Supplementary Figures

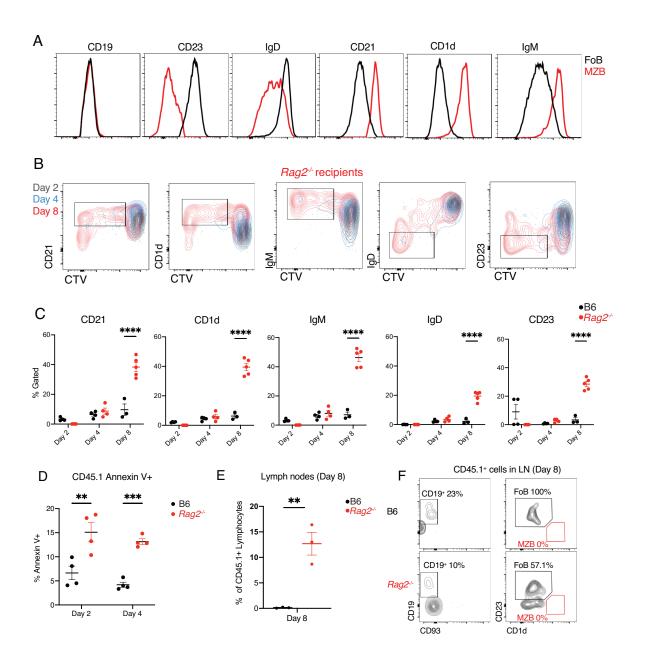


Figure S1, related to Figure 1 and 2. The acquisition of a marginal zone B-like phenotype precedes proliferation and is exclusive to the spleen. (A) Histograms depicting cell surface expression of markers indicated above in resting follicular (FoB, black) and marginal zone B cells (MZB, red); (B) Cell surface phenotype change in 2, 4, or 8 days (black, grey, and red respectively) following adoptive transfer with respect to cell trace violet (CTV) dilution in $Rag2^{-/-}$ recipients. One representative of 4 is shown; (C) Percentage of divided cells positive for the indicated cell surface marker on day 2, 4, or 8 in B6 (black) and $Rag2^{-/-}$ (red) recipients. Error bars depict SEM; (D) Apoptosis was measured at 2 and 4 days post adoptive transfer of FoB into B6 (black) and $Rag2^{-/-}$ (red) recipients. %Annexin-V⁺ fraction (including both 7-AAD⁻ and 7-

AAD⁺) is shown as a measure of total apoptosis. **(E)** CD45.1⁺ B cells detected in lymph nodes on day 8 after adoptive transfer of FoB cells into B6 and $Rag2^{-/-}$ recipients; **(F)** Percentage of CD45.1⁺ lymphocytes detected in lymph nodes of B6 (black) or $Rag2^{-/-}$ recipients (red) on day 8. (C,E) **p<0.01 ****p<0.001 ****p<0.0001: two-way ANOVA **(C,D)**, Student's t test **(E)**. Each data point represents an individual mouse.

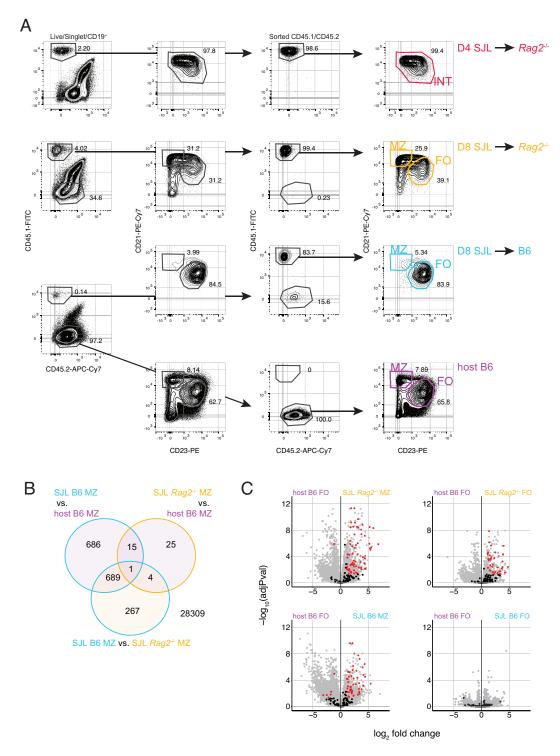


Figure S2, related to Figure 4. Follicular B cells adoptively transferred into a lymphopenic environment attain a full marginal zone B cell transcriptome. (A) Representative sort purification gates are shown for all samples in Figure 4. Samples were initially sorted based on CD45.1/2 expression (first column). A flow cytometry plot derived from this gate is shown for reference only in column 2. A second sort was then performed on purified CD45.1⁺ and CD45.2⁺ samples with sorting based on CD23/CD21 expression into intermediate (day 4 only) as well as MZB and FoB cell gates (day 8), as shown. CD45.2⁺ host B cells at day 8 were purified as

equivalents to normal MZB and FoB cells; (**B**) Venn diagrams displaying differential gene testing results of the indicated comparisons. Shown are the number of genes differential (adj-P value <0.01, log2 fold change >2) without respect to direction; (**C**) Volcano plots indicating the magnitude and significance of gene expression changes between indicated groups for all (grey), and highlighted (black/red) empirically defined MZB cell Notch2-regulated genes (genes significantly downregulated in MZB cells after 24 hours anti-Notch2 antibody blockade) (Gaudette et al., *J Clin Invest* 131(20), e151975, 2021). Red color indicates significance (adj-P < .05; log2 fold change >1).

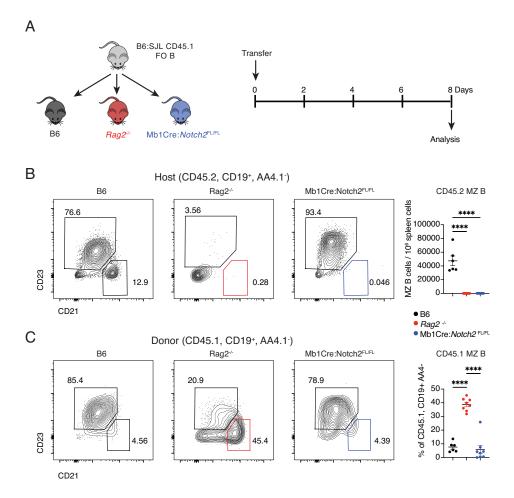


Figure S3. Lack of host marginal zone B cells alone is not sufficient to drive the transdifferentiation of adoptively transferred follicular B cells into MZ-like B cells. Congenically marked FoB cells were labeled with CellTrace Violet and adoptively transferred into either B6, Rag2^{-/-}, or Mb1-Cre;Notch2^{f/f} recipients, which selectively lack MZB cells. Flow cytometric analysis of splenocytes performed 8 days post transfer. (A) Schematic depiction of the experimental design. (B) Representative FoB vs. MZB cell gating is shown for the host spleen B cell compartment in each genotype (gated on live, singlet, CD45.2⁺ CD19⁺ AA4.1⁻) and MZB cells are quantified per spleen (right) for all animals. (C) Representative FoB and MZB cell gating is shown for the donor B cell compartment (gated on live, singlet, CD45.1⁺ CD19⁺ AA4.1⁻) in each genotype. The MZB cell phenoype as a percentage of donor-derived B cells is shown on the right. (B,C) Pooled results from two independent experiments are shown as individual animal data points (n=6-8 per group, mean + SEM). *****p<.0001 one way ANOVA, Tukey's correction for multiple comparisons.