Supplemental Material
Implication of the IL-10-expression signature in the pathogenicity of Leptospira-infected
macrophages
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- 21 Running Head: IL-10 expression in *Leptospiral*-infected macrophages
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26 This article contains the following supplemental material

## 27 FIG S1. Biological characterization of *Leptospira*-infected bone marrow-derived macrophages

28 (BMDMs). (A) Changes in the density and shape of mitochondria in *Leptospira*-infected BMDMs. Representative transmission electron microscopy images of BMDMs infected with L. biflexa (left) or 29 30 L. interrogans (right) for 2 h. Magnification: (upper row) 27,000x, scale bar: 2.0 µm; (lower row) 31 110,000x, scale bar: 1.0 µm, area delineated by a rectangle in the top row. Black arrow and M indicate 32 mitochondria; N-Nucleus; LBP-L. biflexa; LIC- L. interrogans. (B) Transmission electron 33 microscopy images of intracellular leptospiral localization patterns in infected macrophages. Images 34 show L. biflexa-infected BMDMs at 2 h (upper row) and (lower row) 24 h post-infection (scale bar, 35 1.0 µm), and L. interrogans-infected BMDMs at 2 h (upper low) and (lower row) 24 h post-infection 36 (scale bar, 500 nm and 1.0 µm, respectively). Black arrowheads show L. biflexa, arrows show L. 37 interrogans, and M\* and V indicate damaged mitochondria and vacuoles, respectively. (C) Amount 38 of L. biflexa in infected BMDMs was larger than the amount of L. interrogans. Quantification of the 39 total fluorescent area (upper row) and fluorescence intensity (lower row) in an infected cell population 40 at the single-cell level. An algorithm for measuring the total fluorescent area and assessing the total 41 cell number on an image was developed in Python 3.10.0 using OpenCV (Open-Source Computer 42 Vision, Intel Corp.). The total fluorescence intensity was measured using MetaMorph software. Cells 43 within 4 images (approximately 80 cells) were analyzed. P: L. biflexa-infected BMDMs, and C: L. 44 interrogans-infected BMDMs. BMDM, bone marrow-derived macrophage. (D) Induction of 45 apoptosis in BMDMs infected with *Leptospira* spp. After infection with leptospires for (upper row) 2 h and (lower row) 24 h, the percentage of apoptotic cells was detected using a fluorescein 46 47 isothiocyanate (FITC)-annexin V/propidium iodide (PI) double staining method and analyzed using 48 flow cytometry. Non-apoptotic cells (lower left quadrant); early apoptotic cells (lower right quadrant); 49 late apoptotic/necrotic cells (upper right quadrant).

50 FIG S2. Macrophage responses after 24 h of infection with pathogenic Leptospira spp. Plots obtained from gene set enrichment analysis of "HALLMARK UV RESPONSE UP" and 51 52 "HALLMARK PI3K AKT MTOR SIGNALING" are shown in L. interrogans-infected BMDMs 53 vs. uninfected BMDM groups but not in L. biflexa-infected BMDMs vs. uninfected BMDM groups. 54 The green curve refers to calculation of the enrichment score. GSEA, gene set enrichment analysis; NES, normalized enrichment score; FDR, false discovery rate. Left panels show GSEA enrichment 55 56 plots (score curves) and the heatmap on the right side of each panel shows the core enrichment genes 57 (positively enriched associated molecules) contributing the most to the enriched pathway. Increased 58 expression (red) and decreased expression (blue). C24: L. interrogans-infected BMDMs at 24 h post-59 infection and E24: uninfected BMDMs at 24 h. BMDM, bone marrow-derived macrophage.

60 FIG S3. Representative flow cytometry plot showing the gating strategy to identify classically activated M1 or alternatively activated M2 macrophage subsets in leptospiral-infected BMDMs. 61 62 The cells were first morphologically gated using the forward light scatter area (FSC-A) and side light 63 scatter area (SSC-A) to exclude cell debris. Viability gating was performed to select cells that were 64 negative for the eBioscience fixable viability dye stain (Thermo Fisher Scientific, 65-0865-14), and 65 cell doublets were excluded based on the FSC-A and FSH-height (FSH-H). Live cells were gated on 66 CD11b<sup>+</sup>F4/80<sup>+</sup> cells (R1). Next, CD11b<sup>+</sup>F4/80<sup>+</sup> cells were analyzed for the expression of CD80 and CD206: R2: CD11b+F4/80+CD80+ (classically activated M1) and R3: CD11b+F4/80+CD260+ 67 68 (alternatively activated M2).

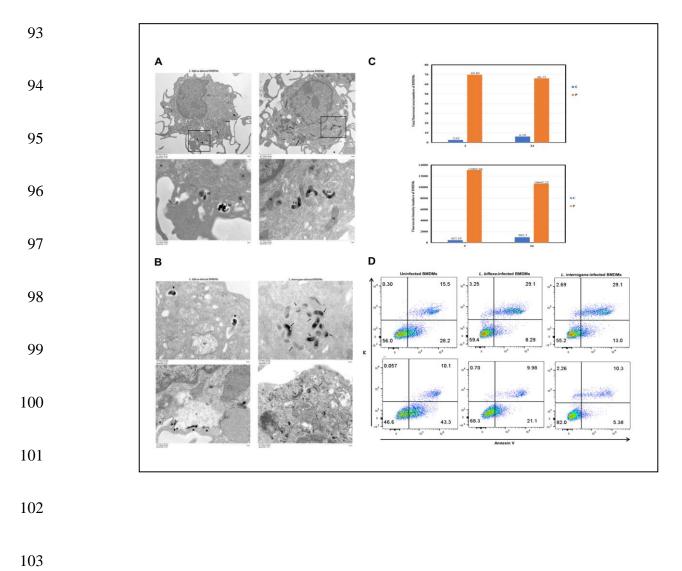
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73	Tables	and	their	legends
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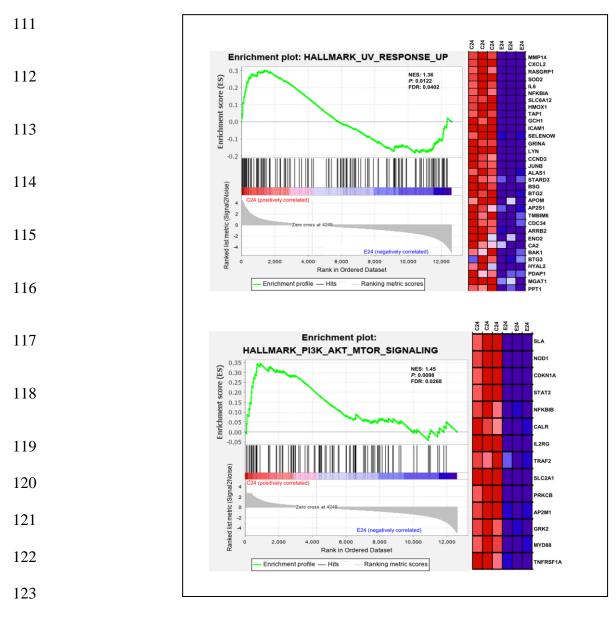
- 74 Table S1. Summary of sequencing data and alignment statistics
- 75 Table S2. Enriched canonical pathways of DEGs using Ingenuity Pathway Analysis. The
- respective to the significance of canonical pathways was determined using a default threshold of *p*-values < 1E-5.
- 77 Table S3. Candidate genes for Taqman probe ID.
- Table S4. At 24 h after infection, pathogenic *Leptospira*-infected macrophages showed lower
  expression levels of gene transcripts compared to those in nonpathogenic *Leptospira*-infected
  macrophages.
- 81 Table S5. At 24 h after infection, pathogenic *Leptospira*-infected macrophages showed higher 82 expression levels of gene transcripts compared to those in nonpathogenic *Leptospira*-infected 83 macrophages.
- 84 Table S6. Pathways significantly enriched in Gene Set Enrichment Analysis.
- 85 **Table S7. Core enrichment genes**
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**FIG S1.** 

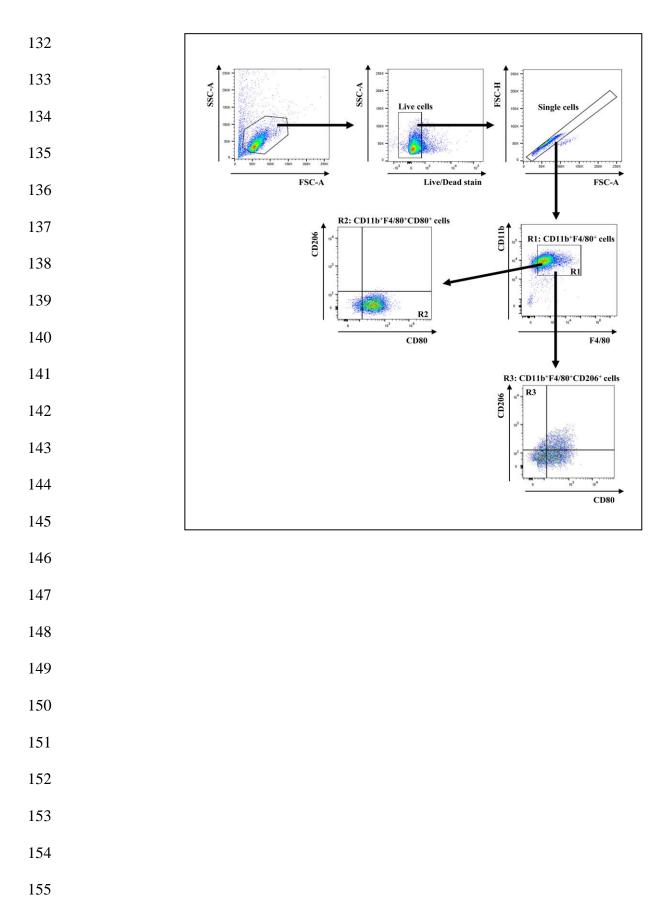


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Groups	Infection	Samples	Total number of	Total number of	Total number of	Total mapping	Total gene
	times <sup>a</sup>		clean reads <sup>b</sup>	clean reads <sup>c</sup>	mapped reads <sup>d</sup>	ratio (%) <sup>e</sup>	number
Uninfected BMDMs	2	1	60.81	60,792,594	59,212,335	97.40	24,495
		2	57.26	57,241,093	55,611,226	97.15	24,468
		3	65.97	65,943,574	64,229,828	97.40	24,661
	24	1	62.26	62,255,382	60,667,230	97.45	32,874
		2	60.39	60,388,092	58,725,491	97.25	33,194
		3	79.18	79,172,384	77,212,919	97.53	34,304
L. biflexa-infected	2	1	60.58	60,563,231	59,205,948	97.76	23,406
BMDMs		2	61.90	61,876,066	60,205,229	97.30	23,725
		3	63.74	63,706,692	62,162,068	97.58	23,411
	24	1	70.52	70,520,481	68,275,316	96.82	34,588
		2	81.37	81,363,336	78,895,826	96.97	33,625
		3	62.55	62,547,567	60,181,836	96.22	32,388
L. interrogans-infected	2	1	55.45	55,429,633	54,046,002	97.50	22,984
BMDMs		2	59.81	59,785,235	58,571,055	97.97	22,919
		3	55.90	55,877,445	54,412,993	97.38	23,238
	24	1	47.95	47,951,630	46,287,713	96.53	32,695
		2	57.20	57,192,970	55,195,931	96.51	31,560
		3	53.23	53,231,811	51,516,773	96.78	31,140

156 **Table S1. Summary of sequencing data and alignment statistics** 

<sup>*a*</sup> Infection times, hours

<sup>b</sup> Total number of raw reads: The reads amount before filtering, Unit: Mb

<sup>158</sup> <sup>c</sup> Total number of clean reads: The number of clean reads after filtering; removal of low quality sequence (Quality value  $\geq 20$ ; sequencing reads

trimming and quality control using CLC Genomics Workbench v10.1 (CLC Bio, Denmark)

<sup>d</sup> Total clean reads alignment and mapping on to the reference genome (*Mus musculus* GRCm38)

161	<sup>e</sup> Total mapping ratio (%): The percentage of mapped reads
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Gene symbol	Gene name	TaqMan assay ID	
NLRP3	NLR family, pyrin domain containing 3	Mm00840904_m1	
TLR1	Toll-like receptor 1	Mm00446095_m1	
TLR2	Toll-like receptor 2	Mm00442346_m1	
TNF-a	Tumor necrosis factor-alpha	Mm99999068_m1	
IL-10	Interleukin 10	Mm00439614_m1	
CCL2	Chemokine (C-C motif) ligand 2; MCP-1	Mm00441242_m1	
IL-6	Interleukin 6	Mm01210733_m1	
CXCL1	Chemokine (C-X-C motif) ligand 1	Mm04207460_m1	
CCR1	Chemokine (C-C motif) receptor 1	Mm00438260_s1	
Emr1	EGF-like module containing, mucin-like, hormone; F4/80	Mm00802529_m1	
HIF-1a	Hypoxia inducible factor 1, alpha subunit	Mm01283761_g1	
GAPDH	Glyceraldehyde-3-phosphate dehydrogenase	Mm99999915_g1	

**Table S3. List of candidate genes for Taqman probe ID.**