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## **Supplemental information**

### ***In vivo* generation of CAR**

#### **T cells in the presence of human myeloid cells**

**Naphang Ho, Shiwani Agarwal, Michela Milani, Alessio Cantore, Christian J. Buchholz, and Frederic B. Thalheimer**

## Supplemental Information

**Table S1: Particle concentration and transducing units of applied LV stocks.**

Vector	Particle/mL	<sup>a</sup> T.u./mL
<sup>b</sup> CD4-LV	$1.3 \times 10^{12}$	<sup>d</sup> $1.4 \times 10^7$
<sup>b</sup> CD8-LV	$1 \times 10^{12}$	<sup>e</sup> $1.2 \times 10^7$
<sup>b</sup> CD4-LV	$6 \times 10^{11}$	<sup>d</sup> $3 \times 10^6$
<sup>b</sup> CD8-LV	$5.7 \times 10^{11}$	<sup>f</sup> $5.5 \times 10^6$
<sup>c</sup> CD4-LV	$4.3 \times 10^{11}$	<sup>f</sup> $1.7 \times 10^7$
<sup>c</sup> CD4-LV <sup>sh</sup>	$9.9 \times 10^{11}$	<sup>f</sup> $6.5 \times 10^7$
<sup>c</sup> CD8-LV	$7.3 \times 10^{11}$	<sup>f</sup> $2.6 \times 10^6$
<sup>c</sup> CD8-LV <sup>sh</sup>	$1.8 \times 10^{12}$	<sup>f</sup> $3.7 \times 10^6$

<sup>a</sup>(T.u.) Transducing units.

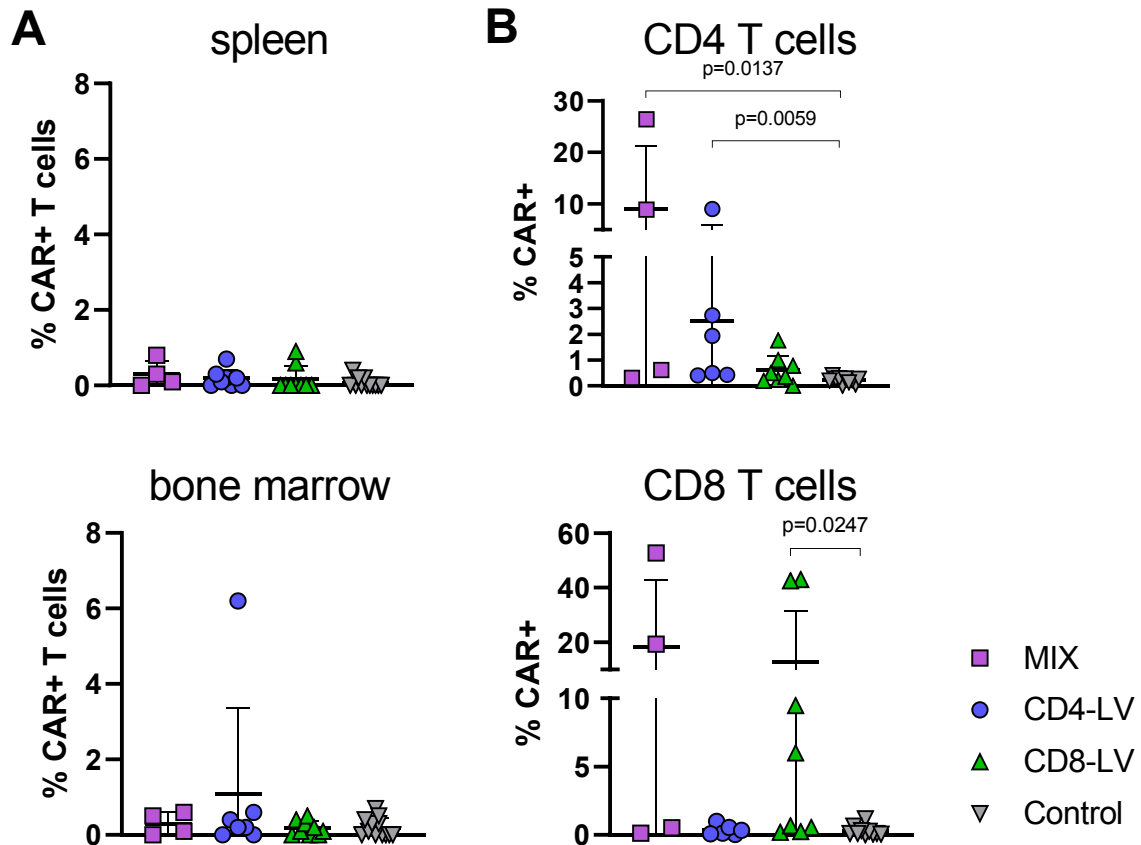
<sup>b</sup>stock used in Figure 1.

<sup>c</sup>stock used in Figure 5.

<sup>d</sup>t.u. determined on A301 cells.

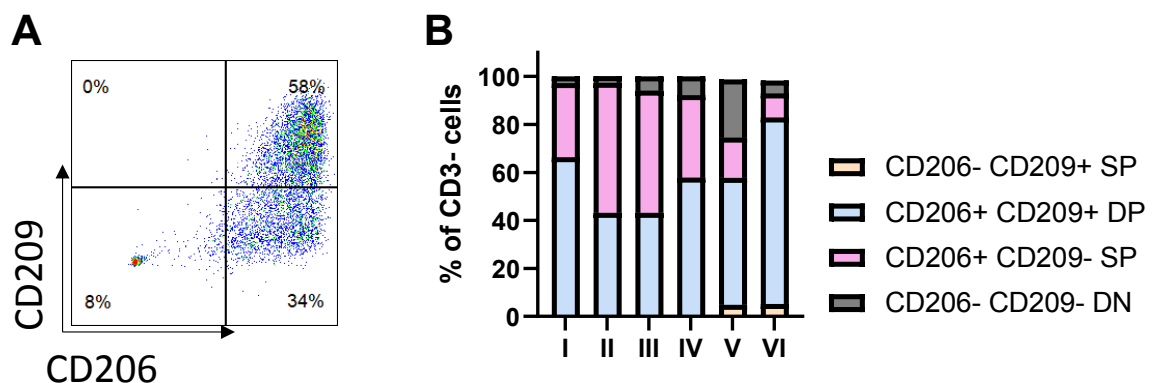
<sup>e</sup>t.u. determined on J76S8ab cells.

<sup>f</sup>t.u. determined on PBMCs.



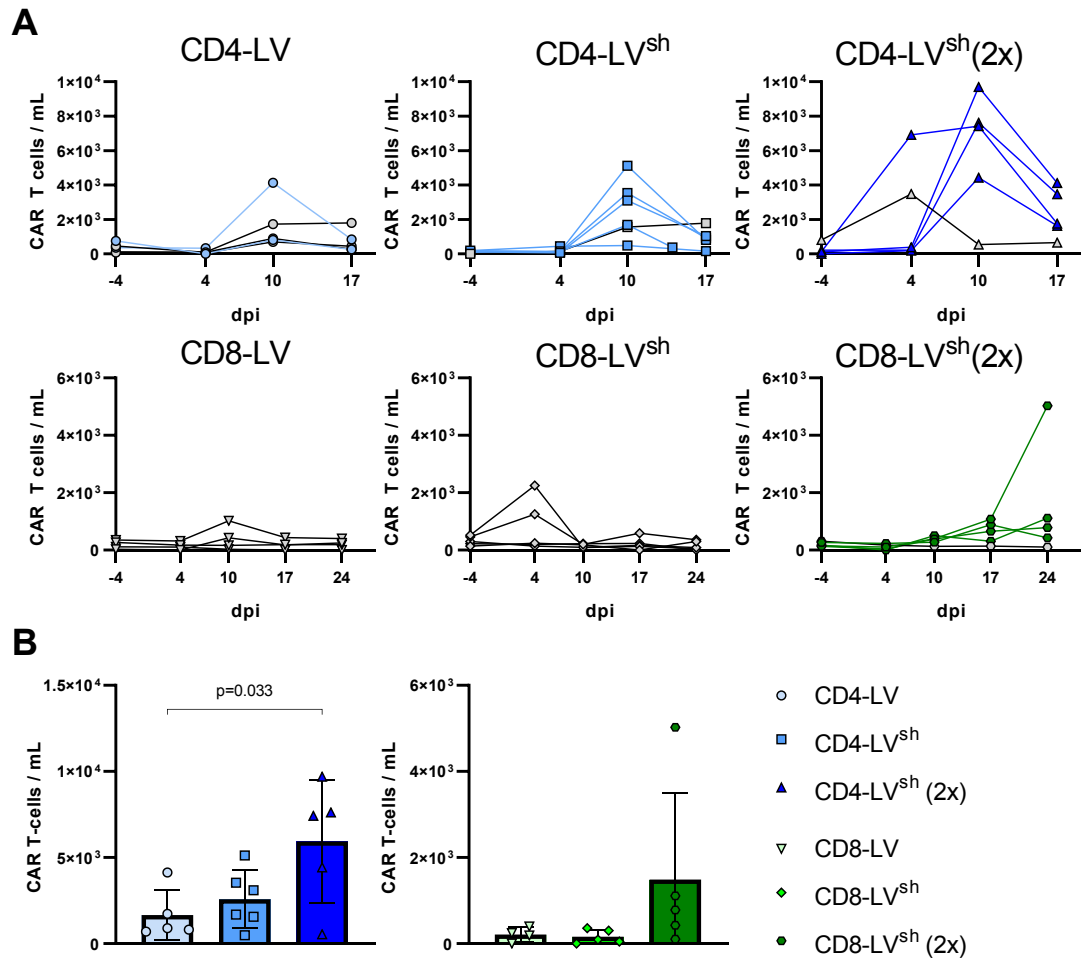
**Figure S1: CAR T cells in organs and co-culture.**

**A)** Percentage of CAR T cells in the spleen and bone marrow is shown for the different vector groups of the experiment in Fig. 1. **B)** *In vitro* expanded CAR+ T cells after further cultivation of isolated splenocytes with irradiated Raji cells. Organ data are shown for each mouse with mean and standard deviation (SD) of the group. n= 4 (MIX), 8 (CD4-LV), 9 (CD8-LV) and 12 (Control) of two experiments. Co-culture data are shown for each mouse, performed in technical triplicates, with mean and SD of the group. n= 4 (MIX), 6 (CD4-LV), 8 (CD8-LV) and 10 (Control) of two experiments. Statistics were determined by non-parametric Kruskal-Wallis ANOVA with Dunn's multiple comparisons test and significant p-values are indicated.



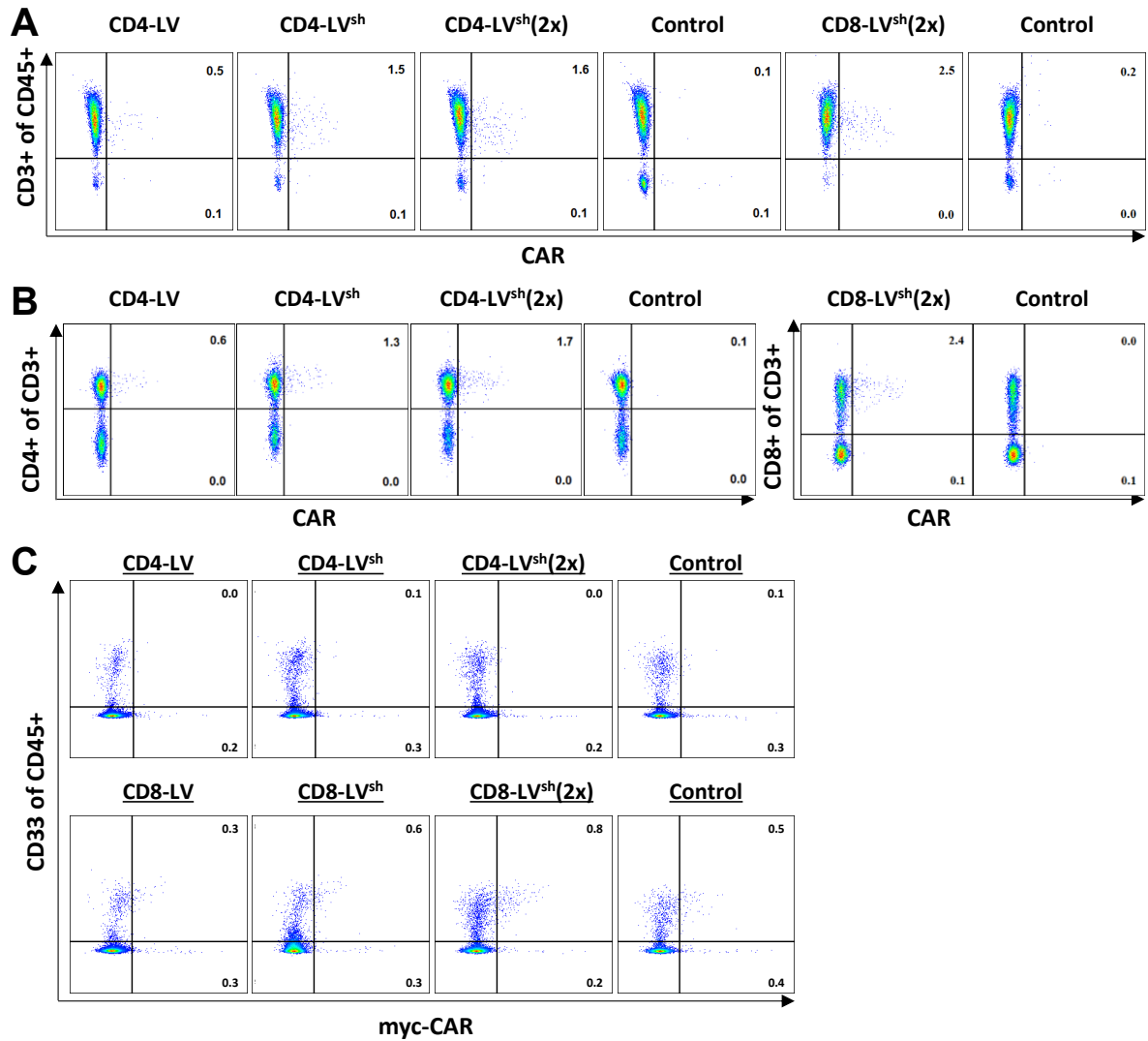
**Figure S2: Characterization of polarized macrophages.**

Macrophages, polarized from monocytes with GM-CSF, were characterized for surface expression of CD206 and CD209 by FACS. **A)** A representative FACS plot for macrophage polarization. **B)** Marker expression on polarized macrophages of five different donors used in four independent experiments. (SP) single positive, (DP) double positive, (DN) double negative.



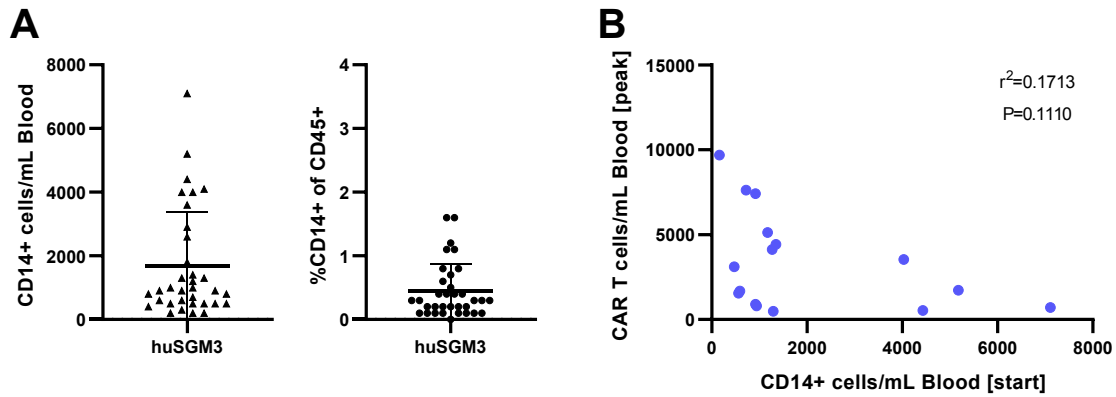
**Figure S3: Absolute *in vivo* CAR T cell numbers in blood of huSGM3 mice.**

See Figure 5 for experimental set up. **A)** Kinetic of CAR<sup>+</sup> T cells in blood shown for each mouse. **B)** Side-by-side comparison of CAR T cell count for the CD4-LV group on day 10 and for the CD8-LV group on day 24. Each data point represents an individual mouse with mean and standard deviation of the group. Statistics for **(B)** were determined by one-way ANOVA with Turkey's multiple comparisons test and significant p-values indicated. n= 5 (CD4-LV), 6 (CD4-LV<sup>sh</sup>), 5 (CD4-LV<sup>sh</sup>(2x)), 4 (CD8-LV), 5 (CD8-LV<sup>sh</sup>), 5 (CD8-LV<sup>sh</sup>(2x)) in one experiment. dpi: days post injection.



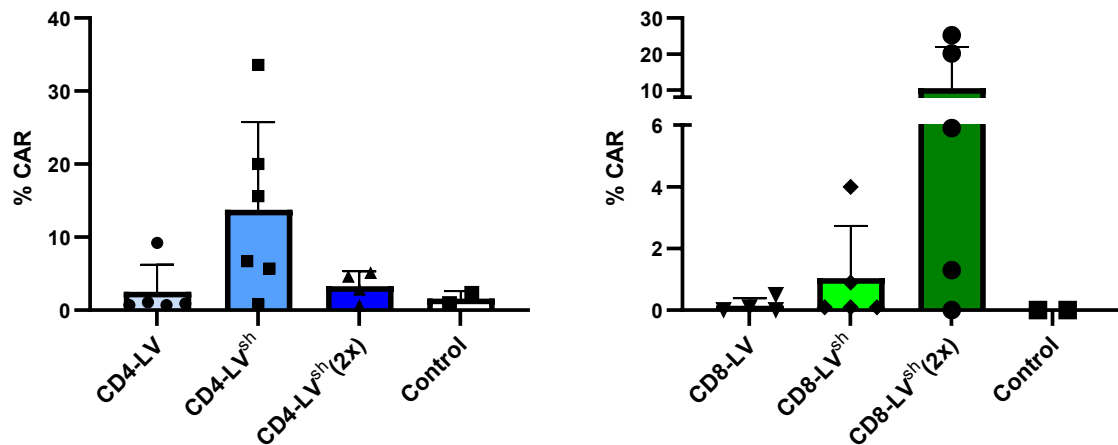
**Figure S4: FACS plots of CAR signal in blood and spleen.**

See Figure 5 for experimental set up. **A)** Representative FACS plots of *in vivo* generated CAR T cells in blood for human T cell specific transduction are presented as CD3<sup>+</sup> of human CD45<sup>+</sup> cells positive for CAR, detected via its myc-tag. **B)** Representative FACS plots from blood for specific transduction in T cell subtype are shown as CD4<sup>+</sup> or CD8<sup>+</sup> of CD3<sup>+</sup> cells positive for CAR. **C)** Representative FACS plots from spleen for background signal in CD33<sup>+</sup> of human CD45<sup>+</sup> cells. Frequency of CAR signal as percent is indicated in the plots.



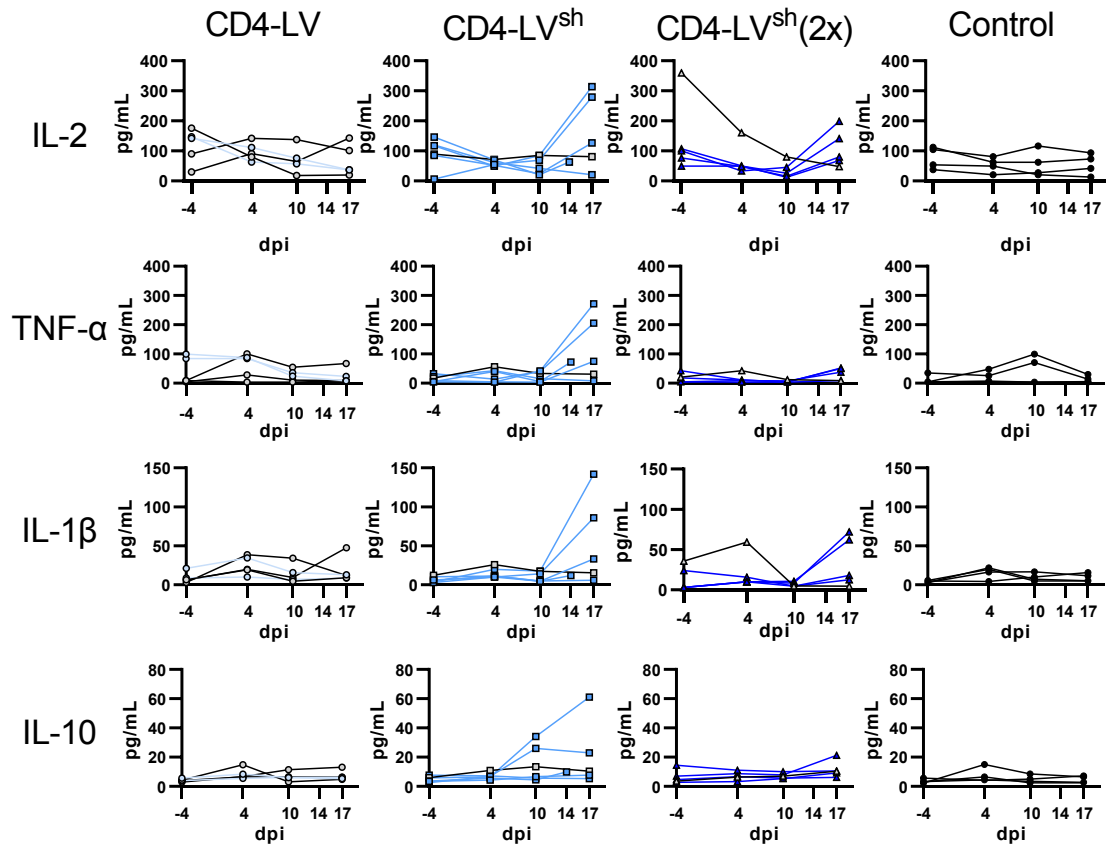
**Figure S5: Correlation of monocyte level with *in vivo* CAR T cell generation by CD4-LV.**

**A)** Blood of all huSGM3 mice from Figure 5 were analyzed for human CD14<sup>+</sup> cells before experiment start (day -4) by flow-cytometry analysis. CD14<sup>+</sup> Cells were pre-gated for human CD45<sup>+</sup>, human CD3<sup>-</sup> and human CD19<sup>-</sup> cells. Individual mice are shown as data point with mean and standard deviation for n=34. **B)** Correlation of CD14<sup>+</sup> cell number in the blood before experiment start (day -4) with *in vivo* generated CAR T cell numbers in the blood on day 10 for CD4 targeted LVs. Individual mice are shown as data point for n=16. Correlation in **(B)** was determined by Pearson correlation analysis with indicated  $r^2$  and P value.



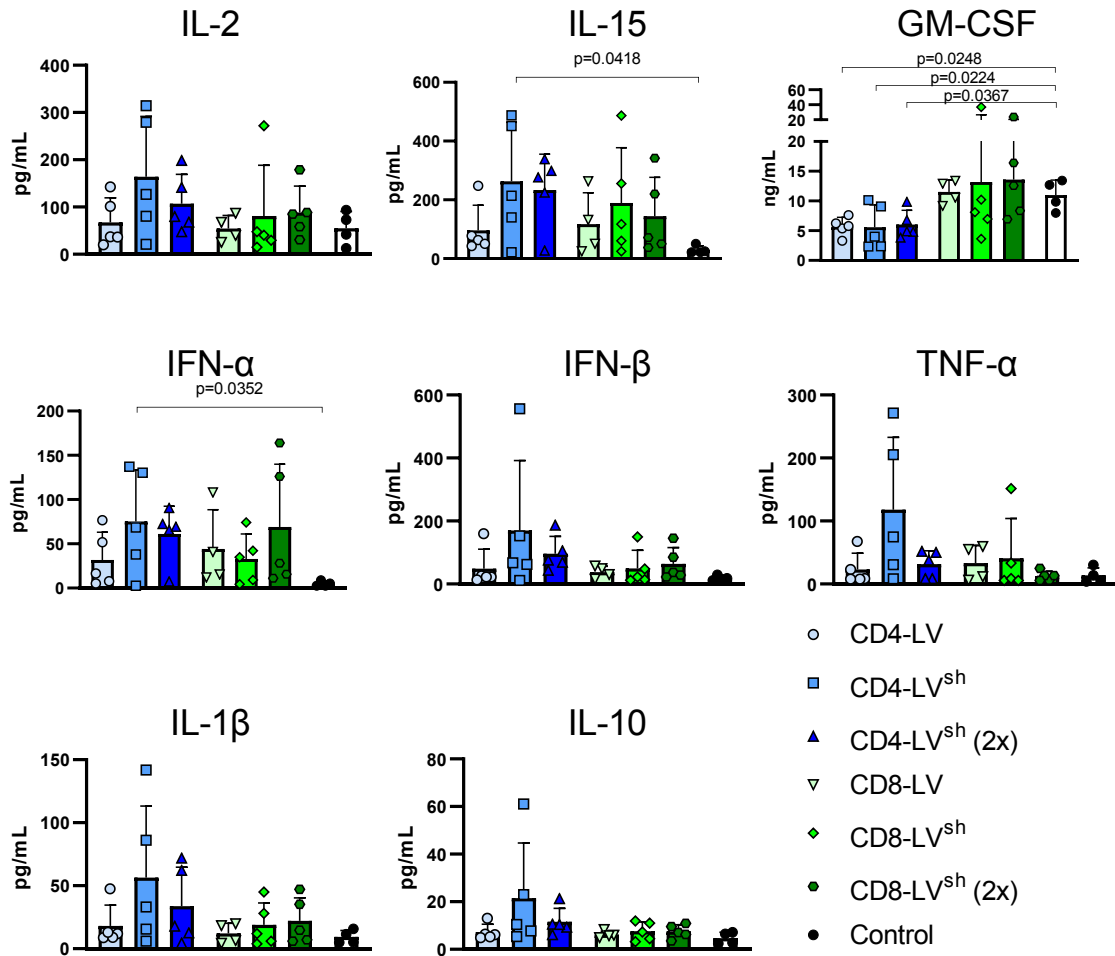
**Figure S6: *In vitro* CAR T cell expansion from splenocytes.**

See Figure 5 for experimental set up. Expansion of CAR T cells from splenocytes of LV injected huSGM3 mice upon co-culture with irradiated Nalm6 tumor cells for seven days. Percentage of expanded CAR<sup>+</sup> T cells of the respective subpopulation is shown for each mouse as data point with mean and standard deviation of the group. Statistics were determined by one-way ANOVA with Dunnett's multiple comparisons test.



**Figure S7: Kinetic of further plasma cytokines in CD4-LV injected huSGM3 mice.**

See Figure 5 for experimental set up. Plasma cytokines of huSGM3 mice injected with the indicated LVs were determined by bead-based multi-analysis kit. The concentrations for each cytokine over time are shown for each mouse. Mice determined as CAR negative in blood are depicted in grey with black connecting lines. n= 5 (CD4-LV), 6 (CD4-LV<sup>sh</sup>), 5 (CD4-LV<sup>sh</sup>(2x)) and 4 (Control) from one experiment. dpi: days post injection.



**Figure S8: Plasma cytokines in LV injected huSGM3 mice on day 17.**

See Figure 5 for experimental set up. Plasma cytokines of huSGM3 mice on day 17 after vector application were measured by bead-based multi-analysis kit. Individual mice are shown as data point with mean and standard deviation of the group. Statistics were determined by one-way ANOVA with Dunnett's multiple comparisons test with indicated significant p-values. n= 5 (CD4-LV), 6 (CD4-LV<sup>sh</sup>), 5 (CD4-LV<sup>sh</sup> (2x)), 4 (CD8-LV), 5 (CD8-LV<sup>sh</sup>), 5 (CD8-LV<sup>sh</sup>(2x)) and 4 (Control) from one experiment.