Supporting Information

Mechanistic Insights into the Reaction of Amidines With 1,2,3-Triazines and 1,2,3,5-Tetrazines

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I. General Methods

General Methods. All reagents and solvents were used as supplied without further purification unless otherwise noted. Preparative TLC (PTLC) and column chromatography were conducted using Millipore SiO₂ 60 F_{254} PTLC (0.5 mm) and Zeochem ZEOprep 60 ECO SiO₂ (40–63 μ m), respectively. Analytical TLC was conducting using Millipore SiO₂ 60 F254 TLC (0.250 mm) plates. ¹H and ¹³C NMR spectra were obtained using a Bruker Avance III HD 600 MHz spectrometer equipped with either a 5 mm QCI or 5 mm CPDCH probe. Mass spectrometry analysis was performed by direct sample injection on an Agilent G1969A ESI-TOF mass spectrometer.

II. ¹⁵N labeling experiments

Preparation of doubly ¹⁵N-labeled amidines 4a-e

4a was prepared according to a literature procedure.¹

$$R \xrightarrow{15}NH_4CI, K_2CO_3 \\ CI \xrightarrow{15}NH_2O, 25 \ ^\circC, 20 \ h} R \xrightarrow{0}_{15}NH_2$$

A solution of the corresponding acyl chloride (3 mmol, 1 equiv) in Et₂O (6 mL) was treated with a solution of K₂CO₃ (621 mg, 4.5 mmol, 1.5 equiv) and ¹⁵NH₄Cl (246 mg, 4.5 mmol, 1.5 equiv) in H₂O (6 mL). The mixture was stirred at 25 °C for 20 h before 2M KOH (20 mL) was added. The mixture was extracted by EtOAc (40 mL × 3). The organic layers were combined, dried over Na₂SO₄, then concentrated to provide corresponding ¹⁵N-labeled amides as white solids: R = cyclohexyl, reaction scaled up by 2-fold, 414 mg, 54%; R = *p*-MeOC₆H₄, 252 mg, 44%; R = *p*-CF₃C₆H₄, 547 mg, 84%.

$$\begin{array}{c} & \mathsf{Me_3O^+ BF_4^-, CH_2Cl_2} \\ O \\ \mathbb{R}^{-15}\mathsf{NH_2} \end{array} \xrightarrow{ \begin{array}{c} 25 \ ^\circ \mathsf{C}, \ 24 \ \mathsf{h} \end{array} \xrightarrow{ \begin{array}{c} 15} \mathsf{NH_2^+} \\ \mathbb{R}^{-15}\mathsf{NH_2} \end{array} \xrightarrow{ \begin{array}{c} \mathsf{BF_4^-} \end{array} \xrightarrow{ \begin{array}{c} \mathsf{EtOH}, \ 25 \ ^\circ \mathsf{C}, \ 24 \ \mathsf{h} \end{array} \xrightarrow{ \begin{array}{c} 15} \mathsf{NH} \\ \mathbb{R}^{-15}\mathsf{NH_2} \end{array} \xrightarrow{ \begin{array}{c} \mathsf{BF_4^-} \end{array} \xrightarrow{ \begin{array}{c} \mathsf{EtOH}, \ 25 \ ^\circ \mathsf{C}, \ 24 \ \mathsf{h} \end{array} \xrightarrow{ \begin{array}{c} \mathsf{NH} \\ \mathbb{R}^{-15}\mathsf{NH_2} \end{array}$$

In a flame-dried reaction vessel, corresponding ¹⁵N-labeled amide (1 mmol, 1 equiv) was added to a suspension of trimethyloxonium tetrafluoroborate (222 mg, 1.5 mmol, 1.5 equiv) in anhydrous CH_2Cl_2 (1 mL). The mixture was stirred under argon atmosphere at 25 °C for 24 h. Solvent was then removed to provide crude imidate salt. This crude was dissolved in EtOH (6 mL) and the mixture was treated with K_2CO_3 (414 mg, 3 mmol, 3 equiv) and ¹⁵NH₄Cl (163.5 mg, 3 mmol, 3 equiv). The mixture was stirred at 25 °C for 24 h and was then concentrated. 1M HCl (40 mL) was added to the residue and the aqueous phase was washed with CH₂Cl₂ (50 mL × 2). The aqueous phase was then basified by 2M KOH (50 mL) and extracted with CH₂Cl₂ (50 mL × 3). The organic layers were combined, dried over Na₂SO₄, then concentrated to provide corresponding doubly ¹⁵N-labeled amidines:

[¹⁵N]₂-Cyclohexylformamidine (4b)



4b (70 mg, 55% over two steps, viscous oil): ¹H NMR (500 MHz, CDCl₃, 298K) δ 2.07–2.03 (m, 1H), 1.93–1.89 (m, 2H), 1.84–1.80 (m, 2H), 1.72–1.69 (m, 1H), 1.32–1.27 (m, 4H), 1.23–1.18 (m, 1H); ¹³C NMR (125 MHz, CDCl₃, 298K) δ 172.4 (t, *J* = 11 Hz), 44.8, 30.9, 29.7, 26.0; HRMS ESI-TOF *m*/*z* 129.1172 ([M + H]⁺, C₇H₁₄[¹⁵N]₂ + H⁺ requires 129.1170).

[¹⁵N]₂-Benzamidine (4c)



4c (73 mg, 60% over two steps, viscous oil): ¹H NMR (500 MHz, CDCl₃, 298K) δ 7.60 (dd, 2H, J = 8.0, 1.5 Hz), 7.46 (t, 1H, J = 7.0 Hz), 7.42 (t, 2H, J = 7.0 Hz); ¹³C NMR (125 MHz, CDCl₃, 298K) δ 166.0 (t, J = 10 Hz), 136.4, 130.7, 128.9, 126.1; HRMS ESI-TOF *m*/*z* 123.0702 ([M + H]⁺, C₇H₈[¹⁵N]₂ + H⁺ requires 123.0701).

[¹⁵N]₂-4-Methoxybenzamidine (4d)



4d (108 mg, 71% over two steps, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 7.57 (d, 2H, J = 8.6 Hz), 6.92 (d, 2H, J = 8.6 Hz), 3.84 (s, 3H); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 165.4 (t, J = 9 Hz), 161.5, 128.8, 127.7, 114.1, 55.5; HRMS ESI-TOF m/z 153.0808 ([M + H]⁺, C₈H₁₀O[¹⁵N]₂ + H⁺ requires 153.0807).

^{[15}N]₂-4-Trifluoromethylbenzamidine (4e)



4e (36 mg, 20% over two steps, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 7.74 (d, 2H, J = 8.0 Hz), 7.69 (d, 2H, J = 8.0 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 163.7 (t, J = 16 Hz), 139.3, 131.9 (q, J = 33 Hz), 126.1, 125.3, 123.2 (q, J = 272 Hz); HRMS ESI-TOF *m*/*z* 191.0585 ([M + H]⁺, C₈H₇F₃[¹⁵N]₂ + H⁺ requires 191.0575).



Figure S1. Reactions between 1,2,3,5-tetrazine (1) or 1,2,3-triazines (2 and 3) with labeled amidines (4a-e)

Reactions between 4,6-diphenyl-1,2,3,5-tetrazine (1) and labeled amidines (4a-e)

A 40 mM solution of **1** (4.68 mg, 0.02 mmol, 1 equiv) in anhydrous CH_3CN (0.5 mL) was treated with the corresponding double ¹⁵N-labelled amidine **4a-e** (0.03 mmol, 1.5 equiv). The mixture was stirred at 25 °C for 2 h, and the solvent was removed under a gentle stream of N₂. The residue was purified by column chromatography (SiO₂, EtOAc/hexanes) or PTLC (EtOAc/hexanes) to provide **5a-e** as desired products.

[¹⁵N]-2-Benzyl-4,6-diphenyl-1,3,5-triazine and [¹⁵N]₂-2-Benzyl-4,6-diphenyl-1,3,5-triazine (5a)



Detailed in ref. S2.²

[¹⁵N]-2-Cyclohexyl-4,6-diphenyl-1,3,5-triazine and [¹⁵N]₂-2-Cyclohexyl-4,6-diphenyl-1,3,5-triazine (5b)



5b (6.24 mg, 99%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 8.67 (dd, 4H, *J* = 8.3 Hz, 1.4 Hz), 7.58 (t, 2H, *J* = 7.2 Hz), 7.54 (t, 4H, *J* = 7.2 Hz), 2.97–2.91 (m, 1H), 2.17–2.12 (m, 2H), 1.94–1.89 (m, 2H), 1.84–1.76 (m, 3H), 1.49 (qt, 2H, *J* = 12.9 Hz, 3.3 Hz), 1.38 (qt, 1H, *J* = 13.4 Hz, 3.3 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 183.1 (d, *J* = 2 Hz), 171.3 (m), 136.5 (t, *J* = 4 Hz), 132.4, 129.0, 128.7, 47.3 (d, *J* = 8 Hz), 31.4, 26.21, 26.19; HRMS ESI-TOF *m/z* 317.1782 ([M + H]⁺, C₂₁H₂₁N₂[¹⁵N] + H⁺ requires 317.1779).

[¹⁵N]-2,4,6-triphenyl-1,3,5-triazine and [¹⁵N]₂-2,4,6-triphenyl-1,3,5-triazine (5c)



5c (5.50 mg, 90%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 8.79 (d, 6H, J = 7.4 Hz), 7.63 (t, 3H, J = 7.1 Hz), 7.59 (t, 6H, J = 7.3 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 171.8 (m), 136.4 (m), 132.8, 129.3 (m), 128.8; HRMS ESI-TOF m/z 311.1309, 312.1297 ([M + H]⁺, C₂₁H₁₅N₂[¹⁵N] + H⁺ requires 311.1312, C₂₁H₁₅N[¹⁵N]₂ + H⁺ requires 312.1279).

[¹⁵N]-2-(4-Methoxyphenyl)-4,6-diphenyl-1,3,5-triazine and [¹⁵N]₂-2-(4-Methoxyphenyl)-4,6-diphenyl-1,3,5-triazine (5d)



5d (6.68 mg, 98%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 8.77 (dd, 4H, *J* = 8.2, 1.4 Hz), 8.75 (d, 2H, *J* = 8.9 Hz), 7.61 (t, 2H, *J* = 7.2 Hz), 7.57 (t, 4H, *J* = 7.2 Hz), 7.07 (d, 2H, *J* = 8.9 Hz), 3.92 (s, 3H); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 171.5 (m), 171.3 (m), 163.5, 136.6 (m), 132.5, 131.0 (m), 129.0 (m), 128.9 (m), 128.7, 114.1, 55.6; HRMS ESI-TOF *m/z* 341.1421, 342.1403 ([M + H]⁺, C₂₂H₁₇ON₂[¹⁵N] + H⁺ requires 341.1415, C₂₂H₁₇ON[¹⁵N]₂ + H⁺ requires 342.1385).

[¹⁵N]₂-2-(4-



5e (7.55 mg, 99%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 8.88 (d, 2H, J = 8.2 Hz), 8.77 (d, 4H, J = 7.1 Hz), 7.83 (d, 2H, J = 8.2 Hz), 7.64 (t, 2H, J = 7.2 Hz), 7.59 (t, 4H, J = 7.2 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 172.1 (m), 170.6 (m), 139.7 (m), 136.0 (m), 134.0 (q, J = 32 Hz), 132.9, 129.4 (m), 129.2 (m), 128.9, 125.7 (q, J = 4 Hz), 124.2 (q, J = 273 Hz); HRMS ESI-TOF m/z 379.1186, 380.1165 ([M + H]⁺, C₂₂H₁₄F₃N₂[¹⁵N] + H⁺ requires 379.1183, C₂₂H₁₄F₃N[¹⁵N]₂ + H⁺ requires 380.1153).

Reactions between 5-methoxycarbonyl-1,2,3-triazine (2) and labelled amidines (4a-e)

A 60 mM solution of **2** (8.4 mg, 0.06 mmol, 1.5 equiv) in anhydrous CH₃CN (1 mL) was treated with the corresponding double ¹⁵N-labeled amidine **2a-e** (0.04 mmol, 1 equiv). The mixture was stirred at 25 °C for 2 h, and the solvent was removed under a gentle stream of N₂. The residue was purified by column chromatography (SiO₂, EtOAc/hexanes) or PTLC (EtOAc/hexanes) to provide **6a-e** as desired products.

¹⁵N]-Methyl 2-benzylpyridine-5-carboxylate (6a)



Detailed in ref. S3.³

[¹⁵N]-Methyl 2-cyclohexylpyridine-5-carboxylate (6b)



6b (7.62 mg, 86%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.20–9.17 (m, 2H), 3.96 (s, 3H), 2.96 (tq, 1H, *J* = 11.8 Hz, 2.4 Hz), 2.01 (d, 2H, *J* = 13.0 Hz), 1.86 (dt, 2H, *J* = 13.2 Hz, 3.2 Hz), 1.75 (d, 1H, *J* = 12.8 Hz), 1.64 (qd, 2H, *J* = 12.4 Hz, 3.4 Hz), 1.42 (qt, 2H, *J* = 13.0 Hz, 3.3 Hz), 1.31 (qt, 1H, *J* = 12.8 Hz, 3.5 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 178.3 (d, *J* = 3 Hz), 164.7, 158.3 (m), 121.4 (d, *J* = 2 Hz), 52.7, 47.9 (d, *J* = 8 Hz), 31.9, 26.3, 26.0; HRMS ESI-TOF *m/z* 222.1263 ([M + H]⁺, C₁₂H₁₆O₂N[¹⁵N] + H⁺ requires 222.1255).

[¹⁵N]-Methyl 2-phenylpyridine-5-carboxylate (6c)



6c (7.85 mg, 92%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.34–9.31 (m, 2H), 8.53 (d, 2H, J = 6.3 Hz), 7.57–7.48 (m, 3H), 4.00 (s, 3H); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 167.4 (d, J = 2 Hz), 164.6, 158.6 (m), 136.7 (d, J = 9 Hz), 132.0, 129.1 (d, J = 2 Hz), 128.9, 121.6, 52.7; HRMS ESI-TOF *m*/*z* 216.0789 ([M + H]⁺, C₁₂H₁₀O₂N[¹⁵N] + H⁺ requires 216.0786).

¹⁵N]-Methyl 2-(4-methoxyphenyl)pyridine-5-carboxylate (6d)



6d (9.71 mg, 99%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.29–9.25 (m, 2H), 8.49 (d, 2H, *J* = 8.0 Hz), 7.01 (d, 2H, *J* = 8.0 Hz), 3.98 (s, 3H), 3.89 (s, 3H); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 167.1 (d, *J* = 2 Hz), 164.8, 163.0, 158.5 (m), 131.0 (d, *J* = 2 Hz), 129.4 (d, *J* = 9 Hz), 120.8 (d, *J* = 2 Hz), 114.3, 55.6, 52.6; HRMS ESI-TOF *m*/*z* 246.0894 ([M + H]⁺, C₁₃H₁₂O₃N[¹⁵N] + H⁺ requires 246.0891).

[¹⁵N]-Methyl 2-(4-trifluoromethylphenyl)pyridine-5-carboxylate (6e)



6e (10.81 mg, 96%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.37–9.33 (m, 2H), 8.66 (d, 2H, *J* = 7.8 Hz), 7.78 (d, 2H, *J* = 7.8 Hz), 4.01(s, 3H); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 166.0, 164.3, 158.5 (m), 139.9 (d, *J* = 9 Hz), 133.4 (q, *J* = 32 Hz), 129.4 (d, *J* = 2 Hz), 125.8 (q, *J*

= 4 Hz), 124.1 (q, J = 272 Hz), 122.4 (d, J = 2 Hz), 52.9; HRMS ESI-TOF m/z 284.0662 ([M + H]⁺, C₁₃H₉F₃O₂N[¹⁵N] + H⁺ requires 284.0659).

Reactions between 5-nitro-1,2,3-triazine (3) and labelled amidines (4a-e)

A 60 mM solution of **3** (7.5 mg, 0.06 mmol, 1.5 equiv) in anhydrous CH_3CN (1 mL) was treated with the corresponding double ¹⁵N-labeled amidine **4a-e** (0.04 mmol, 1 equiv). The mixture was stirred at 25 °C for 10 min, and the solvent was removed under a gentle stream of N₂. The residue was purified by column chromatography (SiO₂, CH₂Cl₂) to provide **7a-e** as desired products.

[¹⁵N]-2-Benzyl-5-nitropyridine (7a)



7a (8.19 mg, 95%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.42–9.40 (m, 2H), 7.36 (d, 2H, *J* = 7.0 Hz), 7.33 (t, 2H, *J* = 7.6 Hz), 7.26 (t, 1H, *J* = 7.6 Hz), 4.43 (d, 2H, *J* = 2.6 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 175.1 (d, *J* = 4 Hz), 152.9 (m), 140.8, 136.7, 129.4, 129.0, 127.4, 46.2 (d, *J* = 9 Hz); HRMS ESI-TOF *m*/*z* 217.0739 ([M + H]⁺, C₁₁H₉O₂N₂[¹⁵N] + H⁺ requires 217.0738).

^{[15}N]-2-Cyclohexyl-5-nitropyridine (7b)



7b (7.76 mg, 93%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.42–9.39 (m, 2H), 3.06 (tq, 1H, *J* = 11.8 Hz, 3.0 Hz), 2.06–2.01 (m, 2H), 1.91–1.87 (m, 2H), 1.80–1.76 (m, 1H), 1.66 (qd, 2H, *J* = 12.5 Hz, 3.4 Hz), 1.44 (qt, 2H, *J* = 13.0 Hz, 3.5 Hz), 1.32 (qt, 1H, *J* = 12.8 Hz, 3.6 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 180.1 (d, *J* = 4 Hz), 152.5 (m), 140.6 (d, *J* = 2 Hz), 47.9 (d, *J* = 8 Hz), 31.9, 26.1, 25.9; HRMS ESI-TOF *m*/*z* 209.1049 ([M + H]⁺, C₁₀H₁₃O₂N₂[¹⁵N] + H⁺ requires 209.1051).

[¹⁵N]-5-Nitro-2-phenyl-pyridine (7c)



7c (7.80 mg, 97%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.54–9.51 (m, 2H), 8.53 (dd, 2H, *J* = 8.4 Hz, 1.2 Hz), 7.60 (t, 2H, *J* = 7.3 Hz), 7.55 (t, 1H, *J* = 7.3 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 168.4 (d, *J* = 2 Hz), 153.0 (m), 140.7, 135.7 (d, *J* = 9 Hz), 133.1, 129.8 (d, *J* = 3 Hz), 129.1; HRMS ESI-TOF *m*/*z* 203.0580 ([M + H]⁺, C₁₀H₇O₂N₂[¹⁵N] + H⁺ requires 203.0581).

[¹⁵N]-2-(4-methoxyphenyl)-5-nitro-pyridine (7d)



7d (8.86 mg, 95%, yellow solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.46–9.44 (m, 2H), 8.52 (d, 2H, *J* = 8.2 Hz), 7.03 (d, 2H, *J* = 8.2 Hz), 3.91 (s, 3H); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 168.0 (d, *J* = 2 Hz), 163.9, 153.0 (m), 140.0, 131.9 (d, *J* = 3 Hz), 128.3 (d, *J* = 9 Hz), 114.6, 55.7; HRMS ESI-TOF *m*/*z* 233.0696 ([M + H]⁺, C₁₁H₉O₃N₂[¹⁵N] + H⁺ requires 233.0687).

[¹⁵N]-5-Nitro-2-(4-trifluoromethylphenyl)-pyridine (7e)



7e (10.03 mg, 93%, white solid): ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.58–9.56 (m, 2H), 8.69 (d, 2H, *J* = 9.0 Hz), 7.81 (d, 2H, *J* = 9.0 Hz); ¹³C NMR (151 MHz, CDCl₃, 298K) δ 167.0 (d, *J* = 2 Hz), 153.2 (m), 141.1, 138.7 (d, *J* = 10 Hz), 134.3 (q, *J* = 32 Hz), 130.1 (d, *J* = 3 Hz), 126.0 (q, *J* = 4 Hz), 123.9 (q, *J* = 273 Hz); HRMS ESI-TOF *m*/*z* 271.0455 ([M + H]⁺, C₁₁H₆F₃O₂N₂[¹⁵N] + H⁺ requires 271.0455).

Determination of product ratio by HRMS

Products of the isotope labeling experiments were subjected to HRMS analysis, where the ratio of signal intensity (peak area) of $M + H^+$ (single ¹⁵N-labeled product) and $M + 1 + H^+$ (double ¹⁵N-labeled product) was used to determine the ¹⁵N content in the reaction product with the existence of natural occurring isotopes taken into account following the equation below. Two parallel experiments are conducted to provide the average, summarized in Table S1. Data for products **5a** and **6a** were included in our previous reports.^{2,3}

Second ¹⁵N (%) = $\frac{Intensity_{M+1+H} - Intensity_{M+H} + \times Natural Abundance_{M+1+H} + Intensity_{M+H+} + Intensity_{M+H+} + Natural Abundance_{M+1+H} + Intensity_{M+H+} + Intensity_{M+H+} + Natural Abundance_{M+1+H+} + Intensity_{M+H+} + Intensity_{M+H+} + Natural Abundance_{M+1+H+} + Intensity_{M+H+} + Intensity_{M+H+} + Natural Abundance_{M+1+H+} + Intensity_{M+H+} + Intensity_{M+} + Intensity_{M+H+} + Intensity_{M+} + Intensity_{M+} + Intensity_{M+} + Intensity_{M+H+} + Intensity_{M+} + Intensity_{M+} + Intensity$

Product	Natural isotope abundance (M + 1 + H ⁺)	Intensity (M + H ⁺)	Intensity (M + 1 + H ⁺)	Second ¹⁵ N incorporation
5b		23861	89161	7.8%
50	23.9%	30194	112419	7.8%
Fo	22.0%	7111	8592	49.2%
90	23.8%	4844	5784	48.9%
Ed	24.0%	70523	89454	50.5%
au	24.9%	73127	93429	50.7%
Fa	24.0%	1448	2245	56.6%
ъe	24.8%	1117	1681	55.7%
6h	10.0%	64006	8249	< 0%
60	13.8%	51947	6546	< 0%
6c		15869	1975	< 0%
	13.7%	39190	4730	< 0%
6d	14.8%	39011	5043	< 0%
		36330	4515	< 0%
0.	14 70/	7460	1098	< 0%
0e	14.7%	9267	1290	< 0%
7-	10.0%	19840	2779	1.2%
/ d	12.8%	12654	1791	1.3%
76	10.0%	75492	10647	3.1%
70	12.9%	85246	12109	2.8%
7.	11.0%	23947	3617	0.8%
/C	11.9%	17232	2550	0.4%
7d	11 00/	49238	6189	0.7%
70	11.0%	58995	7225	0.2%
70	12.0%	4363	595	1.2%
7e	12.9%	2323	305	1.3%

Table S1. Determination of second incorporation ratio in products 5-7 by HRMS

Determination of product ratio of 5a-e by ¹³C NMR

Data for product 5a was included in our previous report.²

[¹⁵N]-2-cyclohexyl-4,6-diphenyl-1,3,5-triazine (5b)



[¹⁵N]-2,4,6-triphenyl-1,3,5-triazine (5c)



Single ¹⁵N labeled product would give a doublet and a singlet, resulting in three peaks (ratio 1 : 1 : 1), double ¹⁵N labeled product would give a doublet and a triplet, resulting in five peaks (ratio 1 : 4 : 2 : 4 : 1). The spectra observed reveals a mixture of single ¹⁵N and double ¹⁵N labelled product.



[¹⁵N]-2-(4-methoxyphenyl)-4,6-diphenyl-1,3,5-triazine (5d)



[¹⁵N]-2-(4-trfluoromethylphenyl)-4,6-diphenyl-1,3,5-triazine (5e)





III. Kinetic studies

The reactions between 4,6-diphenyl-1,2,3,5-tetrazine **1** (4 mM, 1.0 equiv) and amidines **8a-e** (8 mM, 2.0 equiv) were conducted in CD₃CN-CDCl₃ (V/V = 1:1) at 298 K. The conversion of starting materials and the formation of the products were monitored by ¹H NMR spectra. Clean conversion from reactant to product was observed with no intermediates or byproducts. The data points at low conversion range were observed to fit second-order reaction kinetics, and rate constants were fitted according to the following equation:

$$y = \frac{1}{[tetrazine]_0 - [amidine]_0} ln \frac{[amidine]_0([tetrazine]_0 - [triazine])}{[tetrazine]_0([amidine]_0 - [triazine])} = kt$$



Figure S2. Kinetic studies of reaction between 4,6-diphenyl-1,2,3,5-tetrazine **1** and *p*-methoxybenzamidine **8a** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = (7.57 \pm 0.10) \times 10^{-2} M^{-1} \cdot s^{-1} (R^2 = 0.997)$



Figure S3. Kinetic studies of reaction between 4,6-diphenyl-1,2,3,5-tetrazine 1 and *p*-methylbenzamidine **8b** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = (6.18 \pm 0.08) \times 10^{-2} M^{-1} \cdot s^{-1} (R^2 = 0.998)$



Figure S4. Kinetic studies of reaction between 4,6-diphenyl-1,2,3,5-tetrazine 1 and benzamidine 8c (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = (3.45 \pm 0.04) \times 10^{-2} \text{ M}^{-1} \cdot \text{s}^{-1} (\text{R}^2 = 0.998)$





Figure S5. Kinetic studies of reaction between 4,6-diphenyl-1,2,3,5-tetrazine 1 and *p*-bromobenzamidine **8d** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = (1.79 \pm 0.01) \times 10^{-2} \text{ M}^{-1} \cdot \text{s}^{-1} (\text{R}^2 = 0.998)$



Figure S6. Kinetic studies of reaction between 4,6-diphenyl-1,2,3,5-tetrazine **1** and *p*-trifluoromethylbenzamidine **8e** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = (0.76 \pm 0.01) \times 10^{-2} \text{ M}^{-1} \cdot \text{s}^{-1} (\text{R}^2 = 0.998)$

The reactions between 5-methoxycarbonyl-1,2,3-triazine **2** (0.68 mM, 1.0 equiv) and amidines **8ae** (1.36 mM, 2.0 equiv) were conducted in CD₃CN-CDCl₃ (V/V = 1:1) at 298 K. The conversion of starting materials and the formation of the products were monitored by ¹H NMR spectra. Clean conversion from reactant to product was observed with no intermediates or byproducts. The data points at low conversion range were observed to fit second-order reaction kinetics, and rate constants were fitted according to the following equation:



Figure S7. Kinetic studies of reaction between 5-methoxycarbonyl-1,2,3-triazine **2** and *p*-methoxybenzamidine **8a** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = 3.93 \pm 0.10 \text{ M}^{-1} \cdot \text{s}^{-1} (\text{R}^2 = 0.996)$



Figure S8. Kinetic studies of reaction between 5-methoxycarbonyl-1,2,3-triazine **2** and *p*-methylbenzamidine **8b** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = 2.22 \pm 0.05 \text{ M}^{-1} \cdot \text{s}^{-1} (R^2 = 0.997)$



Figure S9. Kinetic studies of reaction between 5-methoxycarbonyl-1,2,3-triazine **2** and benzamidine **8c** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = 1.50 \pm 0.04 \text{ M}^{-1} \cdot \text{s}^{-1}$ (R² = 0.994)



Figure S10. Kinetic studies of reaction between 5-methoxycarbonyl-1,2,3-triazine **2** and *p*-bromobenzamidine **8d** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = 0.55 \pm 0.02 \text{ M}^{-1} \cdot \text{s}^{-1} (\text{R}^2 = 0.994)$



Figure S11. Kinetic studies of reaction between 5-methoxycarbonyl-1,2,3-triazine **2** and *p*-trifluorobenzamidine **8e** (top: NMR spectra; bottom left: reaction progress over time; bottom right: linear fitting). $k = 0.22 \pm 0.01 \text{ M}^{-1} \cdot \text{s}^{-1} (R^2 = 0.995)$

IV. Kinetic Isotope Effect Studies



Preparation of methyl 4-methyl-1,2,3-triazine-5-carboxylate-6-d (6-d-triazine 9)

Partially deuterated dimethylformamide dimethyl acetal was prepared according to a literature procedure.⁴ A solution of dimethylformamide dimethyl acetal (10 mL, 76 mmol, 1 equiv) in mesitylene (8 mL) in a 500 mL flame-dried round-bottom flask was treated with CH₃OD (20 mL, 500 mmol, 7 equiv). The mixture was stirred at 25 °C for 3 h, and CH₃OH/CH₃OD was distilled off (bath T = 120 °C). The mixture was cooled to 25 °C, and methyl acetoacetate (8.8 g, 76 mmol, 1 equiv) was added. The reaction mixture was then warmed at 110 °C for 3 h and cooled to 25 °C. The resulting mixture was treated with hydrazine hydrate (3.7 mL, 76 mmol, 1 equiv) in EtOH (150 mL), and stirred at reflux (bath temp = 110 °C). The reaction mixture was cooled to 25 °C and concentrated. Water (100 mL) was added to the residue and the mixture was extracted by EtOAc (100 mL × 3). The organic layers were combined, dried over Na₂SO₄, and concentrated. The residue was purified by column chromatography (SiO₂, 60% EtOAc/hexanes) to provide methyl 3-methyl-1*H*-pyrazole-4-carboxylate-5-*d* (1.50 g, 13% over 3 steps) as a white solid with 46% deuterium incorporation at C5 as determined by ¹H NMR: ¹H NMR (600 MHz, CDCl₃, 298K) δ 9.32 (s, br, 1H), 7.97 (s, 0.54H), 3.84 (s, 3H), 2.57 (s, 3H).

Methyl 4-methyl-1,2,3-triazine-5-carboxylate-6-*d* (6-*d*-triazine **9**) was prepared according to a literature preparation⁵ of triazine **9** from methyl 3-methyl-1*H*-pyrazole-4-carboxylate-5-*d* to provide a purple oil (620 mg, 58%) with 44.63% deuterium incorporation at C6 as determined by quantitative ¹H NMR: ¹H NMR (600 MHz, CD₃CN, 298K) δ 9.23 (s, 0.55H), 3.96 (s, 3H), 2.91 (s, 3H).

Determination of ¹²C/¹³C Kinetic Isotope Effect in the Reaction of 9 with Benzamidine (8c)



Large scale reaction between 9 and 8c:

Methyl 4-methyl-1,2,3-triazine-5-carboxylate **9** (765 mg, 5 mmol, 1 equiv) was dissolved in CH₃CN (100 mL). Anisole (0.5 mL) was added as an internal standard. A solution of benzamidine (**8c**, 4 mmol, 480 mg, 0.8 equiv) in CH₃CN (5 mL) was added dropwise, and the mixture was stirred at 25 °C for 1 h when full conversion was observed. LC-MS analysis of the mixture before and after the reaction was conducted to determine the conversion. The mixture was concentrated and the residue was purified by column chromatography (SiO₂, 75% Et₂O/hexanes) to provide recovered **9** (123 mg, 16%) as a purple oil for NMR analysis.

Quantitative ¹³C NMR measurement:

The NMR samples of starting and recovered material were prepared identically, filling 10 mm NMR tubes to a constant height of 5 cm with a CDCl₃ solution of 100 mg material. T_1 determination by the inversion-recovery method was carried out on each NMR sample. The ¹³C spectra were recorded at 150.90 MHz on a Bruker Avance III HD 600 MHz spectrometer at room temperature, with inverse gated decoupling and 65 s (>5T₁) delays between calibrated pulses. 6 independent spectra of each sample were recorded, and integrations were determined numerically using a constant region for each peak that was 10 times the peak width at half height distance on either side of the peak. The integrations of the methyl carbon on the ester were set at 1000. The average integrations for the other carbons are shown in Table S2.

Chemical shift (ppm)	163.7	159.1	147.7	119.0	53.4	21.4
Carbon	Ca	C4	C6	C5	methoxy	Cb
		Starting ma	terial (n = 6)			
Intensity	977.8	1001.8	981.7	983.5	1000	1047.2
Standard deviation	0.15%	0.18%	0.13%	0.22%	0.00%	0.23%
	Reactio	on 1 (n = 6) (co	onversion: 65	% ± 2%)	-	
Intensity	984.7	1006.2	1019.4	992.1	1000	1038.8
Standard deviation	0.48%	0.37%	0.45%	0.31%	0.00%	0.27%
		R	/R ₀			
Value	1.007	1.004	1.038	1.009	1.000	0.991
Standard deviation	0.51%	0.41%	0.46%	0.38%	0.00%	0.36%
		¹² C/ ¹²	³ C KIE		-	-
KIE	1.007	1.004	1.039	1.009	1.000	0.992
Standard deviation	0.006	0.004	0.007	0.004	0.000	0.004
	Reactio	on 2 (n = 6) (co	onversion: 80	% ± 2%)		
Intensity	982.7	1014.4	1039.8	998.6	1000	1030.7
Standard deviation	0.22%	0.17%	0.12%	0.16%	0.00%	0.18%
		R	/R ₀		-	-
Value	1.005	1.012	1.059	1.015	1.000	0.984
Standard deviation	0.26%	0.25%	0.17%	0.27%	0.00%	0.29%
¹² C/ ¹³ C KIE						
KIE	1.003	1.008	1.037	1.009	1.000	0.990
Standard deviation	0.003	0.003	0.005	0.004	0.000	0.004
¹² C/ ¹³ C KIE (average)						
KIE (average)	1.005	1.006	1.038	1.009	1.000	0.991
Standard deviation	0.006	0.004	0.007	0.004	0.000	0.005

 Table S2. ¹²C/¹³C kinetic isotope effect determination.

Calculation of KIE: $KIE = \frac{\ln (1-F)}{\ln \left[\frac{(1-F)R}{R_0}\right]}$

Where R is the ¹³C proportion of the recovered starting material, R_0 is ¹³C proportion of the starting material, and F is the conversion of the material.

Determination of ¹H/²H Kinetic Isotope Effect for the Reaction of 9 with Benzamidine (8c)



Methyl 4-methyl-1,2,3-triazine-5-carboxylate-6-*d* (6-*d*-triazine **9**, 30.8 mg, 0.2 mmol, 1 equiv, 44.63% deuterium incorporation at C6) was dissolved in 1.5 mL CD₃CN. A solution of benzamidine (**8c**, 16.8 mg, 0.14 mmol, 0.7 equiv) in CD₃CN was added dropwise at 25 °C. The mixture was stirred at 25 °C for 1 h when full conversion was observed. The mixture was directly subjected to ¹H NMR analysis.

The NMR samples of starting material and reaction mixture were prepared and filling 10 mm NMR tubes to a constant height of 5 cm. T_1 determination by the inversion-recovery method was carried out on each NMR sample. The ¹H spectra were recorded at 600.13 MHz on a Bruker Avance III HD 600 MHz spectrometer at room temperature, with 40 s (>5 T_1) delays between calibrated pulses. Six (6) independent spectra of each sample were recorded, and the integrations were determined numerically using a constant region for each peak that was 10 times the peak width at half height distance on either side of the peak. The integration of the methyl protons on the triazine were set at 3000. The average integrations for the aromatic proton of starting material and reaction mixture are shown in Table S3. The conversion was determined through the integration of aromatic protons of 1,2,3-triazine and pyrimidine.

Starting material (n = 6)				
Signal	Triazine aromatic pro	oton Triazine	methyl proton	
	(δ = 9.23)	(δ	5 = 2.91)	
Intensity	553.68		3000	
Standard Deviation	0.49		0	
Deuterium Proportion		44.63%		
Standard Deviation		0.05%		
	Reaction 1	(n = 6)		
Signal	Triazine aromatic	Pyrimidine aromatic	Triazine methyl	
	proton (δ = 9.23)	proton (δ = 9.15)	proton (δ = 2.91)	
Intensity	606.95	1743.91	3000	
Standard Deviation	2.02	10.5	0	
Conversion		74.2%		
Standard Deviation		0.8%		
Deuterium Proportion		39.33%		
Standard Deviation		0.20%		
R/R ₀		0.8033		
Standard Deviation	0.0040			
KIE	0.861			
Standard Deviation	0.005			
	Reaction 2	(n = 6)		
Signal	Triazine aromatic	Pyrimidine aromatic	Triazine methyl	
	proton (δ = 9.23)	proton (δ = 9.15)	proton (δ = 2.91)	
Intensity	624.81	3020.70	3000	
Standard Deviation	4.40	12.1	0	
Conversion		82.9%		
Standard Deviation		0.6%		
Deuterium Proportion		37.52%		
Standard Deviation		0.44%		
R/R ₀		0.7449		
Standard Deviation		0.0074		
KIE		0.857		
Standard Deviation	0.006			
KIE (average)		0.859		
Standard Deviation		0.008		

Table S3. ¹H/²H kinetic isotope effect determination.

Calculation of KIE:
$$KIE = \frac{\ln (1-F)}{\ln \left[\frac{(1-F)R}{R_0}\right]}$$

Where R is the deuterium proportion of the recovered starting material, R_0 is deuterium proportion of the starting material, and F is the conversion of the protium proportion of the material.

IV. Computational Studies

Methods

DFT calculations were performed using Gaussian 09 Rev D.01.⁶ The M06-2X density functional⁷ was employed in combination with D3 empirical dispersion correction,⁸ as proposed by Grimme and coworkers.⁹ Single imaginary frequencies corresponding to the desired reaction coordinates were obtained only in the case of transition state (TS) calculations. No imaginary frequencies were obtained for all other structures. A quasi-harmonic correction to entropy was applied by setting all frequencies below 100 cm⁻¹ to 100 cm⁻¹ using GoodVibes.¹⁰

Conformer searches of all structures, including transition states, were performed using CREST 2.7.1 with the iMTD-GC workflow and the GFN2-xTB method.¹¹⁻¹² Conformers found by CREST were then optimized using M06-2X-D3/def2-TZV-SMD(acetonitrile). The lowest energy conformer was reoptimized using M06-2X-D3/def2-TZVP-SMD(acetonitrile). Structures were visualized using CYLview.¹³

Tunneling behavior was estimated with the correction by Skodje and coworkers¹⁴ using the PyTUN utility (<u>https://github.com/SJ-Ang/PyTUN</u>). The tunneling correction was converted into an apparent ΔG lowering using the Eyring equation at 298.15 K.

For computational determination of KIEs the atom was interest was substituted by the heavier isotope in both the starting materials and the transition state and a frequency/thermodynamic analysis was performed. Barrier heights were calculated and compared to the parent system without isotope substitution. $\Delta\Delta G^{\ddagger}$ were converted into relative rates using the Eyring equation at 298.15 K.

Additional retro-Diels-Alder pathways



Figure S12. Alternative retro-Diels–Alder pathways in the reaction between 1 and 4c. Energies relative to starting materials.



Figure S13. Alternative retro-Diels–Alder pathways in the reaction between 3 and 4c. Energies relative to starting materials.

Proton tunneling in TS5, TS7, and TS14



Figure S14. Tunneling corrections for TS5, TS7, and TS14.

Kinetic isotope effects



Figure S15. Energy barriers for A. 4c + 9 and B. 4c + 2.

Table S4: M06-2X-D3/def2-TZVP-SMD(acetonitrile) calculated Gibbs free energies for determination of KIEs on key atoms in the reactions 4c + 9 and 4c + 2.

Nucleophilic attack 4c + 9					
	4c	9	TS-S7		
	G(298.15) / hartree	G(298.15) / hartree	G(298.15) / hartree		
no heavy isotopes	-380.956545	-547.399970	-928.320158		
C4- ¹³ C	-380.956545	-547.400240	-928.320156		
C5- ¹³ C	-380.956545	-547.400237	-928.320109		
C6-13C	-380.956545	-547.400229	-928.320173		
C_a -13C	-380.956545	-547.400258	-928.320126		
C _b - ¹³ C	-380.956545	-547.400207	-928.320158		
C6- ² H	-380.956545	-547.403421	-928.323489		

	IM-S1	TS-S8
	G(298.15) / hartree	G(298.15) / hartree
no heavy isotopes	-928.336975	-928.293484
C4- ¹³ C	-928.337242	-928.293752
C5- ¹³ C	-928.337239	-928.293753
C6- ¹³ C	-928.337236	-928.293744
C_a -13C	-928.337259	-928.293768
C _b - ¹³ C	-928.337202	-928.293713
C6- ² H	-928.340613	-928.297121

Diels–Alder ringclosure 4c + 9

Nucleophilic attack **4c** + **2**

	4c	2	TS-S9
	G(298.15) / hartree	G(298.15) / hartree	G(298.15) / hartree
no heavy isotopes	-380.956545	-508.110381	-889.033774
C4- ¹³ C	-380.956545	-889.033994	-508.110643
C5- ¹³ C	-380.956545	-889.034042	-508.110653
C6- ¹³ C	-380.956545	-889.034031	-508.110641
C _a - ¹³ C	-380.956545	-889.034061	-508.110673
C4- ² H	-380.956545	-889.037206	-508.113867
C6- ² H	-380.956545	-889.037342	-508.113836

Diels–Alder ringclosure 4c + 2

	IM-S2	TS-S10
	G(298.15) / hartree	G(298.15) / hartree
no heavy isotopes	-889.050347	-889.004328
C4- ¹³ C	-889.050607	-889.004588
C5- ¹³ C	-889.050611	-889.004597
C6- ¹³ C	-889.050602	-889.004587
C _a -13C	-889.050631	-889.004615
C4- ² H	-889.053967	-889.007934
C6- ² H	-889.053749	-889.007686

Computational Data

The calculated energies for the lowest energy conformers are summarized in Table S5. Coordinates of lowest energy conformers are provided as *.xyz files.

Table S5: M06-2X-D3/def2-TZVP-SMD(acetonitrile) calculated energies of lowest energy conformers. Quasiharmonic correction was applied for Gibbs free energies.

Structure	E / hartree	ZPE / hartree	H / hartree	G(298.15) / hartree
1	-758.454580	0.215342	-758.225136	-758.278679
3	-484.801746	0.067153	-484.727014	-484.765965
4c	-381.065388	0.140748	-380.916149	-380.956540
5a	-1012.847178	0.337968	-1012.488897	-1012.554563
5c	-973.540656	0.309410	-973.213179	-973.274777
7c	-699.888118	0.161904	-699.714101	-699.763204
N ₂	-109.530714	0.005748	-109.521662	-109.543381
NH₃	-56.556070	0.034178	-56.518081	-56.539922
IM1	-1139.522252	0.360037	-1139.140155	-1139.208971
IM2	-1139.492609	0.360220	-1139.111053	-1139.177762
IM3	-1030.086497	0.350689	-1029.715321	-1029.781077
IM4	-1139.524463	0.360105	-1139.142267	-1139.211227
IM5	-1139.522601	0.360451	-1139.139913	-1139.209207
IM6	-1139.529006	0.360097	-1139.146550	-1139.215850
IM7	-1030.070869	0.347799	-1029.701611	-1029.769286
IM8	-1030.073121	0.348927	-1029.702999	-1029.769936
IM9	-1069.382510	0.376241	-1068.983416	-1069.053639
IM10	-1069.381985	0.377267	-1068.983259	-1069.051112
IM11	-1069.403235	0.378833	-1069.002690	-1069.070592
IM12	-1069.396575	0.378174	-1068.996521	-1069.064573
IM13	-865.889554	0.211882	-865.663126	-865.717412
IM14	-865.842470	0.211864	-865.615773	-865.669874
IM15	-756.440992	0.202429	-756.224838	-756.276818
IM16	-865.891369	0.211899	-865.663539	-865.720542
IM17	-865.889711	0.213108	-865.661178	-865.717183
IM18	-865.887076	0.212788	-865.658748	-865.715019
IM19	-756.431147	0.200332	-756.215887	-756.270860
IM20	-756.421580	0.199090	-756.207183	-756.262579
IM21	-756.423722	0.200336	-756.208574	-756.262970
IM22	-756.452900	0.203025	-756.235978	-756.288815
TS1	-1139.508266	0.356724	-1139.129152	-1139.198745
TS2	-1139.486759	0.358704	-1139.106959	-1139.173470
TS3	-1139.490542	0.357370	-1139.111708	-1139.178878
TS4	-1139.510319	0.355925	-1139.131860	-1139.201518
TS5	-1030.051975	0.343769	-1029.687628	-1029.753896
TS6	-1030.051641	0.346910	-1029.683947	-1029.750313
TS7	-1069.361625	0.371828	-1068.967627	-1069.036806

TS8	-1069.367137	0.375398	-1068.969817	-1069.038034
TS9	-1069.362397	0.375122	-1068.965122	-1069.033951
TS10	-865.863188	0.209012	-865.638312	-865.695018
TS11	-865.838135	0.211518	-865.612247	-865.665773
TS12	-865.841961	0.210449	-865.616907	-865.670624
TS13	-865.873996	0.209961	-865.648517	-865.704725
TS14	-756.411305	0.195829	-756.201374	-756.254525
TS15	-756.414431	0.198744	-756.201556	-756.254241
TS16	-756.404174	0.199671	-756.190401	-756.243647
TS-S1	-1139.482714	0.356057	-1139.104076	-1139.173981
TS-S2	-1139.499852	0.357533	-1139.120168	-1139.189161
TS-S3	-1139.504309	0.357343	-1139.124827	-1139.193804
TS-S4	-865.842482	0.208626	-865.618034	-865.674782
TS-S5	-865.867505	0.208181	-865.643310	-865.700281
TS-S6	-865.870417	0.209787	-865.645108	-865.701275

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VI. Spectra







7.61 7.61 7.60 7.59 7.45 7.45 7.45 7.45 7.42 7.42 7.42





^{220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0} f1 (ppm)



$\begin{array}{c} 8.8\\ 8.8.7\\ 7.5.6\\ 8.8.7\\ 7.5.6\\ 8.8.7\\ 7.5.6\\ 8.8.7\\ 7.5.6\\ 8.8.7\\ 7.5.6\\ 8.8.7\\ 7.5.6\\ 8.8.7\\ 7.5.6\\ 7.5.$







8.80 8.80 8.78 8.78 8.78 7.64 7.61 7.61 7.60 7.59 7.57















8.88 8.87 8.78 8.77 8.77 8.77 7.82 7.65 7.65 7.65 7.65 7.65 7.59 7.56 7.56



































$\begin{array}{c} 9.54\\ 9.53\\ 9.53\\ 9.53\\ 9.57\\ 8.57\\ 7.60\\ 7.60\\ 7.56\\ 7.55\\$







-3.91



















12.5 12.0 11.5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 -1. fl (ppm)