## SUPPLEMENTAL INFORMATION

## for the article

## Efficient discrimination against RNA-containing primers by human DNA polymerase $\epsilon$

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Tahir H. Tahirov Email: <u>ttahirov@unmc.edu</u> Table S1. Oligonucleotides used in this study

Sequence	Description	Application	Length
5'-AATGTTTCTAGGCAGCTCGGAGTCC a	Template (DNA)	kinetic studies	25
5'-/Cy3/GGACTCCGAGCTGCC (D15)	Primer (DNA)		15
5'-/Cy3/ <u>GGACTCCGAGCTGC</u> C <sup>b</sup> (D14-R1)	Primer (chimeric)		15
5'-/Cy3/GGACUCCGAGCUGCC (R15)	Primer (RNA)	kinetic studies	15
5'-/Cy3/GGACUCCGAGCUGC <u>C</u> (R14-D1)	Primer (chimeric)		15
5'-/Cy3/GGACUCCGAGCU <u>GCC</u> (R12-D3)	Primer (chimeric)		15
5'-/Cy3/GGACUCCGAG <u>CTGCC</u> (R10-D5)	Primer (chimeric)		15
5'-/Cy3/GGACUCCG <u>AGCTGCC</u> (R8-D7)	Primer (chimeric)		15
5'-/BiotinTEG/AATACATAAGCGCTCCAGGCAAT	Template (DNA)	Octet K2	23
5'-GCCUGGAGCGC (R11)	Primer (RNA)		11
5'-GCCUGGAGCG/ddC/ (R10-D1)	Primer (chimeric)		11
5'-GCCUGGAG <u>CG/ddC/</u> (R8-D3)	Primer (chimeric)		11
5'-GCCUGG <u>AGCG/ddC/</u> (R6-D5)	Primer (chimeric		11
5'-GCCU <u>GGAGCG/ddC/</u> (R4-D7)	Primer (chimeric)		11
5'-GCCTGGAGCG/3ddC/ (D11)	Primer (DNA)		11
5'-GCCTGGAGCG/3dC/ (D10-R1)	Primer (chimeric)		11
5'-ATTATGGCAGCTCGGAGTCC	Template (DNA)	EMGA	20
5'-/Cy3/GGACTCCGAGCTGCC (D15)	Primer (DNA)	EMSA	15

<sup>a</sup> The template regions complementary to a primer are in italics.
<sup>b</sup> Deoxy- and dideoxy- nucleotides in the chimeric primer sequences are underlined.



**Figure S1.** Analysis of hPoleco purity. Proteins were separated by 8% SDS-PAGE and stained by Coomassie Brilliant Blue R-250. M – molecular weight markers. The dashed line indicates splicing of the original image.

Homo sapiens	832	AG <mark>I</mark> VCFTGAN <mark>IITQARELIEQIGR</mark> PLELDTDGIWC <mark>VLPNSFPENFVF</mark> KTTNVKK	885
Mus musculus	832	AG <mark>I</mark> VCFTG <mark>ANIITQARELIEQIGR</mark> PLELDTDGIWC <mark>VLPNSFPENF</mark> VIKTTNAKK	885
Drosophila melanogaster	830	AG <mark>IVCLTGSNIITKAREIIERVGR</mark> PLELDTDGIWC <mark>ILP</mark> GSFPQEFT <mark>I</mark> HTSHEKK	883
Arabidopsis thaliana	805	AG <mark>VVTYTGA</mark> KIIQNARLLIERIGKPLELDTDGIWCCLPGSFPENFTFKTIDMKK	858
Aspergillus nidulans	816	AG <mark>V</mark> TCL <mark>TGARIIQMARELVERIGR</mark> PLELDTDGIWCMLP <mark>GTFPENFSF</mark> TLKNGKK	869
Saccharomyces cerevisiae	847	AG <mark>I</mark> TCL <mark>TGA</mark> TIIOMARALVERVGRPLELDTDGIWCILPKSFPETYFFTLENGKK	900

**Figure S2. Thr861 of hPole is conserved.** Amino acid sequence alignment was performed using ClustalW. The conserved and similar residues are highlighted with red and yellow, respectively. The conserved threonines are boxed.



Figure S3. Kinetics of hPoleco/DNA complex formation and dissociation in the presence of 50  $\mu$ M dTTP (*A*) or 5 mM MgCl<sub>2</sub> (*B*). Binding studies were conducted in the buffer containing 30 mM Tris-Hepes, pH 7.8, 100 mM NaCl, 2 mM TCEP, and 0.002% Tween 20. Enzyme concentration is indicated on each graph. DNA is attached to the streptavidin coated biosensor (SAX, Sartorius AG). Each binding cycle consists of next steps: baseline, association, and dissociation. The blue line depicts the actual binding curve. The red line represents the model curve build by the Data Analysis HT software (Sartorius AG) with use of global fitting.







Supp. Fig. S1.