

1

**Supplementary Information for**

2

3 **Ecological networks of dissolved organic matter and microorganisms under global**

4

**change**

5

6 **The file includes:**

7

Supplementary Methods

8

Supplementary Tables (S1–S5)

9

Supplementary Figures (S1–S21)

10

## 11 **Supplementary Methods**

### 12 **Considerations of the experimental design**

13 In our study, we took advantage of elevational temperature gradients as a natural  
14 experiment. Natural climatic gradients along latitudinal and elevational gradients have  
15 played a pivotal role in testing the importance of climatic factors on biological  
16 community composition in the history of biogeography not only for macroorganisms, but  
17 also for microorganisms and biogeochemical cycling, and experimental manipulations  
18 along elevational or biogeographical gradients have been recognised as an important tool  
19 for disentangling different underlying drivers on community structure and ecosystem  
20 processes. For instance, De Sassi *et al.* <sup>1</sup> used a natural temperature gradient along  
21 elevations, combined with experimental nitrogen fertilization, to investigate the effects of  
22 elevated temperature and globally increasing anthropogenic nitrogen deposition on the  
23 structure and phenology of a grassland herbivore assemblage. Thus, using manipulative  
24 experiments along broad environmental gradients can help tease apart the relative  
25 importance of the interactive effects of local-scale factors and broad-scale climatic  
26 variation in shaping biological communities and ecosystem processes.

27 Temperature generally correlates strongly negatively with elevation, which was  
28 also true in our study showing significantly negatively correlation between measured  
29 water temperature and elevation in both China and Norway ( $R^2 = 0.98$ ,  $P < 0.05$ , and  $R^2 =$   
30  $0.96$ ,  $P < 0.05$ , respectively). Specifically, we used the common sediment from Taihu  
31 Lake at each elevation with ten nutrient levels, and microbes from the local species pool  
32 of each elevation could be inoculated into the common sediment. As such, the covariance  
33 between climate and natural environments such as nutrients could be eliminated, which  
34 could disentangle the independent effects of temperature versus nutrient enrichment and  
35 also their interactive effects on the associations between DOM and microbes. Similar  
36 achievements are also reported in previous literature by experiments on mountainsides.  
37 For instance, in a translocation experiment, microbial decomposers were deliberately  
38 inoculated onto common leaf litter to disentangle the effects of microbial community  
39 versus temperature on litter decomposition rates and reevaluate the role of microbial  
40 community composition in its decomposition responses to climate <sup>2</sup>.

41           However, for in-situ experiments, it is inevitable that there are some unmeasured  
42 environmental variables which might have some effects on biological communities and  
43 ecosystem processes. This will not be a major problem if our main focus was to  
44 investigate the major environmental variables which are playing pivotal roles in  
45 structuring biological and organic carbon compositions. Our data support that  
46 temperature and nutrients were key factors in explaining chemodiversity and bacterial  
47 community composition<sup>3</sup>. In our study, we considered as many important environmental  
48 variables as possible, such as chlorophyll *a*, bacterial abundance and nutrients. Due to  
49 practical and logistical issues, we were unable to measure all potential variables, such as  
50 UV radiation, across all elevations. This is largely because we have no clear evidence that  
51 these other variables have a dominant role in affecting the sediment bacterial and DOM  
52 compositions we studied here. More importantly, the bottom of our microcosm was  
53 buried into the local soils by 10% of the bottle height, and so sediments of each bottle  
54 were below the ground surface. It is thus unlikely that UV radiation and the related  
55 photodegradation would have stronger effects on bacteria and DOM in the sediments than  
56 temperature and nutrients.

57           In addition to relying on natural temperature gradients, controlled temperature  
58 experiments are another option to study climate effects on biological communities or  
59 organic matter decomposition. So far, there are two primary tools for simulating warming:  
60 passive greenhouses and active heating devices including soil, water and aerial arrays.  
61 Generally, controlling water temperatures in the field will encounter serious practical  
62 issues on mountainsides along large geographical regions. For instance, to prevent  
63 malfunction in a long-term run, heating devices are needed to be checked and replaced  
64 regularly, which is not possible in remote mountains and high elevations as in our current  
65 study. To obtain desired temperatures is also not easy in the field due to the variations  
66 such as in wind velocities, sun shine and forest shading along elevations. We do not see  
67 clearly how to practically and efficiently control for water temperatures in such a large-  
68 scale field study, and thus relied on natural temperature gradients.

69           It should be noted that our aims were considered to be as realistic as possible by  
70 combining field experiments and natural gradients, and by considering only the most

71 important and generally most interesting factors (that is, nutrients and temperature) for  
72 robust findings. Compared to field experiments, researchers can much more easily control  
73 virtually all aspects important for biological communities and organic matter in  
74 laboratory experiments. But laboratory experiments can be also easily criticized by some  
75 researchers to be far from realistic and results may not be applicable for real natural  
76 communities <sup>4</sup>. Different from laboratory manipulations, we think that these experiments  
77 are closer to nature environments, and expect such experiments to bridge between  
78 laboratory experiments and field observations. The communities in the microcosms  
79 should be dynamically driven by ecological processes, such as species dispersal, growth  
80 and extinction, and a time-series sampling would be ideally helpful to capture these  
81 processes. However, considering the harsh field situations in remote mountains or high  
82 elevations, the nature of destructive sampling, the large sample volumes required for  
83 repeat sampling, and the limited resources, it is unlikely we could have sampled  
84 biological communities and DOM at more than one time-point. As a tradeoff among  
85 logistics issue, heavy workload and robust results, we only sampled once after a one-  
86 month field incubation, and are confident that a final number of 300 DOM or bacterial  
87 samples across two mountains, 10 elevations and 10 nutrient levels was large enough for  
88 solid statistical analyses. We welcome future studies to undertake more intensive  
89 experiments if more resources could be allocated.

90

91

92 **Supplementary Tables**

93

94 **Table S1.** Variables used to explain the specialization of DOM-bacteria interactions.

95 These variables were considered based on the hypothetical casual relationships in Fig. 6a:

96 climate change, human impacts and contemporary nutrient variables as distal drivers, and

97 energy supply, biodiversity, chemodiversity and DOM traits as proximal drivers. NMDS:

98 non-metric multidimensional scaling.

Drivers	Group	Subgroup	Variable	Description	
Distal drivers	Environment	Climate change	Temp	Water temperature <sup>#1</sup>	
		Human impacts	ADD.NO <sub>3</sub>	Nutrient enrichment <sup>#2</sup>	
		Contemporary nutrient	TN.sedi	Sediment total nitrogen (TN)	
		Contemporary nutrient	TP.sedi	Sediment total phosphorus (TP)	
		Contemporary nutrient	NO <sub>x</sub> .sedi	Sediment NO <sub>x</sub> <sup>-</sup>	
		Contemporary nutrient	NO <sub>2</sub> .sedi	Sediment NO <sub>2</sub> <sup>-</sup>	
		Contemporary nutrient	NH <sub>4</sub> .sedi	Sediment NH <sub>4</sub> <sup>+</sup>	
		Contemporary nutrient	PO <sub>4</sub> .sedi	Sediment PO <sub>4</sub> <sup>3-</sup>	
		Contemporary nutrient	NO <sub>3</sub>	Water NO <sub>3</sub> <sup>-</sup>	
		Contemporary nutrient	NO <sub>2</sub>	Water NO <sub>2</sub> <sup>-</sup>	
		Contemporary nutrient	NH <sub>4</sub>	Water NH <sub>4</sub> <sup>+</sup>	
Contemporary nutrient	PO <sub>4</sub>	Water PO <sub>4</sub> <sup>3-</sup>			
Proximal drivers	Energy supply	Energy supply	TC.sedi	Sediment total organic carbon (TOC)	
		Energy supply	DOC.sedi	Sediment dissolved organic carbon (DOC)	
		Energy supply	pH	Water pH	
		Energy supply	Chla.sedi	Sediment Chlorophyll <i>a</i> (Chl <i>a</i> )	
	Diversity	Biodiversity	bac.rich	Species richness of bacteria	
		Biodiversity	bac.nmids1	NMDS axis 1 of bacterial composition	
		Biodiversity	bac.nmids2	NMDS axis 2 of bacterial composition	
		Chemodiversity	chemo.rich	Chemical richness of molecules	
		Chemodiversity	mol.nmids1	NMDS axis 1 of molecular composition	
	Molecular traits	Molecular weight	Chemodiversity	mol.nmids2	NMDS axis 2 of molecular composition
			Molecular weight	Mass	The mass to charge ratio (m/z)
			Molecular weight	C	The number of carbon
		Molecular weight	kdefect <sub>CH2</sub>	Kendrick Defect	
		Stoichiometry	O/C	O/C ratio	
		Stoichiometry	H/C	H/C ratio	
Stoichiometry		N/C	N/C ratio		
Stoichiometry		P/C	P/C ratio		
Stoichiometry		N/P	N/P ratio		
Stoichiometry	S/C	S/C ratio			

	Chemical structure	AI <sub>Mod</sub>	The modified aromaticity index
	Chemical structure	DBE	Double bond equivalence
	Chemical structure	DBE <sub>O</sub>	Double bond equivalence minus oxygen
	Chemical structure	DBE <sub>AI</sub>	Double bond equivalence minus aromaticity index
	Oxidation state	GFE	Gibbs free energy
	Oxidation state	NOSC	Nominal oxidation state of carbon
	Carbon use efficiency	Y <sub>met</sub>	Carbon use efficiency

99 #<sup>1</sup> Water temperature was used to represent climatic variables due to its strong relations

100 with elevation in both mountain ranges (Wang *et al.* 2016).

101 #<sup>2</sup> Nitrate addition (ADD.NO<sub>3</sub>) was used to represent nutrient enrichment as the ratio

102 between nitrate and phosphorus in the initial overlying water was constant.

103

104 **Table S2.** Variables used to explain DOM features. These variables were categorised into  
 105 three groups: Environment (climate change, human impacts and contemporary nutrient),  
 106 energy supply and biodiversity. NMDS: non-metric multidimensional scaling.

Group	Subgroup	Variable	Description
Environment	Climate change	Temp	Water temperature
	Human impacts	ADD.NO <sub>3</sub>	Nutrient enrichment
	Contemporary nutrient	TN.sedi	Sediment total nitrogen (TN)
	Contemporary nutrient	TP.sedi	Sediment total phosphorus (TP)
	Contemporary nutrient	NO <sub>x</sub> .sedi	Sediment NO <sub>x</sub> <sup>-</sup>
	Contemporary nutrient	NO <sub>2</sub> .sedi	Sediment NO <sub>2</sub> <sup>-</sup>
	Contemporary nutrient	NH <sub>4</sub> .sedi	Sediment NH <sub>4</sub> <sup>+</sup>
	Contemporary nutrient	PO <sub>4</sub> .sedi	Sediment PO <sub>4</sub> <sup>3-</sup>
	Contemporary nutrient	NO <sub>3</sub>	Water NO <sub>3</sub> <sup>-</sup>
	Contemporary nutrient	NO <sub>2</sub>	Water NO <sub>2</sub> <sup>-</sup>
	Contemporary nutrient	NH <sub>4</sub>	Water NH <sub>4</sub> <sup>+</sup>
	Contemporary nutrient	PO <sub>4</sub>	Water PO <sub>4</sub> <sup>3-</sup>
	Energy supply	Energy supply	TC.sedi
Energy supply		DOC.sedi	Sediment dissolved organic carbon (DOC)
Energy supply		pH	Water pH
Energy supply		Chla.sedi	Sediment Chlorophyll <i>a</i> (Chl <i>a</i> )
Biodiversity	Biodiversity	bac.rich	Species richness of bacteria
	Biodiversity	bac.nm1	NMDS axis 1 of bacterial composition
	Biodiversity	bac.nm2	NMDS axis 2 of bacterial composition

108 **Table S3.** Formulae to calculate composite variables for structure equation models of the  
 109 specialization  $H_2'$  of DOM-bacteria interactions. We constructed four bipartite networks,  
 110 that is, the negative and positive interaction networks in China or Norway. The obtained  
 111 composite variables were used in Fig. 6. The abbreviations of included variables are  
 112 listed in Table S1.

Response	Region	Network type	Composite	Formula
$H_2'$	China	Negative	Contemporary nutrient	$-0.606 \times \text{NO}_3 + -0.222 \times \text{NO}_2 + 0.221 \times \text{PO}_4 + -0.294 \times \text{TN.sedi} + 0.184 \times \text{PO}_4.\text{sedi} + 0.296 \times \text{NH}_4.\text{sedi} + 0.219 \times \text{NO}_2.\text{sedi}$
			Energy supply	$-0.589 \times \text{pH} + 0.493 \times \text{DOC.sedi}$
			Biodiversity	$-0.278 \times \text{bac.nmids1} + 0.160 \times \text{bac.nmids2}$
			Chemodiversity	$-0.451 \times \text{chemo.rich} + 0.481 \times \text{mol.nmids1}$
			Molecular traits	$66.076 \times \text{Mass} + -64.710 \times \text{C} + -1.402 \times \text{AI}_{\text{Mod}} + -15.166 \times \text{DBE} + -20.154 \times \text{DBE}_O + 61.402 \times \text{DBE}_{\text{AI}} + -3.279 \times \text{GFE} + -23.908 \times \text{kdefect}_{\text{CH}_2} + -1.378 \times \text{O/C} + 2.627 \times \text{H/C} + 0.652 \times \text{N/P}$
$H_2'$	China	Positive	Contemporary nutrient	$0.339 \times \text{NO}_2 + -0.442 \times \text{PO}_4 + 0.443 \times \text{PO}_4.\text{sedi} + -0.173 \times \text{NO}_2.\text{sedi}$
			Energy supply	$0.525 \times \text{pH} + 0.253 \times \text{Chla.sedi} + -0.102 \times \text{TC.sedi} + -0.262 \times \text{DOC.sedi}$
			Biodiversity	$-0.293 \times \text{bac.rich} + 0.718 \times \text{bac.nmids1}$
			Chemodiversity	$-0.110 \times \text{chemo.rich} + -0.591 \times \text{mol.nmids1} + 0.416 \times \text{mol.nmids2}$
			Molecular traits	$-50.009 \times \text{Mass} + 48.984 \times \text{C} + 1.728 \times \text{AI}_{\text{Mod}} + 3.075 \times \text{DBE}_O + -26.770 \times \text{DBE}_{\text{AI}} + 3.904 \times \text{GFE} + 24.822 \times \text{kdefect}_{\text{CH}_2} + 1.204 \times \text{O/C} + -5.311 \times \text{H/C}$
$H_2'$	Norway	Negative	Contemporary nutrient	$-0.381 \times \text{NO}_3 + 0.365 \times \text{PO}_4.\text{sedi}$
			Energy supply	$0.210 \times \text{DOC.sedi}$
			Chemodiversity	$-0.359 \times \text{chemo.rich} + -0.186 \times \text{mol.nmids1} + -0.120 \times \text{mol.nmids2}$
			Molecular traits	$-1.316 \times \text{Mass} + 1.062 \times \text{AI}_{\text{Mod}} + 3.571 \times \text{DBE} + 2.524 \times \text{DBE}_O + -4.638 \times \text{DBE}_{\text{AI}} + 2.193 \times \text{H/C} + -0.387 \times \text{N/C} + -0.372 \times \text{N/P}$
$H_2'$	Norway	Positive	Contemporary nutrient	$-0.338 \times \text{NO}_3 + 0.280 \times \text{NO}_2 + 0.338 \times \text{PO}_4 + 0.350 \times \text{PO}_4.\text{sedi} + -0.105 \times \text{NO}_2.\text{sedi}$
			Energy supply	$-0.327 \times \text{pH} + 0.231 \times \text{TC.sedi} + -0.350 \times \text{DOC.sedi}$
			Biodiversity	$-0.510 \times \text{bac.nmids1}$
			Chemodiversity	$-0.169 \times \text{mol.nmids1} + -0.480 \times \text{mol.nmids2}$
			Molecular traits	$1.872 \times \text{Mass} + -1.825 \times \text{DBE} + 2.282 \times \text{DBE}_{\text{AI}} + -2.674 \times \text{GFE} + -1.431 \times \text{kdefect}_{\text{CH}_2} + 1.288 \times \text{H/C} + -0.877 \times \text{N/C} + 1.653 \times \text{P/C}$

113

114



115 **Table S4.** Summary of the model fit statistics evaluated for standardized structural  
 116 equation model (SEM). We explored the potential links between predictor variables and  
 117 the specialization  $H_2'$  of the negative and positive bipartite networks in China or Norway,  
 118 and the best-fitting models are shown in Fig. 6. We constructed the full SEM models  
 119 based on the hypothetical casual relationships (Fig. 6a), and further performed sequential  
 120 models by dropping non-significant paths from the full models.  $\chi^2$ : Chi-square.  $P$ : p-value  
 121 of chi-square test. df: Degrees of freedom. CFI: Comparative fit index. SRMR:  
 122 Standardized root mean squared residual. AICc: Second-order Akaike information  
 123 criterion.  $\Delta$ AICc: Delta AICc.

SEM models	Omitted paths	df	$\chi^2$	$P$	CFI	SRMR	AICc	$\Delta$ AICc
<b>China; Negative</b>								
1 <sup>a</sup>		1	0.563	0.453	1	0.004	1818.8	10.75
2	Human impacts -> $H_2'$	2	1.267	0.531	1	0.005	1816.6	8.53
3	Human impacts -> $H_2'$ Human impacts -> Chemodiversity	4	2.924	0.571	1	0.017	1812.5	4.46
4	Human impacts -> $H_2'$ Human impacts -> Chemodiversity Human impacts -> Biodiversity	5	4.335	0.502	1	0.017	1811.2	3.08
5	Human impacts -> $H_2'$ Human impacts -> Chemodiversity Human impacts -> Biodiversity Human impacts -> Energy	6	6.201	0.401	1	0.017	1810.3	2.19
6	Human impacts -> $H_2'$ Human impacts -> Chemodiversity Human impacts -> Biodiversity Human impacts -> Energy Nutrient -> DOM traits	7	8.631	0.280	1	0.018	1810.0	1.92
7	Human impacts -> $H_2'$ Human impacts -> Chemodiversity Human impacts -> Biodiversity Human impacts -> Energy Nutrient -> DOM traits Biodiversity -> DOM traits	8	10.579	0.227	1	0.020	1809.3	1.20
8 <sup>b</sup>	Human impacts -> $H_2'$ Human impacts -> Chemodiversity Human impacts -> Biodiversity Human impacts -> Energy Nutrient -> DOM traits Biodiversity -> DOM traits Climate change -> DOM traits	7	6.714	0.459	1	0.018	1808.1	0
<b>China; Positive</b>								
1 <sup>a</sup>		1	0.760	0.383	1	0.004	1966.3	15.04

2	Nutrient -> Biodiversity	2	0.768	0.681	1	0.004	1963.4	12.11
3	Nutrient -> Biodiversity Energy -> $H_2'$	3	0.869	0.833	1	0.004	1960.6	9.33
4	Nutrient -> Biodiversity Energy -> $H_2'$ Human impacts -> $H_2'$	4	1.086	0.896	1	0.005	1958.0	6.71
5	Nutrient -> Biodiversity Energy -> $H_2'$ Human impacts -> $H_2'$ Nutrient -> DOM traits	5	1.477	0.916	1	0.006	1955.6	4.31
6	Nutrient -> Biodiversity Energy -> $H_2'$ Human impacts -> $H_2'$ Nutrient -> DOM traits Human impacts -> Biodiversity	6	2.526	0.866	1	0.009	1953.9	2.61
7	Nutrient -> Biodiversity Energy -> $H_2'$ Human impacts -> $H_2'$ Nutrient -> DOM traits Human impacts -> Biodiversity Climate change -> $H_2'$	7	3.888	0.793	1	0.010	1952.6	1.26
8 <sup>b</sup>	Nutrient -> Biodiversity Energy -> $H_2'$ Human impacts -> $H_2'$ Nutrient -> DOM traits Human impacts -> Biodiversity Climate change -> $H_2'$ Climate change -> Chemodiversity	8	5.291	0.726	1	0.013	1951.3	0
9	Nutrient -> Biodiversity Energy -> $H_2'$ Human impacts -> $H_2'$ Nutrient -> DOM traits Human impacts -> Biodiversity Climate change -> $H_2'$ Climate change -> Chemodiversity Human impacts -> DOM traits	9	8.220	0.512	1	0.019	1951.6	0.30
10	Nutrient -> Biodiversity Energy -> $H_2'$ Human impacts -> $H_2'$ Nutrient -> DOM traits Human impacts -> Biodiversity Climate change -> $H_2'$ Climate change -> Chemodiversity Human impacts -> DOM traits Nutrient -> $H_2'$	10	11.134	0.347	1	0.023	1951.9	0.63
<b>Norway; Negative</b>								
1 <sup>a</sup>		0	0	0	1	0	1225.2	11.51
2	Energy -> Chemodiversity	1	0.120	0.729	1	0.004	1222.6	8.96

3	Energy -> Chemodiversity Climate change -> Energy	2	0.263	0.877	1	0.006	1220.1	6.48
4	Energy -> Chemodiversity Climate change -> Energy Energy -> $H_2'$	3	0.656	0.884	1	0.007	1218.0	4.29
5	Energy -> Chemodiversity Climate change -> Energy Energy -> $H_2'$ Nutrient -> DOM traits	4	1.097	0.895	1	0.011	1215.8	2.18
6	Energy -> Chemodiversity Climate change -> Energy Energy -> $H_2'$ Nutrient -> DOM traits Climate change -> Nutrient	5	2.097	0.836	1	0.022	1214.3	0.67
7 <sup>b</sup>	Energy -> Chemodiversity Climate change -> Energy Energy -> $H_2'$ Nutrient -> DOM traits Climate change -> Nutrient Climate change -> $H_2'$	6	3.893	0.691	1	0.024	1213.7	0
8	Energy -> Chemodiversity Climate change -> Energy Energy -> $H_2'$ Nutrient -> DOM traits Climate change -> Nutrient Climate change -> $H_2'$ Chemodiversity -> DOM traits	7	6.735	0.457	1	0.038	1214.1	0.41
<b>Norway; Positive</b>								
1 <sup>a</sup>		0	0	0	1	0	1751.2	19.79
2	Human impacts -> Chemodiversity	1	0.097	0.756	1	0.002	1748.3	16.91
3	Human impacts -> Chemodiversity Biodiversity -> DOM traits	2	0.225	0.894	1	0.002	1745.5	14.11
4	Human impacts -> Chemodiversity Biodiversity -> DOM traits Energy -> DOM traits	3	0.391	0.942	1	0.003	1742.8	11.39
5	Human impacts -> Chemodiversity Biodiversity -> DOM traits Energy -> DOM traits Energy -> $H_2'$	4	0.802	0.938	1	0.006	1740.4	8.97
6	Human impacts -> Chemodiversity Biodiversity -> DOM traits Energy -> DOM traits Energy -> $H_2'$ Climate change -> $H_2'$	5	1.286	0.936	1	0.008	1738.0	6.66

7	Human impacts -> Chemodiversity Biodiversity -> DOM traits Energy -> DOM traits Energy -> $H_2'$ Climate change -> $H_2'$ Human impacts -> Biodiversity	6	1.892	0.929	1	0.009	1735.9	4.51
8	Human impacts -> Chemodiversity Biodiversity -> DOM traits Energy -> DOM traits Energy -> $H_2'$ Climate change -> $H_2'$ Human impacts -> Biodiversity Nutrient -> Energy	7	2.513	0.926	1	0.011	1733.8	2.43
9	Human impacts -> Chemodiversity Biodiversity -> DOM traits Energy -> DOM traits Energy -> $H_2'$ Climate change -> $H_2'$ Human impacts -> Biodiversity Nutrient -> Energy Chemodiversity -> $H_2'$	8	3.949	0.862	1	0.012	1732.6	1.2
10 <sup>b</sup>	Human impacts -> Chemodiversity Biodiversity -> DOM traits Energy -> DOM traits Energy -> $H_2'$ Climate change -> $H_2'$ Human impacts -> Biodiversity Nutrient -> Energy Chemodiversity -> $H_2'$ Nutrient -> $H_2'$	9	5.377	0.800	1	0.013	1731.4	0
11	Human impacts -> Chemodiversity Biodiversity -> DOM traits Energy -> DOM traits Energy -> $H_2'$ Climate change -> $H_2'$ Human impacts -> Biodiversity Nutrient -> Energy Chemodiversity -> $H_2'$ Nutrient -> $H_2'$ Human impacts -> DOM traits	10	8.458	0.584	1	0.017	1731.9	0.5

124 <sup>a</sup> Full SEM models; <sup>b</sup> Best-fitting models shown in red.

125

126 **Table S5.** The hypothesized causal relationships and path coefficients in the structural  
 127 equation model (Fig. 6a).

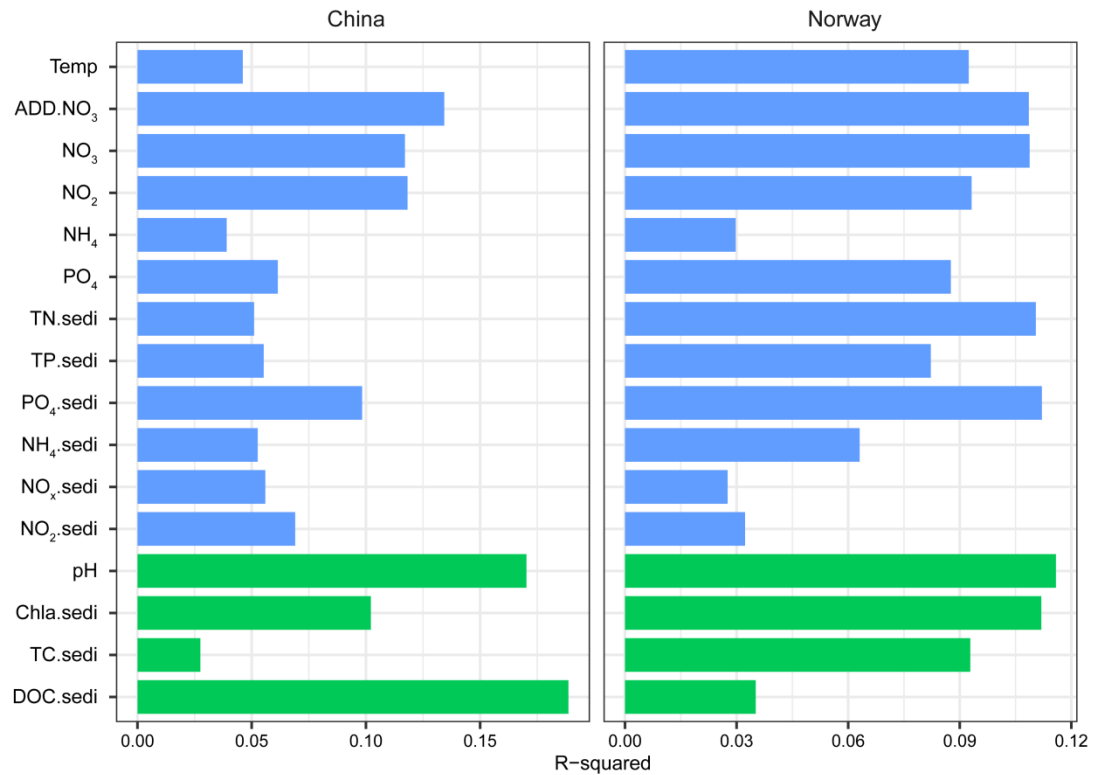
Relationship	Path coefficients
Climate change -> Nutrient	$\lambda_{\text{nut,temp}}$
Human impacts -> Nutrient	$\lambda_{\text{nut,N}}$
Climate change -> Energy	$\lambda_{\text{energy,temp}}$
Human impacts -> Energy	$\lambda_{\text{energy,N}}$
Nutrient -> Energy	$\lambda_{\text{energy,nut}}$
Climate change -> Biodiversity	$\lambda_{\text{biodiv,temp}}$
Human impacts -> Biodiversity	$\lambda_{\text{biodiv,N}}$
Nutrient -> Biodiversity	$\lambda_{\text{biodiv,nut}}$
Energy -> Biodiversity	$\lambda_{\text{biodiv,energy}}$
Climate change -> Chemodiversity	$\lambda_{\text{chemodiv,temp}}$
Human impacts -> Chemodiversity	$\lambda_{\text{chemodiv,N}}$
Nutrient -> Chemodiversity	$\lambda_{\text{chemodiv,nut}}$
Energy -> Chemodiversity	$\lambda_{\text{chemodiv,energy}}$
Climate change -> DOM traits	$\lambda_{\text{trait,temp}}$
Human impacts -> DOM traits	$\lambda_{\text{trait,N}}$
Nutrient -> DOM traits	$\lambda_{\text{trait,nut}}$
Energy -> DOM traits	$\lambda_{\text{trait,energy}}$
Biodiversity -> DOM traits	$\lambda_{\text{trait,biodiv}}$
Chemodiversity -> DOM traits	$\lambda_{\text{trait,chemodiv}}$
Climate change -> $H_2'$	$\lambda_{H_2,temp}$
Human impacts -> $H_2'$	$\lambda_{H_2,N}$
Nutrient -> $H_2'$	$\lambda_{H_2,nut}$
Energy -> $H_2'$	$\lambda_{H_2,energy}$
Biodiversity -> $H_2'$	$\lambda_{H_2,biodiv}$
Chemodiversity -> $H_2'$	$\lambda_{H_2,chemodiv}$
DOM traits -> $H_2'$	$\lambda_{H_2,trait}$

128

129

130 **Supplementary Figures**

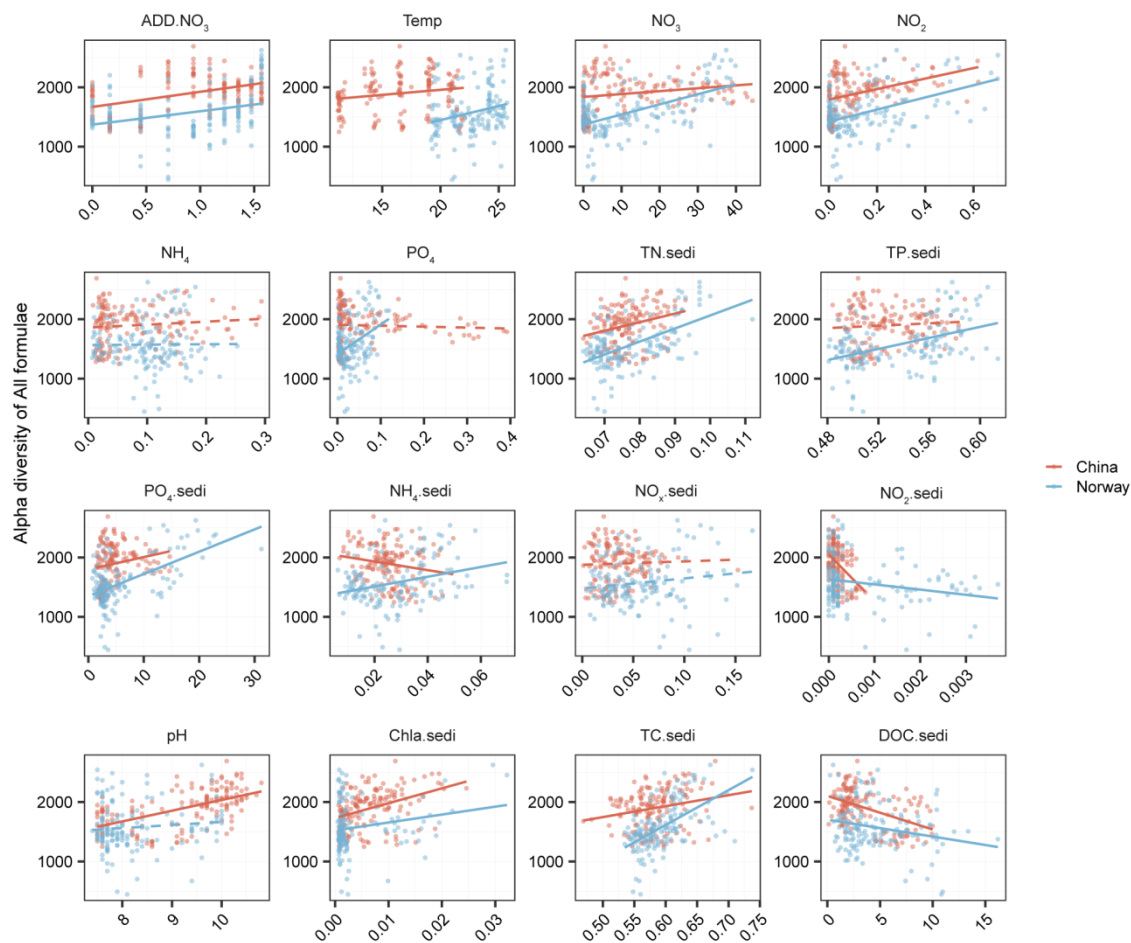
131



132

133 **Figure S1.** Bar plots showing R-squared to identify the effects of environmental (blue)  
134 and energy supply (green) variables on molecular composition of DOM in China and  
135 Norway. R-squared was determined by permutational multivariate analysis of variance  
136 (PERMANOVA) with 999 permutations and was statistically significant ( $P \leq 0.001$ ) for  
137 explanatory variables. R-squared for each explanatory variable stands for the explained  
138 variations in the differences of DOM molecular composition among the samples. Nitrate  
139 addition (ADD.NO<sub>3</sub>) was used to represent nutrient enrichment as the ratio between  
140 nitrate and phosphorus in the initial overlying water was constant. The abbreviations of  
141 explanatory variables are detailed in Table S2.

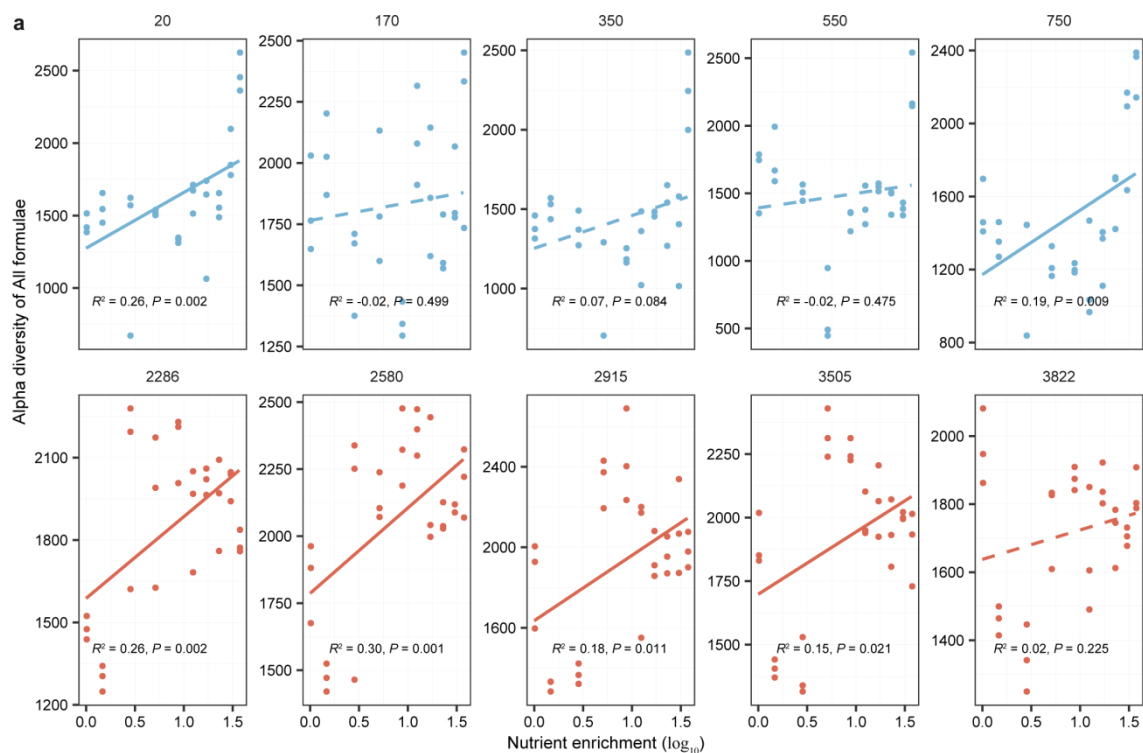
142



143

144 **Figure S2.** The relationships between DOM alpha diversity (i.e., molecular richness) and  
 145 explanatory variables in China (red lines) and Norway (blue lines). We plotted alpha  
 146 diversity against the variables relevant to environment and energy supply (Table S2). The  
 147 relationships are indicated by solid ( $P \leq 0.05$ ) and dotted ( $P > 0.05$ ) lines estimated using  
 148 linear models with one-sided F-statistics.

149

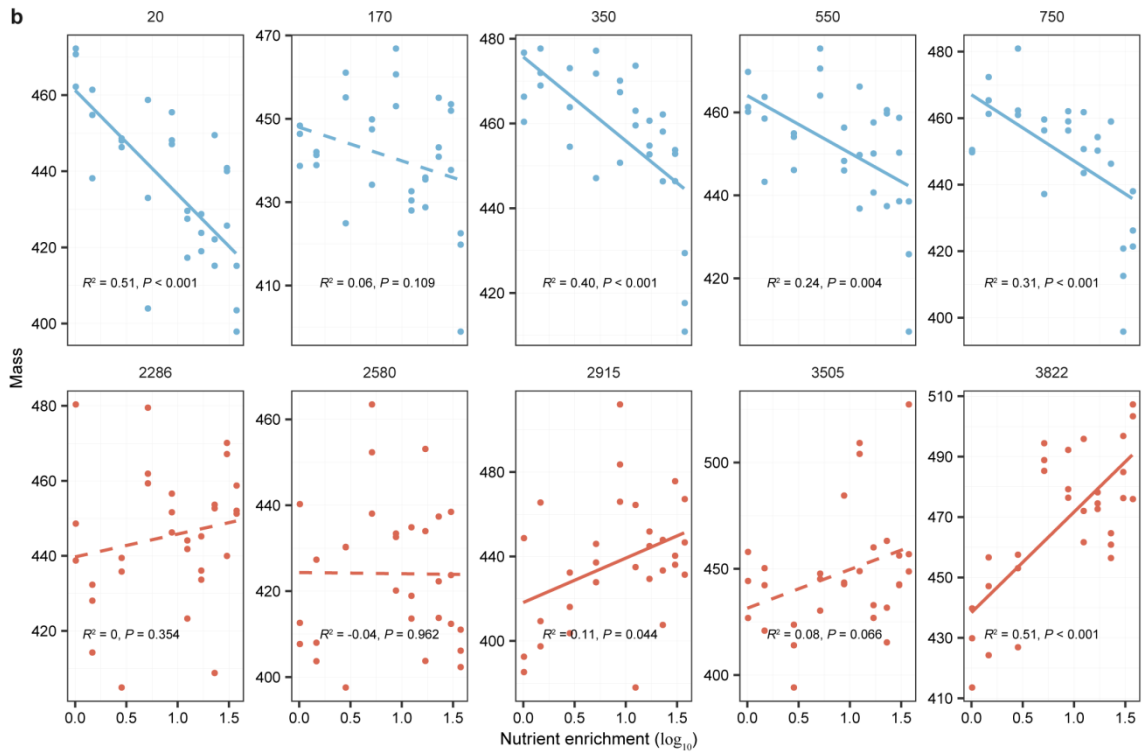


150

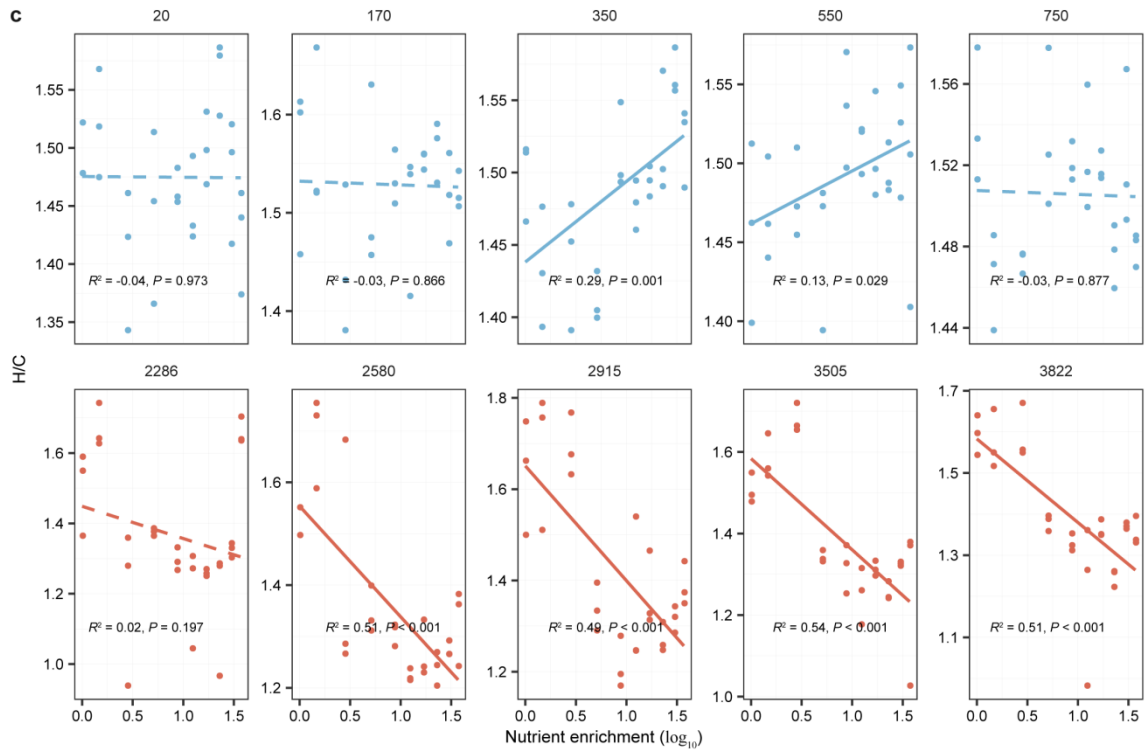
151 **Figure S3.** The relationships between nutrient enrichment and DOM alpha diversity or  
 152 molecular traits in China (red lines) and Norway (blue lines) at different elevations (20 to  
 153 3,822 m a.s.l.). We considered richness for all formulae (a) and also molecular traits such  
 154 as weighted means of mass (b), H/C ratio (c), O/C ratio (d) and AI<sub>Mod</sub> (e) in China (red  
 155 lines and dots) and Norway (blue lines and dots). We used nitrate addition to represent  
 156 nutrient enrichment as the ratio between nitrate and phosphorus in the initial overlying  
 157 water was constant. We plotted the richness or traits against the nutrient gradient of  
 158 nitrate, and their relationships are indicated by solid ( $P \leq 0.05$ ) and dotted ( $P > 0.05$ ) lines  
 159 using linear models with one-sided F-statistics.

160





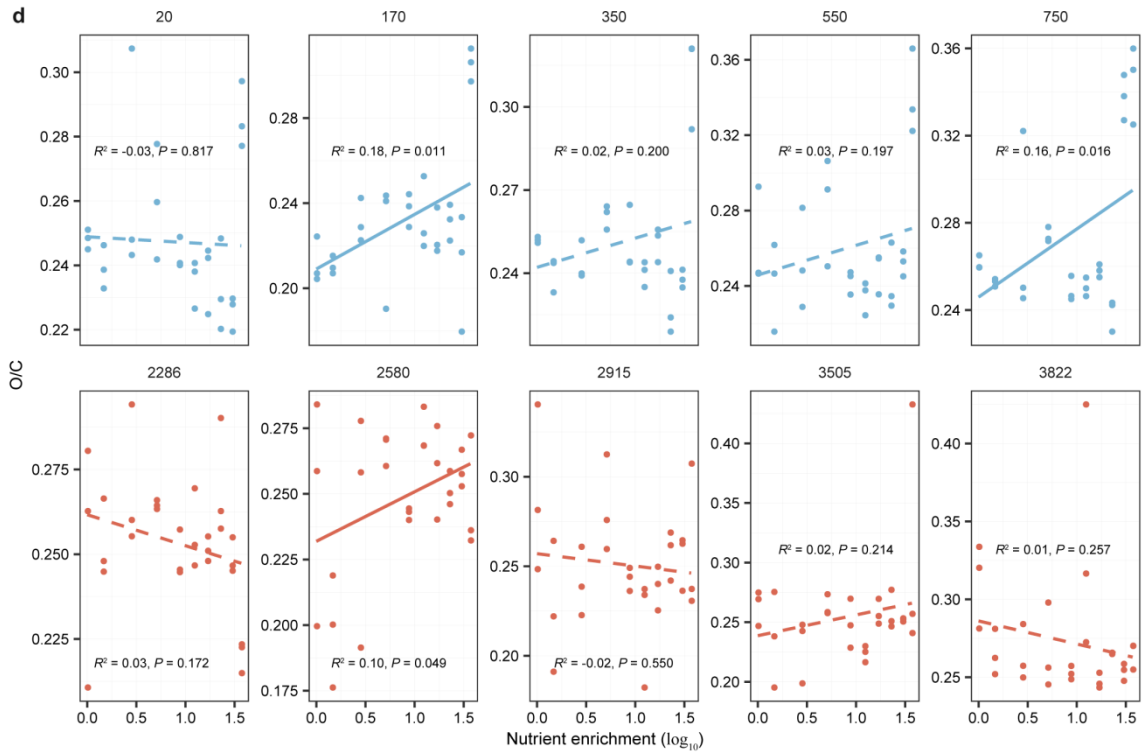
161



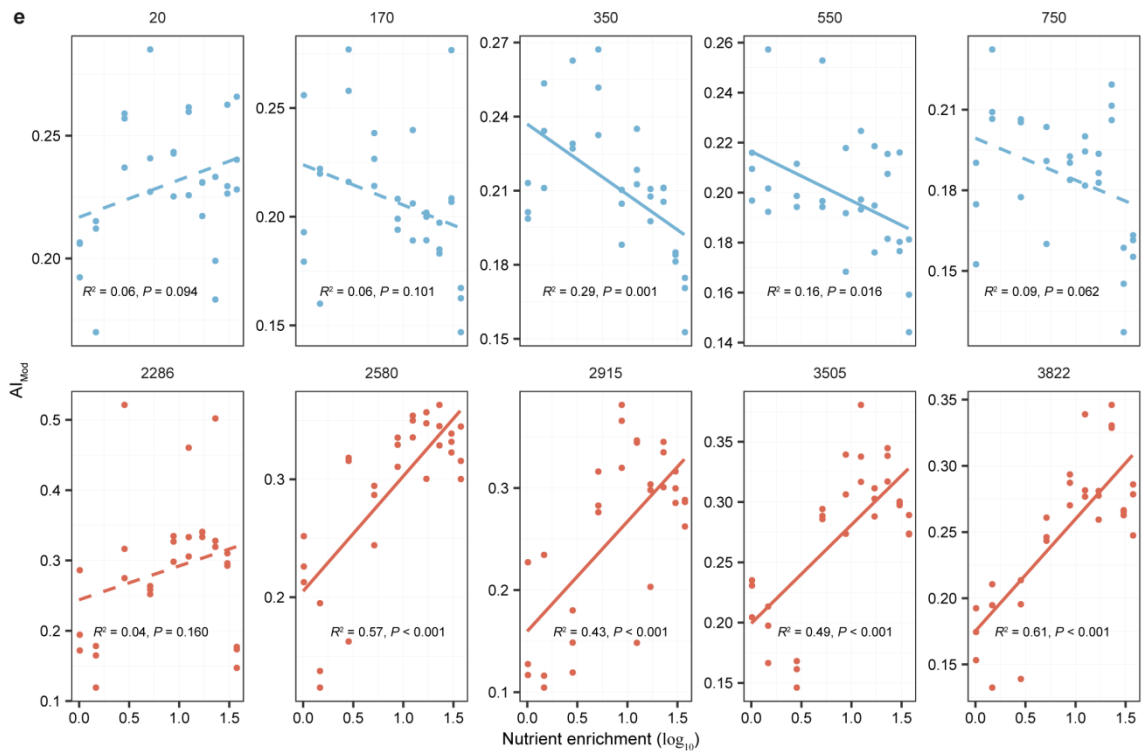
162

163 **Figure S3.** Continued. Weighted means of mass (b), and H/C ratio (c).

164



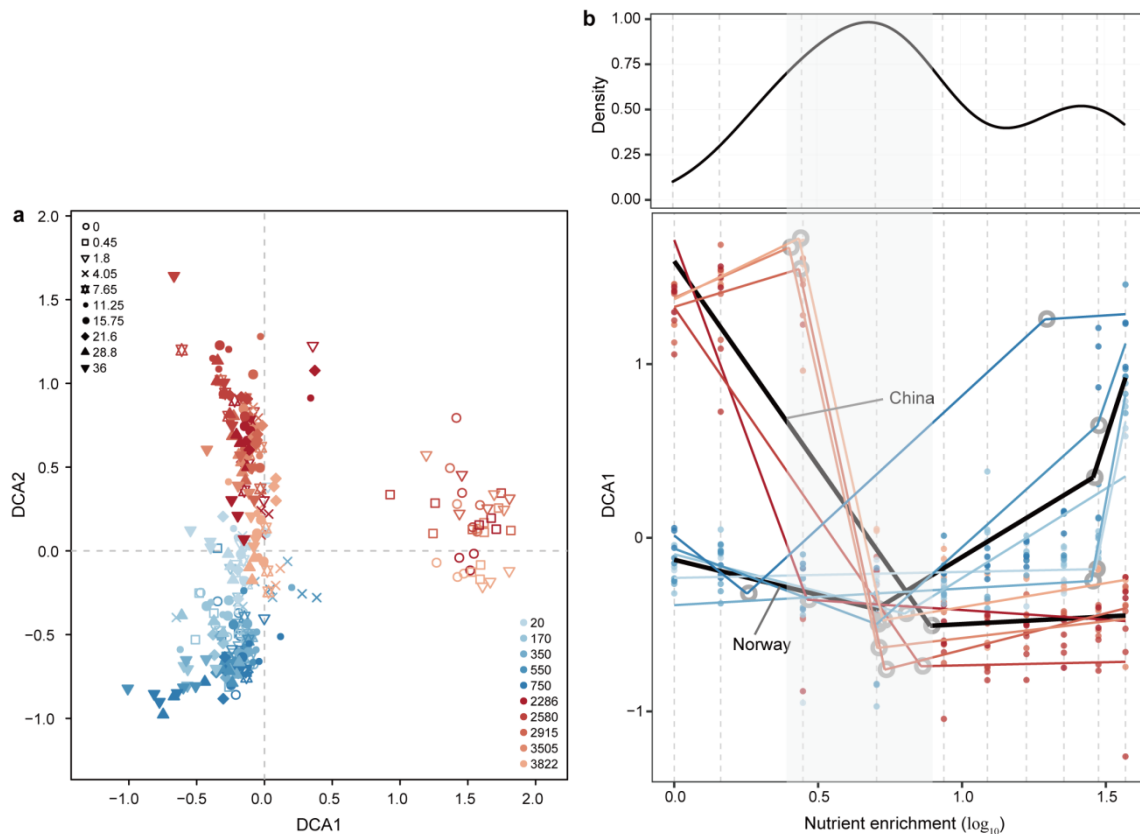
165



166

167 **Figure S3.** Continued. Weighted means of O/C ratio (d) and  $AI_{Mod}$  (e).

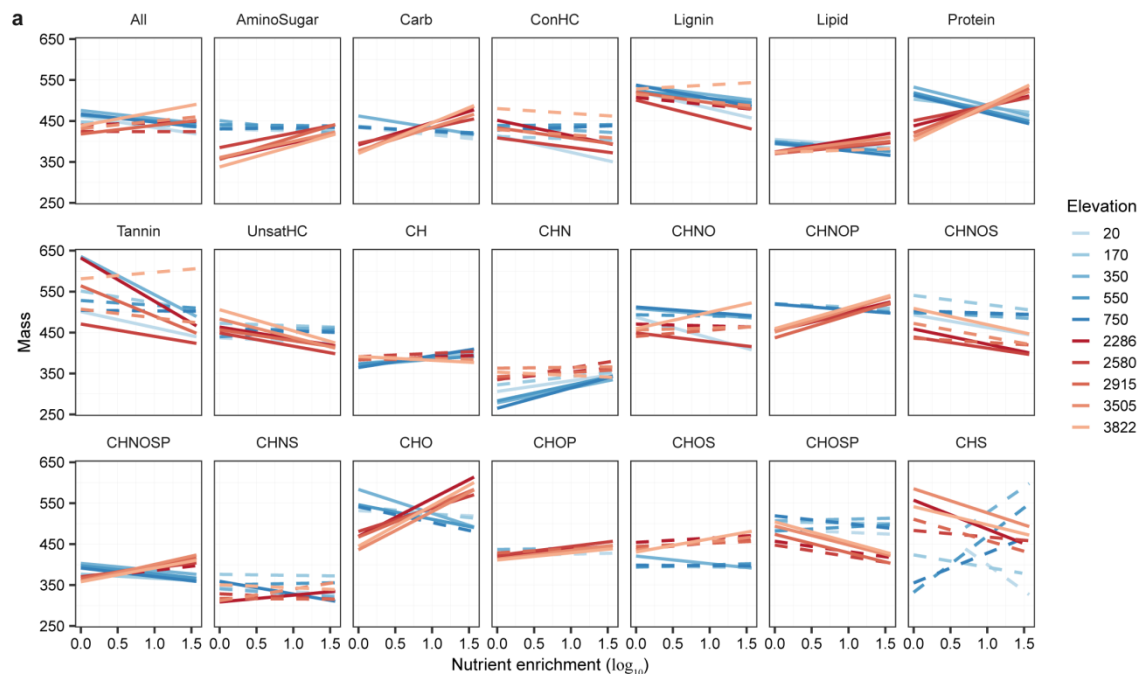
168



169

170 **Figure S4.** Variations in DOM compositions along the nutrient gradient of nitrate. (a)  
 171 Detrended correspondence analyses (DCA) of DOM compositions. (b) Nutrient  
 172 breakpoint estimation of the first axis of the DCA of DOM composition for each  
 173 elevation using piecewise regression analysis with Bayesian Information Criteria  
 174 statistics (Muggeo, 2008). The upper panel is a density plot of the distribution of  
 175 breakpoints. The black thick lines in the lower panel indicate each region (i.e., China and  
 176 Norway), and the colored dots or lines indicate the elevations of the two regions, which  
 177 are consistent with the figure legend of Fig. S4a. The gray open circles indicate nutrient  
 178 breakpoints. The vertical gray lines indicate the ten experimental nutrient levels. We  
 179 found that, along the nutrient gradient, there were breakpoints of the first axis of the DCA  
 180 mostly occurring between 1.80 and 4.05 mg N L<sup>-1</sup> (indicated by gray shade) especially in  
 181 China.

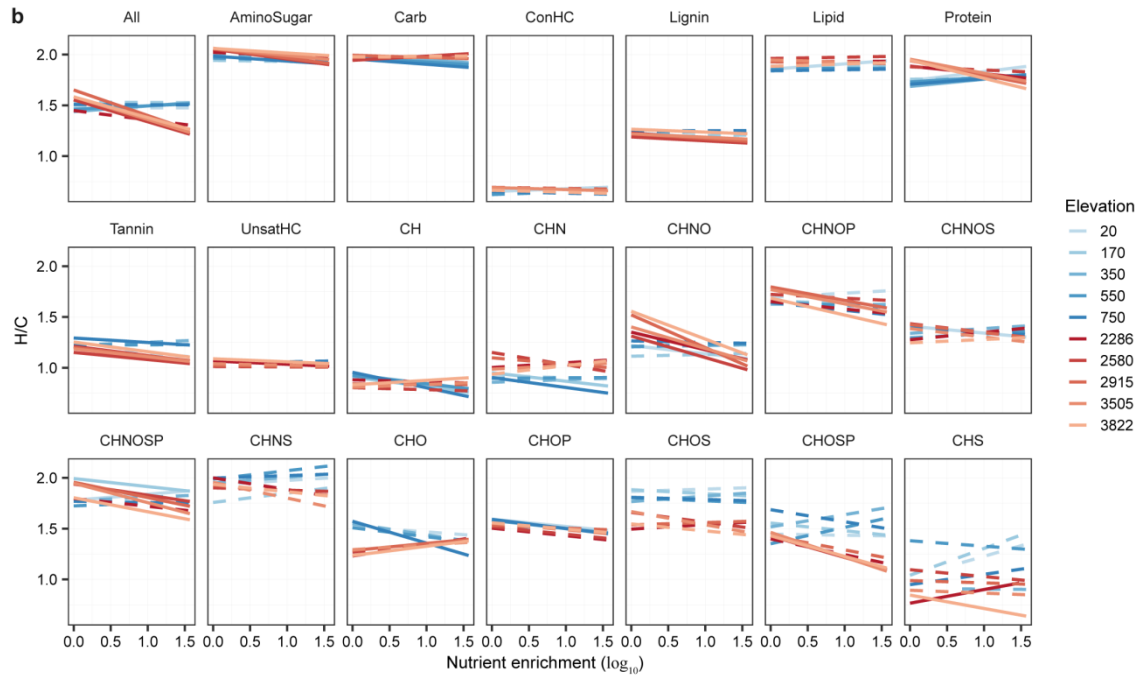
182



183

184 **Figure S5.** Effects of nutrient enrichment on DOM traits for all formulae and subsets of  
 185 formulae within compound classes or elemental combinations across different elevations  
 186 in China (red lines) and Norway (blue lines). We considered molecular traits such as  
 187 weighted means of mass (a), H/C ratio (b) and  $AI_{Mod}$  (c), which were plotted against the  
 188 nutrient gradient of nitrate, and their relationships are indicated by the solid ( $P \leq 0.05$ ) or  
 189 dotted ( $P > 0.05$ ) lines using linear models with one-sided F-statistics. The details of  
 190 abbreviations of DOM traits are available in Table S1. We found that nutrient enrichment  
 191 increased the weighted means of molecular mass more strongly at higher elevations in  
 192 China (with maximal 495 Da at 3,822 m a.s.l.), but decreased more strongly at lower  
 193 elevations in Norway (with minimal 405 Da at 20 m a.s.l.). This finding implies that  
 194 nutrient enrichment leads to an increase in the molecular mass especially at colder  
 195 temperatures in subtropical regions, but a decline at the warmer temperatures in subarctic  
 196 regions.

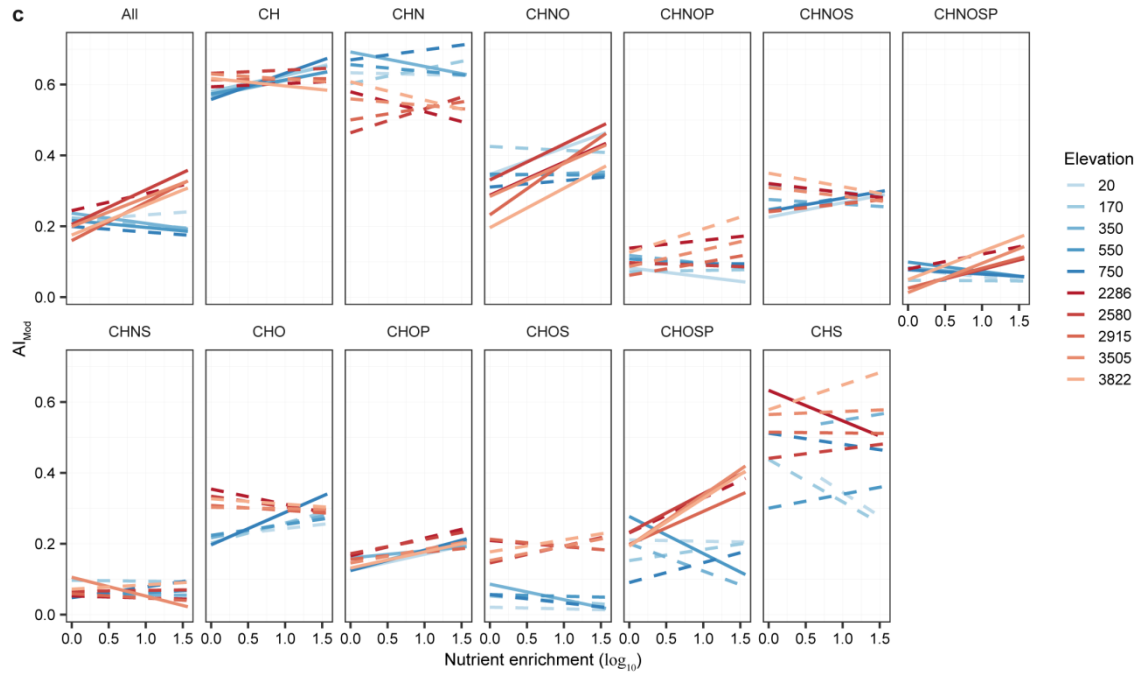
197



198

199 **Figure S5.** Continued. Weighted means of H/C ratio (b).

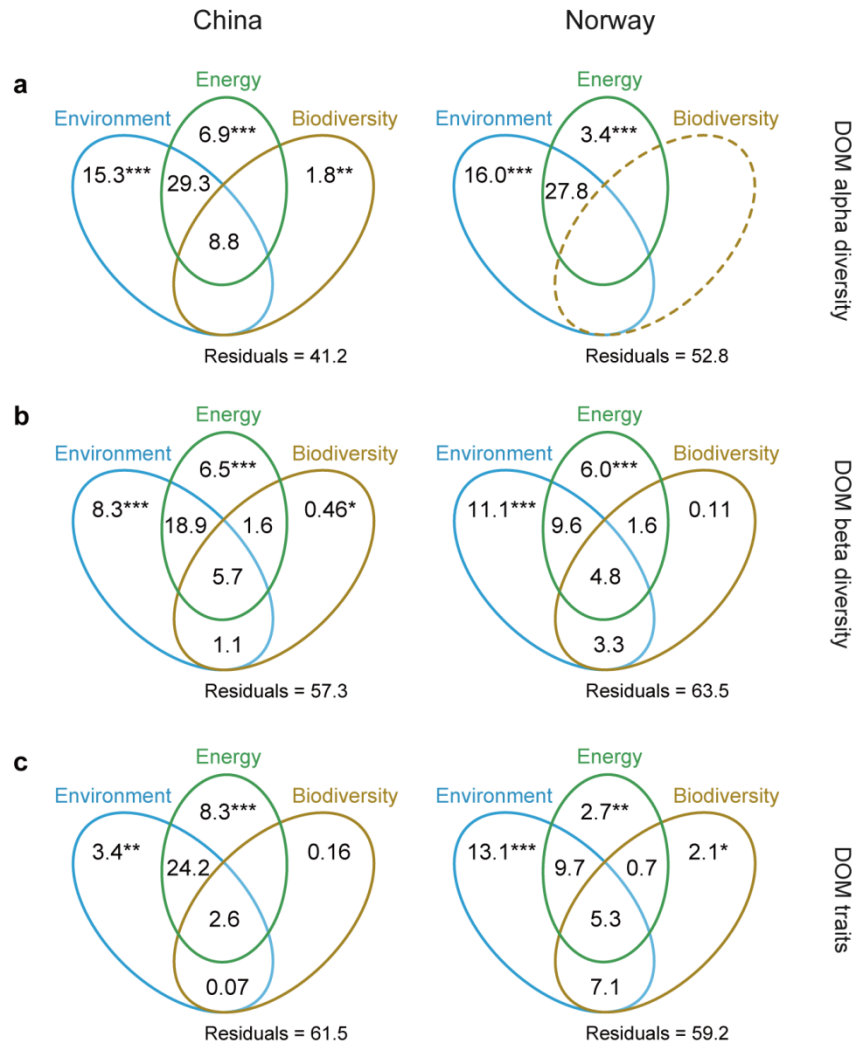
200



201

202 **Figure S5.** Continued. Weighted means of  $AI_{Mod}(c)$ .

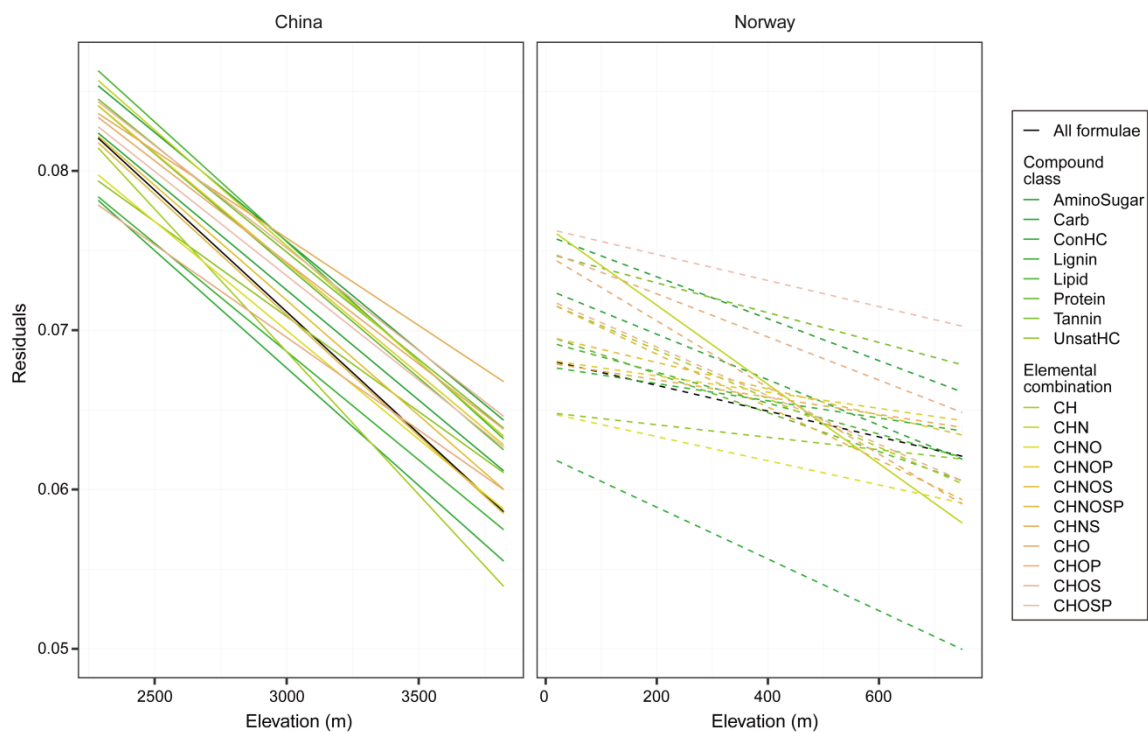
203



204

205 **Figure S6.** The roles of microbes in explaining the alpha diversity (upper panel), beta  
 206 diversity (middle panel) and molecular traits (lower panel) of DOM using variation  
 207 partitioning analysis. The numbers indicate the variance explained (%) by environments,  
 208 energy supply and bacterial biodiversity which were described in detail in Table S2. The  
 209 significance was examined using one-sided F-statistics and asterisks represent statistically  
 210 significant effects at \*\*\*,  $P \leq 0.001$ ; \*\*,  $P \leq 0.01$ ; \*,  $P \leq 0.05$ .

211

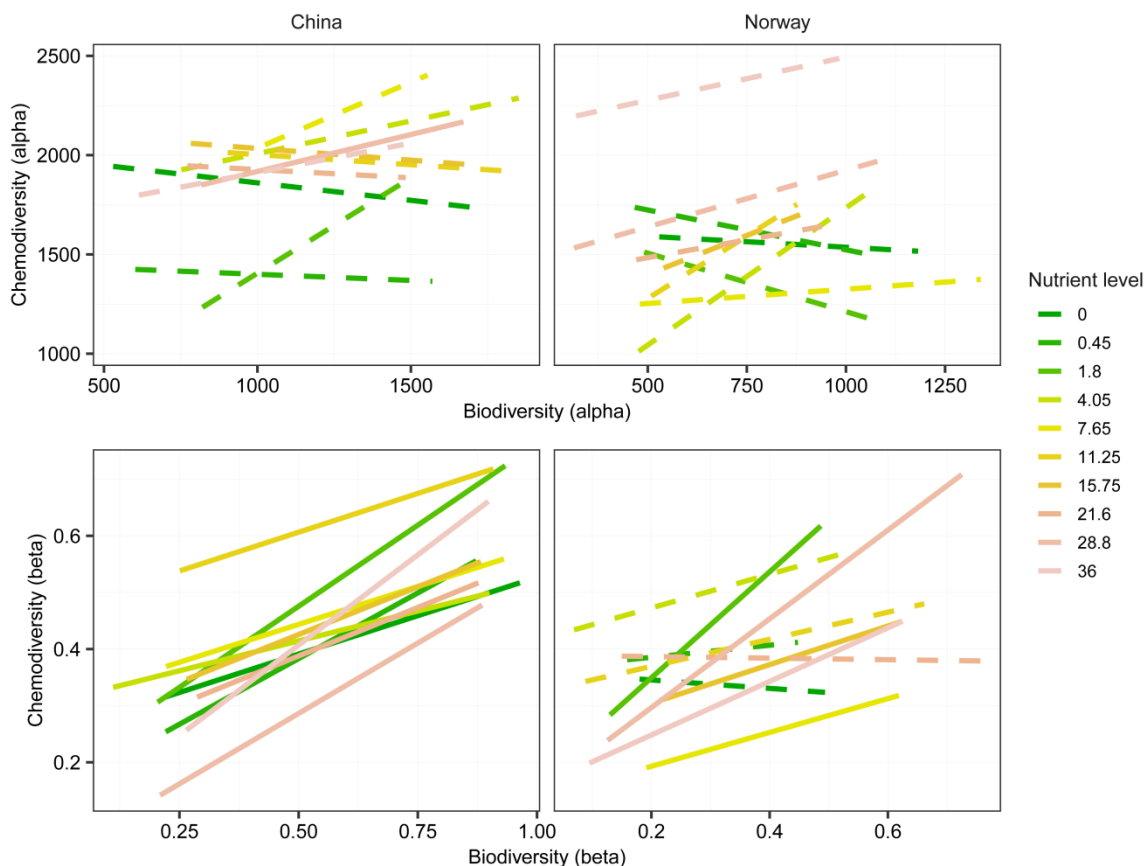


212

213 **Figure S7.** The effects of elevation on DOM-microbe associations indicated by  
 214 Procrustes residuals, that is, the difference in composition between DOM and bacteria for  
 215 each microcosm. These effects are indicated by solid ( $P \leq 0.05$ ) or dotted ( $P > 0.05$ ) lines  
 216 estimated using linear models with one-sided F-statistics. The colours of the lines indicate  
 217 the DOM composition for all formulae and categories of compound classes or elemental  
 218 combinations.

219

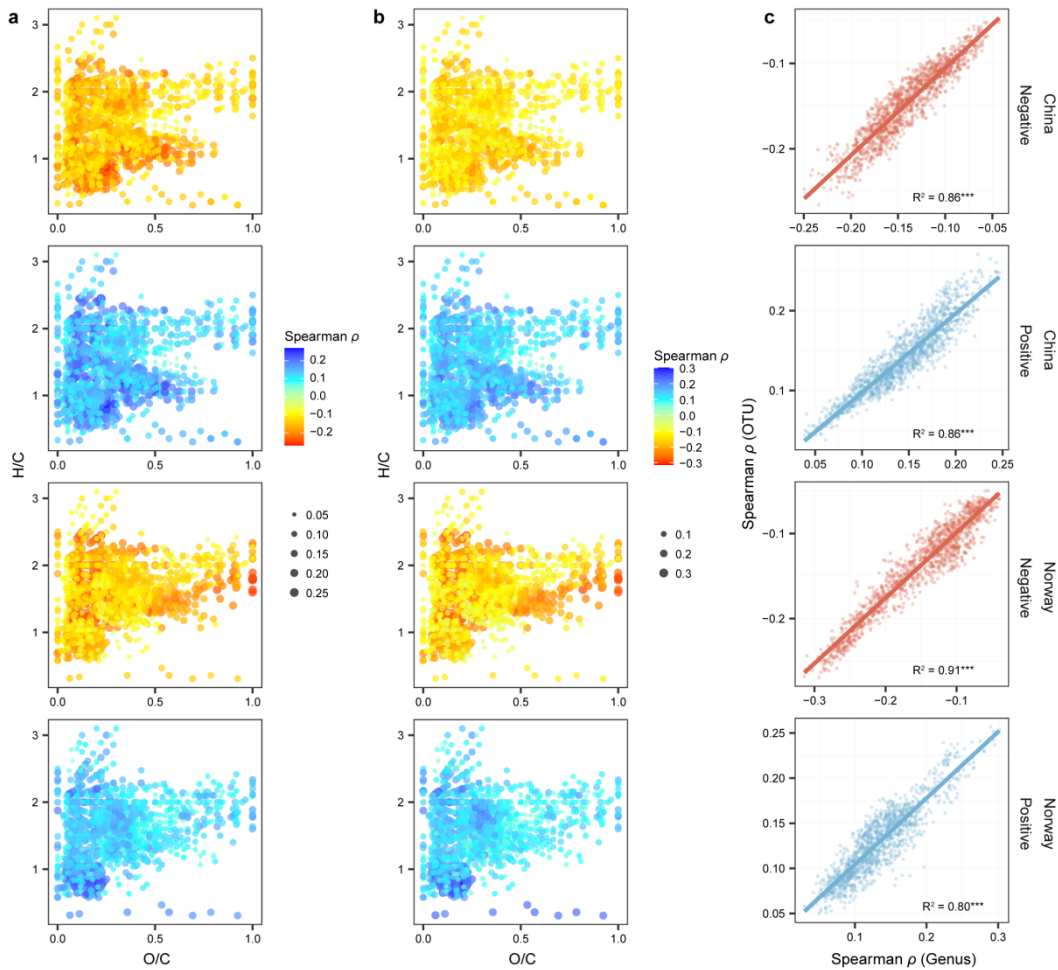




220

221 **Figure S8.** The relationships between chemodiversity and bacterial diversity in China  
 222 and Norway along the nutrient gradient of nitrate. Upper panel: Alpha diversity (richness)  
 223 of DOM molecular formulae and bacterial OTUs. Lower panel: Beta diversity determined  
 224 by the Bray–Curtis dissimilarity index for the mixtures of molecular formulae and the  
 225 communities of OTUs. Each line visualises the relationship of alpha or beta diversity  
 226 between DOM and bacteria across 15 samples at each nutrient level in China or Norway.  
 227 The relationships are indicated by solid ( $P \leq 0.05$ ) and dotted ( $P > 0.05$ ) lines using linear  
 228 models, and the significance was determined by ANOVA with one-sided F-statistics  
 229 (upper panels) or two-sided Mantel test with 999 permutations (lower panels).

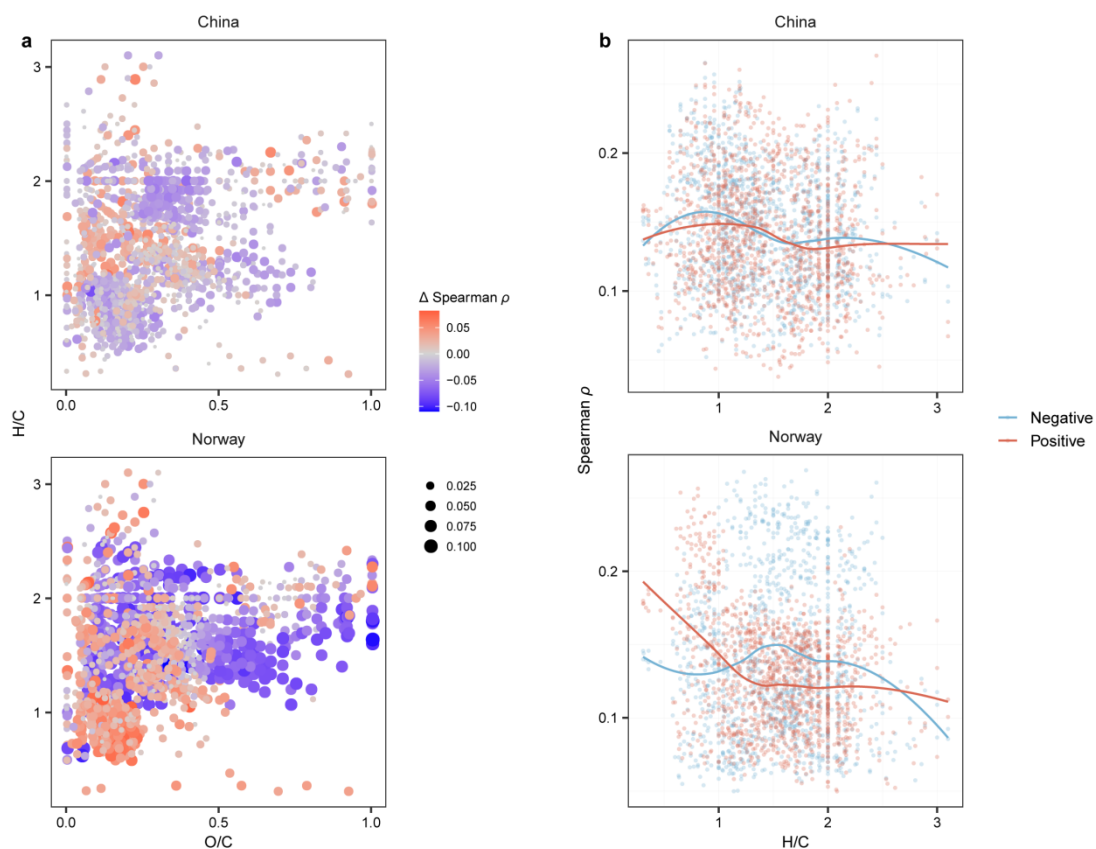
230



231

232 **Figure S9.** Molecular formulae correlating with bacterial OTUs (a) or genera (b) in  
 233 China and Norway using Spearman's rank correlation ( $\rho$ ). Each molecule is colored by  
 234 the mean  $\rho$  value of negative or positive correlations across all bacterial OTUs, and the  
 235 absolute value of mean  $\rho$  is indicated by the dot size. (c) The relationships in Spearman  $\rho$   
 236 between bacterial OTU and genus levels for both negative and positive correlations in  
 237 China and Norway. Solid lines indicate significant linear fits with one-sided F-statistics  
 238 ( $P \leq 0.05$ ).

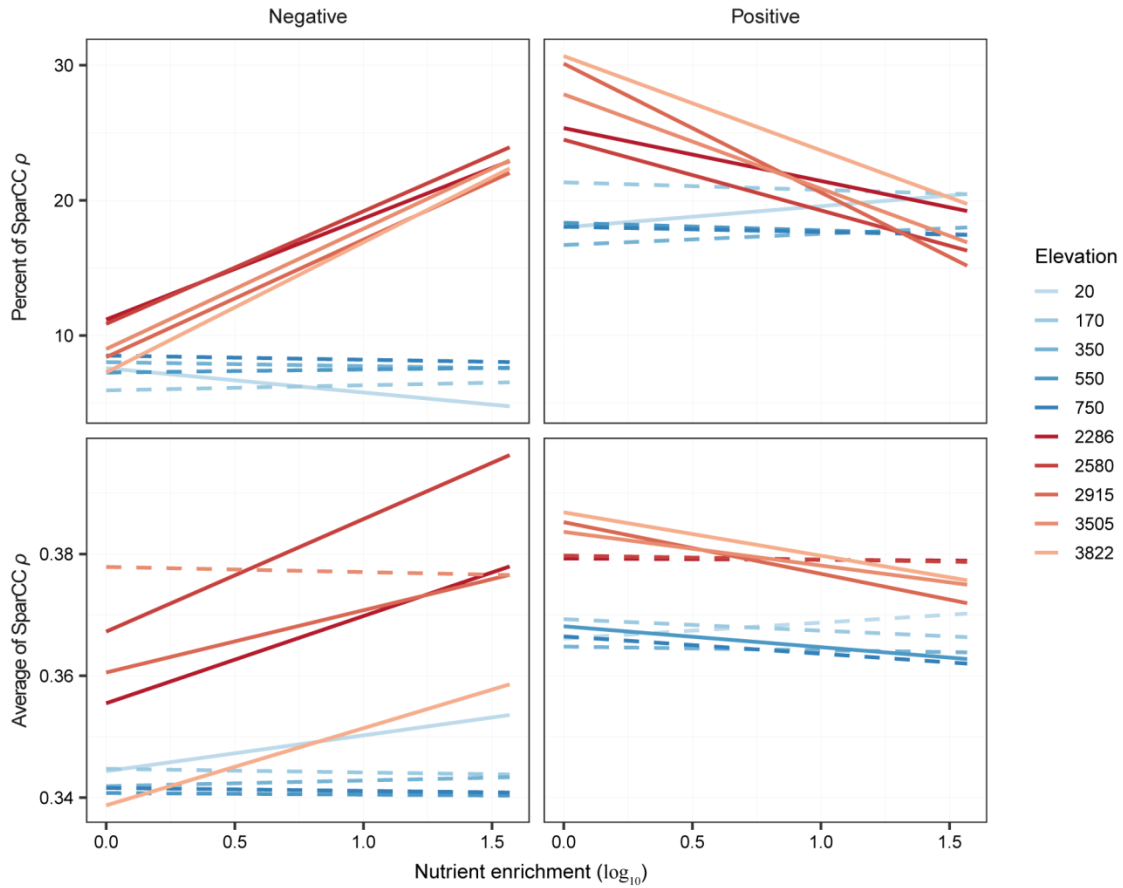
239



240

241 **Figure S10.** Correlations between DOM and bacteria regarding molecular traits. (a)  
 242 Molecular formulae correlating with bacterial OTUs in China and Norway using  
 243 Spearman's rank correlation ( $\rho$ ). Each molecule is colored by difference in the absolute  
 244 Spearman  $\rho$  ( $\Delta\rho$ ). The difference for each molecule was calculated by subtracting the  
 245 mean absolute  $\rho$  value of the negative correlations across all bacterial OTUs from that of  
 246 the positive correlations, and the absolute value of  $\rho$  difference is indicated by the dot  
 247 size. (b) The patterns of absolute  $\rho$  values of positive and negative correlations along the  
 248 gradient of H/C ratio visualized with loess regression models.

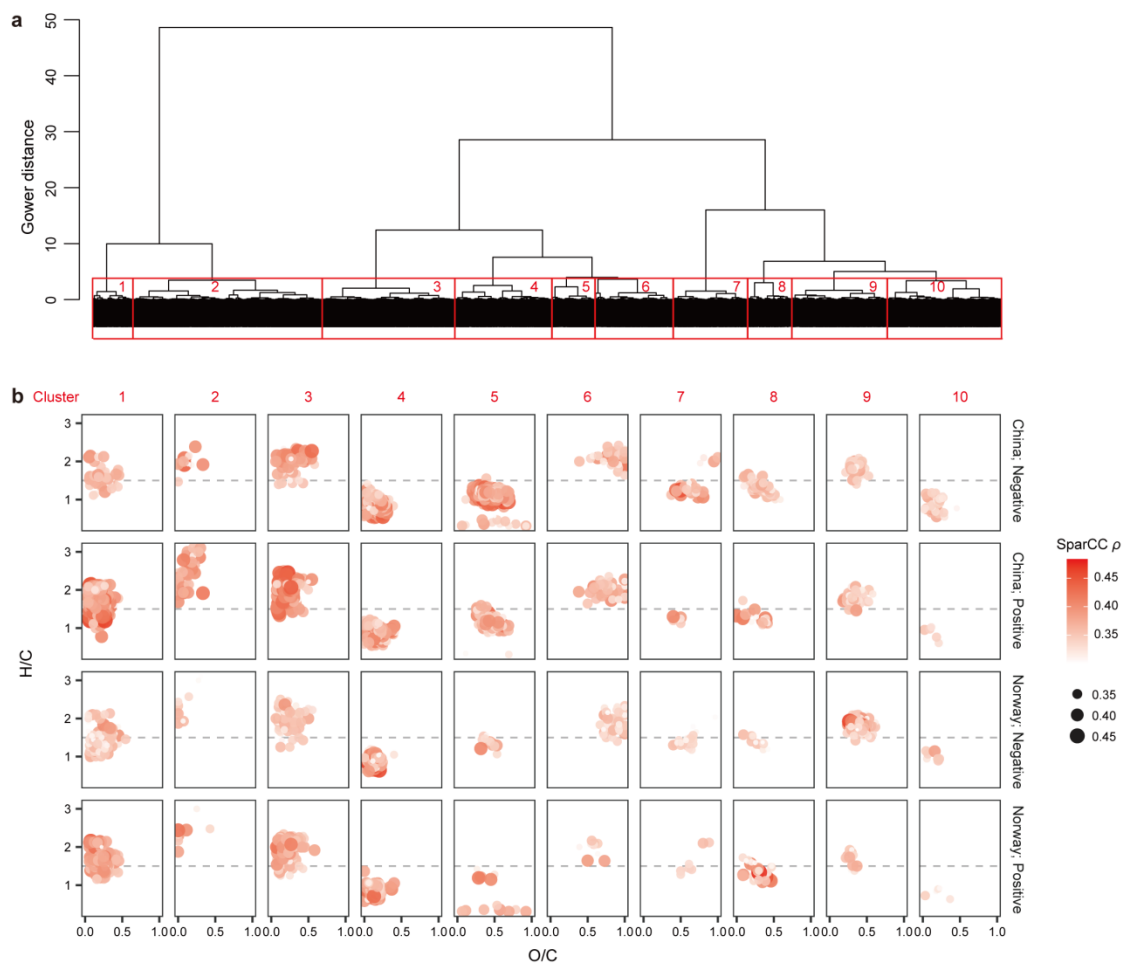
249



250

251 **Figure S11.** The effects of nutrient enrichment on weighted means of indices of DOM-  
 252 bacteria bipartite networks. The network indices include the percent and average of  
 253 strong correlations ( $|\text{SparCC } \rho| \geq 0.3$ ) of negative or positive networks. We plotted these  
 254 indices against the nutrient gradient of nitrate for both negative (left panel) and positive  
 255 (right panel) networks for each elevation in China (red lines) or Norway (blue lines), and  
 256 their relationships are indicated by solid ( $P \leq 0.05$ ) or dotted ( $P > 0.05$ ) lines using linear  
 257 models with one-sided F-statistics.

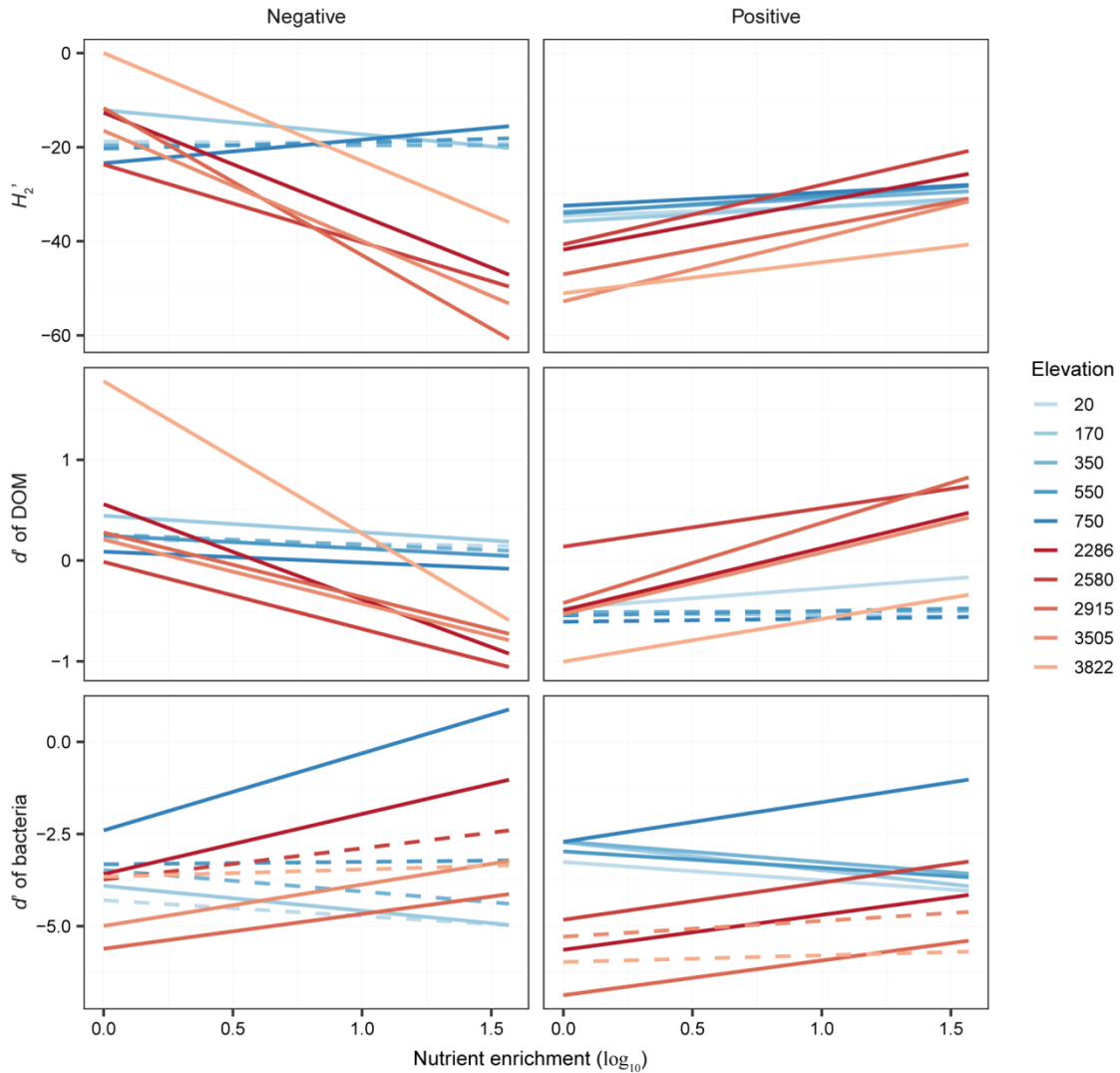
258



259

260 **Figure S12.** Correlations between DOM and bacteria regarding molecular traits. (a)  
 261 Cluster analysis identified ten molecular sub-mixtures based on 16 molecular traits (Table  
 262 S1). (b) Location of the ten clusters in Van Krevelen space with colour- and size-coded  
 263 correlations between molecule-specific intensities and the relative abundance of bacterial  
 264 genera using SparCC (Sparse Correlations for Compositional data). For each molecule,  
 265 we showed the mean absolute SparCC  $\rho$  values of negative or positive correlations across  
 266 all bacterial OTUs. We considered only strong correlations ( $|\text{SparCC } \rho| \geq 0.3$ ).

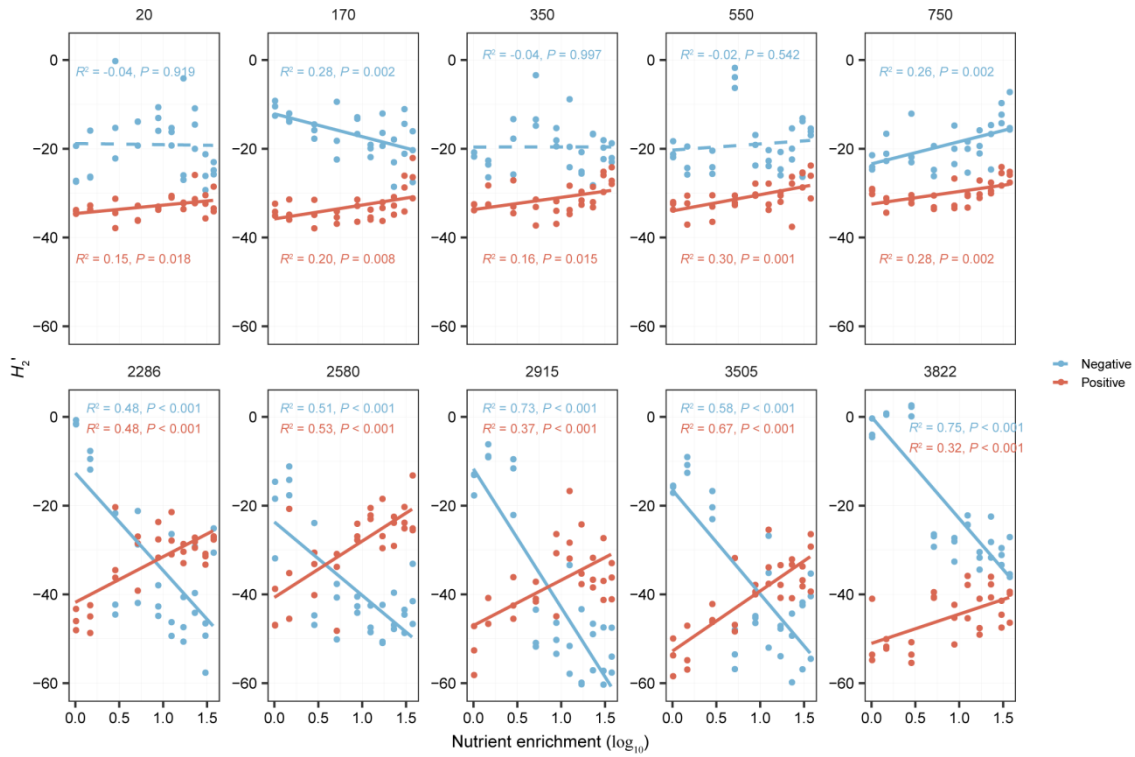
267



268

269 **Figure S13.** The effects of nutrient enrichment on specialization indices of DOM-  
 270 bacteria bipartite networks. The specialization indices include network-level  
 271 specialization  $H_2'$ , and the weighted means of specialization  $d'$  for DOM molecules and  
 272 bacterial genera. We plotted these indices against the nutrient gradient of nitrate for both  
 273 negative (left panel) and positive (right panel) networks for each elevation in China (red  
 274 lines) or Norway (blue lines), and their relationships are indicated by solid ( $P \leq 0.05$ ) or  
 275 dotted ( $P > 0.05$ ) lines using linear models with one-sided F-statistics.

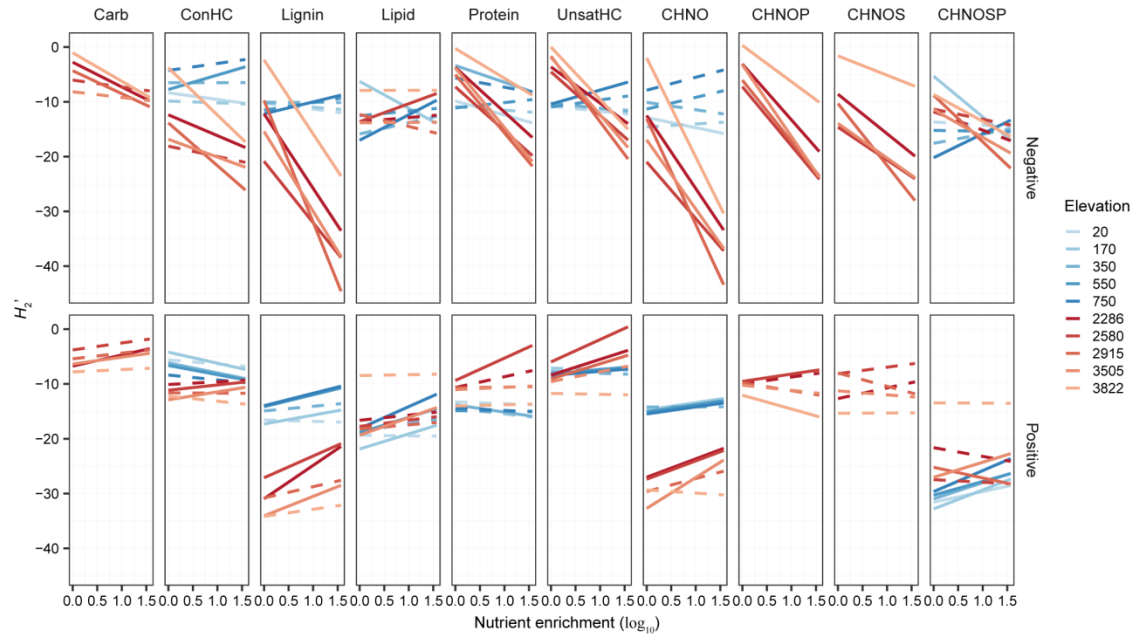
276



277

278 **Figure S14.** The effect of nutrient enrichment on the specialization  $H_2'$  of DOM-bacteria  
 279 bipartite networks. We plotted the  $H_2'$  against the nutrient gradient of nitrate for both  
 280 negative (blue lines) and positive (red lines) networks for each elevation in China or  
 281 Norway, and their relationships are indicated by solid ( $P \leq 0.05$ ) or dotted ( $P > 0.05$ )  
 282 lines using linear models with one-sided F-statistics.

283

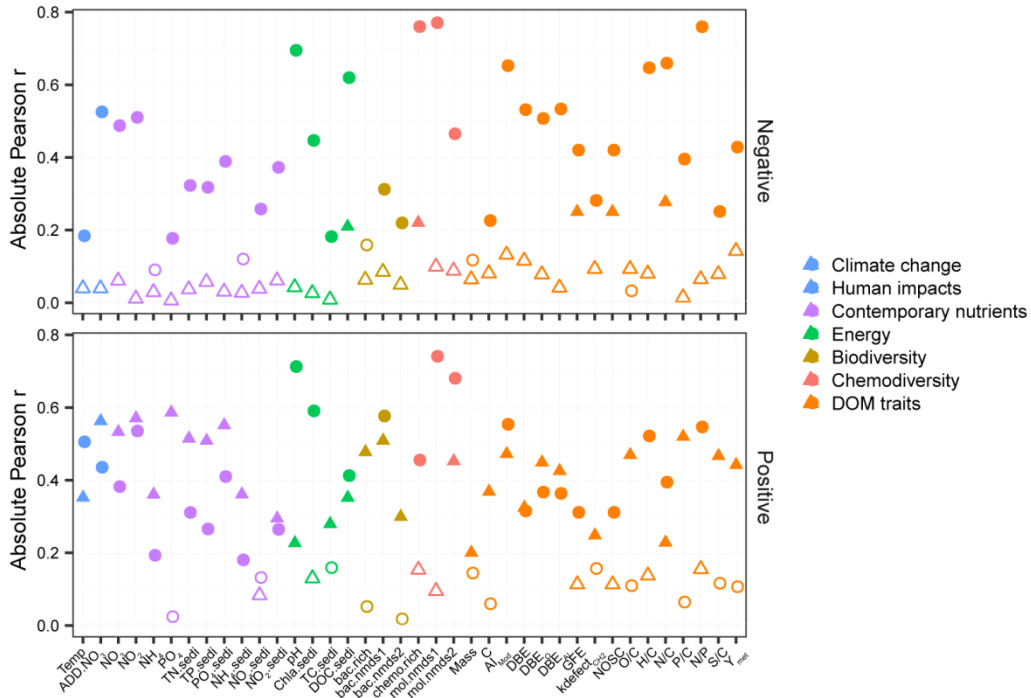


284

285 **Figure S15.** The effects of nutrient enrichment on the specialization  $H_2'$  of DOM-  
 286 bacteria bipartite networks for all formulae and subsets of formulae within the category of  
 287 compound classes or elemental combinations. We plotted the  $H_2'$  against the nutrient  
 288 gradient of nitrate for both negative (upper panel) and positive (lower panel) networks for  
 289 each elevation in China (red lines) or Norway (blue lines), and their relationships are  
 290 indicated by solid ( $P \leq 0.05$ ) or dotted ( $P > 0.05$ ) lines using linear models with one-sided  
 291 F-statistics.

292

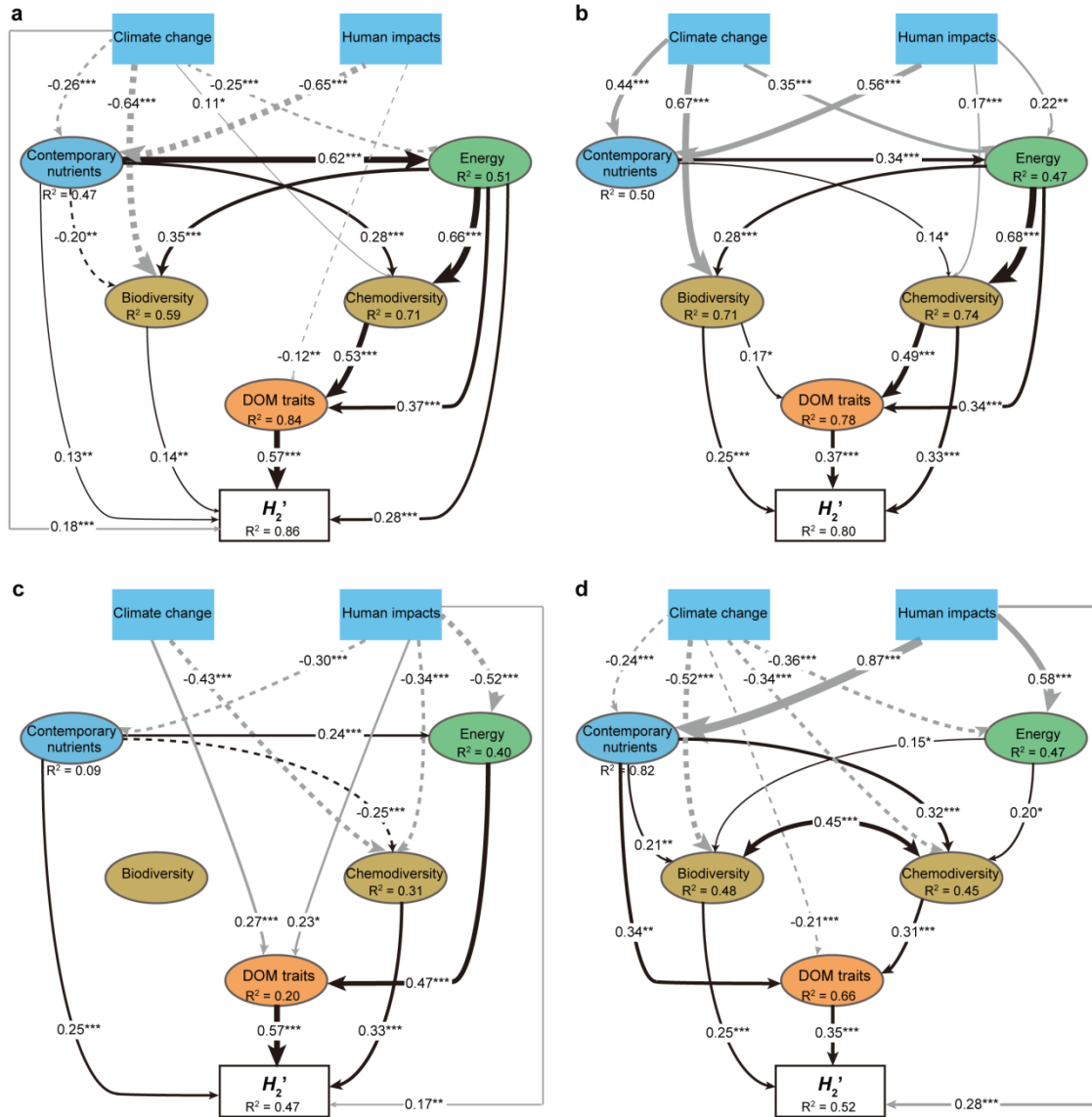




293

294 **Figure S16.** The relative influence of explanatory variables on the specialization  $H_2'$  of  
 295 negative (upper panel) and positive (lower panel) DOM-bacteria bipartite networks using  
 296 Pearson correlation analysis. Each circle and triangle are the absolute values of Pearson  $r$   
 297 for individual explanatory variable in China and Norway, respectively. Solid and open  
 298 circles or triangles indicate the significant ( $P \leq 0.05$ ) and non-significant ( $P > 0.05$ ) two-  
 299 sided Pearson  $r$ , respectively. The details of abbreviations of explanatory variables are  
 300 available in Table S1.

301

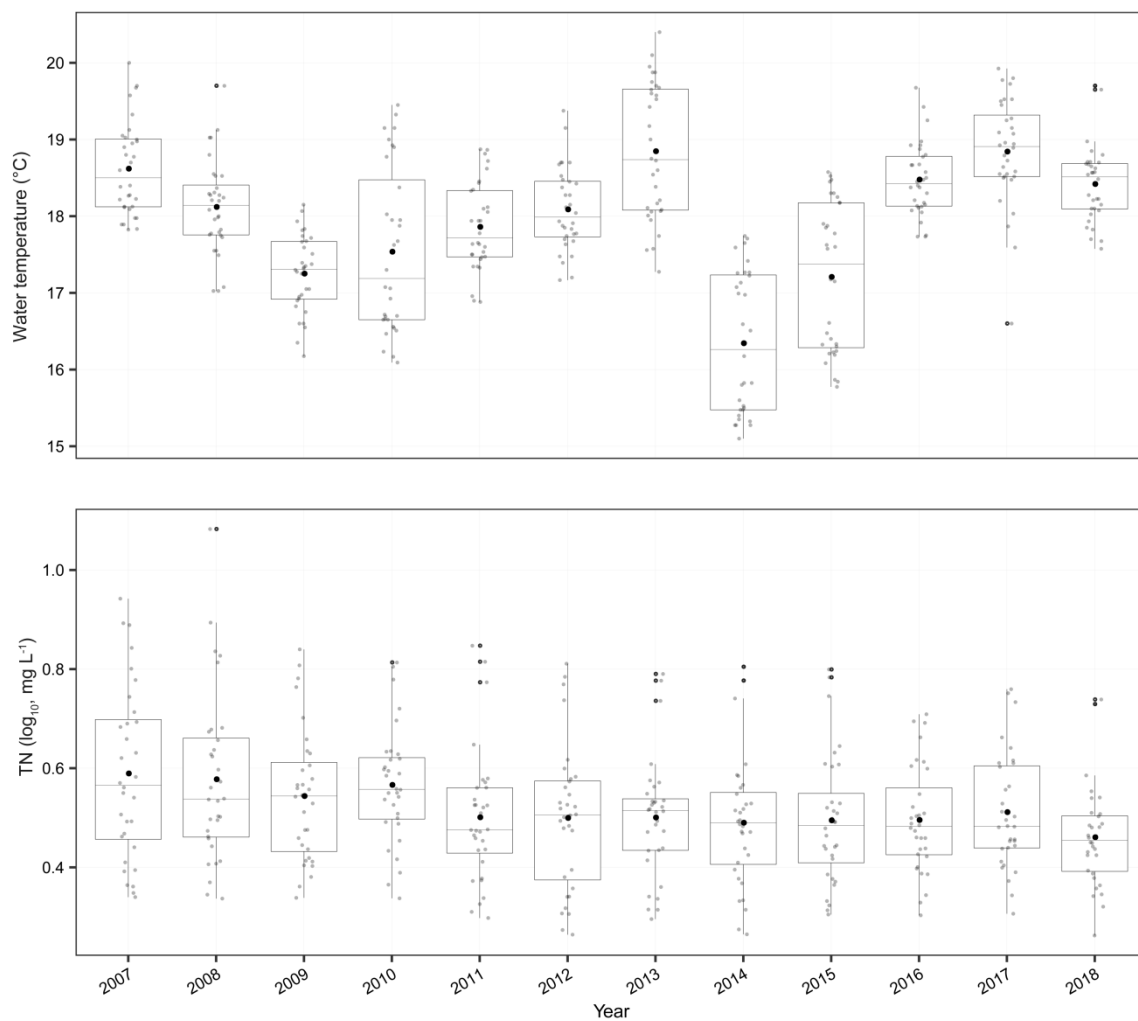


302

303 **Figure S17.** Structural equation models to explain specialization of DOM-bacteria  
 304 bipartite networks. Best-fitting models illustrate the effects of predictor variables on the  
 305  $H_2'$  of negative (a, c) and positive (b, d) bipartite networks in China (a-b) or Norway (c-  
 306 d). Predictor variables were grouped by climate change, human impacts, contemporary  
 307 nutrients, energy supply, biodiversity, chemodiversity and DOM traits, and described in  
 308 detail in Table S3.  $R^2$  denotes the proportion of variance explained for the endogenous  
 309 variables. Dotted and solid arrows indicate the two-sided statistically significantly  
 310 negative and positive (\*\*\*,  $P \leq 0.001$ ; \*\*,  $P \leq 0.01$ ; \*,  $P \leq 0.05$ ) relationships, respectively,  
 311 with z-statistics. Grey or black arrows indicate the hypothesized relationships among the

312 exogenous or endogenous variables and  $H_2'$ , respectively. Arrow widths and  
313 accompanying numbers are the relative effects (that is, standardized path coefficients) of  
314 modeled relationships. Composite and observed variables are indicated in ovals and  
315 rectangles, respectively. Details of model fit are summarized in Table S4.

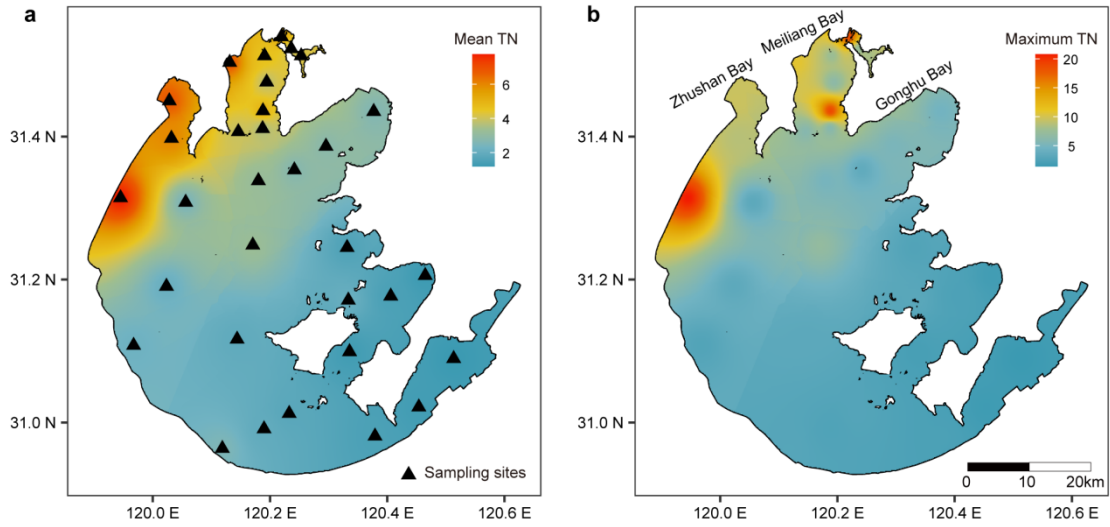
316



317

318 **Figure S18.** Water temperature and total nitrogen (TN) during 2007-2018 in Taihu Lake.  
 319 n = 32 sampling sites across the whole of Taihu Lake (Fig. S19a). The grey dots indicate  
 320 water temperature and TN for individual sites and black dots are the mean values for each  
 321 year. The boxes extend from 25<sup>th</sup> to 75<sup>th</sup> percentile (first and third quartiles), median is  
 322 marked by the line, and the ends of whiskers indicate the minimum and maximum values  
 323 within 1.5× the inter-quartile range from the first and third quartiles, respectively.

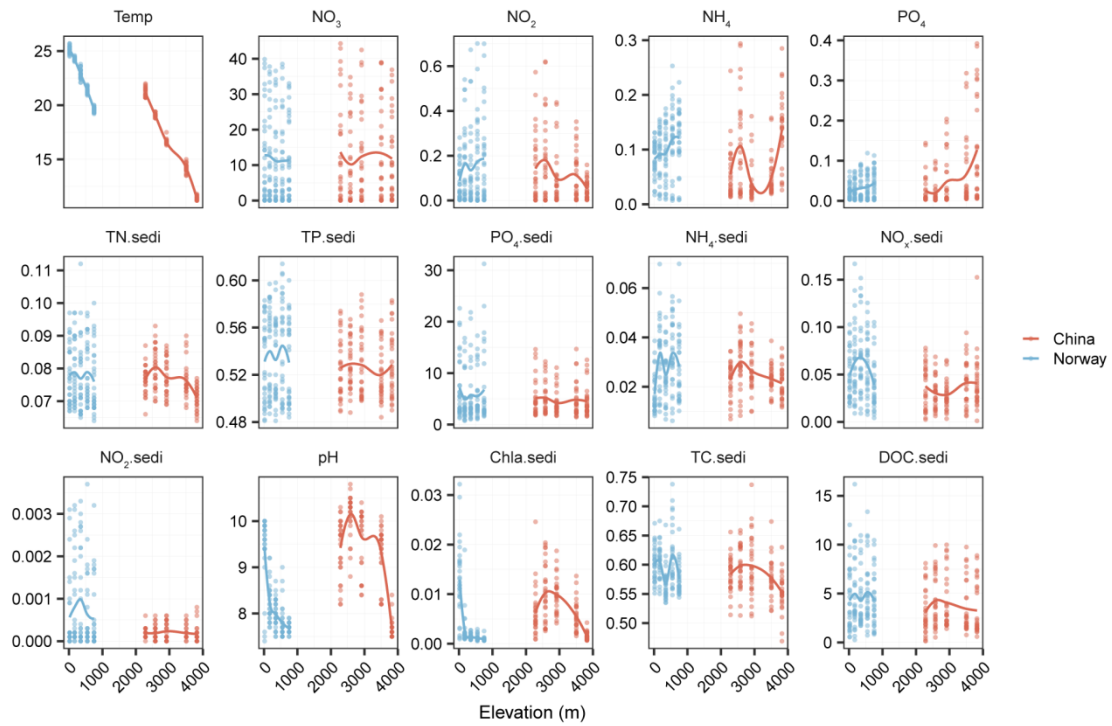
324



325

326 **Figure S19.** The distribution of mean (a) and maximum (b) total nitrogen (TN)  
 327 concentrations ( $\text{mg L}^{-1}$ ) in 2007 across the Taihu Lake. Triangles indicate 32 sampling  
 328 sites.

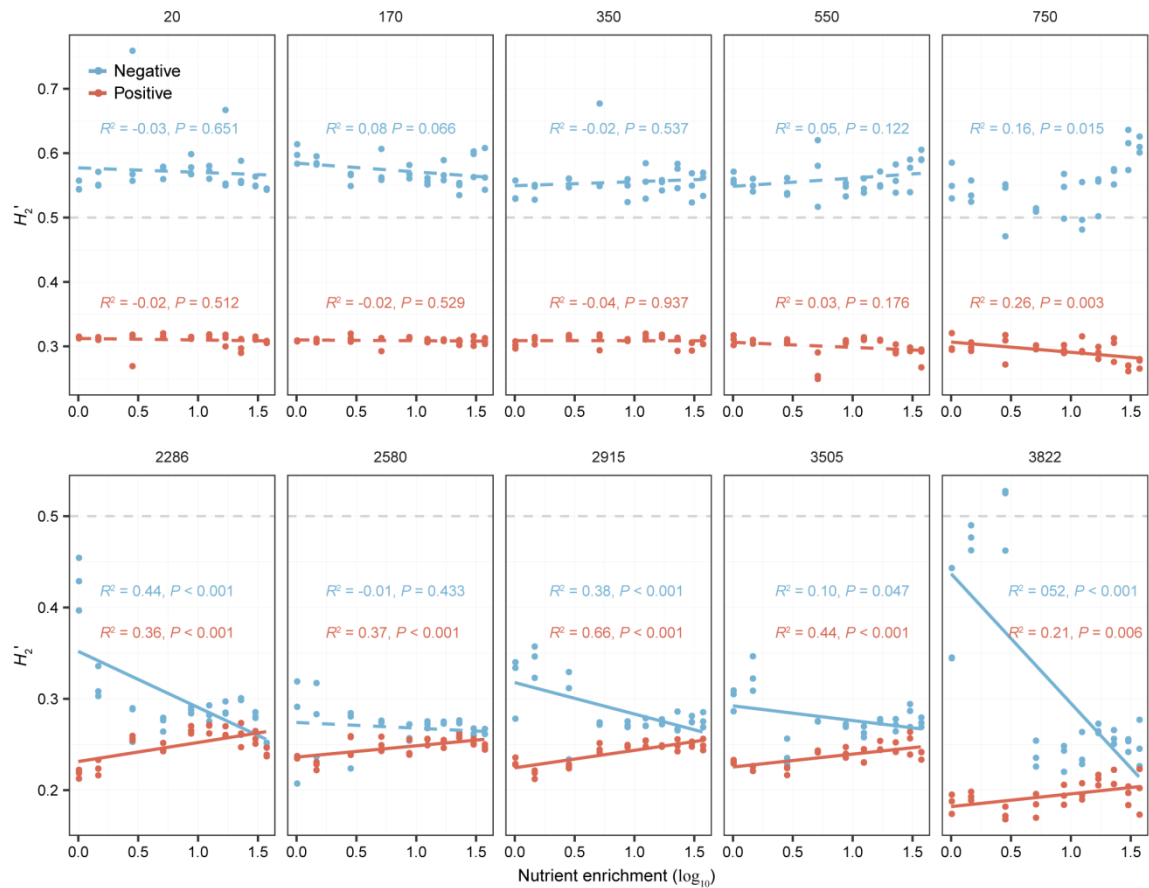
329



330

331 **Figure S20.** Environmental variables along the elevational gradients, as visualized with  
 332 loess regression models. The abbreviations of explanatory variables are detailed in Table  
 333 S1.

334



335

336 **Figure S21.** The effects of nutrient enrichment on the observed values of specialization  
 337  $H_2'$  of DOM-bacteria bipartite networks. We plotted the  $H_2'$  against the nutrient gradient  
 338 of nitrate for both negative (blue lines) and positive (red lines) networks for each  
 339 elevation in China or Norway, and their relationships are indicated by solid ( $P \leq 0.05$ ) or  
 340 dotted ( $P > 0.05$ ) lines using linear models with one-sided F-statistics. The horizontal  
 341 dashed lines indicate more specialized relations with a  $H_2'$  value above 0.5.

342

343 **References**

- 344 1. de Sassi C, Lewis OT, Tylianakis JM. Plant-mediated and nonadditive effects of two  
345 global change drivers on an insect herbivore community. *Ecology* 2012, **93**(8): 1892-  
346 1901.
- 347  
348 2. Glassman SI, Weihe C, Li J, Albright MBN, Looby CI, Martiny AC, *et al.*  
349 Decomposition responses to climate depend on microbial community composition. *Proc*  
350 *Natl Acad Sci U S A* 2018, **115**(47): 11994-11999.
- 351  
352 3. Wang J, Pan F, Soininen J, Heino J, Shen J. Nutrient enrichment modifies temperature-  
353 biodiversity relationships in large-scale field experiments. *Nat Commun* 2016, **7**: 13960.
- 354  
355 4. Jessup CM, Kassen R, Forde SE, Kerr B, Buckling A, Rainey PB, *et al.* Big questions,  
356 small worlds: microbial model systems in ecology. *Trends in Ecology & Evolution*  
357 2004, **19**(4): 189-197.
- 358