

Supplemental information

Figure S1. Study participant characteristics comparing immediate and delayed antiretroviral therapy,

Related to Figure 1.

Figure S2. Batch effect removal by fastMNN, gene expression differences across clusters, and modules identified by WGCNA to differentiate HIV-1 viremic conditions from HIV-1 viral suppression conditions. Related to Figure 1.

Figure S3. Differences related to immediate or deferred initiation of ART. Related to Figure 1, Figure 2, and Figure 3.

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Figure S6. Clone phenotypes and the size of HIV-1 RNA⁺ clones vs HIV-1 RNA⁻ clones. Related to Figure 5 and Figure 6.

Figure S7. Machine learning identification of cellular markers necessary and sufficient to distinguish HIV-1 RNA⁺ T cell clones from HIV-1 RNA⁻ T cell clones by scRFE and flow cytometry validation for HIV-1 p24⁺ cells. Related to Figure 7.

Related to Figure 7.

Table S1 Clinical demographics. Related to Figure 1.

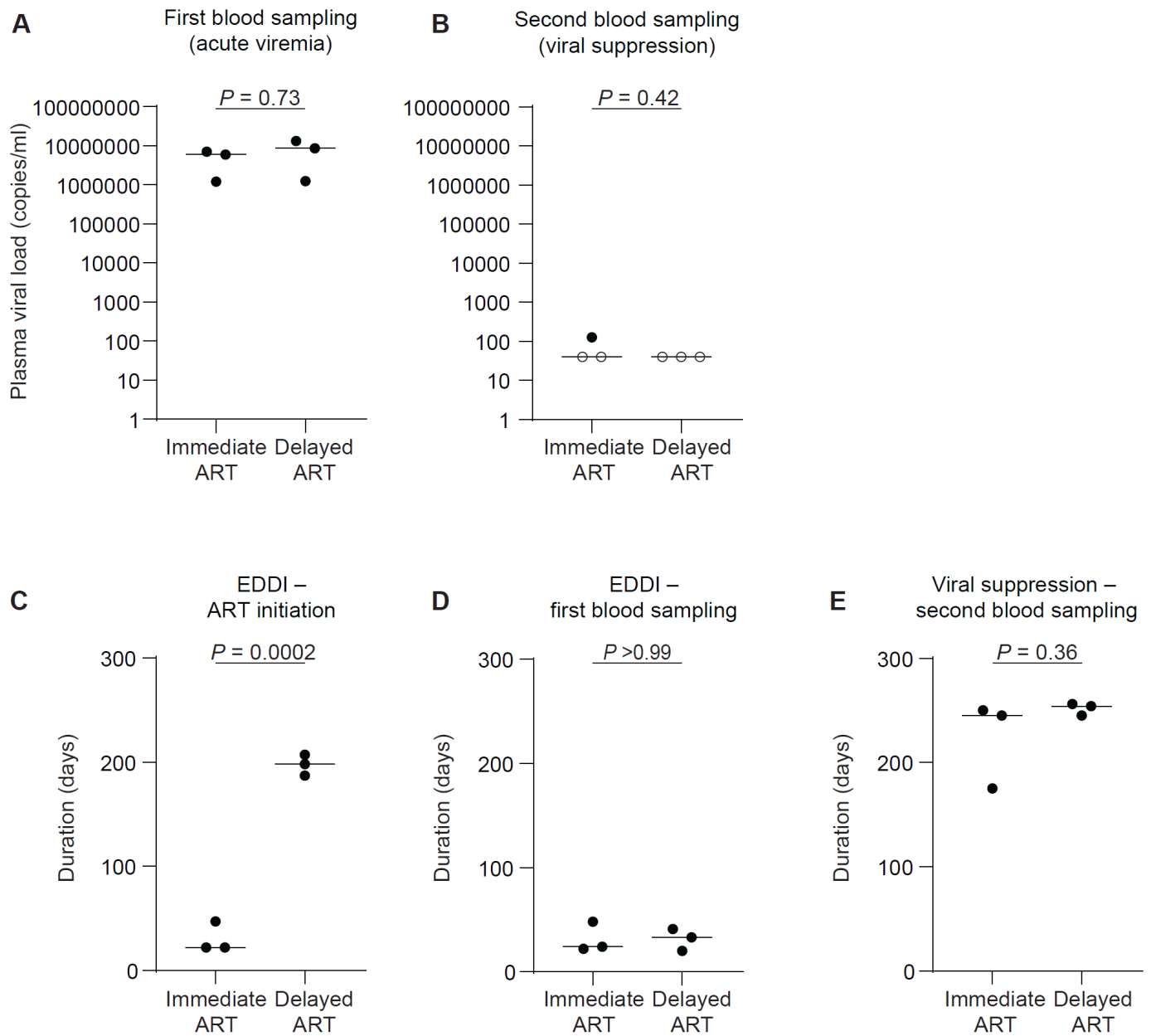


Figure S1. Study participant characteristics comparing immediate and delayed antiretroviral therapy, Related to Figure 1.

Plasma viral loads during the **A**, viremia timepoint and after **B**, viral suppression is achieved. Number of days between the estimated date of infection and the **C**, initiation of antiretroviral therapy or **D**, the time of first blood sampling. **E**, The duration of viral suppression.

P values were determined by two-tailed paired Student's t -test. EDDI, estimated date of detectable infection.

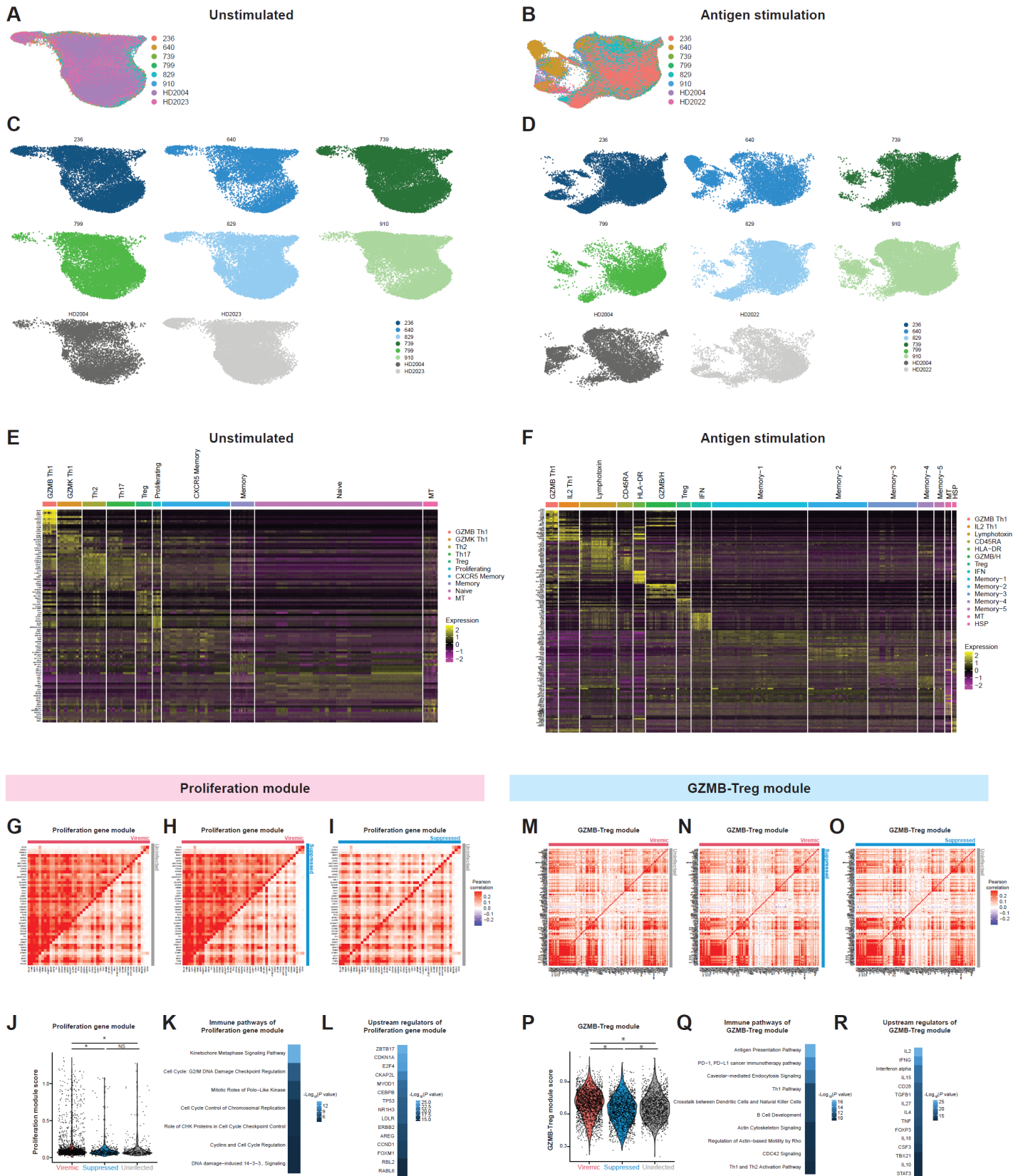


Figure S2. Batch effect removal by FastMNN, Gene expression differences across clusters, and Modules identified by WGCNA to differentiate HIV-1 viremic conditions from HIV-1 viral suppression conditions. Related to Figure 1.

UMAP plots showing cells colored by the participant in unstimulated condition (**A**) and antigen stimulated condition (**B**). Heatmaps showing the top 10 genes for each cluster in unstimulated condition (**C, E**) and antigen-stimulated condition (**D, F**). Of the WGCNA

defined modules, two separated specifically viremia and viral suppression. The proliferation defined module was induced by viremia relative to **G**, uninfected and **H**, virally suppressed individuals, but not in **I**, suppressed individuals relative to uninfected individuals. **J**, Increased expression of this module is restricted to proliferating cells in the viremic condition. Ingenuity pathway analysis (IPA) showing the **K**, immune pathways and **L**, upstream regulators of the proliferation gene module. The GZMB-Treg defined module was not induced by viremia relative to **M**, uninfected or **N**, virally suppressed individuals, and not in **I**, suppressed individuals relative to uninfected individuals. **O**, Increased expression of this module is restricted to Treg cells in the viremic condition relative to suppressed and uninfected, and in suppressed relative to uninfected. IPA analysis showing the **P**, immune pathways and **L**, upstream regulators of the proliferation gene module. **J** and **P**, * denotes $P < 0.05$ by Wilcoxon rank-sum test. **K**, **L**, **Q**, and **R**, Fisher's exact test with Benjamini-Hochberg procedure multiple hypothesis correction.

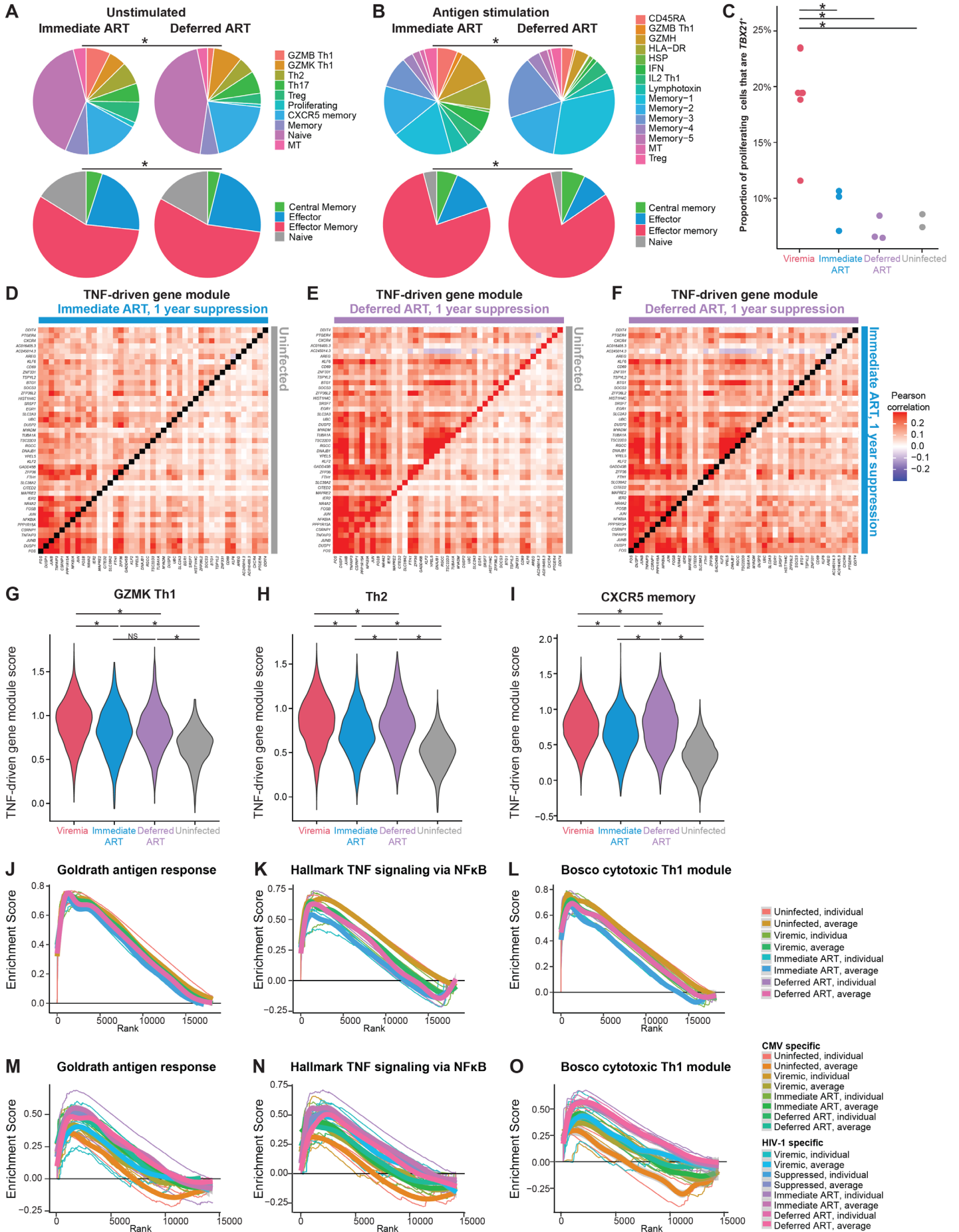


Figure S3. Differences related to immediate or deferred initiation of ART. Related to Figure 1, Figure 2, and Figure 3.

The cluster composition and memory phenotypes as defined by surface protein expression of **A**, unstimulated and **B**, antigen stimulated conditions comparing immediate and delayed antiretroviral therapy initiation. **C**, Proportion of cells in the proliferating cell cluster expressing *TBX21* (encoding Th1 transcription factor Tbet). The module in of Figure 1F–J showing a modest induction in **D**, immediate ART, and an increased induction in deferred ART relative to **E**, uninfected and **F**, immediate ART conditions. This TNF driven module has equivalent expression in immediate and deferred ART conditions in **G**, GZMK Th1, but increased expression in deferred ART conditions in **H**, Th2 and **I**, CXCR5 memory cells. Gene sets identified by GSEA correlating with clone size in unstimulated conditions include: **J**, Goldrath antigen response, **K**, Hallmark TNF signaling via NFκB, and **L**, Bosco cytotoxic Th1 module. Gene sets identified by GSEA correlating with clone size in antigen stimulated conditions include: **M**, Goldrath antigen response, **N**, Hallmark TNF signaling via NFκB, and **O**, Bosco cytotoxic Th1 module. **A** and **B**, $P < 0.05$, Fisher's exact test. **C**, **G**, **H**, $P < 0.05$, Wilcoxon rank-sum test.

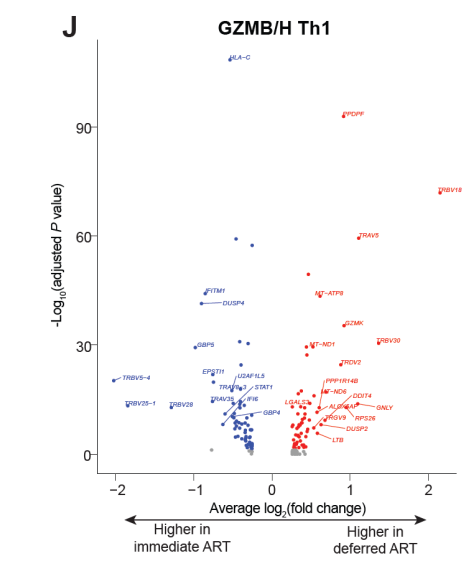
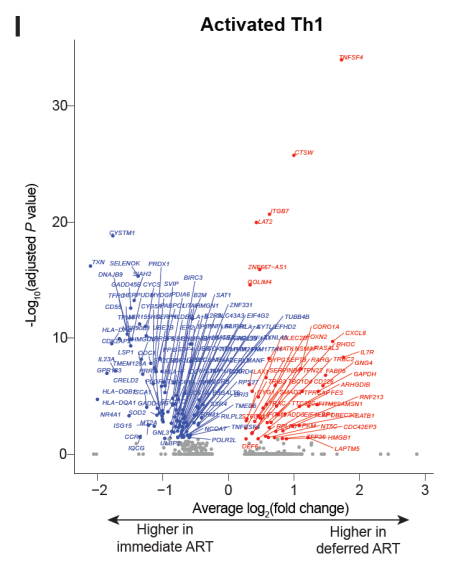
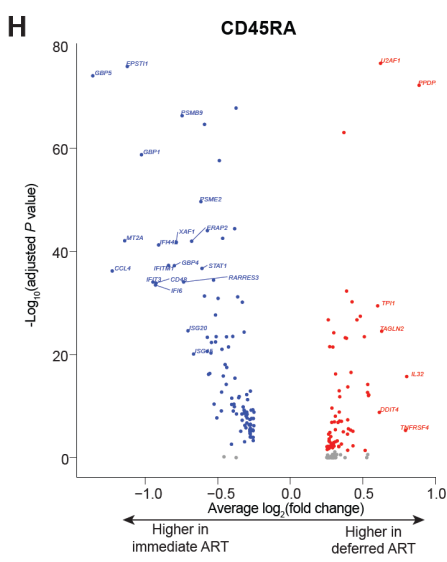
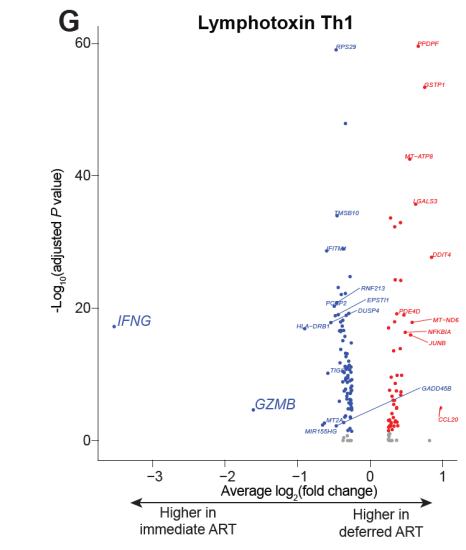
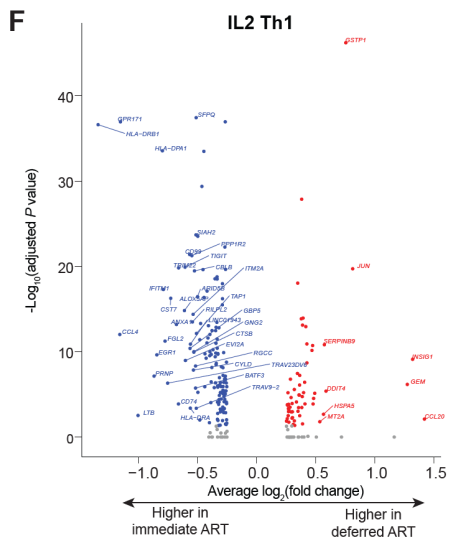
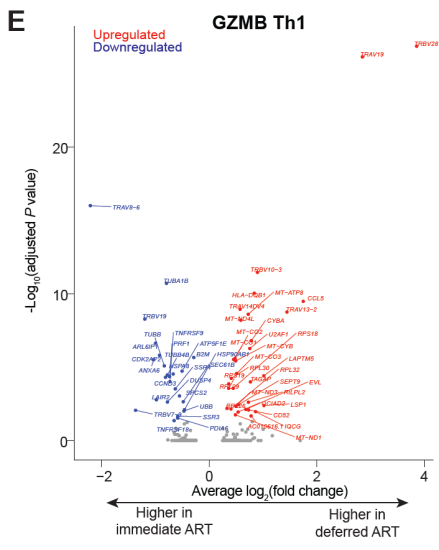
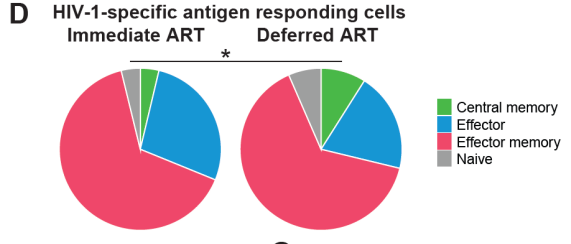
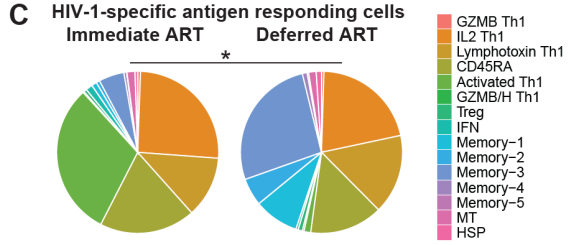
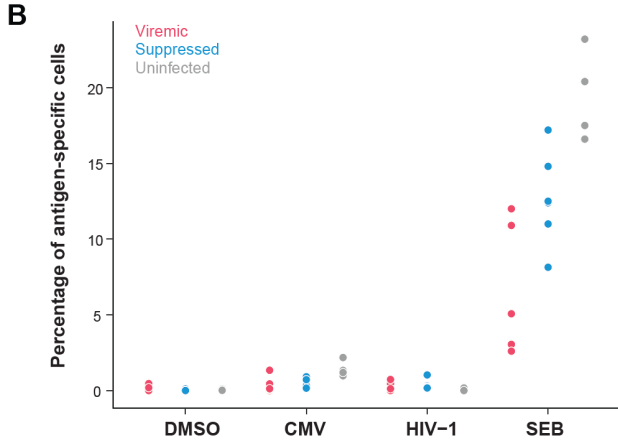
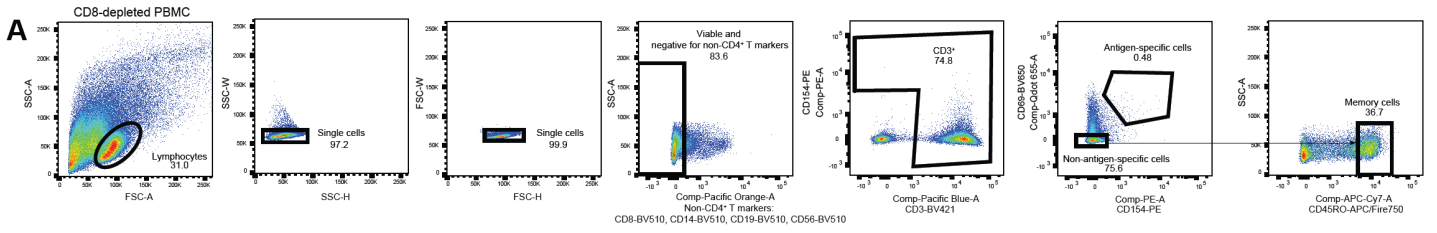


Figure S4. HIV-1-specific antigen responses. Related to Figure 3.

A, CD8⁺ T cells were depleted from PBMC using magnetic depletion before antigen stimulation. CD8-depleted PBMC were gated on flow cytometry to select lymphocytes, single cells, cells that are not CD4⁺ T cells (CD8, CD14, CD19, CD56 negative), and CD3 positive. Antigen-specific cells were defined by CD69 and CD154 double positive expression. Memory cells were defined as CD69 and CD154 double negative and CD45RO positive populations. Representative plots from participant 910 HIV-1 in antigen stimulated condition were shown. **B**, The frequency of CMV-specific and HIV-1-specific cells. SEB, Staphylococcal enterotoxin B. **C**, Cluster distribution and **D**, Memory distribution differences between suppressed immediate and suppressed delayed individuals. Differential expressional analysis comparing deferred suppression to immediate suppression cells which are HIV-1 specific within **E**, GZMB Th1, **F**, IL2 Th1, **G**, Lymphotoxin Th1, **H**, CD45RA, **I**, Activated Th1, and **J**, GZMB/H Th1 cell clusters. **C** and **D**, *, $P < 0.05$, Fisher's exact test. **E–J**, Wilcoxon Rank-Sum test with Benjamini-Hochberg Procedure multiple hypothesis correction.

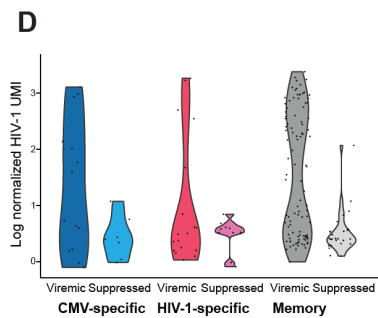
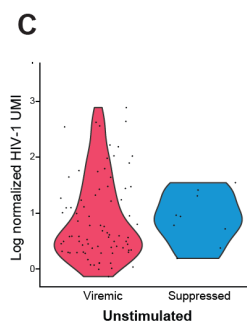
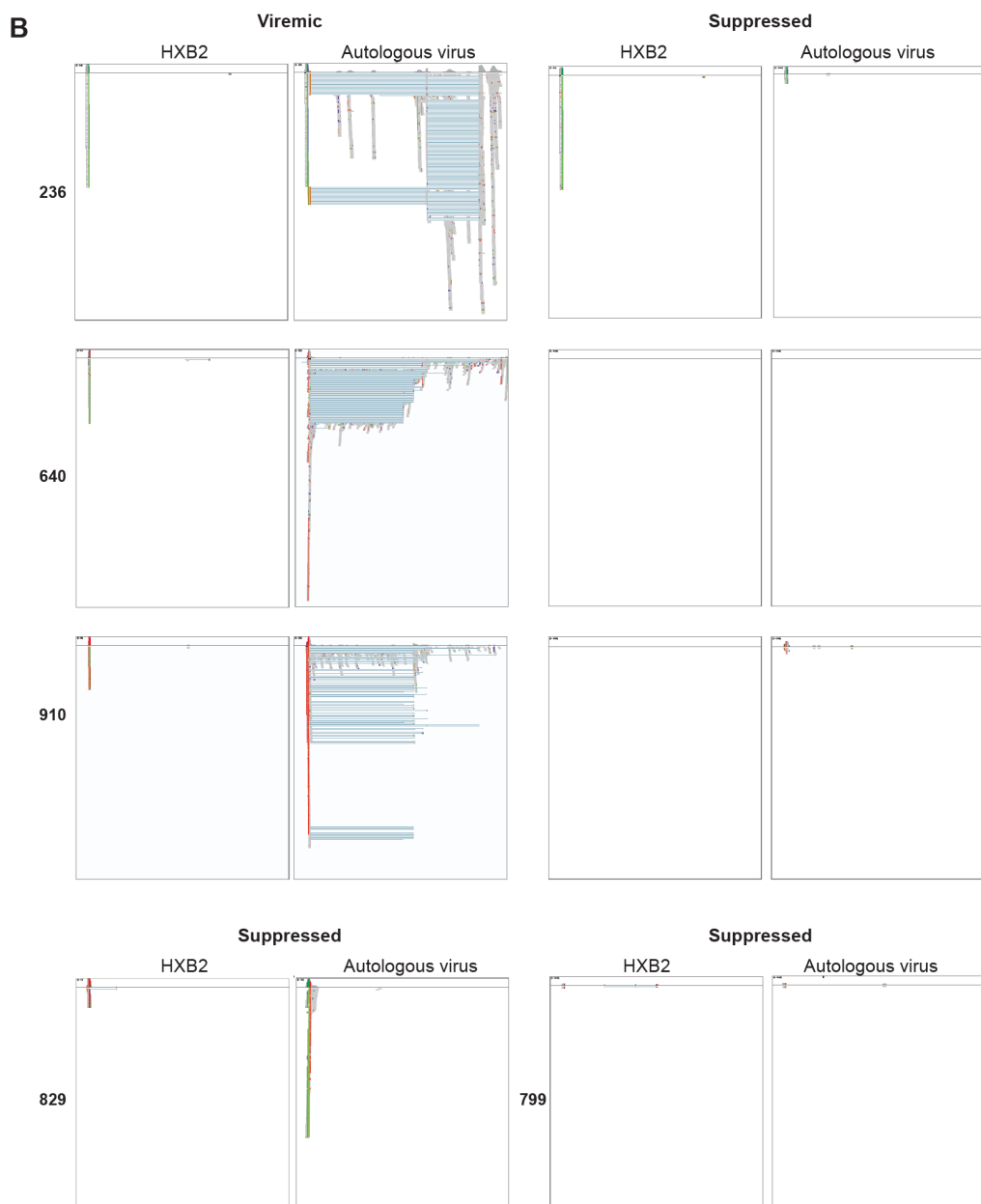
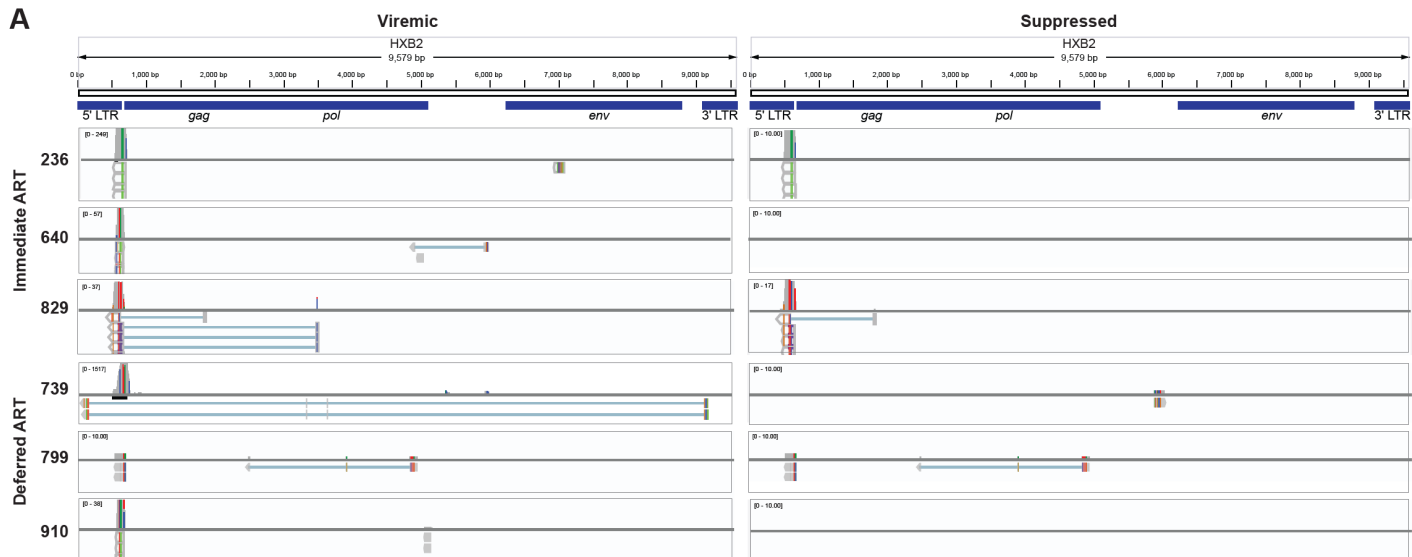


Figure S5. HIV-1 RNA mapping and expression level. Related to Figure 4.

Integrative genomics viewer (IGV) plots of **A**, all six participants to HXB2 both during viremia and suppression and **B**, alignments of select participants comparing HXB2 alignments to alignments to infected individual autologous viral sequences. Violin plot showing the level of HIV-1 RNA expression in HIV-1 RNA⁺ cells in the unstimulated condition during **C**, viremia and after viral suppression and **D**, in antigen stimulated condition during viremia and after viral suppression.

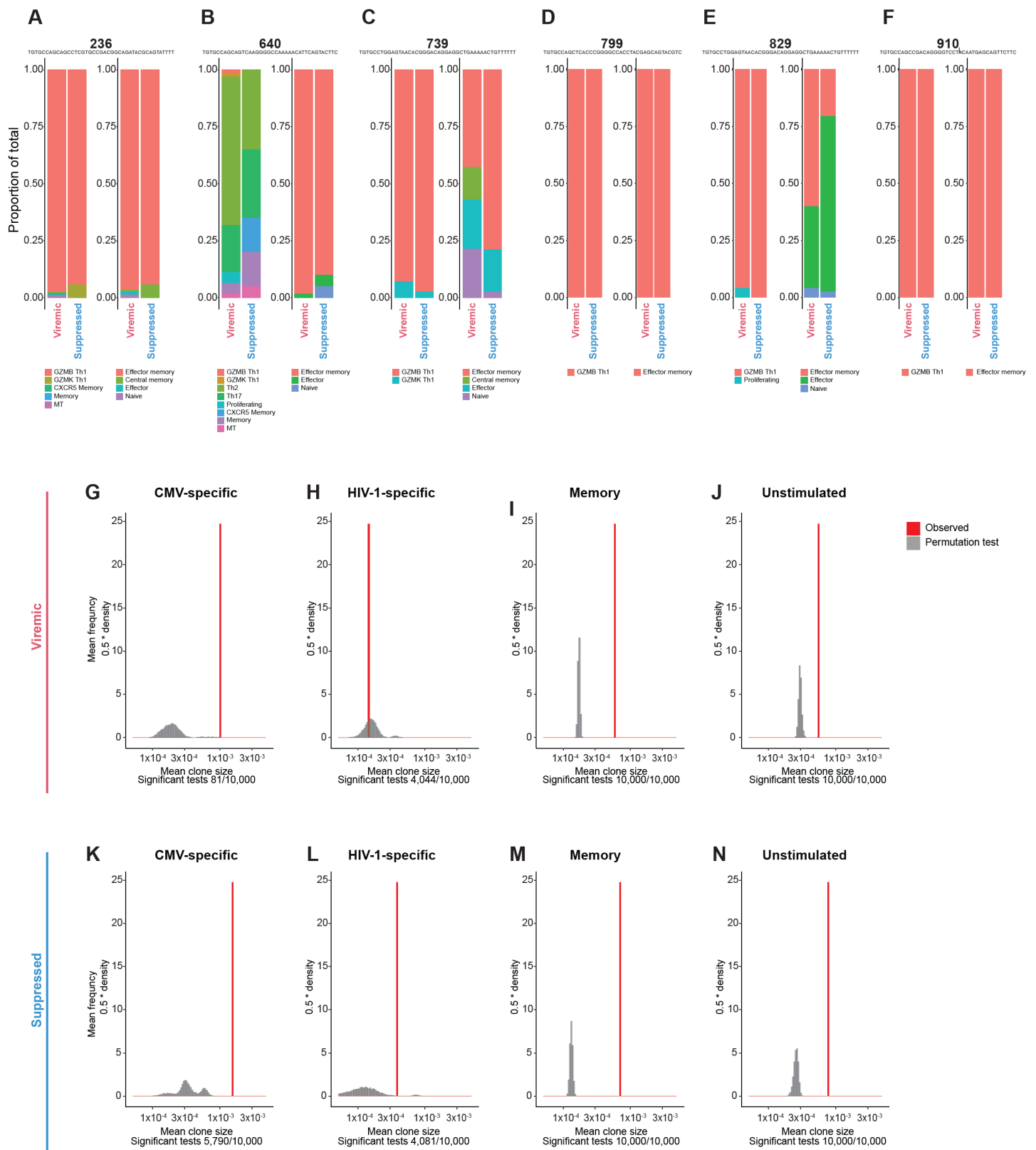


Figure S6. Clone phenotypes and the size of HIV-1 RNA⁺ clones vs HIV-1 RNA⁻ clones. Related to Figure 5 and Figure 6.

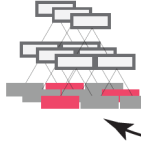
A–F, stacked bar graphs showing the differences in cluster membership and memory phenotype between clone members in viremia and suppression in six example clones. Histograms showing the permutation mean clone size (grey) and the observed HIV-1 RNA⁺ clone mean size (red) for **A**, CMV-specific clones during viremia, **B**, HIV-specific clones during viremia, **C**, clones for the sorted memory

condition during viremia, **D**, clones from the unstimulated condition during viremia, **E**, CMV-specific clones during viral suppression, **F**, HIV-specific clones during viral suppression, **G**, clones for the sorted memory condition during viral suppression, and **H**, clones from the unstimulated condition during viral suppression, $P < 0.05$ in a one-sided (greater) Wilcoxon rank sum test was considered significant.

A Start with gene expression and labels

HIV-1 RNA⁺ T cell clones HIV-1 RNA⁻ T cell clones

Train random forest classifier

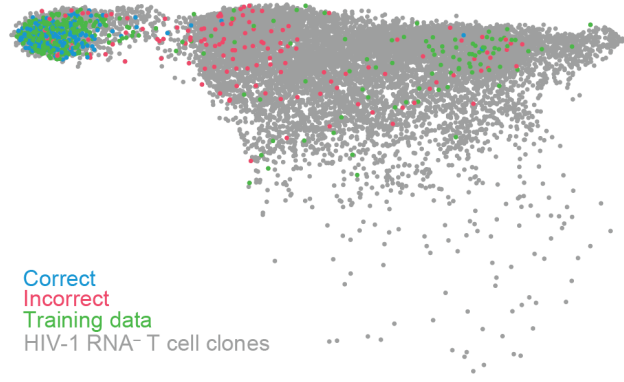


Score genes on usefulness

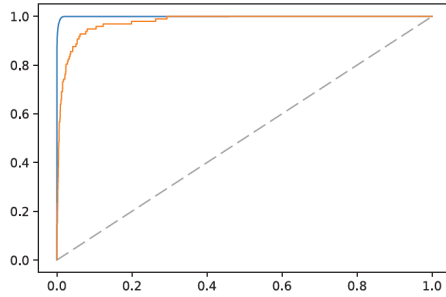
GZMB	0.5
POLR2A	0.2
GAPDH	0
NKG7	0.4

Eliminate least useful genes
Repeat until a minimum number of genes are identified that are necessary and sufficient

B



C



0.9996 Training data AUROC
0.9777 Test data AUROC
Random chance

D

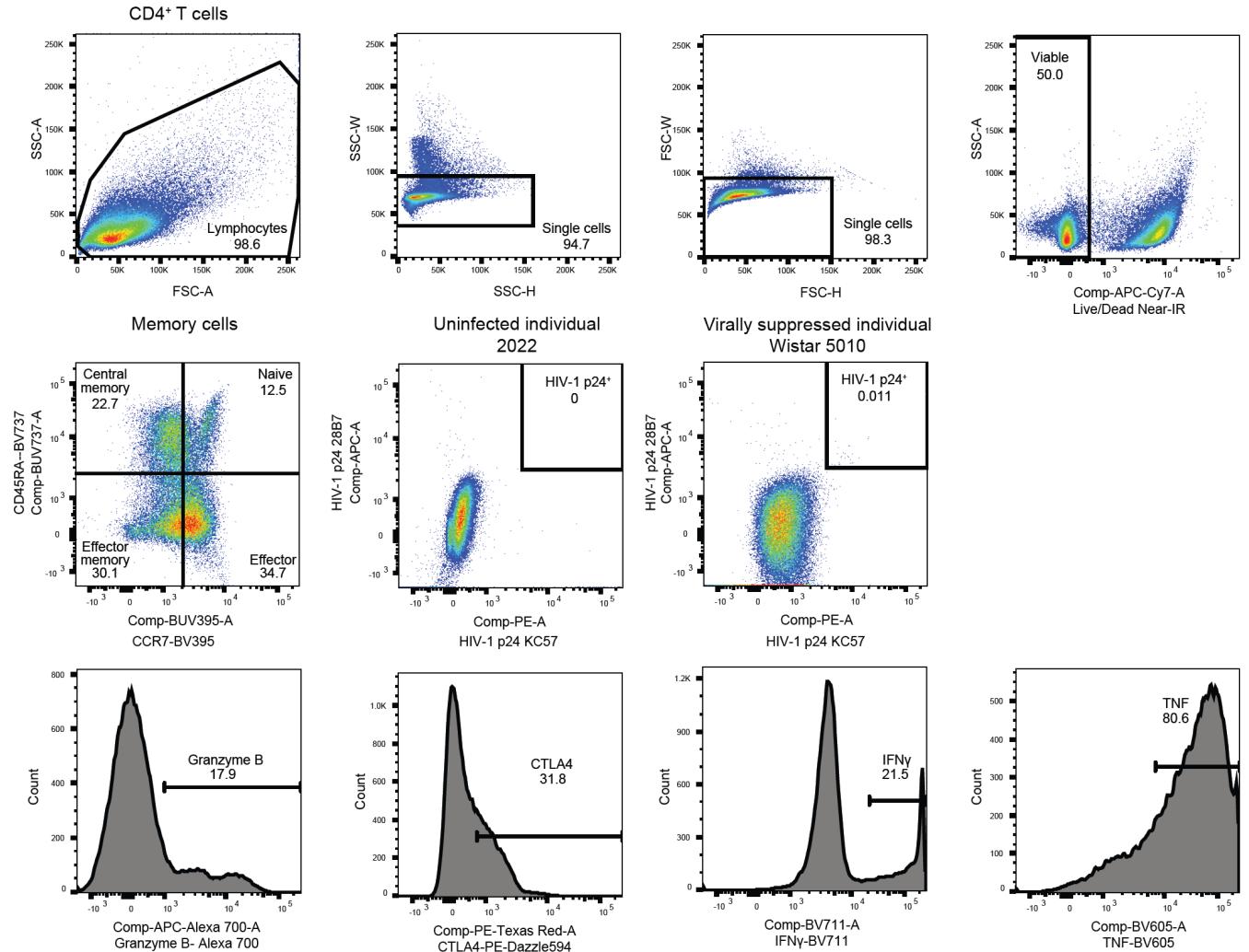


Figure S7. Machine learning identification of cellular markers necessary and sufficient to distinguish HIV-1 RNA⁺ T cell clones from HIV-1 RNA⁻ T cell clones by scRFE and flow cytometry validation for HIV-1 p24⁺ cells. Related to Figure 7.

A, Schematic representation of the process employed for scRFE to identify a subset of genes that were necessary and sufficient to differentiate HIV-1 RNA⁺ T cell clones from HIV-1 RNA⁻ T cell clones. **B**, UMAP plots of clonal cells from HIV-1-infected individuals in the unstimulated condition showing training data (green), cells in the test data correctly identified as HIV-1 RNA⁺ T cell clone (blue), and cells in the test data incorrectly identified as HIV-1 RNA⁻ T cell clone (red). **C**, Receiver operator curve (ROC) of model performance on the training dataset (blue) and the test dataset (orange). **D**, CD4⁺ T cells were isolated from PBMC using magnetic negative selection. CD4⁺ T cells were gated on flow cytometry to select lymphocytes, single cells, and viable cells (Live/Dead near-IR negative). Flow cytometric gating of memory populations (defined by CCR7 and CD45RA expression), HIV-1 p24, granzyme B, CTLA4, IFN γ , and TNF. Gating for p24⁺ cells in an uninfected individual (2022) and in an HIV-1-infected individual (Wistar 5010) was shown.

Table S1. Clinical characteristics of study participants, related to Figure 1.

Participant ID	Study arm	Age	Sex	Ethnicity	ART	Peak viral load (copies/ml)	Duration since EDDI at the viremic time point (days)	Duration from EDDI to ART (days)	Duration from EDDI to suppressed time point (days)	Duration of viral suppression at the viral suppression time point (days)	CD4 count at the viremic time point (/μl)	CD4 count at the viral suppression time point (/μl)
Sabes												
236	Immediate	22	M	Hispanic	FTC/TDF/EFV	7,117,757	24	22	431	384	625	712
640	Immediate	26	M	Hispanic	EVG/c/TDF/FTC	6,023,064	22	22	336	172	586	772
829	Immediate	21	M	Hispanic	EFV/TDF/FTC	1,205,138	48	47	329	245	388	796
739	Delay	23	M	Hispanic	EFV/TDF/3TC	8,604,710	41	207	342	256	467	821
799	Delay	31	M	Hispanic	EFV/TDF/3TC	13,313,300	20	187	329	245	1,203	1,464
910	Delay	24	M	Hispanic	EFV/AZT/3TC	1,257,674	33	198	338	254	375	371
Wistar												
5004	Long-term ART	46	M	White	ABC/DTG/3TC	NA	NA	NA	NA	3,003	NA	737
5005	Long-term ART	41	M	AA	TDF, ABC/DTG/3TC	NA	NA	NA	NA	1,399	NA	498
5007	Long-term ART	40	M	AA	EVG/c/FTC/TAF	NA	NA	NA	NA	5,480	NA	784
5008	Long-term ART	35	M	AA	BIC/FTC/TAF	NA	NA	NA	NA	1,235	NA	797
5009	Long-term ART	62	M	AA	BIC/FTC/TAF	NA	NA	NA	NA	2,052	NA	583
5010	Long-term ART	53	M	AA	BIC/FTC/TAF	NA	NA	NA	NA	1,234	NA	569
5011	Long-term ART	43	M	AA	DTG/3TC	NA	NA	NA	NA	1,444	NA	667
5012	Long-term ART	33	M	AA	EVG/c/FTC/TAF	NA	NA	NA	NA	2,189	NA	1,322
Uninfected												
2004	Uninfected	55	M	AA	NA	NA	NA	NA	NA	NA	NA	NA
2022	Uninfected	59	M	AA	NA	NA	NA	NA	NA	NA	NA	NA
2023	Uninfected	47	M	AA	NA	NA	NA	NA	NA	NA	NA	NA

*AA African American; 3TC, lamivudine; ABC, abacavir; AZT, azidothymidine; BIC, bictegravir; /c, cobicistat; DTG, dolutegravir; EFV, efavirenz; EVG, elvitegravir; FTC, emtricitabine; TAF, tenofovir alafenamide; TDF, tenofovir disoproxil; EDDI, estimated dates of detectable infection; NA, not available.