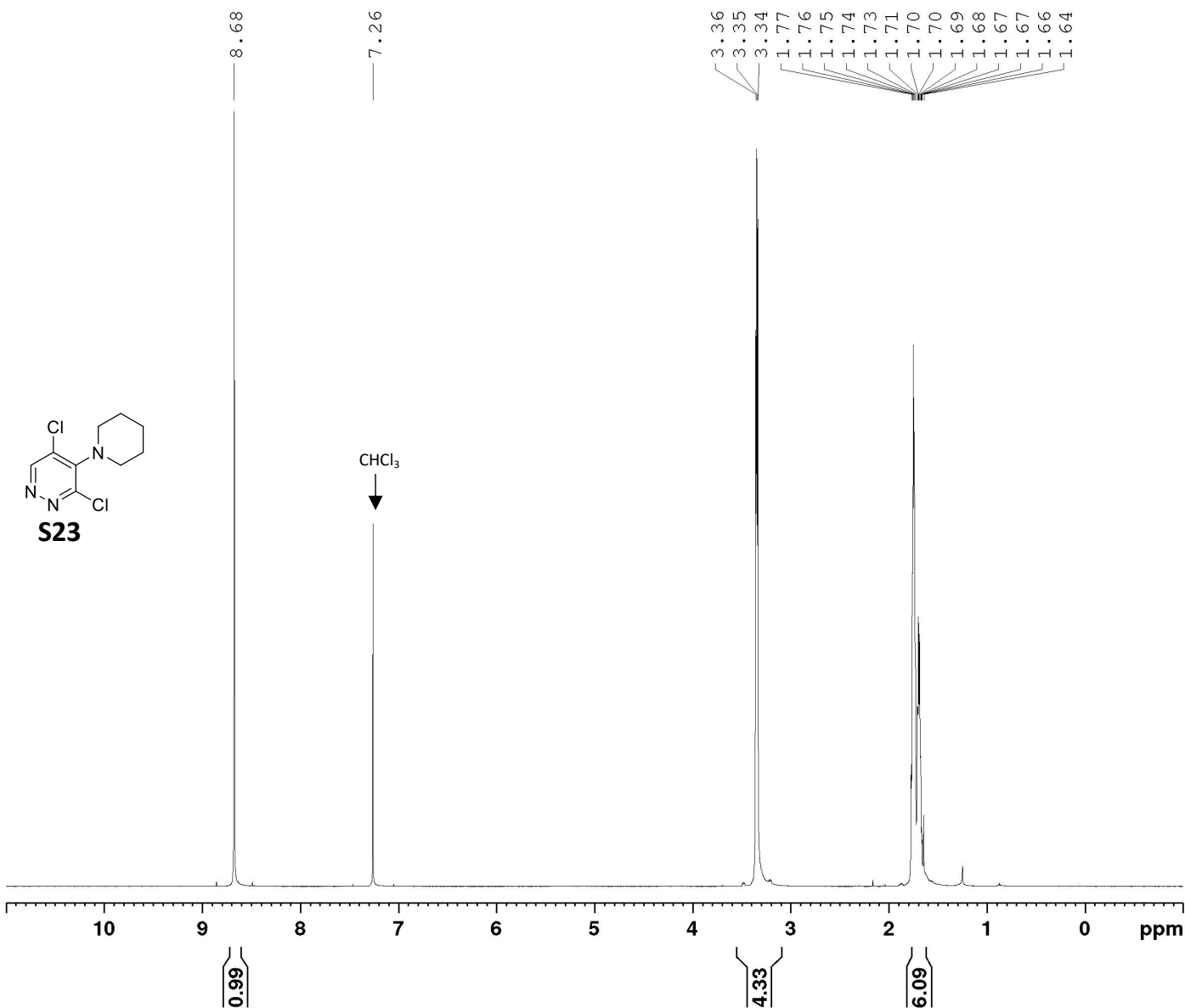
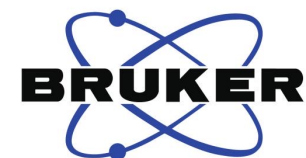
F2 - Processing parameters  
SI 65536  
SF 500.2300118 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME JPN-2-27-1\_repeat  
 EXPNO 17  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211119  
 Time 4.28 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

F2 - Processing parameters  
 SI 32768  
 SF 125.7829267 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
NAME JPN-2-14-1\_C4-subst%2E  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters

Date\_ 20200914  
Time 3.42 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 122.05  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters

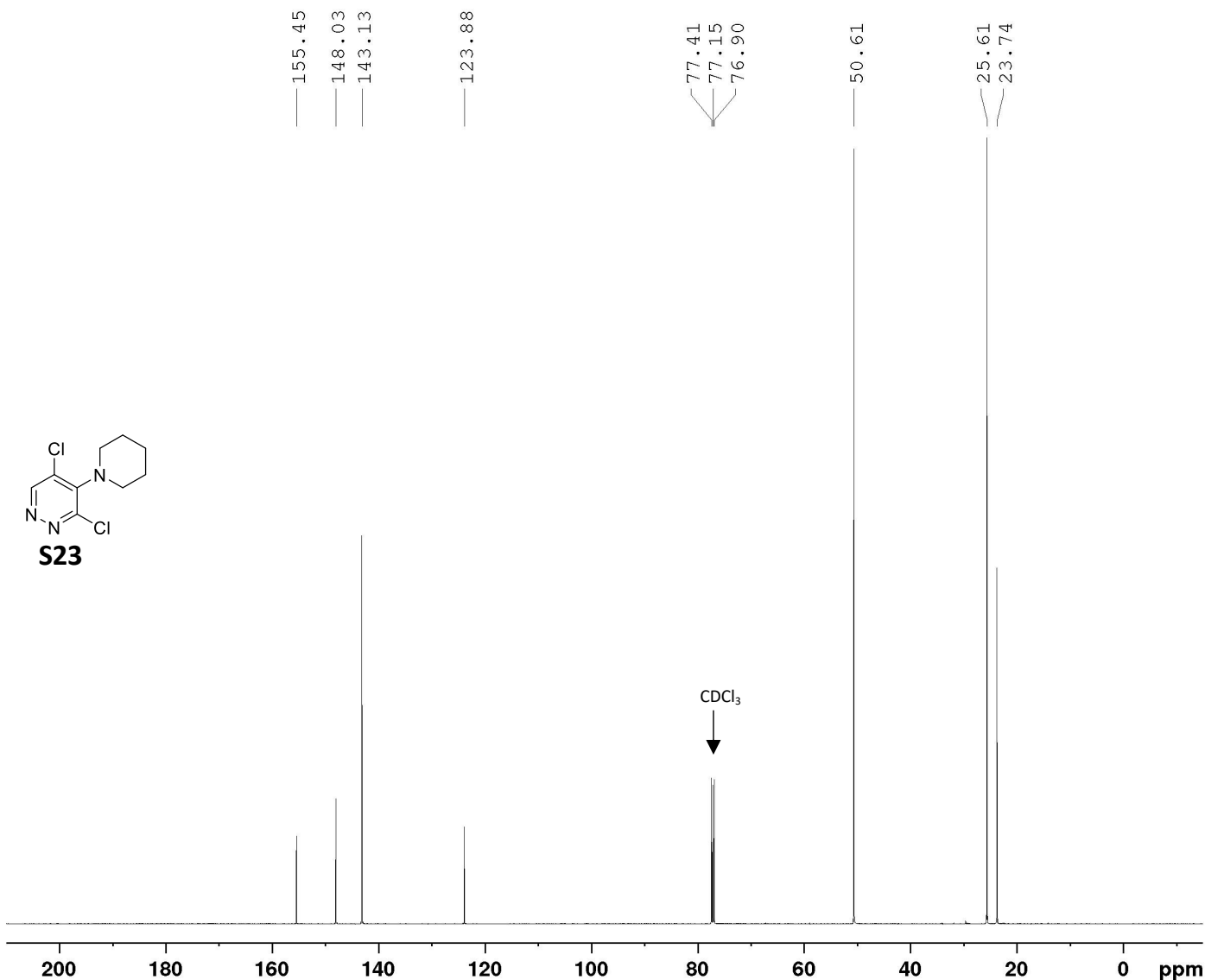
SI 65536  
SF 500.2300122 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME JPN-2-14-1\_(C4-subst.)  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211127  
Time 20.52 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.908261 Hz  
AQ 1.1010048 sec  
RG 190.44  
DW 16.800 usec  
DE 18.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TDO 1  
SFO1 125.7955118 MHz  
NUC1 13C  
P0 3.33 usec  
P1 10.00 usec  
PLW1 56.90299988 W  
SFO2 500.2320009 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 11.44699955 W  
PLW12 0.25756001 W  
PLW13 0.12955000 W

F2 - Processing parameters  
SI 32768  
SF 125.7829371 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

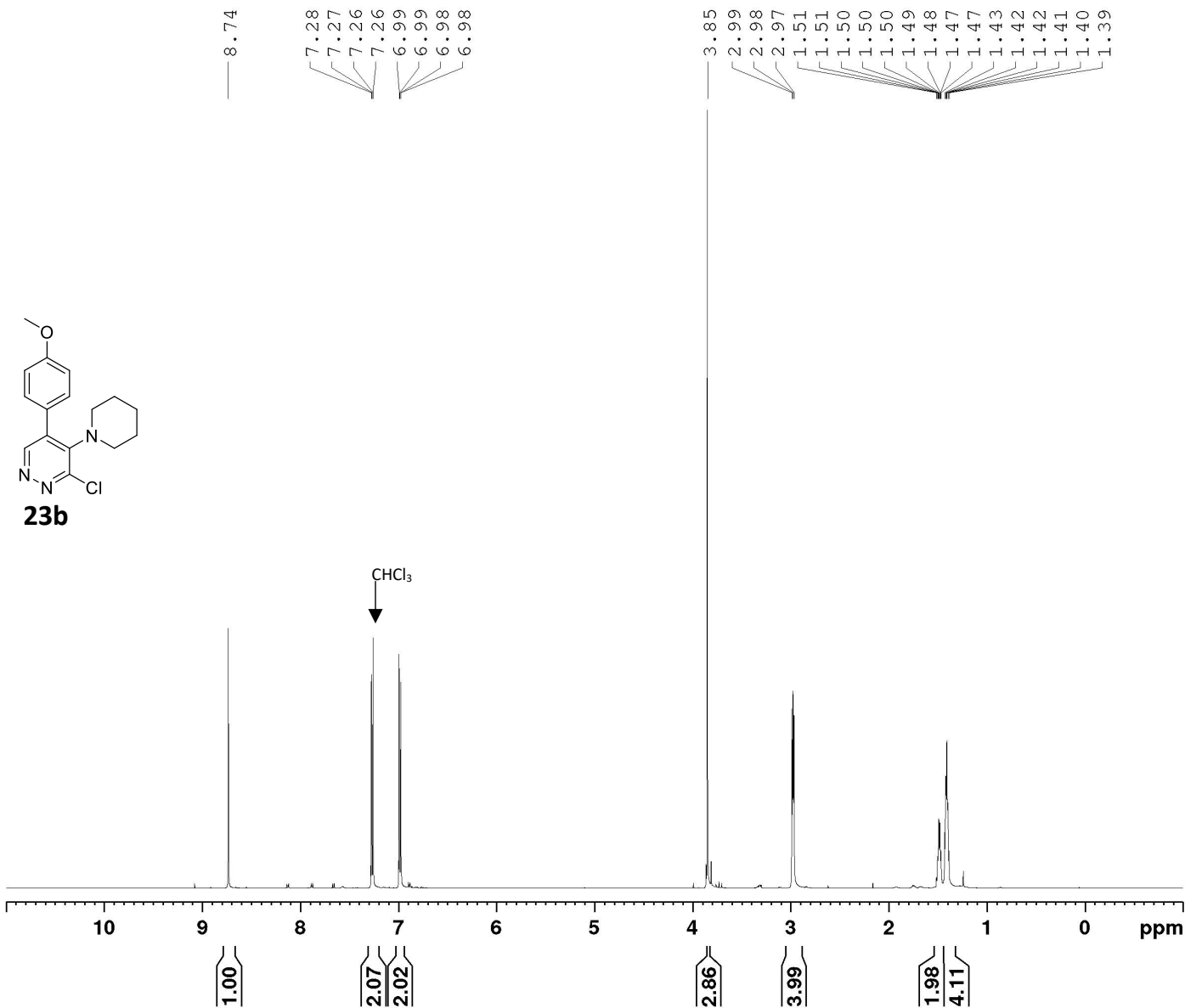


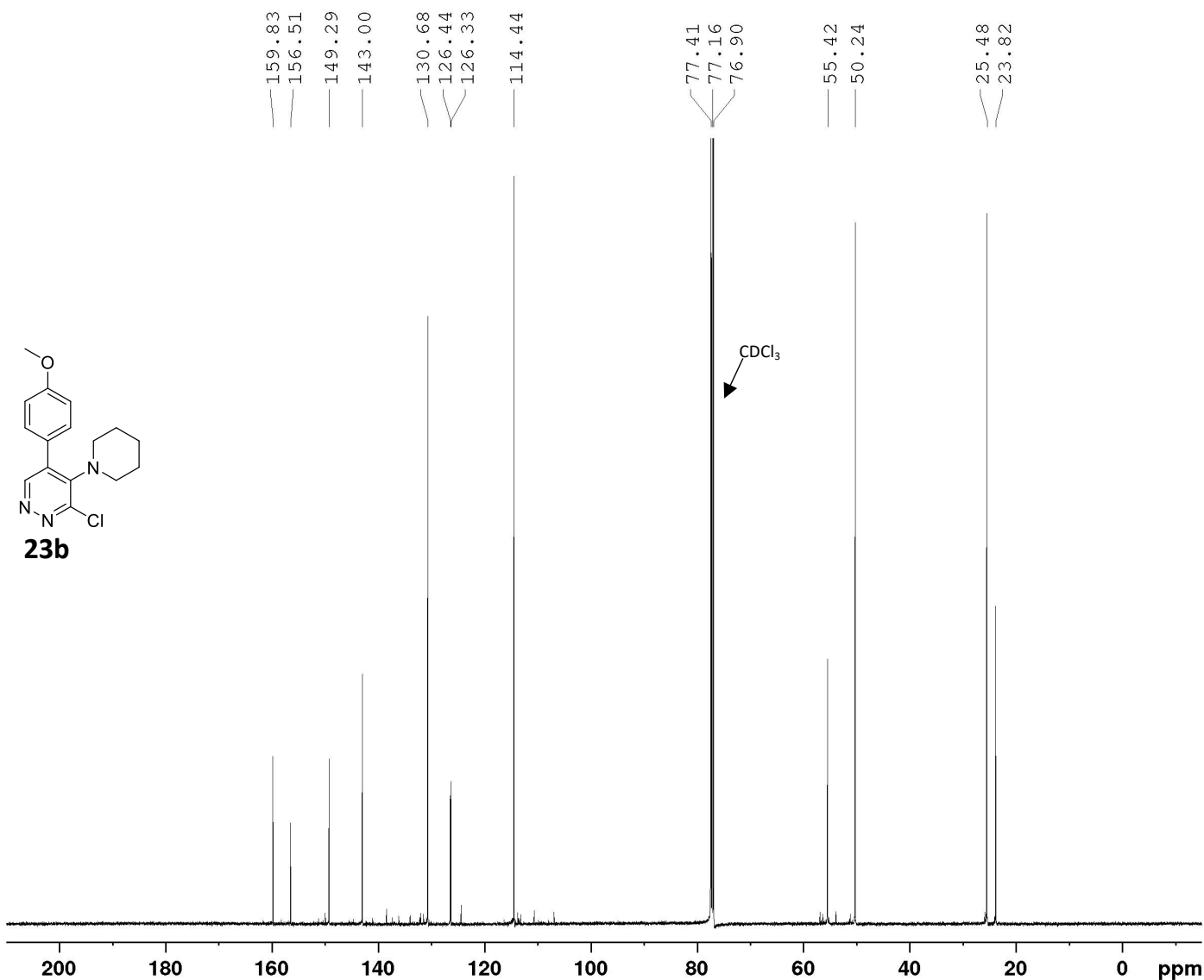
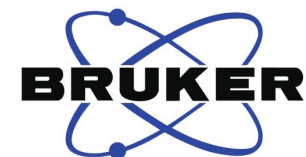


Current Data Parameters  
NAME JPN-2-27-2\_C5-mono  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20201130  
Time 18.00 h  
INSTRUM spect  
PROBHD Z125869\_0055 ( )  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 35.18  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300119 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



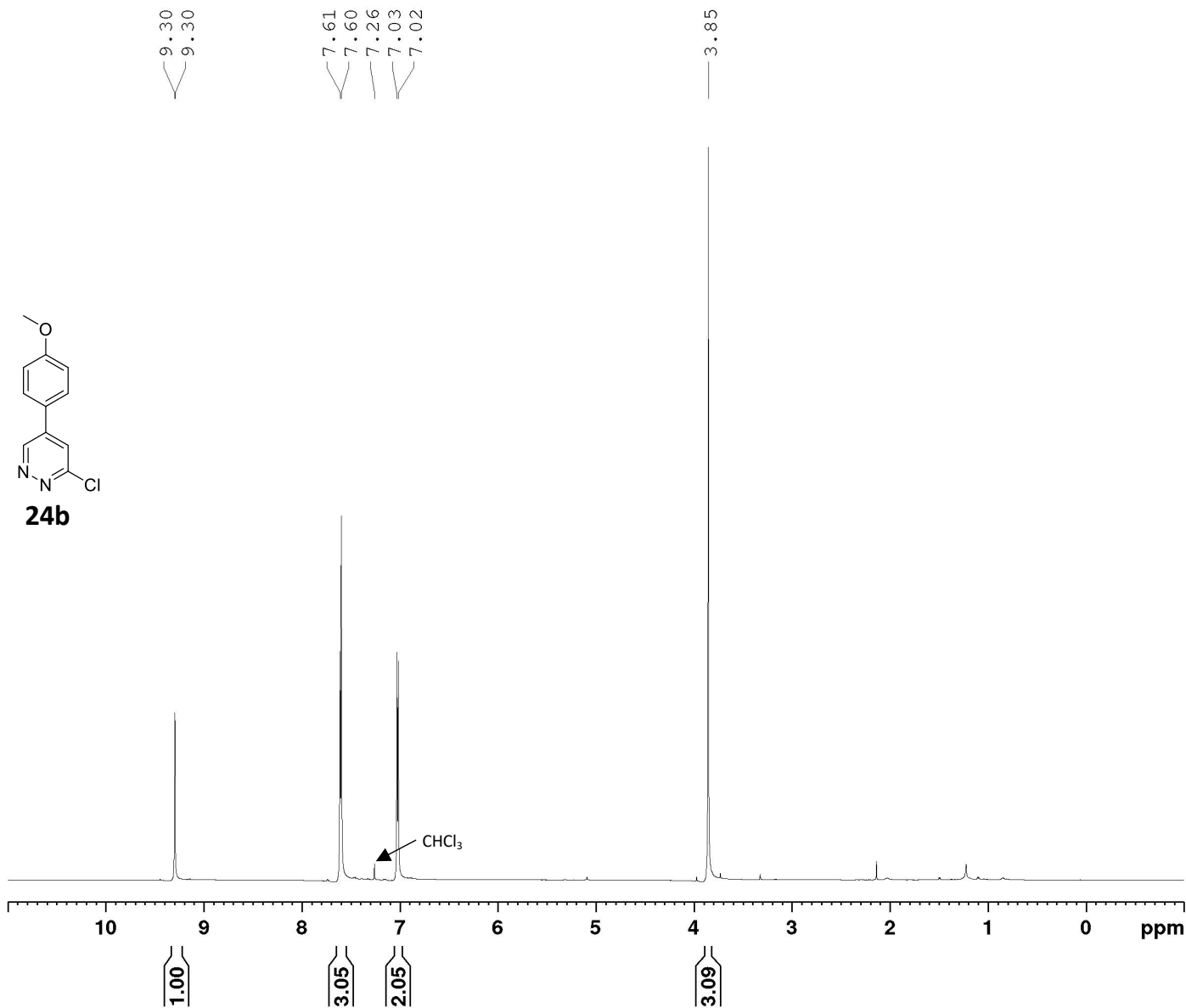
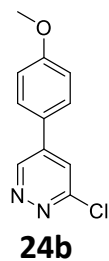


Current Data Parameters  
NAME JPN-2-27-2  
EXPNO 17  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211119  
Time 5.26 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.908261 Hz  
AQ 1.1010048 sec  
RG 190.44  
DW 16.800 usec  
DE 18.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 125.7955118 MHz  
NUC1 13C  
P0 3.33 usec  
P1 10.00 usec  
PLW1 56.90299988 W  
SFO2 500.2320009 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 11.44699955 W  
PLW12 0.25756001 W  
PLW13 0.12955000 W

F2 - Processing parameters  
SI 32768  
SF 125.7829191 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

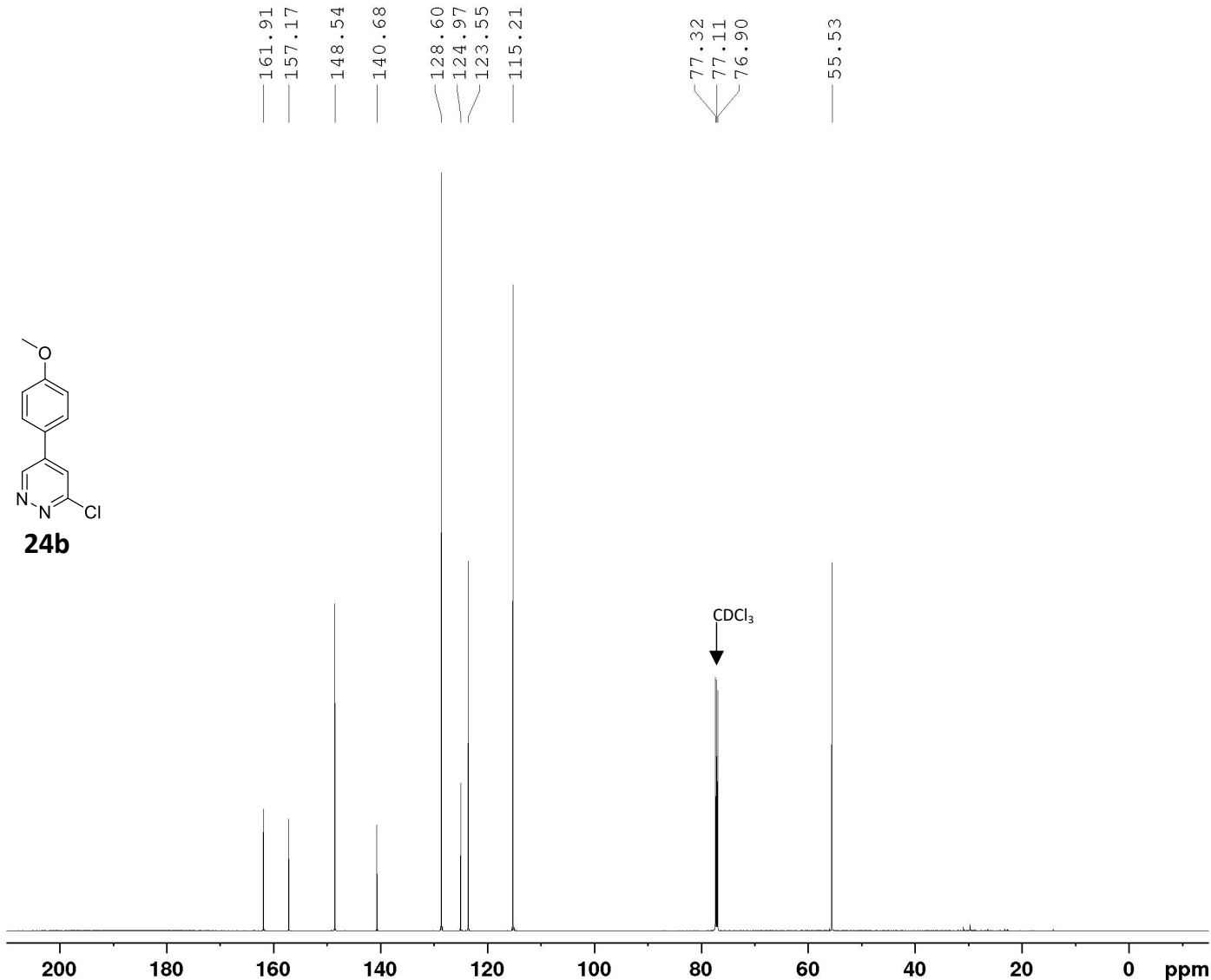
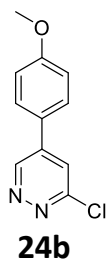




Current Data Parameters  
NAME JPN-1-166-2\_C5-Mono  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200519  
Time 13.52 h  
INSTRUM spect  
PROBHD Z127277\_0002 ( )  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.366798 Hz  
AQ 2.7262976 sec  
RG 4.41  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1  
SFO1 600.1337060 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 5.59999990 W

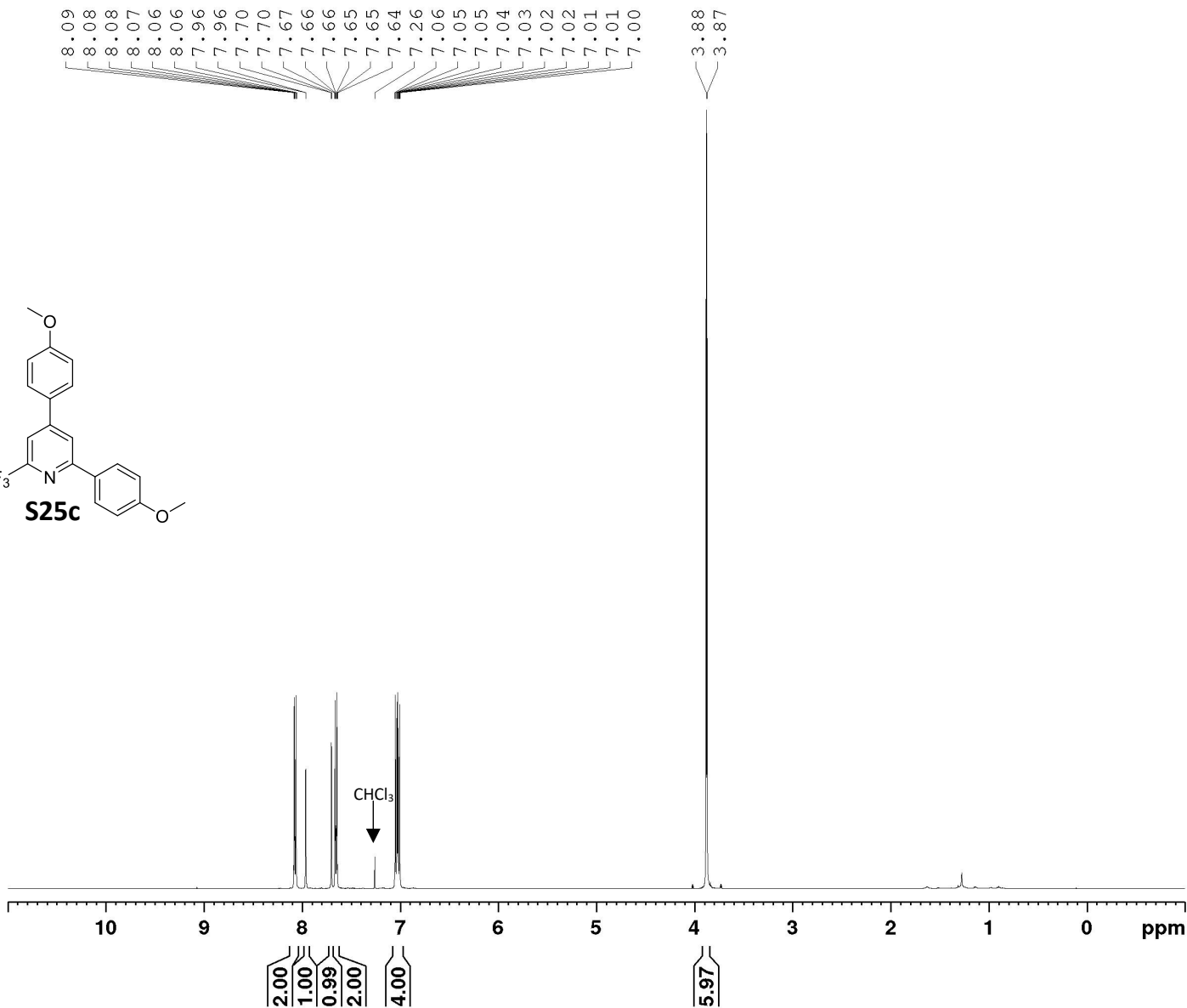
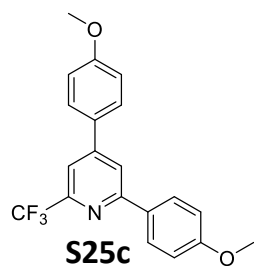
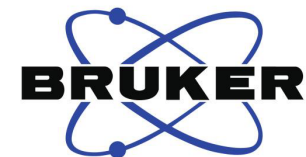
F2 - Processing parameters  
SI 65536  
SF 600.1300144 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME JPN-1-166-2\_C5-Mono  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20200519  
 Time 15.36 h  
 INSTRUM spect  
 PROBHD z127277\_0002 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 36057.691 Hz  
 FIDRES 1.100393 Hz  
 AQ 0.9087659 sec  
 RG 184.4  
 DW 13.867 usec  
 DE 18.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 150.9178981 MHz  
 NUC1 13C  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 91.00000000 W  
 SFO2 600.1324005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 70.00 usec  
 PLW2 5.59999990 W  
 PLW12 0.07314300 W  
 PLW13 0.03679000 W

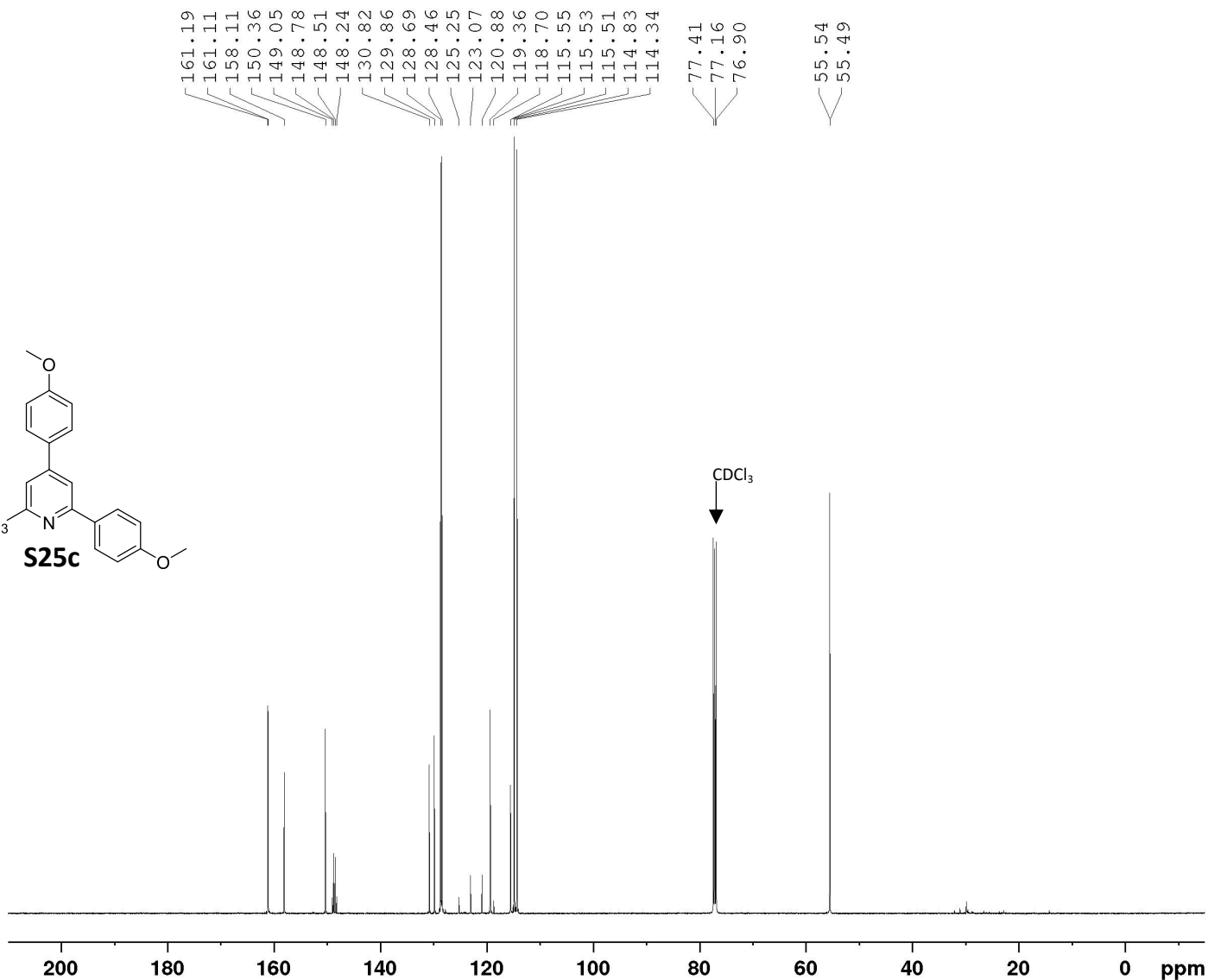
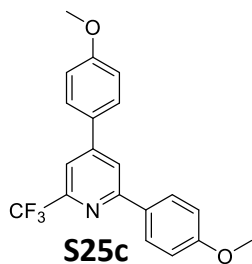
F2 - Processing parameters  
 SI 32768  
 SF 150.9028090 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
NAME JPN-1-187-2\_20-Nov-2021  
EXPNO 20  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211121  
Time 1.37 h  
INSTRUM spect  
PROBHD z125869\_0055 (  
PULPROG zg30  
ID 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 30.54  
DW 50.000 usec  
DE 16.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300119 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME JPN-1-187-2\_DA  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20200812  
 Time 3.15 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDC13  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

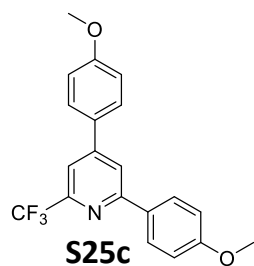
F2 - Processing parameters  
 SI 32768  
 SF 125.7829205 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
NAME JPN-1-187-2  
EXPNO 18  
PROCNO 1

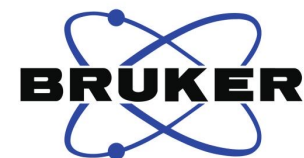
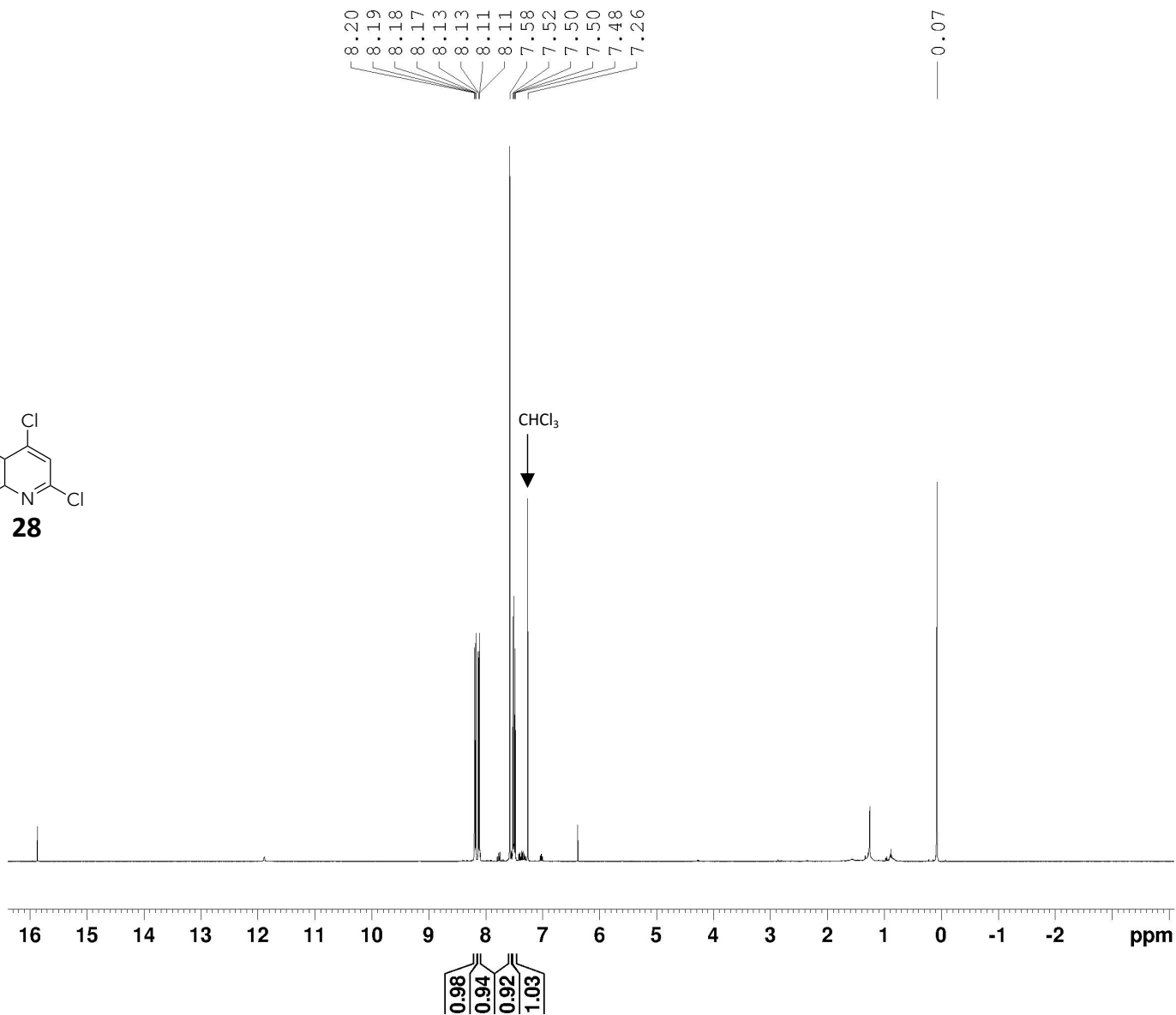
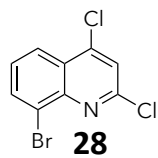
F2 - Acquisition Parameters  
Date\_ 20211120  
Time 3.44 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zgflqn  
TD 131072  
SOLVENT CDC13  
NS 16  
DS 4  
SWH 113636.367 Hz  
FIDRES 1.733953 Hz  
AQ 0.5767168 sec  
RG 15.61  
DW 4.400 usec  
DE 18.00 usec  
TE 298.0 K  
D1 1.00000000 sec  
TD0 1  
SFO1 470.6394024 MHz  
NUC1 19F  
P1 15.00 usec  
PLW1 11.70800018 W

F2 - Processing parameters  
SI 65536  
SF 470.6864712 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



-67.96

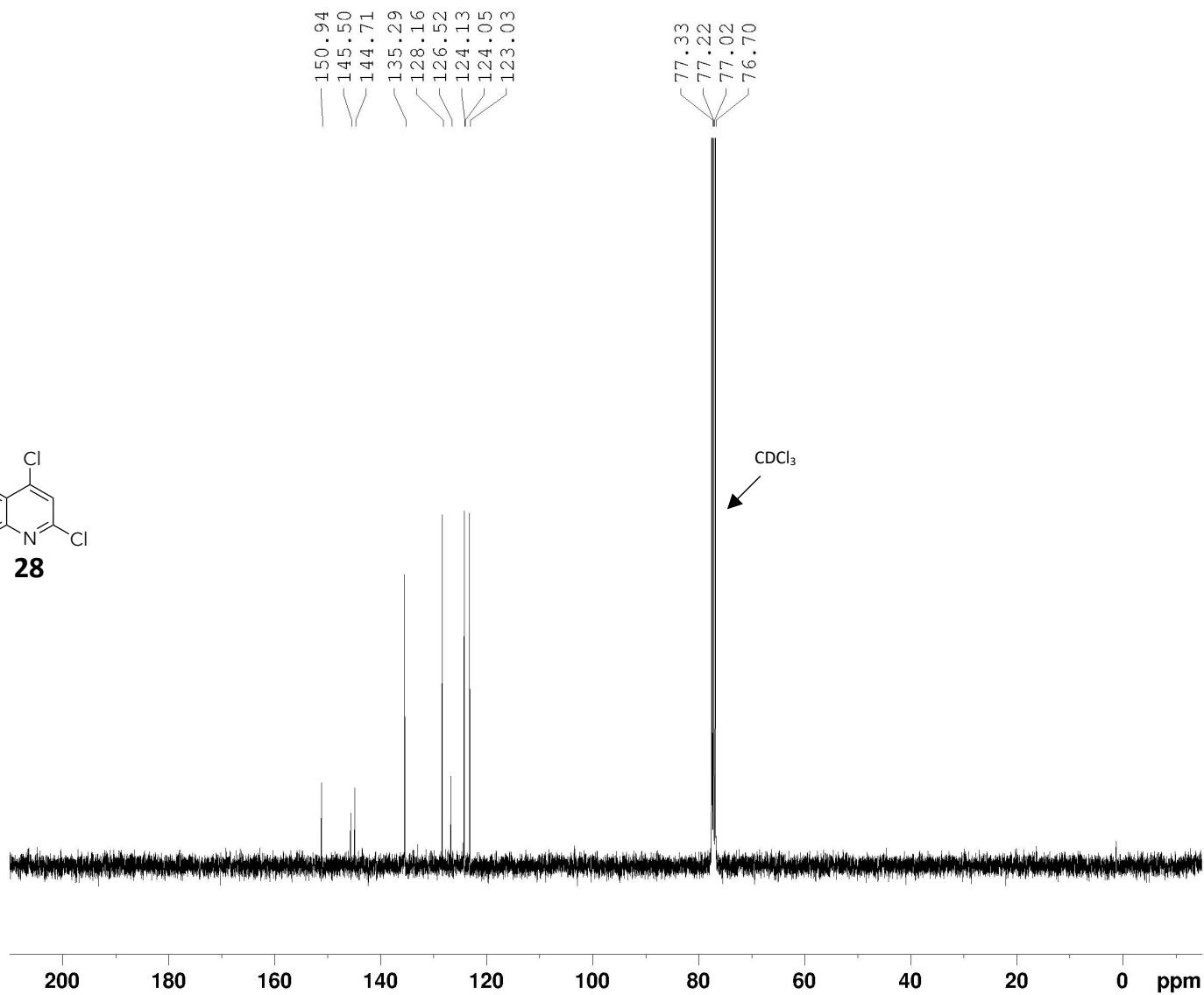
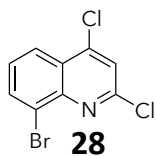
0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 ppm



Current Data Parameters  
 NAME NL-1-117-1H  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211208  
 Time 14.16 h  
 INSTRUM Avance Neo  
 PROBHD z152088\_0031 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 16  
 DS 2  
 SWH 8196.722 Hz  
 FIDRES 0.250144 Hz  
 AQ 3.9976959 sec  
 RG 101  
 DW 61.000 usec  
 DE 13.89 usec  
 TE 298.0 K  
 D1 1.00000000 sec  
 TD0 1  
 SFO1 400.1324708 MHz  
 NUC1 1H  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 24.03499985 W

F2 - Processing parameters  
 SI 65536  
 SF 400.1300098 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



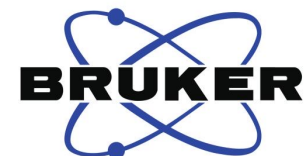
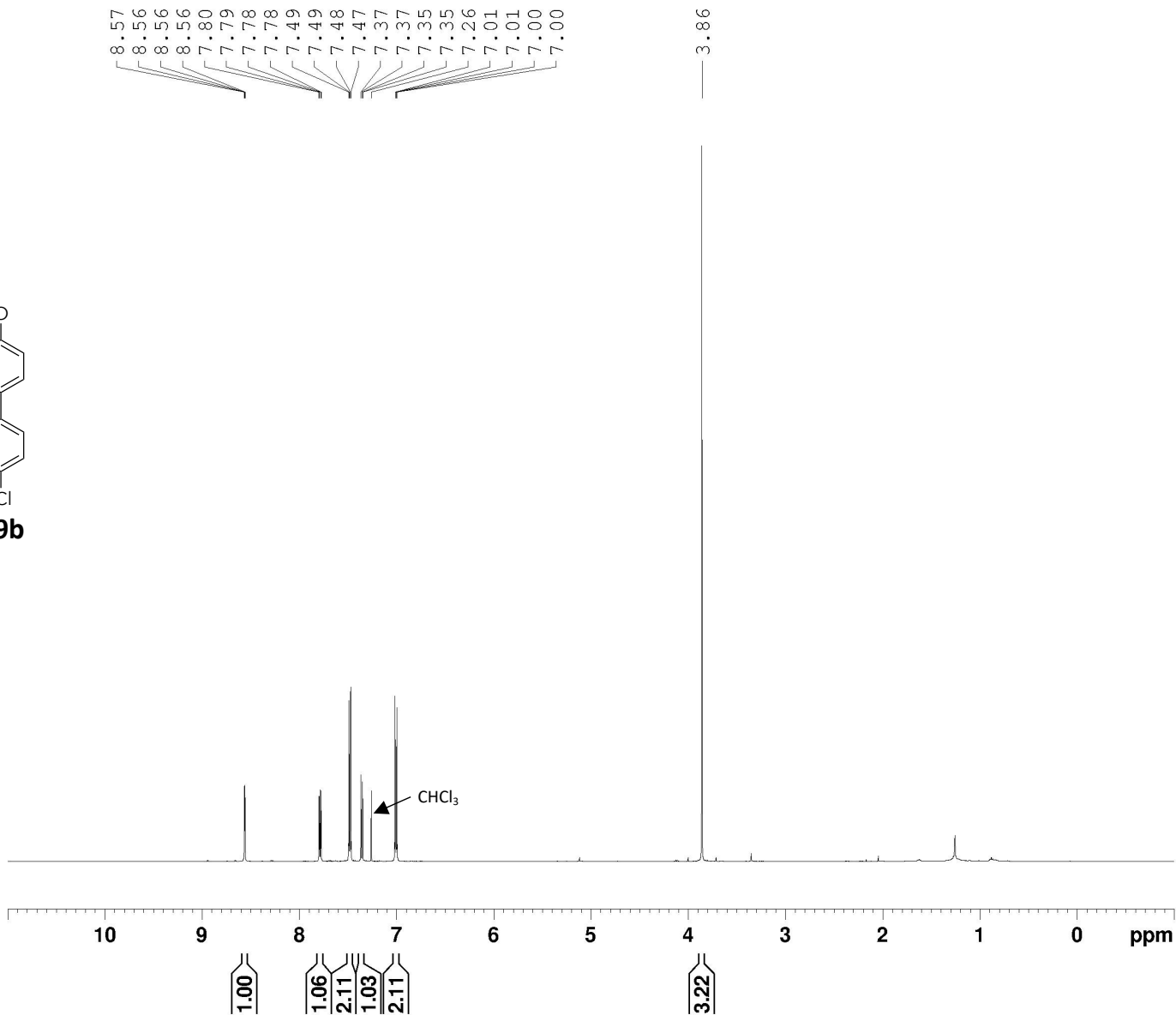
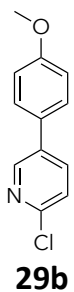
Current Data Parameters  
 NAME NL-1-117  
 EXPNO 14  
 PROCNO 1

F2 - Acquisition Parameters

Date\_ 20211127  
 Time 14.39 h  
 INSTRUM Avance Neo  
 PROBHD z152088\_0031 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 23809.523 Hz  
 FIDRES 0.726609 Hz  
 AQ 1.3762560 sec  
 RG 8.125  
 DW 21.000 usec  
 DE 6.50 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 100.6228298 MHz  
 NUC1 13C  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 86.55400085 W  
 SFO2 400.1316005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz65  
 PCPD2 90.00 usec  
 PLW2 24.03499985 W  
 PLW12 0.18990999 W  
 PLW13 0.09552100 W

F2 - Processing parameters

SI 32768  
 SF 100.6127505 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

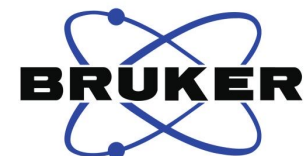
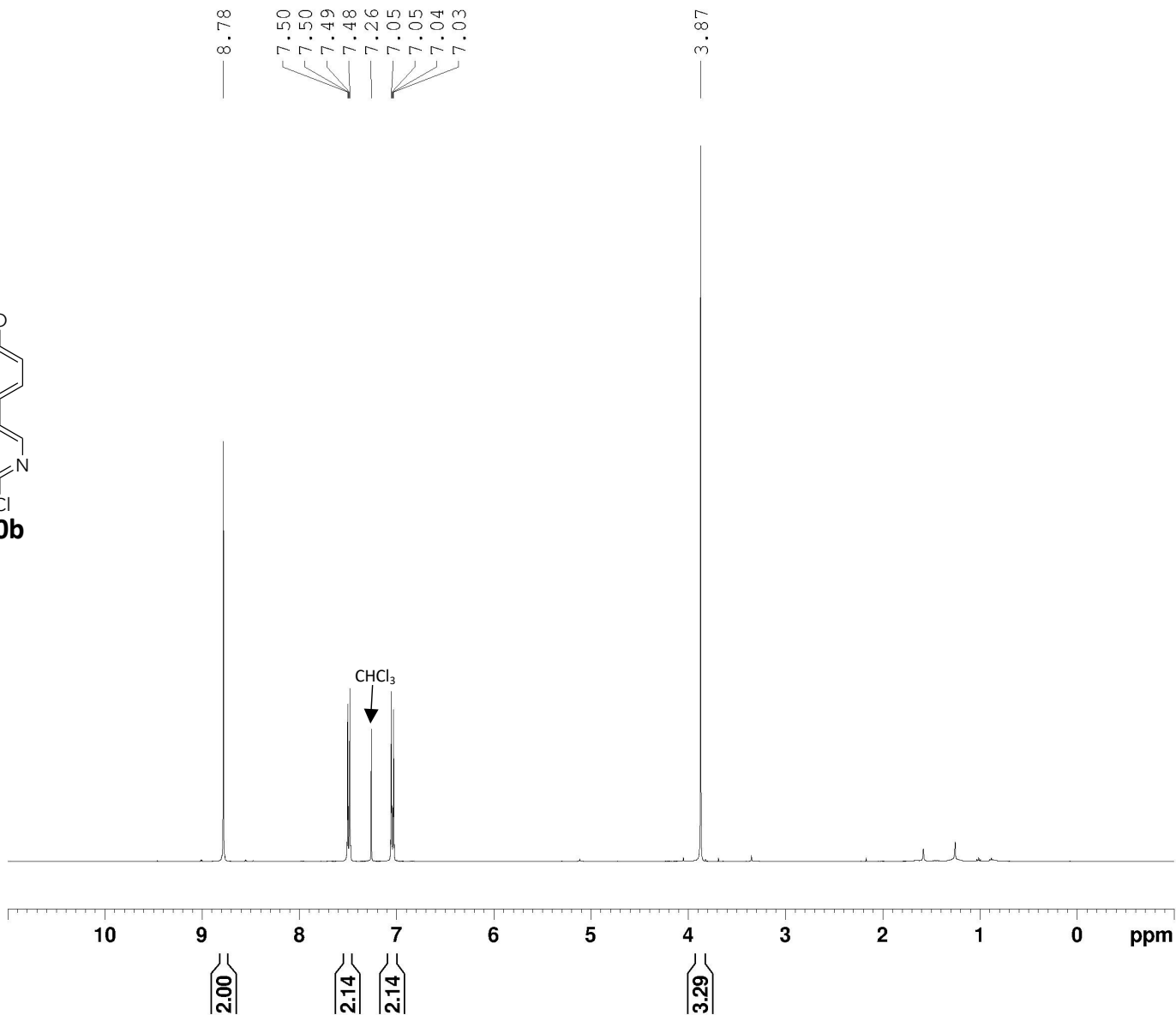
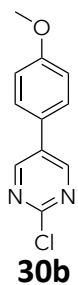


Current Data Parameters  
 NAME NL-1-189-1-C5  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20220317  
 Time 11.49 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 94.51  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

F2 - Processing parameters  
 SI 65536  
 SF 500.2300120 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

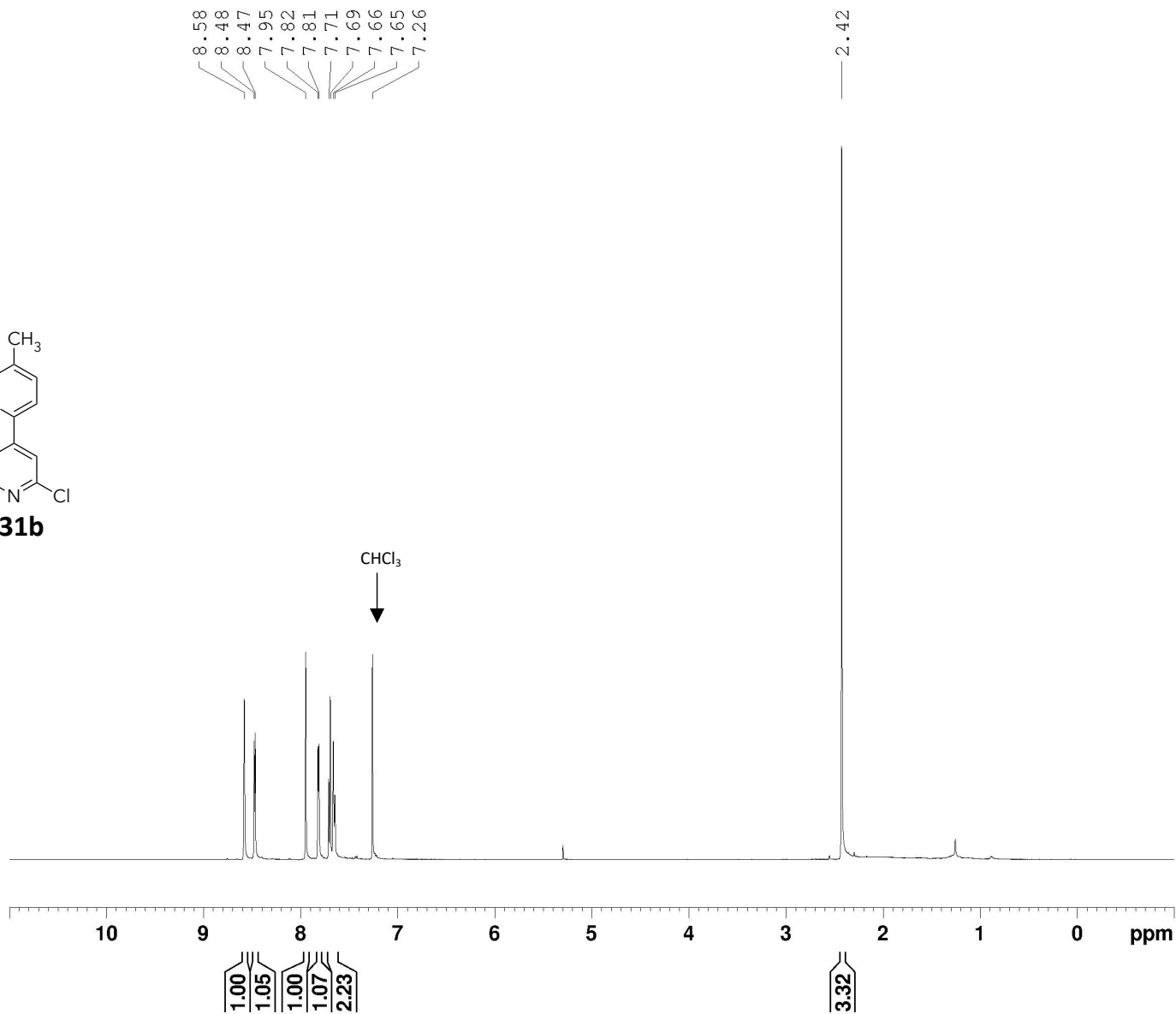
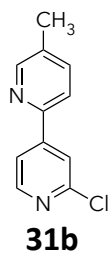




Current Data Parameters  
 NAME NL-1-190-2-C5  
 EXPNO 14  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20220319  
 Time 12.27 h  
 INSTRUM Avance Neo  
 PROBHD z152088\_0031 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 8196.722 Hz  
 FIDRES 0.250144 Hz  
 AQ 3.9976959 sec  
 RG 101  
 DW 61.000 usec  
 DE 13.89 usec  
 TE 298.0 K  
 D1 1.00000000 sec  
 TD0 1  
 SFO1 400.1324708 MHz  
 NUC1 1H  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 24.03499985 W

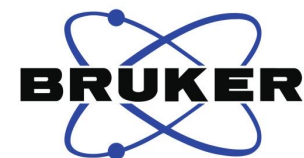
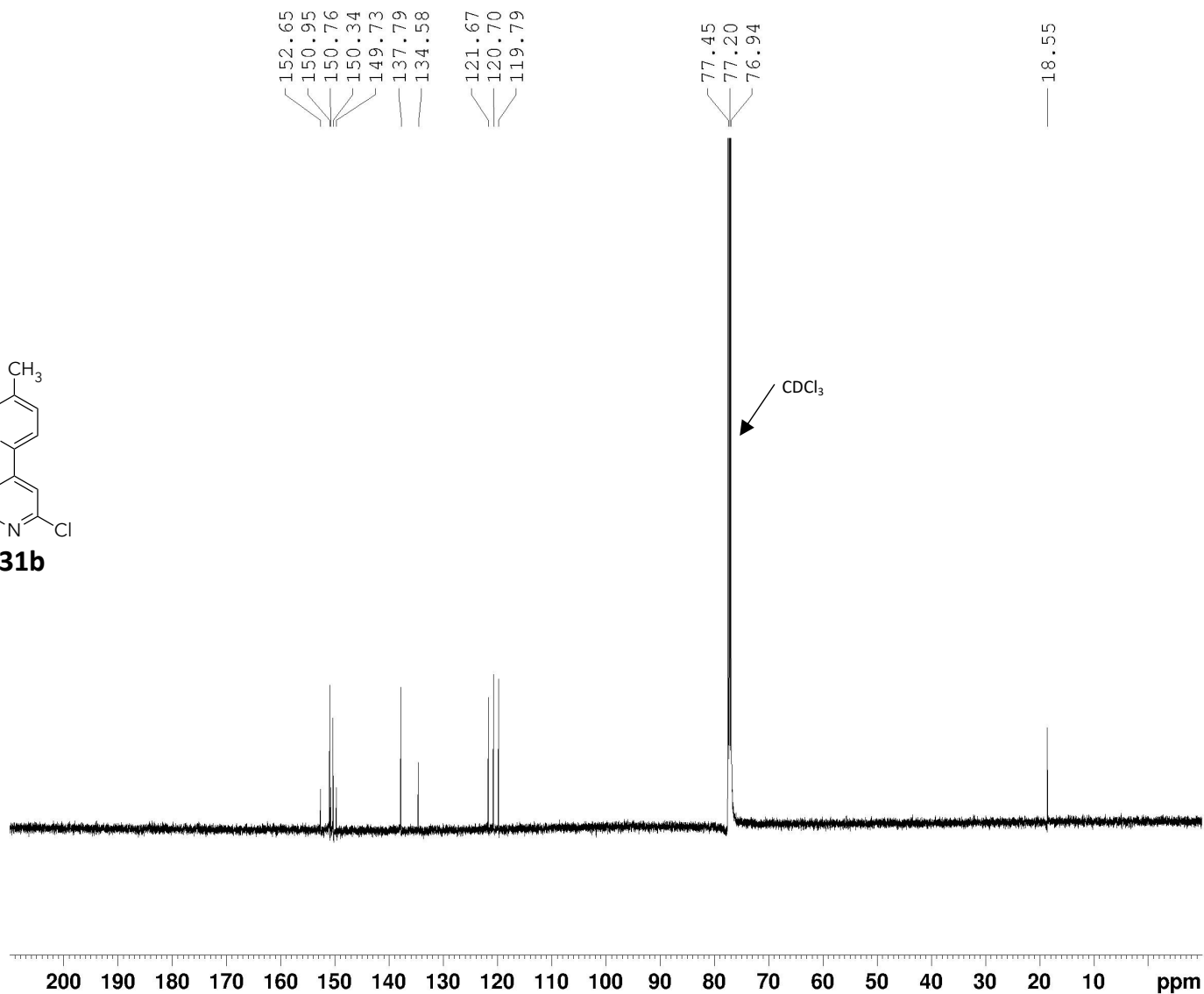
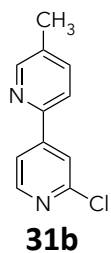
F2 - Processing parameters  
 SI 65536  
 SF 400.1300096 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
 NAME NL-1-6-2d  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210701  
 Time 17.05 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 137.36  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

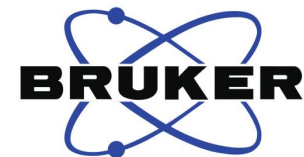
F2 - Processing parameters  
 SI 65536  
 SF 500.2300121 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
 NAME NL-1-6-13Cx2  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211028  
 Time 23.54 h  
 INSTRUM spect  
 PROBHD z125869\_0055 ( )  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

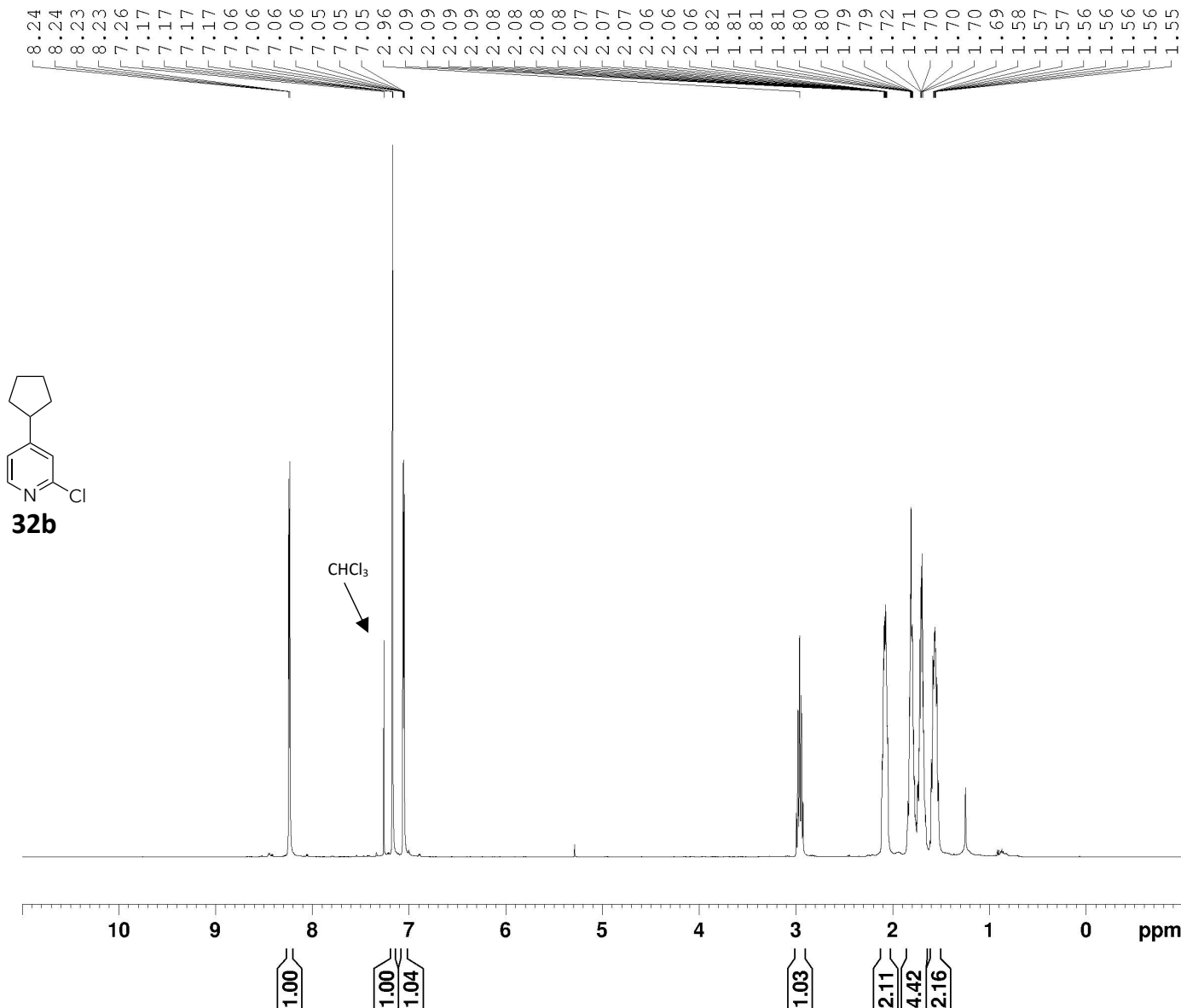
F2 - Processing parameters  
 SI 32768  
 SF 125.7829104 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

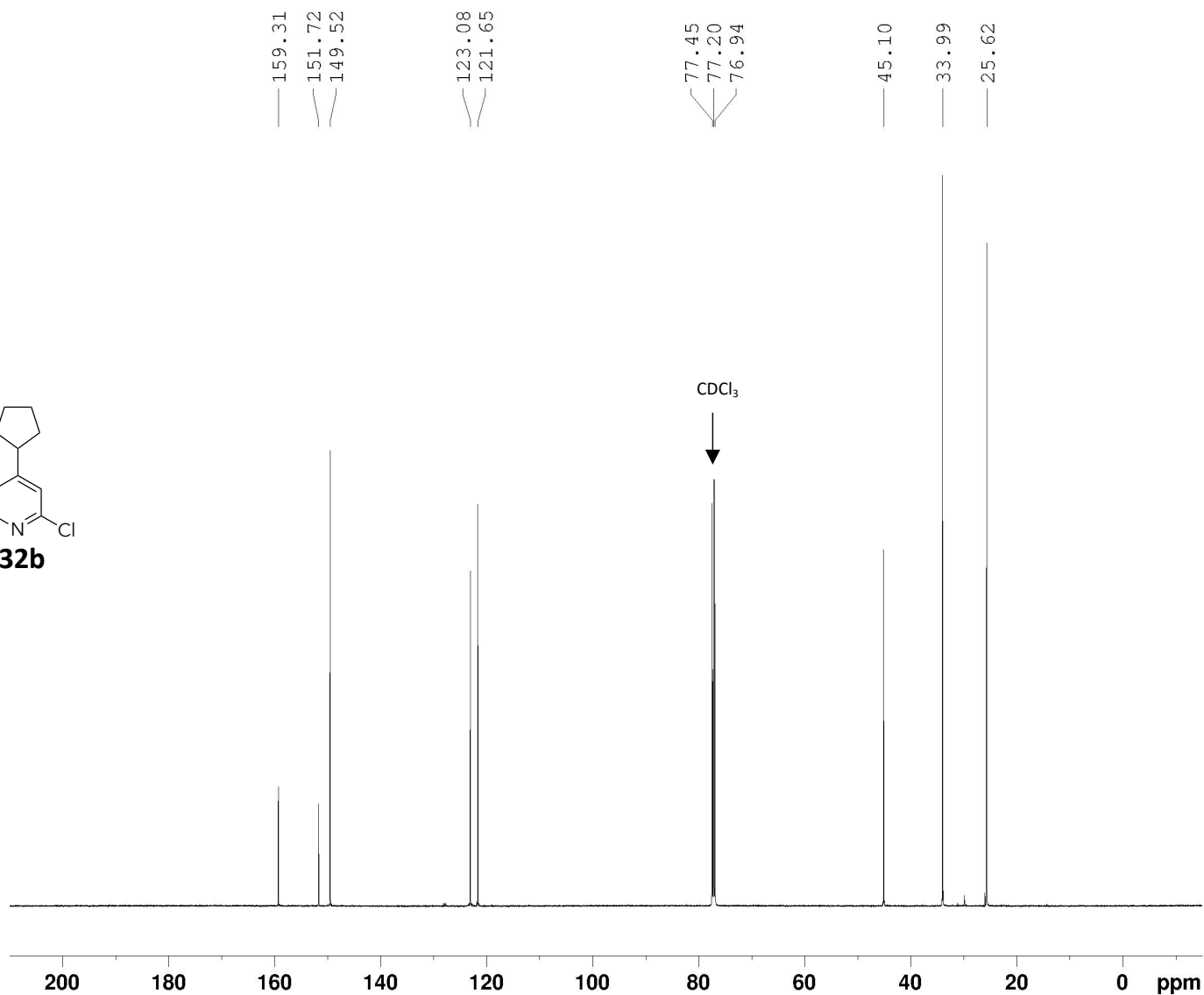
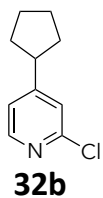


Current Data Parameters  
NAME NL-1-73-C4-1H  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20210621  
Time 13.29 h  
INSTRUM spect  
PROBHD z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 37.93  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300123 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 5.00



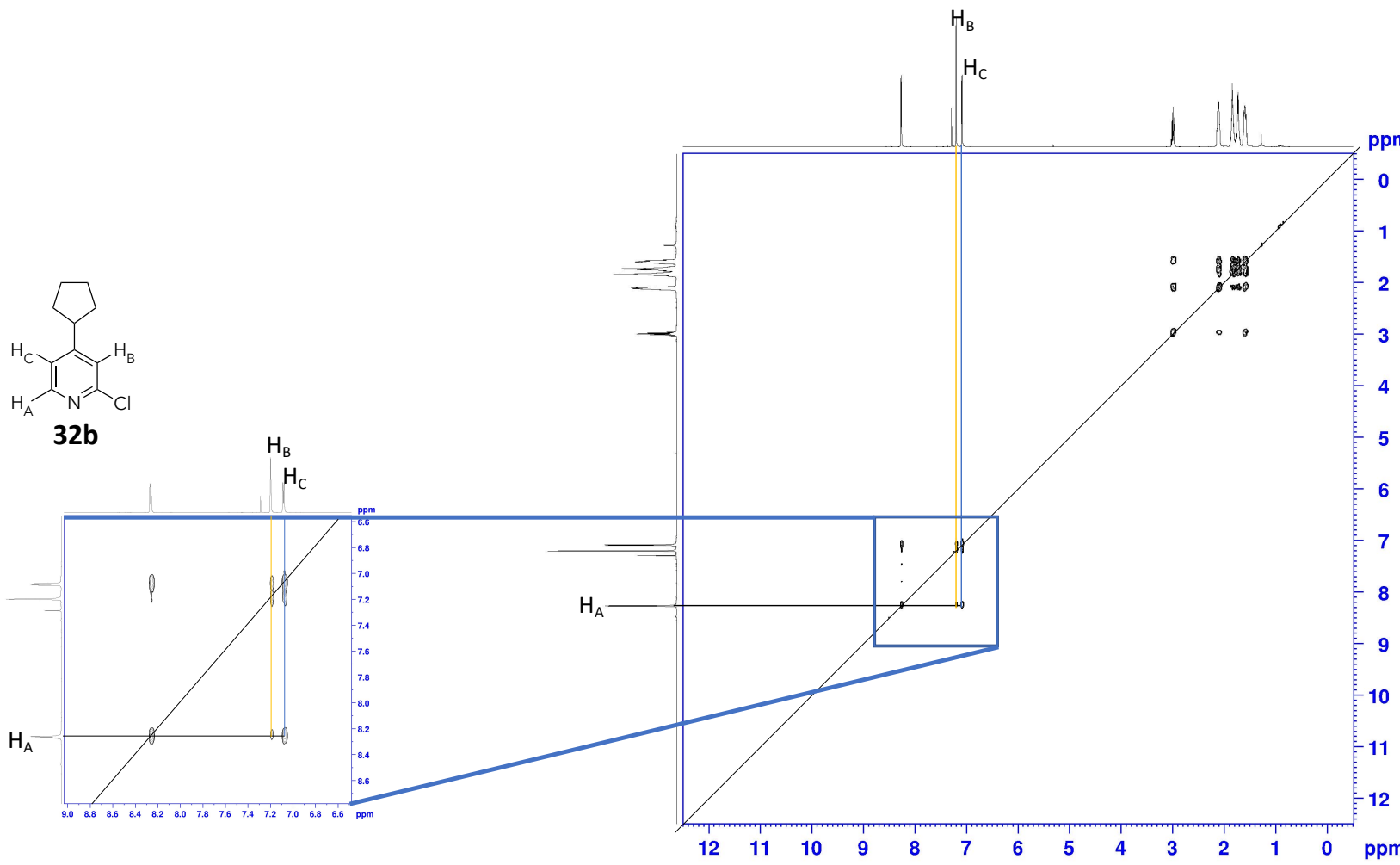
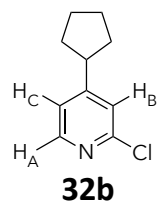


Current Data Parameters  
 NAME NL-1-73-2d  
 EXPNO 20  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210701  
 Time 12.39 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

F2 - Processing parameters  
 SI 32768  
 SF 125.7829141 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

COSY



Current Data Parameters

NAME	ML-1-73-2d
EXPNO	21
PROCNO	1

F2 - Acquisition Parameters

Date_	20210701
Time	12.40 h
INSTRUM	spect
PROBHD	Z125869.0055 f
PULPROG	cosygpmfppgf
TD	4096
SOLVENT	CDCl3
NS	4
DS	16
SWH	6510.417 Hz
FIDRES	3.178914 Hz
AQ	0.3145728 sec
RG	47.3
DW	76.000 usec
DE	16.00 usec
TE	300.0 K
DO	0.00000300 sec
D1	1.12589896 sec
D11	0.03000000 sec
D12	0.00002000 sec
D13	0.00000400 sec
D16	0.00020000 sec
IN0	0.00015380 sec
TDav	1
SFO1	500.2330014 MHz
NUC1	1H
P1	12.00 usec
P17	2500.00 usec
PLW1	11.44699935 W
PLW10	1.83150005 W
GPAM[1]	SMSQ10.100
GPZ1	16.00 %
GPAM[2]	SMSQ10.100
GPZ2	12.00 %
GPAM[3]	SMSQ10.100
GPZ3	40.00 %
P16	1000.00 usec

F1 - Acquisition parameters

TD	256
SFO1	500.233 MHz
FIDRES	50.796490 Hz
SW	12.998 ppm
FMODE	QE

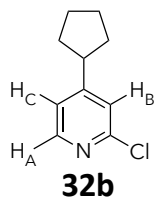
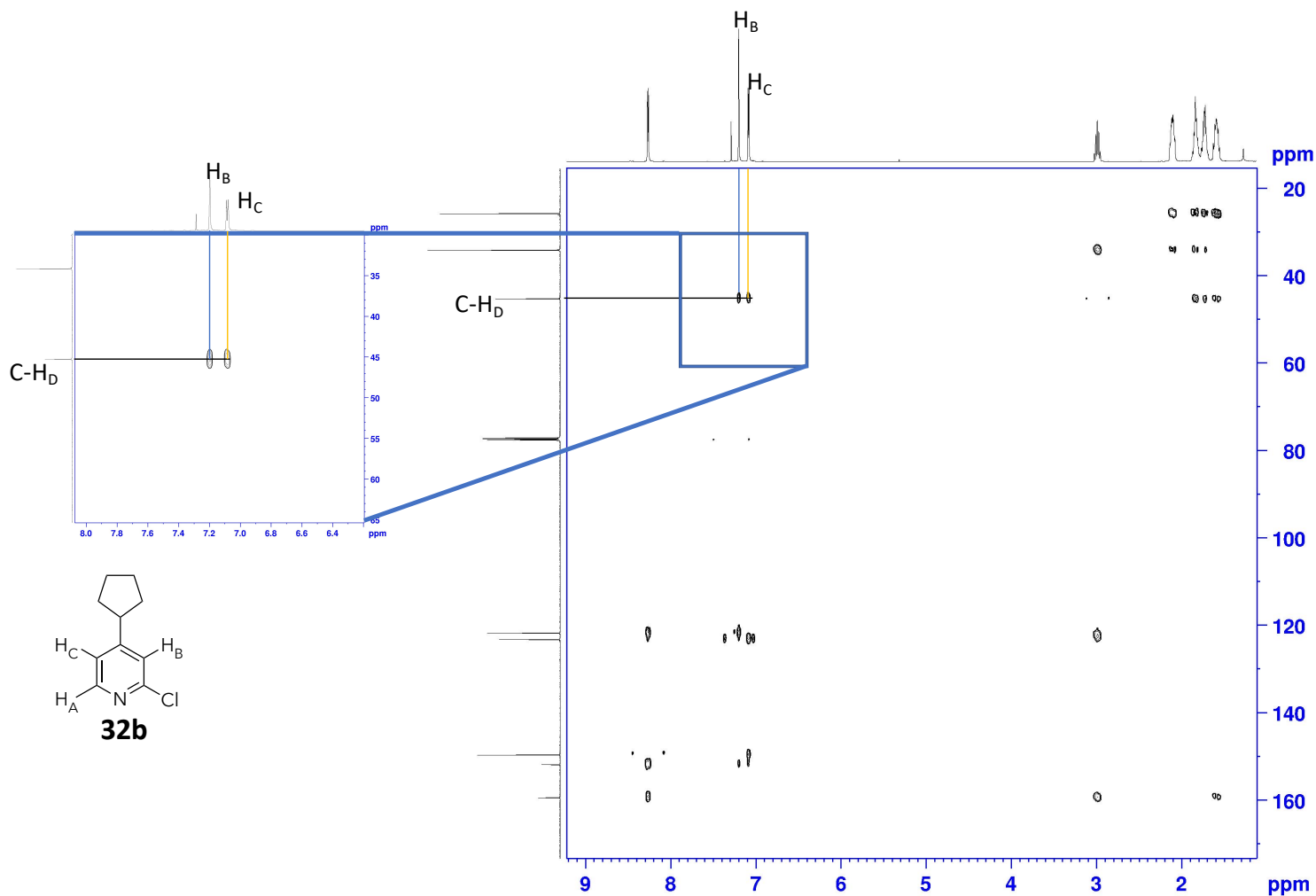
F2 - Processing parameters

SI	2048
SF	500.2300032 MHz
WDW	QSTINE
SSB	0
LB	0 Hz
GB	0
PC	1.40

F1 - Processing parameters

SI	2048
MC2	QE
SF	500.2300032 MHz
WDW	QSTINE
SSB	0
LB	0 Hz
GB	0

# HMBC



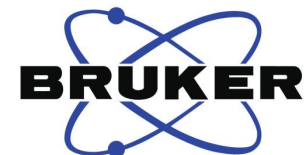
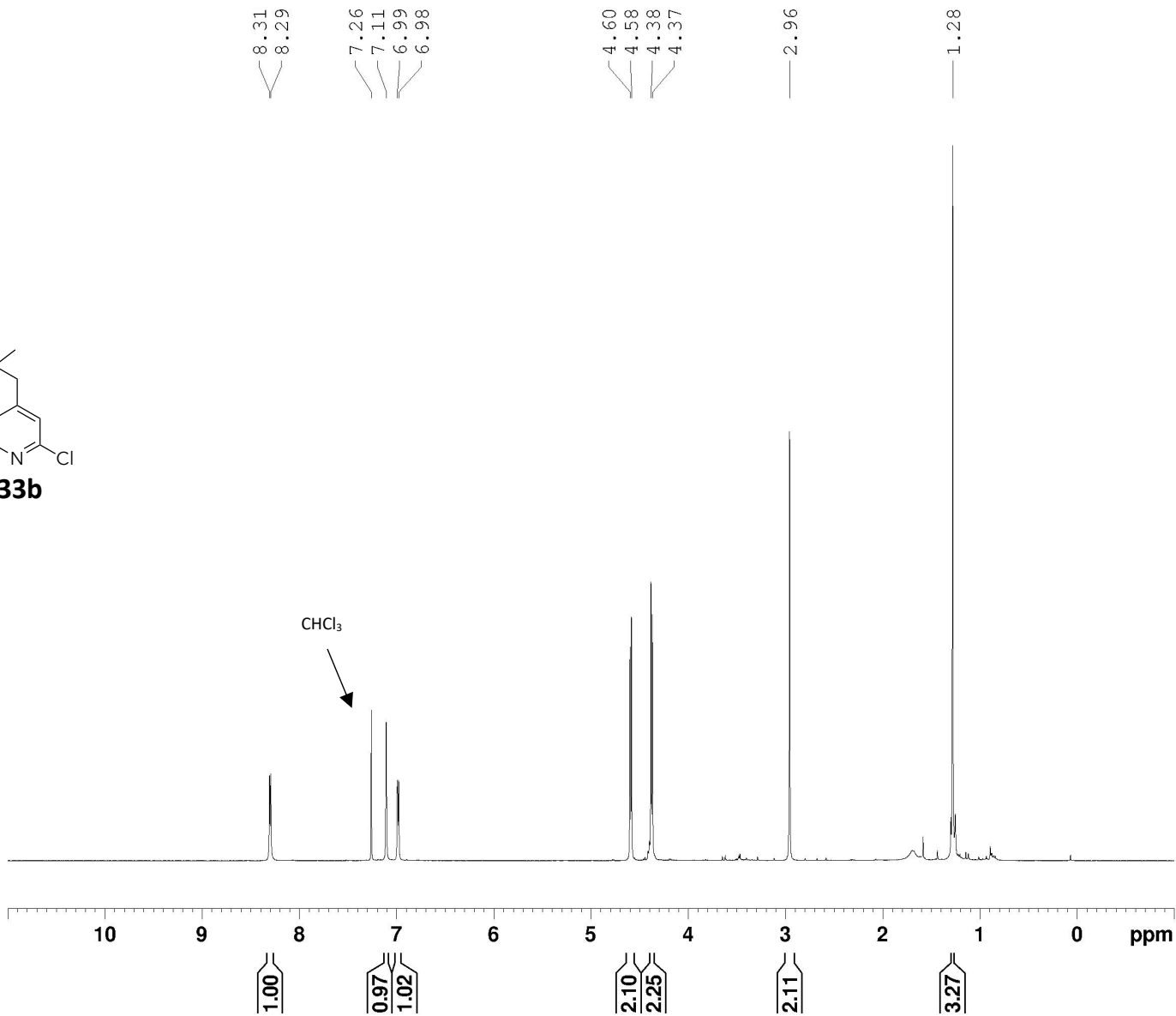
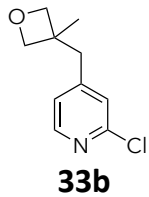
Current Data Parameters  
 NAME NL-1-73-2d  
 EXPNO 23  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210701  
 Time 13.09 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 ( )  
 PULPROG hmbcgp1pndqf  
 TD 2048  
 SOLVENT CDCl3  
 NS 4  
 DS 16  
 SWH 4716.981 Hz  
 FIDRES 4.606427 Hz  
 AQ 0.2170880 sec  
 RG 190.44  
 DW 106.000 usec  
 DE 16.00 usec  
 TE 300.0 K  
 CNST2 145.0000000  
 CNST13 10.0000000  
 D0 0.00000300 sec  
 D1 1.50000000 sec  
 D2 0.00344828 sec  
 D6 0.00000000 sec  
 D16 0.00020000 sec  
 IN0 0.00001810 sec  
 Tdav 1  
 SFO1 500.2322599 MHz  
 NUC1 1H  
 P1 12.00 usec  
 P2 24.00 usec  
 FLW1 11.44699935 W  
 SFO2 125.7955118 MHz  
 NUC2 13C  
 P3 10.00 usec  
 FLW2 56.90299988 W  
 GPNAM[1] SMSQ10.100  
 GPZ1 50.00 %  
 GPNAM[2] SMSQ10.100  
 GPZ2 50.00 %  
 GPNAM[3] SMSQ10.100  
 GPZ3 40.10 %  
 P16 1000.00 usec

F1 - Acquisition parameters  
 TD 128  
 SFO1 125.7955 MHz  
 FIDRES 431.629822 Hz  
 SW 219.897 ppm  
 FhMODE QF

F2 - Processing parameters  
 SI 2048  
 SF 500.2299980 MHz  
 WDW SINE  
 SSB 0  
 LB 0 Hz  
 GB 0  
 PC 1.40

F1 - Processing parameters  
 SI 1024  
 MC2 QF  
 SF 125.7828890 MHz  
 WDW SINE  
 SSB 0  
 LB 0 Hz  
 GB 0

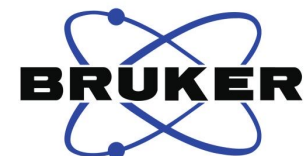
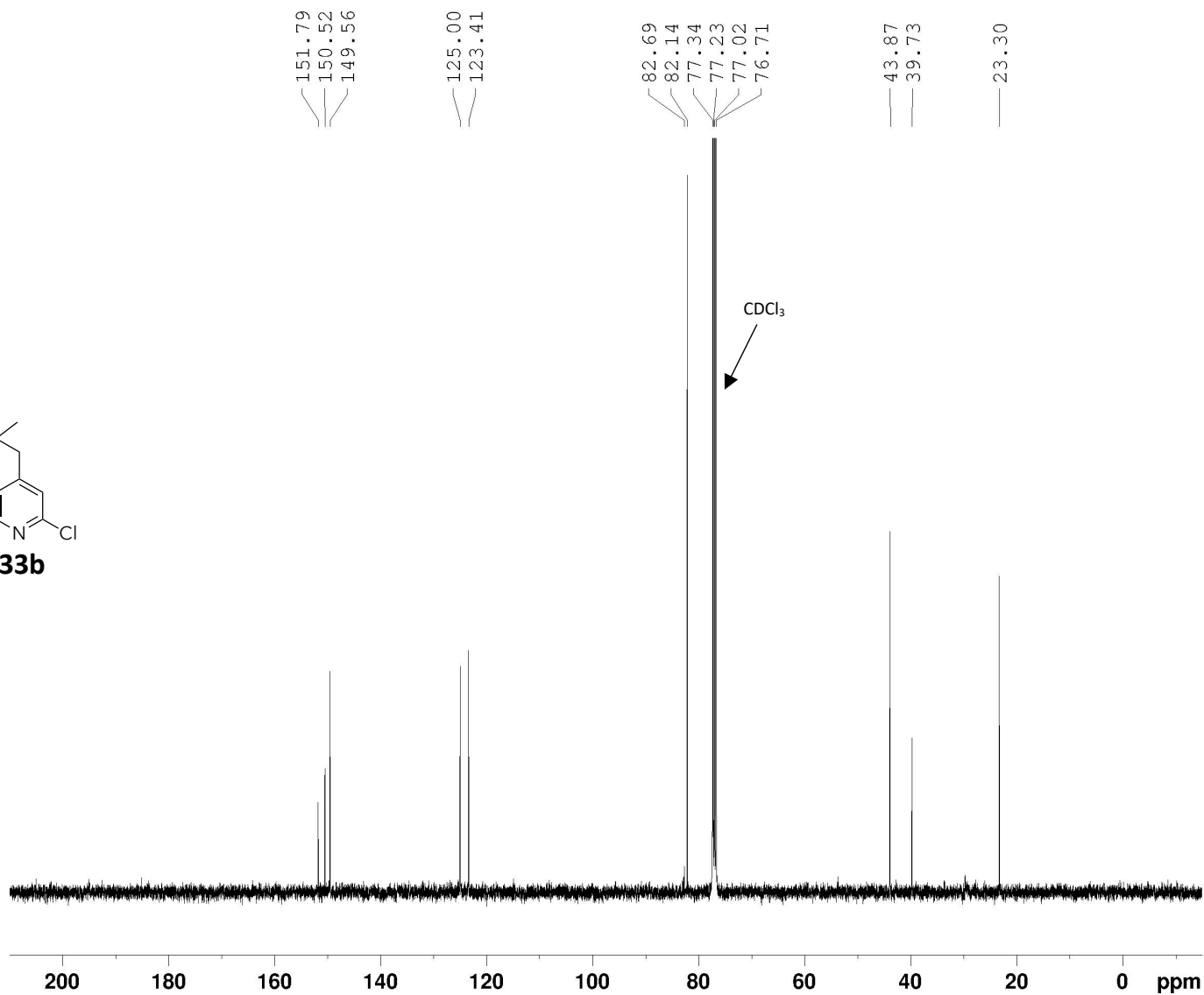
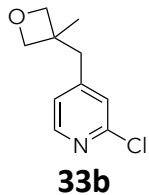


Current Data Parameters  
 NAME NL-1-89-inccnc  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210729  
 Time 18.10 h  
 INSTRUM Avance Neo  
 PROBHD Z152088\_0031 (   
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 16  
 DS 2  
 SWH 8196.722 Hz  
 FIDRES 0.250144 Hz  
 AQ 3.9976959 sec  
 RG 101  
 DW 61.000 usec  
 DE 13.89 usec  
 TE 298.1 K  
 D1 1.00000000 sec  
 TD0 1  
 SFO1 400.1324708 MHz  
 NUC1 1H  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 24.03499985 W

F2 - Processing parameters  
 SI 65536  
 SF 400.1300099 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



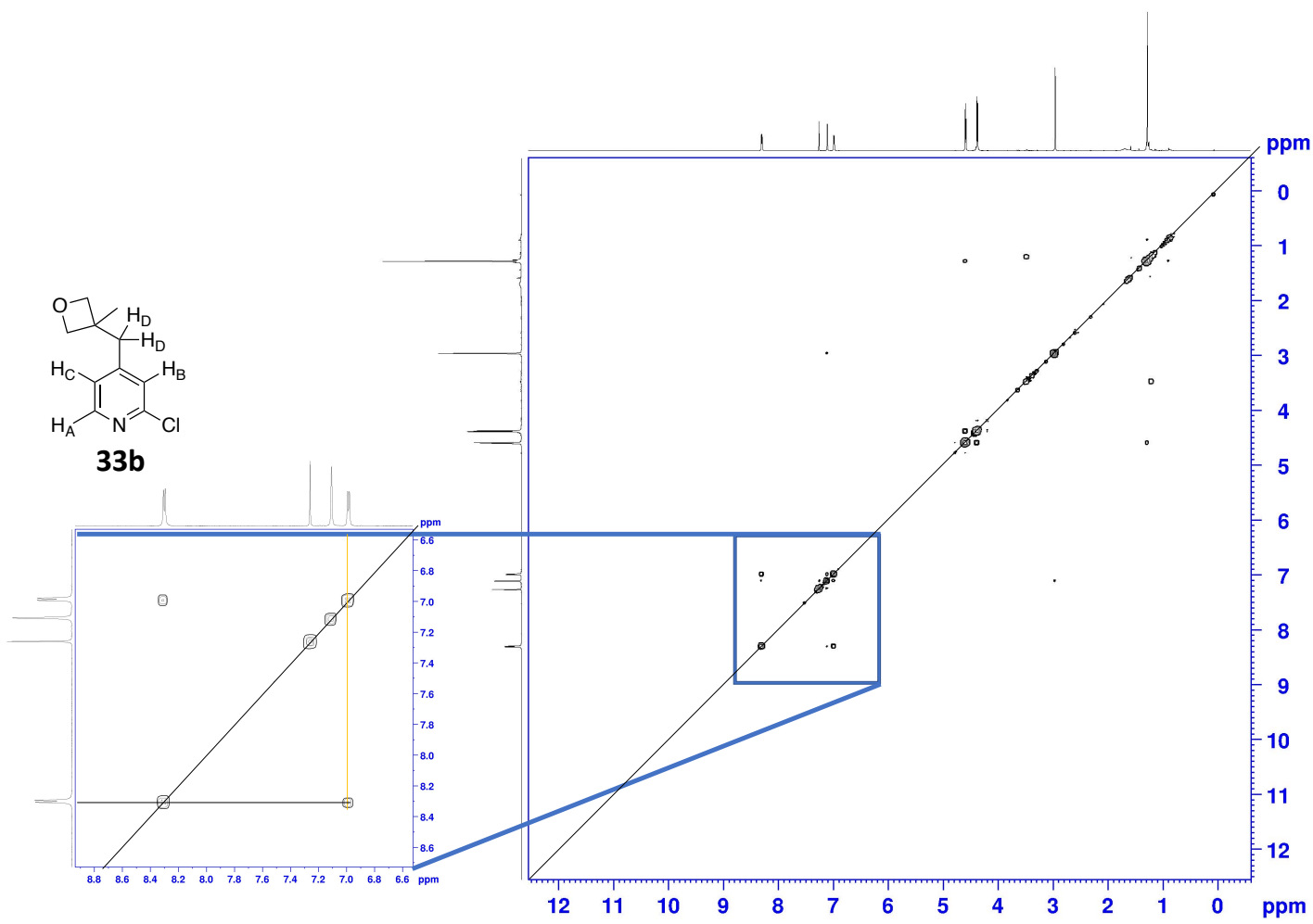
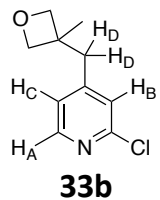


Current Data Parameters  
 NAME NL-1-89-inconc  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210729  
 Time 19.12 h  
 INSTRUM Avance Neo  
 PROBHD z152088\_0031 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 23809.523 Hz  
 FIDRES 0.726609 Hz  
 AQ 1.3762560 sec  
 RG 8.125  
 DW 21.000 usec  
 DE 6.50 usec  
 TE 298.1 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 100.6228298 MHz  
 NUC1 13C  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 86.55400085 W  
 SFO2 400.1316005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz65  
 PCPD2 90.00 usec  
 PLW2 24.03499985 W  
 PLW12 0.18990999 W  
 PLW13 0.09552100 W

F2 - Processing parameters  
 SI 32768  
 SF 100.6127685 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

COSY



```

Current Data Parameters
NAME      NL-1-89d-1H
EXPNO    13
PROCNO   1

F2 - Acquisition Parameters
Date_    20210727
Time     19.20 h
INSTRUM  Avance Neo
PROBHD   Z152088_0031 f
PULPROG  cosypppppf
TD       2048
SOLVENT  CDCl3
NS       8
DS       16
SWH      5263.158 Hz
FIDRES   5.139802 Hz
AQ       0.1945600 sec
RG       101
DW       95.000 usec
DE       8.50 usec
TE       298.1 K
D0       0.00000300 sec
D1       2.00000000 sec
D11      0.03000000 sec
D12      0.00002000 sec
D13      0.00000400 sec
D16      0.00020000 sec
IN0      0.00019000 sec
TD0      1
SFO1     400.1324008 MHz
NUC1     1H
P0       8.00 usec
P1       8.00 usec
P17      2500.00 usec
PLW1     24.03499985 W
PLW10    1.70920002 W
GPM1[1]  SMSq10.100
GP21     10.00 %
P16      1000.00 usec

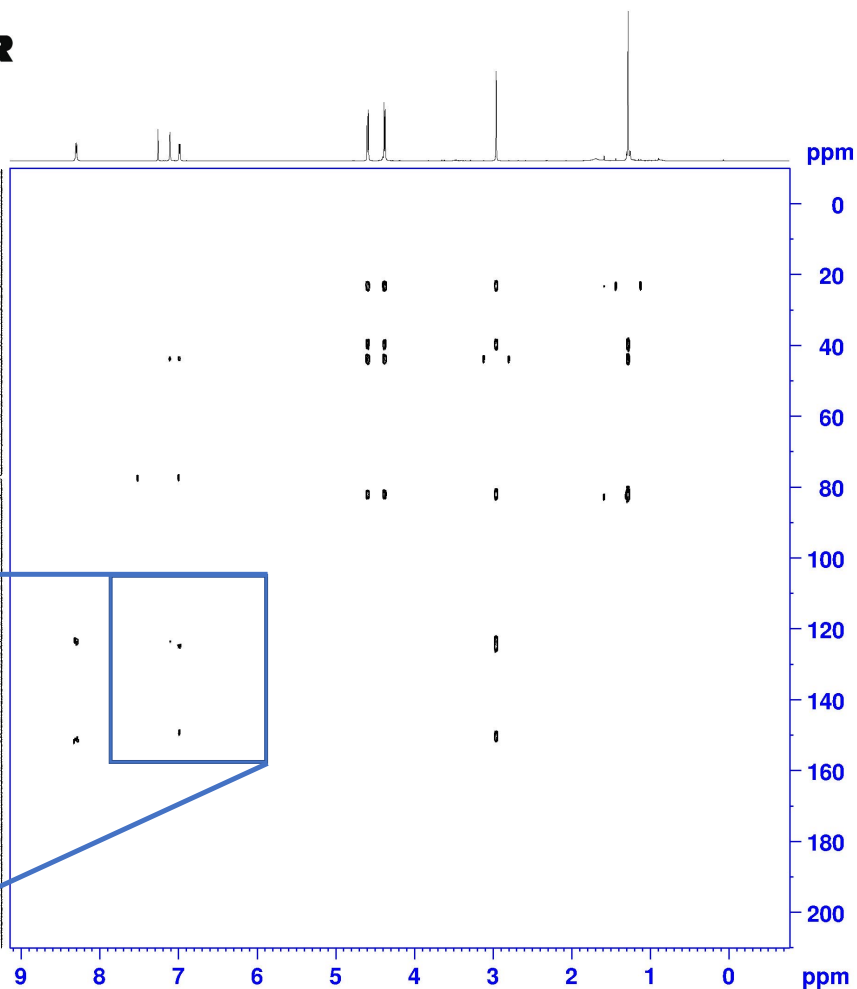
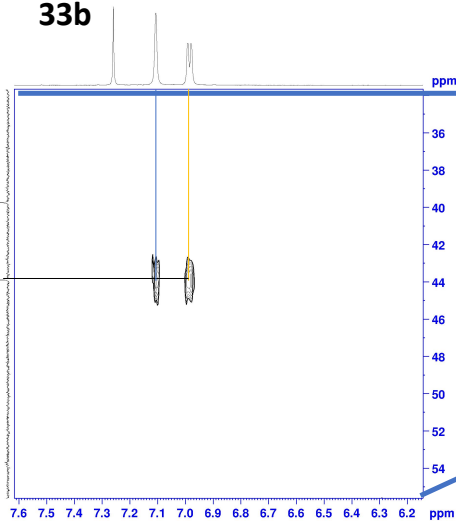
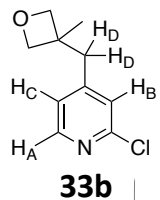
----- F1 INDIRECT DIMENSION -----
td1      128
sw_F1    13.153541

F1 - Acquisition parameters
TD       128
SFO1     400.1324 MHz
FIDRES   62.236839 Hz
SW       13.154 ppm
F1MODE   QF

F2 - Processing parameters
SI       1024
SF       400.1300096 MHz
WDW      QSINE
SSB      0 Hz
LB       0 Hz
GB       0
PC       1.40

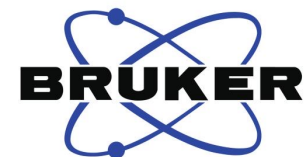
F1 - Processing parameters
SI       1024
MC2      QF
SF       400.1300099 MHz
WDW      QSINE
SSB      0 Hz
LB       0 Hz
GB       0
    
```

# HMBC



```

Current Data Parameters
NAME      NL-1-89-inconc
F2 - Acquisition Parameters
INSTRUM   Avance Neo
PROBHD    Z152088_0031 (
PULPROG   hmbcgp1pndqf
TD         4096
SOLVENT   CDCl3
NS         4
DS         16
SWH        3968.254 Hz
FIDRES     1.937624 Hz
AQ         0.5160960 sec
RG         101
DW         126.000 usec
DE         6.50 usec
TE         298.2 K
CNST2     145.000000
CNST13    10.000000
D0         0.00000300 sec
D1         1.50000000 sec
D2         0.00344828 sec
D6         0.05000000 sec
D16        0.00020000 sec
IN0        0.00002259 sec
TDav       1
SFO1       400.1316847 MHz
NUC1        1H
P1          8.00 usec
P2          16.00 usec
PLW1       24.03499985 W
SFO2       100.6228298 MHz
NUC2        13C
P3          8.00 usec
PLW2       86.55400085 W
GPNAM[1]   SMSQ10.100
GPZ1        50.00 %
GPNAM[2]   SMSQ10.100
GPZ2        30.00 %
GPNAM[3]   SMSQ10.100
GPZ3        40.10 %
P16        1000.00 usec
===== F1 INDIRECT DIMENSION :
td1         128
sw_F1       220.000000
F1 - Acquisition parameters
TD          128
SFO1        100.6228 MHz
FIDRES      345.877136 Hz
SW          219.991 ppm
FnMODE      QF
    
```



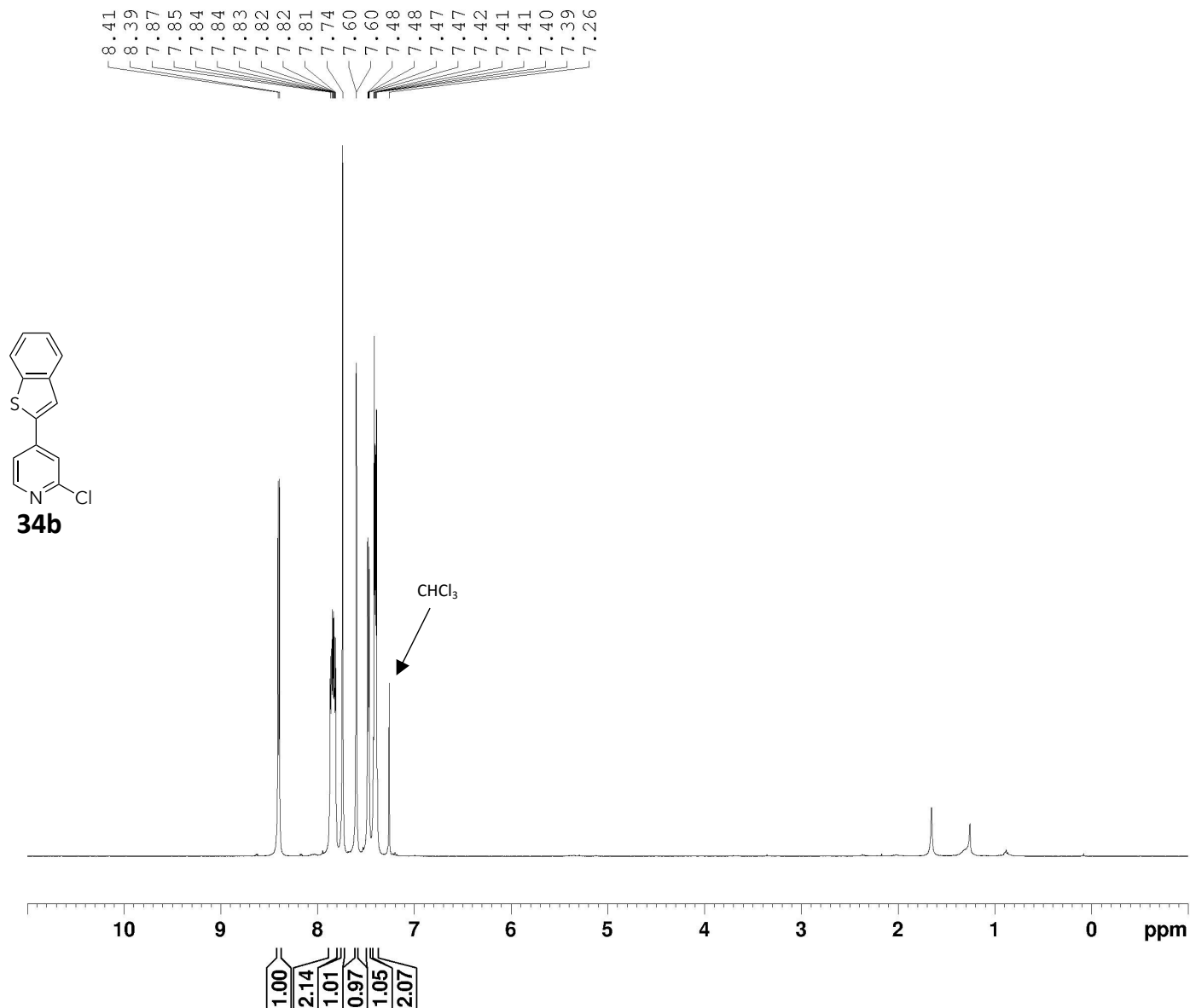
Current Data Parameters  
NAME NL-1-38-C4+  
EXPNO 10  
PROCNO 1

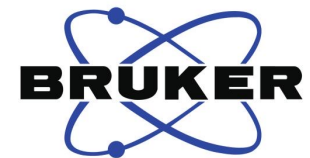
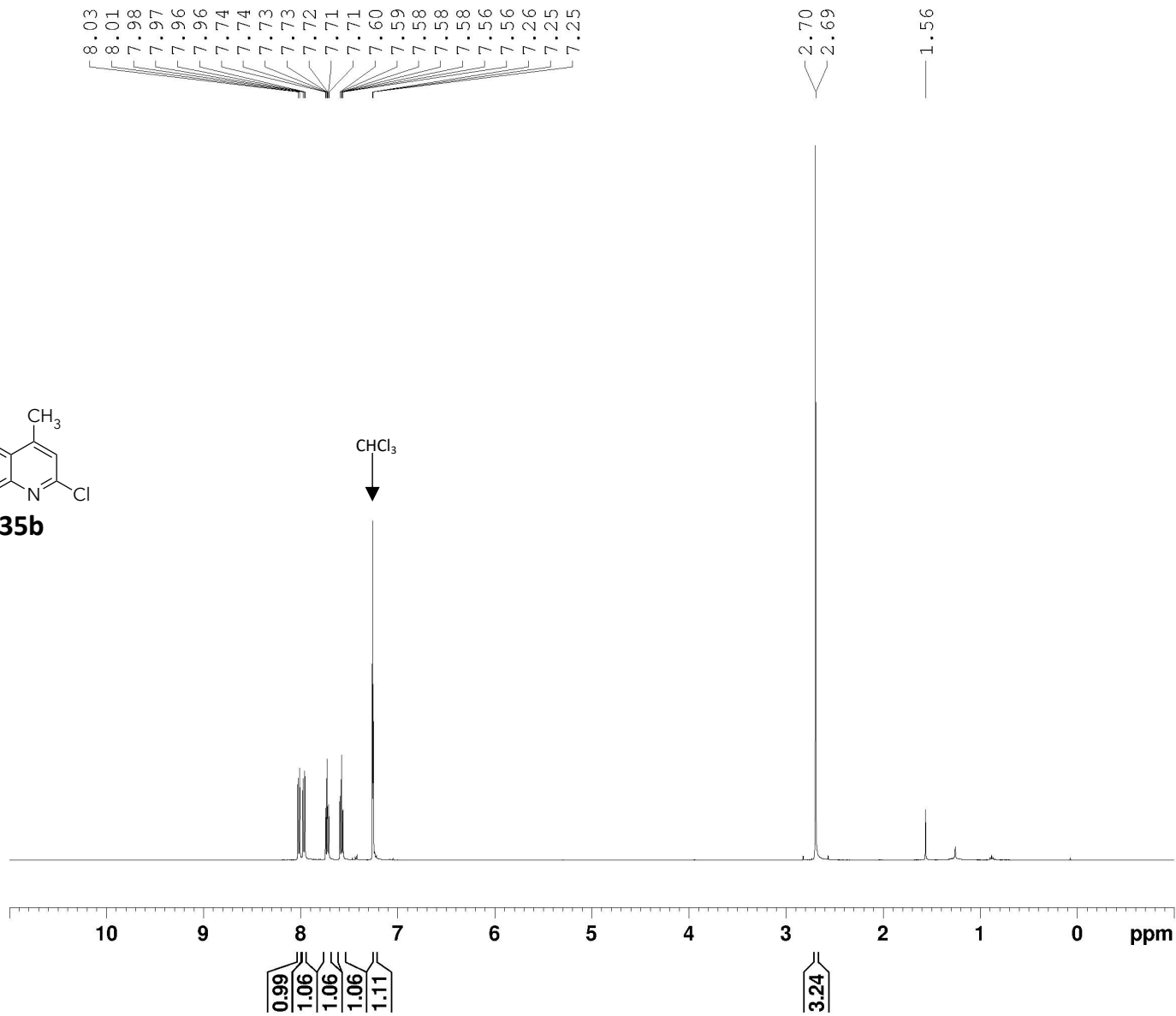
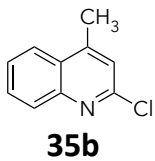
F2 - Acquisition Parameters

Date\_ 20210501  
Time 14.01 h  
INSTRUM Avance Neo  
PROBHD z152088\_0031 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 16  
DS 2  
SWH 8196.722 Hz  
FIDRES 0.250144 Hz  
AQ 3.9976959 sec  
RG 101  
DW 61.000 usec  
DE 13.89 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 400.1324708 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 24.03499985 W

F2 - Processing parameters

SI 65536  
SF 400.1300100 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 10.00

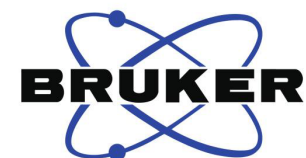
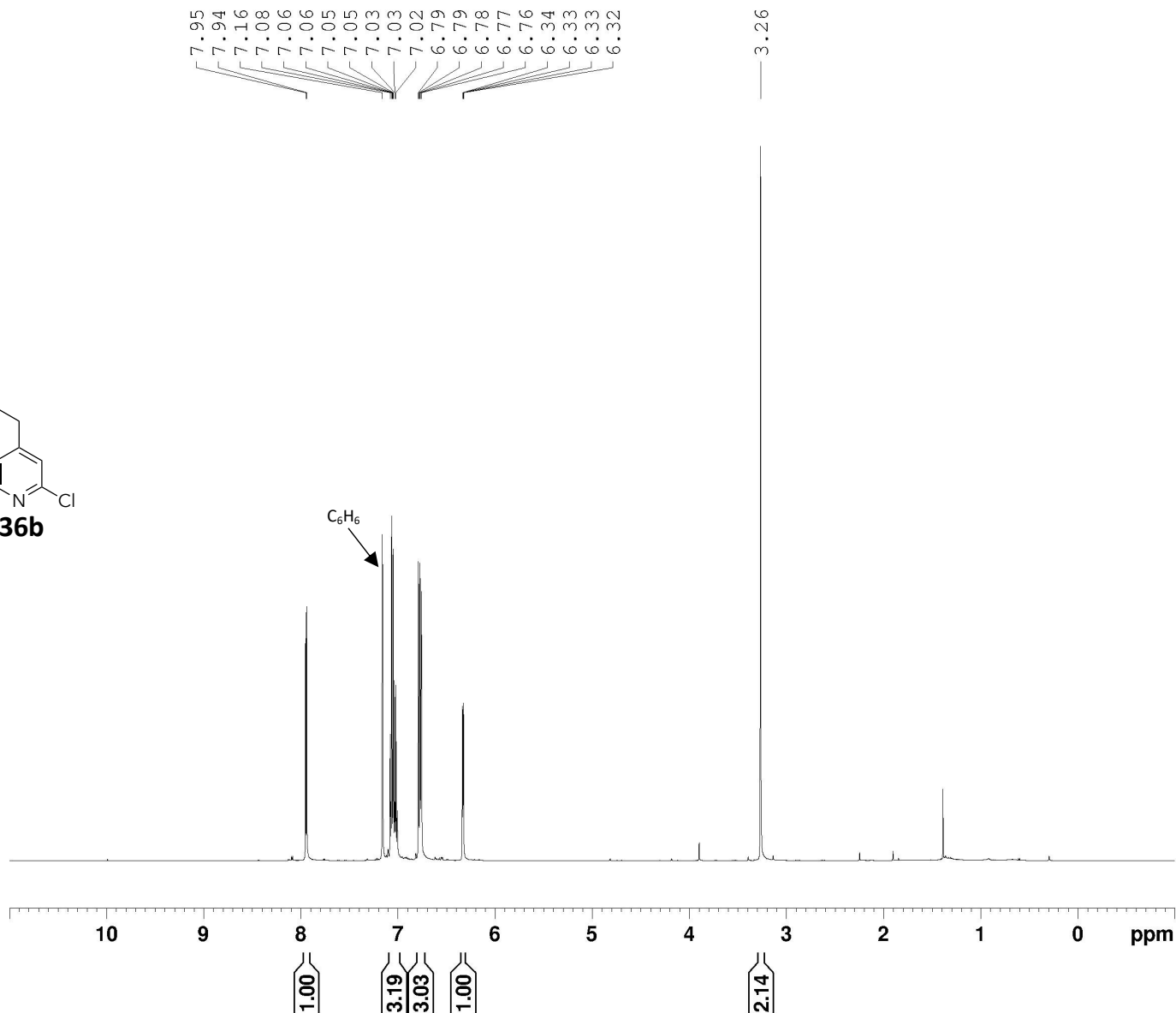
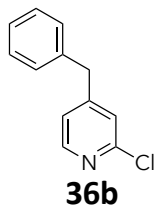




Current Data Parameters  
 NAME NL-1-75-1H  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211028  
 Time 14.50 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 151.18  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

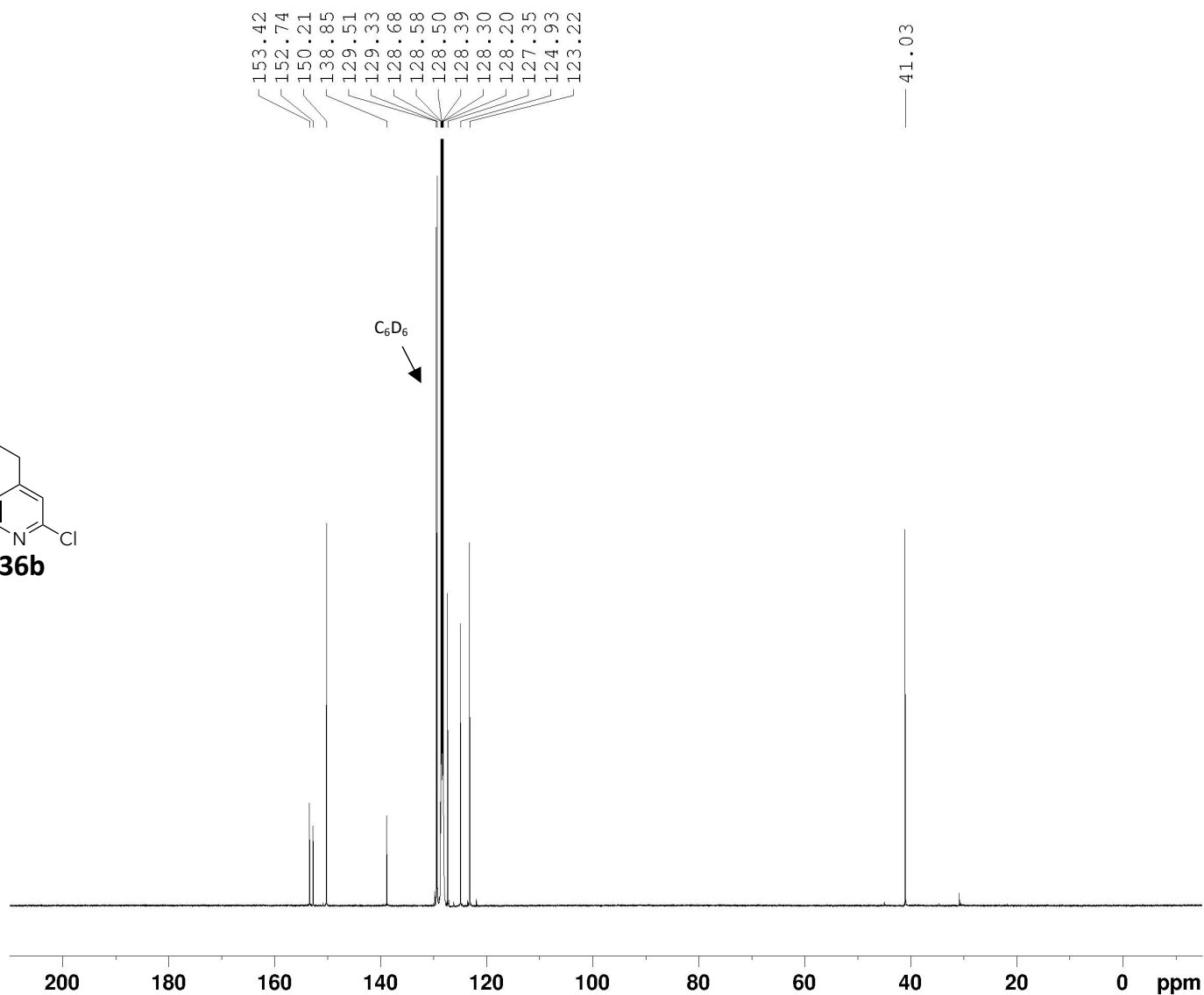
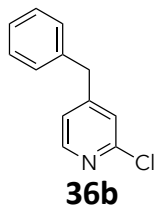
F2 - Processing parameters  
 SI 65536  
 SF 500.2300122 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
 NAME NL-1-61-C4-1H-benzene  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210624  
 Time 14.01 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (zg30)  
 TD 65536  
 SOLVENT C6D6  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 43.72  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

F2 - Processing parameters  
 SI 65536  
 SF 500.2299949 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

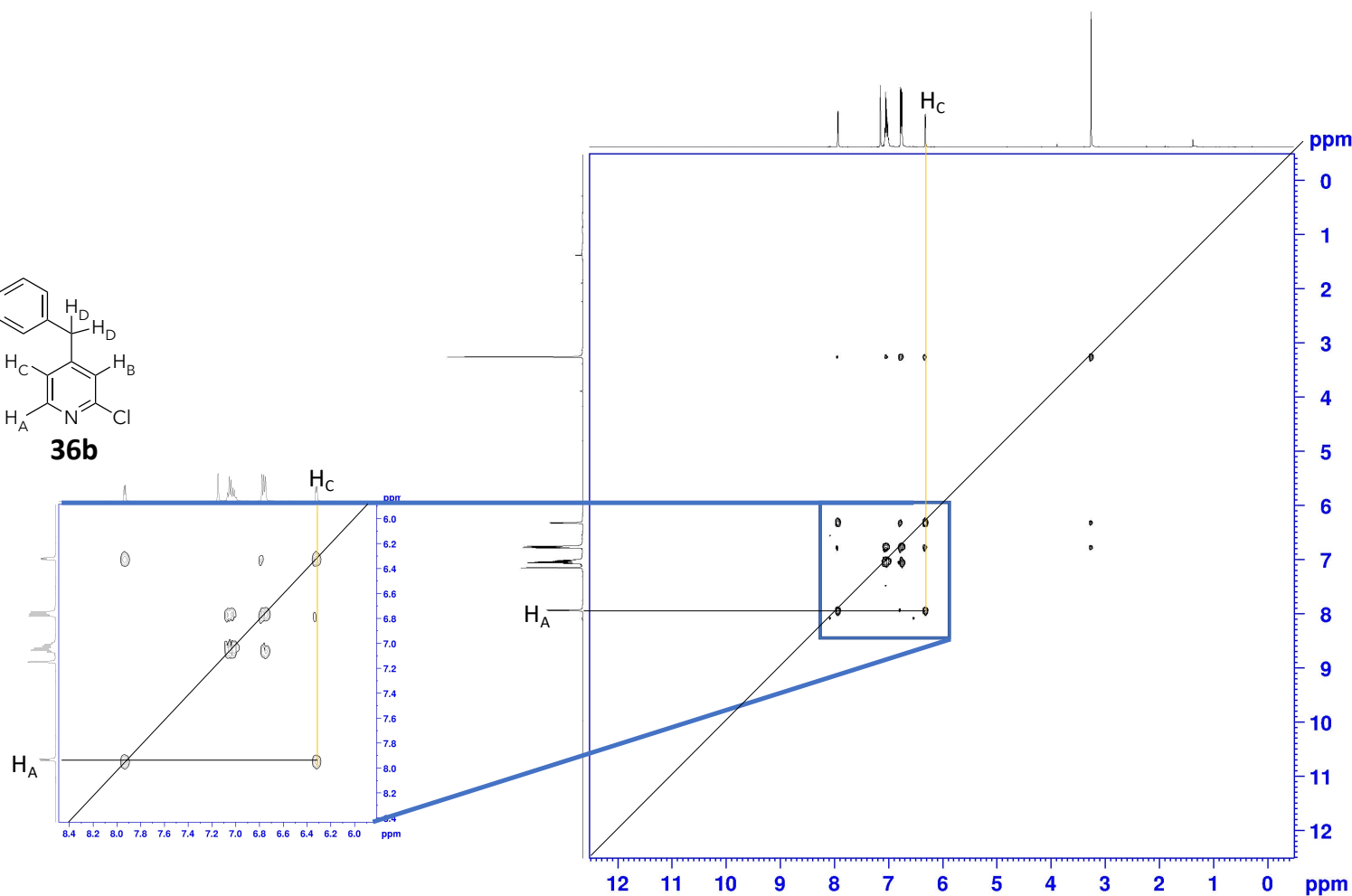
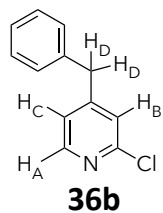


Current Data Parameters  
 NAME NL-1-61-C4-2d  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20210701  
 Time 18.21 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT C6D6  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

F2 - Processing parameters  
 SI 32768  
 SF 125.7828447 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

COSY



```

Current Data Parameters
NAME      NL-1-61-C4-2d
EXPNO    13
PROCNO   1

F2 - Acquisition Parameters
Date_    20210701
Time     18.40 h
INSTRUM  spect
PROBHD   Z125869_0033 (
PULPROG  cosygpmfppqf
TD       4096
SOLVENT  C6D6
NS       16
DS       4
SWH      6510.417 Hz
FIDRES   3.178914 Hz
AQ       0.3145728 sec
RG       53.13
DW       76.800 usec
DE       16.00 usec
TE       300.0 K
D0       0.00000300 sec
D1       1.1289896 sec
D11      0.03000000 sec
D12      0.00002000 sec
D13      0.00000400 sec
D16      0.00020000 sec
IN0      0.00015390 sec
TDav     1
SFO1     500.2330014 MHz
NUC1     1H
P1       12.00 usec
P17      2500.00 usec
PLW1     11.44699955 W
PLW10    1.83150005 W
GPNAM[1] SMSQ10.100
GPZ1     16.00 %
GPNAM[2] SMSQ10.100
GPZ2     12.00 %
GPNAM[3] SMSQ10.100
GPZ3     40.00 %
P16      1000.00 usec

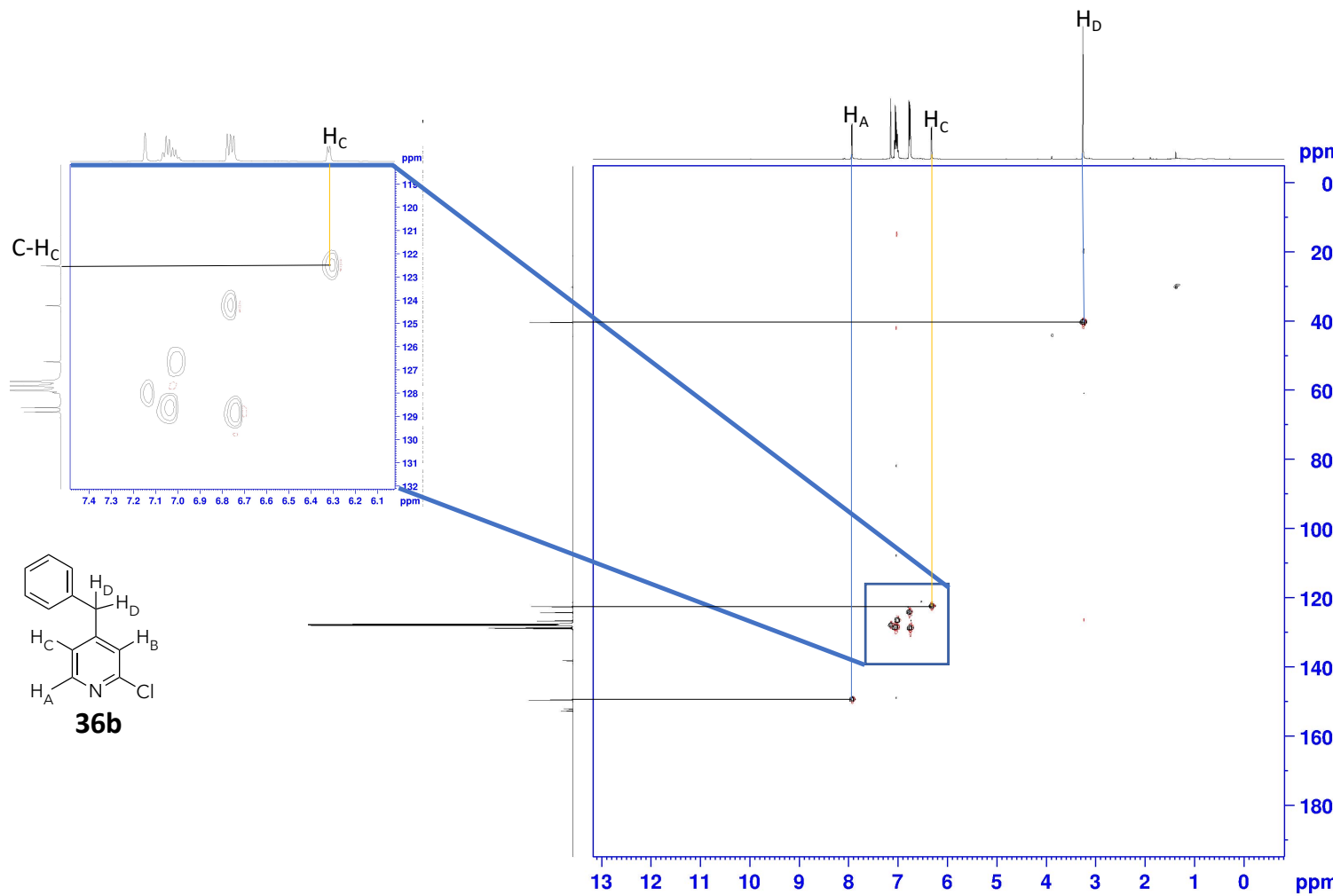
F1 - Acquisition parameters
TD       256
SFO1     500.233 MHz
FIDRES   50.796490 Hz
SW       12.998 ppm
FhMODE   QF

F2 - Processing parameters
SI       2048
SF       500.2299911 MHz
WDW      QSI
SSB      0
LB       0 Hz
GB       0
PC       1.40

F1 - Processing parameters
SI       2048
MC2      QF
SF       500.2299933 MHz
WDW      QSI
SSB      0
LB       0 Hz
GB       0
    
```



# HSQC



Current Data Parameters

NAME	NL-1-61-C4-2d
EXPNO	14
PROCNO	1

F2 - Acquisition Parameters

Date_	20210701
Time	19.07 h
INSTRUM	spect
PROBHD	Z125869_0055 (
PULPROG	hsgcqtgq
TD	1024
SOLVENT	C6D6
NS	8
DS	16
SWH	7002.801 Hz
FIDRES	13.677346 Hz
AQ	0.0731136 sec
RG	190.40
DW	71.400 usec
DE	16.00 usec
TE	300.0 K
CNST2	145.0000000
DO	0.00000300 sec
D1	1.50000000 sec
D4	0.00172414 sec
D11	0.03000000 sec
D16	0.00020000 sec
IND	0.00001990 sec
TDav	1
ZSOP1NS	
SFO1	500.2330889 MHz
MUCL	1H
P1	12.00 usec
P2	24.00 usec
P28	0 usec
PLA1	11.44699955 W
SFO2	125.7948829 MHz
NUC2	13C
CPDPRG2	gssp
P3	10.00 usec
P4	20.00 usec
PCPD2	70.00 usec
PLM2	56.90299983 W
PLW12	1.16129994 W
GFNAM[1]	SMSQ10.100
GF21	80.00 %
GFNAM[2]	SMSQ10.100
GF22	20.10 %
PL6	1000.00 usec

F1 - Acquisition parameters

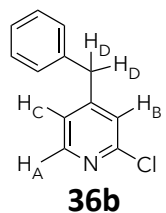
TD	256
SFO1	125.7949 MHz
FIDRES	196.293976 Hz
SW	199.735 ppm
FnMODE	Echo-Antiecho

F2 - Processing parameters

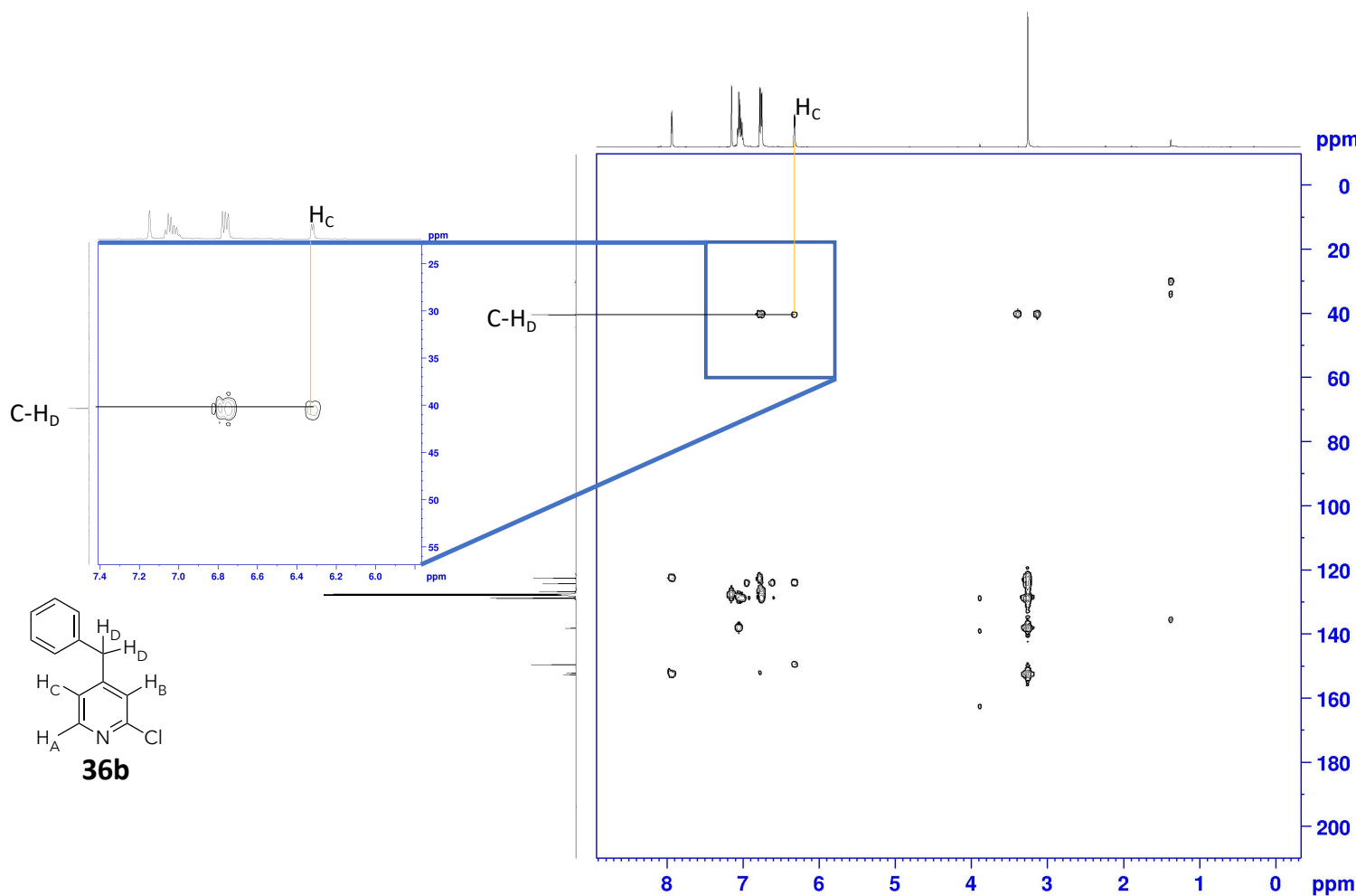
SI	1024
SF	500.2300000 MHz
WDW	Q5INE
SSB	2
LB	0 Hz
GB	0
PC	1.40

F1 - Processing parameters

SI	1024
MC2	echo-antiecho
SF	125.7829335 MHz
WDW	Q5INE
SSB	2
LB	0 Hz
GB	0



# HMBC



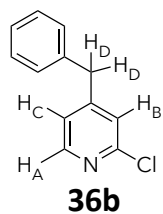
Current Data Parameters  
 NAME NL-1-61-C4-2d  
 EXPNO 12  
 PROCNO 1

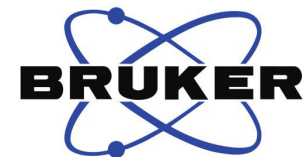
F2 - Acquisition Parameters  
 Date\_ 20210701  
 Time 18.24 h  
 INSTRUM spect  
 PROBHD z125869\_0055 ( hmbcgp1pndgf  
 PULPROG  
 TD 1024  
 SOLVENT c6d6  
 NS 4  
 DS 16  
 SWH 4629.629 Hz  
 FIDRES 9.042245 Hz  
 AQ 0.1105920 sec  
 RG 190.44  
 DW 108.000 usec  
 DE 16.00 usec  
 TE 300.0 K  
 CNST2 145.0000000  
 CNST13 10.0000000  
 DO 0.0000000 sec  
 D1 1.5000000 sec  
 D2 0.00344828 sec  
 D6 0.0500000 sec  
 D16 0.0002000 sec  
 IN0 0.00001810 sec  
 Tdavn 1  
 SFO1 500.2321336 MHz  
 NUC1 1H  
 P1 12.00 usec  
 P2 24.00 usec  
 PLW1 11.44699955 W  
 SFO2 125.7955118 MHz  
 NUC2 13C  
 P3 10.00 usec  
 PLW2 56.90299988 W  
 GENAM[1] SMSQ10.100  
 GEZ1 50.00 %  
 GENAM[2] SMSQ10.100  
 GEZ2 30.00 %  
 GENAM[3] SMSQ10.100  
 GEZ3 40.10 %  
 P16 1000.00 usec

F1 - Acquisition parameters  
 TD 128  
 SFO1 125.7955 MHz  
 FIDRES 431.629822 Hz  
 SW 219.597 ppm  
 FMODE QF

F2 - Processing parameters  
 SI 1024  
 SF 500.2299839 MHz  
 WDW SINE  
 SSB 0  
 LB 0 Hz  
 GB 0  
 PC 1.40

F1 - Processing parameters  
 SI 1024  
 MC2 QF  
 SF 125.7829233 MHz  
 WDW SINE  
 SSB 0  
 LB 0 Hz  
 GB 0

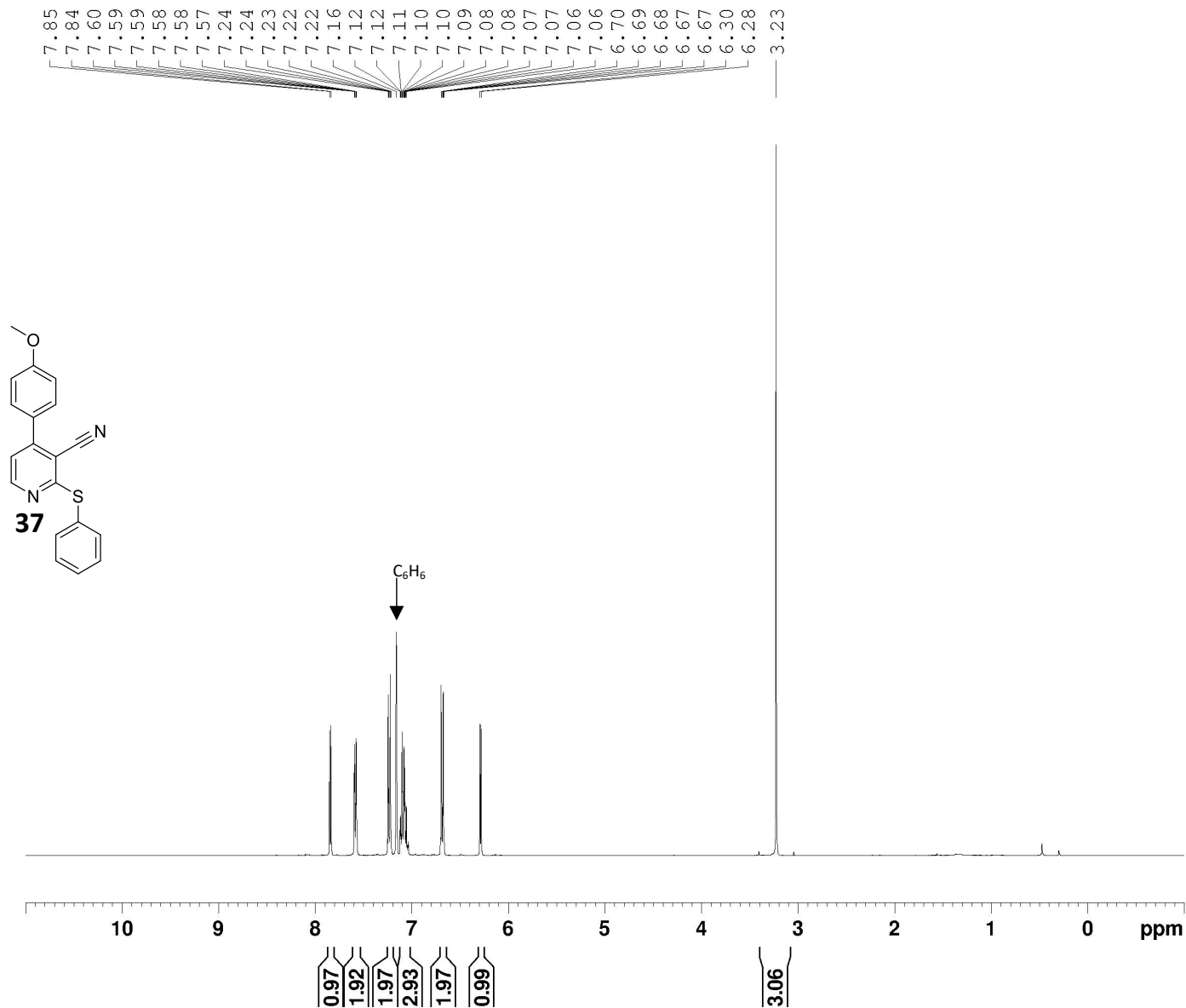


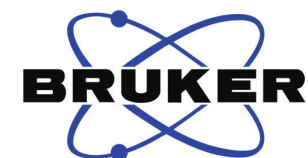
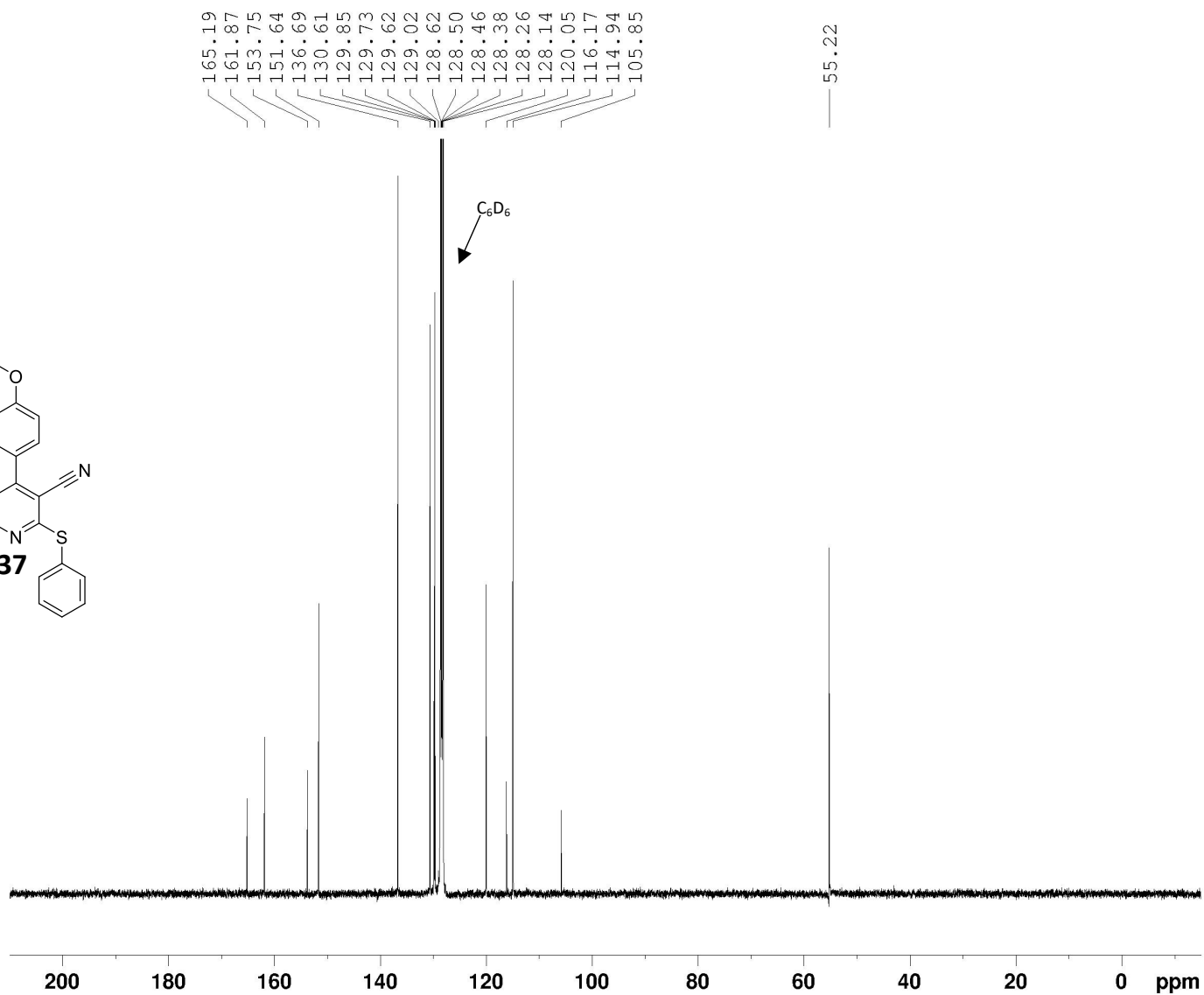
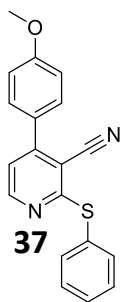


Current Data Parameters  
NAME JPN-1-189-1  
EXPNO 15  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211107  
Time 20.23 h  
INSTRUM Avance Neo  
PROBHD z152088\_0031 (  
PULPROG zg30  
TD 65536  
SOLVENT C6D6  
NS 16  
DS 2  
SWH 8196.722 Hz  
FIDRES 0.250144 Hz  
AQ 3.9976959 sec  
RG 101  
DW 61.000 usec  
DE 13.89 usec  
TE 298.0 K  
D1 1.00000000 sec  
TD0 1  
SFO1 400.1324708 MHz  
NUC1 1H  
P0 2.67 usec  
P1 8.00 usec  
PLW1 24.03499985 W

F2 - Processing parameters  
SI 65536  
SF 400.1299968 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

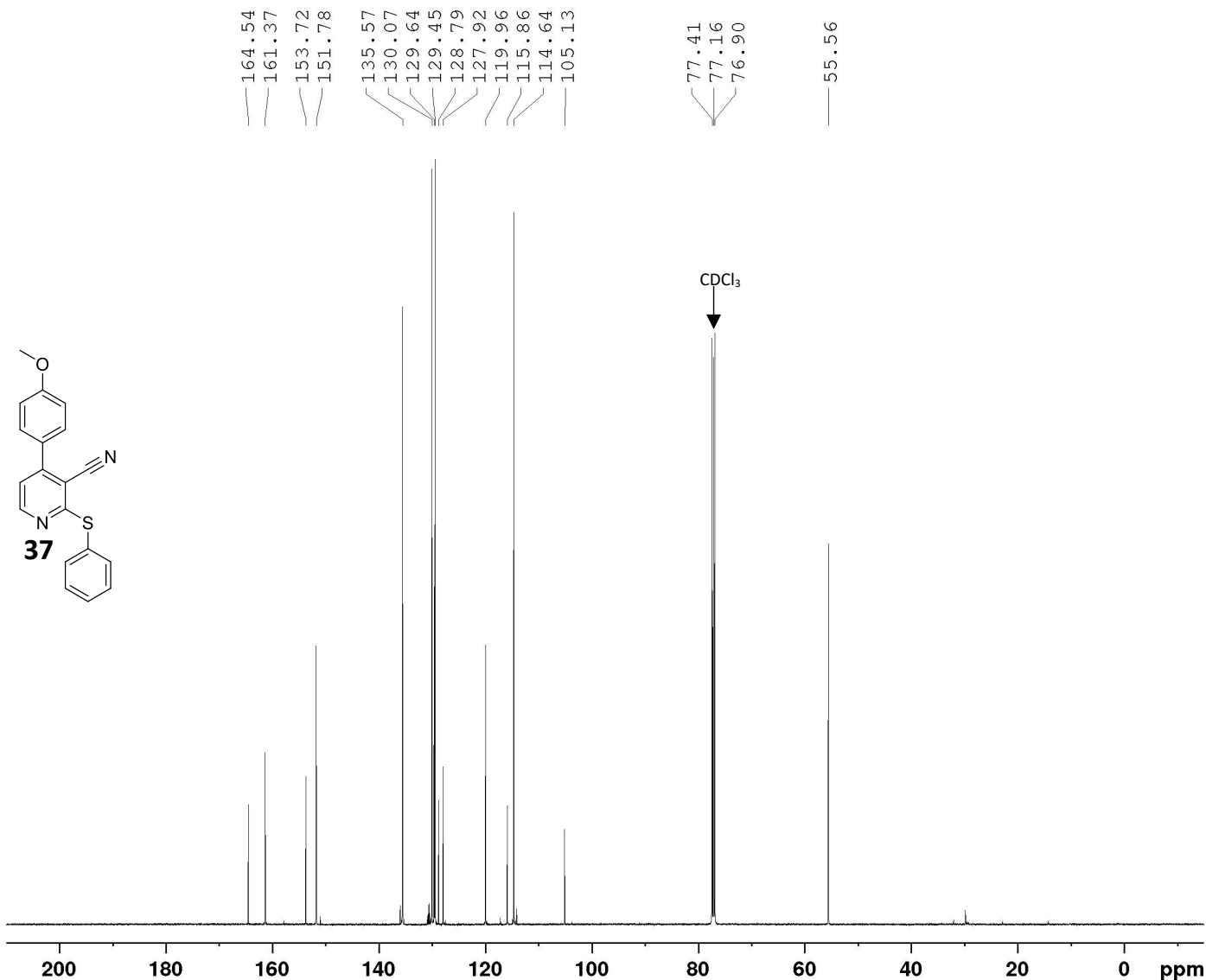
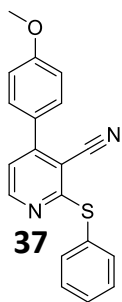




Current Data Parameters  
 NAME JPN-1-189-1  
 EXPNO 14  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211107  
 Time 20.21 h  
 INSTRUM Avance Neo  
 PROBHD z152088\_0031 ( )  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 1024  
 DS 4  
 SWH 23809.523 Hz  
 FIDRES 0.726609 Hz  
 AQ 1.3762560 sec  
 RG 8.125  
 DW 21.000 usec  
 DE 6.50 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 100.6228298 MHz  
 NUC1 13C  
 P0 2.67 usec  
 P1 8.00 usec  
 PLW1 86.55400085 W  
 SFO2 400.1316005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz65  
 PCPD2 90.00 usec  
 PLW2 24.03499985 W  
 PLW12 0.18990999 W  
 PLW13 0.09552100 W

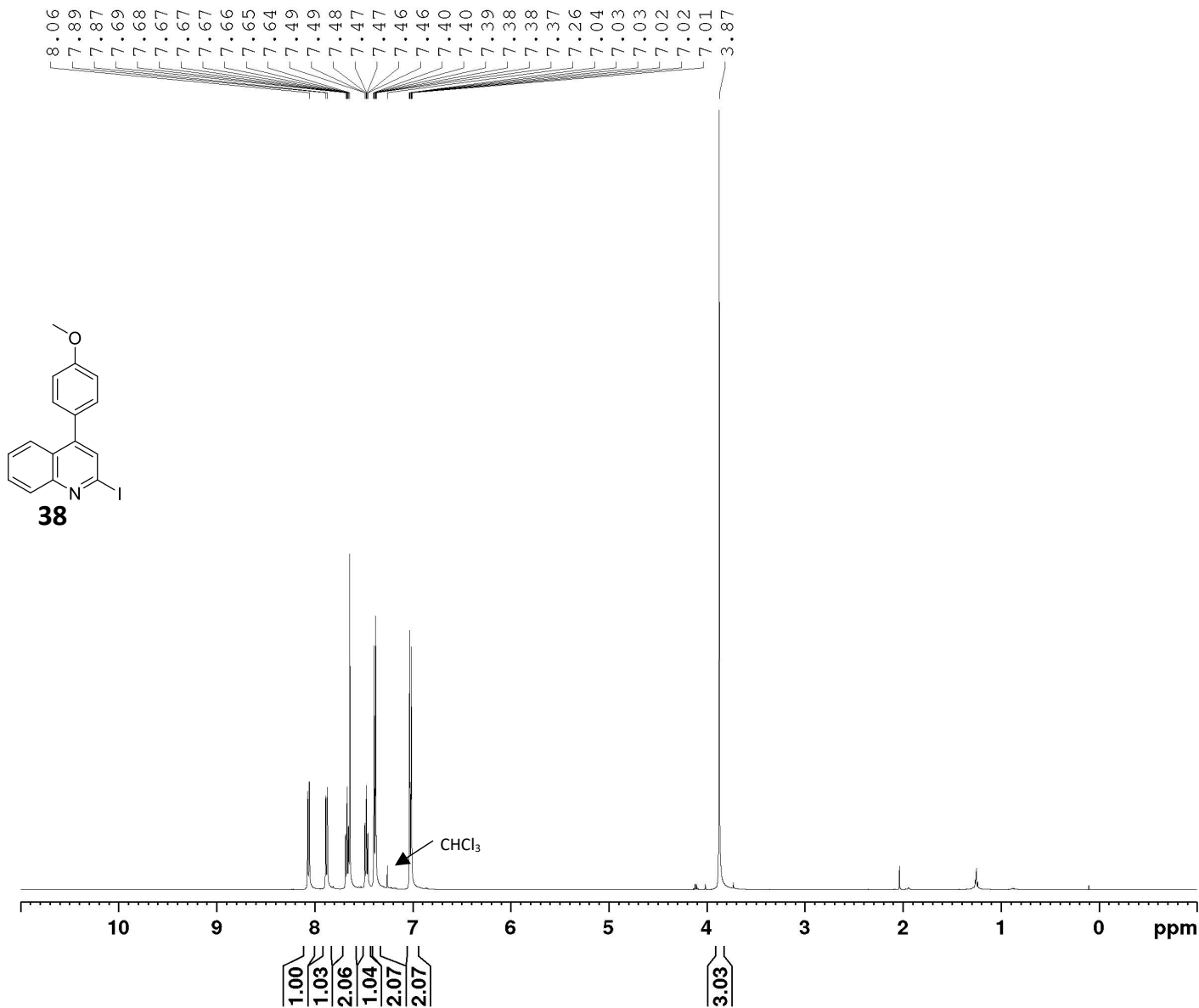
F2 - Processing parameters  
 SI 32768  
 SF 100.6126995 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
 NAME JPN-1-189-1\_CDC13  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211209  
 Time 0.37 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

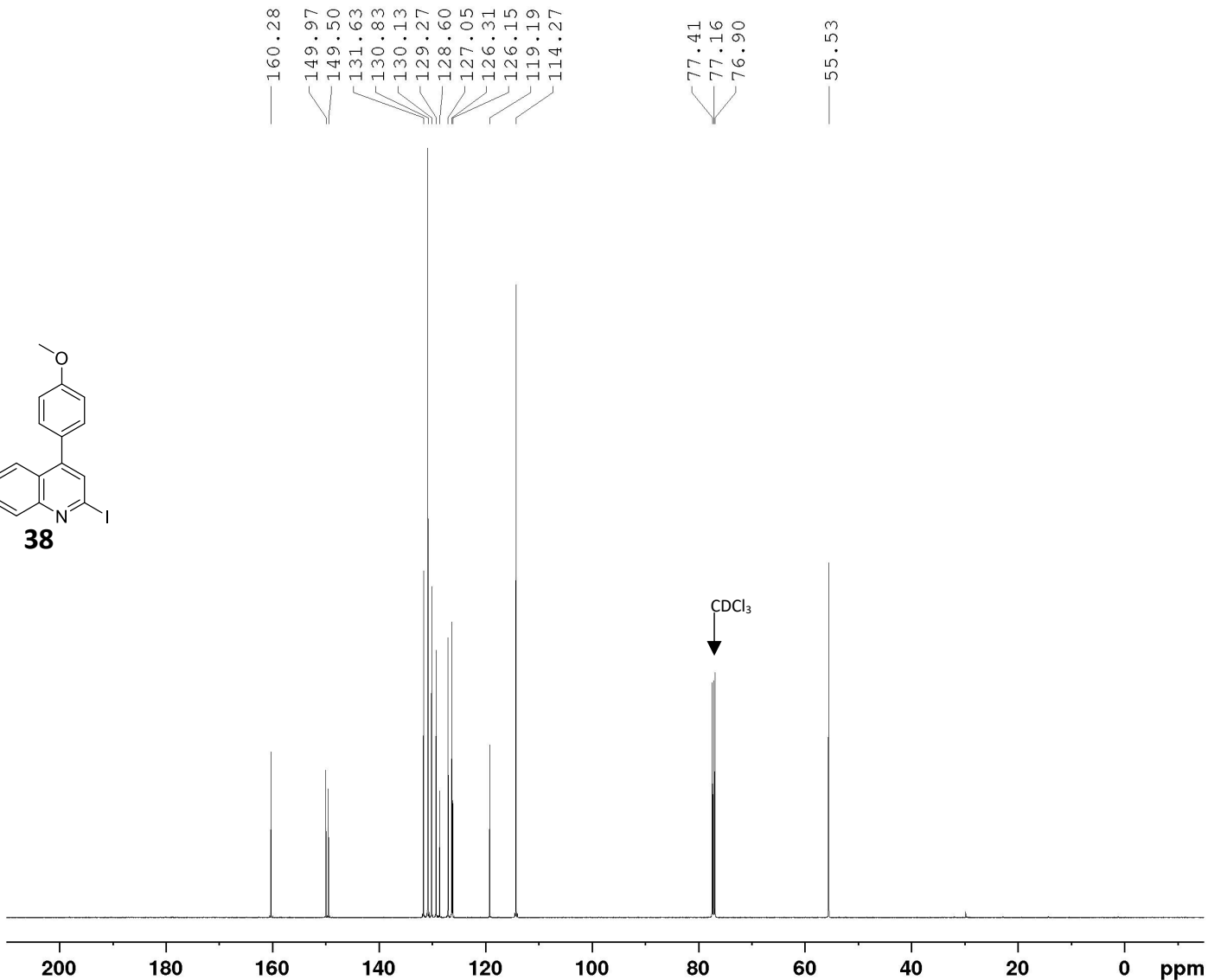
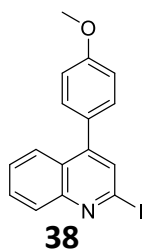
F2 - Processing parameters  
 SI 32768  
 SF 125.7829231 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
NAME JPN-2-25  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20201019  
Time 18.36 h  
INSTRUM spect  
PROBHD Z125869\_0055 ( )  
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 22.16  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

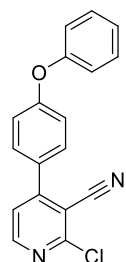
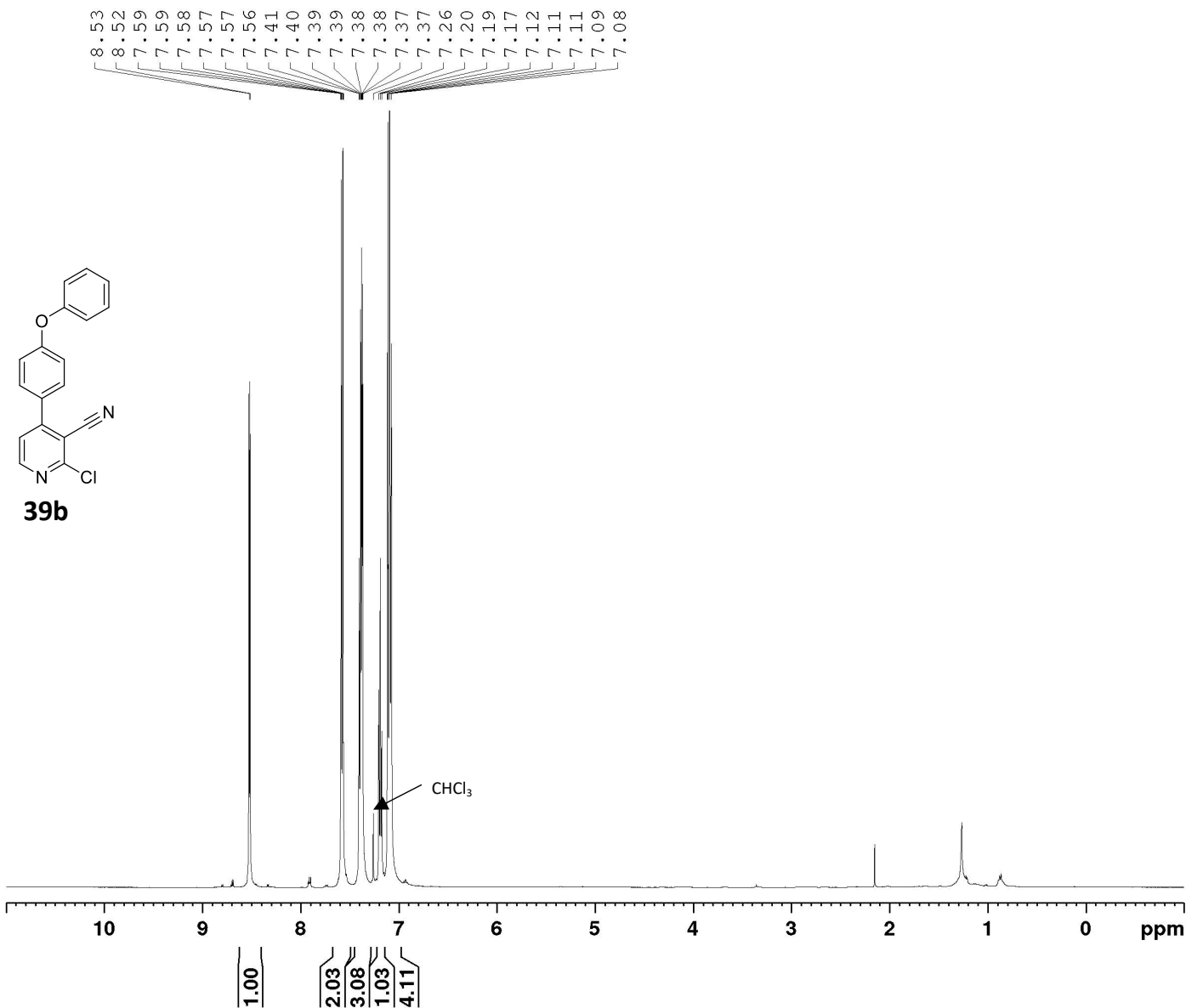
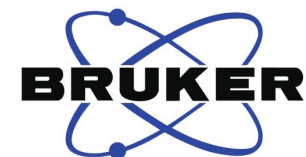
F2 - Processing parameters  
SI 65536  
SF 500.2300124 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME JPN-2-25-1  
 EXPNO 17  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211120  
 Time 4.43 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zgpg30  
 ID 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

F2 - Processing parameters  
 SI 32768  
 SF 125.7829267 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



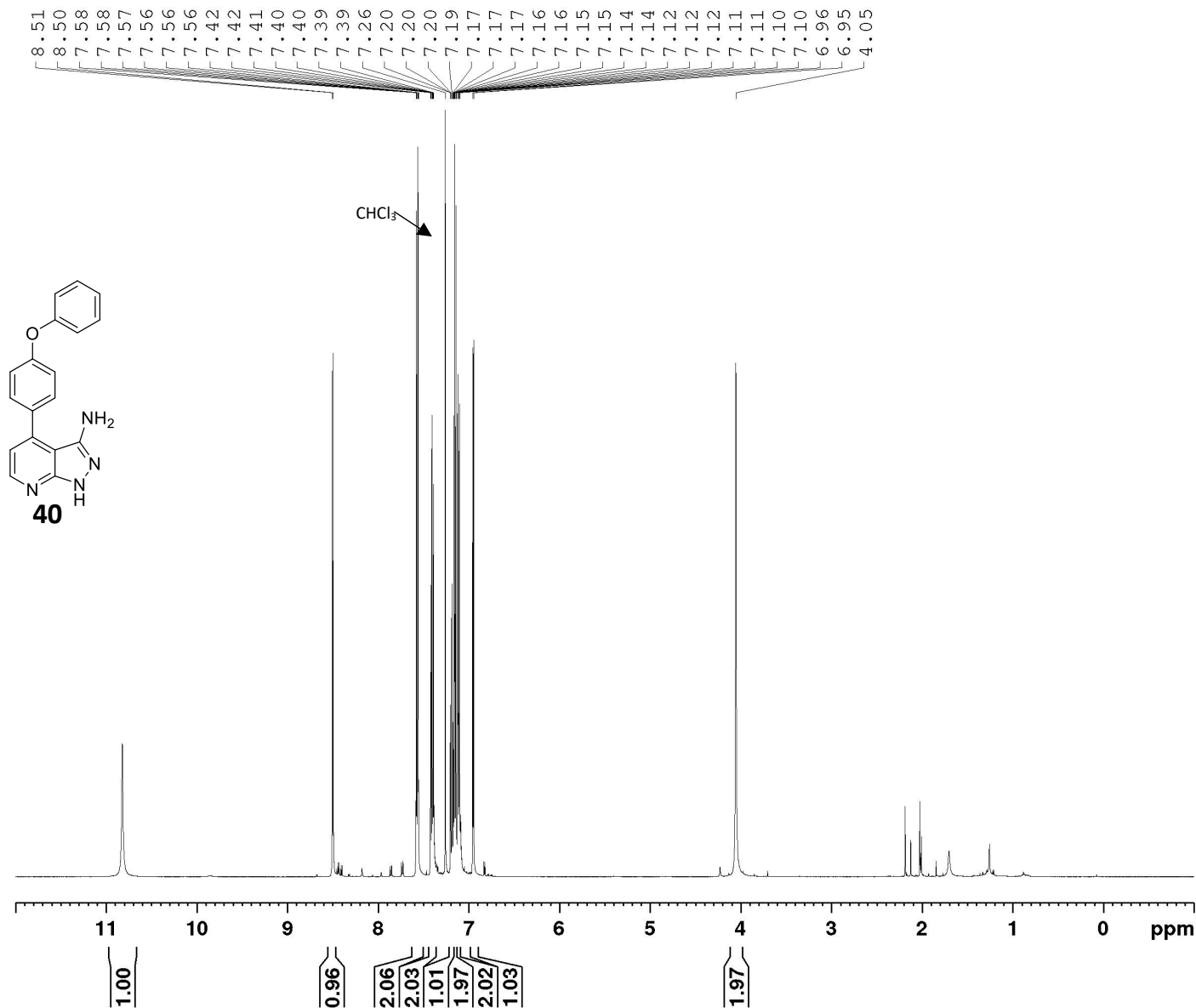
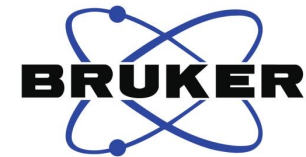
39b

Current Data Parameters  
NAME JPN-2-6-C4-mono\_dried  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20200823  
Time 8.03 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 19.46  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SF01 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300120 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

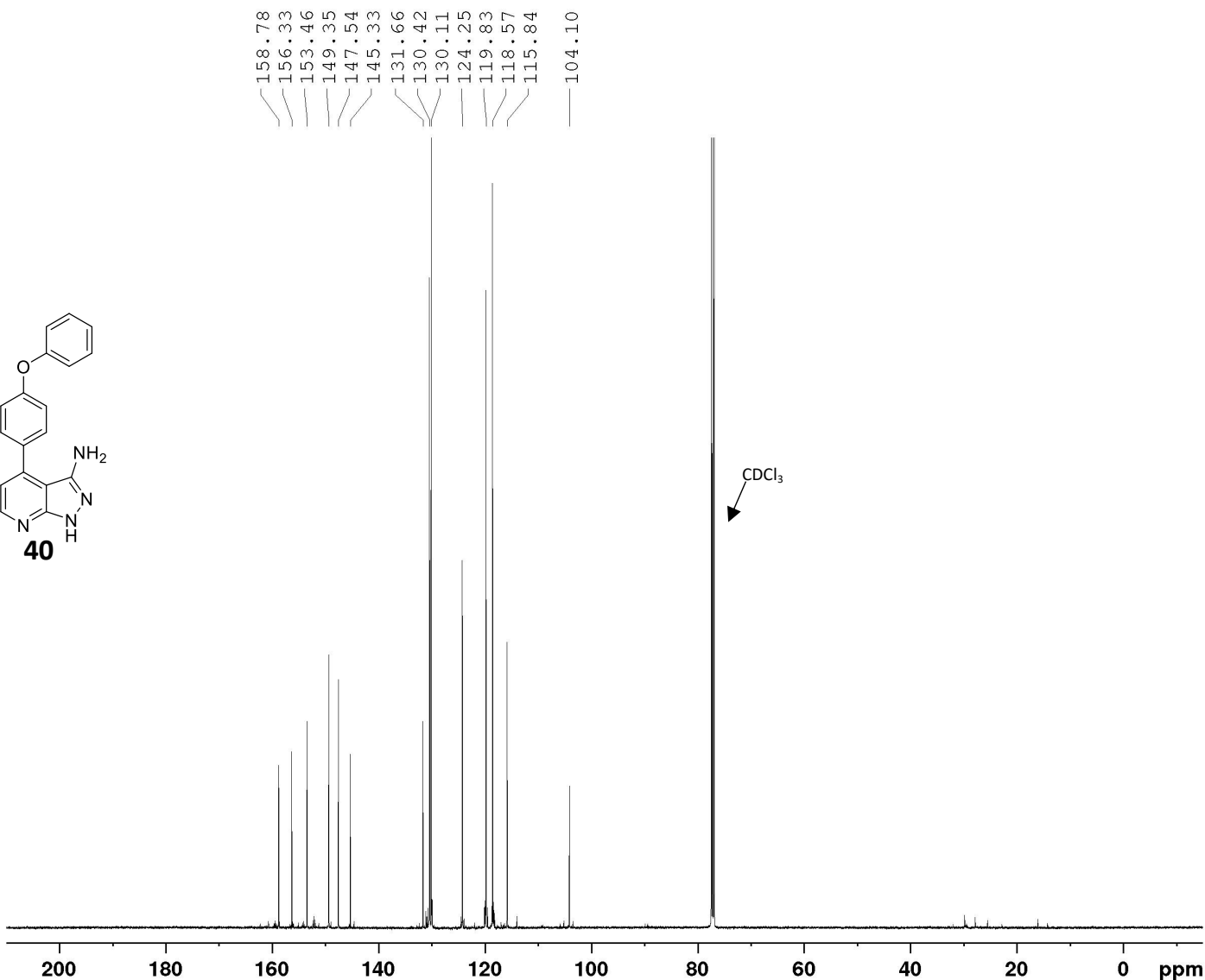
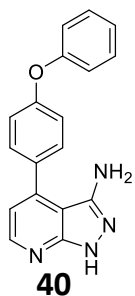




Current Data Parameters  
NAME JPN-2-6-2\_repeat  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20201019  
Time 18.27 h  
INSTRUM spect  
PROBHD Z125869\_0055 (  
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 64  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 151.18  
DW 50.000 usec  
DE 16.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

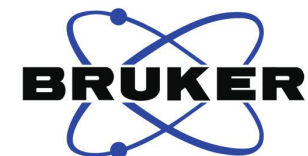
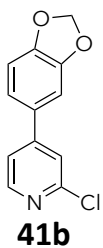
F2 - Processing parameters  
SI 65536  
SF 500.2300122 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME JPN-2-6-2  
 EXPNO 17  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211120  
 Time 5.41 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

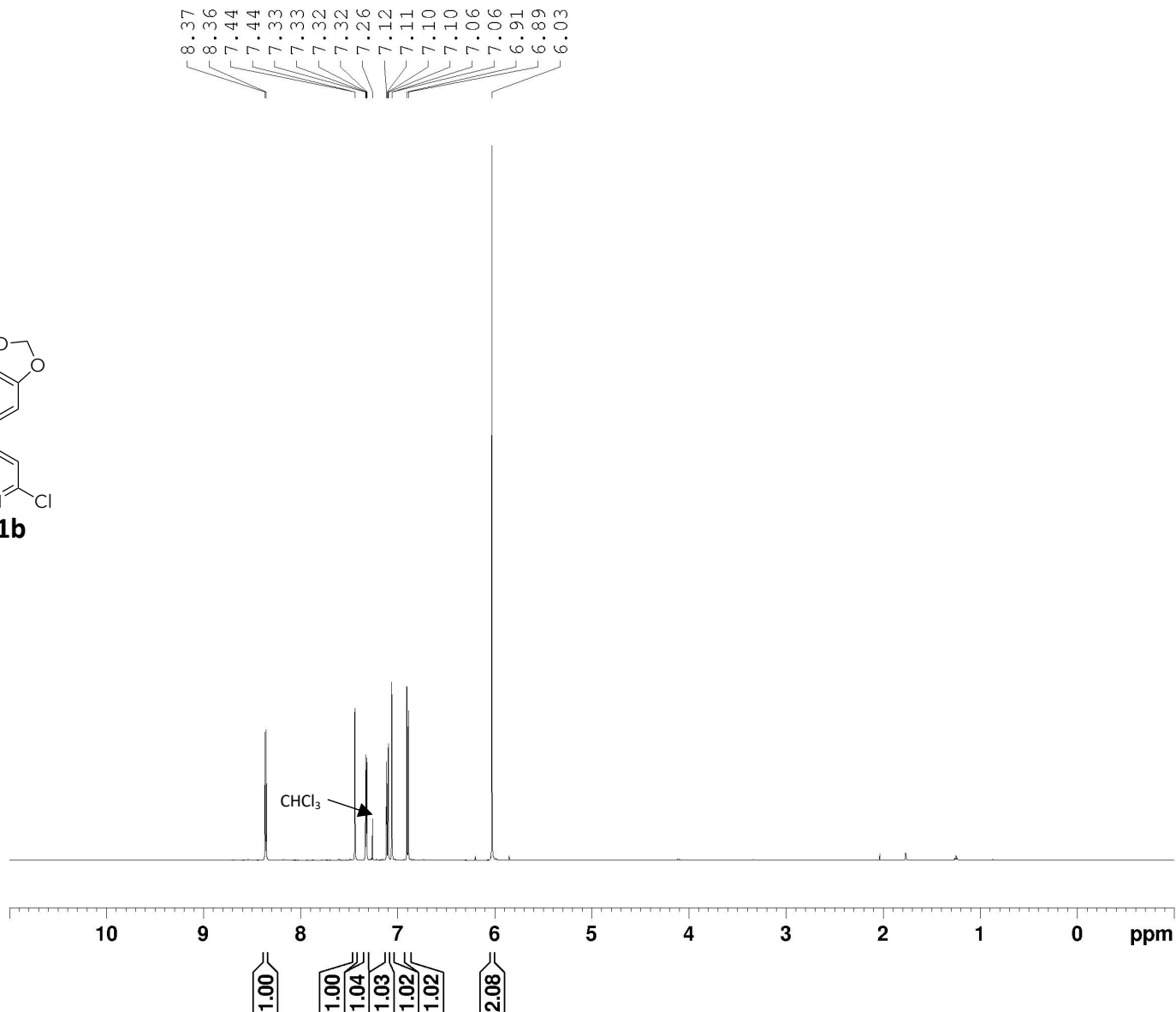
F2 - Processing parameters  
 SI 32768  
 SF 125.7829211 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

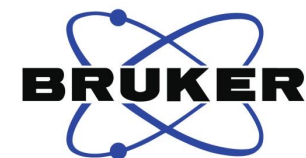
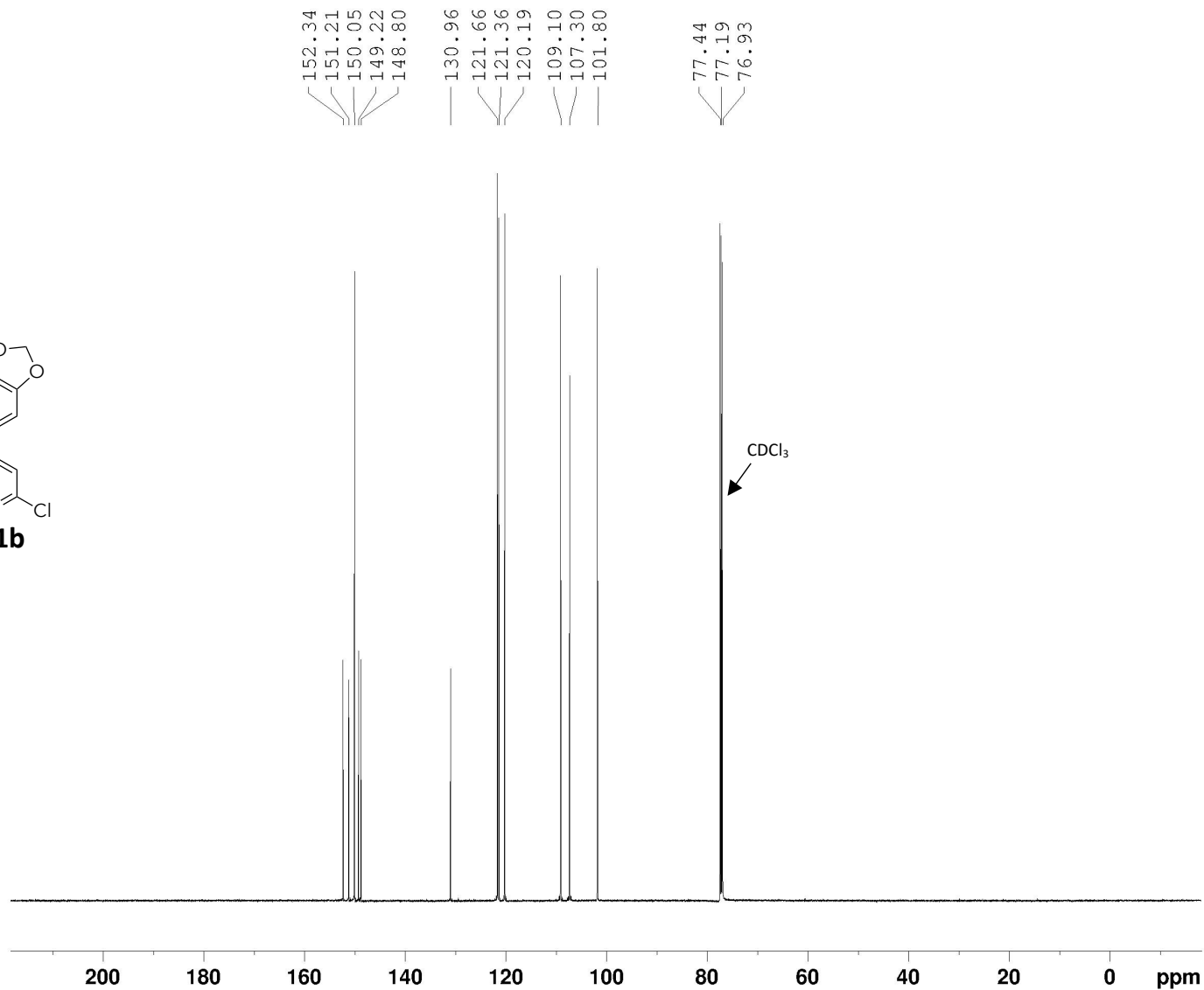
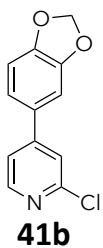


Current Data Parameters  
 NAME JPN-2-153-3  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20220228  
 Time 14.24 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDC13  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 43.72  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

F2 - Processing parameters  
 SI 65536  
 SF 500.2300120 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

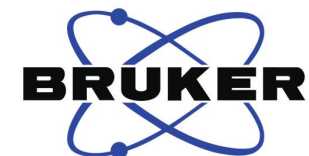
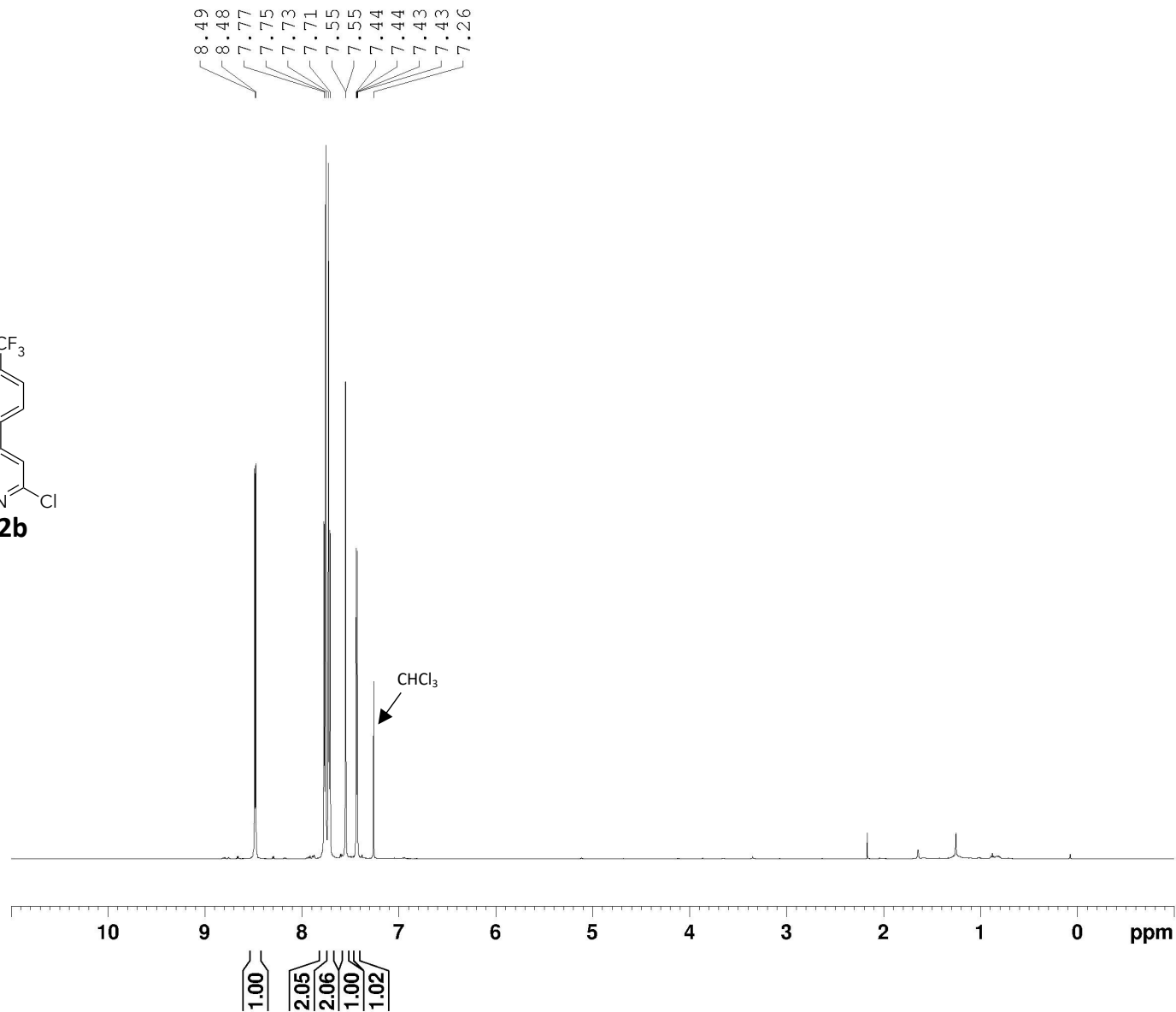
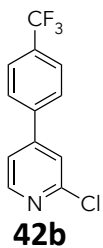




Current Data Parameters  
 NAME JPN-2-153-3  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20220301  
 Time 0.04 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

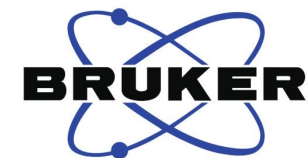
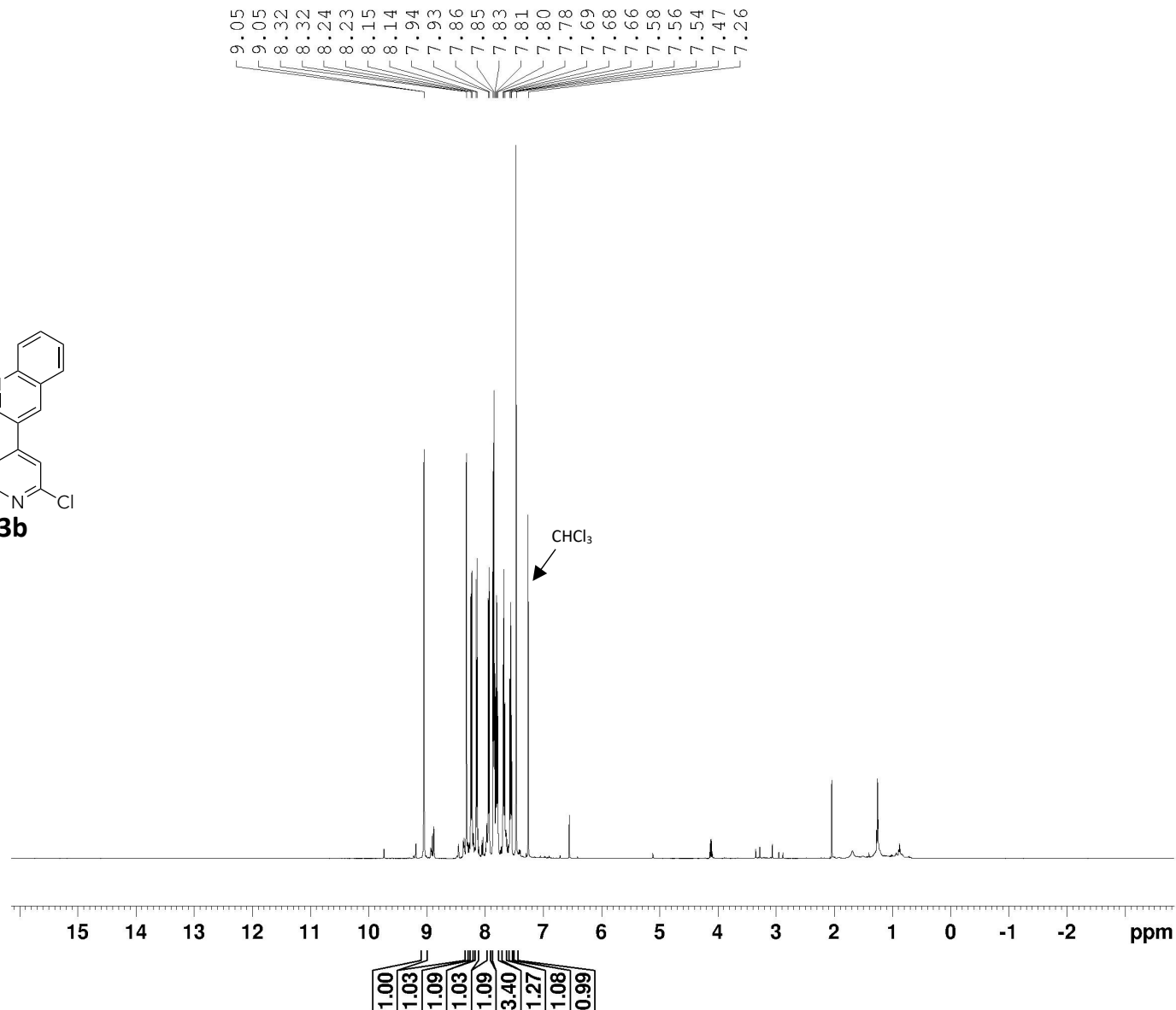
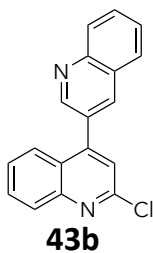
F2 - Processing parameters  
 SI 32768  
 SF 125.7829176 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
 NAME NL-1-186-1-F1  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20220317  
 Time 12.09 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 ( )  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 94.51  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 PC 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

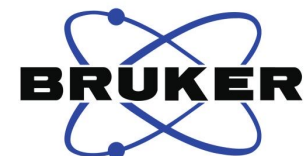
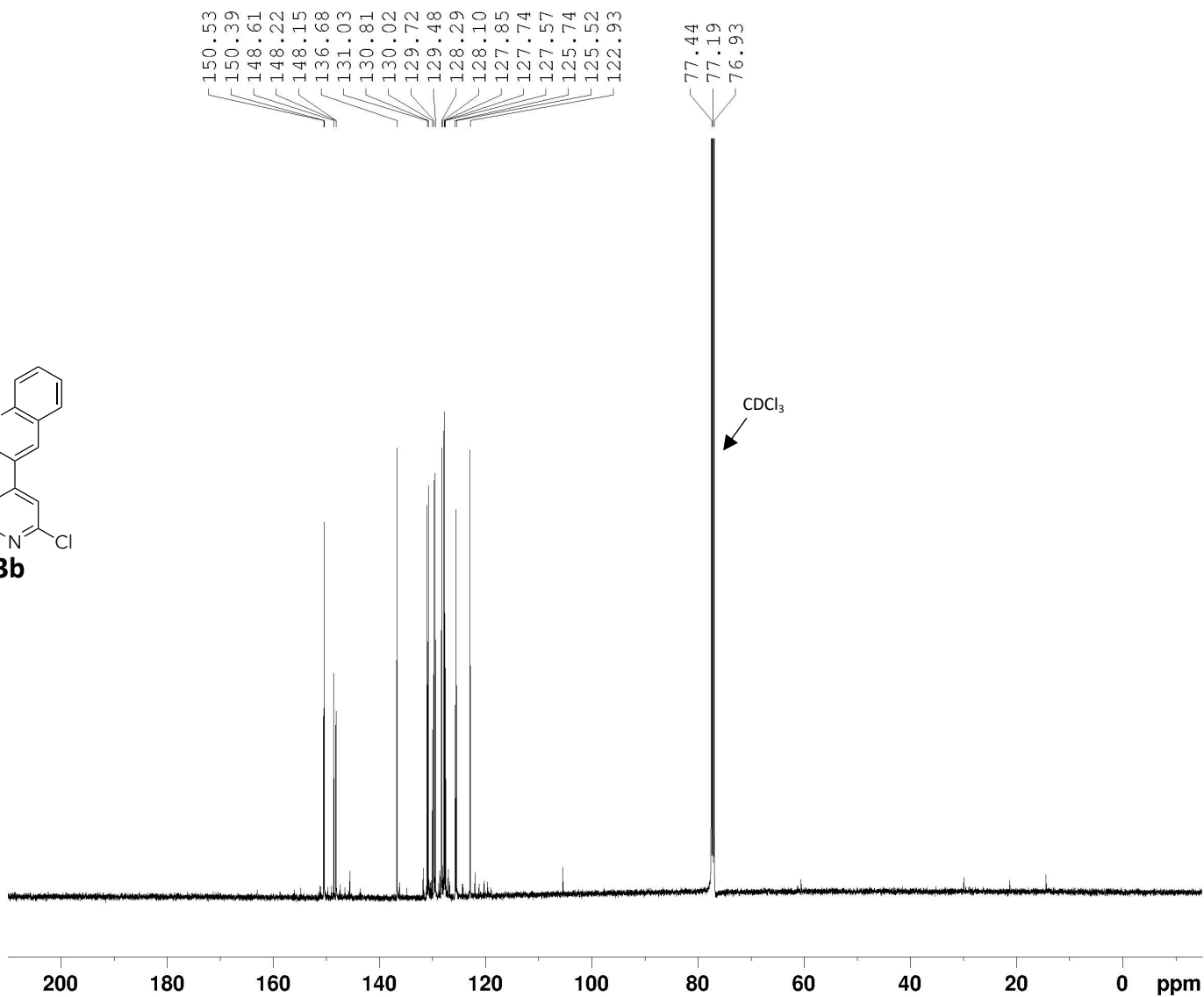
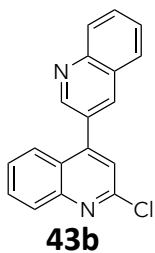
F2 - Processing parameters  
 SI 65536  
 SF 500.2300120 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
 NAME JPN-2-156-2  
 EXPNO 12  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20220317  
 Time 11.59 h  
 INSTRUM spect  
 PROBHD Z125869\_0055 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 151.18  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

F2 - Processing parameters  
 SI 65536  
 SF 500.2300120 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



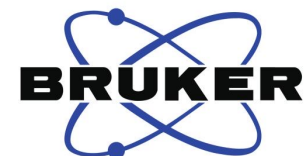
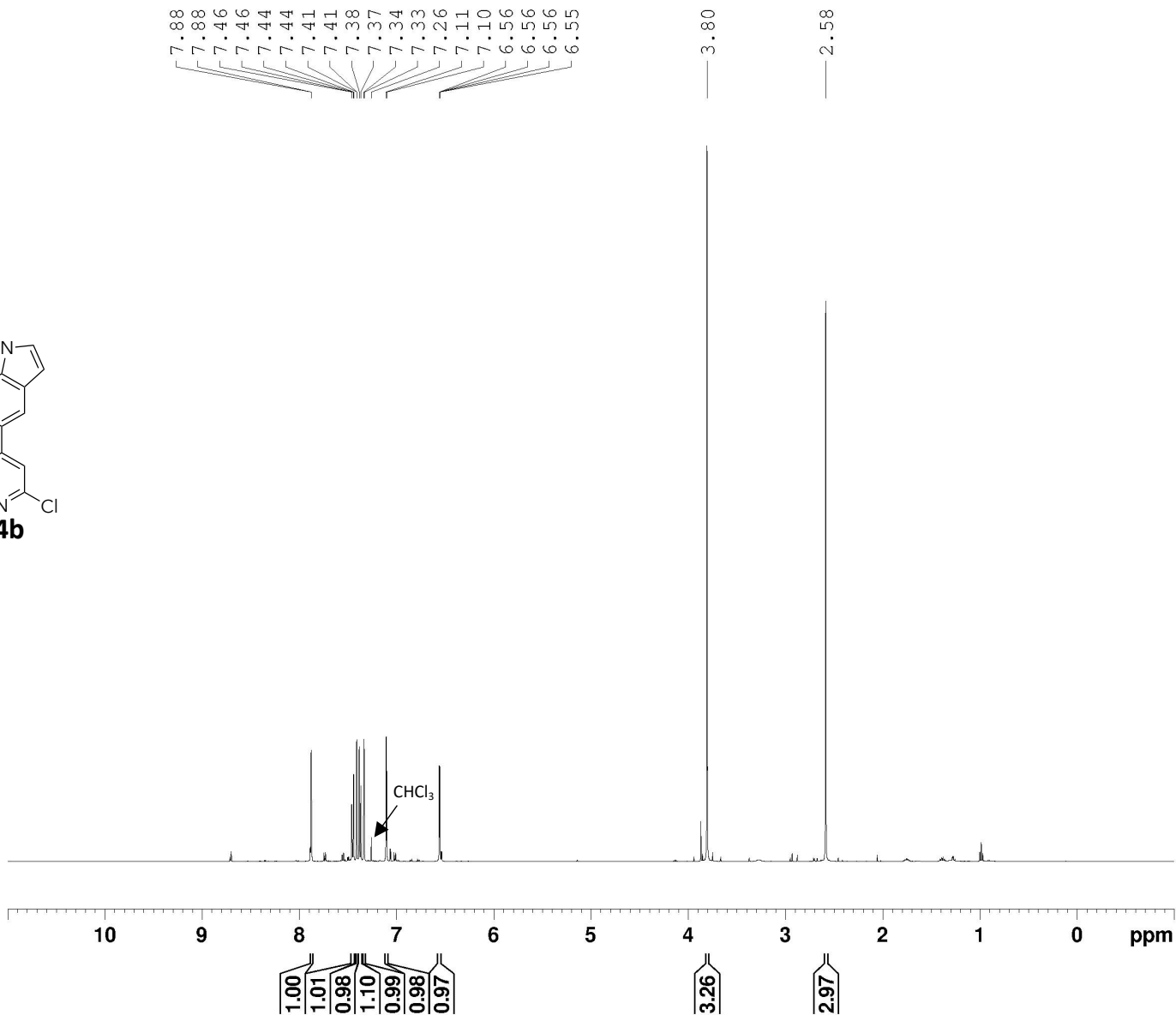
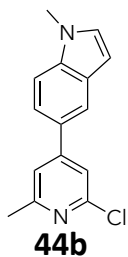
Current Data Parameters  
 NAME JPN-1-156-2  
 EXPNO 13  
 PROCNO 1

F2 - Acquisition Parameters

Date\_ 20220317  
 Time 17.59 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDC13  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

F2 - Processing parameters

SI 32768  
 SF 125.7829137 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

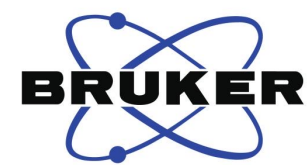
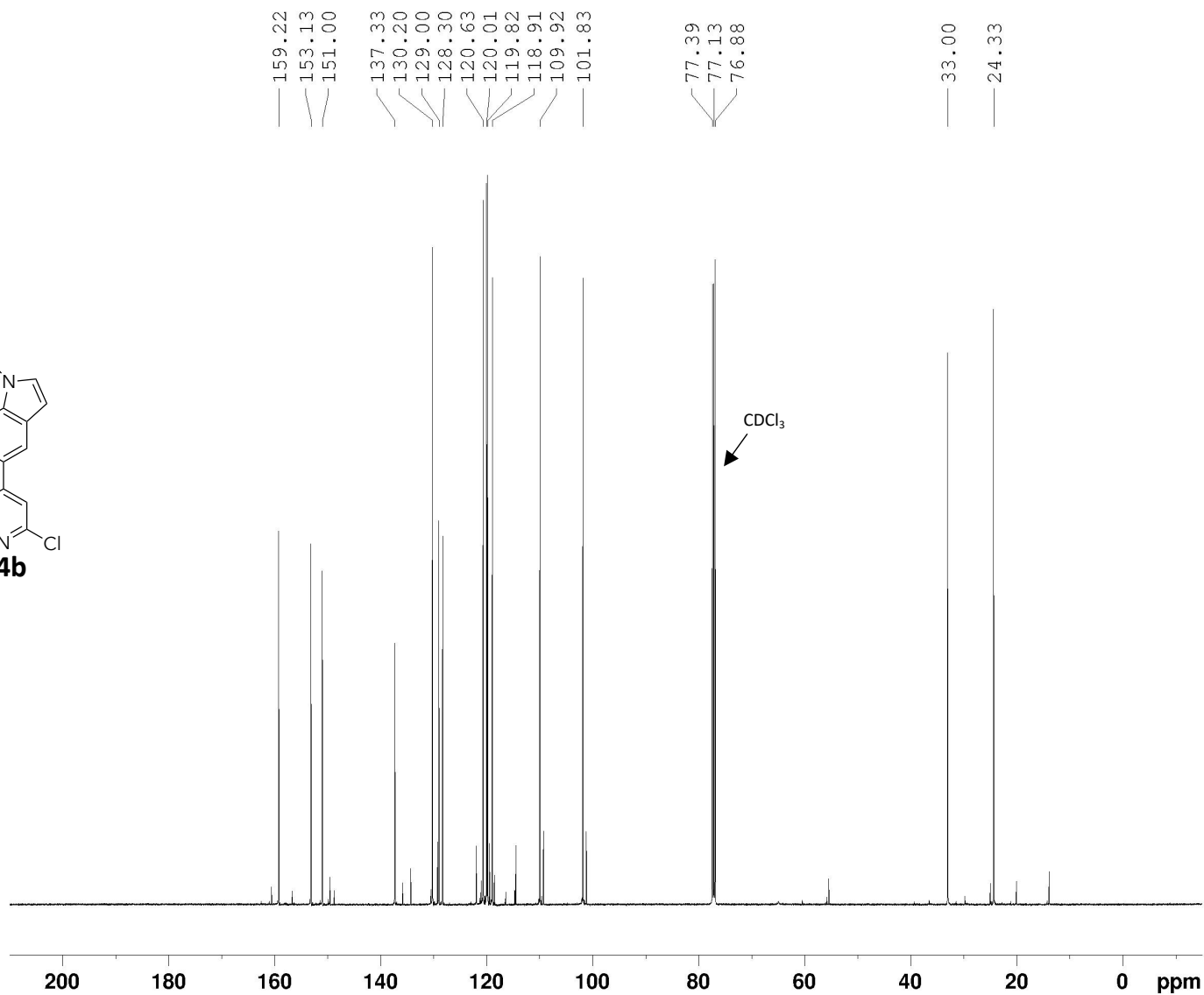
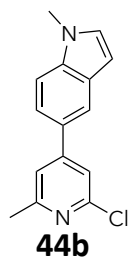


Current Data Parameters  
 NAME JPN-2-156-3  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20220316  
 Time 3.15 h  
 INSTRUM spect  
 PROBHD z125869\_0055 (  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.305176 Hz  
 AQ 3.2767999 sec  
 RG 26.9  
 DW 50.000 usec  
 DE 16.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 TD0 1  
 SFO1 500.2330889 MHz  
 NUC1 1H  
 P0 4.00 usec  
 P1 12.00 usec  
 PLW1 11.44699955 W

F2 - Processing parameters  
 SI 65536  
 SF 500.2300119 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00





Current Data Parameters  
 NAME JPN-2-156-3  
 EXPNO 11  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20220316  
 Time 4.13 h  
 INSTRUM spect  
 PROBHD z125869\_0055 ( )  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 4  
 SWH 29761.904 Hz  
 FIDRES 0.908261 Hz  
 AQ 1.1010048 sec  
 RG 190.44  
 DW 16.800 usec  
 DE 18.00 usec  
 TE 298.0 K  
 D1 2.00000000 sec  
 D11 0.03000000 sec  
 TD0 1  
 SFO1 125.7955118 MHz  
 NUC1 13C  
 P0 3.33 usec  
 P1 10.00 usec  
 PLW1 56.90299988 W  
 SFO2 500.2320009 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 80.00 usec  
 PLW2 11.44699955 W  
 PLW12 0.25756001 W  
 PLW13 0.12955000 W

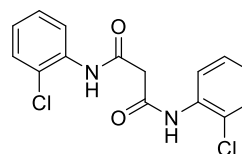
F2 - Processing parameters  
 SI 32768  
 SF 125.7829335 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



Current Data Parameters  
NAME JPN-2-143\_DMSO-D6  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20220211  
Time 14.25 h  
INSTRUM spect  
PROBHD Z125869\_0055 ( )  
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 86.13  
DW 50.000 usec  
DE 16.00 usec  
TE 298.0 K  
D1 2.00000000 sec  
TD0 1  
SFO1 500.2330889 MHz  
NUC1 1H  
P0 4.00 usec  
P1 12.00 usec  
PLW1 11.44699955 W

F2 - Processing parameters  
SI 65536  
SF 500.2300032 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



S48

