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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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St	at	101	ŀπ	$\cap \subseteq$

n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
\boxtimes	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\boxtimes	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection

JANUS G3 Mini built in software (WinPREP); Tecan micorplate readers built in softwares

Data analysis

Data processing and IDentif.Al-x analyses in this study were conducted using basic functions written in Python programming language and in Matlab R2020a (Mathworks, Inc.). IDentif.Al-x analyses were conducted using the built-in "stepwiselm" function in Matlab R2020a as provided in Supplementary Data 1. The custom codes can be shared on reasonable request to the corresponding authors.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our $\underline{\text{policy}}$

All data generated and analyzed during this study are included in this published article and its supplementary information. The %Inhibition and %Cytotoxicity data together with the code for the 6-drug IDentif.Al-x drug combination optimization step, can be found in the Supplementary Data 1. All %Inhibition and %Cytotoxicity predictions generated in IDentif.Al analysis can be found in Supplementary Data 2. The experimental data underlying the monotherapy and validation analyses can be found in Supplementary Data 3 and 4, respectively.

Field-specific reporting					
Life sciences For a reference copy of t	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection. Behavioural & social sciences Ecological, evolutionary & environmental sciences he document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf				
Lite scier	ices study design				
All studies must dis	close on these points even when the disclosure is negative.				
Sample size	l in vitro cellular experiments were performed in at least 3 biological replicates as it is established in the field.				
	IMPORTANT NOTE: This information is already included in the submitted manuscript.				
Data exclusions	In the Monotherapy dose-response curve analysis, we observed that %cytotoxicity >25% had effects on the inhibition assay result, therefore we excluded the %Inhibition data points that had >25% corresponding %Cytotoxicity values (Excluded data are represented by unfilled blue circles in Supplementary Fig S1). No other data were excluded in other analysis.				
IMPORTANT NOTE: This information is already included in the submitted manuscript.					
Replication	All cellular experimental treatments were performed in at least 3 biological replicates. We do not expect any issues with the reproducibility of the results beyond a limited access to the used viral sample.				
Randomization	No randomization was performed.				
Blinding	The laboratory staff performing the experimental work on viral inhibition was blinded to the well content arrangement on the plates.				
-	IMPORTANT NOTE: This information is already included in the submitted manuscript.				
Reportin	g for specific materials, systems and methods				
	on from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, ed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.				
Materials & experimental systems Methods					
n/a Involved in th					
Antibodies	ChIP-seq				
Eukaryotic cell lines Flow cytometry					
Palaeontology and archaeology MRI-based neuroimaging					
Animals and other organisms Human research participants					
Clinical data					
Dual use research of concern					
Eukaryotic c	ell lines				
Policy information	about <u>cell lines</u>				
Cell line source(s) THLE-2 cells: CRL2706, ATCC; AC16 cells: SCC-09, Millipore; Vero E6 cells C1008					

None of the cell lines were authenticated

No commonly misidentified cell lines were used

The cell lines were not tested for mycoplasma contamination

Authentication

Mycoplasma contamination

Commonly misidentified lines (See <u>ICLAC</u> register)

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals

No laboratory animals were used

Wild animals

No wild animals were used

Field-collected samples

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) was previously isolated from a nasopharyngeal swab in early 2020 in Singapore as described before and has undergone several rounds of propagation to form SARS-CoV-2 original, propagated variant used in this study (virus source: Biological Defence Programme, DSO National Laboratories). This propagated variant was identified to have genetic mutations as compared to the original strain. The second viral strain – the B.1.351 (Beta) variant – was isolated from a nasopharyngeal swab in early 2021 in Singapore and was registered in GISAID EpiFlu[™] Database under hCoV-19/Singapore/239/2021 (virus source: National Public Health Laboratory, NCID). The third viral strain – the B.1.617.2. (Delta) variant – was isolated from a nasopharyngeal swab in 2021 in Singapore (virus source: National Public Health Laboratory, NCID).

IMPORTANT NOTE: This information is already included in the submitted manuscript

Ethics oversight

NA

Note that full information on the approval of the study protocol must also be provided in the manuscript.