Supporting Information

for

A Multi-Pronged Evaluation of Aldehyde-Based Tripeptidyl Main Protease Inhibitors as SARS-CoV-2 Antivirals

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*Correspondence should be addressed to Henry Ji: <u>hji@sorrentotherapeutics.com</u>, Shiqing Xu: <u>shiqing.xu@tamu.edu</u> and Wenshe Ray Liu: <u>wsliu2007@tamu.edu</u> **Materials.** We purchased yeast extract from Thermo Fisher Scientific, tryptone from Gibco, Sub3 from Bachem, HEK 293T/17 cells from ATCC, DMEM with GlutaMax from Gibco, FBS from Gibco, polyethyleneimine from Polysciences, the trypsin-EDTA solution from Gibco. Chemicals used in this work were acquired from Sigma Aldrich, Chem Impex, Ambeed, A2B, etc.

M^{Pro} Expression and Purification. The expression plasmid pET28a-His-SUMO-M^{Pro} was constructed in a previous study. We used this construct to transform E. coli BL21(DE3) cells. A single colony grown on a LB plate containing 50 µg/mL kanamycin was picked and grown in 5 mL LB media supplemented with 50 µg/mL kanamycin overnight. We inoculated this overnight culture to 6 L 2YT media with 50 μ g/mL kanamycin. Cells were grown to OD₆₀₀ as 0.8. At this point, we added 1 mM IPTG to induce the expression of His-SUMO-M^{Pro}. Induced cells were let grown for 3 h and then harvested by centrifugation at 12,000 rpm, 4 °C for 30 min. We resuspended cell pellets in 150 mL lysis buffer (20 mM Tris-HCl, 100 mM NaCl, 10 mM imidazole, pH 8.0) and lysed the cells by sonication on ice. We clarified the lysate by centrifugation at 16,000 rpm, 4 °C for 30 min. We decanted the supernatant and mixed with Ni-NTA resins (GenScript). We loaded the resins to a column, washed the resins with 10 volumes of lysis buffer, and eluted the bound protein using elution buffer (20 mM Tris-HCl, 100 mM NaCl, 250 mM imidazole, pH 8.0). We exchanged buffer of the elute to another buffer (20 mM Tris-HCl, 100 mM NaCl, 10 mM imidazole, 1 mM DTT, pH 8.0) using a HiPrep 26/10 desalting column (Cytiva) and digested the elute using 10 units SUMO protease overnight at 4 °C. The digested elute was subjected to Ni-NTA resins in a column to remove His-tagged SUMO protease, His-tagged SUMO tag, and undigested His-SUMO-M^{Pro}. We loaded the flow-through onto a Q-Sepharose column and purified M^{Pro} using FPLC by running a linear gradient from 0 to 500 mM NaCl in a buffer (20 mM Tris-HCl, 1 mM DTT, pH 8.0). Fractions eluted from the Q-Sepharose column was concentrated and loaded onto a HiPrep 16/60 Sephacryl S-100 HR column and purified using a buffer containing 20 mM Tris-HCl, 100 mM NaCl, 1 mM DTT, and 1 mM EDTA at pH 7.8. The final purified was concentrated and stored in a -80 °C freezer.

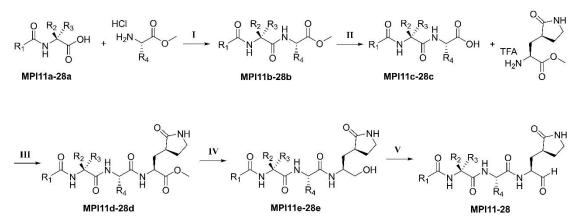
In Vitro M^{Pro} Inhibition Potency Characterizations of MPIs. For most MPIs, we conducted the assay using 20 nM M^{Pro} and 10 µM Sub3. For MPI13-14, 10 nM M^{Pro} was used. We dissolved all inhibitors in DMSO as 10 mM stock solutions. Sub3 was dissolved in DMSO as a 1 mM stock solution and diluted 100 times in the final assay buffer containing 10 mM Na_xH_yPO₄, 10 mM NaCl, 0.5 mM EDTA, and 1.25% DMSO at pH 7.6. We incubated M^{Pro} and an inhibitor in the final assay buffer for 30 min before adding the substrate to initiate the reaction catalyzed by M^{Pro} . The production format was monitored in a fluorescence plate reader with excitation at 336 nm and emission at 455 nm. More assay details can be found in a previous study.¹

X-Ray Crystallography Analysis of M^{Pro}-Inhibitor Complexes. The production of crystals of M^{pro}-inhibitor complexes wasfollowing the previous protocols.¹ The data of M^{pro} with MPI11, MPI12 and MPI24 were collected on a Rigaku R-AXIS IV++ image plate detector. The data of M^{pro} with MPI16, MPI18, MPI19, MPI22, MPI23, MPI25 and MPI27 were collected on a Bruker Photon II detector. The data of M^{pro} with MPI14, MPI20, MPI21, MPI26 and MPI28 were collected at the Advanced Light Source (ALS) beamline 5.0.2 using a Pilatus3 6M detector. The diffraction data were indexed, integrated and scaled with iMosflm or PROTEUM3.² All the structures were determined by molecular replacement using the structure model of the free enzyme of the SARS-CoV-2 M^{pro} [Protein Data Bank (PDB) ID code 7JPY] as the search model

using Phaser in the Phenix package.^{1,3} *JLigand* and *Sketcher* from the CCP4 suite were employed for the generation of PDB and geometric restraints for the inhibitors. The inhibitors were built into the Fo-Fc density by using *Coot*.⁴ Refinement of all the structures was performed with Real-space Refinement in Phenix.³ Details of data quality and structure refinement are summarized in Table S1. All structural figures were generated with PyMOL (https://www.pymol.org).

Cellular MPro Inhibition Potency Characterizations of MPIs. We grew HEK 293T/17 cells in high-glucose DMEM with GlutaMAX supplement and 10% FBS in 10 cm culture plates under 37 °C and 5% CO₂ to 80-90% confluency and then transfected cells with the pLVX-M^{Pro}-eGFP-2 plasmid. 30 mg/mL polyethyleneimine and the total of 8 µg of the plasmid in 500 µL opti-MEM media were used for transfection. We incubated transfected cells overnight. On the second day, we collected cells using 0.05% trypsin-EDTA to detach them from plates, resuspended collected cells in the original growth media, adjusted the cell density to $5 \cdot 10^5$ cells/mL, added 500 μ L adjusted cells to each well of a 48-well plate, and then added 100 μ L of a drug solution in DMEM. We incubated treated cells under 37 °C and 5% CO₂ for 72 h. After 72 h incubation, cells were collected using trypsinization and centrifugation. We resuspended collected cells in 200 (L PBS and analyzed cells with fluorescence using a Cytoflex Research Flow Cytometer based on the size scatters (SSC-A and SSC-H) and forward scatter (FSC-A). We gated cells based on SSC-A and FSC-A then with SSC-A and SSC-H. Fluorescence was detected with excitation at 488 nm and emission at 525 nm. All collected data were converted to csv files and analyzed using a selfprepared MATLAB script for massive data processing. We sorted the FITC-A column from lowest to highest. A 10⁶ cutoff was set to separate the column to two groups with higher than 10^6 as positive and lower than 10^6 as negative. We integrated the positive group and divided the total integrated fluorescence intensity by the total cell positive cell counts as Flu. Int. shown in all graphs. The standard deviation of positive fluorescence was calculated as well. All processed data were plotted and fitted to a fourparameter Hill equation in GraphPad 9.0 to obtain determined EC₅₀ values.

The Synthesis of MPIs. All reagents and solvents for the synthesis were purchased from commercial sources and used without purification. All glassware was flame-dried prior to use. Thin-layer chromatography (TLC) was carried out on aluminum plates coated with 60 F254 silica gel. TLC plates were visualized under UV light (254 or 365 nm) or stained with 5% phosphomolybdic acid. Normal phase column chromatography was carried out using a Yamazen Small Flash AKROS system. Analytical reverse phase HPLC was carried out on a Shimazu LC20 HPLC system with an analytical C18 column. Semipreparative HPLC was carried out the same system with a semipreparative C18 column. The mobile phases were H₂O with 0.1% formic acid (A) and acetone with 0.1% formic acid (B). NMR spectra were recorded on a Bruker AVANCE Neo 400 MHz or Varian INOVA 300 MHz spectrometer in specified deuterated solvents. High-resolution electrospray mass spectrometry was carried out on a Thermo Scientific Q Exactive Focus system. The purity of all compounds was confirmed by NMR and analytic HPLC-UV as ϵ 95%.



Scheme S1. General synthetic route to MPI11-28. Reagents and conditions: (I) HATU, DIPEA, DMF; (II) LiOH•H₂O, THF/H₂O; (III) HATU, DIPEA, DMP; (IV) LiBH₄, THF; (V) DMP, DCM

2-((S)-2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-4,4-(S)-methyl dimethylpentanoate (MPI11b). Methyl (S)-2-amino-4,4-dimethylpentanoate hydrochloride (0.4 g, 2.27 mmol, 1.1 equiv) and N-Cbz-L-valine (0.52 g, 2.07 mmol, 1.0 equiv) were dissolved in dry DMF (20 mL) and the reaction was cooled to 0 °C. HATU (1.01 g, 2.69 mmol, 1.3 equiv) and DIPEA (1.46 mL, 8.28 mmol, 4.0 equiv) were added, and the reaction mixture was allowed warm up to room temperature and stirred for 12 h. The mixture was then poured into water (50 mL) and extracted with ethyl acetate $(4 \times 20 \text{ mL})$. The organic layer was washed with aqueous hydrochloric acid 10% v/v (2×20 mL), saturated aqueous NaHCO₃ (2×20 mL), brine (2×20 mL) and dried over Na₂SO₄. The organic phase was evaporated to dryness and the crude material purified by silica gel column chromatography (15-50% EtOAc in n-hexane as the eluent) to afford pure **MPI11b** as a white solid (0.6 g, 73%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.36 (m, 5H), 6.39 (m, 1H), 5.43 (t, *J* = 10.6 Hz, 1H), 5.12 (s, 2H), 4.65 (td, J = 8.5, 3.6 Hz, 1H), 4.04 (q, J = 7.2 Hz, 1H), 3.73 (s, 3H), 2.24 - 2.01 (m, 1H), 1.80 (dd, J = 14.2, 3.5 Hz, 1H), 1.50 (dd, J = 14.3, 8.7 Hz, 1H), 1.07 - 0.90 (m, 15H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 173.5, 170.9, 156.4, 136.26, 128.5, 128.2, 128.0, 77.4, 77.0, 76.7, 67.0, 60.3, 52.3, 49.8, 46.0, 31.1, 30.7, 29.5, 19.1, 17.9.

(S)-2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-4,4dimethylpentanoic acid (MPI11c). The MPI11b (600 mg, 1.53 mmol, 1.0 equiv) was dissolved in THF/H₂O (1:1, 10 mL). LiOH•H₂O (153 mg, 3.82 mmol, 2.5 equiv) was added at 0 °C. The mixture was stirred at room temperature overnight. Then THF was removed on vacuum and the aqueous layer was acidified with 1 M HCl and extracted with dichloromethane (3 x 10 mL). The organic layer was dried over anhydrous Na₂SO₄ and concentrated to yield **MPI11c** as a white solid (415 mg, yield 70%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.42 (s, 1H), 7.26 (tdd, *J* = 9.1, 5.5, 2.5 Hz, 5H), 6.69 (d, *J* = 8.2 Hz, 1H), 5.82 (d, *J* = 9.3 Hz, 1H), 5.10 – 4.96 (m, 2H), 4.54 (td, *J* = 8.6, 3.2 Hz, 1H), 4.13 – 3.88 (m, 1H), 1.83 – 1.73 (m, 1H), 1.43 (dd, *J* = 14.7, 8.9 Hz, 1H), 1.18 (dt, *J* = 12.4, 7.1 Hz, 1H), 0.91 (s, 3H), 0.85 (m, 12H).

(5S,8S,11S)-methyl 5-isopropyl-8-neopentyl-3,6,9-trioxo-11-(((S)-2oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate (MPI11d). The methyl (S)-2-amino-3-((S)-2-oxopyrrolidin-3-yl)propanoate hydrochloride (110 mg, 0.494 mmol, 1.1 equiv) and the MPI11c (170 mg, 0.45 mmol, 1.0 equiv) were dissolved in anhydrous DMF (10 mL) and the reaction was cooled to 0 °C. HATU (222 mg, 0.585 mmol, 1.3 equiv) and DIPEA (0.32 mL, 1.81 mmol, 4.0 equiv) were added, and the reaction mixture was allowed to warm up to room temperature and stirred for 12 h. The mixture was then poured into water (20 mL) and extracted with ethyl acetate (4×20 mL). The organic layer was washed with aqueous hydrochloric acid 10% v/v (2×20 mL), saturated aqueous NaHCO₃ (2×20 mL), brine (2×20 mL) and dried over Na₂SO₄. The organic phase was evaporated to dryness and the crude material purified by silica gel column chromatography to afford pure **MPI11d** as a white solid (200 mg, yield 81%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.72 (d, *J* = 7.7 Hz, 1H), 7.32 – 7.21 (m, 5H), 6.88 – 6.60 (m, 2H), 5.47 (d, *J* = 9.2 Hz, 1H), 5.00 (s, 2H), 4.58 - 4.56 (m, 1H), 4.46 – 4.43 (m, 1H), 3.91 – 3.88 (m, 1H), 3.61 (s, 3H), 3.27 – 3.24 (m, 2H), 2.34 – 2.24 (m, 2H), 2.15 – 2.12 (m, 1H), 2.01 – 1.97 (m, 1H), 1.78 – 1.75 (m, 3H), 1.19 – 1.11 (m, 1H), 0.91 – 0.81 (m, 16H).

((S)-1-(((S)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-Benzyl yl)amino)-4,4-dimethyl-1-oxopentan-2-yl)amino)-3-methyl-1-oxobutan-2yl)carbamate (MPI11e). To a stirred solution of MPI11d (150 mg, 0.27 mmol, 1.0 equiv) in anhydrous THF (8 mL) was added LiBH₄ (2.0 M in THF, 0.412 mL, 0.54 mmol, 2.0 equiv) in several portions at 0 °C. The reaction mixture was then allowed to warm up to room temperature and stirred for an additional 2 h. The reaction was quenched by the dropwise addition of 1.0 M HCl (aq) (1.2 mL) with cooling in an ice bath. Removed THF in vacuo, and the mixture was diluted with H₂O and extracted with EtOAc, washed with sat. NaCl, dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (6% MeOH in CH₂Cl₂ as the eluent) to afford the pure product MPI11e as a white solid (102 mg, yield 71%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.84 (d, J = 9.3 Hz, 1H), 7.71 (d, J = 7.5 Hz, 1H), 7.28 – 7.24 (m, 5H), 6.86 (d, J = 4.4 Hz, 1H), 5.66 (d, J = 8.7 Hz, 1H), 5.07 – 4.94 (m, 2H), 4.60 – 4.58 (m, 1H), 4.08 (t, J = 8.2 Hz, 1H), 3.89 (dq, J = 11.9, 5.6 Hz, 1H), 3.77 (s, 1H), 3.59 (d, J = 10.9 Hz, 1H), 3.32 - 3.06 (m, 2H), 2.46 - 2.14 (m, 3H), 1.99 - 1.96 (m, 2H), 1.72-1.69 (m, 2H), 1.64 - 1.40 (m, 2H), 0.94 - 0.77 (m, 15H). ¹³C NMR (100 MHz, CDCl₃): δ 180.8, 173.6, 171.0, 156.6, 136.2, 128.5, 128.2, 128.0, 67.1, 65.3, 60.3, 51.1, 50.4, 46.6, 40.6, 38.2, 32.2, 31.4, 30.6, 29.6, 28.2, 19.1, 18.3.

((S)-1-(((S)-4,4-dimethyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-Benzvl yl)propan-2-yl)amino)pentan-2-yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (MPI11). To a solution of MPI11e (90 mg, 0.17 mmol, 1.0 equiv) in anhydrous CH₂Cl₂ (6 mL) was added NaHCO₃ (60 mg, 0.68 mmol, 4 equiv) and the Dess-Martin reagent (225 mg, 0.51 mmol, 3 equiv). The resulting mixture was stirred at RT for 12 h. Then the reaction was quenched with a saturated NaHCO₃ solution containing 10 % Na₂S₂O₃. The layers were separated. The organic layer was then washed with saturated brine solution, dried over anhydrous Na₂SO₄ and concentrated on vacuum. The residue was purified by column chromatography (6% MeOH in CH₂Cl₂ as the eluent) to afford the pure product MPI11 as a white solid (58 mg, yield 65%). ¹H NMR (400 MHz, Chloroform-d): δ 9.42 (s, 1H), 7.32 – 7.23 (m, 5H), 6.64 – 6.41 (m, 1H), 5.91 (d, J = 26.3 Hz, 1H), 5.43 (d, J = 8.9 Hz, 1H), 5.31 (d, J = 8.6 Hz, 1H), 5.03 (d, J = 7.8 Hz, 2H), 4.51 (dt, J = 8.6, 4.4 Hz, 1H), 4.26 (d, J = 7.0 Hz, 1H), 4.18 – 4.01 (m, 1H), 3.92 (dd, J = 8.4, 6.3 Hz, 1H), 3.29-3.26 (m, 2H), 2.44 - 2.25 (m, 2H), 2.16 - 2.05 (m, 1H),1.91 – 1.87 (m, 2H), 1.80 – 1.73 (m, 1H), 1.58 – 1.54 (m, 2H), 1.02 – 0.73 (m, 15H). ¹³C NMR (100 MHz, Chloroform-d): δ 199.5, 180.0, 173.8, 171.1, 156.6, 136.2, 128.5, 128.2, 128.0, 67.1, 60.6, 57.2, 50.9, 46.4, 40.6, 37.9, 31.0, 30.6, 29.9, 29.6, 28.2, 19.2, 18.1.

(S)-methyl 2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-4methylpent-4-enoate (MPI12b). MPI112b was prepared with methyl (S)-2-amino-4methylpent-4-enoate hydrochloride and *N*-Cbz-L-valine as a white solid following a similar procedure to MPI11b (yield 40%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.41-7.28 (m, 5H), 6.19 (t, *J* = 5.8 Hz, 1H), 5.33 (d, *J* = 9.2 Hz, 1H), 5.11 (s, 2H), 4.83 (s, 1H), 4.73 (s, 1H), 4.67 (td, *J* = 8.0, 5.6 Hz, 1H), 4.08 – 3.95 (m, 1H), 3.72 (s, 3H), 2.55 (dd, *J* = 14.1, 5.4 Hz, 1H), 2.40 (dd, *J* = 14.0, 8.4 Hz, 1H), 2.14 (dt, *J* = 13.4, 7.0 Hz, 1H), 1.71 (s, 3H), 1.69 -1.66 (m, 1H), 0.95 (dd, *J* = 18.8, 6.7 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 172.3, 170.9, 156.3, 140.2, 136.2, 128.6, 128.2, 128.1, 114.9, 67.1, 60.2, 52.3, 50.5, 40.4, 31.2, 21.8, 21.1, 19.1, 17.7.

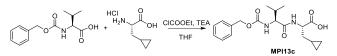
(S)-2-((S)-2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-4-methylpent-4-enoic acid (MPI12c). MPI12c was prepared as a white solid following a similar procedure to MPI11c (yield 83%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.41 – 7.28 (m, 5H), 6.41 (d, *J* = 7.3 Hz, 1H), 5.44 (d, *J* = 9.1 Hz, 1H), 5.11 (s, 2H), 4.83 (s, 1H), 4.75 (s, 1H), 4.73 – 4.63 (m, 1H), 2.62 (dd, *J* = 14.2, 5.2 Hz, 1H), 2.43 (dd, *J* = 14.2, 8.7 Hz, 1H), 1.94 – 1.84 (m, 1H), 1.72 (s, 3H), 0.94 (dd, *J* = 16.9, 6.8 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 174.8, 171.7, 156.6, 140.2, 136.1, 128.6, 128.3, 128.0, 127.0, 114.9, 68.0, 67.2, 60.3, 50.5, 40.0, 31.0, 25.6, 21.8, 19.1, 17.9.

(5S,8S,11S)-methyl 5-isopropyl-8-(2-methylallyl)-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate (MPI12d). MPI12d was prepared as a white solid following a similar procedure to MPI11d (yield 63%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.82 (d, *J* = 7.7 Hz, 1H), 7.41 – 7.28 (m, 5H), 7.06 (d, *J* = 8.5 Hz, 1H), 6.67 (s, 1H), 5.45 (d, *J* = 8.8 Hz, 1H), 5.09 (s, 2H), 4.75 (s, 1H), 4.71 (s, 1H), 4.54 (ddd, *J* = 11.6, 7.8, 3.4 Hz, 1H), 4.05 – 3.95 (m, 1H), 3.70 (s, 3H), 3.31 (p, *J* = 8.1, 6.7 Hz, 2H), 2.58 (dd, *J* = 14.1, 4.8 Hz, 1H), 2.45 – 2.28 (m, 3H), 2.25 – 2.05 (m, 2H), 1.90 – 1.73 (m, 2H), 1.72 (s, 3H), 0.91 (dd, *J* = 18.8, 6.8 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 179.8, 172.0, 171.8, 171.2, 156.5, 140.9, 136.2, 128.6, 128.2, 128.0, 114.3, 67.1, 60.6, 52.4, 51.3, 41.1, 40.5, 38.3, 33.3, 30.9, 28.1, 21.9, 19.2, 18.6, 17.3.

Benzyl ((S)-1-(((S)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-4-methyl-1-oxopent-4-en-2-yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (MPI12e). MPI12e was prepared as a white solid following a similar procedure to MPI11e (yield 70%). ¹H NMR (400 MHz, Methanol- d_4) δ 7.36 – 7.15 (m, 5H), 5.01 (q, *J* = 12.5 Hz, 2H), 4.69 (d, *J* = 8.8 Hz, 2H), 4.41 (dd, *J* = 9.5, 5.6 Hz, 1H), 3.84 (dd, *J* = 16.3, 7.9 Hz, 2H), 3.44 (dd, *J* = 11.2, 5.4 Hz, 1H), 3.36 (dd, *J* = 11.0, 6.4 Hz, 1H), 3.20 – 3.12 (m, 2H), 2.50 – 2.16 (m, 4H), 2.00 – 1.80 (m, 2H), 1.65 (s, 4H), 1.45 (ddd, *J* = 14.2, 11.1, 3.1 Hz, 1H), 0.84 (dd, *J* = 11.7, 6.8 Hz, 6H). ¹³C NMR (100 MHz, Methanol- d_4) δ 181.2, 172.8, 172.6, 157.5, 141.0, 136.8, 128.1, 127.6, 127.5, 112.9, 66.5, 64.1, 61.0, 52.0, 49.3, 40.1, 39.4, 38.1, 32.2, 30.4, 27.6, 20.8, 18.3, 17.1.

Benzyl ((S)-3-methyl-1-(((S)-4-methyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)pent-4-en-2-yl)amino)-1-oxobutan-2-yl)carbamate (MPI12). MPI12 was prepared as a white solid following a similar procedure to MPI11 (yield 60%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.48 (s, 1H), 8.33 – 8.20 (m, 1H), 7.42 – 7.29 (m, 5H), 6.63 (d, *J* = 7.8 Hz, 1H), 6.05 (s, 1H), 5.34 (d, *J* = 8.2 Hz, 1H), 5.10 (s, 2H), 4.82 (t, *J* = 1.7 Hz, 1H), 4.75 (s, 1H), 4.68 (td, *J* = 8.5, 5.7 Hz, 1H), 4.33 (d, *J* = 8.5 Hz, 1H), 4.02 (dd, *J* = 8.2, 5.8 Hz, 1H), 3.33 (dt, *J* = 8.9, 4.3 Hz, 2H), 2.62 (dd, *J* = 14.1, 5.5 Hz, 1H), 2.53 – 2.32 (m, 3H), 2.21 – 2.09 (m, 1H), 2.05 – 1.88 (m, 2H), 1.87 – 1.82 (m, 1H), 1.75 (s, 3H), 0.93 (dd, *J* = 23.8, 6.8 Hz, 6H).

¹³C NMR (100 MHz, Chloroform-*d*): δ 199.7, 180.0, 172.4, 171.4, 156.6, 140.8, 136.2, 128.5, 128.2, 128.0, 114.4, 67.1, 60.6, 57.5, 51.4, 41.0, 40.6, 38.0, 30.9, 29.9, 28.3, 21.9, 19.2, 17.7.



(S)-2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-3cyclopropylpropanoic acid (MPI13c). *N*-Cbz-L-valine (500 mg, 2.0 mmol, 1.0 equiv), ethyl chloroformate (268 μ L, 2.8 mmol, 1.4 equiv) and TEA (836 μ L, 6.0 mmol, 3.0 equiv) were dissolved in anhydrous THF (30 mL). After stirring at 0 °C for 0.5 h, L-cyclopropylalanine (386 mg, 4.0 mmol, 1.5 equiv) in water was added. The reaction mixture was stirred for 0.5 h at 0 °C. The THF was removed in vacuo and the aqueous layer was acidified with 1.0 M HCl. The aqueous layer was extracted with ethyl acetate. The combined organic layers were dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was recrystallized from CHCl₃ and hexane to yield the pure product **MPI13c** as a white solid (380 mg, yield 52%).

Methyl (5S,8S,11S)-8-(cyclopropylmethyl)-5-isopropyl-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate (MPI13d). MPI13d was prepared as a white solid following a similar procedure to MPI11d (yield 51%).

Benzyl ((S)-1-(((S)-3-cyclopropyl-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3yl)propan-2-yl)amino)-1-oxopropan-2-yl)amino)-3-methyl-1-oxobutan-2yl)carbamate (MPI13e). MPI13e was prepared as a white solid following a similar procedure to MPI11e (yield 75%). ¹H NMR (400 MHz, Methanol-*d*4) δ 7.3 – 7.1 (m, 5H), 5.0 (d, *J* = 4.1 Hz, 2H), 4.3 (t, *J* = 7.3 Hz, 1H), 3.9 (d, *J* = 7.3 Hz, 2H), 3.5 – 3.3 (m, 2H), 3.1 (dt, *J* = 16.3, 5.5 Hz, 2H), 2.4 (dd, *J* = 14.7, 6.6 Hz, 1H), 2.2 (dt, *J* = 15.7, 7.8 Hz, 1H), 2.0 – 1.8 (m, 2H), 1.6 (dq, *J* = 20.5, 7.7, 7.0 Hz, 2H), 1.5 – 1.3 (m, 2H), 0.8 (dd, *J* = 9.4, 6.7 Hz, 6H), 0.7 (p, *J* = 6.9 Hz, 1H), 0.3 (t, *J* = 10.5 Hz, 2H), 0.0 (dq, *J* = 7.5, 4.0 Hz, 2H). ¹³C NMR (100 MHz, Methanol-*d*4) δ 181.3, 172.9, 172.7, 157.4, 136.8, 128.1, 127.6, 127.5, 66.4, 64.1, 60.7, 54.3, 49.1, 40.1, 38.1, 36.6, 32.2, 30.6, 27.6, 18.4, 17.3, 7.3, 3.8, 3.6.

Benzyl ((S)-1-(((S)-3-cyclopropyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)propan-2-yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (MPI13). MPI13 was prepared as a white solid following a similar procedure to MPI11 (yield 50%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.4 (s, 1H), 8.3 (d, *J* = 6.5 Hz, 1H), 7.2 (s, 5H), 7.2 (s, 1H), 6.6 (d, *J* = 23.9 Hz, 1H), 5.7 – 5.5 (m, 1H), 5.1 – 4.9 (m, 2H), 4.6 (dq, *J* = 18.1, 9.9, 8.6 Hz, 1H), 4.4 – 4.2 (m, 1H), 4.0 (h, *J* = 6.9, 5.7 Hz, 1H), 3.3 (s, 1H), 3.2 (d, *J* = 10.8 Hz, 1H), 2.4 (t, *J* = 8.4 Hz, 1H), 2.3 (s, 1H), 2.1 (s, 1H), 2.0 (dp, *J* = 25.6, 9.2, 7.8 Hz, 2H), 1.9 – 1.6 (m, 2H), 1.5 – 1.4 (m, 1H), 0.8 (dd, *J* = 12.2, 6.6 Hz, 6H), 0.7 – 0.5 (m, 1H), 0.3 (t, *J* = 9.5 Hz, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 199.5, 180.0, 172.7, 171.3, 156.6, 136.2, 128.5, 128.2, 128.0, 67.1, 60.5, 53.7, 50.7, 40.6, 38.0, 37.6, 31.1, 29.8, 28.4, 19.2, 17.9, 7.3, 4.5, 4.4.

Methyl (S)-2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-3-(furan-2-yl)propanoate (MPI14b). MPI14b was prepared with methyl (S)-2-amino-3-(furan-2-yl)propanoate hydrochloride and *N*-Cbz-L-valine as a white solid following a similar procedure to MPI11b (yield 80%). ¹H NMR (400 MHz, Methanol-*d*₄) δ 7.32 – 7.09 (m, 6H), 6.15 (dd, *J* = 3.2, 1.9 Hz, 1H), 6.02 (d, *J* = 3.2 Hz, 1H), 4.99 (s, 2H), 4.59 (dd, *J* = 8.1, 5.5 Hz, 1H), 3.84 (d, *J* = 7.1 Hz, 1H), 3.59 (s, 3H), 3.14 – 2.90 (m, 2H), 2.02 - 1.77 (m, 1H), 0.82 (dd, J = 11.5, 6.8 Hz, 6H). ¹³C NMR (100 MHz, Methanol- d_4) δ 172.73, 171.40, 150.58, 141.76, 128.09, 127.46, 109.92, 107.20, 48.25, 48.04, 47.82, 47.61, 47.40, 47.19, 46.97, 30.61, 29.49, 18.25.

(S)-2-((S)-2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-3-(furan-2-yl)propanoic acid (MPI14c). MPI14c was prepared as a white solid following a similar procedure to MPI11c (yield 88%). ¹H NMR (400 MHz, Methanol- d_4) δ 7.30 – 7.14 (m, 6H), 6.19 – 6.08 (m, 1H), 6.02 (d, J = 3.2 Hz, 1H), 4.98 (s, 2H), 4.58 (dd, J = 8.2, 5.0 Hz, 1H), 3.92 – 3.76 (m, 1H), 3.11 (dd, J = 15.3, 5.1 Hz, 1H), 2.97 (dd, J = 15.3, 8.2 Hz, 1H), 1.99 – 1.82 (m, 1H), 0.82 (dd, J = 14.3, 6.8 Hz, 6H). ¹³C NMR (100 MHz, Methanol- d_4) δ 172.61, 172.49, 150.87, 141.63, 128.10, 127.63, 127.45, 109.89, 107.09, 66.31, 60.59, 48.27, 48.06, 47.84, 47.63, 47.42, 47.21, 46.99, 30.63, 18.32.

Methyl (5S,8S,11S)-8-(furan-2-ylmethyl)-5-isopropyl-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate (MPI14d). MPI14d was prepared as a white solid following a similar procedure to MPI11d (yield 80%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.67 (d, J = 7.2 Hz, 1H), 7.38 – 7.22 (m, 5H), 6.90 (d, J = 7.9 Hz, 1H), 6.23 – 6.11 (m, 1H), 6.03 (d, J = 3.2 Hz, 1H), 5.86 (s, 1H), 5.26 (d, J = 8.0 Hz, 1H), 5.04 (d, J = 5.1 Hz, 2H), 4.74 (d, J = 7.3 Hz, 1H), 4.45 (s, 1H), 3.95 (t, J = 7.1 Hz, 1H), 3.64 (s, 3H), 3.24 (d, J = 8.8 Hz, 1H), 3.17 – 3.01 (m, 2H), 2.31 (s, 1H), 2.19 (s, 1H), 2.07 (dd, J = 13.3, 6.9 Hz, 2H), 1.86 – 1.68 (m, 2H), 1.60 (s, 3H), 1.41 (d, J = 22.6 Hz, 4H), 0.96 – 0.74 (m, 6H).

Benzyl ((S)-1-(((S)-3-(furan-2-yl)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl) propan-2-yl)amino)-1-oxopropan-2-yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (MPI14e). MPI14e was prepared as a white solid following a similar procedure to MPI11e (yield 37%). ¹H NMR (400 MHz, Methanol-*d*₄) δ 7.25 (s, 6H), 6.24 - 6.14 (m, 1H), 6.05 (d, *J* = 3.1 Hz, 1H), 5.09 - 4.86 (m, 2H), 4.52 (d, *J* = 7.9 Hz, 2H), 3.80 (t, *J* = 9.6 Hz, 2H), 3.48 - 3.25 (m, 2H), 3.11 - 2.88 (m, 2H), 2.39 - 2.02 (m, 2H), 2.03 - 1.73 (m, 2H), 1.73 - 1.55 (m, 1H), 1.51 - 1.32 (m, 1H), 1.26 (dd, *J* = 6.8, 2.3 Hz, 1H), 0.80 (dd, *J* = 9.1, 6.5 Hz, 6H). ¹³C NMR (100 MHz, Methanol-*d*₄) δ 181.23, 172.74, 171.62, 157.61, 151.02, 141.73, 136.74, 128.10, 127.66, 127.57, 109.96, 107.24, 66.57, 64.01, 61.10, 52.89, 48.26, 48.05, 47.83, 47.62, 47.41, 47.20, 46.98, 40.11, 38.05, 32.13, 29.99 (d, *J* = 65.0 Hz), 27.59, 18.23, 17.08.

Benzyl ((S)-1-(((S)-3-(furan-2-yl)-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)propan-2-yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (MPI14). MPI14 was prepared as a white solid following a similar procedure to MPI11 (yield 80%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.34 (s, 1H), 8.22 (d, *J* = 6.8 Hz, 1H), 7.27 (q, *J* = 7.8, 5.8 Hz, 5H), 7.02 (d, *J* = 8.1 Hz, 1H), 6.16 (d, *J* = 2.5 Hz, 2H), 6.03 (d, *J* = 3.1 Hz, 1H), 5.40 (t, *J* = 11.0 Hz, 1H), 5.03 (d, *J* = 6.8 Hz, 2H), 4.79 (q, *J* = 6.7 Hz, 1H), 4.22 (d, *J* = 10.4 Hz, 1H), 4.00 – 3.88 (m, 1H), 2.25 (s, 2H), 2.17 – 2.02 (m, 1H), 1.76 (d, *J* = 35.2 Hz, 5H), 0.84 (dd, *J* = 25.3, 6.8 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 199.96, 179.97, 171.32, 171.18, 150.71, 142.04, 136.11, 128.60, 128.29, 128.11, 110.41, 108.07, 67.26, 52.41, 50.86, 40.56, 30.70, 28.57, 19.26, 17.59.

Methyl (S)-2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-3-(thiophen-2-yl)propanoate (MPI15b). MPI15b was prepared with methyl (S)-2amino-3-(thiophen-2-yl)propanoate and *N*-Cbz-L-valine as a white solid following a similar procedure to MPI11b (yield 72%). ¹H NMR (400 MHz, Methanol- d_4) δ 7.37 – 7.13 (m, 5H), 7.11 – 7.03 (m, 1H), 6.78 (dd, J = 4.6, 2.9 Hz, 2H), 4.99 (d, J = 2.9 Hz, 2H), 4.58 (dd, J = 7.9, 5.3 Hz, 1H), 3.90 – 3.74 (m, 1H), 3.60 (s, 3H), 3.27 (dd, J =14.9, 5.3 Hz, 1H), 3.21 – 3.12 (m, 3H), 1.91 (s, 1H), 0.82 (dd, J = 8.4, 6.7 Hz, 6H). ¹³C NMR (100 MHz, Methanol- d_4) δ 172.79, 171.25, 138.12, 136.86, 128.10, 127.63, 127.47, 66.30, 60.62, 53.75, 51.40, 48.26, 48.04, 47.83, 47.62, 47.41, 47.19, 46.98, 31.05, 30.62, 18.28, 17.13.

(S)-2-(((benzyloxy)carbonyl)amino)-3-methylbutanamido)-3-(thiophen-2-yl)propanoic acid (MPI15c). MPI15c was prepared as a white solid following a similar procedure to MPI11c (yield 60%). ¹H NMR (400 MHz, Methanol- d_4) δ 7.44 – 7.29 (m, 5H), 7.19 (dd, J = 4.9, 1.5 Hz, 1H), 6.97 – 6.83 (m, 2H), 5.12 (d, J = 2.6 Hz, 2H), 4.69 (dd, J = 7.9, 4.9 Hz, 1H), 3.99 (d, J = 7.1 Hz, 1H), 3.43 (dd, J = 15.0, 4.8 Hz, 1H), 3.31 – 3.22 (m, 1H), 2.14 – 1.93 (m, 1H), 1.08 – 0.86 (m, 6H).

Methyl (5S,8S,11S)-5-isopropyl-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-8-(thiophen-2-ylmethyl)-2-oxa-4,7,10-triazadodecan-12-oate (MPI15d). MPI15d was prepared as a white solid following a similar procedure to MPI11d (yield 90%). ¹H NMR (400 MHz, Methanol- d_4) δ 7.35 – 7.18 (m, 5H), 7.08 (dd, J = 5.0, 1.3 Hz, 1H), 6.84 – 6.70 (m, 1H), 5.03 – 4.95 (m, 2H), 4.58 – 4.48 (m, 1H), 4.44 – 4.32 (m, 1H), 3.88 – 3.73 (m, 1H), 3.60 (d, J = 6.3 Hz, 3H), 3.14 (d, J = 3.4 Hz, 2H), 2.39 – 2.25 (m, 1H), 2.24 – 2.09 (m, 1H), 2.10 – 2.01 (m, 1H), 1.99 – 1.83 (m, 1H), 1.67 (s, 2H), 1.19 (d, J = 3.6 Hz, 2H), 0.89 (dd, J = 9.1, 6.8 Hz, 2H), 0.78 (dd, J = 8.4, 6.7 Hz, 4H). ¹³C NMR (100 MHz, Methanol- d_4) δ 180.21, 172.65, 172.01, 171.69, 157.27, 138.44, 136.84, 128.14, 127.66, 127.51, 126.54, 126.36, 124.01, 66.40, 51.58, 50.54, 48.32, 48.11, 47.90, 47.69, 47.47, 47.26, 47.05, 32.51, 31.41, 27.30, 18.40, 17.21.

Benzyl ((S)-1-(((S)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-1-oxo-3-(thiophen-2-yl)propan-2-yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (MPI15e). MPI15e was prepared as a white solid following a similar procedure to MPI11e (yield 53%). ¹H NMR (400 MHz, Methanol-*d*₄) δ 7.32 – 7.15 (m, 5H), 7.10 (d, *J* = 4.8 Hz, 1H), 6.80 (d, *J* = 4.9 Hz, 2H), 5.08 – 4.88 (m, 2H), 4.48 (s, 1H), 3.80 (d, *J* = 6.7 Hz, 2H), 3.30 (d, *J* = 23.7 Hz, 2H), 3.17 – 3.00 (m, 3H), 2.24 (dd, *J* = 24.9, 14.9 Hz, 2H), 2.02 – 1.69 (m, 2H), 1.70 – 1.54 (m, 1H), 1.50 – 1.24 (m, 1H), 1.19 (s, 1H), 0.78 (t, *J* = 7.2 Hz, 6H).

Benzyl ((S)-3-methyl-1-oxo-1-(((S)-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3yl)propan-2-yl)amino)-3-(thiophen-2-yl)propan-2-yl)amino)butan-2yl)carbamate (MPI15). MPI15 was prepared as a white solid following a similar procedure to MPI11 (yield 60%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.30 (s, 1H), 8.24 (s, 1H), 7.38 – 7.22 (m, 4H), 7.04 (d, J = 5.0 Hz, 1H), 6.91 (d, J = 8.4 Hz, 1H), 6.87 – 6.65 (m, 2H), 5.97 (s, 1H), 5.33 (d, J = 8.2 Hz, 1H), 5.03 (s, 2H), 4.80 (d, J = 7.7Hz, 1H), 4.20 (s, 1H), 3.97 (t, J = 7.2 Hz, 1H), 3.49 – 3.03 (m, 4H), 2.24 (s, 2H), 2.14 – 1.93 (m, 2H), 1.76 (d, J = 33.7 Hz, 3H), 0.83 (dd, J = 25.9, 6.7 Hz, 6H).

Methyl (S)-2-((2S,3R)-2-(((benzyloxy)carbonyl)amino)-3-(tertbutoxy)butanamido)-4,4-dimethylpentanoate (MPI16b). MPI16b was prepared with Z-Thr(tBu)-OH and methyl (S)-2-amino-4,4-dimethylpentanoate hydrochloride as a white solid following a similar procedure to **MPI11b** (yield 83%).¹H NMR (400 MHz, Chloroform-*d*) δ 7.5 (d, J = 8.1 Hz, 1H), 7.3 (d, J = 4.2 Hz, 5H), 5.9 (d, J = 4.3Hz, 1H), 5.1 – 4.9 (m, 2H), 4.5 (td, J = 9.0, 3.2 Hz, 1H), 4.1 (d, J = 5.0 Hz, 2H), 3.6 (s, 3H), 1.7 (dd, J = 14.5, 3.2 Hz, 1H), 1.4 (dd, J = 14.5, 9.4 Hz, 1H), 1.1 (d, J = 74.2 Hz, 12H), 0.9 (s, 9H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.2, 169.2, 156.1, 136.3, 128.5, 128.1, 127.9, 75.6, 66.9, 66.8, 58.4, 52.2, 50.3, 45.7, 30.7, 29.7, 28.2, 16.2. (S)-2-((2S,3R)-2-(((benzyloxy)carbonyl)amino)-3-(tert-butoxy)butanamido)-4,4-dimethylpentanoic acid (MPI16c). MPI16c was prepared as a white solid following a similar procedure to MPI11c (yield 82%).

Methyl (5S,8S,11S)-5-((R)-1-(tert-butoxy)ethyl)-8-(cyclopropylmethyl)-3,6,9trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10triazadodecan-12-oate (MPI16d). MPI16d was prepared as a white solid following a similar procedure to MPI11d (yield 66%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.9 (d, J = 7.5 Hz, 1H), 7.5 (d, J = 8.4 Hz, 1H), 7.2 (q, J = 8.0, 6.1 Hz, 5H), 7.2 (s, 1H), 6.0 (d, J = 5.5 Hz, 1H), 5.0 (q, J = 12.3 Hz, 2H), 4.5 (td, J = 8.4, 4.1 Hz, 1H), 4.4 (ddd, J =11.9, 7.4, 3.5 Hz, 1H), 4.1 (p, J = 4.2 Hz, 2H), 3.6 (s, 3H), 3.2 (dd, J = 20.8, 9.1 Hz, 2H), 2.3 (qd, J = 9.9, 3.9 Hz, 1H), 2.2 – 2.1 (m, 2H), 1.8 – 1.6 (m, 3H), 1.4 (dd, J =14.5, 8.6 Hz, 1H), 1.3 – 0.9 (m, 12H), 0.9 (s, 9H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 179.6, 172.5, 172.0, 168.9, 155.9, 136.0, 128.3, 128.0, 127.8, 75.3, 66.7, 66.7, 58.6, 52.0, 50.9, 50.7, 45.8, 40.3, 38.0, 32.8, 30.2, 29.6, 28.0, 27.6, 16.8.

Benzyl ((2S,3R)-3-(tert-butoxy)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-4,4-dimethyl-1-oxopentan-2-yl)amino)-1-oxobutan-2-yl)carbamate (MPI16e). MPI16e was prepared as a white solid following a similar procedure to MPI11e (yield 66%). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.1 (d, *J* = 8.7 Hz, 1H), 7.5 (d, *J* = 8.0 Hz, 1H), 7.3 – 7.1 (m, 6H), 6.8 (s, 1H), 5.1 – 4.9 (m, 2H), 4.4 (q, *J* = 6.9 Hz, 2H), 4.1 (dd, *J* = 6.3, 3.8 Hz, 1H), 4.0 – 3.9 (m, 1H), 3.9 (q, *J* = 10.1, 9.2 Hz, 1H), 3.6 – 3.4 (m, 2H), 3.1 (dtd, *J* = 17.2, 9.3, 8.7, 5.0 Hz, 2H), 2.4 – 2.1 (m, 3H), 1.8 – 1.6 (m, 2H), 1.3 (dd, *J* = 14.3, 6.9 Hz, 2H), 1.2 (s, 9H), 0.9 (d, *J* = 6.3 Hz, 3H), 0.8 (s, 9H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 181.7, 172.7, 169.0, 156.3, 136.3, 128.4, 128.0, 127.9, 75.6, 67.2, 66.7, 65.2, 58.6, 53.4, 51.2, 49.1, 45.4, 40.5, 38.5, 33.9, 30.2, 29.5, 28.1, 16.5.

Benzyl ((2S,3R)-3-(tert-butoxy)-1-(((S)-4,4-dimethyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)pentan-2-yl)amino)-1-oxobutan-2-yl)carbamate (MPI16). MPI16 was prepared as a white solid following a similar procedure to MPI11 (yield 40%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.4 (s, 1H), 8.1 (d, *J* = 6.6 Hz, 1H), 7.5 (d, *J* = 8.3 Hz, 1H), 7.3 – 7.2 (m, 5H), 6.8 (s, 1H), 5.9 (d, *J* = 5.1 Hz, 1H), 5.0 (q, *J* = 12.2 Hz, 2H), 4.5 (td, *J* = 8.5, 4.1 Hz, 1H), 4.3 (dq, *J* = 10.8, 5.4, 4.8 Hz, 1H), 4.1 (d, *J* = 5.8 Hz, 2H), 3.3 – 3.1 (m, 2H), 2.3 (p, *J* = 8.3 Hz, 1H), 2.3 – 2.2 (m, 1H), 1.9 (td, *J* = 12.4, 10.2, 5.7 Hz, 1H), 1.8 (ddt, *J* = 11.8, 8.3, 3.9 Hz, 2H), 1.7 (tt, *J* = 12.3, 6.2 Hz, 1H), 1.4 (dd, *J* = 14.4, 8.6 Hz, 1H), 1.1 (d, *J* = 81.8 Hz, 12H), 0.9 (d, *J* = 7.3 Hz, 9H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 199.6, 180.0, 173.3, 169.3, 156.2, 136.2, 128.6, 128.2, 128.1, 75.6, 67.0, 66.8, 58.9, 57.3, 51.2, 46.1, 40.5, 37.8, 30.5, 29.9, 29.8, 29.7, 28.2, 17.0.

Methyl (S)-2-((2S,3R)-2-(((benzyloxy)carbonyl)amino)-3-(tertbutoxy)butanamido)-3-cyclopropylpropanoate (MPI17b). MPI17b was prepared with Z-Thr(tBu)-OH and methyl (S)-2-amino-3-cyclopropylpropanoate hydrochloride as a white solid following a similar procedure to **MPI11b** (yield 79%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.7 (d, J = 7.5 Hz, 1H), 7.3 (d, J = 4.3 Hz, 5H), 5.9 (d, J = 5.3Hz, 1H), 5.1 – 5.0 (m, 2H), 4.6 (q, J = 6.3 Hz, 1H), 4.2 – 4.1 (m, 2H), 3.7 (s, 3H), 1.6 (ddt, J = 27.6, 13.9, 7.1 Hz, 2H), 1.2 (s, 8H), 1.0 (d, J = 6.4 Hz, 4H), 0.6 (p, J = 6.3 Hz, 1H), 0.4 (d, J = 8.1 Hz, 2H), -0.0 (q, J = 8.7, 7.3 Hz, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 172.2, 169.0, 156.0, 136.2, 128.4, 128.0, 127.9, 75.4, 66.8, 66.7, 58.5, 52.7, 52.0, 36.9, 28.1, 16.5, 6.9, 4.2, 4.0. (S)-2-((2S,3R)-2-(((benzyloxy)carbonyl)amino)-3-(tert-butoxy)butanamido)-3cyclopropylpropanoic acid (MPI17c). MPI17c was prepared as a white solid following a similar procedure to MPI11c (yield 90%).

Methyl (5S,8S,11S)-5-((R)-1-(tert-butoxy)ethyl)-8-(cyclopropylmethyl)-3,6,9trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10triazadodecan-12-oate (MPI17d). MPI17d was prepared as a white solid following a similar procedure to MPI11d (yield 60%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.8 (d, *J* = 7.4 Hz, 1H), 7.6 (d, *J* = 7.9 Hz, 1H), 7.2 (d, *J* = 12.4 Hz, 5H), 7.0 (s, 1H), 5.9 (d, *J* = 6.0 Hz, 1H), 5.0 (t, *J* = 10.8 Hz, 2H), 4.5 (q, *J* = 7.1 Hz, 1H), 4.4 (ddd, *J* = 11.4, 7.7, 3.8 Hz, 1H), 4.1 – 4.0 (m, 2H), 3.6 (s, 3H), 3.2 (dt, *J* = 16.5, 9.1 Hz, 2H), 2.3 (ddq, *J* = 28.6, 13.9, 7.6 Hz, 2H), 2.1 (ddd, *J* = 15.6, 11.7, 4.2 Hz, 1H), 1.8 – 1.6 (m, 2H), 1.6 (q, *J* = 7.7 Hz, 2H), 1.2 (s, 8H), 1.0 (d, *J* = 6.2 Hz, 4H), 0.7 (td, *J* = 8.0, 4.2 Hz, 1H), 0.4 (dd, *J* = 8.1, 5.0 Hz, 2H), 0.0 (d, *J* = 4.9 Hz, 2H). ¹³C NMR (100 MHz, Chloroform*d*) δ 179.8, 172.1, 171.7, 169.2, 156.1, 136.2, 128.5, 128.1, 128.0, 75.2, 66.8, 58.9, 53.6, 53.5, 52.2, 50.9, 40.4, 38.2, 37.6, 33.0, 28.2, 27.9, 17.3, 7.1, 4.4, 4.2.

Benzyl ((2S,3R)-3-(tert-butoxy)-1-(((S)-3-cyclopropyl-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-1-oxopropan-2-yl)amino)-1-oxobutan-2-yl)carbamate (MPI17e). MPI17e was prepared as a white solid following a similar procedure to MPI11e (yield 85%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.6 (t, *J* = 7.0 Hz, 2H), 7.3 (d, *J* = 4.3 Hz, 5H), 7.0 (s, 1H), 6.1 (d, *J* = 6.3 Hz, 1H), 5.1 – 4.9 (m, 2H), 4.4 (t, *J* = 7.2 Hz, 1H), 4.2 (d, *J* = 6.6 Hz, 1H), 4.2 – 4.0 (m, 2H), 4.0 (ddt, *J* = 13.4, 9.3, 4.5 Hz, 1H), 3.6 – 3.4 (m, 2H), 3.2 (s, 2H), 2.4 – 2.2 (m, 2H), 2.0 (d, *J* = 12.4 Hz, 1H), 1.8 – 1.4 (m, 4H), 1.1 (d, *J* = 66.6 Hz, 12H), 0.6 (t, *J* = 7.0 Hz, 1H), 0.4 (q, *J* = 8.3, 7.4 Hz, 2H), 0.0 (s, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 181.0, 171.8, 169.4, 156.2, 136.1, 128.4, 128.1, 128.0, 75.1, 67.8, 66.8, 65.1, 59.1, 54.0, 49.7, 40.5, 38.2, 37.3, 32.4, 28.1, 28.0, 17.6, 7.2, 4.3, 4.3.

Benzyl ((2S,3R)-3-(tert-butoxy)-1-(((S)-3-cyclopropyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)propan-2-yl)amino)-1-oxobutan-2-yl)carbamate (MPI17). MPI17 was prepared as a white solid following a similar procedure to MPI11 (yield 54%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.4 (s, 1H), 8.1 (d, J = 6.3 Hz, 1H), 7.5 (t, J = 8.6 Hz, 1H), 7.3 (s, 5H), 6.4 – 6.1 (m, 1H), 5.9 (d, J = 5.4 Hz, 1H), 5.0 (q, J = 12.3 Hz, 2H), 4.4 (dt, J = 34.9, 6.6 Hz, 1H), 4.3 (p, J = 5.6, 4.6 Hz, 1H), 4.1 (t, J = 5.9 Hz, 2H), 3.3 (d, J = 8.6 Hz, 1H), 3.2 – 3.2 (m, 1H), 2.3 (d, J = 14.0 Hz, 1H), 2.2 (s, 2H), 2.0 – 1.9 (m, 1H), 1.9 (t, J = 6.8 Hz, 1H), 1.7 (dt, J = 15.4, 8.5 Hz, 1H), 1.6 (dd, J = 12.9, 6.3 Hz, 1H), 1.1 (d, J = 72.7 Hz, 12H), 0.7 (d, J = 9.1 Hz, 1H), 0.4 (q, J = 8.5 Hz, 2H), 0.0 (d, J = 5.1 Hz, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 199.7, 180.1, 172.4, 169.6, 156.3, 136.2, 128.7, 128.4, 128.2, 75.5, 67.1, 66.9, 59.2, 57.7, 55.0, 40.6, 38.0, 37.5, 29.9, 28.6, 28.3, 17.7, 7.4, 4.7, 4.5.

(S)-methyl 2-((S)-2-(((benzyloxy)carbonyl)amino)-3,3-dimethylbutanamido)-4,4-dimethylpentanoate (MPI18b). MPI18b was prepared with Cbz-L-tert-leucine and methyl (S)-2-amino-4,4-dimethylpentanoate hydrochloride as a white solid following a similar procedure to MPI11b (yield 81%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.40 – 7.19 (m, 5H), 6.51 – 6.17 (m, 1H), 5.64 – 5.46 (m, 1H), 5.09 (s, 2H), 4.74 – 4.54 (m, 1H), 4.06 – 3.96 (m, 1H), 3.70 (d, *J* = 2.5 Hz, 3H), 1.83 – 1.74 (m, 1H), 1.52 – 1.38 (m, 1H), 1.00 (d, *J* = 2.4 Hz, 9H), 0.91 (d, *J* = 2.6 Hz, 9H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 173.5, 170.2, 156.4, 136.3, 128.5, 128.1, 127.92, 127.90, 66.9, 60.4, 52.3, 49.7, 46.1, 34.8, 30.7, 29.4, 26.4, 21.1. (S)-2-(((benzyloxy)carbonyl)amino)-3,3-dimethylbutanamido)-4,4dimethylpentanoic acid (MPI18c). MPI18c was prepared as a white solid following a similar procedure to MPI11c (yield 79%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.40 – 7.27 (m, 5H), 7.03 – 6.56 (m, 1H), 5.85 (s, 1H), 5.08 (q, *J* = 12.5 Hz, 2H), 4.64 – 4.42 (m, 1H), 4.14 – 3.94 (m, 1H), 1.85 – 1.72 (m, 1H), 1.59 – 1.43 (m, 1H), 0.94 (s, 9H), 0.89 (s, 9H).

(5S,8S,11S)-methyl 5-(tert-butyl)-8-neopentyl-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate (MPI18d). MPI18d was prepared as a white solid following a similar procedure to MPI11d (yield 60%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.73 (d, *J* = 8.2 Hz, 1H), 7.60 (d, *J* = 9.0 Hz, 1H), 7.38 – 7.27 (m, 5H), 7.18 – 7.08 (m, 1H), 5.62 (d, *J* = 9.8 Hz, 1H), 5.06 (d, *J* = 2.4 Hz, 2H), 4.75 – 4.61 (m, 1H), 4.61 – 4.50 (m, 1H), 4.00 (dd, *J* = 9.6, 5.3 Hz, 1H), 3.67 (s, 3H), 3.41 – 3.21 (m, 2H), 2.51 – 2.32 (m, 2H), 2.29 – 2.16 (m, 1H), 1.94 – 1.74 (m, 3H), 1.49 – 1.44 (m, 1H), 0.96 (s, 9H), 0.89 (s, 9H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 179.9, 173.0, 171.9, 170.2, 156.6, 136.3, 128.5, 128.2, 128.1, 127.9, 67.0, 62.7, 55.5, 52.3, 50.8, 50.5, 46.8, 40.6, 38.2, 34.6, 30.6, 29.5, 27.8, 26.5, 26.4.

Benzyl ((S)-1-(((S)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-4,4-dimethyl-1-oxopentan-2-yl)amino)-3,3-dimethyl-1-oxobutan-2-yl)carbamate (MPI18e). MPI18e was prepared as a white solid following a similar procedure to MPI11e (yield 67%). ¹H NMR (400 MHz, Chloroform-*d*): δ 8.73 – 8.45 (m, 1H), 8.15 – 7.94 (m, 1H), 7.62 – 7.43 (m, 1H), 7.42 – 7.25 (H), 5.83 – 5.63 (m, 1H), 5.07 (d, J = 4.2 Hz, 2H), 4.93 – 4.75 (m, 1H), 4.49 – 4.33 (m, 1H), 4.01 – 3.85 (m, 1H), 3.85 – 3.67 (m, 1H), 3.51 – 3.22 (m, 3H), 3.22 – 3.03 (m, 1H), 2.35 – 2.23 (m, 1H), 22.20 – 2.10 (m, 1H), 1.90 – 1.55 (m, 4H), 1.55 – 1.37 (m, 1H), 0.93 (s, 9H), 0.90 (s, 9H).¹³C NMR (100 MHz, Chloroform-*d*): δ 180.3, 173.5, 169.8, 156.5, 136.2, 128.6, 128.5, 128.2, 128.1, 128.0, 67.2, 65.0, 61.7, 53.4, 50.5, 47.2, 40.5, 38.2, 35.3, 34.8, 30.6, 29.6, 26.5.

Benzyl ((S)-1-(((S)-4,4-dimethyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)pentan-2-yl)amino)-3,3-dimethyl-1-oxobutan-2-yl)carbamate (MPI18). MPI18 was prepared as a white solid following a similar procedure to MPI11 (yield 67%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.50 (s, 1H), 8.16 – 7.99 (m, 1H), 7.42 – 7.28 (m, 5H), 6.90 – 6.74(m, 1H), 5.55 (dd, *J* = 22.5, 9.6 Hz, 1H), 5.08 (d, *J* = 2.7 Hz, 2H), 4.72 – 4.58 (m, 1H), 4.38 (dd, *J* = 10.6, 5.9 Hz, 1H), 3.96 (dd, *J* = 13.1, 9.4 Hz, 1H), 3.47 – 3.25 (m, 2H), 2.51 – 2.29 (m, 2H), 2.09 – 1.77 (m, 4H), 1.53 – 1.43 (m, 1H), 1.13 – 0.71 (m, 18H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 199.3, 18.0, 173.7, 170.3, 156.5, 136.3, 128.5, 128.2, 127.9, 67.1, 62.6, 57.0, 50.8, 46.6, 40.5, 37.8, 36.5, 34.7, 30.6, 29.9, 29.5, 28.0, 26.5.

Methyl (S)-2-((S)-2-(((benzyloxy)carbonyl)amino)-3,3-dimethylbutanamido)-3cyclopropylpropanoate (MPI19b). MPI19b was prepared with Cbz-L-tert-leucine and methyl (S)-2-amino-3-cyclopropylpropanoate hydrochloride as a colorless oil following a similar procedure to MPI11b (yield 73%).

(S)-2-((S)-2-(((benzyloxy)carbonyl)amino)-3,3-dimethylbutanamido)-3cyclopropylpropanoic acid (MPI19c). MPI19c was prepared as a white solid following a similar procedure to MPI11c (yield 82%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.41 – 7.28 (m, 5H), 6.89 (d, *J* = 7.6 Hz, 1H), 5.85 (d, *J* = 9.7 Hz, 1H), 5.10 (s, 2H), 4.67 (q, *J* = 7.7, 6.3 Hz, 1H), 4.16 (d, *J* = 9.7 Hz, 1H), 1.72 (dtd, *J* = 27.3, 13.9, 6.4 Hz, 2H), 0.99 (s, 9H), 0.67 (pd, *J* = 7.5, 3.7 Hz, 1H), 0.42 (td, *J* = 8.1, 5.0 Hz, 2H), 0.07 (td, *J* = 4.7, 2.3 Hz, 2H).

Methyl (5S,8S,11S)-5-(tert-butyl)-8-(cyclopropylmethyl)-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate

(MPI19d). MPI19d was prepared as a white solid following a similar procedure to MPI11d (yield 64%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.80 (d, *J* = 7.4 Hz, 1H), 7.41 – 7.28 (m, 5H), 6.96 (d, *J* = 8.3 Hz, 1H), 6.33 (d, *J* = 9.9 Hz, 1H), 5.55 (t, *J* = 10.4 Hz, 1H), 5.09 (s, 2H), 4.67 – 4.48 (m, 2H), 3.96 (t, *J* = 9.2 Hz, 1H), 3.71 (s, 3H), 3.43 – 3.27 (m, 2H), 2.53 – 2.32 (m, 2H), 2.17 (ddd, J = 16.5, 12.2, 4.8 Hz, 1H), 1.87 (dddd, J = 17.6, 11.4, 8.6, 2.9 Hz, 1H), 1.80 – 1.68 (m, 2H), 1.60 (dt, *J* = 13.9, 7.0 Hz, 1H), 0.99 (s, 9H), 0.73 (td, *J* = 7.5, 3.9 Hz, 1H), 0.42 (ddq, *J* = 11.4, 7.7, 3.8 Hz, 2H), 0.16 – 0.01 (m, 2H).

Benzyl ((S)-1-(((S)-3-cyclopropyl-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-1-oxopropan-2-yl)amino)-3,3-dimethyl-1-oxobutan-2-yl)carbamate (MPI19e). MPI19e was prepared as a white solid following a similar procedure to MPI11e. It was used in the next step without further purification.

Benzyl ((S)-1-(((S)-3-cyclopropyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-3,3-dimethyl-1-oxobutan-2-

yl)carbamate (MPI19). MPI19 was prepared as a white solid following a similar procedure to **MPI11** (yield 48%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.51 (s, 1H), 8.20 (d, *J* = 6.7 Hz, 1H), 7.32 (d, *J* = 4.2 Hz, 5H), 7.17 (d, *J* = 8.3 Hz, 1H), 6.67 (s, 1H), 5.68 (d, J = 9.4 Hz, 1H), 5.08 (s, 2H), 4.67 (q, J = 7.3 Hz, 1H), 4.42 (p, *J* = 4.9 Hz, 1H), 4.02 (d, *J* = 9.4 Hz, 1H), 3.31 (dq, *J* = 17.4, 9.5 Hz, 2H), 2.44 (p, *J* = 8.1 Hz, 1H), 2.34 (dt, *J* = 14.9, 8.5 Hz, 1H), 2.07 – 1.86 (m, 2H), 1.86 – 1.74 (m, 1H), 1.65 (ddt, J = 20.5, 13.6, 6.9 Hz, 2H), 0.98 (s, 9H), 0.70 (h, *J* = 7.1, 6.6 Hz, 1H), 0.43 (d, *J* = 8.0 Hz, 2H), 0.08 (d, *J* = 5.1 Hz, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 199.3, 180.0, 172.6, 170.4, 156.6, 136.3, 128.5, 128.2, 128.0, 77.3, 67.1, 62.9, 57.4, 53.7, 40.6, 37.9, 37.7, 34.7, 29.9, 28.4, 26.5, 7.3, 4.5, 4.3.

(S)-methyl 2-((S)-2-(((benzyloxy)carbonyl)amino)-2-cyclopropylacetamido)-4,4dimethylpentanoate (MPI20b). MPI20b was obtained from methyl (S)-2-amino-4,4dimethylpentanoate hydrochloride and (S)-2-(((benzyloxy)carbonyl)amino)-2cyclopropylacetic acid following a similar procedure to MPI11b. The crude product was purified by silica gel column chromatography (15-50% EtOAc in n-hexane) to afford the MPI20b as a white solid (yield 80%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.42 – 7.28 (m, 5H), 6.53 – 6.37 (m, 1H), 5.63 – 5.44 (m, 1H), 5.09 (s, 2H), 4.64 (td, J = 8.7, 3.5 Hz, 1H), 3.71 (s, 3H), 3.68 – 3.60 (m, 1H), 1.77 (dd, J = 14.4, 3.6 Hz, 1H), 1.48 (dd, J = 14.4, 8.8 Hz, 1H), 1.22 – 1.07 (m, 1H), 0.93 (s, 9H), 0.72 – 0.41 (m, 4H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 173.59, 170.61, 156.26, 136.20, 128.54, 128.17, 128.03, 67.06, 60.43, 52.39, 49.90, 46.06, 30.72, 29.50, 14.21, 3.60, 2.99.

(S)-2-(((benzyloxy)carbonyl)amino)-2-cyclopropylacetamido)-4,4dimethylpentanoic acid (MPI20c). MPI20c was prepared as a white solid following a similar procedure to MPI11c (yield 83%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.41 – 7.29 (m, 5H), 6.61 (d, *J* = 8.2 Hz, 1H), 5.72 (d, *J* = 7.6 Hz, 1H), 5.09 (s, 2H), 4.62 (td, *J* = 8.7, 3.0 Hz, 1H), 3.72 – 3.55 (m, 1H), 1.91 – 1.81 (m, 1H), 1.51 (dd, *J* = 14.5, 9.1 Hz, 1H), 1.20 – 1.06 (m, 1H), 0.94 (s, 9H), 0.69 – 0.36 (m, 4H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 176.37, 171.31, 156.49, 136.06, 128.56, 128.24, 128.06, 67.97, 67.24, 50.01, 45.68, 30.77, 29.49, 25.61, 13.90, 3.66, 3.15. (58,88,118)-methyl 5-cyclopropyl-8-neopentyl-3,6,9-trioxo-11-(((8)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate

(MPI20d). MPI20d was prepared as a white solid following a similar procedure to MPI11d (yield 49%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.89 – 7.74 (m, 1H), 7.30 – 7.19 (m, 5H), 6.75 (s, 1H), 5.85 – 5.67 (m, 1H), 5.00 (d, *J* = 5.2 Hz, 2H), 4.55 (td, *J* = 8.7, 3.5 Hz, 1H), 4.49 – 4.34 (m, 1H), 3.61 (s, 3H), 3.60 – 3.52 (m, 1H), 3.28 – 3.14 (m, 2H), 2.39 – 2.19 (m, 2H), 2.11 (td, *J* = 12.7, 11.7, 4.1 Hz, 1H), 1.87 – 1.61 (m, 3H), 1.12 – 0.98 (m, 1H), 0.85 (s, 9H), 0.57 – 0.23 (m, 4H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 179.90, 173.11, 172.07, 170.88, 156.30, 136.24, 128.52, 128.15, 128.00, 67.02, 58.86, 55.27, 52.36, 50.89, 45.93, 40.57, 38.33, 30.55, 29.60, 28.01, 12.47, 3.30, 3.18.

Benzyl ((S)-1-cyclopropyl-2-(((S)-1-((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-4,4-dimethyl-1-oxopentan-2-yl)amino)-2-

oxoethyl)carbamate (MPI20e). MPI20e was prepared as a white solid following a similar procedure to **MPI11e** (yield 79%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.70 – 7.42 (m, 2H), 7.38 – 7.21 (m, 5H), 6.79 (d, *J* = 5.1 Hz, 1H), 6.16 – 5.91 (m, 1H), 5.05 (q, *J* = 12.4 Hz, 2H), 4.62 – 4.43 (m, 1H), 4.06 – 3.87 (m, 1H), 3.63 – 3.54 (m, 2H), 3.24 (t, *J* = 9.2 Hz, 2H), 2.44 – 2.22 (m, 2H), 1.91 – 1.68 (m, 2H), 1.62 – 1.44 (m, 2H), 1.17 – 1.02 (m, 1H), 0.90 (s, 9H), 0.57 – 0.42 (m, 3H), 0.42 – 0.30 (m, 1H).

Benzyl ((S)-1-cyclopropyl-2-(((S)-4,4-dimethyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)pentan-2-yl)amino)-2-oxoethyl)carbamate (MPI20). MPI20 was prepared as a white solid following a similar procedure to MPI11 (yield 62%). ¹H NMR (400 MHz, Chloroform-*d*): δ 9.48 (s, 1H), 8.22 (s, 1H), 7.40 – 7.27 (m, 5H), 7.06 – 6.87 (m, 1H), 6.51 – 6.29 (m, 1H), 5.78 (s, 1H), 5.07 (s, 2H), 4.68 – 4.52 (m, 1H), 4.36 – 4.24 (m, 1H), 3.56 (ddd, J = 9.2, 7.0, 2.4 Hz, 1H), 3.36 – 3.19 (m, 2H), 2.50 – 2.28 (m, 2H), 2.07 – 1.84 (m, 4H), 1.84 – 1.72 (m, 1H), 1.21 – 1.07 (m, 1H), 0.95 (s, 9H), 0.66 – 0.35 (m, 4H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 199.77, 180.13, 173.80, 171.01, 156.44, 136.16, 128.55, 128.21, 128.04, 67.10, 59.12, 57.47, 51.03, 45.90, 40.59, 38.03, 30.62, 30.01, 29.64, 28.29, 13.85, 3.29, 3.22.

$$H_2N$$
 H_2N H_2N

(S)-2-(((benzyloxy)carbonyl)amino)-2,3-dimethylbutanoic acid (MPI21a). H-L-(α Me)Val-OH (1.0 g, 7.63 mmol, 1.0 equiv) was dissolved in a 1:1 mixture of 1N NaOH (20 mL) : acetone (20 mL) and kept at 0 °C . Z-Cl (1.25 mL, 0.877 mmol, 0.1 equiv), diluted in acetone (5 mL), was added drop wise in 40 min and the pH was adjusted to 10.8-11.0 with 2N NaOH. After stirring the reaction mixture at room temperature for 5 h, Acetone was evaporated under reduce pressure and NaOH 2 N was added (20 mL). The unreacted Z-Cl was extracted with Et₂O. The aqueous solution was acidified with KHSO₄, the product was extracted with EtOAc (4x30 mL). The organic phase was washed with water (3x30 mL), and the solvent was evaporated at reduced pressure. The solid colorless product was obtained in 76% yield. ¹H NMR (400 MHz, Chloroform-*d*): δ 10.38, 7.38, 7.37, 7.36, 7.36, 7.35, 7.34, 7.33, 5.46, 5.12, 1.61, 1.03, 1.01, 0.99, 0.98.

(S)-methyl 2-((S)-2-(((benzyloxy)carbonyl)amino)-2,3-dimethylbutanamido)-4,4-dimethylpentanoate (MPI21b). MPI21b was obtained from methyl (S)-2-amino-4,4-dimethylpentanoate hydrochloride and MPI21a following a similar procedure to MPI11b. The crude product was purified by silica gel column chromatography (1550% EtOAc in n-hexane) to afford the **MPI21b** as a white solid (yield 78%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.53 – 7.30 (m, 5H), 6.93 (s, 1H), 5.24 (s, 1H), 5.13 – 5.01 (m, 2H), 4.68 – 4.51 (m, 1H), 3.68 (s, 3H), 1.77 (dd, *J* = 14.4, 3.6 Hz, 1H), 1.50 (dd, *J* = 14.5, 8.9 Hz, 1H), 1.45 (s, 3H), 1.08 – 0.67 (m, 15H).

(S)-2-((S)-2-(((benzyloxy)carbonyl)amino)-2,3-dimethylbutanamido)-4,4dimethylpentanoic acid (MPI21c). MPI21c was prepared as a white solid following a similar procedure to MPI11c. The crude MPI21c was used in the next step without further purification.

(5S,8S,11S)-methyl 5-isopropyl-5-methyl-8-neopentyl-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate (MPI21d). MPI21d was prepared as a white solid following a similar procedure to MPI11d (yield 63%). ¹H NMR (400 MHz, Methanol- d_4) δ 8.35 (d, J = 8.1 Hz, 1H), 7.67 (d, J = 8.0 Hz, 1H), 7.37 – 7.13 (m, 5H), 6.93 (s, 1H), 5.14 – 4.93 (m, 2H), 4.43 – 4.19 (m, 2H), 3.60 (s, 3H), 3.20 – 3.10 (m, 2H), 2.45 – 2.33 (m, 1H), 2.28 – 2.18 (m, 1H), 1.97 – 1.87 (m, 1H), 1.77 – 1.61 (m, 3H), 1.50 (dd, J = 14.4, 8.3 Hz, 1H), 1.32 (s, 3H), 0.93 – 0.62 (m, 15H). ¹³C NMR (100 MHz, Methanol- d_4): δ 180.36, 174.44, 174.38, 172.08, 156.83, 136.80, 128.18, 128.09, 127.65, 127.62, 66.51, 62.93, 54.47, 51.65, 51.37, 50.73, 44.60, 42.43, 40.04, 38.06, 30.08, 28.71, 17.34, 17.12, 16.55, 16.16, 15.90.

Benzyl ((S)-1-(((S)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-4,4-dimethyl-1-oxopentan-2-yl)amino)-2,3-dimethyl-1-oxobutan-2-yl)carbamate (MPI21e). MPI21e was prepared as a white solid following a similar procedure to MPI11e (yield 70%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.41 – 7.27 (m, 5H), 6.56 (d, *J* = 6.3 Hz, 1H), 5.99 (s, 1H), 5.47 (s, 1H), 5.22 – 5.07 (m, 2H), 4.23 (ddd, *J* = 9.1, 6.2, 2.4 Hz, 1H), 4.03 (s, 1H), 3.74 – 3.64 (m, 1H), 3.63 – 3.54 (m, 1H), 3.36 – 3.19 (m, 2H), 2.46 – 2.29 (m, 2H), 2.22 – 1.96 (m, 2H), 2.01 – 1.88 (m, 2H), 1.76 (dd, *J* = 12.2, 9.0 Hz, 1H), 1.41 (s, 3H), 1.29 (dd, *J* = 14.7, 9.6 Hz, 1H), 1.02 – 0.77 (m, 15H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 180.44, 173.62, 173.41, 156.25, 135.91, 128.67, 128.46, 128.37, 67.62, 65.70, 63.37, 53.46, 53.04, 50.63, 45.80, 40.33, 35.36, 32.36, 30.69, 29.69, 28.38, 17.45, 17.33, 17.11, 17.00.

Benzyl ((S)-1-(((S)-4,4-dimethyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)pentan-2-yl)amino)-2,3-dimethyl-1-oxobutan-2yl)carbamate (MPI21). MPI21 was prepared as a white solid following a similar procedure to MPI11 (yield 78%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.52 (s, 1H), 8.07 (d, J = 7.7 Hz, 1H), 7.50 – 7.26 (m, 5H), 6.77 (d, J = 7.4 Hz, 1H), 6.39 – 6.20 (m, 1H), 5.47 (s, 1H), 5.07 (d, J = 3.2 Hz, 2H), 4.57 – 4.37 (m, 1H), 4.33 – 4.16 (m, 1H), 3.35 – 3.18 (m, 2H), 2.44 – 2.29 (m, 2H), 2.14 – 2.05 (m, 2H), 1.98 – 1.92 (m, 1H), 1.90 – 1.81 (m, 1H), 1.74 (t, J = 9.8 Hz, 1H), 1.40 (s, 3H), 0.97 – 0.85 (m, 15H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 200.59, 173.83, 173.43, 173.17, 156.23, 136.06, 128.70, 128.63, 128.35, 128.16, 67.48, 67.28, 63.36, 57.12, 51.83, 45.78, 40.35, 37.64, 34.88, 30.65, 30.24, 29.67, 28.12, 17.44, 17.36, 17.09, 17.04.

Methyl (S)-2-(2-(((benzyloxy)carbonyl)amino)-2-methylpropanamido)-4,4dimethylpentanoate (MPI22b). MPI22b was obtained from methyl (S)-2-amino-4,4dimethylpentanoate hydrochloride and Z-Aib-OH following a similar procedure to MPI11b. The crude product was purified by silica gel column chromatography (15-50% EtOAc in hexanes as the eluent) to afford the MPI22b as a colorless oil (yield 79%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.34 (s, 5H), 6.68 (s, 1H), 5.31 (s, 1H), 5.08 (s, 2H), 4.57 (t, *J* = 8.4 Hz, 1H), 3.69 (s, 3H), 1.91 – 1.64 (m, 2H), 1.53 (d, *J* = 6.0 Hz, 6H), 0.94 (s, 9H).

(S)-2-(2-(((benzyloxy)carbonyl)amino)-2-methylpropanamido)-4,4dimethylpentanoic acid (MPI22c). MPI22c was prepared as a white solid following a similar procedure to MPI11c (yield 85%).

Methyl(8S,11S)-5,5-dimethyl-8-neopentyl-3,6,9-trioxo-11-(((S)-2-
oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate(MPI22d). MPI22d was prepared as a white solid following a similar procedure to
MPI11d (yield 66%). ¹H NMR (400 MHz, Chloroform-d) δ 7.81 (d, J = 7.8 Hz, 1H),
7.41 – 7.29 (m, 5H), 6.67 (d, J = 8.2 Hz, 1H), 5.77 (s, 1H), 5.43 (s, 1H), 5.17 – 5.01 (m,
2H), 4.63 – 4.43 (m, 2H), 3.71 (s, 3H), 3.40 – 3.16 (m, 2H), 2.48 – 2.32 (m, 2H), 2.30
– 2.16 (m, 1H), 2.15 – 1.95 (m, 1H), 1.95 – 1.82 (m, 2H), 1.52 (d, J = 11.7 Hz, 6H),
1.49 – 1.36 (m, 1H), 0.95 (s, 9H).

Benzyl (1-(((S)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-4,4-dimethyl-1-oxopentan-2-yl)amino)-2-methyl-1-oxopropan-2-yl)carbamate (MPI22e). MPI22e was prepared as a white solid following a similar procedure to MPI11e. The crude MPI22e was used in the next step without further purification.

Benzyl (1-(((S)-4,4-dimethyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)pentan-2-yl)amino)-2-methyl-1-oxopropan-2-yl)carbamate (MPI22). MPI22 was prepared as a white solid following a similar procedure to MPI11 (yield 50%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.52 (s, 1H), 8.15 (d, *J* = 7.6 Hz, 1H), 7.33 (s, 5H), 6.83 (d, *J* = 8.0 Hz, 1H), 6.12 (s, 1H), 5.71 (s, 1H), 5.05 (q, *J* = 12.2 Hz, 2H), 4.58 – 4.42 (m, 1H), 4.23 (ddd, *J* = 11.3, 7.6, 3.9 Hz, 1H), 3.33 – 3.16 (m, 2H), 2.49 – 2.29 (m, 2H), 2.13 – 1.97 (m, 2H), 1.96 – 1.82 (m, 2H), 1.82 – 1.68 (m, 1H), 1.47 (d, *J* = 6.8 Hz, 6H), 0.94 (s, 9H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 200.5, 180.0, 173.8, 156.0, 136.0, 128.6, 128.3, 128.1, 67.2, 57.3, 57.1, 51.6, 45.5, 40.4, 37.7, 30.7, 30.2, 29.6, 28.3, 26.0.

Methyl (S)-2-(1-(((benzyloxy)carbonyl)amino)cyclopropane-1-carboxamido)-3cyclohexylpropanoate (MPI23b). MPI23b was obtained from H-Cha-OMe hydrochloride and 1-(((benzyloxy)carbonyl)amino)cyclopropane-1-carboxylic acid following a similar procedure to MPI11b. The crude product was purified by silica gel column chromatography (50-100% EtOAc in hexanes as the eluent) to afford the MPI23b. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.29 (d, *J* = 3.9 Hz, 5H), 6.72 (s, 1H), 5.05 (d, *J* = 12.0 Hz, 2H), 4.60 – 4.33 (m, 1H), 3.63 (s, 3H), 1.78 – 0.66 (m, 18H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.61, 171.53, 135.97, 128.63, 52.27, 50.58, 40.06, 34.09, 33.38, 32.57, 26.35, 26.15, 25.98.

(S)-2-(1-(((benzyloxy)carbonyl)amino)cyclopropane-1-carboxamido)-3cyclohexylpropanoic acid (MPI23c). MPI23c was prepared as a white solid following a similar procedure to MPI11c. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.26 (s, 5H), 6.84 (d, *J* = 11.3 Hz, 1H), 5.80 (s, 1H), 5.25 – 4.94 (m, 2H), 4.49 (s, 1H), 1.59 (dd, *J* = 13.3, 6.7 Hz, 6H), 1.51 – 1.24 (m, 3H), 1.20 (s, 2H), 1.06 – 0.91 (m, 4H), 0.81 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 176.01, 172.64, 128.63, 128.35, 67.96, 50.83, 34.07, 33.40, 32.43, 29.72, 26.34, 26.13, 25.95, 25.60.

Methyl (S)-2-((S)-2-(1-(((benzyloxy)carbonyl)amino)cyclopropane-1carboxamido)-3-cyclohexylpropanamido)-3-((S)-2-oxopyrrolidin-3yl)propanoate (MPI23d). MPI23d was prepared as a white solid following a similar procedure to **MPI11d** (yield 62%). ¹H NMR (400 MHz, Methanol- d_4) δ 7.32 – 7.13 (m, 5H), 5.14 – 4.94 (m, 2H), 4.47 – 4.26 (m, 2H), 3.62 (s, 3H), 3.13 (d, J = 7.5 Hz, 2H), 2.39 (d, J = 10.3 Hz, 1H), 2.22 – 2.02 (m, 2H), 1.82 – 1.43 (m, 9H), 1.43 – 1.30 (m, 2H), 1.17 (d, J = 12.4 Hz, 2H), 1.11 (q, J = 8.4, 7.6 Hz, 2H), 0.96 (d, J = 8.3 Hz, 2H), 0.89 – 0.70 (m, 2H).

Benzyl (1-(((S)-3-cyclohexyl-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-1-oxopropan-2-yl)carbamoyl)cyclopropyl)carbamate (MPI23e). MPI23e was prepared as a white solid following a similar procedure to MPI11e (yield 61%). ¹H NMR (400 MHz, Methanol- d_4) δ 7.32 – 7.09 (m, 5H), 5.09 (d, J = 12.4 Hz, 1H), 4.98 (d, J = 12.4 Hz, 1H), 4.35 (dd, J = 10.4, 4.9 Hz, 1H), 3.96 – 3.74 (m, 1H), 3.42 (dd, J = 11.5, 5.9 Hz, 2H), 2.37 – 2.14 (m, 2H), 1.87 (t, J = 12.5 Hz, 1H), 1.70 – 1.41 (m, 9H), 1.42 – 1.28 (m, 2H), 1.28 – 1.19 (m, 3H), 1.19 – 0.97 (m, 4H), 0.97 – 0.72 (m, 3H). ¹³C NMR (100 MHz, Methanol- d_4) δ 181.30, 173.80, 173.46, 136.60, 128.14, 127.59, 66.73, 64.06, 48.26, 48.05, 47.84, 47.63, 47.41, 47.20, 46.99, 40.16, 38.24, 35.21, 34.08, 33.57, 27.55, 26.19, 26.04, 25.80.

Benzyl (1-(((S)-3-cyclohexyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3yl)propan-2-yl)amino)propan-2-yl)carbamoyl)cyclopropyl)carbamate (MPI23). MPI23 was prepared as a white solid following a similar procedure to MPI11 (yield 41%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.43 (d, *J* = 24.1 Hz, 1H), 8.30 (s, 1H), 7.27 (d, *J* = 4.2 Hz, 5H), 6.86 (d, *J* = 56.9 Hz, 1H), 6.25 (s, 1H), 5.88 (d, *J* = 34.9 Hz, 1H), 5.18 – 4.90 (m, 2H), 4.56 (q, *J* = 4.1 Hz, 1H), 4.51 – 4.35 (m, 1H), 4.20 (s, 1H), 3.24 (d, *J* = 10.0 Hz, 2H), 2.45 – 2.20 (m, 2H), 2.03 – 1.66 (m, 7H), 1.52 – 1.35 (m, 3H), 1.22 (d, *J* = 9.4 Hz, 1H), 1.14 – 1.00 (m, 4H), 0.98 – 0.67 (m, 4H).

(S)-methyl 2-((S)-2-(((benzyloxy)carbonyl)amino)-2-cyclopropylacetamido)-3cyclopropylpropanoate (MPI24b). MPI24b was obtained from methyl (S)-2-amino-3-cyclopropylpropanoate hydrochloride and (S)-2-(((benzyloxy)carbonyl)amino)-2cyclopropylacetic acid following a similar procedure to MPI11b. The crude product was purified by silica gel column chromatography (15-50% EtOAc in hexanes as the eluent) to afford the MPI24b (yield 80%). ¹H NMR (400 MHz, Chloroform-*d*): δ 7.58 – 7.21 (m, 5H), 6.77 (s, 1H), 5.68 – 5.49 (m, 1H), 5.10 (s, 2H), 4.66 (q, *J* = 6.4 Hz, 1H), 3.75 (s, 3H), 3.71 – 3.59 (m, 1H), 1.70 (q, *J* = 6.4 Hz, 2H), 1.20 – 1.04 (m, 1H), 0.68 – 0.41 (m, 6H), 0.20 – -0.02 (m, 2H).¹³C NMR (100 MHz, Chloroform-*d*): δ 172.51, 171.10, 156.27, 136.21, 128.52, 128.15, 127.98, 67.05, 58.58, 52.79, 52.36, 36.91, 14.30, 6.82, 4.20, 4.10, 3.57, 3.16.

(S)-2-(((benzyloxy)carbonyl)amino)-2-cyclopropylacetamido)-3cyclopropylpropanoic acid (MPI24c). MPI24c was prepared as a white solid following a similar procedure to MPI11c (yield 85%). ¹H NMR (400 MHz, DMSO- d_6) δ 7.96 (d, J = 7.9 Hz, 1H), 7.43 (d, J = 8.2 Hz, 1H), 7.34 – 7.16 (m, 5H), 4.96 (s, 2H), 4.23 (td, J = 7.8, 5.3 Hz, 1H), 3.57 (t, J = 8.3 Hz, 1H), 1.64 – 1.35 (m, 2H), 1.07 – 0.87 (m, 1H), 0.82 – 0.64 (m, 1H), 0.50 – 0.19 (m, 6H), 0.12 – -0.09 (m, 2H). ¹³C NMR (100 MHz, DMSO- d_6): δ 173.78, 171.30, 156.15, 137.51, 128.78, 128.22, 128.13, 65.83, 58.15, 52.78, 36.62, 14.20, 8.04, 4.85, 4.53, 3.54, 3.00.

(5S,8S,11S)-methyl 5-cyclopropyl-8-(cyclopropylmethyl)-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-1-phenyl-2-oxa-4,7,10-triazadodecan-12-oate (MPI24d). MPI24d was prepared as a white solid following a similar procedure to MPI11d (yield 50%). ¹H NMR (400 MHz, Chloroform-*d*): δ 8.03 (d, *J* = 7.2 Hz, 1H), 7.37 - 7.26 (m, 5H), 7.19 (d, *J* = 8.0 Hz, 1H), 6.53 (s, 1H), 5.83 (d, *J* = 7.4 Hz, 1H),

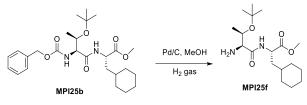
5.08 (d, J = 3.0 Hz, 2H), 4.63 (q, J = 7.1 Hz, 1H), 4.57 – 4.43 (m, 1H), 3.70 (s, 4H),

3.40 – 3.18 (m, 2H), 2.51 – 2.28 (m, 2H), 2.26 – 2.09 (m, 1H), 1.95 – 1.76 (m, 2H), 1.76 – 1.55 (m, 2H), 1.52 – 1.38 (m, 1H), 1.20 – 1.03 (m, 1H), 0.79 – 0.66 (m, 1H), 0.65 – 0.33 (m, 6H), 0.17 – -0.03 (m, 2H). ¹³C NMR (100 MHz, Chloroform-*d*): δ 179.99, 172.09, 171.94, 170.97, 156.32, 136.26, 128.53, 128.15, 128.03, 67.01, 54.81, 53.65, 52.43, 51.53, 40.60, 38.51, 37.45, 33.14, 28.31, 14.36, 7.10, 4.49, 4.22, 3.49, 3.20.

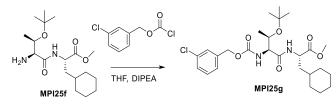
Benzyl ((S)-1-cyclopropyl-2-(((S)-3-cyclopropyl-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-1-oxopropan-2-yl)amino)-2-oxoethyl)carbamate (MPI24e). MPI24e was prepared as a white solid following a similar procedure to MPI11e (yield 79%). ¹H NMR (400 MHz, Methanol- d_4): δ 7.34 – 6.98 (m, 5H), 5.06 – 4.87 (m, 2H), 4.23 (d, J = 7.3 Hz, 1H), 3.85 (d, J = 10.5 Hz, 1H), 3.48 – 3.27 (m, 3H), 3.16 – 3.09 (m, 1H), 2.41 – 2.25 (m, 1H), 2.27 – 2.13 (m, 1H), 1.88 – 1.77 (m, 1H), 1.70 – 1.31 (m, 4H), 1.05 – 0.90 (m, 1H), 0.73 – 0.59 (m, 1H), 0.53 – 0.20 (m, 6H), 0.15 – -0.11 (m, 2H).

oxoethyl)carbamate (**MPI24**). **MPI24** was prepared as a white solid following a similar procedure to **MPI11** (yield 66%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 9.35 (s, 1H), 8.38 (d, *J* = 7.7 Hz, 1H), 7.91 (dd, *J* = 22.3, 7.8 Hz, 1H), 7.57 (s, 1H), 7.52 – 7.42 (m, 1H), 7.35 – 7.16 (m, 5H), 4.94 (s, 2H), 4.34 – 4.20 (m, 1H), 4.20 – 4.07 (m, 1H), 3.57 – 3.41 (m, 1H), 3.15 – 2.91 (m, 2H), 2.30 – 2.14 (m, 1H), 2.14 – 1.99 (m, 1H), 1.81 (q, *J* = 16.5, 14.7 Hz, 1H), 1.64 – 1.27 (m, 4H), 1.04 – 0.85 (m, 1H), 0.74 – 0.55 (m, 1H), 0.48 – 0.16 (m, 6H), 0.08 – -0.11 (m, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 201.20, 178.70, 172.56, 171.22, 156.23, 137.50, 128.79, 128.24, 128.15, 65.84, 58.23, 56.67, 53.57, 37.63, 29.81, 27.74, 14.07, 8.05, 4.88, 4.69, 3.59, 2.87.

(S)-methyl 2-((2S,3R)-2-(((benzyloxy)carbonyl)amino)-3-(tertbutoxy)butanamido)-3-cyclohexylpropanoate (MPI25b). MPI25b was obtained methvl (S)-2-amino-3-cyclohexylpropanoate hydrochloride from and N-((benzyloxy)carbonyl)-O-(tert-butyl)-L-allothreonine following a similar procedure to MPI11b. The crude product was purified by silica gel column chromatography (15-50% EtOAc in hexanes as the eluent) to afford the **MPI25b** (yield 84%).¹H NMR (400 MHz, Chloroform-d) δ 7.65 (d, J = 7.7 Hz, 1H), 7.43 – 7.29 (m, 5H), 5.94 (d, J = 5.0Hz, 1H), 5.27 – 4.90 (m, 2H), 4.52 (td, J = 8.4, 5.0 Hz, 1H), 4.27 – 4.16 (m, 2H), 3.72 (s, 3H), 1.79 – 1.61 (m, 7H), 1.60 – 1.47 (m, 1H), 1.30 (s, 9H), 1.22 – 1.14 (m, 3H), 1.11 (d, J = 6.0 Hz, 3H), 1.01 – 0.85 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 173.07, 169.43, 156.15, 136.28, 128.55, 128.15, 127.96, 75.57, 66.83, 60.42, 58.37, 52.16, 50.51, 39.76, 34.30, 33.51, 32.56, 28.20, 26.32, 26.18, 25.99, 21.07, 16.41.



(S)-methyl 2-((2S,3R)-2-amino-3-(tert-butoxy)butanamido)-3cyclohexylpropanoate (MPI25f). To a solution of MPI25b (2.73 mmol, 1.3 g) in methanol (20 mL) was added 10 % Pd/C (150 mg). The mixture was then stirred with hydrogen balloon at room temperature for 12 h. The catalyst was then filtered off and the solution was evaporated on vacuum to afford MPI25f as white solid, which was used without purification. ¹H NMR (400 MHz, Methanol- d_4) δ 4.49 (dd, J = 9.4, 5.4 Hz, 1H), 4.01 - 3.90 (m, 1H), 3.73 (s, 3H), 3.41 - 3.28 (m, 1H), 3.20 (d, J = 4.5 Hz, 1H), 1.90 - 1.59 (m, 7H), 1.51 - 1.37 (m, 1H), 1.23 (d, J = 9.9 Hz, 15H), 1.11 - 0.89 (m, 2H). ¹³C NMR (100 MHz, MeOD): δ 174.31, 173.15, 73.93, 68.77, 59.84, 51.23, 50.10, 38.97, 33.92, 33.35, 32.03, 27.59, 26.10, 25.94, 25.70, 18.68.



(S)-methyl

2-((2S,3R)-3-(tert-butoxy)-2-((((3-

chlorobenzyl)oxy)carbonyl)amino)butanamido)-3-cyclohexylpropanoate (MPI25g). To 3,5-dichlorobenzyl alcohol (0.201 g, 1.39 mmol) in THF (5 mL) were added K₂CO₃ (193 mg, 1.39 mmol) and Triphosgene (166 mg, 0.56 mmol) and the mixture was stirred at room temperature for 1 h. The mixture was then poured into water (10 mL) and extracted with ethyl acetate (2×20 mL), combine organic layers and dried over Na₂SO₄. The organic phase was evaporated to dryness and the crude material was used directly in the next step. 3,5-Dichlorobenzyl Chloroformate in THF (5 mL) was added to drop wise to a mixture of compound MPI25f (400 mg,1.39mmol) and DIPEA (0.47 ml, 2.78mmol). The reaction mixture stirred for 12 h. The mixture was then poured into water (30 mL) and extracted with ethyl acetate (4×20 mL). The organic layer was washed with aqueous hydrochloric acid 10% v/v (2×20 mL), saturated aqueous NaHCO₃ (2×20 mL), brine (2×20 mL) and dried over Na₂SO₄. The organic phase was evaporated to dryness and the crude material purified by silica gel column chromatography (15-50% EtOAc in n-hexane as the eluent) to afford MPI25g white solid (410 mg, 70%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.69 (d, J = 7.7 Hz, 1H), 7.44 – 7.22 (m, 4H), 6.00 (d, J = 4.9 Hz, 1H), 5.19 – 5.03 (m, 2H), 4.55 (td, J = 8.4, 5.1 Hz, 1H), 4.22 (q, J = 6.9, 5.5 Hz, 2H), 3.75 (s, 3H), 1.88 – 1.60 (m, 7H), 1.46 – 1.11 (m, 16H), 1.04 – 0.89 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 173.04, 171.18, 169.33, 155.86, 138.37, 134.42, 129.83, 128.24, 127.85, 125.81, 75.62, 66.84, 65.85, 60.42, 58.34, 52.17, 50.54, 39.75, 34.68, 34.31, 33.51, 32.56, 31.61, 28.19, 26.32, 26.18, 16.36.

(S)-2-((2S,3R)-3-(tert-butoxy)-2-((((3-

chlorobenzyl)oxy)carbonyl)amino)butanamido)-3-cyclohexylpropanoic acid (MPI25c). MPI25c was prepared as a white solid following a similar procedure to MPI11c (yield 82%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.57 (d, *J* = 7.3 Hz, 1H), 7.39 – 7.04 (m, 4H), 6.09 (d, *J* = 5.8 Hz, 1H), 5.16 – 4.83 (m, 2H), 4.36 (td, *J* = 8.1, 7.6, 4.7 Hz, 1H), 4.20 – 4.01 (m, 2H), 1.72 – 1.46 (m, 7H), 1.33 – 0.95 (m, 16H), 0.95 – 0.76 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 177.40, 170.06, 156.11, 138.37, 134.39, 129.82, 128.21, 127.85, 125.84, 75.56, 67.03, 65.96, 58.65, 39.41, 34.33, 33.65, 32.35, 28.20, 26.34, 26.20, 25.97, 16.82.

 $(5S,8S,11S) - methyl \\ 5-((R)-1-(tert-butoxy)ethyl)-1-(3-chlorophenyl)-8-(cyclohexylmethyl)-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-2-oxa-$

4,7,10-triazadodecan-12-oate (**MPI25d**). **MPI25d** was prepared as a white solid following a similar procedure to **MPI11d** (yield 64%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.48 (dd, *J* = 52.5, 7.6 Hz, 2H), 7.36 – 7.01 (m, 4H), 6.30 (s, 1H), 5.90 (d, *J* = 5.1 Hz, 1H), 5.01 (q, *J* = 12.6 Hz, 2H), 4.54 – 4.33 (m, 2H), 4.11 (d, *J* = 5.7 Hz, 2H), 3.65 (s, 3H), 3.39 – 3.18 (m, 2H), 2.46 – 2.25 (m, 2H), 2.20 – 2.03 (m, 1H), 1.87 – 1.40 (m, 9H), 1.11 (d, *J* = 72.3 Hz, 16H), 0.97 – 0.79 (m, 2H). ¹³C NMR (100 MHz,

CDCl₃) δ 179.92, 172.23, 172.12, 169.47, 155.95, 138.30, 134.42, 129.86, 128.28, 127.93, 125.93, 75.51, 66.73, 65.97, 58.82, 52.43, 51.49, 51.19, 40.55, 39.95, 38.64, 38.24, 34.10, 33.62, 33.13, 32.66, 28.30, 28.23, 26.39, 26.23, 26.07, 17.15.

3-chlorobenzyl ((2S,3R)-3-(tert-butoxy)-1-(((S)-3-cyclohexyl-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-1-oxopropan-2-yl)amino)-1oxobutan-2-yl)carbamate (MPI25e). MPI25e was prepared as a white solid following a similar procedure to MPI11e (yield 67%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.54 (d, *J* = 7.6 Hz, 1H), 7.34 – 7.10 (m, 4H), 6.10 (d, *J* = 5.4 Hz, 1H), 5.89 (s, 1H), 5.01 (q, *J* = 12.6 Hz, 2H), 4.32 (q, *J* = 7.7 Hz, 1H), 4.18 – 4.03 (m, 2H), 3.94 (s, 1H), 3.55 (t, *J* = 4.0 Hz, 2H), 3.36 – 3.12 (m, 2H), 2.44 – 2.23 (m, 4H), 2.05 – 1.90 (m, 1H), 1.78 – 1.41 (m, 8H), 1.11 (d, *J* = 68.1 Hz, 15H), 0.93 – 0.77 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 180.95, 172.81, 169.73, 156.11, 138.27, 134.42, 129.88, 128.31, 127.97, 125.97, 75.52, 66.73, 66.06, 65.92, 59.13, 51.89, 40.52, 39.74, 38.21, 34.25, 33.62, 32.61, 28.63, 28.24, 26.36, 26.23, 26.03, 17.44.

3-chlorobenzyl ((2S,3R)-3-(tert-butoxy)-1-(((S)-3-cyclohexyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)propan-2-yl)amino)-1oxobutan-2-yl)carbamate (MPI25). MPI25 was prepared as a white solid following a similar procedure to MPI11 (yield 72%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.44 (s, 1H), 7.99 (d, *J* = 6.4 Hz, 1H), 7.41 (d, *J* = 7.8 Hz, 1H), 7.29 (s, 1H), 7.22 (d, *J* = 4.6 Hz, 2H), 7.16 (t, *J* = 4.5 Hz, 1H), 6.11 (s, 1H), 5.89 (d, *J* = 5.1 Hz, 1H), 5.01 (q, *J* = 12.7 Hz, 2H), 4.48 – 4.37 (m, 1H), 4.35 – 4.25 (m, 1H), 4.11 (d, *J* = 5.5 Hz, 2H), 3.36 – 3.19 (m, 2H), 2.44 – 2.26 (m, 2H), 1.98 – 1.86 (m, 2H), 1.78 – 1.70 (m, 2H), 1.60 – 1.47 (m, 3H), 1.30 – 1.15 (m, 12H), 1.16 – 1.08 (m, 3H), 1.02 (d, *J* = 6.0 Hz, 3H), 0.96 – 0.82 (m, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 199.57, 179.96, 172.90, 169.62, 155.99, 138.26, 134.43, 129.88, 128.32, 127.95, 125.95, 75.51, 66.72, 66.02, 58.93, 57.58, 51.50, 40.54, 40.03, 37.92, 34.24, 33.65, 32.62, 29.81, 29.71, 28.62, 28.24, 26.35, 26.22, 26.02, 17.29.

(S)-methyl 2-((2S,3R)-2-(((benzyloxy)carbonyl)amino)-3-(tertbutoxy)butanamido)-4-methylpentanoate (MPI26b). MPI26b was obtained from methyl L-leucinate hydrochloride and N-((benzyloxy)carbonyl)-O-(tert-butyl)-Lallothreonine following a similar procedure to MPI11b. The crude product was purified by silica gel column chromatography (15-50% EtOAc in hexanes as the eluent) to afford the MPI26b (yield 78%).¹H NMR (400 MHz, Chloroform-*d*) δ 7.66 (d, *J* = 7.8 Hz, 1H), 7.51 – 7.31 (m, 5H), 5.94 (d, *J* = 4.9 Hz, 1H), 5.26 – 4.95 (m, 2H), 4.57 – 4.42 (m, 1H), 4.28 – 4.16 (m, 2H), 3.72 (s, 3H), 1.75 – 1.52 (m, 3H), 1.30 (s, 9H), 1.11 (d, *J* = 6.2 Hz, 3H), 0.94 (dd, *J* = 6.1, 3.8 Hz, 6H).¹³C NMR (100 MHz, CDCl₃): δ 172.95, 169.43, 156.11, 136.28, 128.54, 128.14, 127.95, 75.57, 66.89, 66.82, 60.40, 58.36, 52.15, 51.07, 41.27, 28.18, 25.01, 22.80, 21.95, 21.06, 16.39, 14.21.

(S)-methyl 2-((2S,3R)-2-amino-3-(tert-butoxy)butanamido)-4methylpentanoate (MPI26f). MPI26f was prepared as a white solid following a similar procedure to MPI125f. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.88 (d, *J* = 8.1 Hz, 1H), 4.61 – 4.43 (m, 1H), 4.19 – 4.06 (m, 1H), 3.72 (s, 3H), 3.21 (d, *J* = 3.2 Hz, 1H), 1.75 – 1.51 (m, 3H), 1.18 (s, 12H), 1.04 – 0.86 (m, 6H). ¹³C NMR (100 MHz, CDCl₃): δ 173.53, 173.48, 74.14, 67.85, 59.58, 52.11, 50.73, 41.23, 28.53, 24.90, 22.85, 21.91, 19.27.

(S)-methyl 2-((2S,3R)-3-(tert-butoxy)-2-((((3chlorobenzyl)oxy)carbonyl)amino)butanamido)-4-methylpentanoate (MPI126g). MPI26g was prepared as a white solid following a similar procedure to MPI125g (yield 51%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.61 (d, *J* = 7.8 Hz, 1H), 7.36 – 7.09 (m, 4H), 5.91 (d, *J* = 5.0 Hz, 1H), 5.11 – 4.93 (m, 2H), 4.52 – 4.38 (m, 1H), 4.24 – 3.98 (m, 2H), 3.66 (s, 3H), 1.70 – 1.54 (m, 2H), 1.51 (q, *J* = 9.0, 8.4 Hz, 1H), 1.24 (s, 9H), 1.05 (d, *J* = 6.2 Hz, 3H), 0.87 (dd, *J* = 6.1, 3.6 Hz, 6H).¹³C NMR (100 MHz, CDCl₃): δ 172.95, 169.36, 155.83, 138.36, 134.41, 129.83, 128.23, 127.85, 125.81, 75.64, 66.83, 65.84, 58.32, 52.19, 51.10, 41.24, 28.16, 25.02, 22.81, 21.95, 16.34.

(S)-2-((2S,3R)-3-(tert-butoxy)-2-((((3-

chlorobenzyl)oxy)carbonyl)amino)butanamido)-4-methylpentanoic acid (MPI126c). MPI26c was prepared as a white solid following a similar procedure to MPI11c. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.63 (d, *J* = 7.5 Hz, 1H), 7.37 – 7.09 (m, 4H), 5.97 (d, *J* = 5.4 Hz, 1H), 5.01 (q, *J* = 12.7 Hz, 2H), 4.44 (q, *J* = 7.2, 6.0 Hz, 1H), 4.25 – 4.07 (m, 2H), 1.72 – 1.59 (m, 2H), 1.59 – 1.43 (m, 1H), 1.22 (s, 9H), 1.02 (d, *J* = 6.3 Hz, 3H), 0.95 – 0.80 (m, 6H). ¹³C NMR (100 MHz, CDCl₃): δ 177.19, 169.79, 155.91, 138.31, 134.41, 129.83, 128.25, 127.86, 125.82, 75.70, 67.95, 66.91, 65.92, 58.35, 51.08, 40.96, 28.14, 25.03, 22.84, 21.84, 16.42.

(5S,8S,11S)-methyl 5-((R)-1-(tert-butoxy)ethyl)-1-(3-chlorophenyl)-8-isobutyl-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-2-oxa-4,7,10-triazadodecan-12oate (MPI126d). MPI26d was prepared as a white solid following a similar procedure to MPI11d (yield 60%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.61 (d, *J* = 7.4 Hz, 1H), 7.42 (d, *J* = 7.9 Hz, 1H), 7.33 – 7.12 (m, 4H), 6.58 (s, 1H), 5.88 (d, *J* = 5.3 Hz, 1H), 5.01 (q, *J* = 12.6 Hz, 2H), 4.54 – 4.33 (m, 2H), 4.10 (d, *J* = 5.5 Hz, 2H), 3.65 (s, 3H), 3.36 – 3.18 (m, 2H), 2.52 – 2.25 (m, 2H), 2.17 – 2.05 (m, 1H), 1.87 – 1.71 (m, 2H), 1.67 – 1.59 (m, 2H), 1.49 (q, *J* = 8.9, 8.4 Hz, 1H), 1.20 (s, 9H), 1.00 (d, *J* = 6.0 Hz, 3H), 0.88 (dd, *J* = 11.7, 6.0 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃): δ 179.96, 172.16, 172.07, 169.48, 155.92, 138.30, 134.41, 129.87, 128.29, 127.93, 125.94, 75.50, 66.73, 65.97, 58.83, 54.58, 52.43, 41.51, 40.82, 38.35, 33.08, 28.21, 24.78, 22.86, 22.12, 18.53, 17.18, 17.14.

3-chlorobenzyl ((2S,3R)-3-(tert-butoxy)-1-(((S)-1-(((S)-1-hydroxy-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)-4-methyl-1-oxopentan-2-yl)amino)-1-oxobutan-2-yl)carbamate (MPI126e). MPI26e was prepared as a white solid following a similar procedure to MPI11e (yield 73%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.66 (d, *J* = 7.9 Hz, 1H), 7.40 (d, *J* = 7.8 Hz, 1H), 7.33 – 7.10 (m, 4H), 6.28 – 6.06 (m, 2H), 5.08 – 4.88 (m, 2H), 4.34 (td, *J* = 8.2, 5.2 Hz, 1H), 4.14 – 4.06 (m, 1H), 4.02 – 3.80 (m, 1H), 3.74 (t, *J* = 6.4 Hz, 1H), 3.61 – 3.50 (m, 2H), 3.30 – 3.13 (m, 2H), 2.33 (dt, *J* = 8.6, 5.1 Hz, 2H), 2.15 – 1.96 (m, 1H), 1.82 – 1.68 (m, 1H), 1.64 – 1.32 (m, 4H), 1.20 (s, 9H), 1.00 (d, *J* = 6.3 Hz, 3H), 0.84 (dd, *J* = 9.0, 6.1 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃): δ 181.06, 172.62, 169.58, 156.06, 138.34, 134.40, 129.86, 128.27, 127.94, 125.94, 75.52, 66.85, 65.97, 65.85, 58.98, 52.37, 50.31, 41.29, 40.55, 38.29, 32.66, 28.47, 28.21, 24.90, 22.77, 22.14, 17.21.

3-chlorobenzyl ((2S,3R)-3-(tert-butoxy)-1-(((S)-4-methyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)pentan-2-yl)amino)-1-oxobutan-2-yl)carbamate (MPI126). MPI26 was prepared as a white solid following a similar procedure to MPI11 (yield 79%). ¹H NMR (400 MHz, Chloroform-*d***) \delta 9.50 (s, 1H), 8.39 – 7.96 (m, 1H), 7.47 (d,** *J* **= 7.6 Hz, 1H), 7.35 (s, 1H), 7.29 (d,** *J* **= 4.7 Hz, 2H), 7.23 (s, 1H), 6.19 – 5.86 (m, 2H), 5.07 (q,** *J* **= 12.6 Hz, 2H), 4.55 – 4.34 (m, 2H), 4.18 (d,** *J* **= 5.3 Hz, 2H), 3.37 – 3.26 (m, 2H), 2.56 – 2.26 (m, 3H), 2.01 – 1.92 (m, 1H), 1.87 – 1.77 (m, 1H), 1.77 – 1.64 (m, 2H), 1.59 (t,** *J* **= 9.0 Hz, 1H), 1.27 (s, 9H), 1.08 (d,** *J* **= 5.9 Hz, 3H), 0.96 (dd,** *J* **= 10.9, 5.7 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) \delta 199.64,**

172.82, 169.54, 169.33, 155.98, 138.29, 134.42, 129.88, 128.31, 127.95, 125.96, 75.53, 66.74, 66.00, 58.91, 57.66, 52.11, 41.48, 40.61, 28.60, 28.22, 24.93, 22.92, 22.08, 17.21.

(((3-chlorobenzyl)oxy)carbonyl)-L-valine (MPI27a). MPI27a was prepared as a white solid following a similar procedure to MPI125g and followed by MPI111c protocol. ¹H NMR (400 MHz, DMSO- d_6) δ 12.58 (s, 1H), 7.55 (d, *J* = 8.6 Hz, 1H), 7.47 – 7.25 (m, 4H), 5.18 – 4.98 (m, 2H), 3.87 (dd, *J* = 8.6, 5.8 Hz, 1H), 2.15 – 1.96 (m, 1H), 0.89 (t, *J* = 6.5 Hz, 6H).

Methyl(S)-2-((S)-2-((((3-chlorobenzyl)oxy)carbonyl)amino)-3-methylbutanamido)-3-cyclohexylpropanoate (MPI127b). MPI27b was prepared asa white solid following a similar procedure to MPI11b (yield 56%). ¹H NMR (400MHz, DMSO- d_6) δ 8.25 (d, J = 7.5 Hz, 1H), 7.48 – 7.22 (m, 5H), 5.11 – 4.95 (m, 2H),4.33 (q, J = 7.7, 7.2 Hz, 1H), 3.88 (t, J = 8.1 Hz, 1H), 3.60 (s, 3H), 2.01 – 1.89 (m, 1H),1.72 – 1.44 (m, 7H), 1.34 (s, 1H), 1.22 – 1.00 (m, 3H), 0.98 – 0.69 (m, 8H).

(S)-2-((S)-2-((((3-chlorobenzyl)oxy)carbonyl)amino)-3-methylbutanamido)-3cyclohexylpropanoic acid (MPI127c). MPI27c was prepared as a white solid following a similar procedure to MPI11c (yield 83%).

Methyl (5S,8S,11S)-1-(3-chlorophenyl)-8-(cyclohexylmethyl)-5-isopropyl-3,6,9-trioxo-11-(((S)-2-oxopyrrolidin-3-yl)methyl)-2-oxa-4,7,10-triazadodecan-12-oate (MPI127d). MPI27d was prepared as a white solid following a similar procedure to MPI11d (yield 56%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.81 (d, *J* = 7.2 Hz, 1H), 7.35 (s, 1H), 7.32 – 7.18 (m, 3H), 6.60 (d, *J* = 8.4 Hz, 1H), 6.02 (s, 1H), 5.42 (d, *J* = 8.6 Hz, 1H), 5.15 – 5.00 (m, 2H), 4.58 (td, *J* = 8.8, 5.5 Hz, 1H), 4.49 (d, *J* = 6.6 Hz, 1H), 3.99 (dd, *J* = 8.6, 6.2 Hz, 1H), 3.72 (s, 3H), 3.41 – 3.24 (m, 2H), 2.48 – 2.35 (m, 2H), 2.23 – 2.05 (m, 2H), 2.01 – 1.58 (m, 8H), 1.56 – 1.46 (m, 1H), 1.40 – 1.04 (m, 5H), 1.04 – 0.77 (m, 8H).

3-chlorobenzyl ((S)-1-(((S)-3-cyclohexyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)propan-2-yl)amino)-3-methyl-1-oxobutan-**2-yl)carbamate** (MPI127). MPI27 was prepared as a white solid following a similar procedure of MPI11e followed by MPI11 procedure (yield 52%). ¹H NMR (400 MHz, Chloroform-*d*) δ 9.49 (s, 1H), 8.25 (d, *J* = 6.6 Hz, 1H), 7.39 – 7.11 (m, 4H), 7.02 (d, *J* = 8.3 Hz, 1H), 6.56 (s, 1H), 5.64 (d, *J* = 8.5 Hz, 1H), 5.06 (q, *J* = 12.6 Hz, 2H), 4.74 – 4.61 (m, 1H), 4.37 (s, 1H), 4.03 (t, *J* = 7.6 Hz, 1H), 3.41 – 3.22 (m, 2H), 2.50 – 2.27 (m, 2H), 2.19 – 2.07 (m, 1H), 2.07 – 1.86 (m, 3H), 1.86 – 1.48 (m, 8H), 1.39 – 1.04 (m, 5H), 1.04 – 0.81 (m, 8H). ¹³C NMR (100 MHz, CDCl₃) δ 199.5, 180.0, 173.3, 171.2, 156.4, 138.2, 134.4, 129.9, 128.3, 128.0, 126.0, 66.2, 60.6, 57.5, 51.1, 40.6, 40.2, 38.0, 34.2, 33.5, 32.5, 31.0, 29.9, 29.7, 28.4, 26.3, 26.2, 26.0, 19.2, 17.8.

Methyl (S)-2-((S)-2-((S)-2-amino-3-methylbutanamido)-4methylpentanamido)-3-((S)-2-oxopyrrolidin-3-yl)propanoate (MPI128f). MPI28f was prepared as a white solid following a similar procedure to MPI125f (yield 90%).

Methyl (S)-2-((S)-2-(1H-indole-2-carboxamido)-3-methylbutanamido)-4methylpentanamido)-3-((S)-2-oxopyrrolidin-3-yl)propanoate (MPI128d). MPI28d was prepared as a white solid following a similar procedure to MPI11d (yield 53%). ¹H NMR (400 MHz, Methanol-44) δ 7.62 (d, J = 8.0 Hz, 1H), 7.49 – 7.39 (m, 1H), 7.22 (ddd, J = 8.2, 7.0, 1.2 Hz, 1H), 7.18 (d, J = 0.8 Hz, 1H), 7.11 – 7.01 (m, 1H), 4.53 (dd, J = 11.8, 3.9 Hz, 1H), 4.42 (dd, J = 13.8, 7.7 Hz, 2H), 3.72 (s, 3H), 2.56 (qd, J = 10.4, 4.0 Hz, 1H), 2.35 – 2.14 (m, 3H), 1.87 – 1.69 (m, 3H), 1.63 (t, J = 7.2 Hz, 2H), 1.04 (d, *J* = 6.7 Hz, 6H), 0.94 (dd, *J* = 19.4, 6.5 Hz, 6H).

N-((S)-3-methyl-1-(((S)-4-methyl-1-oxo-1-(((S)-1-oxo-3-((S)-2-oxopyrrolidin-3-yl)propan-2-yl)amino)pentan-2-yl)amino)-1-oxobutan-2-yl)-1H-indole-2-carboxamide (MPI128). MPI28 was prepared as a white solid following a similar procedure of **MPI11e** followed by **MPI11** procedure (yield 60%). ¹H NMR (400 MHz, DMSO-d₆) δ 11.59 (d, J = 2.2 Hz, 1H), 9.42 (s, 1H), 8.48 (d, J = 7.8 Hz, 1H), 8.23 (dd, J = 19.8, 8.1 Hz, 2H), 7.71 – 7.57 (m, 2H), 7.50 – 7.39 (m, 1H), 7.29 (d, J = 2.1 Hz, 1H), 7.18 (ddd, J = 8.2, 6.9, 1.2 Hz, 1H), 7.03 (ddd, J = 8.0, 6.9, 1.0 Hz, 1H), 4.44 – 4.32 (m, 2H), 4.24 (ddd, J = 11.6, 7.8, 3.9 Hz, 1H), 3.21 – 3.11 (m, 1H), 3.07 (td, J = 9.2, 7.0 Hz, 1H), 2.31 (qd, J = 10.3, 3.9 Hz, 1H), 2.13 (dt, J = 14.3, 7.9 Hz, 2H), 1.96 – 1.85 (m, 1H), 1.73 – 1.58 (m, 3H), 1.58 – 1.43 (m, 2H), 1.02 – 0.79 (m, 12H). ¹³C NMR (100 MHz, DMSO-d₆) δ 201.2, 178.8, 173.1, 171.5, 161.5, 137.0, 131.8, 127.5, 123.9, 122.0, 120.2, 112.7, 104.2, 58.8, 56.6, 51.6, 41.2, 37.6, 30.9, 29.8, 27.7, 24.7, 23.3, 22.3, 19.6, 19.2

Table S1. Statistics of crystallographic analysis of M^{Pro} in complexed with different inhibitors.

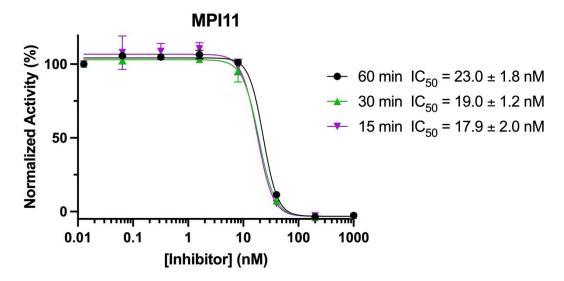
Protein/Ligand (PDI entry)	MPI11 (7RVM)	MPI12 (7RVN)	MPI13 (7RVO)	MPI14 (7RVP)
Data Collection		(/KVN)	(7800)	(/KVP)
Space group	C121	C121	P1	P1
cell dimensions	CIZI	CIZI	F1	FI
a, b, c (Å)	98.71, 81.35, 51.98	98.92, 81.18, 51.92	55.51, 60.54, 63.15	55.58, 60.80, 63.45
	98.71, 81.35, 51.98	98.92, 81.18, 51.92 90.00, 115.15, 90.00	79.98, 68.24, 70.25	80.06, 68.16, 70.12
α, β, γ (°)			Contraction of the second s	
Resolution (Å)	46.93-1.95 (2.00-1.95)	47.00-1.63 (1.66-1.63)	44.34-1.80 (1.84-1.80)	58.81-1.90 (1.94-1.90)
R _{merge}	9.2 (72.0)	6.6 (77.6)	20.8 (57.9)	10.4 (145.9)
l/σl	9.3 (1.5)	12.5 (1.6)	3.5 (1.6)	5.9 (0.9)
Completeness (%)	97.4 (95.8)	95.8 (90.5)	89.8 (90.6)	95.5 (94.5)
Redundancy	6.9 (6.9)	5.9 (5.5)	3.7 (3.9)	3.6 (3.7)
Refinement				
Resolution (Å)	46.93-1.95	47.00-1.63	44.34-1.80	43.48-1.90
No. Reflections	26267 (2594)	44233 (4238)	59690 (6029)	54436 (5351)
R _{work} /R _{free}	0.2055/0.2383	0.1972/0.2124	0.2444/0.2750	0.2368/0.2736
No. atoms				
Protein	2400	2426	4798	4802
Water	230	183	246	173
B factors				
Protein	23.652	40.122	31.900	44.602
Water	30.233	43.562	36.757	44.891
R.m.s deviations				
Bond lengths (Å)	0.009	0.009	0.010	0.010
Bond angles (°)	1.67	1.38	2.38	1.41
Protein/Ligand	MPI16	MPI18	MPI19	MPI20
(PDI entry)	(7RVQ)	(7RVR)	(7RVS)	(7RVT)
Data Collection	· · ·		A	
Space group	P1	P1	C121	P 1
	P1	P1	C121	P 1
cell dimensions				
cell dimensions a, b, c (Å)	56.34, 62.85, 63.57	54.70, 61.66, 62.10	98.80, 81.80, 52.03	55.51, 60.70, 63.34
cell dimensions a, b, c (Å) α, β, γ (°)	56.34, 62.85, 63.57 80.34, 68.56, 69.65	54.70, 61.66, 62.10 81.13, 69.31, 69.71	98.80, 81.80, 52.03 90.00, 115.5, 90.00	55.51, 60.70, 63.34 80.42, 68.42, 70.59
cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å)	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47)	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46)	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85)	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10)
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge}	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6)	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3)	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7)	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2)
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} I/σI	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3)	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5)	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6)	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6)
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} I/σI Completeness (%)	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8)	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8)	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0)	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3)
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} I/σI Completeness (%) Redundancy	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3)	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5)	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6)	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6)
cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) <i>R_{merge}</i> <i>I/σI</i> Completeness (%) Redundancy Refinement	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1)	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8)	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5)	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1)
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} I/σI Completeness (%) Redundancy Refinement Resolution (Å)	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} I/σI Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018)	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758)	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128)	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014)
zell dimensions a, b, c (Å) a, β, γ (°) Resolution (Å) Remerge 1/a/ Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections Rework/Riree	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10
zell dimensions a, b, c (Å) a, β, γ (°) Resolution (Å) Remerge Vol Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections R _{work} /R _{free} No. atoms	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018) 0.2386/0.3222	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758) 0.2529/0.3231	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128) 0.2000/0.2265	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014) 0.2702/0.3076
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} t/σl Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections R _{work} /R _{free} No. atoms Protein	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018) 0.2386/0.3222 2431	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758) 0.2529/0.3231 2401	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128) 0.2000/0.2265 2427	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014) 0.2702/0.3076 2433
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} t/σl Completeness (%) Redundancy Redundancy Refinement Resolution (Å) No. Reflections R _{work} /R _{free} No. atoms Protein Water	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018) 0.2386/0.3222	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758) 0.2529/0.3231	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128) 0.2000/0.2265	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014) 0.2702/0.3076
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} t/σl Completeness (%) Redundancy Redundancy Refinement Resolution (Å) No. Reflections R _{work} /R _{free} No. atoms Protein Water	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018) 0.2386/0.3222 2431	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758) 0.2529/0.3231 2401	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128) 0.2000/0.2265 2427	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014) 0.2702/0.3076 2433
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} t/σl Completeness (%) Redundancy Redundancy Refinement Resolution (Å) No. Reflections R _{work} /R _{free} No. atoms Protein Water	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018) 0.2386/0.3222 2431	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758) 0.2529/0.3231 2401	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128) 0.2000/0.2265 2427	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014) 0.2702/0.3076 2433
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} //σ/ Completeness (%) Redundancy Redundancy Refinement Resolution (Å) No. Reflections <i>R</i> work/ <i>R</i> free No. atoms Protein Water <i>B</i> factors	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018) 0.2386/0.3222 2431 45	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758) 0.2529/0.3231 2401 37	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128) 0.2000/0.2265 2427 176	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014) 0.2702/0.3076 2433 144
cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R _{merge} I/σl Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections R _{work} /R _{free} No. atoms Protein Water B factors Protein Water	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018) 0.2386/0.3222 2431 45 50.449	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758) 0.2529/0.3231 2401 37 50.903	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128) 0.2000/0.2265 2427 176 37.652	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014) 0.2702/0.3076 2433 144 31.883
α, β, γ (°) Resolution (Å) R _{merge} I/σ/ Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections R _{work} /R _{free} No. atoms Protein Water B factors Protein	56.34, 62.85, 63.57 80.34, 68.56, 69.65 24.91-2.47 (2.58-2.47) 41.3 (419.6) 3.5 (0.3) 98.1 (84.8) 8.9 (5.1) 24.91-2.49 13243 (1018) 0.2386/0.3222 2431 45 50.449	54.70, 61.66, 62.10 81.13, 69.31, 69.71 24.27-2.46 (2.56-2.46) 21.7 (231.3) 8.0 (0.5) 98.4 (86.8) 13.1 (7.8) 24.27-2.46 12359 (758) 0.2529/0.3231 2401 37 50.903	98.80, 81.80, 52.03 90.00, 115.5, 90.00 24.41-1.85 (1.89-1.85) 6.9 (118.7) 17.8 (1.6) 99.9 (100.0) 10.1 (7.5) 24.41-1.85 31512 (3128) 0.2000/0.2265 2427 176 37.652	55.51, 60.70, 63.34 80.42, 68.42, 70.59 58.83-2.10 (2.16-2.10) 30.0 (54.2) 3.8 (2.6) 95.8 (96.3) 2.9 (3.1) 32.85 - 2.10 40069 (4014) 0.2702/0.3076 2433 144 31.883

Table S1 – continued

Protein/Ligand	MPI21	MPI22 (7RVV)	MPI23	MPI24
(PDI entry) Data Collection	(7RVU)	(/KVV)	(7RVW)	(7RVX)
Space group	P1	P1	C1	P1
cell dimensions	F1	F1	CI	F1
a, b, c (Å)	55.60, 60.93, 62.87	46.46, 52.26, 64.12	95.91, 81.31, 54.47	56.06, 61.15, 63.76
	79.27, 68.84, 69.73	108.82, 97.3, 98.14	90.00, 116.98, 90.00	79.94, 68.33, 70.17
α, β, γ (°)				
Resolution (Å)	57.76-2.50 (2.60-2.50)	48.52-3.00 (3.18-3.00)	24.27-1.85 (1.89-1.85)	49.69-1.85 (1.89-1.85)
R _{merge}	47.1 (91.8)	10.5 (170.0)	13.6 (98.3)	8.3 (51.4)
l/σl	2.1 (0.8)	6.1 (4.0)	13.3 (2.1)	7.6 (1.5)
Completeness (%)	98.9 (98.7)	93.2 (95.9)	99.9 (100.0)	89.1 (85.9)
Redundancy	5.4 (5.6)	2.8 (2.9)	11.4 (9.1)	3.8 (3.8)
Refinement				
Resolution (Å)	43.51-2.50	48.52-3.00	24.27-1.85	49.69-1.85
No. Reflections	12573 (1249)	10372 (1108)	31811 (3152)	56153 (5444)
R _{work} /R _{free}	0.2515/0.3285	0.3414/0.4279	0.1898/0.2175	0.2353/0.2651
No. atoms				
Protein	2378	2376	2401	4840
Water	43	0	223	229
B factors				
Protein	29.165	22.608	30.173	40.269
Water	26.273	0	34.473	42.174
R.m.s deviations				
Bond lengths (Å)	0.010	0.010	0.009	0.012
Bond angles (°)	1.28	1.38	1.24	1.50
Protein/Ligand	MPI25	MPI26	MPI27	MPI28
(PDI entry)				
(FDI entry)	(7RVY)	(7RVZ)	(7RV0)	(7RV1)
	(7RVY)	(7RVZ)	(7RV0)	(7RV1)
Data Collection	(7RVY) C1	(7RVZ)	(7RV0) C1	(7RV1) P1
Data Collection Space group		•••••••••••		
Data Collection Space group cell dimensions	C1	C121	C1	P1
Data Collection Space group cell dimensions a, b, c (Å)	C1 96.50, 81.24, 54.48	C121 98.33, 80.95, 51.82	C1 95.66, 80.81, 54.44	P1 55.64, 61.09, 63.87
Data Collection Space group cell dimensions a, b, c (Å) α, β, γ (°)	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å)	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85)	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90)	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85)	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50)
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) R _{merge}	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7)	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9)	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8)	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8)
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) R _{merge} I/σl	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5)	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3)	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7)	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8)
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/σI</i> Completeness (%)	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3)	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6)	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9)	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4)
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/σI</i> Completeness (%) Redundancy	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5)	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3)	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7)	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8)
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) R _{merge} I/ol Completeness (%) Redundancy Refinement	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7)	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5)	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0)	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1)
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) R _{merge} I/ol Completeness (%) Redundancy Refinement Resolution (Å)	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/σI</i> Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613)	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745)	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102)	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442)
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/σI</i> Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections <i>R</i> _{work} / <i>R</i> _{free}	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/σI</i> Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections <i>R</i> _{work} / <i>R</i> _{free} No. atoms	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613) 0.2309/0.2680	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745) 0.1911/0.2199	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102) 0.2220/0.2534	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442) 0.2566/0.3160
Data Collection Space group cell dimensions <i>a, b, c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/σI</i> Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections <i>R</i> _{work} / <i>R</i> _{free} No. atoms Protein	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613) 0.2309/0.2680 2407	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745) 0.1911/0.2199 2358	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102) 0.2220/0.2534 2403	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442) 0.2566/0.3160 4722
Data Collection Space group cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R_{merge} I/of Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections R_{work}/R_{free} No. atoms Protein Water	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613) 0.2309/0.2680	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745) 0.1911/0.2199	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102) 0.2220/0.2534	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442) 0.2566/0.3160
Data Collection Space group cell dimensions a, b, c (Å) α, β, γ (°) Resolution (Å) R_{merge} $1/\sigma l$ Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections R_{work}/R_{free} No. atoms Protein Water B factors	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613) 0.2309/0.2680 2407 232	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745) 0.1911/0.2199 2358 103	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102) 0.2220/0.2534 2403 200	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442) 0.2566/0.3160 4722 122
Data Collection Space group cell dimensions <i>a</i> , <i>b</i> , <i>c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/dI</i> Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections <i>R</i> _{work} / <i>R</i> _{free} No. atoms Protein Water <i>B</i> factors Protein	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613) 0.2309/0.2680 2407 232 22.457	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745) 0.1911/0.2199 2358 103 44.839	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102) 0.2220/0.2534 2403 200 28.397	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442) 0.2566/0.3160 4722 122 16.575
Data Collection Space group cell dimensions <i>a</i> , <i>b</i> , <i>c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/dI</i> Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections <i>R</i> _{work} / <i>R</i> _{free} No. atoms Protein Water <i>B</i> factors Protein Water	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613) 0.2309/0.2680 2407 232	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745) 0.1911/0.2199 2358 103	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102) 0.2220/0.2534 2403 200	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442) 0.2566/0.3160 4722 122
Data Collection Space group cell dimensions <i>a</i> , <i>b</i> , <i>c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/dI</i> Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections <i>R</i> _{work} / <i>R</i> _{free} No. atoms Protein Water <i>B</i> factors Protein Water <i>R</i> .m.s deviations	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613) 0.2309/0.2680 2407 232 22.457 25.228	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745) 0.1911/0.2199 2358 103 44.839 44.799	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102) 0.2220/0.2534 2403 200 28.397 31.384	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442) 0.2566/0.3160 4722 122 16.575 15.514
Data Collection Space group cell dimensions <i>a</i> , <i>b</i> , <i>c</i> (Å) α, β, γ (°) Resolution (Å) <i>R</i> _{merge} <i>I/dI</i> Completeness (%) Redundancy Refinement Resolution (Å) No. Reflections <i>R</i> _{work} / <i>R</i> _{free} No. atoms Protein Water <i>B</i> factors Protein	C1 96.50, 81.24, 54.48 90.00, 117.06, 90.00 24.21-1.85 (1.89-1.85) 6.5 (38.7) 12.2 (2.5) 92.6 (81.3) 4.2 (3.7) 24.21-1.85 29597 (2613) 0.2309/0.2680 2407 232 22.457	C121 98.33, 80.95, 51.82 90.00, 115.19, 90.00 59.88-1.90 (1.94-1.90) 10.1 (141.9) 8.7 (1.3) 96.1 (95.6) 6.4 (6.5) 43.45-1.90 27763 (2745) 0.1911/0.2199 2358 103 44.839	C1 95.66, 80.81, 54.44 90.00, 116.96, 90.00 24.26-1.85 (1.89-1.85) 12.2 (90.8) 8.8 (1.7) 99.8 (98.9) 6.8 (4.0) 24.26-1.85 31429 (3102) 0.2220/0.2534 2403 200 28.397	P1 55.64, 61.09, 63.87 79.27, 68.06, 69.37 59.13-2.50 (2.60-2.50) 26.2 (37.8) 4.7 (2.8) 97.1 (97.4) 2.9 (3.1) 46.10 - 2.50 24135 (2442) 0.2566/0.3160 4722 122 16.575

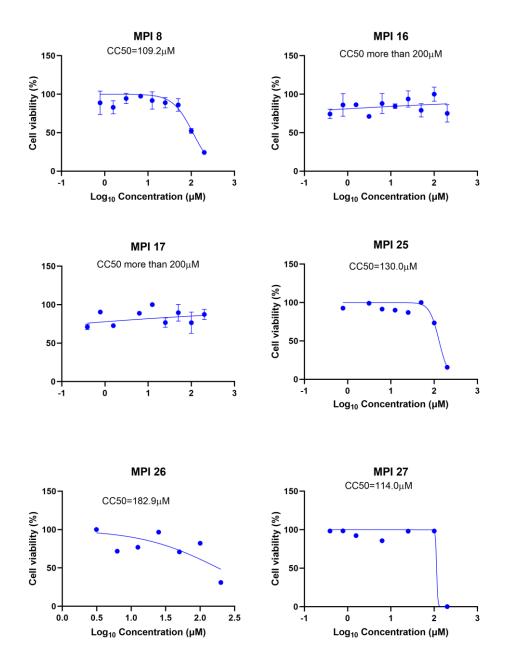
Supplementary Figure 1

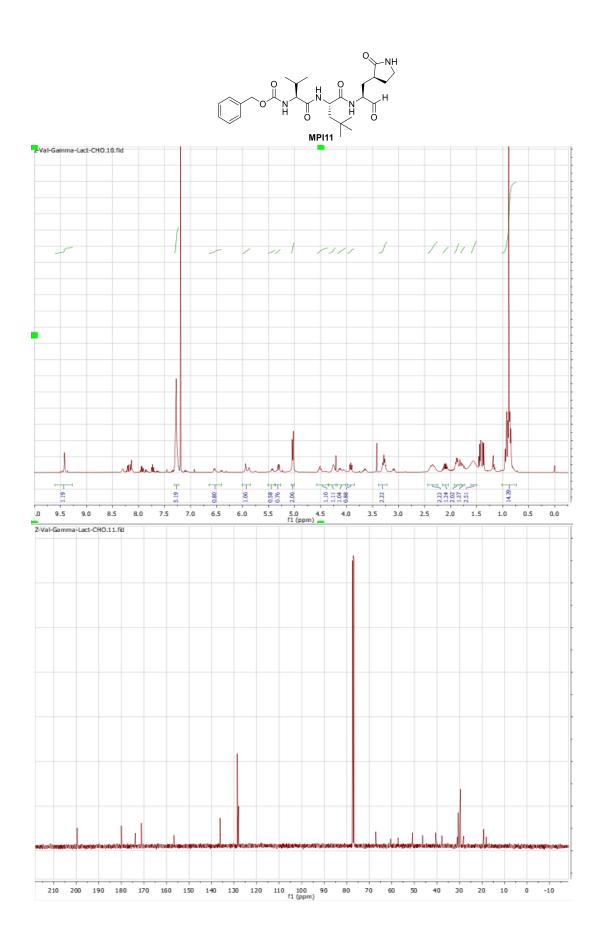
M^{Pro} was incubated with different concentrations of MPI11 for 15, 30 and 60 min and then its activity was determined by adding Sub3.

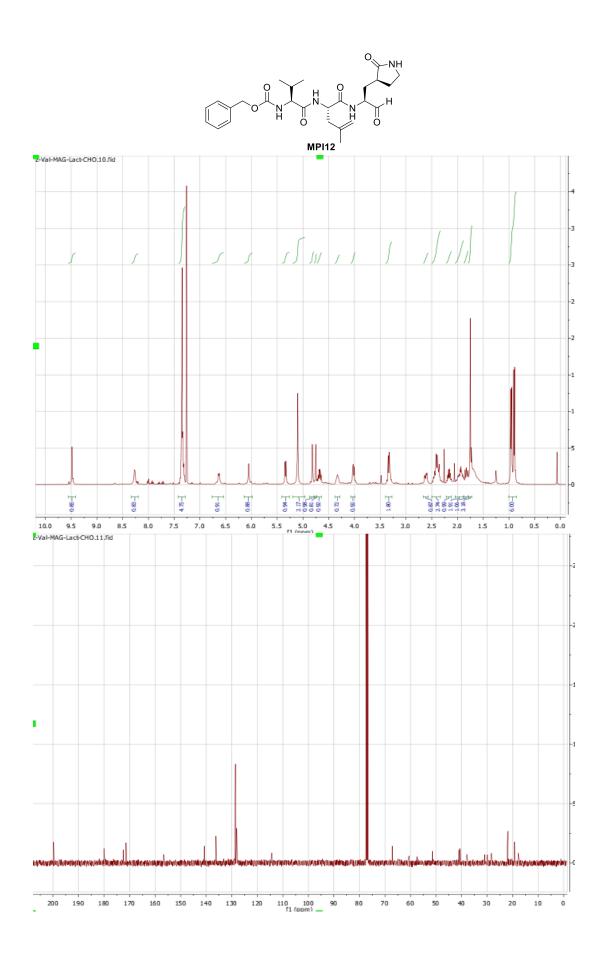


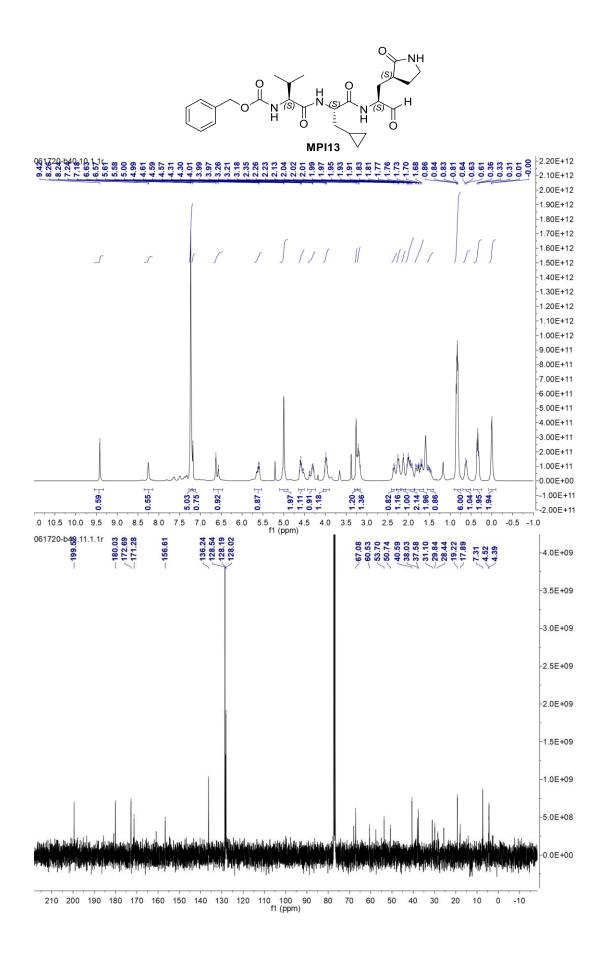
Supplementary Figure S2

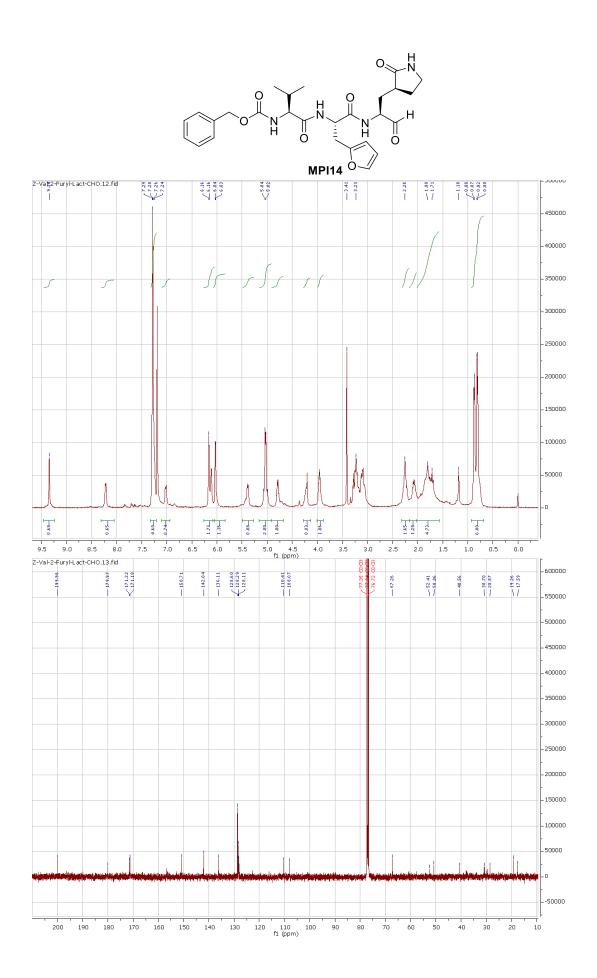
To assess the half-maximal cytotoxic concentration (CC₅₀), stock solutions of the tested compounds were dissolved in DMSO and diluted further to the working solutions with DMEM. Briefly, the 293T cells were seeded in 96 well-plates and incubated at 37 °C and 5% CO₂ for 24 h. After that, the cells were treated with different concentrations (200 μ M, 100 μ M, 50 μ M, 25 μ M, 12.5 μ M, 6.25 μ M, 3.125 μ M, 1.5625 μ M, 0.78125 μ M, 0 μ M) of the tested compounds in triplicates for 48 h. Cell viability was assessed by MTT assay to determine the CC₅₀. All data reported was normalized on a per-plate basis to wells that contained cells in the presence of 200 μ M doxorubincin (0% cell viability). The concentration caused a 50% cytotoxicity (CC₅₀) was obtained by plotting the normalization % cell viability versus log10 sample concentration.

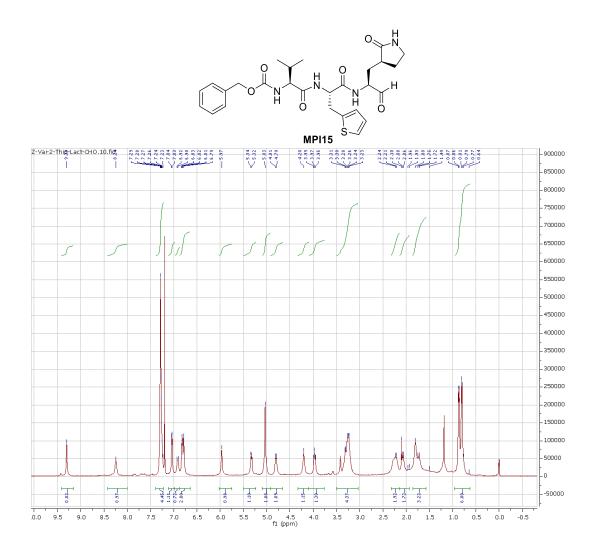


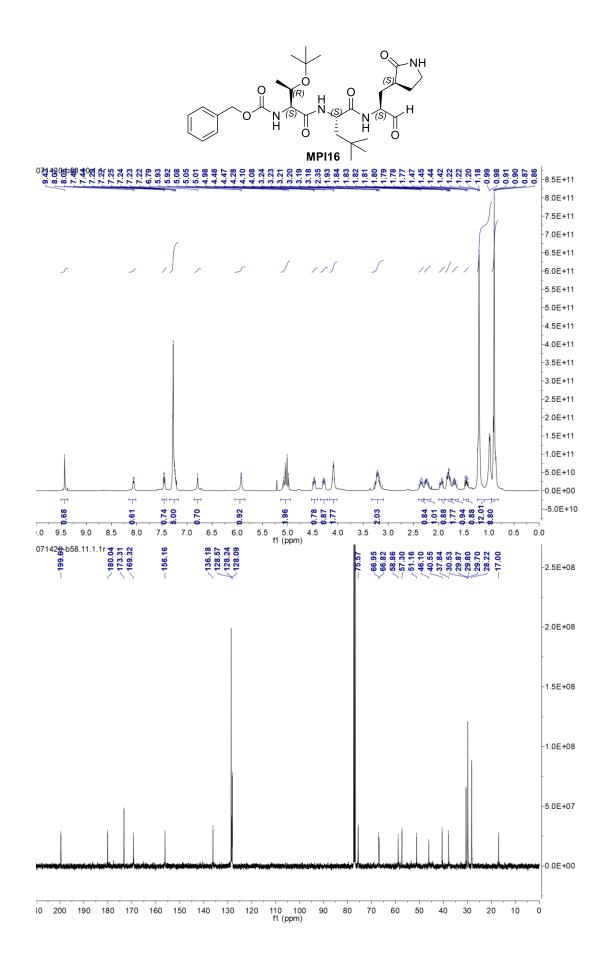


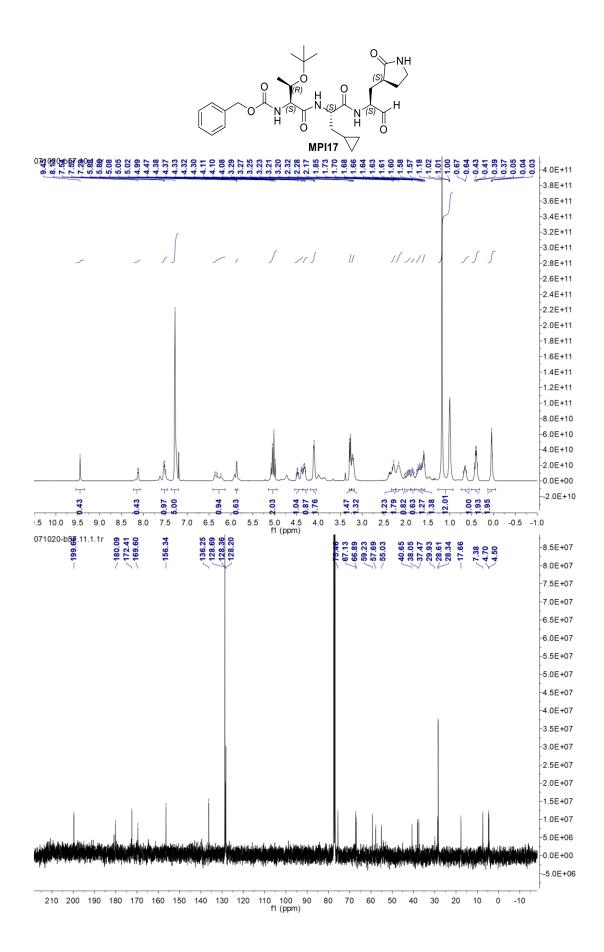


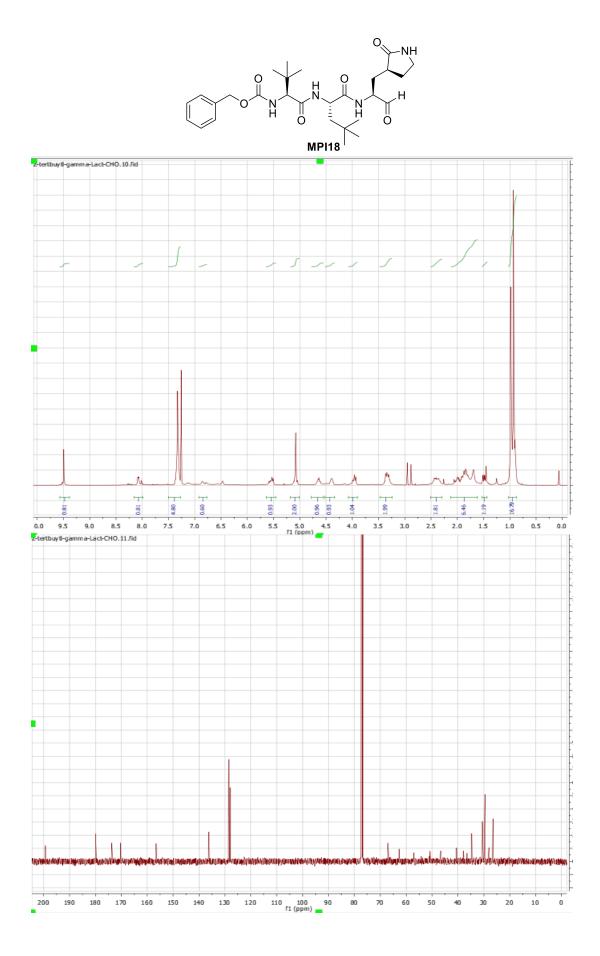


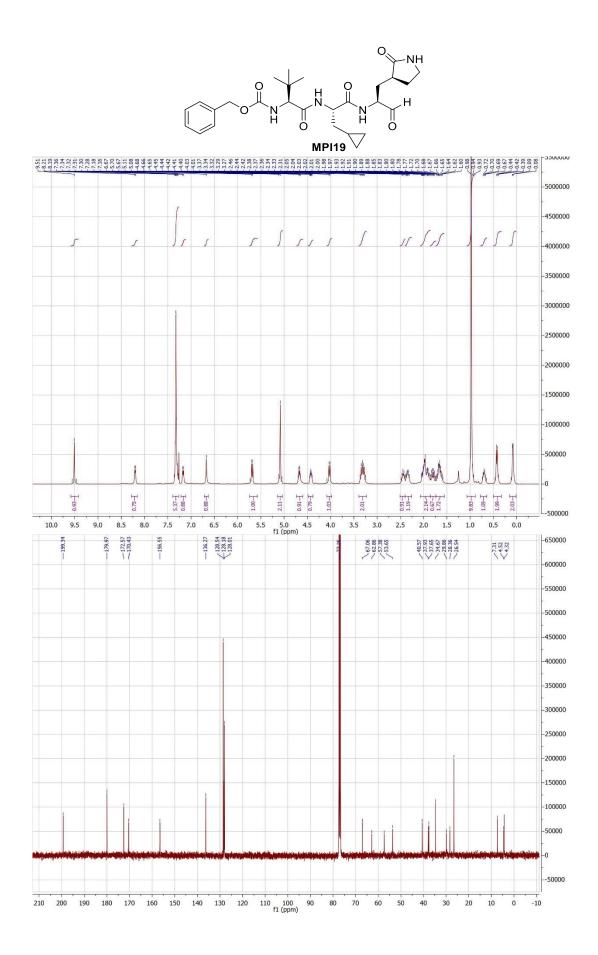


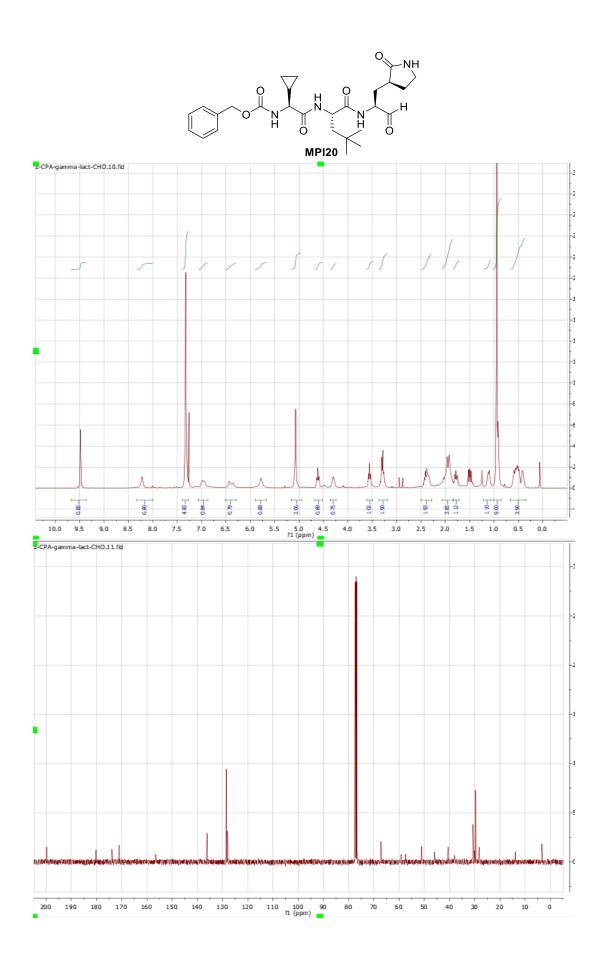


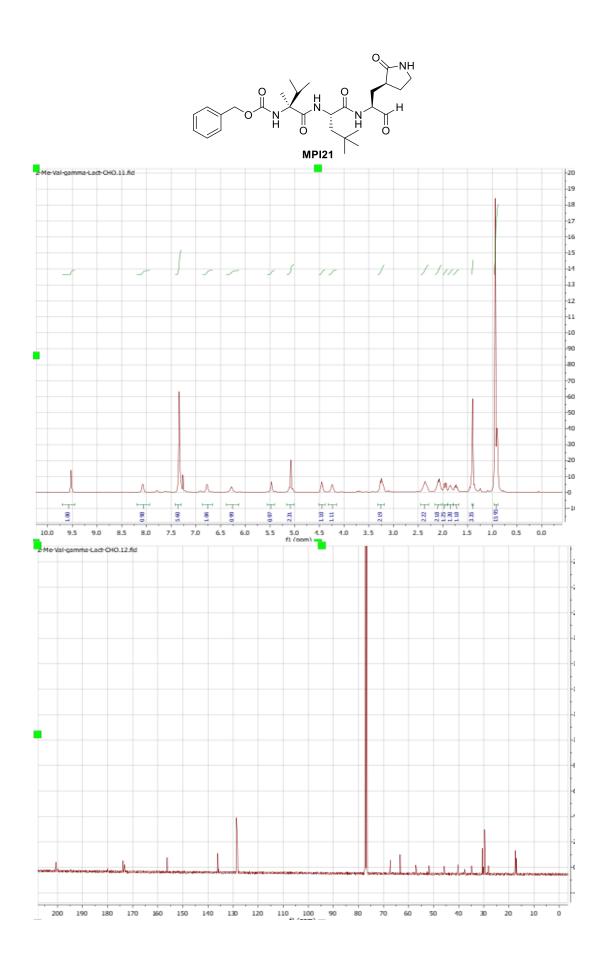


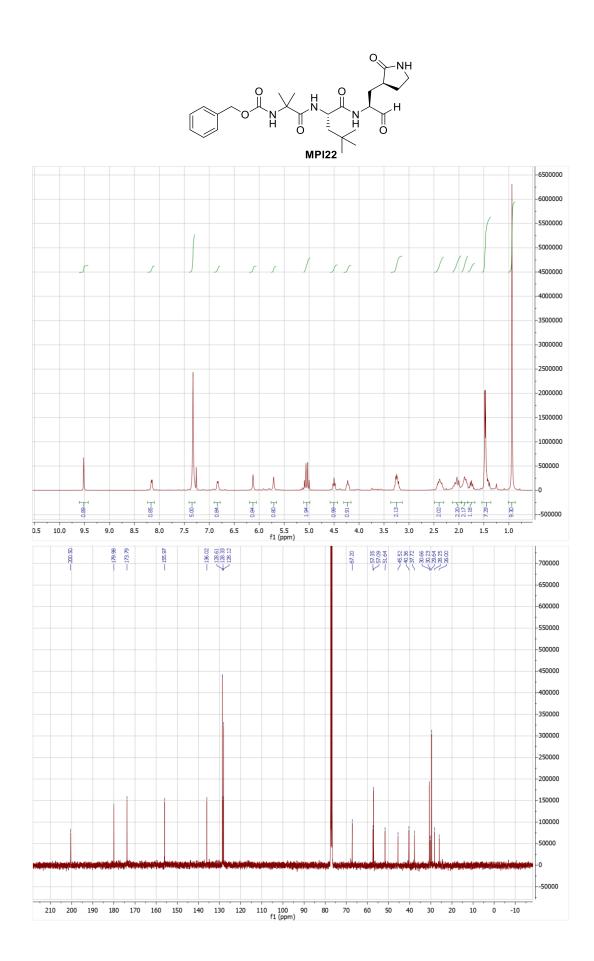


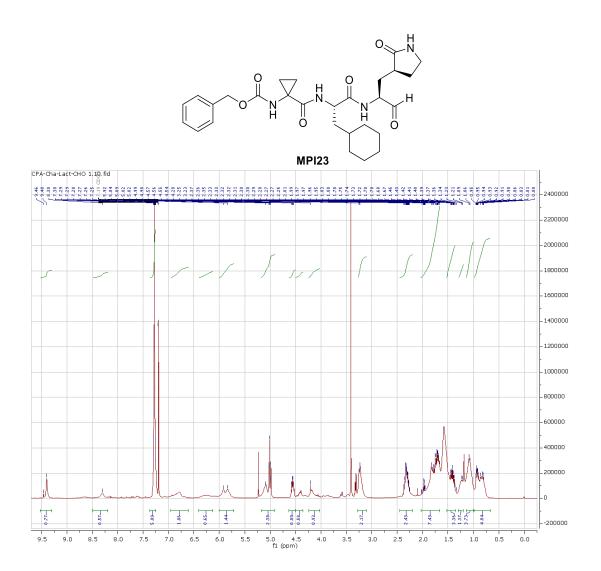


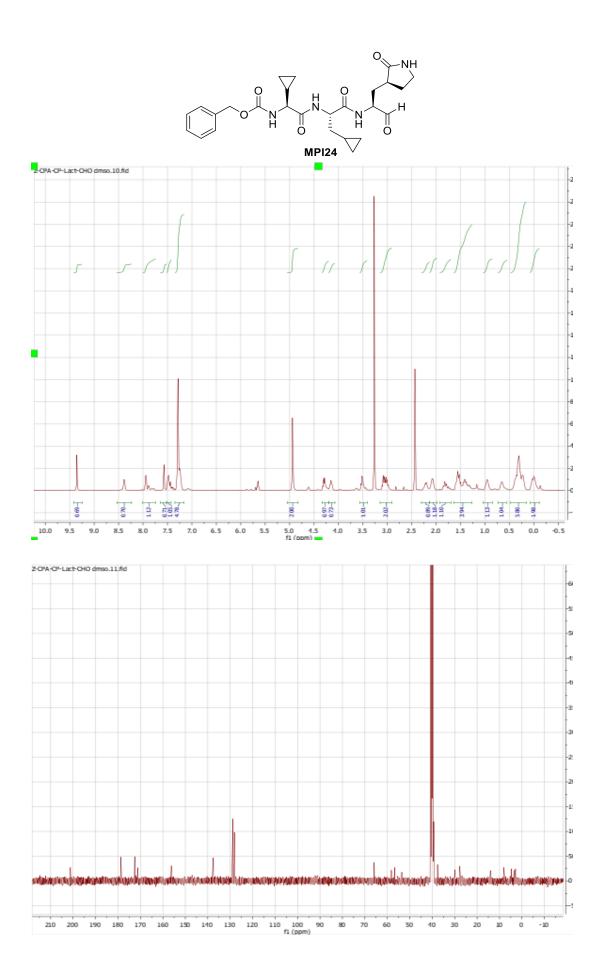


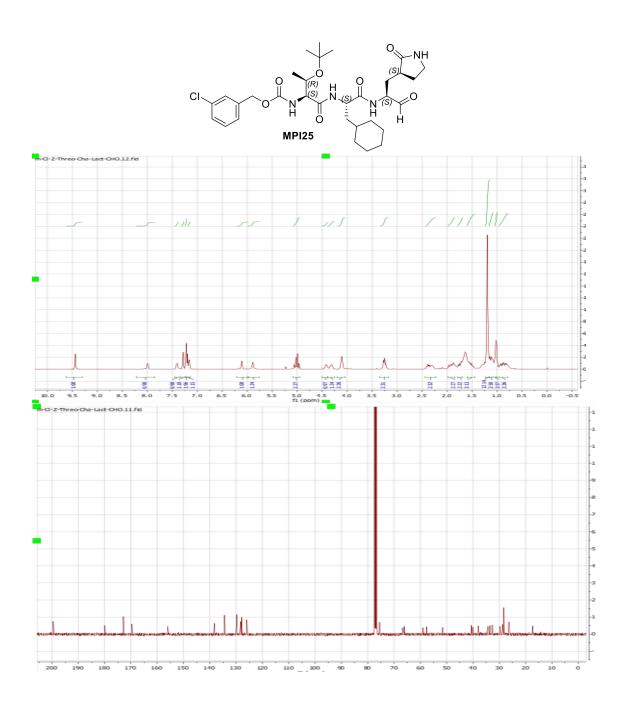


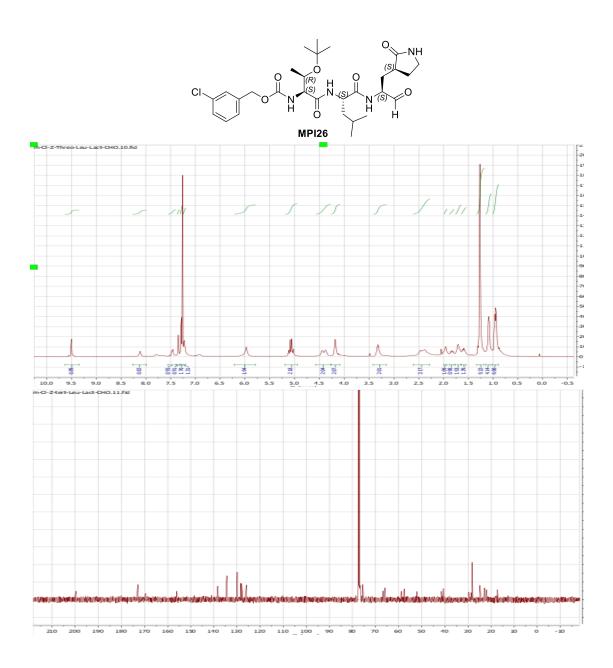


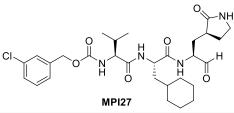


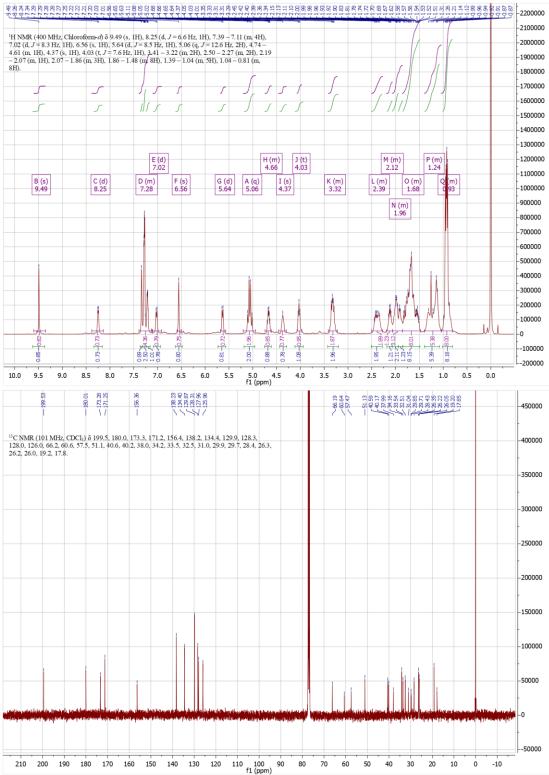


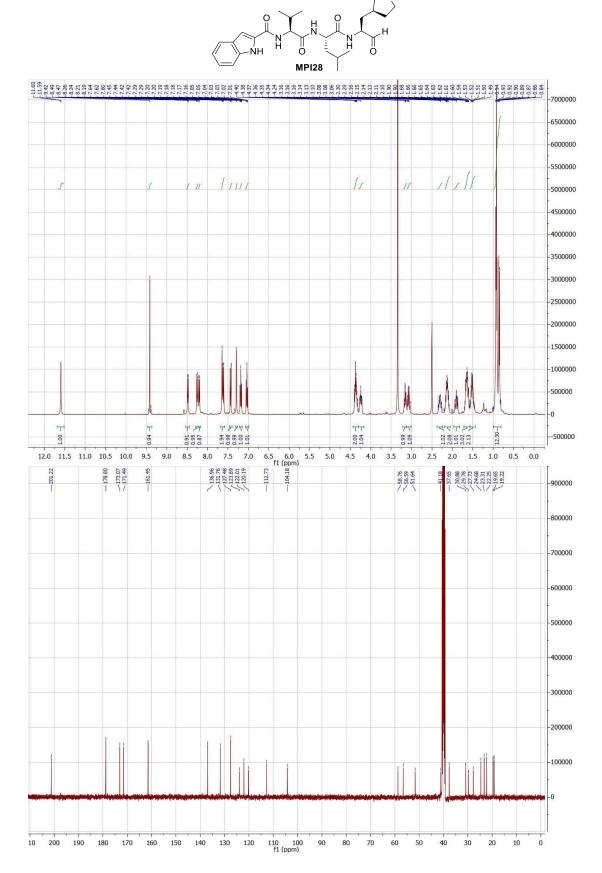




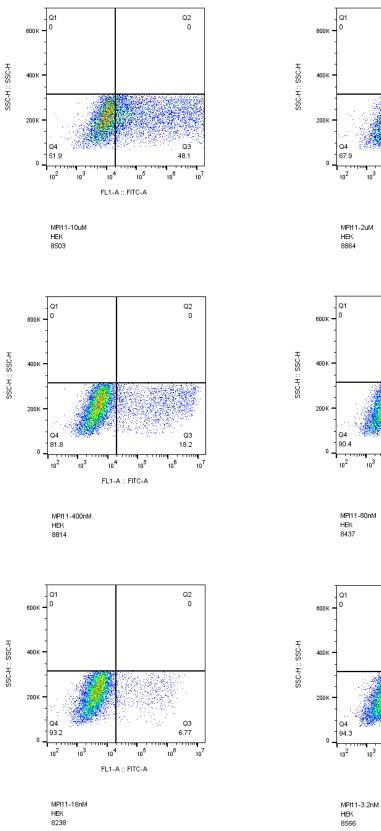


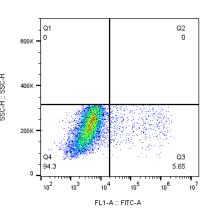




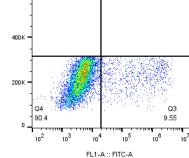


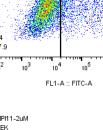
Flow cytometry images for MPI11, compound concentration labeled.











Q3 32.1

Q2 0

107

10⁶







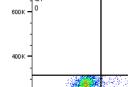


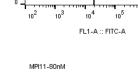


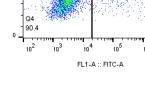


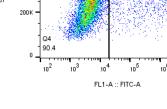
Q4 67.9

MPI11-2uM HEK 8864

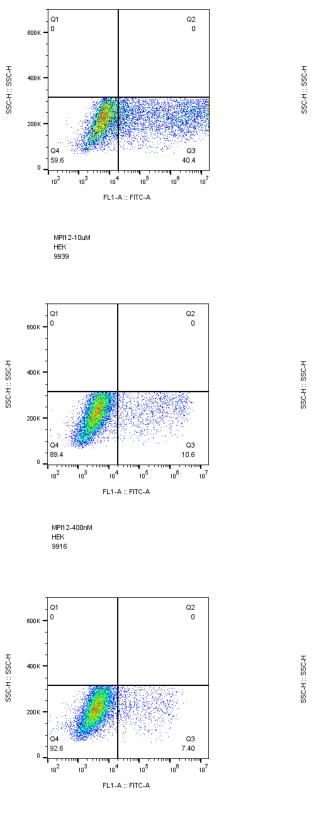




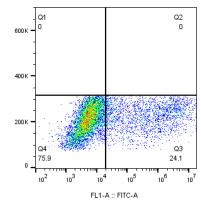




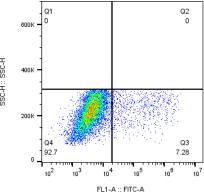
MPI12



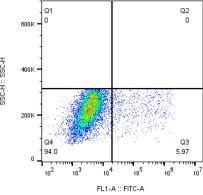




MPI12-2uM HEK 10342

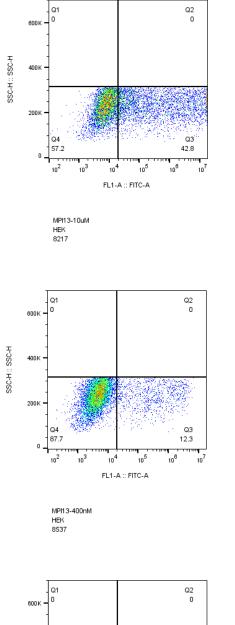


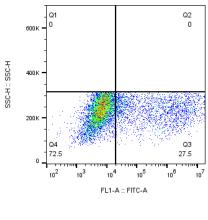


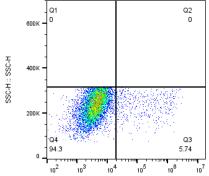


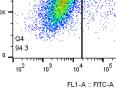


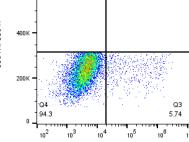
MPI13

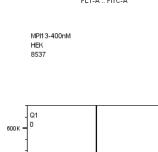












SSC-H :: SSC-H

400 K

200 K

0

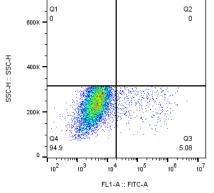
Q4 94.4

10²

10³

MPI13-16nM HEK 8397







MPI13-3.2nM HEK 8191



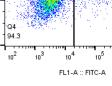












Q1 0

MPI13-2uM HEK 8105

FL1-A :: FITC-A

10⁴

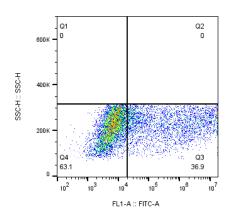
10⁵

Q3 5.57

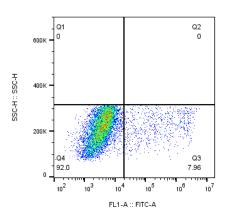
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107

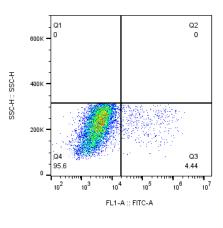
10⁶



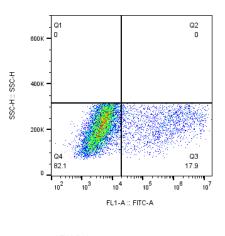




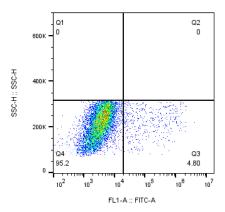




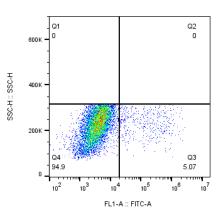
MPI14-16nM HEK 8937



MPI14-2uM HEK 9318



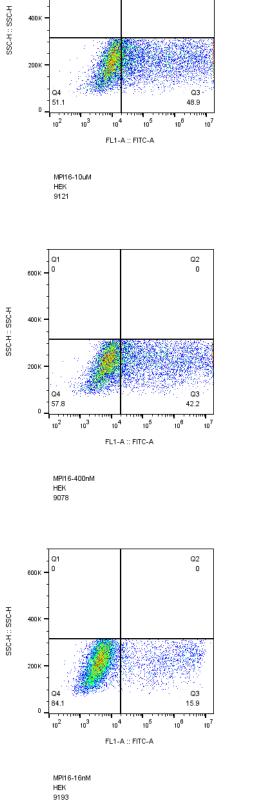


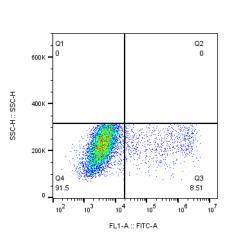




MPI16

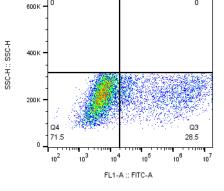
MPI17







MPI16-3.2nM HEK 8670

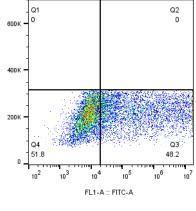


Q2 0



Q1 0

SSC-H :: SSC-H

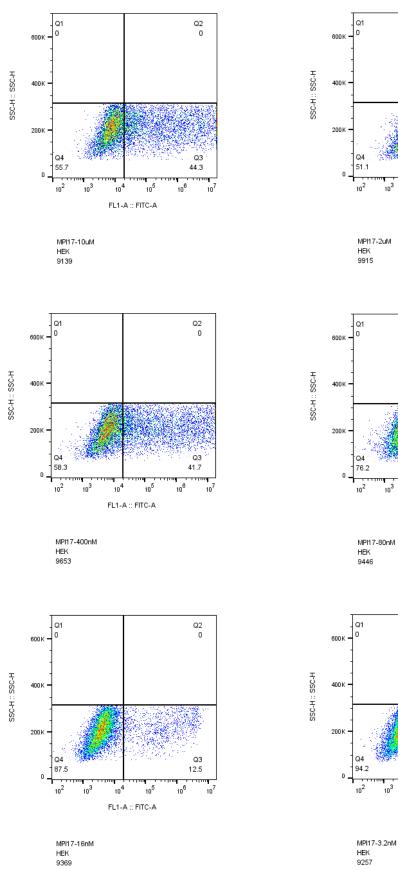


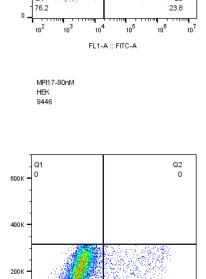
Q1 0

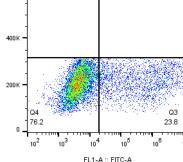
600 K

Q2 0

MPI18







Q2 0

Q3 5.81

10⁶

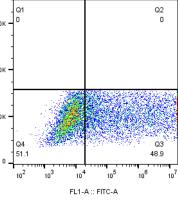
10⁵

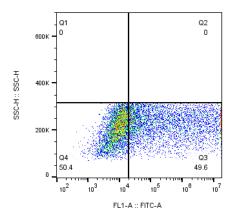
10⁴

FL1-A :: FITC-A

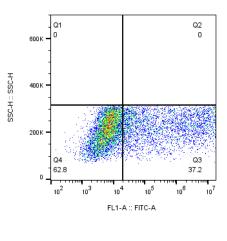
107









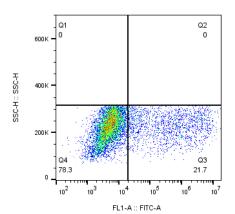




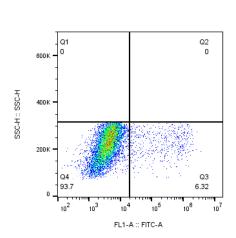
600 K

400 K

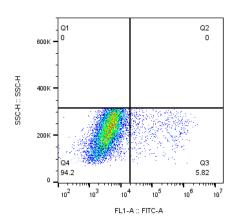
SSC-H :: SSC-H



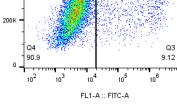




MPI18-80nM HEK 8317

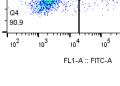






Q2 0

10⁷





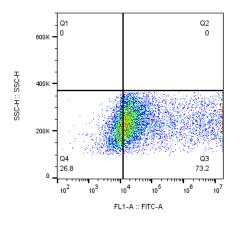
MPI18-3.2nM HEK 8421



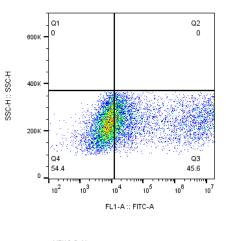




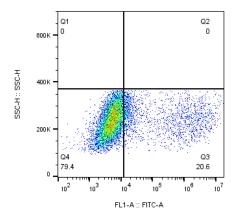




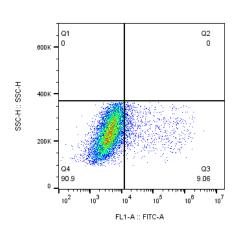




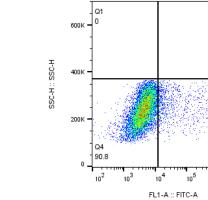


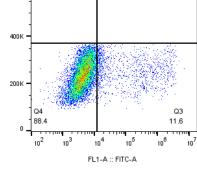


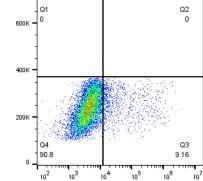




















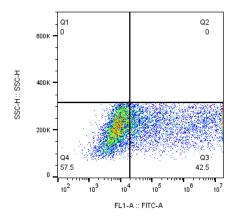


Q1 0 600K

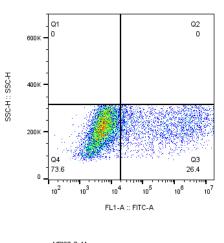
SSC-H :: SSC-H

MPI19-3.2nM HEK 9501

MPI20



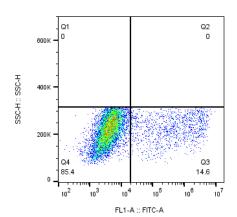




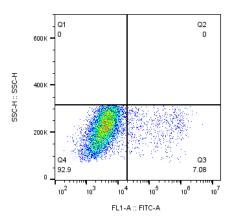
MPI20-2uM HEK 9242

600

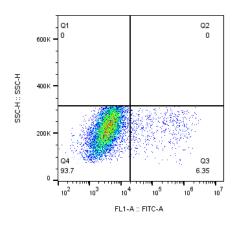
SSC-H :: SSC-H







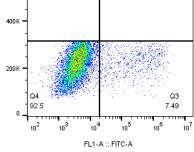
MPI20-16nM HEK 8988





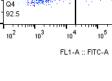






Q2 0







MPI20-3.2nM HEK 8886

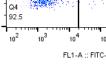




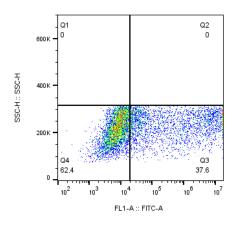




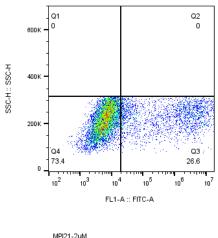




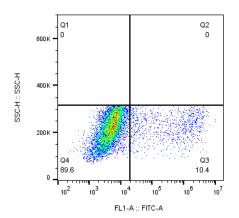




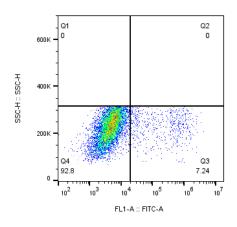




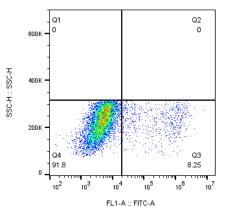




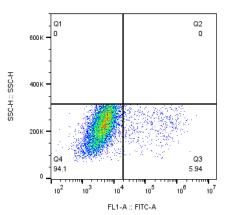




MPI21-16nM HEK 8162

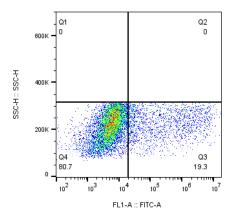




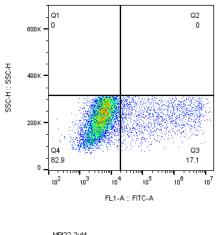






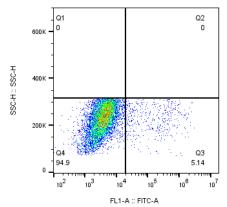


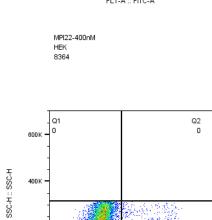






SSC-H :: SSC-H





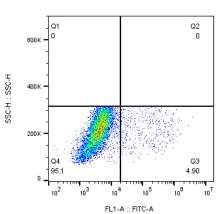
200 K

Q4 95.4 0

10²

10³

MPI22-16nM HEK 8647

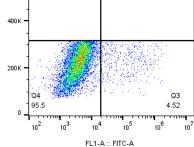




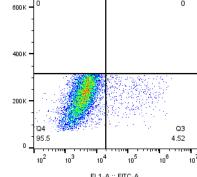


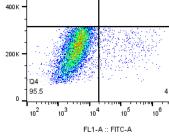






Q2 0

















MPI23

Q3 4.56

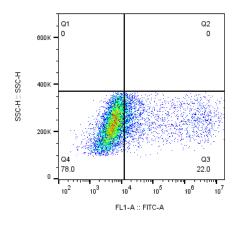
10⁷

10⁶

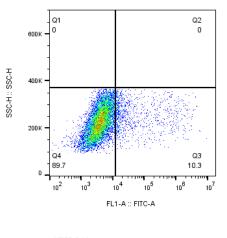
10⁵

10⁴

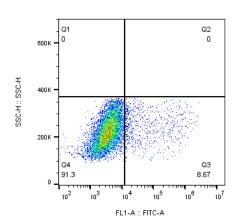
FL1-A :: FITC-A



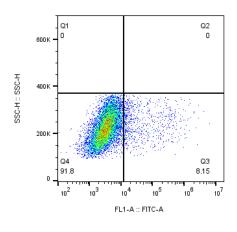




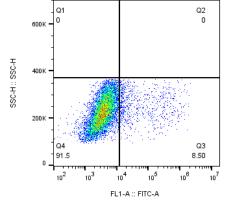
MPI23-2uM HEK 9530



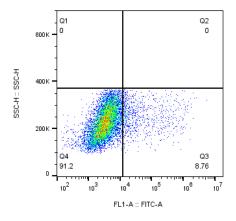




MPI23-16nM HEK 9504



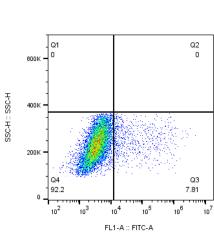


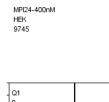


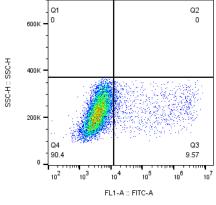
MPI23-3.2nM HEK 9690

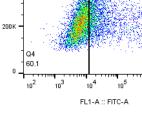












Q3 39.9

10⁶

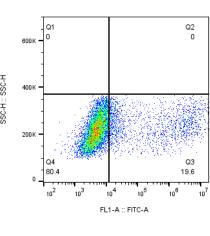
107



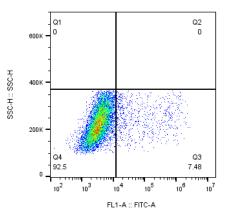
Q1 0 600 K

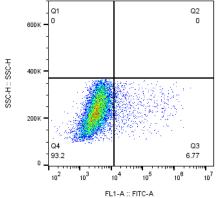
SSC-H :: SSC-H

400 K



MPI24-2uM HEK 9423







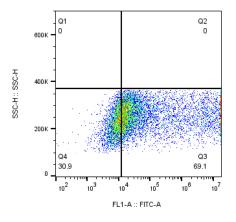
MPI24-3.2nM HEK 9495



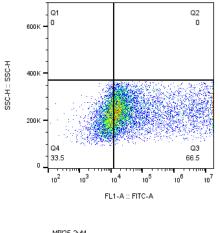


MPI24-16nM HEK 9710

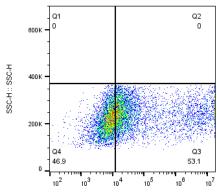
SSC-H :: SSC-H

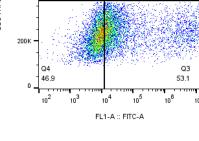






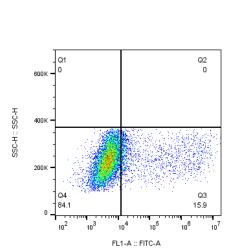
MPI25-2uM HEK 9793



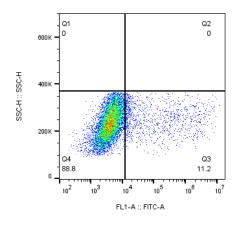




MPI25-16nM HEK 10032







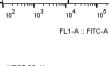


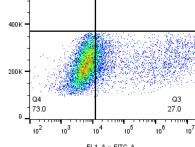


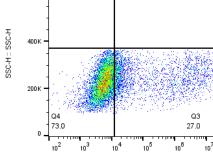


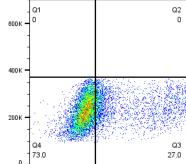
MPI25-3.2nM HEK 9797



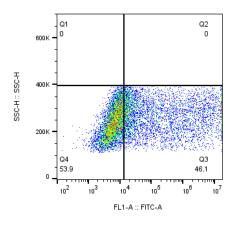




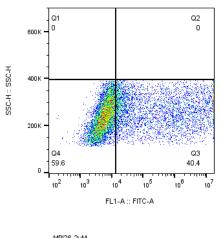




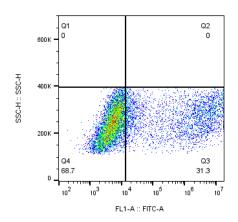




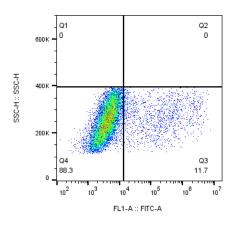




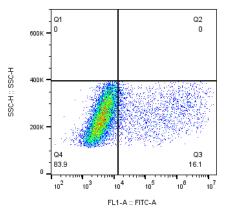
MPI26-2uM HEK 11229



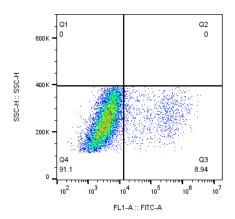




MPI26-16nM HEK 11129





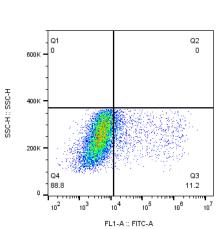


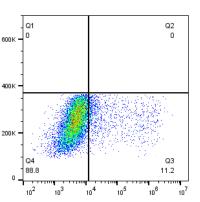






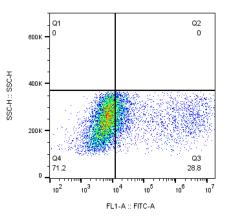








MPI27-16nM HEK 9631





10³

10

FL1-A :: FITC-A

105

Q4

25.4

10²

Q1

0

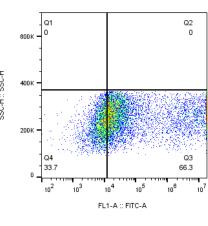
6001

400 K

200 K

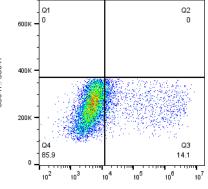
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SSC-H:: SSC-H

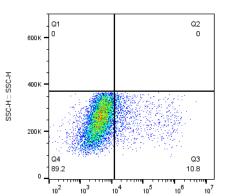










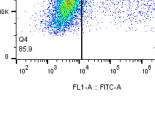


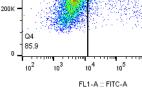
FL1-A :: FITC-A



MPI27-3.2nM HEK 9598

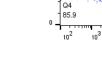


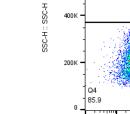












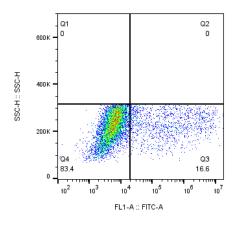


Q2 0

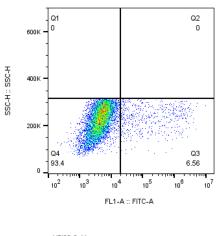
Q3 74.6

10⁶

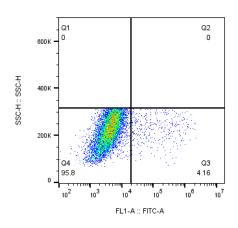
107



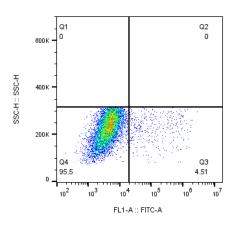




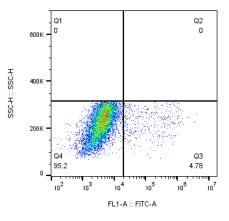
MPI28-2uM HEK 8242



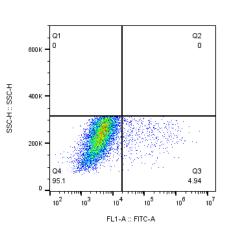




MPI28-16nM HEK 8027







MPI28-3.2nM HEK 8466

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