

Supplemental information

***PDGF Signaling Inhibits Mitophagy in Glioblastoma Stem Cells through N⁶-
methyladenosine***

Deguan Lv, Ryan C. Gimple, Cuiqing Zhong, Qiulian Wu, Kailin Yang, Briana C. Prager, Bhaskar Godugu, Zhixin Qiu, Linjie Zhao, Guoxin Zhang, Deobrat Dixit, Derrick Lee, Jia Shen, Xiqing Li, Qi Xie, Xiuxing Wang, Sameer Agnihotri, Jeremy N. Rich

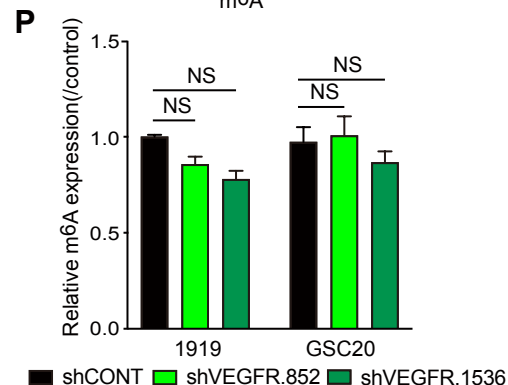
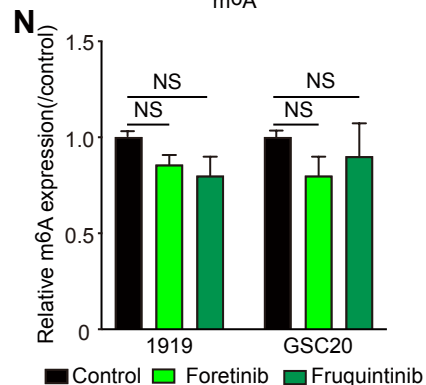
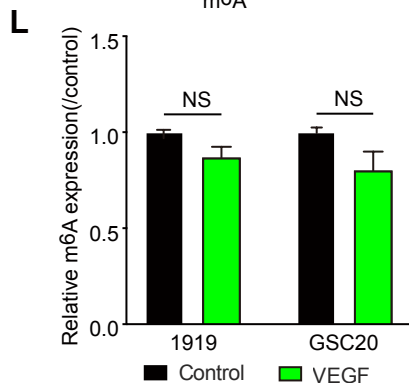
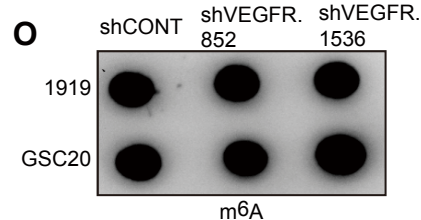
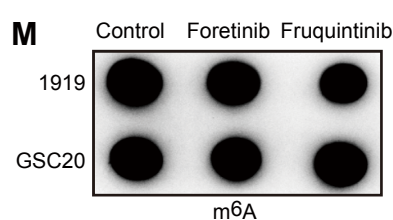
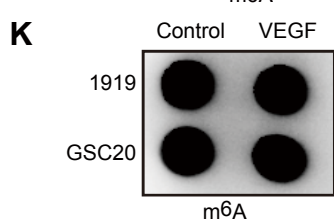
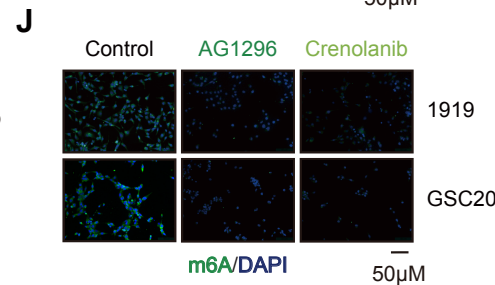
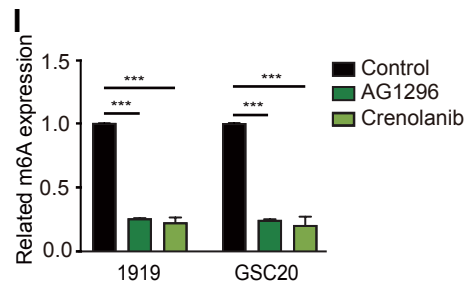
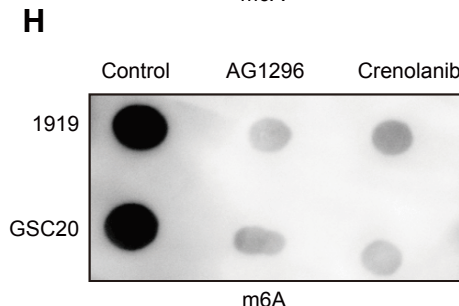
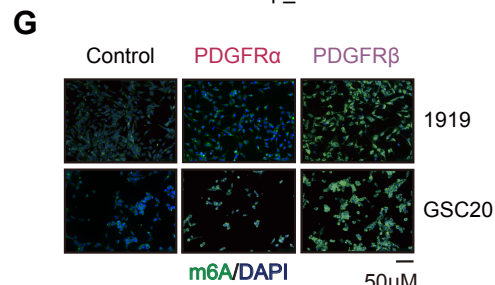
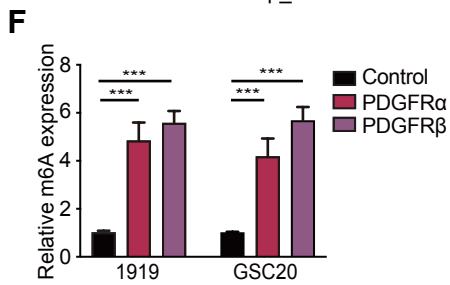
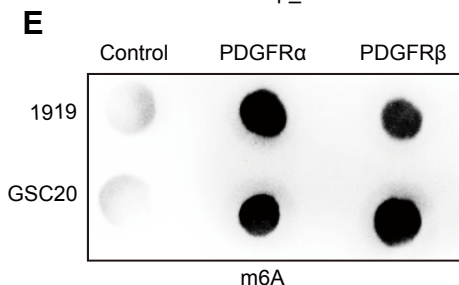
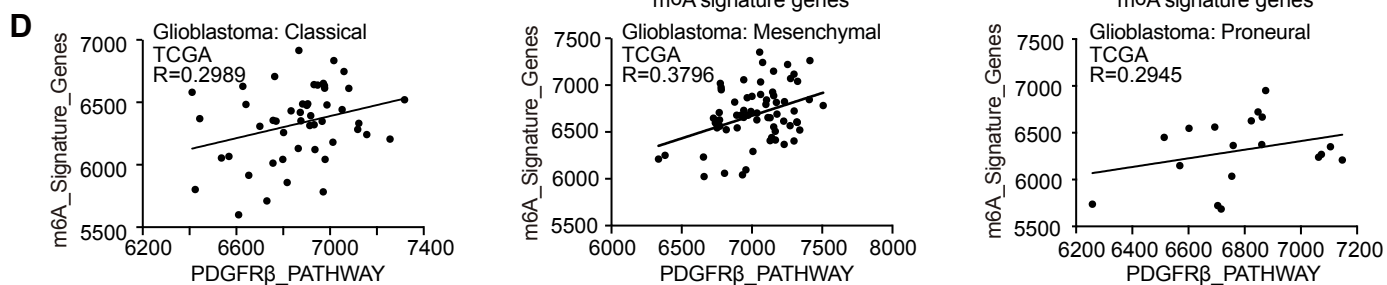
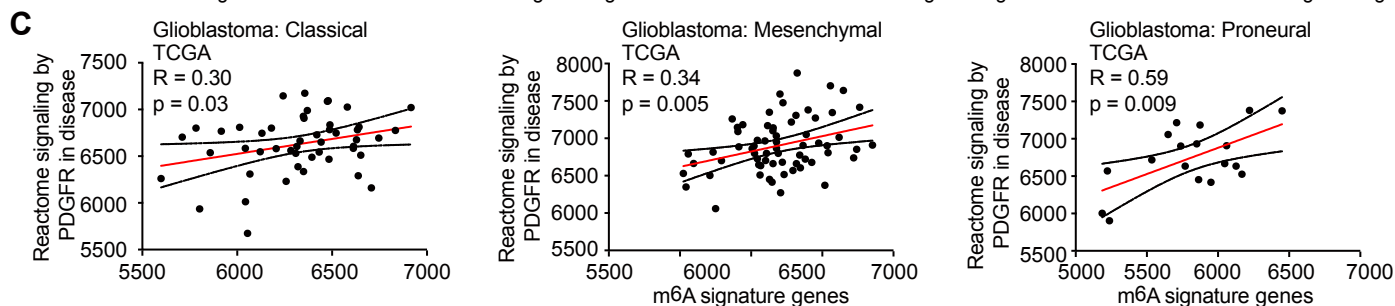
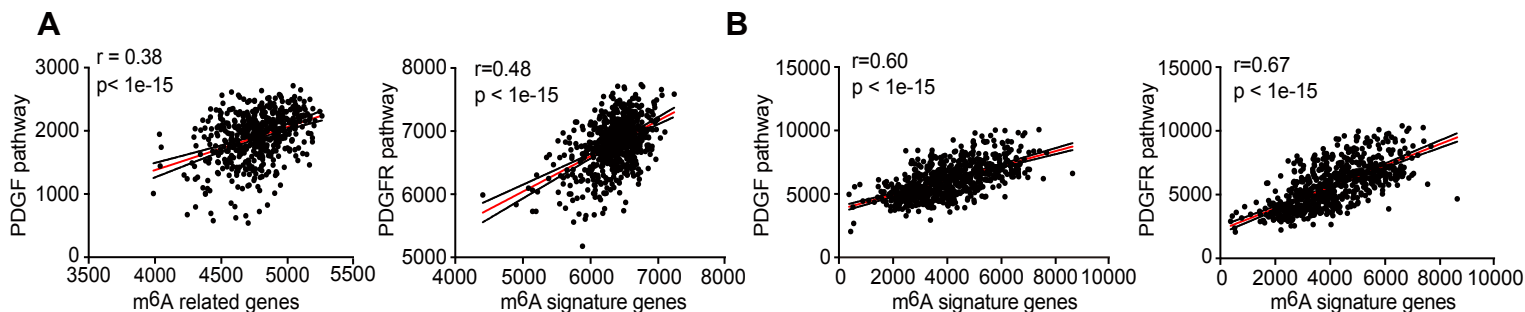


Figure S1. VEGF signaling does not affect m⁶A abundance in GSCs, related to Figure 1

(A) PDGF (left) and PDGFR (right) signaling positively correlates with m⁶A signature in glioblastoma from TCAG dataset. (B) PDGF (left) and PDGFR (right) signaling positively correlates with m⁶A signature in glioblastoma from CGGA dataset. (C) PDGFR signaling positively correlates with m⁶A signature in different glioblastoma subtypes from TCGA dataset. (D) Correlative analysis was performed on the TCGA glioblastoma dataset with PDGFRB mRNA levels and an m⁶A regulator signature in different glioblastoma subtypes. (E-G) PDGFR overexpression increases global m⁶A levels in GSC. Verification of the m⁶A abundance in GSC RNA by dot blot (E), colorimetric estimation (F) and immunofluorescence (G). (H-J) PDGFR inhibitors treatment decreases global m⁶A levels in GSC. Verification of the m⁶A abundance in GSC RNA by dot blot (H), colorimetric estimation (I) and immunofluorescence (J). (K and L) VEGF treatment has no effects on RNA global m⁶A levels in GSC. Verification of the m⁶A abundance in GSC RNA by dot blot (K), colorimetric estimation (L). (M and N) Global m⁶A levels of RNA assay in GSC after VEGFR inhibitors treatment. Verification of the m⁶A abundance in GSC RNA by dot blot (M), colorimetric estimation (N). (O and P) Global m⁶A levels of RNA assay in GSC with VEGFR2 knockdown (shVEGFR2.852, shVEGFR2.1536). Verification of the m⁶A abundance in GSC RNA by dot blot (O), colorimetric estimation (P).

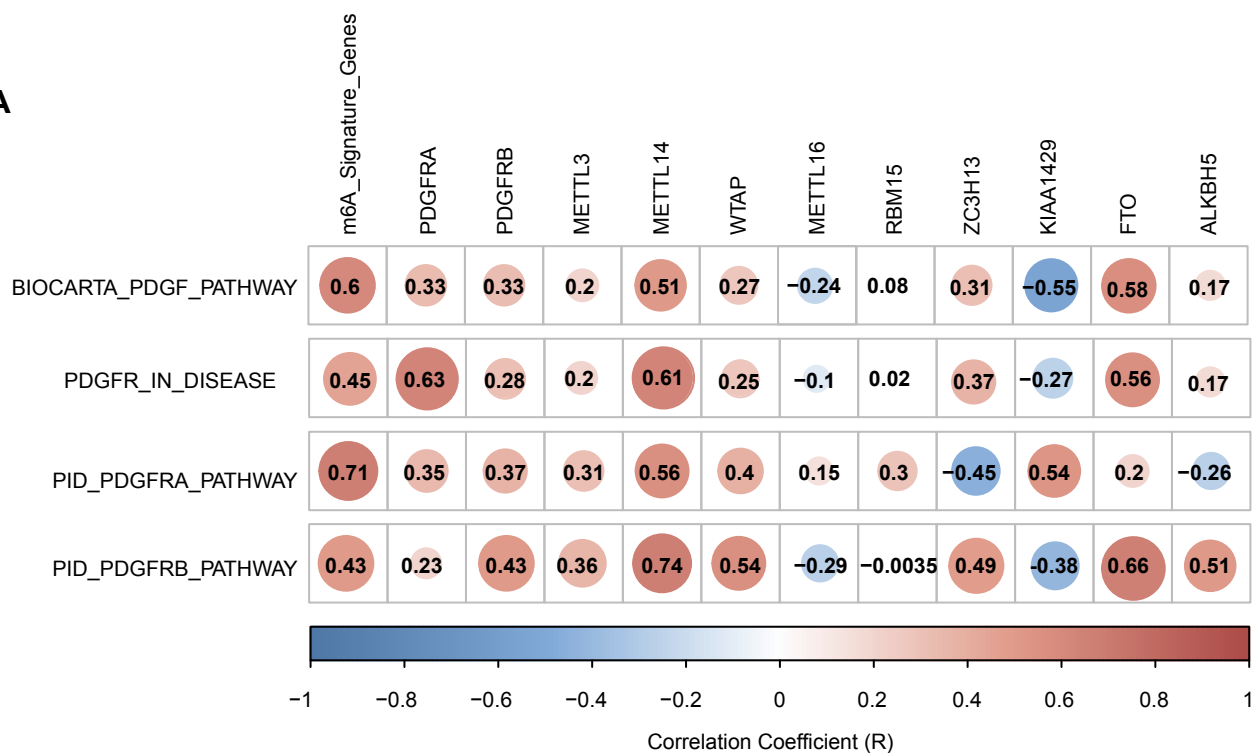
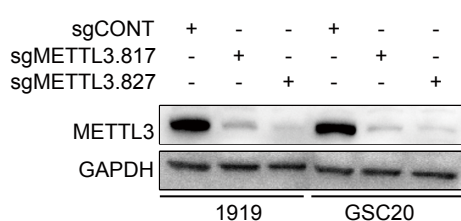
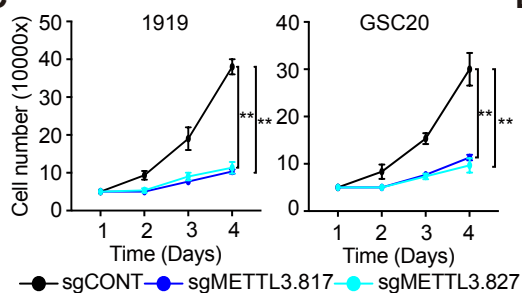
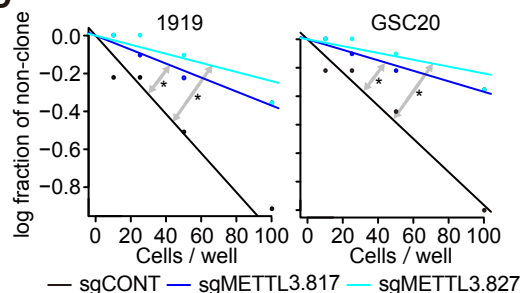
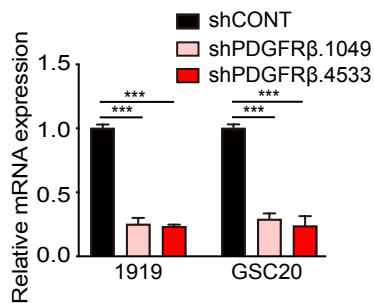
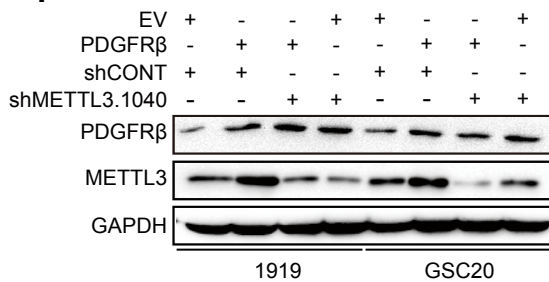
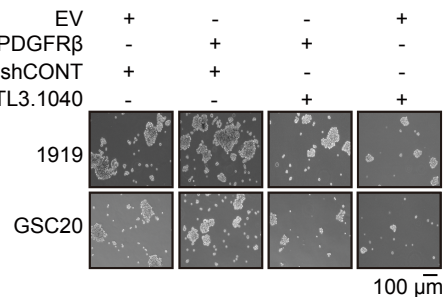
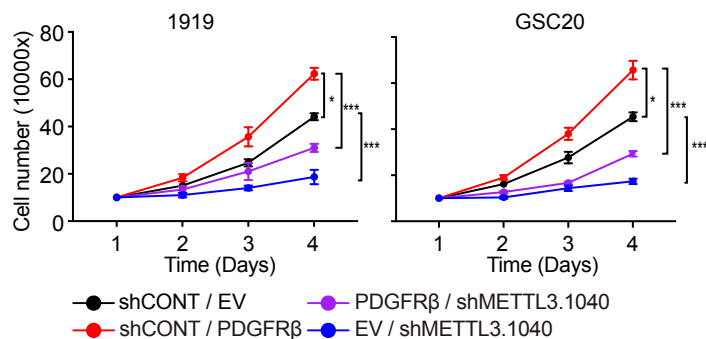
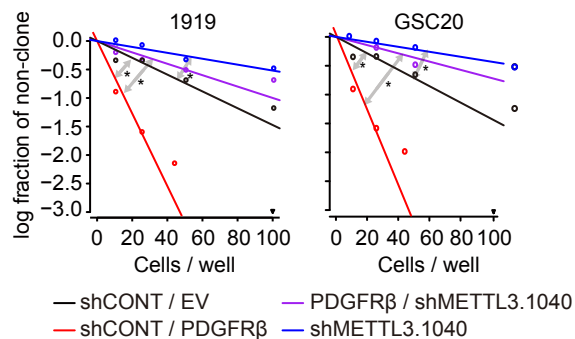
A**B****C****D****E****F****G****H****I**

Figure S2. METTL3 knockout impairs PDGF-induced GSCs proliferation and clone formation, related to Figure 2

(A) Correlations between PDGF/PDGFR signatures and various m⁶A signature genes in CGGA datasets. (B) WB assays of METTL3 protein levels in 1919 and GSC20 cells following METTL3 knockout with two different sgMETTL3 (sgMETTL3.817, sgMETTL3.827) or a non-targeting sgRNA (sgCONT). (C) Cell viability of METTL3 knockout GSCs 1919 (left) and GSC20 (right), up to 3 days of post-transduction. **, $p < 0.01$. Error bars show standard deviation. (D) *In vitro* limiting dilution assay in 1919 (left) and GSC20 (right) following knockout with two independent sgRNAs targeting METTL3 or a non-targeting sgRNA (sgCONT). 10 wells were quantified for each condition and 3 biologic replicates for the assay. *, $p < 0.05$. (E) qPCR analysis of METTL3 mRNA expression following knockdown with two independent, non-overlapping shRNAs targeting PDGFR β (shPDGFR β .2164 and shPDGFR β .2682) or a non-targeting shRNA (shCONT). Three technical replicates were used for each condition. Statistical significance was assessed using a two-way ANOVA with Sidak multiple test correction, ***, $p < 0.001$. (F) WB assays of METTL3 knockdown in 1919 and GSC20 cells with, or without PDGFR β overexpression. (G) Spheres formation of 1919 and GSC20 following transduction with METTL3 knockdown (shMETTL3.915 and shMETTL3.1040) with or without PDGFR β overexpression. (H) Cell viability of METTL3 knockdown GSCs, in the presence or absence of PDGFR β overexpression, up to 3 days of post-transduction. Error bars show standard deviation. *, $p < 0.05$, ***, $p < 0.001$. (H) *In vitro* limiting dilution assay in 1919

and GSC20 following knockdown with two independent, non-overlapping shRNAs targeting METTL3 or a non-targeting shRNA (shCONT) with or without PDGFR β overexpression. 10 wells were quantified for each condition and 3 biologic replicates for the assay. *, $p < 0.05$.

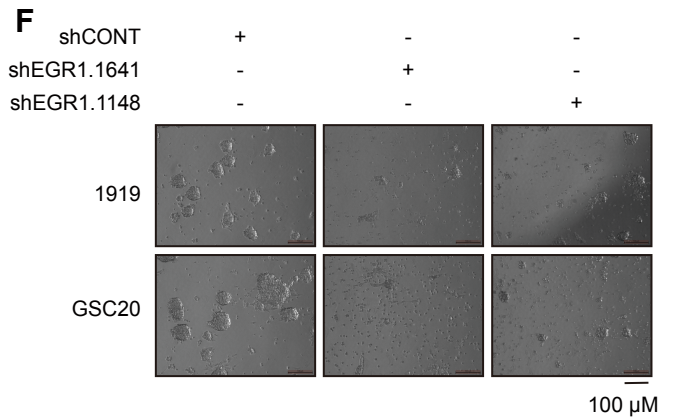
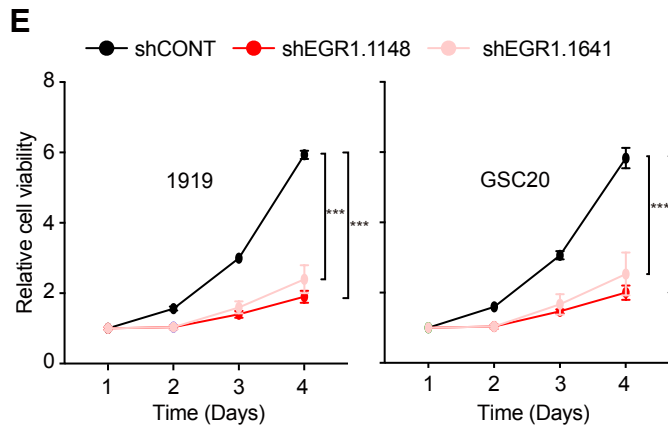
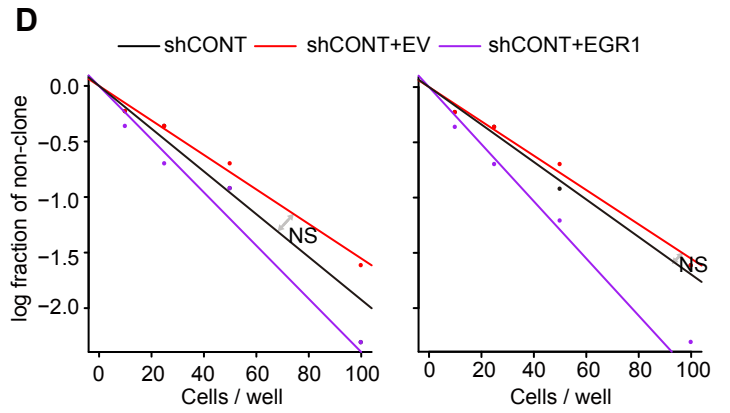
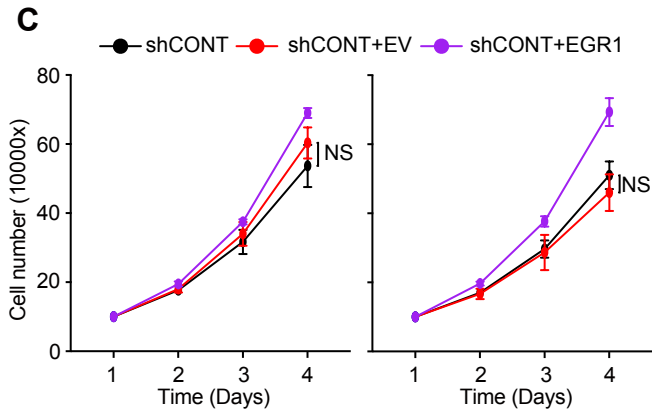
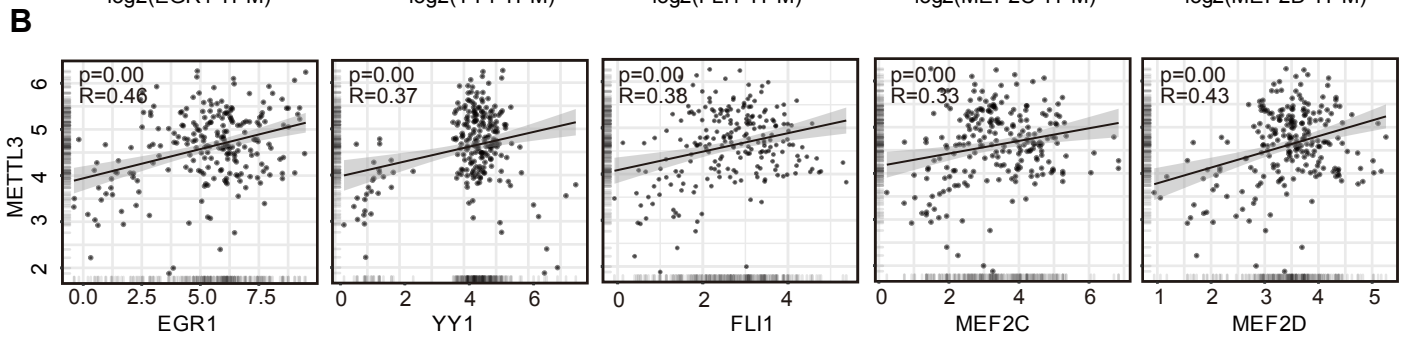
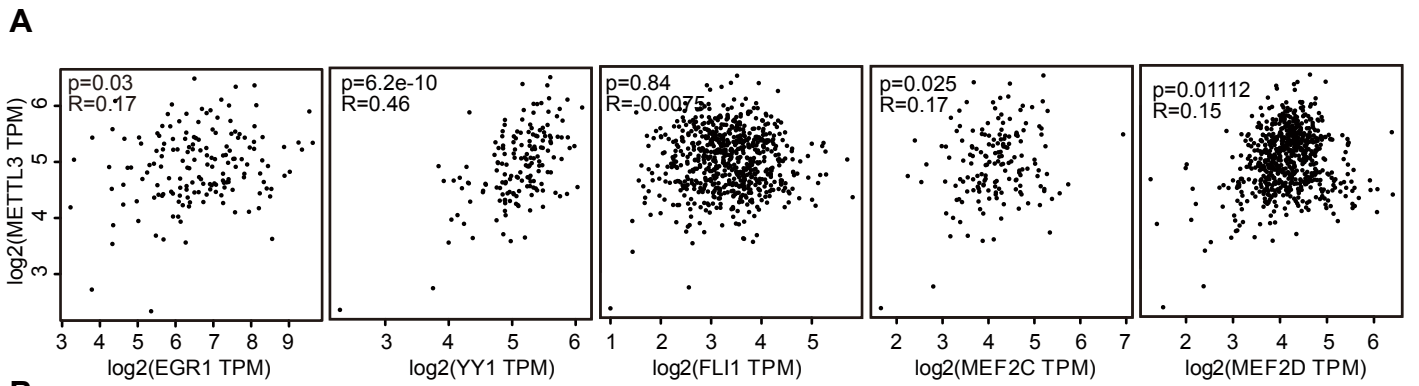


Figure S3. EGR1 is an oncogene in glioblastoma, related to Figure 3

(A and B) The correlation analysis of TFs and METTL3 with TCGA dataset (A) and CGGA dataset (B). (C) Cell viability of EV or EGR1-transduced GSCs 1919 and GSC20 in shCONT. NS, no statistical significance. Error bars show standard deviation. (D) In vitro limiting dilution assay in 1919 and GSC20 following overexpressed EV or EGR1 with non-targeting shRNA (shCONT) knockdown. 10 wells were quantified for each condition and 3 biologic replicates for the assay. NS, not statistically significant. (E) Cell viability in the 1919 (left) and GSC20 (right) cell models over a 3-day time course following knockdown with two independent, non-overlapping shRNAs targeting EGR1 (shEGR1.1641 and shEGR1.1148) or a non-targeting shRNA (shCONT). Three technical replicates were performed for each group. One-way ANOVA was used for statistical analysis with Dunnett's multiple hypothesis test correction with three technical replicates. ***, $p < 0.001$. (F) Functional assays of self-renewal (serial neurosphere passage), and tumor propagation using in vitro limiting dilution after EGR1 knockdown with two independent, non-overlapping shRNAs targeting EGR1 (shEGR1.1641 and shEGR1.1148) or a non-targeting shRNA (shCONT). Scale bar: 100 μ M.

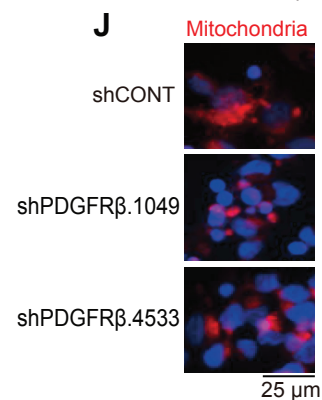
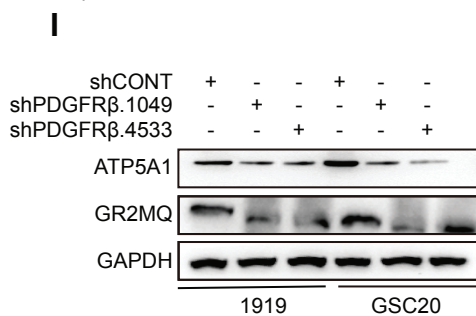
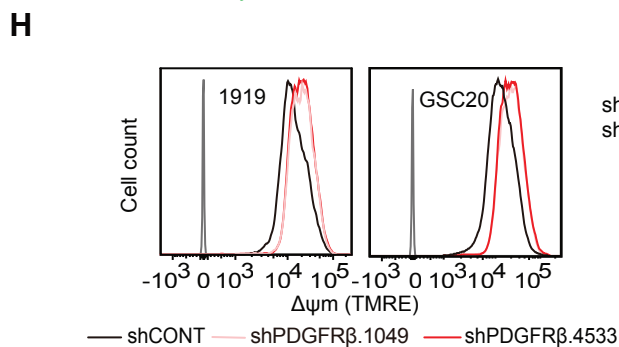
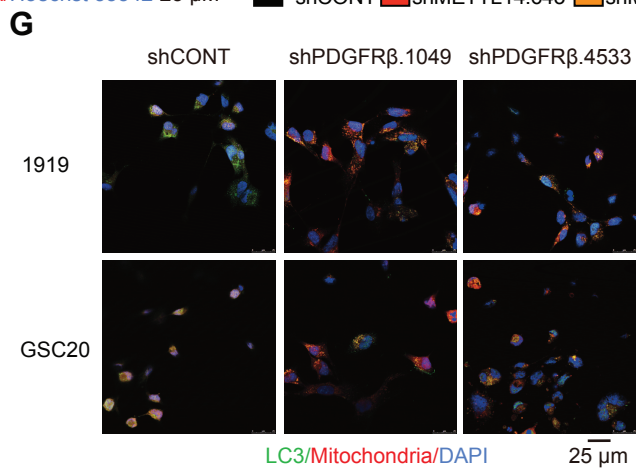
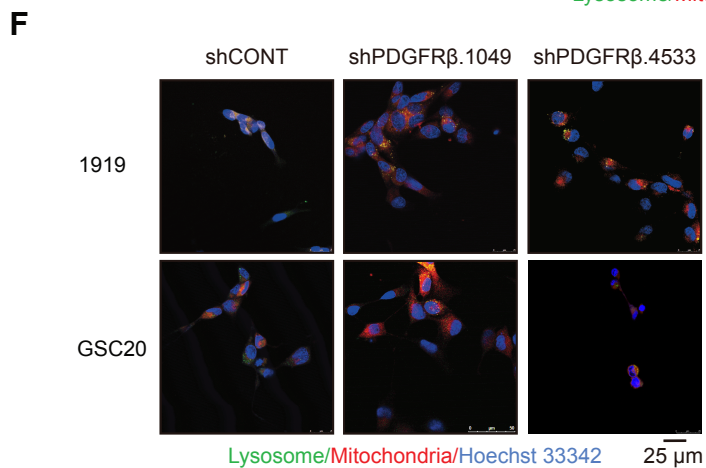
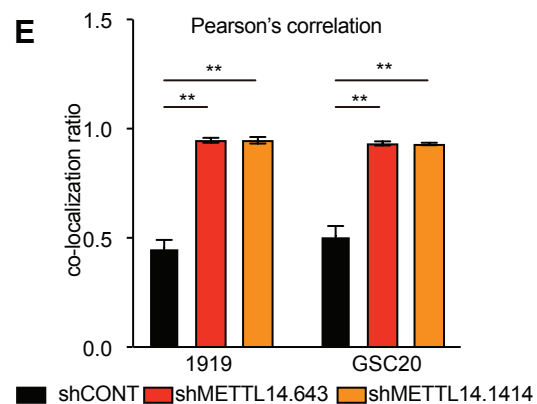
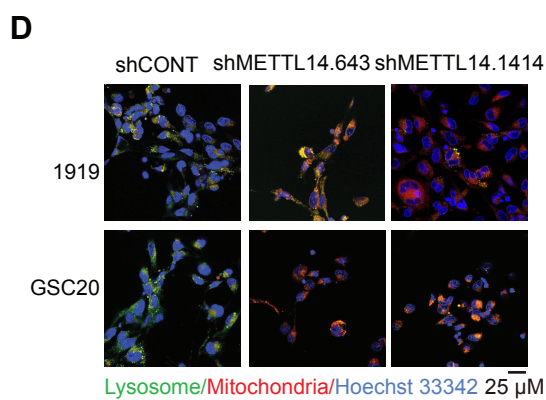
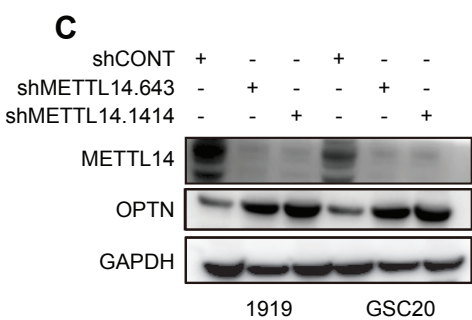
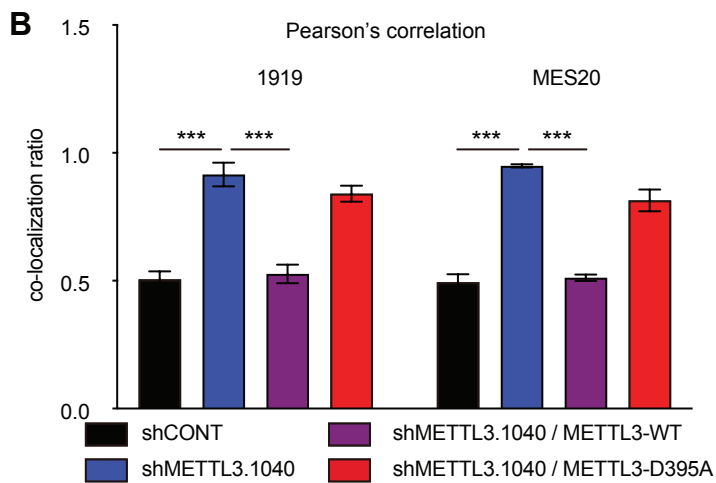
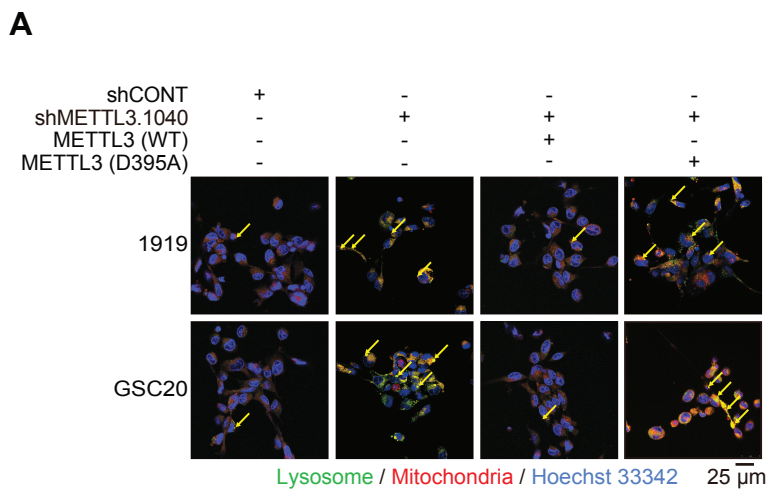


Figure S4. PDGFR β affects mitochondrial function via mitophagy, related to Figure 4

(A) Cellular colocalization of lysosome and mitochondria in 1919 (above) and GSC20 (below) after shRNA-resistant METTL3 (WT) or (D395A) overexpression with, or without METTL3 knockdown (shMETTL3.1040) by immunofluorescence staining using lysosomal and mitochondrial probe. Yellow arrows designate mitochondria that are being degraded. Scale bar: 25 μ m. (B) Pearson's correlation coefficient is shown in bar graph format from three independent experiments in METTL3 rescue GSCs. Error bars show standard deviation. ***, $p < 0.001$. (C) Two patient-derived GSCs, 1919 and GSC20, were transduced with either one of two non-overlapping shRNAs targeting METTL14 or shCONT. WB assays of METTL14 and OPTN protein levels were performed. (D) Cellular colocalization of lysosome and mitochondria in 1919 (top) and GSC20 (bottom) after METTL14 knockdown, as demonstrated by immunofluorescence staining using lysosomal and mitochondrial probes. Scale bar: 25 μ m. (E) Pearson's correlation coefficient is shown in bar graph format for (D). Error bars represents the standard deviation. **, $p < 0.01$. (F) Cellular colocalization of lysosome and mitochondria in 1919 (left) and GSC20 (right) as demonstrated by immunofluorescence staining using lysosomal and mitochondrial probe. Scale bar: 25 μ m. (G) Cellular colocalization and levels of LC3 and mitochondria in 1919 (left) and GSC20 (right) as demonstrated by immunofluorescence staining using LC3 antibody and mitochondrial probe. Scale bar: 25 μ m. (H) PDGFR β knockdown with two independent, non-overlapping shRNAs (shPDGFR β .1049 and shPDGFR β .4533) increases 1919 (left) and GSC20 (right) cell mitochondrial

membrane potential by flow cytometry analysis. (I) METTL3 knockdown with two independent, non-overlapping shRNAs (shPDGFR β .1049 and shPDGFR β .4533) decreases number of mitochondria in GSC1919. (J) PDGFR β knockdown with two independent, non-overlapping shRNAs (shPDGFR β .1049 and shPDGFR β .4533) decreases number of mitochondria in GSC1919.

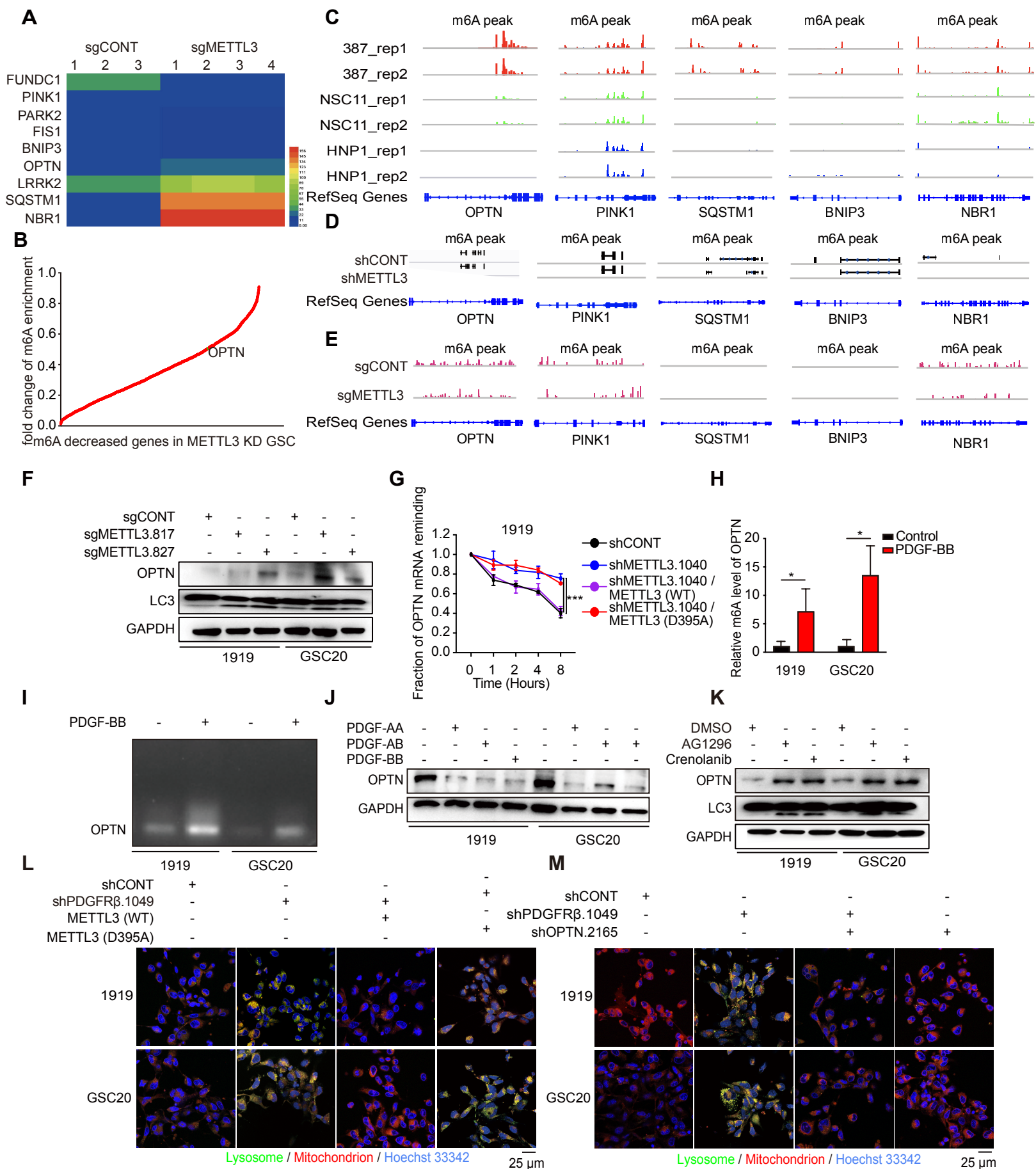


Figure S5. PDGF-METTL3 inhibits mitophagy via regulating OPTN m⁶A modification in GSCs, related to Figure 5

(A) Heatmap of mitophagy related genes expression in METTL3 knockout cells. (B) m⁶A abundance assay in GSC after METTL3 knockout. (C) m⁶A signal of OPTN, PINK1, SQSTM1, BNIP3, and NBR1 in GSC and NSCs at the gene locus. (D and E) m⁶A signal in OPTN locus after METTL3 knockout (D) and knockdown (E). (F) WB assays of OPTN and LC3 level in GSC with METTL3 knockout. (G) METTL3 promotes OPTN mRNA decay in GSCs. qPCR analysis of mRNA expression in 1919 after METTL3-WT or METTL3-mutant overexpression in METTL3 knockdown cell line treated with actinomycin D. Student's t-test with Holm-Sidak multiple test correction. ***, $p < 0.001$. (H) Verification of the m⁶A abundance in 1919 and GSC20 RNA with PDGFR β knockdown by MeRIP-qPCR. ***, $p < 0.001$. (I) Agarose gel image of MeRIP-qPCR result from H. (J) WB assays of OPTN expression in 1919 and GSC20 after PDGF treatment. (K) WB assays of LC3 and OPTN expression in 1919 and GSC20 after PDGFR inhibitors treatment. (L) Cellular colocalization of mitochondrion and lysosome in METTL3 WT or D395A rescued 1919 and GSC20 cells. Scale bar: 25 μ m. (M) Cellular colocalization of lysosome and mitochondria in 1919 and GSC20 with PDGF β and OPTN knockdown as demonstrated by immunofluorescence staining using lysosomal and mitochondrial probe. Scale bar: 25 μ m.

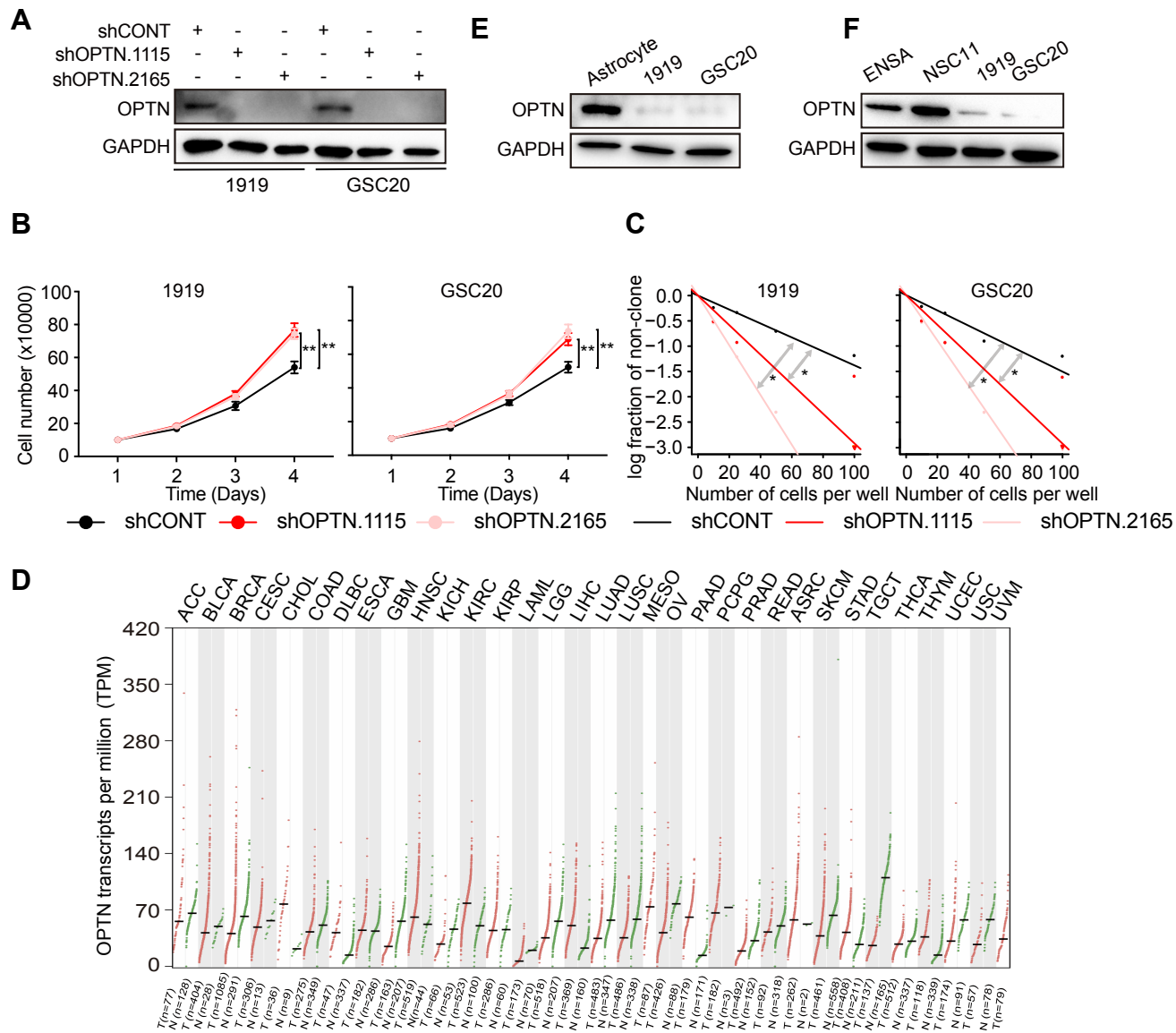


Figure S6. OPTN is a novel suppressor in glioblastoma, related to Figure 6

(A) Cell viability of METTL3 knockdown GSCs, up to 3 days of post-transduction. Error bars show standard deviation. *, $p < 0.05$. (B) In vitro limiting dilution assay in 1919 and GSC20 following knockdown with two independent, non-overlapping shRNAs targeting OPTN or a non-targeting shRNA (shCONT). 10 wells were quantified for each condition and 3 biologic replicates for the assay. *, $p < 0.05$. (C) WB assays of OPTN expression following knockdown with two independent, non-overlapping shRNAs targeting OPTN (shOPTN.1115 and shOPTN.2165) or a non-targeting shRNA (shCONT). (D) Landscape of OPTN expression in pan-cancers. (E) WB assays of OPTN expression in astrocyte and GSCs. (F) WB assays of OPTN expression in NSCs (ENSA and NSC11) and GSCs.

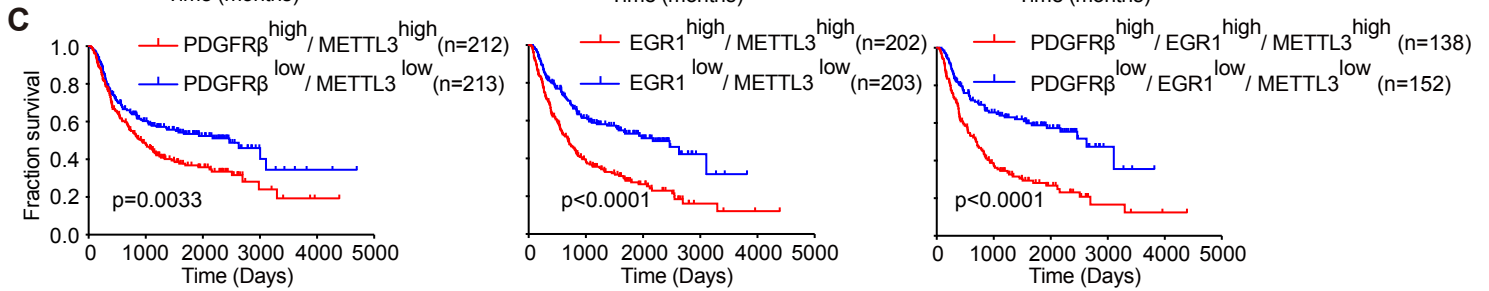
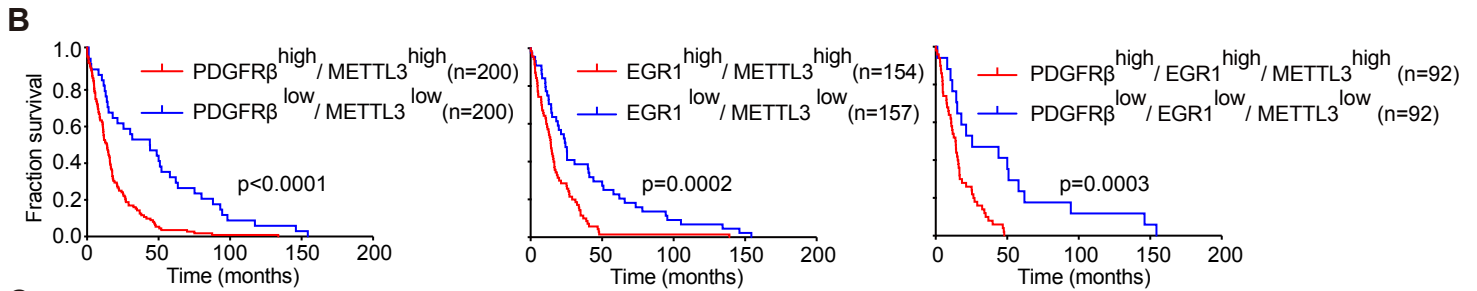
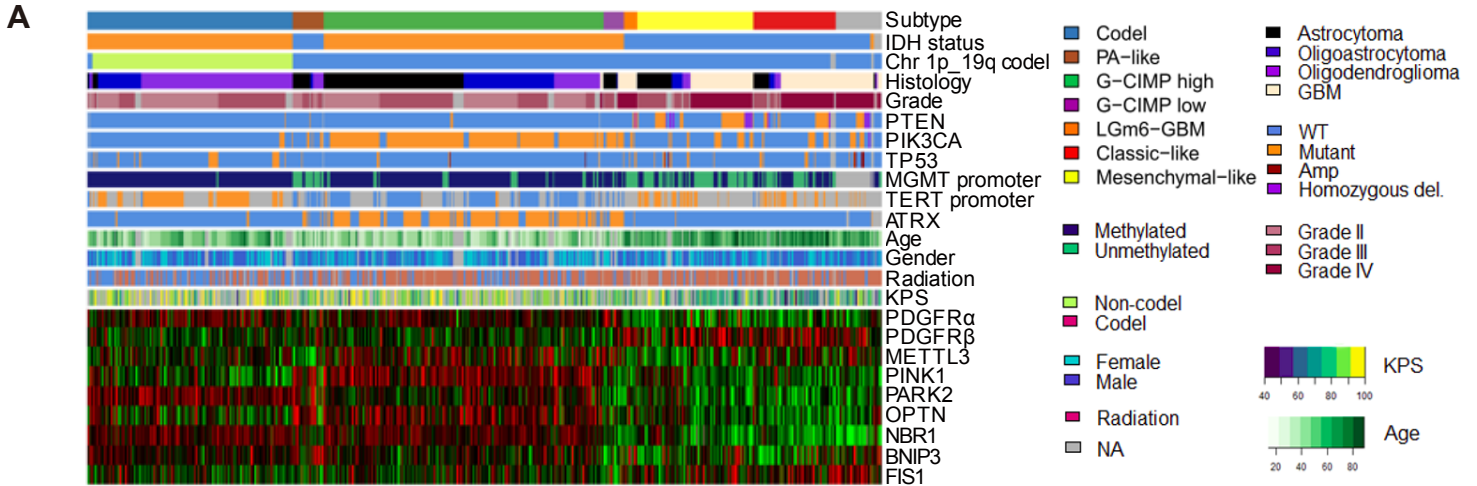


Figure S7. PDGF-EGR1-METTL3 axis is clinically prognostic, related to Figure 7

(A) Heatmaps of TCGA glioma samples displaying mRNA levels of PDGFR β -METTL3 along with clinical and genetic variant information for each sample. (B) Kaplan-Meier curve showing patient survival based on PDGFR β , EGR1 and METTL3 mRNA expression in glioma patients from the TCGA dataset. Log-rank test was used for statistical analysis. PDGFR β^{high} METTL3 $^{\text{high}}$ vs. PDGFR β^{low} METTL3 $^{\text{low}}$ ($p < 0.0001$), EGR1 $^{\text{high}}$ METTL3 $^{\text{high}}$ vs. EGR1 $^{\text{low}}$ METTL3 $^{\text{low}}$ ($p = 0.0002$), PDGFR β^{high} EGR1 $^{\text{high}}$ METTL3 $^{\text{high}}$ vs. PDGFR β^{low} EGR1 $^{\text{low}}$ METTL3 $^{\text{low}}$ ($p = 0.0003$). (C) Kaplan-Meier curve showing patient survival based on PDGFR β , EGR1 and METTL3 mRNA expression in glioma patients from the CGGA dataset. Log-rank test was used for statistical analysis. PDGFR β^{high} METTL3 $^{\text{high}}$ vs. PDGFR β^{low} METTL3 $^{\text{low}}$ ($p = 0.0033$), EGR1 $^{\text{high}}$ METTL3 $^{\text{high}}$ vs. EGR1 $^{\text{low}}$ METTL3 $^{\text{low}}$ ($p < 0.0001$), PDGFR β^{high} EGR1 $^{\text{high}}$ METTL3 $^{\text{high}}$ vs. PDGFR β^{low} EGR1 $^{\text{low}}$ METTL3 $^{\text{low}}$ ($p < 0.0001$). (D) The correlation analysis of PDGFR and METTL3, METTL3 and OPTN in breast invasive carcinoma, lung adenocarcinoma, colon adenocarcinoma, prostate adenocarcinoma, liver hepatocellular carcinoma, stomach adenocarcinoma, and glioblastoma with TCGA dataset. (E and F) WB assays of METTL3 and OPTN expression in A549, MD231, and HepG2 cells with or without PDGF and PDGFR inhibitors treatment. (G) WB assays of PDGFR β expression in A549, MD231, and HepG2 cells. (H) WB assays of METTL3 expression in GSC and DGC pairs derived from 1919 and GSC20 xenografts. (I) RNA m⁶A methylation levels measured by colorimetric assay from RNA samples from 1919 and GSC20 GSCs and the differentiated glioma cells. (J) GSC 1919 and GSC20

differentiation decreases OPTN m⁶A levels. Verification of the m⁶A abundance in 1919 and GSC20 RNA by MeRIP-qPCR.