iScience, Volume 25

Supplemental information

A Tead1-Apelin axis directs paracrine

communication from myogenic

to endothelial cells in skeletal muscle

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SUPPLEMENTARY MATERIALS

Supplementary Figure 1.

Characterization of the *ApIn* promoter and identification of transcription factors in Yeast 1 Hybrid screen, Related to Figure 1.

Supplementary Figure 2.

Effects of Zfp319 and Zdhhc9 on ApIn expression in C2C12 cells, Related to Figure 1.

Supplementary Figure 3. Expression profiles of single-cell RNA-sequencing clusters, Related to Figure 3.

Supplementary Figure 4.

Expression profiles of six transcription factors in Y1H from single-cell and nucleus RNA-sequencing datasets, Related to Figure 3.

Supplementary Table 1. Primers used for Chromatin Immunoprecipitation (ChIP) qPCR Assay, Related to Figure 1.



Supplementary Figure 1. Characterization of the *Apln* promoter and identification of transcription factors in Yeast 1 Hybrid screen, Related to Figure 1. (A) Epigenomic annotation near the TSS of apelin precursor gene (*Apln*) in mouse ES-Bruce4 cells from the WashU Epigenome Browser. Tag densities (blue) of proximal promoter markers H3K4me3, H3K9ac, EP300, and POLR2A by ChIP-seq are highlighted (top). Placental Con represents the conservation score between 20 mammalian species available from WashU Epigenome Browser (Miller et al., 2007) (**B**) Nano luciferase activity of four putative *Apln* promoter regions and no-promoter vector transfected into C2C12 cells, normalized by co-transfected firefly luciferase activity. Mean \pm s.e.m. of n = 4 cell culture replicates per group. (**C**) Y1H screen growth cluster results using -400/-1bp fragment of *Apln* promoter mated with two TF libraries. (**D**) Z-score-normalized Y1H spot intensities for all 745 TF-promoter interactions using -400/-1bp promoter fragments.



Supplementary Figure 2. Effects of Zfp319 and Zdhhc9 on *Apln* expression in C2C12 cells, Related to Figure 1. (A-B) *Zfp319* (A) and *Apln* (B) mRNA expression measured by RT-qPCR in C2C12 myoblasts transfected with scrambled control or *Zfp319* targeted siRNA for 3 d, n = 16 cell culture replicates per condition. (C) Apln peptide concentration in culture medium measured by enzyme immunoassay (EIA) after 3 d culture from the same experiment. n = 20 cell culture supernatants per condition. (D-E) *Zdhhc9* (D) and *Apln* (E) mRNA expression in C2C12 myoblasts transfected with scrambled control or *Zdhhc9* targeted siRNA for 3 d, n = 16 cell culture replicates per condition. (F) Apln peptide concentration in culture medium measured by enzyme immunoassay (EIA) after 3 d culture from the same experiment. n = 20 cell culture replicates per condition. (F) Apln peptide concentration in culture medium measured by enzyme immunoassay (EIA) after 3 d culture from the same experiment. n = 20 from one experiment. For all panels, data are presented as mean \pm s.e.m., and *P* values are reported from two-tailed, unpaired t-tests between conditions.



Supplementary Figure 3. Expression profiles of single-cell RNA-sequencing clusters, Related to Figure 3. (A) Dot plots representing marker gene expression used to characterize cell clusters in the single-cell RNA-sequencing muscle regeneration data set from Figure 3C-E. Cell clustering was performed using SNN and annotations were manually determined based on these gene expression patterns. For each gene/cluster combination, the average expression level is shown in red color and the fraction of cells with non-zero expression is shown as the dot size. Some clusters shown here are not plotted in Figure 3E if not consistently observed at different time-points in the dataset.



Supplementary Figure 4. Expression profiles of six transcription factors in Y1H from single-cell and nucleus RNA-sequencing datasets, Related to Figure 3. (A) Dot plots representing cell-specific TF gene expression from integrated 102 publically-available single-cell or nucleus RNA-sequencing datasets by harmony batch-correction (McKellar et al., 2020).

Gene	Forward Primer	Reverse Primer	Genomic Coordinates
Ankrd	GAGGGGAGGACAAGCTAACC	CGATGTGATCACCACCAAAG	Chr10:92681001– 92681083
Ctgf	GCCAATGAGCTGAATGGAGT	CAATCCGGTGTGAGTTGATG	Chr6:132272566- 132272653
Apln	СТСТСССТСТТССТВССТС	TTTTGTAGCTTGTGGTTTGGC	Chr X:48034975 - 48034875
Neg- control	ACCAACACTCTTCCCTCAGC	TTATTTTGGTTCAGGTGGTTGA	Chr10:60902566- 60902665

Supplementary Table 1. Primers used for Chromatin Immunoprecipitation (ChIP) qPCR Assay, Related to Figure 1. Forward and reverse primers to detect *Ankrd1, Ctgf, Apln, Neg* promoters in ChIP-qPCR detection.