Supplementary Text

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Table A. *C. albicans* strains used in this study.

C. albicans strain name	Parent	Genotype	Strain background /construction	Reference
SC5314		Wild type		[1]
SN95		*arg4Δ/arg4Δ his1Δ/his1Δ IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*		[2]
JKC917	SN95	*hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*		[3]
JKC1361	JKC917	*hisΔ1/his1Δ:: tetR-FRT arg4/ARG4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC917 transformed with PCR product of fjk1184 & rjk1186 using Candida cDNA library as template	[3]
JKC1713	JKC917	*hisΔ1/his1Δ:: tetR-FRT arg4/ARG4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC917 transformed with PCR product of fjk1184 & rjk1186 using Candida cDNA library as template	This work
JKC1347	JKC917	*tor1::ARG4/TOR1 hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC917 transformed with PCR product of fjk1142 & rjk1143 using pFA-ARG4 as template	[3]
JKC1345	JKC917	*tor1::ARG4/TOR1 hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	*tor1/TOR1* heterozygote constructed like JKC1347, independent isolate.	This work
JKC1346	JKC917	*tor1::ARG4/TOR1 hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	*tor1/TOR1* heterozygote constructed like JKC1347, independent isolate.	This work
JKC1441	JKC1347	*tor1::ARG4/tor1::FRT-tetO-TOR1-Del381 hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC1347 transformed with StuI/NcoI digested pJK1189 to have *TOR1-Del381* under *tetO (OFF)* promoter, after inducing FLP	[3]
JKC1442	JKC1345	*tor1::ARG4/tor1::FRT-tetO-TOR1-Del381 hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	*tor1/tetO-TOR1-Del381* constructed like JKC1441, distinct lineage from JKC1345.	This work
JKC1445	JKC1346	*tor1::ARG4/tor1::FRT-tetO-TOR1-Del381 hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	*tor1/tetO-TOR1-Del381* constructed like JKC1441, distinct lineage from JKC1346.	This work
JKC1549	JKC1347	*tor1::ARG4/tor1::FRT-tetO-TOR1-FL hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC1347 transformed with StuI/NcoI digested pJK1236 to have *TOR1-FL* under *tetO (OFF)* promoter, after inducing FLP	[3]
JKC1543	JKC1345	*tor1::ARG4/tor1::FRT-tetO-TOR1-FL hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	*tor1/tetO-TOR1-FL* constructed like JKC1549, distinct lineage from JKC1345.	This work
JKC1546	JKC1346	*tor1::ARG4/tor1::FRT-tetO-TOR1-FL hisΔ1/his1Δ:: tetR-FRT arg4/arg4 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	*tor1/tetO-TOR1-FL* constructed like JKC1549, distinct lineage from JKC1346.	This work
TETG25B	CAI4	*ADH1/adh1::tetO(ON)-GFP*	CAI4 transformed with pTET25 tetracycline inducible GFP cassette	[4]
JKC2616	JKC1713	*hisΔ1/his1Δ:: tetR-FRT arg4/ARG4 MAL2/pMAL2-GFP-FRT-pMAL2-MAL2 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC1713 transformed with BsrGI digested pJK1489 to have GFP under *pMAL2* promoter, after inducing FLP	This work
JKC2620	JKC1347	*tor1::ARG4/tor1::FRT-tetO-TOR1-Del381 hisΔ1/his1Δ:: tetR-FRT arg4/arg4 MAL2/pMAL2-GFP-FRT-pMAL2-MAL2 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC1347 transformed with BsrGI digested pJK1489 to have GFP under *pMAL2* promoter, after inducing FLP	This work
JKC2624	JKC1441	*tor1::ARG4/tor1::FRT-tetO-TOR1-Del381 hisΔ1/his1Δ:: tetR-FRT arg4/arg4 MAL2/pMAL2-GFP-FRT-pMAL2-MAL2 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC1441 transformed with BsrGI digested pJK1489 to have GFP under *pMAL2* promoter, after inducing FLP	This work
JKC2628	JKC1549	*tor1::ARG4/tor1::FRT-tetO-TOR1-FL hisΔ1/his1Δ:: tetR-FRT arg4/arg4 MAL2/pMAL2-GFP-FRT-pMAL2-MAL2 IRO1/iro1Δ::λimm^{434} URA3/ura3Δ::λimm^{434}*	JKC1549 transformed with BsrGI digested pJK1489 to have GFP under *pMAL2* promoter, after inducing FLP	This work

Table B. Plasmids used in this study.

Plasmid	Description	Source (Reference)
pJK1000	*FLP-NAT1 tetO-PES1* construct, vector backbone is pLitmus28 (New England Biolabs)	[5]
pJK1027	pAU34 with *URA3* disrupted by $Ag_{promoter}$*TEF1-NAT1-*$Ag_{terminator}$*TEF1*	[6,7]
pJK1189	*FLP-NAT1 tetO-TOR1-Del381* construct, derived from pJK1000.	[3]
pJK1236	*FLP-NAT1 tetO-TOR1-FL* construct, derived from pJK1000.	[3]
pAU15	*pMAL2* expression vector	[6]
pGFP-HIS1	*GFP-HIS1* fusion construct	[8]
pJK1482	*pMAL2-GFP* construct, derived from pAU15. Product of fjk2036 & rjk1633 using pGFP-*HIS1* as template was ligated into pAU15 using SalI/XmaI sites.	This work
pJK1489	*FLP-NAT1 pMAL2-GFP* construct, derived from pJK1482. *URA3* marker in pJK1482 was replaced with the '*FLP-NAT1*' cassette by blunt cloning.	This work

Table C. Oligonucleotides used in this study.

Primer name	Purpose	Sequence 5' to 3' (lower cases - restriction enzyme recognition sites)
fjk2062	Forward primer for *TOR1* quantitative real-time PCR	GCTTAGTTTTATCAGGCAAGGGA
rjk2063	Reverse primer for *TOR1* quantitative real-time PCR	ACTCATCCCCGTGTCTCTAG
fjk1400	Forward primer for *ACT1* quantitative real-time PCR	TGGTGATGGTGTTACTCACG
rjk1401	Reverse primer for *ACT1* quantitative real-time PCR	GACAATTTCTCTTTCAGCAC
fjk1184	Forward primer to amplify the *ARG4* marker	GAATCCACAATCGTATATGAAC
rjk1186	Reverse primer to amplify the *ARG4* marker	GAATATAGTGATGATGAGGATG
fjk1185	Forward primer to confirm the 5'end integration of *ARG4* marker in *Candida* strains.	GACATATTGACCGACATAATTC
rjk1187	Reverse primer to confirm the 5'end integration of *ARG4* marker in *Candida* strains	GTCGTTTCACCGGTGCCACTG
fjk1188	Forward primer to confirm the 3'end integration of *ARG4* marker in *Candida* strains	CAGTACCACCAATAGCATCTC
rjk1199	Reverse primer to confirm the 3'end integration of *ARG4* marker in *Candida* strains	GGTAGTCTCCGATTATGATTC
fjk2036	Forward primer to amplify GFP sequence.	CCTGCTgtcgacATGTCTAAAGGTGAAGAATTAT
rjk1633	Reverse primer to amplify GFP sequence and to confirm the 5'end integration of *pMAL2-GFP* in *Candida* strains	GCAGCTcccgggTTATTTGTATAATTCATCCATACCATGG
fjk2045	Forward primer to confirm the 5'end integration of *pMAL2-GFP* in *Candida* strains	CATTGTGTTGAGCTGCGACT
fjk1517	Forward primer to confirm the 3'end integration of *pMAL2-GFP* in *Candida* strains	GGAATTGTGAGCGGATAAC
rjk2046	Reverse primer to confirm the 3'end integration of *pMAL2-GFP* in *Candida* strains	CAAGGTCCCGTATTTGCTGT

Table D. Antibodies used in this study.

Purpose	Antigen recognized	Species	Source or Reference
primary	P-Mkc1	rabbit	Cell Signaling Technology, #4370P
primary	P-S6	rabbit	Cell Signaling Technology, #9611L
primary	S6	sheep	R&D Systems, #AF5436
primary	P-Hog1	rabbit	Cell Signaling Technology, #4511S
primary	P-eIF2a	rabbit	Cell Signaling Technology, #3597S
primary	GFP	mouse	Roche, #11814460001
loading control	PSTAIRE (Cdc2)	rabbit	Santa Cruz Biotechnology, #sc-53
loading control	Tubulin	rat	Abcam, #ab6161
secondary	Rabbit IgG	goat	Cell Signaling Technology, #7074S
secondary	Sheep IgG	donkey	Santa Cruz Biotechnology, #sc-2473
secondary	Mouse IgG	horse	Cell Signaling Technology, #7076S
secondary	Rat IgG	goat	Abcam, #ab97057

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