

Supplementary Tables and Table Legends

Table S1. Subsets of Nucleolar Assembly Factors

Subsets of Nucleolar Assembly Factors						
Subset *	Operative Distribution	Latent Distribution	Suggested Role	Fraction Essential **	Non-Essential AFs ^x	Orthologs Implicated in Disease ^{+ o}
SSU-In	Inner layer	Inner layer	Initiator ?	2/5	Efg1, Nop6, Nsr1	
SSU-F	Inner layer	Outer layer	SSU knob formation	50/52	Fyv7, Rrt14	BUD23 [Bud23] Cirhin [Utp4] DHX37 [Dhr1] ESF1 [Esf1] UTP14 [Utp14] WDR36 [Utp21] WDR43 [Utp5]
LSU-Ou	Outer layer	Outer layer	LSU knob formation	38/55	Abl1, Arx1, Bmt2, Bud20, Bud23, Cic1, Dbp7, Fpr3, Fpr4, Mrt4, Nop12, Nop13, Nop16, Puf6, Rae1/Gle2, Rcm1, Ssf1, Ssf2, Tri1	DCAF13 [Sof1] DNAJC21 [Jjj1]
LSU-F	Outer layer	Inner layer	Initiator ?	3/4	Rsa3	
snoRNP proteins	Inner layer	Inner layer	modify rRNA	7/7	none	DKC1 [Cbf5] SNORD118 #
Rnt1, nuclease	Inner layer	Inner layer	removal of SSU knobs			nuclease: LAS1L RMRP #
Rcl1, nuclease	Outer layer	Outer layer	release of transcripts			
Rrp5	Inner layer	Outer layer	binds ITS1			
rDNAPs	Axis	Axis	Packaging of rDNA ?		Csm1, Fob1, Hmo1, Lrs4, Sir2, Tof2, Top1	
Other (Pol A, Dbp3, Prp43)	Inner layer	Inner layer	transcription, early steps			TCOF1, POLR1A [Rpa190] POLR1C/D [RPA/C19; RPA/C40]

Table S1 Subsets of Assembly Factors - related to Table 1

In addition to the information summarized in Table 1 and Table S2, this table specifies the suggested role of each subset of AFs and the fraction of the AFs in each group that are essential for mitotic growth [2]. We also list those for which homologs in man have been implicated in disease. The group of non-essential AFs in yeast includes proteins associated with the 5'-ETS (Bud21, Nan1/Utp17), other proteins required for making the SSU (Bud22, Cms1, Efg1, Fyv7, Nsr1, Rrp8, Rrt14, Slx9, Utp30) and proteins required for making the LSU (Bmt2, Bud20, Cgr1, Cic1/Nsa3, Dbp7, Fpr3, Fpr4, Jjj1, Loc1, Mrt4, Ngl2, Nop12, Nop13, Nop16, Puf6, Rrp6, Ssf1, Ssf2, Tri1).

Collection of data. The *Saccharomyces* Genome Database (SGD) was used to gather information on the abundance, molecular weight and amino acid sequence of assembly factors. For that database, BioPerl and CODONW were used to calculate isoelectric points and abundance data were averaged from multiple quantitative analyses of cells under standard growth conditions [5]. Predictor of Natural Disordered Regions (PONDR) was used to predict overall disorder based on the amino acid sequence (<http://www.pondr.com/>). FoldIndex was also used to make such estimates and the two algorithms gave similar estimates.

For human proteins, we used UniProt Knowledgebase (UniProtKB) and ExPASy to retrieve molecular weight and isoelectric point information. PONDR was again used to predict order.

Master List of Assembly Factors and Their Physical Properties											
name	name	MW (kDa)	MW (kDa)	IEP	IEP	predicted disorder (%)	predicted disorder (%)	abundance mean	abundance STD	subset	tripartite domain
Nan1/Utp17 *	WDR75	101.22	94.49	6.31	5.65	17.58	17.31	6811	3486	SSU-F	5'-ETS
Utp5 *	WDR43	71.96	74.89	4.56	5.37	34.53	27.62	6874	7312	SSU-F	5'-ETS
Utp6	UTP6	52.43	41.53	9.33	7.59	20.68	16.25	4712	2273	SSU-F	5'-ETS
Utp8		80.16		4.78		18.93		7512	3299	SSU-F	5'-ETS
Utp9		65.23		4.48		24.7		8516	7706	SSU-F	5'-ETS
Utp13 *	TBL3	91.00	89.03	5.06	6.44	21.05	21.41	6417	4007	SSU-F	5'-ETS
Utp15 *	UTP15	57.70	58.41	9.76	9.18	28.65	23.94	4549	3327	SSU-F	5'-ETS
Utp18 *	UTP18	66.39	62.00	4.61	8.93	31.99	33.63	6638	7492	SSU-F	5'-ETS
Utp21 *	WDR36	104.79	105.32	7.89	7.33	23	19.24	4129	2115	SSU-F	5'-ETS
Bud21/Utp16		24.38		6.55		83.64		3559	2746	unknown	5'-ETS
Dip2/Utp12 *	WDR3	106.32	106.09	5.89	6.2	21	24.18	5172	3868	unknown	5'-ETS
Fcf2	DNTTIP2	25.64	84.46	10.17	5.86	71.43	60.58	3755	1318	unknown	5'-ETS
Imp3 #	IMP3	21.89	21.85	10.33	9.54	22.4	30.43	5499	5796	unknown	5'-ETS
Imp4 +	IMP4	33.48	21.85	9.5	9.54	31.38	30.43	4976	2363	unknown	5'-ETS
Mpp10	MPHOSPH10	66.89	78.86	4.27	4.77	66.27	59.18	6676	3779	unknown	5'-ETS
Pwp2/Utp1 *	PWP2	103.94	102.45	4.71	5.76	17.44	17.3	5498	4011	unknown	5'-ETS
Rrp9 *	RRP9	65.04	51.84	5.43	7.97	34.73	38.53	4905	1950	unknown	5'-ETS
Sas10/Utp3	UTP3	70.19	54.55	4.32	5.5	59.18	64.93	6159	3201	unknown	5'-ETS
Sof1 *	DCAF13	56.80	51.40	9.91	9.3	36.61	17.53	4982	2024	unknown	5'-ETS
Utp4 *	CIRH1A	87.79	76.89	6.52	9.03	18.17	4.37	4299	2439	unknown	5'-ETS
Utp7/Kre31 *	WDR46	62.33	68.07	10.02	9.69	45.31	52.3	6067	2569	unknown	5'-ETS
Utp10 -	HEAR1	200.06	242.37	6.44	6.11	13.23	18.24	8060	3856	unknown	5'-ETS
Utp11	UTP11L	29.76	30.00	10.91	10.08	76.4	64.8	2954	1720	unknown	5'-ETS
Dbp8 !	DDX49	47.89	54.22	10.24	9.21	21.63	29.24	3904	3732	SSU-F	pre-40S
Dhr2 !		82.73		9.22		33.47		1734	1002	SSU-F	pre-40S
Fyv7		18.21		10.99		87.42		2403	1324	SSU-F	pre-40S
Noc4/Utp19 -	NOC4L	63.62	58.46	6.01	7.08	14.31	26.16	3607	1572	SSU-F	pre-40S
Rrp7	RRP7A	34.47	32.33	9.35	9.61	36.36	51.07	5813	2489	SSU-F	pre-40S
Utp30	RSL1D1	31.63	54.97	10.15	10.13	24.82	49.59	3195	1655	SSU-F	pre-40S
Efg1		27.13		10.15		66.09		4774	2174	SSU-In	pre-40S
Hca4/Dbp4/											
Ecm24 !	DDX10	87.19	100.88	7.47	8.72	39.09	42.4	6606	7745	SSU-In	pre-40S
Nop6 #		25.23		10.87		64.44		5962	3007	SSU-In	pre-40S
Nop9 # &	NOP9	77.72	69.43	8.38	6.86	22.07	46.23	4330	1859	SSU-In	pre-40S
Nsr1/She5 #	NCL	44.51	76.61	4.58	4.6	63.53	55.49	25614	21560	SSU-In	pre-40S
Bfr2	AATF	61.16	63.13	4.41	4.82	37.88	55.89	7791	3997	unknown	pre-40S
Bms1 ^	BMS1	135.56	145.80	6.79	6.04	48.77	42.59	4889	3601	unknown	pre-40S
Bud22	SRFBP1	60.06	48.63	9.28	9.59	48.55	59.44	3797	1934	unknown	pre-40S
Bud23 %	WBSCR22	30.74	31.88	9.48	8.95	34.18	44.13	4328	2802	unknown	pre-40S
Cms1	CMSS1	33.39	31.88	8.56	9.26	35.05	38.35	4906	3267	unknown	pre-40S
Dim1 %	PNO1	35.96	27.92	10.16	9.75	28.3	48.02	5449	3874	unknown	pre-40S
Dim2 #	PNO1	30.33	27.92	9.71	9.75	52.55	48.02	5172	2826	unknown	pre-40S
Ecm16/Dhr1 !	DHX37	144.93	129.54	6.26	8.36	40.49	46.59	4312	1955	unknown	pre-40S
Emg1/Nep1 %	EMG1	27.89	26.72	8.58	9.28	20.24	27.46	7148	5216	unknown	pre-40S
Enp1/Meg1	BYSL	55.10	49.60	4.56	8.19	39.96	47.83	7222	4442	unknown	pre-40S
Enp2 *	NOL10	81.72	80.30	6.54	8.64	48.09	31.54	5078	2812	unknown	pre-40S
Esf2/Abt1 #		36.40		8.37		67.41		4489	3189	unknown	pre-40S
Faf1		38.89		10.02		73.41		2182	1168	unknown	pre-40S
Fal1 !	EIF4A3	45.22	46.87	9.49	6.3	16.79	30.41	2165	629	unknown	pre-40S
Kre33/Rra1	NAT10	119.35	115.72	7.94	8.5	32.43	31.28	8767	5717	unknown	pre-40S
Kri1	KRI1	68.62	82.30	4.74	5.06	57.06	60.77	9342	12381	unknown	pre-40S
Krr1 #	KRR1	37.17	43.66	10	9.78	49.05	43.31	6288	3601	unknown	pre-40S
Lcp5	NGDN	40.79	35.89	6.91	9.57	62.59	61.13	2006	1325	unknown	pre-40S

Table S2 (contd.)

Master List of Assembly Factors and Their Physical Properties											
name	name	MW (kDa)	MW (kDa)	IEP	IEP	predicted disorder (%)	predicted disorder (%)	abundance mean	abundance STD	subset	tripartite domain
Mrd1 #	RBM19	101.11	107.33	6.5	6.13	46.34	35.34	4887	3492	unknown	pre-40S
Nop14/Utp2	NOP14	94.30	97.66	7.44	7.33	47.04	55.19	5441	3311	unknown	pre-40S
Rio1/Rrp10	RIOK1	56.11	65.45	6.53	5.84	36.57	44.27	2556	1383	unknown	pre-40S
Rok1 !	DDX52	63.67	67.49	9.73	9.69	49.38	28.05	7571	8349	unknown	pre-40S
Rrp3 !	DDX47	55.98	50.64	9.71	9.18	31.54	39.34	5173	2608	unknown	pre-40S
Rrp12 -	RRP12	137.50	143.70	7.02	8.97	25.81	34.77	7240	3716	unknown	pre-40S
Rrt14		23.86		10.56		82.04		3228	2867	unknown	pre-40S
Sgd1 &	NOM1	102.84	96.25	5.74	8.28	36.93	53.72	2544	1238	unknown	pre-40S
Slx9		24.08		10.73		59.52		4639	1530	unknown	pre-40S
Utp14	UTP14A	102.97	87.97	4.68	7.67	73.08	71.47	5126	2989	unknown	pre-40S
Utp14	UTP14C	102.97	87.18	4.68	6.7	71.87	63.17	5126	2989	unknown	pre-40S
Utp20 &	UTP20	287.55	318.38	7.03	7.07	12.99	21.17	3802	2320	unknown	pre-40S
Utp22	NOL6	140.49	127.59	8.72	7.42	28.91	15.82	7828	4461	unknown	pre-40S
Utp23	UTP23	28.82	28.40	10.74	10.08	50	33.73	3903	1531	unknown	pre-40S
Brx1 +	BRIX1	33.54	44.40	9.81	9.92	28.52	47.88	10449	4015	LSU-Ou	pre-LSU
Cic1/Nsa3	RSL1D1	42.53	54.97	9.39	10.13	27.13	49.59	11772	9346	LSU-Ou	pre-LSU
Fpr3/Npi46	FPR3	46.49	39.96	4.09	8.12	79.08	11.9	10680	4750	LSU-Ou	pre-LSU
Mak5 !	DDX24	87.05	96.33	8.76	9.14	42.69	44.7	4105	2344	LSU-Ou	pre-LSU
Mak11 *	PAK1IP1	53.67	43.96	8.66	8.66	19.13	18.59	4910	2542	LSU-Ou	pre-LSU
Mak21/Noc1 &	CEBPZ	116.62	120.97	4.63	5.65	35.41	37.76	7700	3421	LSU-Ou	pre-LSU
Mrt4	MRT04	27.06	27.56	8.89	8.34	33.05	33.05	12109	5199	LSU-Ou	pre-LSU
Noc2 -	NOC2L	81.60	84.91	8.85	5.46	36.62	39.12	10327	7258	LSU-Ou	pre-LSU
Noc3 -	NOC3L	75.57	92.54	5.71	9.22	44.95	41.88	5027	2925	LSU-Ou	pre-LSU
Nog1 ^		74.42		9.18		41.58		14566	7766	LSU-Ou	pre-LSU
Nop2/Yna1 %	NOP2	69.78	89.30	4.68	9.27	52.91	60.84	11196	4039	LSU-Ou	pre-LSU
Nop4/Nop77 #	RBM28	77.84	85.73	9.67	9.26	63.21	61.26	5443	2689	LSU-Ou	pre-LSU
Nop7/Yph1	PES1	69.86	68.00	5.26	6.93	44.46	41.84	10235	6330	LSU-Ou	pre-LSU
Nop13 #		45.62		9.94		59.06		6924	3734	LSU-Ou	pre-LSU
Nop15 #		25.45		9.91		53.18		7498	2435	LSU-Ou	pre-LSU
Nop16	NOP16	51.74	46.67	8.71	6.72	27.4	23.84	6417	2633	LSU-Ou	pre-LSU
Puf6 #&		75.10		6.88		42.23		9678	6137	LSU-Ou	pre-LSU
Rix7	NVL	93.05	95.05	5	6.11	46.24	40.54	5682	4006	LSU-Ou	pre-LSU
Rrp1	RRP1	33.19	52.83	4.92	9.39	24.1	64.21	5406	2431	LSU-Ou	pre-LSU
Rrp17	NOL12	27.98	24.66	10.03	10.22	59.15	87.32	3792	2128	LSU-Ou	pre-LSU
Ssf1 +		51.77		9.96		59.16		5117	2126	LSU-Ou	pre-LSU
Tif6/Cdc95	EIF6	26.44	26.55	4.27	4.56	52.65	28.57	12902	7262	LSU-Ou	pre-LSU
Ytm1/Cst14	WDR12	51.35	47.70	6.96	5.57	35	15.82	6698	3329	LSU-Ou	pre-LSU
Dbp6 !	DDX51	68.07	72.45	9.72	8.43	30.3	31.83	4861	2833	LSU-F	pre-LSU
Rsa3		24.64		4.51		40.91		4878	1995	LSU-F	pre-LSU
Urb1/Npa1	URB1/NPA1	203.28	254.38	7.35	6.03	9.52	23.43	4666	1877	LSU-F	pre-LSU
Urb2/Npa2	URB2/NPA2	135.09	170.54	6.33	6.97	9.63	12.93	3932	2647	LSU-F	pre-LSU
Bmt2 %	BMT2	38.57	46.32	10.28	5.75	26.71	27.65	803	687	unknown	pre-LSU
Bud20	ZNF93	18.52	15.19	9.3	9.9	50.6	79.85	5349	2142	unknown	pre-LSU
Cgr1		14.43		11.2		97.5		4596	2094	unknown	pre-LSU
Dbp7 !	DDX31	83.33	94.08	9.84	10.01	22.91	40.54	2715	1711	unknown	pre-LSU
Dbp9 !	DDX56	68.07	61.58	9.72	9.34	33.52	32.8	7865	7383	unknown	pre-LSU
Dbp10 !	DDX54	112.96	98.59	9.71	10.03	38.77	51.38	5311	3228	unknown	pre-LSU
Drs1 !	DDX27	84.83	89.83	5.41	9.33	55.98	46.48	5785	2903	unknown	pre-LSU
Ebp2	EBNA1BP2	49.72	34.85	6.34	10.1	68.85	50	11139	10585	unknown	pre-LSU
Erb1 *		91.66		4.8		37.79		8395	4787	unknown	pre-LSU
Fpr4		43.86		4.31		66.07		10074	6507	unknown	pre-LSU
Ipi1 &		37.88		10.14		18.26		3250	1614	unknown	pre-LSU
Ipi3 *		61.76		5.5		15.5		5697	2575	unknown	pre-LSU

Table S2 (contd.)

Master List of Assembly Factors and Their Physical Properties											
name	name	MW (kDa)	MW (kDa)	IEP	IEP	predicted disorder (%)	predicted disorder (%)	abundance mean	abundance STD	subset	tripartite domain
Jjj1	DNAJC21	68.77	62.02	6.11	5.34	47.29	56.69	2082	1449	unknown	pre-LSU
Loc1		23.62		11.07		79.9		5272	4337	unknown	pre-LSU
Mak16	MAK16	35.68	35.36	4.9	5.27	48.37	60	4068	1908	unknown	pre-LSU
Ngl2	PDE12	59.54	67.35	7.96	6.11	29.32	26.27	4050	2114	unknown	pre-LSU
Nip7	NIP7	20.38	20.64	9.75	8.66	12.15	11.1	8306	2906	unknown	pre-LSU
Nop8 #		56.95		5.75		45.87		7413	12383	unknown	pre-LSU
Nop12 #	RBM34	51.96	48.56	10.13	10.11	45.11	51.32	7772	3542	unknown	pre-LSU
Nop53	NOP53	52.57	54.38	10.04	10.32	57.36	83.26	5690	3023	unknown	pre-LSU
Nsa1	WDR74	51.90	42.44	8.06	8.64	23.11	29.87	5501	2937	unknown	pre-LSU
Nsa2	NSA2	29.75	30.06	10.97	10.28	58.24	56.54	35483	35752	unknown	pre-LSU
Nug1 ^	NUGGC	57.71	91.13	9.44	8.85	42.88	39.07	10176	5823	unknown	pre-LSU
Rcm1 %		56.18		9.02		25.1		2016	1355	unknown	pre-LSU
Rea1/Mdn1	MDN1	559.08	632.82	4.63	5.46	24.99	28.43	5887	5392	unknown	pre-LSU
Rix1/Ipi2	PELP1	86.71	119.69	5.34	4.29	18.35	61.33	4644	2074	unknown	pre-LSU
Rlp7		36.57		10.14		53.42		8857	3068	unknown	pre-LSU
Rlp24	RSL24D1	23.98	19.62	10.21	9.99	70.85	47.24	6004	4065	unknown	pre-LSU
Rpf1 +	RPF1	35.13	40.11	10.08	10.01	31.19	40.4	6121	3435	unknown	pre-LSU
Rpf2 +	RPF2	39.60	35.58	9.57	10	39.83	33.99	9388	7843	unknown	pre-LSU
Rrb1 *	GRWD1	52.21	49.41	4.23	4.82	40.12	30.94	9170	5803	unknown	pre-LSU
Rrp6	EXOSC10	84.03	100.83	7.15	8.68	30.42	30.73	4148	2743	unknown	pre-LSU
Rrp8 %		46.00		10.1		39.03		4016	1322	unknown	pre-LSU
Rrp15	RRP15	28.20	31.48	5.43	5.39	65.6	74.82	5969	1870	unknown	pre-LSU
Rrs1	RRS1	22.97	41.19	10.14	10.69	58.62	43.96	8448	3397	unknown	pre-LSU
Rsa4 *	NLE1	57.03	53.32	9.18	6.92	16.89	15.88	4313	2097	unknown	pre-LSU
Sda1 &		86.61		6.93		47.98		4189	3101	unknown	pre-LSU
Spb1 %	FTSJ3	96.48	96.55	8.29	8.53	50.89	61.98	5451	3109	unknown	pre-LSU
Spb4 !	DDX55	69.43	68.54	9.54	9.32	28.22	26.17	3067	1157	unknown	pre-LSU
Sqt1 *		47.12		4.09		22.74		7749	3699	unknown	pre-LSU
Ssf2 +		51.86		9.69		56.73		2659	1604	unknown	pre-LSU
Tri1		26.48		10.16		50.44		2300	1066	unknown	pre-LSU
Fcf1/Utp24	FCF1	21.63	23.36	10.48	9.74	37.04	49.49	2689	1920	nuclease	N/A
Las1	LAS1L	59.35	83.06	8.05	4.64	25.3	35.69	1146	597	nuclease	N/A
Nob1	NOB1	51.74	46.67	8.71	6.72	43.14	29.13	5287	3149	nuclease	N/A
Rcl1	RCL1	40.17	40.84	9.23	9.36	34.33	23.32	6025	2731	nuclease	N/A
Rnt1		54.07		9.06		36.52		2800	1291	nuclease	N/A
Dbp3		58.84		9.73		20.84		13426	7719	shared	N/A
Has1	DDX18	56.73	75.40	9.88	9.52	23.98	31.04	12037	4084	shared	N/A
Prp43/Ja1	DHX15	87.54	90.93	6.38	7.12	39.11	40.13	8852	3735	shared	N/A
Rrp5	PCDCD11	193.11	208.70	6.07	8.99	32.97	26.56	15799	9124	shared	N/A
Cbf5	DKC1	54.71	57.67	9.34	9.46	29.81	36.19	18964	8567	snoRNP	N/A
Gar1	GAR1	21.50	22.34	11.99	10.91	37.56	31.34	19463	10558	snoRNP	N/A
Nhp2	NHP2	17.13	17.20	10.45	8.48	35.26	50.33	16956	6672	snoRNP	N/A
Nop1/Lot3	FBL	34.48	33.78	10.85	10.18	59.33	44.24	31821	28378	snoRNP	N/A
Nop10	DKCB1	6.64	7.70	10.78	10.01	63.79	42.19	14475	10128	snoRNP	N/A
Nop56/Sik1	NOP56	56.87	66.04	9.43	9.24	28.57	44.78	30862	19939	snoRNP	N/A
Nop58/Nop5	NOP58	56.96	59.57	9.55	9.03	44.23	36.86	27653	9644	snoRNP	N/A
Snu13	SNU13	13.57	14.17	8.04	8.72	18.25	20.31	50898	59087	snoRNP	N/A
Csm1		21.74		4.82		8.95		2655	2061	rDNAP	N/A
Fob1		65.30		6.1		39.58		1734	1079	rDNAP	N/A
Hmo1		27.54		9.11		47.56		16564	14005	rDNAP	N/A
Lrs4		39.36		10.21		62.54		1473	917	rDNAP	N/A
Top1		90.00		9.22		34.98		5012	3280	rDNAP	N/A
Top2		164.21		7.04		27.94		5856	4193	rDNAP	N/A

Table S2 (contd.)

Master List of Assembly Factors and Their Physical Properties											
name	name	MW (kDa)	MW (kDa)	IEP	IEP	predicted disorder (%)	predicted disorder (%)	abundance mean	abundance STD	subset	tripartite domain
Rrn3		72.34		4.34		27.43		469	554	RRN	N/A
Rrn5		41.71		6.85		47.38		811	568	RRN	N/A
Rrn6		102.01		4.81		34		542	739	RRN	N/A
Rrn7		60.36		8.25		12.06		1192	1122	RRN	N/A
Rrn9		42.79		5.02		41.92		638	516	RRN	N/A
Rrn10		16.49		4.85		27.59		1408	1842	RRN	N/A
Rrn11		59.24		5.85		36.29		706	456	RRN	N/A
Rpa12								10655	5328	RPA	N/A
Rpa14								7820	10363	RPA	N/A
Rpa34								7722	3534	RPA	N/A
Rpa43								7471	3109	RPA	N/A
Rpa49								9948	3503	RPA	N/A
Rpa135								12623	6060	RPA	N/A
Rpa190								16112	8843	RPA	N/A
Rpb1								16726	11053	RPB	N/A
Rpb2								14925	6189	RPB	N/A
Rpb3								8911	4777	RPB	N/A
Rpb7								6897	3986	RPB	N/A
Rpb8								8439	4971	RPB	N/A
Rpb9								5395	2749	RPB	N/A
Rpb11								10860	6845	RPB	N/A
Ret1								4745	2562	RPC	N/A
Rpc17								3431	1714	RPC	N/A
Rpc19								15166	16514	RPC	N/A
Rpc37								4592	3392	RPC	N/A
Rpc40								15108	4005	RPC	N/A
Rpc53								3911	1354	RPC	N/A
Rpc82								4435	2123	RPC	N/A
HHF1								73439	141532	histone	N/A
HHF2								119590	176858	histone	N/A
HHT1								64338	101323	histone	N/A
HHT2								57624	99029	histone	N/A
HTA1								20958	21428	histone	N/A
HTA2								64804	39933	histone	N/A
HTB1								197857	267074	histone	N/A
HTB2								117817	122347	histone	N/A

¹ DExD/H-box proteins [#] RNA-binding proteins + brix domain % methyl transferases * WD repeats - HEAT repeats & ARM repeats ^ GTPases

Table S2 Master List of Nucleolar Assembly Factors and their Physical Properties

The listed AFs are strongly concentrated in the nucleolus and are essentially absent from the nucleoplasm and cytoplasm [1]. The shaded columns correspond to yeast values while the non-shaded columns correspond to human values. The following AFs were not included: Pfa1/Sqs1, Rrp36, Sen1/Cik3 and Utp25.

Table S3. Physical Properties of Subgroups of Assembly Factors

Motif Groupings	Subunit	MW	IEP
ARM Motif	Ipi1	LSU	38
	Nop9	pre-40S	78
	Puf6	LSU	75
	Sgd1	LSU	103
	Utp20	SSU	287
	mean +/- S.D.		116 +/- 88
Brix Domain	Brx1	LSU	33
	Imp4	5'-ETS	33
	Rpf1	LSU	35
	Ssf1	LSU	52
	Ssf2	LSU	52
	mean +/- S.D.		41 +/- 9
DExD/H-box	Dbp3	pre-40S & LSU	59
	Dbp4/Hca4	pre-40S	87
	Dbp6	LSU	70
	Dbp7	LSU	83
	Dbp8	SSU	47
	Dbp9	LSU	68
	Dbp10	LSU	113
	Dhr1	SSU	145
	Dhr2	SSU	83
	Drs1	LSU	84
	Fal1	SSU	45
	Has1	pre-40S & LSU	57
	Mak5	LSU	87
	Prp43	pre-40S & LSU	88
	Rok1	pre-40S	63
	Rrp3	SSU	55
	Spb4	LSU	69
	mean +/- S.D.		77 +/- 24
GTPases or putative GTPases	Bms1	SSU	135
	Efl1	pre-LSU	124
	Lsg1	pre-LSU	73
	Nog1	pre-LSU	74
	Nog2	pre-LSU	55
	Nug1	pre-LSU	58
	mean +/- S.D.		80 +/- 32
HEAT Repeat	Noc2	LSU	81
	Noc3	LSU	75
	Noc4/Utp19	SSU	63
	Rrp12	SSU	137
	Sda1	LSU	87
	Utp10	5'-ETS	200
	mean +/- S.D.		107 +/- 47
Methyl Transfer	Bmt2	LSU	39
	Bud23	SSU	32
	Dim1	SSU	36
	Emg1	pre-40S	28

Table S3. Physical Properties of Subgroups of Assembly Factors

Motif Groupings	Subunit	MW	IEP
RNA-binding Proteins (Motifs)	Nop1	snoRNP/5'-ETS	34
	Nop2	LSU	70
	Rcm1	LSU	56
	Rrp8	LSU	46
	Spb1	LSU	96
	mean +/- S.D.	49 +/- 20	9.0 +/- 1.7
WD Repeat Proteins	Dim2/Pno1 (KH)	pre-40S	30
	Esf2 (RRM)	pre-40S	36
	Imp3 (S4 domain)	5'-ETS	21
	Krr1 (KH)	pre-40S	37
	Mrd1 (RRM)	pre-40S	101
	Nop4 (RRM)	LSU	77
	Nop6 (RRM)	pre-40S	25
	Nop8 (RRM)	LSU	57
	Nop9 (pumilio)	pre-40S	77
	Nop12 (RRM)	LSU	51
	Nop13 (RRM)	LSU	46
	Nop15 (RRM)	LSU	25
	Nsr1 (RRM)	pre-40S	44
	Puf6 (pumilio)	LSU	75
	mean +/- S.D.	50 +/- 23	8.7 +/- 2.0
snoRNP Proteins	Enp2	pre-40S	82
	Erb1	LSU	92
	Ipi3	LSU	62
	Mak11	LSU	53
	Rrb1	LSU	52
	Rrp9	5'-ETS	65
	Rsa4	LSU	57
	Sof1	5'-ETS	57
	Utp1	5'-ETS	103
	Utp4	5'-ETS	88
	Utp5	5'-ETS	72
	Utp7	5'-ETS	62
	Utp12	5'-ETS	106
	Utp13	5'-ETS	91
	Utp15	5'-ETS	57
	Utp17	5'-ETS	101
	Utp18	5'-ETS	66
	Utp21	5'-ETS	104
	Ytm1	LSU	51
	mean +/- S.D.	75 +/- 19	6.6 +/- 1.9
snoRNP Proteins	Cbf5	pre-40S	54
	Gar1	pre-40S	21
	Nhp2	pre-40S	17
	Nop1	snoRNP/5'-ETS	34
	Nop10	snoRNP/pre-40S	6
	Nop56/Sik1	snoRNP/5'-ETS	56
	Nop58	snoRNP/5'-ETS	57

Table S3. Physical Properties of Subgroups of Assembly Factors			
Motif Groupings	Subunit	MW	IEP
Snu13 (U3)	snoRNP/5'-ETS	13	8.0
mean +/- S.D.		32 +/- 19	10.0 +/- 1.1

Table S3 Physical Properties of Subgroups of Assembly Factors

The quadrants illustrated in Figure 3B/C are differentially enriched in subsets of AFs.

This table lists the physical properties of the relevant groups.

Table S4A. Isoelectric Point and Predicted Disorder Extremes						
	most acidic (<25 th percentile)		most basic (>75 th percentile)		most disordered (> 60%)	
	yeast AFs	human AFs	yeast AFs	human AFs	yeast AFs	human AFs
5'-ETS	(< 4.72)	(< 5.81)	(> 9.69)	(> 9.24)	(> 60%)	(> 60%)
	<i>Pwp2</i>	<i>PWP2</i>	<i>Utp15</i>	<i>DCAF13</i>	<i>Mpp10</i>	<i>DNTTIP2</i>
	<i>Utp18</i>	<i>WDR75</i>	<i>Sof1</i>	<i>IMP4</i>	<i>Fcf2</i>	<i>UTP11L</i>
	<i>Utp5</i>	<i>UTP3</i>	<i>Utp7</i>	<i>IMP3</i>	<i>Utp11</i>	<i>UTP3</i>
	<i>Utp9</i>	<i>WDR43</i>	<i>Fcf2</i>	<i>WDR46</i>	<i>Bud21</i>	
	<i>Sas10</i>	<i>MPHOSPH10</i>	<i>Imp3</i>	<i>UTP11L</i>		
	<i>Mpp10</i>		<i>Utp11</i>			
pre-40S	(< 6.53)	(< 6.91)	(> 9.93)	(> 9.49)	(> 60%)	(> 60%)
	<i>Rio1</i>	<i>NOP9</i>	<i>Krr1</i>	<i>NGDN</i>	<i>Lcp5</i>	<i>KRI1</i>
	<i>Mrd1</i>	<i>UTP14C</i>	<i>Faf1</i>	<i>SRFBP1</i>	<i>Nsr1</i>	<i>NGDN</i>
	<i>Ecm16</i>	<i>EIF4A3</i>	<i>Utp30</i>	<i>RRP7A</i>	<i>Nop6</i>	<i>UTP14C</i>
	<i>Noc4</i>	<i>RBM19</i>	<i>Efg1</i>	<i>DDX52</i>	<i>Efg1</i>	<i>UTP14A</i>
	<i>Sgd1</i>	<i>BMS1</i>	<i>Dim1</i>	<i>PNO1</i>	<i>Esf2</i>	
	<i>Kri1</i>	<i>RIOK1</i>	<i>Dbp8</i>	<i>KRR1</i>	<i>Utp14</i>	
	<i>Utp14</i>	<i>KRI1</i>	<i>Rrt14</i>	<i>UTP23</i>	<i>Faf1</i>	
	<i>Nsr1</i>	<i>AATF</i>	<i>Slx9</i>	<i>RSL1D1</i>	<i>Rrt14</i>	
	<i>Enp1</i>	<i>NCL</i>	<i>Utp23</i>		<i>Fyv7</i>	
	<i>Bfr2</i>		<i>Nop6</i>			
			<i>Fyv7</i>			
pre-LSU	(< 5.48)	(< 6.09)	(> 9.85)	(> 9.93)	(> 60%)	(> 60%)
	<i>Rrp15</i>	<i>URB1</i>	<i>Nop15</i>	<i>RSL24D1</i>	<i>Nop4</i>	<i>NOP2</i>
	<i>Drs1</i>	<i>BMT2</i>	<i>Nop13</i>	<i>RPF2</i>	<i>Rrp15</i>	<i>RBM28</i>
	<i>Rix1</i>	<i>CEBPZ</i>	<i>Ssf1</i>	<i>DDX31</i>	<i>Fpr4</i>	<i>PELP1</i>
	<i>Nop7</i>	<i>WDR12</i>	<i>Rrp17</i>	<i>EBNA1BP2</i>	<i>Ebp2</i>	<i>FTSJ3</i>
	<i>Rix7</i>	<i>NOC2L</i>	<i>Nop53</i>	<i>RBM34</i>	<i>Rlp24</i>	<i>RRP1</i>
	<i>Rrp1</i>	<i>MDN1</i>	<i>Rpf1</i>	<i>RSL1D1</i>	<i>Fpr3</i>	<i>RRP15</i>
	<i>Mak16</i>	<i>RRP15</i>	<i>Rrp8</i>	<i>NOL12</i>	<i>Loc1</i>	<i>ZNF93</i>
	<i>Erb1</i>	<i>DNAJC21</i>	<i>Nop12</i>	<i>NSA2</i>	<i>Cgr1</i>	<i>NOP53</i>
	<i>Nop2</i>	<i>MAK16</i>	<i>Ipi1</i>	<i>NOP53</i>		<i>NOL12</i>
	<i>Rea1</i>	<i>GRWD1</i>	<i>Rlp7</i>	<i>RRS1</i>		
	<i>Mak21</i>	<i>EIF6</i>	<i>Rrs1</i>			
	<i>Rsa3</i>	<i>PELP1</i>	<i>Tri1</i>			
	<i>Fpr4</i>		<i>Rlp24</i>			
	<i>Tif6</i>		<i>Bmt2</i>			
	<i>Rrb1</i>		<i>Nsa2</i>			
	<i>Sqt1</i>		<i>Loc1</i>			
	<i>Fpr3</i>		<i>Cgr1</i>			

Table S4B Outliers from the Comparison of Yeast vs Human						
	isoelectric point outlier		predicted disorder outlier		molecular weight outlier	
	yeast homolog	human homolog	yeast homolog	human homolog	yeast homolog	human homolog
5'-ETS	Fcf2	DNTTIP2	Sof1	DCAF13	Utp10	HEAR1
	Utp18	UTP18			Fcf2	DNTTIP2
pre-40S	Utp14	UTP14A	Nop9	NOP9	Utp30	RSL1D1
	Fal1	EIF4A3	Utp30	RSL1D1	Utp20	UTP20
	Enp1	BYSL			Nsr1	NCL
pre-LSU	Ebp2	EBNA1BP2	Rrp1	RRP1	Rix1	PELP1
	Drs1	DDX27	Rix1	PELP1	Nug1	NUGGC
	Fpr3	FPR3	Fpr3	FPR3	Urb2	URB2
	Rrp1	RRP1			Urb1	URB1
	Bmt2	BMT2			Rea1	MDN1
	Nop2	NOP2				

Table S4 Extreme Values of Physical Parameters of Assembly Factors - related to

Figure 2

(A) Isoelectric point and predicted disorder extremes for principal subsets (5'-ETS, pre-40S, pre-LSU) in both yeast and man. AFs are listed from top-down in order of increasing acidity for the most acidic and increasing basicity for the most basic. The entries for predicted disorder are organized in comparable fashion. The italicized AFs are those for which both yeast and human homologs have extreme values. The AFs that appear in multiple lists of extremes are highlighted.

(B) Outliers from the comparison of yeast vs human principal groups of AFs (5'-ETS, pre-40S, pre-LSU). Homolog pairs are listed from top-down in order of increasing difference between yeast and human values. The AFs that appeared in multiple lists of outliers are highlighted.

IQR Rule

The interquartile range (IQR) was calculated by subtracting the first quartile (Q_1) from the third quartile (Q_3) in each set of AF parameters. The boundaries for low and high outliers were then determined using the IQR rule (Equation 1, Equation 2).

$$\text{low outliers} < Q_1 - 1.5 * \text{IQR} \quad \text{Eq. 1}$$

$$\text{high outliers} > Q_3 + 1.5 * \text{IQR} \quad \text{Eq. 2}$$

Table S5.

Assembly Factors with Shared Motif Signatures

Assembly Factors with Shared Motif Signatures						
	Signature Name, Activity	Color code	Prototypes	Number of Motifs in Prototype	Proteins with Variant Motif Signatures	Larger Grouping
1	DExD/H-box	●	Dbp7, [Dbp8], Dbp9	10	Dbp3, Dbp4, Dpb6, Dpb10, [Dhr1], Dhr2, Drs1, Fal1, [Has1], Mak5, [Prp43], Rok1, [Rrp3], Spb4	NTPases
2	GTPase/AAA ATPase	●	Rea1	3	Bms1, [Fun12c], Lsg1c, Nog1, Nog2/Nug2, Nug1, [Sen1]	
3	ATPase	●	Drg1c, Rix7	8		
4	snoRNP	●	Nop56, Nop58	9		
5	HMG-like	●	Nhp2, Snu13	7	(Cic1/Nsa3)	snoRNP proteins
6	Gar1	●	Gar1	1		
7	RRM	●	Mrd1, Nop4, Nop12, Nop13, Nop15, [Nsr1]	5	Esf2, Nop6, Nop8, (Rrp7)	RNA-binding
8	Brix domain	●	Ssf1, Ssf2	4	Brx1, Imp4, Rpf1, Rpf2	
9	Methyl Transferases	●	Nop1	3	Bmt2, Bud23, [Dim1], Emg1, Nop2, Rcm1, Rrp8, Spb1, [YGR283C]	
10	KH domain	●	Dim2, Krr1	3		
11	ARM/HEAT	●	Ipi1, Nop9, Puf6, Rrp12, Sda1, Sgd1, Utp10, Utp20	1	Mak21/Noc1, Noc3, (Noc4/Utp19)	Structural
12	WD repeat and WD-like	●	Nan1/Utp17, Rrp9, Utp4, Utp18	6	Enp2, Erb1, (Esf1), Ipi3, Mak11, (Nsa1), Rrb1, (Rrp5), Rsa4, Sof1, Utp1, (Utp5), Utp6, Utp7, (Utp9), Utp12, Utp13, Utp15, Utp21, Ytm1	
13	Prolyl isomerase	○	Fpr3, Fpr4	8		Other
14	Zn-finger	●	Bud20, Jjj1	5	Rei1	
15	Kinases	○	Rio2c	5	Rio1c	
16	PUA domain	●	Cbf5*, Nip7	3		Various
17	GTP-binding	●	Efl1c	10		
18	Unknown	●	Utp3	2	Lcp5	
19		●	Sdo1	1		

Assembly Factors without Shared Motif Signatures

Alb1, **Arx1**, **Bfr2**, **Bud21**, **Bud22**, **Cgr1**, **Ebp2**, **Efg1**, **Enp1**, **[Fcf2]**, **[Fvy7]**, **Imp3**, **Kre33**, **Kri1**, **Las1**, **Loc1**, **Ltv1c**, **Mak16**, **Mpp10**, **Mrt4**, **Ngl2**, **[Nob1]**, **Noc2**, **Nop7**, **Nop16**, **Npa1/Urb1**, **Npa2**, **Nsa2**, **Pfa1/Sqs1c**, **Rcl1**, **Rix1**, **Rlp7**, **Rlp24**, **Rnt1**, **Rrp1**, **Rrp6**, **Rrp15**, **Rrs1**, **Rrt14**, **Rsa3**, **[Slx9]**, **Snm1**, **Tif6**, **Tri1**, **Utp2**, **Utp8**, **Utp11**, **Utp14**, **Utp22**, **[Utp24/Fcf1]**, **Yvh1**.

Color scheme: Green for 5'-ETS, Blue for pre-40S, Red for pre-18S.

Motif signatures under consideration are listed by the *Saccharomyces* Genome Database. If a motif is found in only a single AF, that AF is considered to lack shared motifs.

Variant motif signatures can have more or fewer of the motifs by comparison to the prototypes.

Variant motif signatures can have more or fewer of the motifs by comparison to the primary motif. Entries in parentheses are marginally related to the groups in which they are included.

Entries in parentheses are marginally-related to the groups in which they are included.

* Cbf5 is the pseudouridine synthase of H/ACA snoRNPs. These proteins have been recovered in association with the pre-40S segment of rRNA, unlike Nop1 and other box C/D snoRNP proteins that are recovered with the 5' ETS [6, 7].

The suffix *c* signifies that the AE localizes largely to the cytoplasm.

@ Low-stringency BLAST searches detect the following relations: Mrt4 with Rpp0, Tri1 with Uaf30 and Enp1, Fcf1/Utp24 with Utp23, and Tfcf1 with Ufcf1, Blp7 with Blp24, and Tfcf1 with Blp17 and Blp124 [6], and Blp1 with Blp11.

The colors indicate the sequence of rRNA with which the AEs are associated. Green = 5'ETS; Blue = 16S; Red = 18S; Purple = 28S.

Table S5 Hierarchic Clustering Motif Signatures - relevant to Figure 3D

AFs with motif(s) that are shared with at least one other AF were tabulated and assigned to 19 groups ("motif signatures"). The lower part of the table lists the AFs that do not share motif signatures with others.

RNA-Binding Proteins Associated with rRNA Domains														
	5'-ETS	pre-40S domains			ITS 1	pre-LSU domains								
RNA domain	ETS	5' Domain	Central	3', Major	3', Minor	ITS1	ITS2-Ssf1	D1	D2	D3	D4	D5	D6	late
Imp3 (10.3) RRM, KH, S4 or pumilio	Esf2 (8.4)	Krl1 (4.7) <u>Krr1</u> (10.1) Nsr1 ^{In} (4.6)	Mrd1 (6.5) Nop6 (10.9) Nop9 ^{In} (8.4)	Pno1 (9.7)	Rrp5 (6.1)		Nop12 (10.1) Nop15 (9.9) Rrp5 (6.1)	Nop4 (9.7)		Puf6 (6.9)		<u>Nop8</u> ^F (5.7)	Nop13 (9.9)	
		Dbp4 (7.5) Dbp8 (10.2)	Rok1 (9.7)		Dhr1 (6.3) <u>Dhr2</u> (9.2)		Dbp7* (9.8) Dbp9* (9.7) Dbp10 (9.7)	Has1 (9.9)		Drs1 (5.4)				
Imp4 (9.5) Brix domain							Ssf1 (10.0) Ssf2 (9.7)	Brx1 (9.8)	Rpf1 (10.1)			Rpf2 (9.6)		
Me Transferase			Emg1 (8.6)	Rrp8 (10.1)								<u>Nop2</u> (4.7)		
snoRNP proteins	Nop1 (10.8) <u>Nop5</u> 6 (9.4) <u>Nop5</u> 8 (9.5) <u>Snu1</u> 3 (8.0)	Cbf5 (9.3) Gar1 (12.0) <u>Nhp2</u> (10.4) Nop10 (10.8)												

Derived from experiments in which AFs have been recovered along with 3'-truncated segments of rRNA [6, 7, 9, 10].
The superscripts (In, F) designate AFs that belong to the SSU-In or LSU-F subsets.
IEP values from the *Saccharomyces* Genome Database are indicated in parentheses.
The underlined AFs interact with Nop1 on the basis of biochemical and genetic experiments, as is summarized in the *Saccharomyces* Genome Database.

Table S6 Assembly Factors Associated with Successive rRNA Domains - related to Table 3, Figure 4A.

The RNA-binding proteins listed were identified in experiments using 3'-truncated rRNAs that were expressed in yeast, retrieved from lysates and analyzed, as described in the text. Their IEPs are indicated in parentheses. Many additional AFs are also known to contact rRNA judging from cross-linking studies and cryo-EM studies. The column entitled ITS2-Ssf1 corresponds to a very early LSU precursor.

Table S7. Subsets of Assembly Factors in More Primitive Cell Types

Subset of AFs	Orthologs in LECA *	Orthologs in Archaea	Candidates in <i>E. coli</i>
SSU-F	Dbp8, Dhr2, Fyv7, #Nan1/Utp17, Noc4, Rrp7, #Utp5, #Utp6, #Utp13, #Utp15, Utp25		ARB1, BUD23, CDC14, CFD1, DBP5, DBP8, DED1, DIM1, DIS3
SSU-In	Efg1, Nsr1		DRS1, FPR3, Kre33, MDN1, MTR4, NOP2,
LSU-F	Dbp6, Npa1/Urb1, Npa2/Urb2		RCL1, REX2, RIX7, RLI1, RNT1, ROK1, RRP5, RRP6, SKI6
LSU-Ou	Brx1, Cic1, Mak11, Mrt4, Noc2, Noc3, Nop4, Nop7, Nop13, Nop15, Nop16, Puf6, Rrp1, Ytm1, Ssf1	Nog1, Rix7, Tif6	
snoRNP	Gar1, Nhp2, Nop56/Sik1, Nop58	Gar1, Nop1, Nop10, Nop56/Sik1	
shared	Dbp3, Prp43		

Derived from [11]. * We have included only those AFs that we have localized. # These AFs associate with the 5-ETS in yeast: Utp5, Utp6, Utp13, Utp15, Nan1/Utp17.

Table S7 Assembly Factors in Evolutionary Context - related to Table 1

Comparison of each subunit-specific subset with the likely repertoire of AFs in more primitive organisms. Based on [11].

Supplementary Figures and Legends

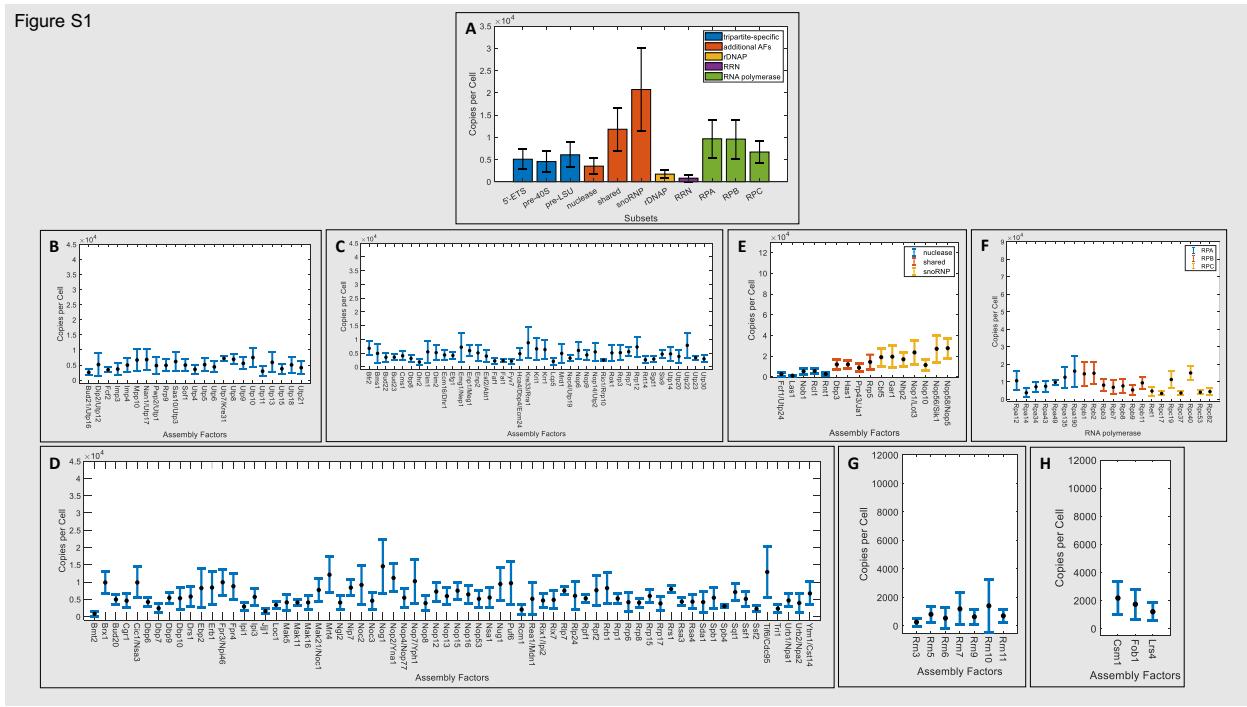


Figure S1 Copy Number per Cell of Single Assembly Factors - related to Figure 2

(A) Copy number estimate bar graphs. Mean values and standard deviations for single AFs as well as proteins associated with rDNA, and RNA polymerases. See Figure S1 and Table S4 for detail. Outlier values for each AF (based on the IQR rule - see Methods) and outlier AFs for each subset (due to unreasonably high standard deviations) were removed before these calculations were made. The following AFs were removed: Nsr1/She5, Nsa2, Hmo1, Top1 and Top2.

The representation is as in Figure 2 but concerns single proteins of **(B) 5'-ETS**, **(C) pre-40S**, **(D) pre-LSU**, **(E) additional AFs**, **(F) RNA polymerase**, **(G) Rrn proteins**, **(H) rDNAPs**. Outlier values for each AF were removed from consideration (Nsr1/She5, Nsa2, Hmo1, Snu13, Top1 and Top2). Nuclease: nucleases that cleave rRNA, Shared:

proteins needed to make both SSU and LSU, RPA/B/C: subunits of RNA polymerase A/B/C, Rrn: regulators of RNA polymerase A, rDNAPs: proteins that colocalize with the rDNA filament. For identification of the subsets of AFs, see Table S1.

Figure S2

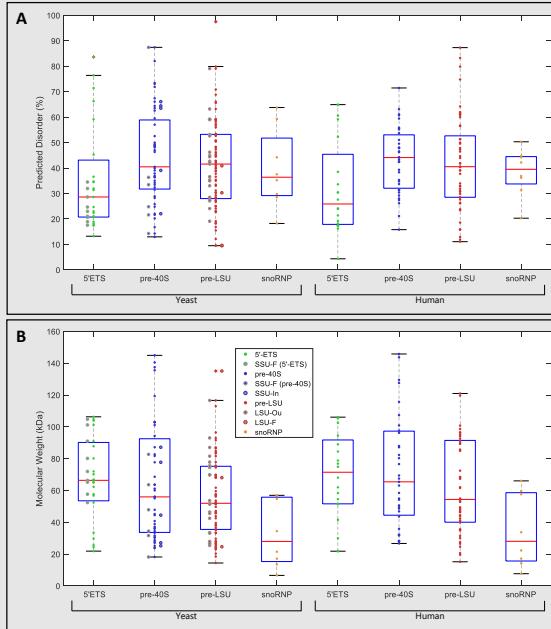


Figure S2 Predicted Disorder and Molecular Weight of Yeast and Human Assembly Factors

Assembly factors were grouped - as in Figure 3A - to illustrate the characteristics of those that associate with the 5'-ETS, the pre-40S segment, or the pre-LSU segment. In each case, the central column of values includes all relevant AFs listed in Table S2. For the 5'-ETS, the points to the left are SSU-F. For the pre-40S, the points to the left are SSU-F and the points to the right are SSU-In. For the pre-LSU, the points to the left are LSU-Ou and the points to the right are LSU-F. Human AFs are subgrouped according to the information for their yeast homologs.

Figure S3

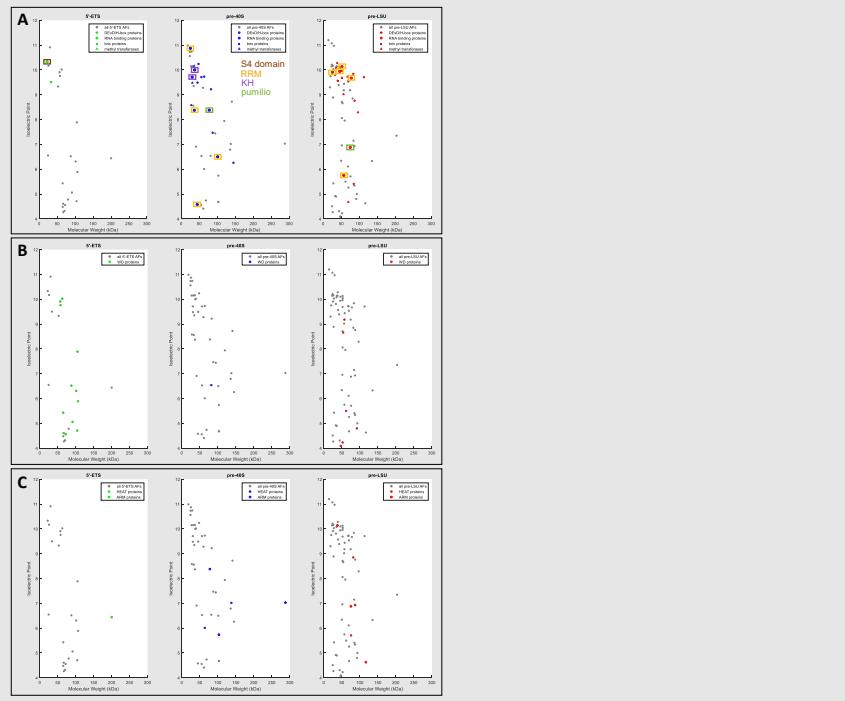


Figure S3 Overlay Representation of Physical Properties for Subsets of Assembly Factors - related to the 2-dimensional plots of Figure 3B/C

All AFs present in the each of the tripartite domains are shown in gray.

(A) RNA-associated proteins. The AFs in the upper quadrants include those with RNA-binding motifs (RRMs, KH and pumilio domains). They have a mean IEP of pH 8.5 and mean size of 64 kDa (pH 8.5, 64 kDa), DExD/H-box proteins (pH 8.7, 77 kDa), the methyl transferases (pH 9.0, 49 kDa), and Brix domain proteins (pH 9.8, 41 kDa). snoRNP proteins, although not illustrated, have the following mean values (pH 10.0, 32 kDa).

(B) WD-repeat proteins. The AFs with WD repeats are found mostly in the lower two quadrants (pH 6.6, 54 kDa).

(C) HEAT/ARM-repeat proteins. The AFs with HEAT repeats (pH 6.8, 107 kDa) and ARM repeats (pH 7.1, 116 kDa) have broadly distributed IEPs.

Figure S4

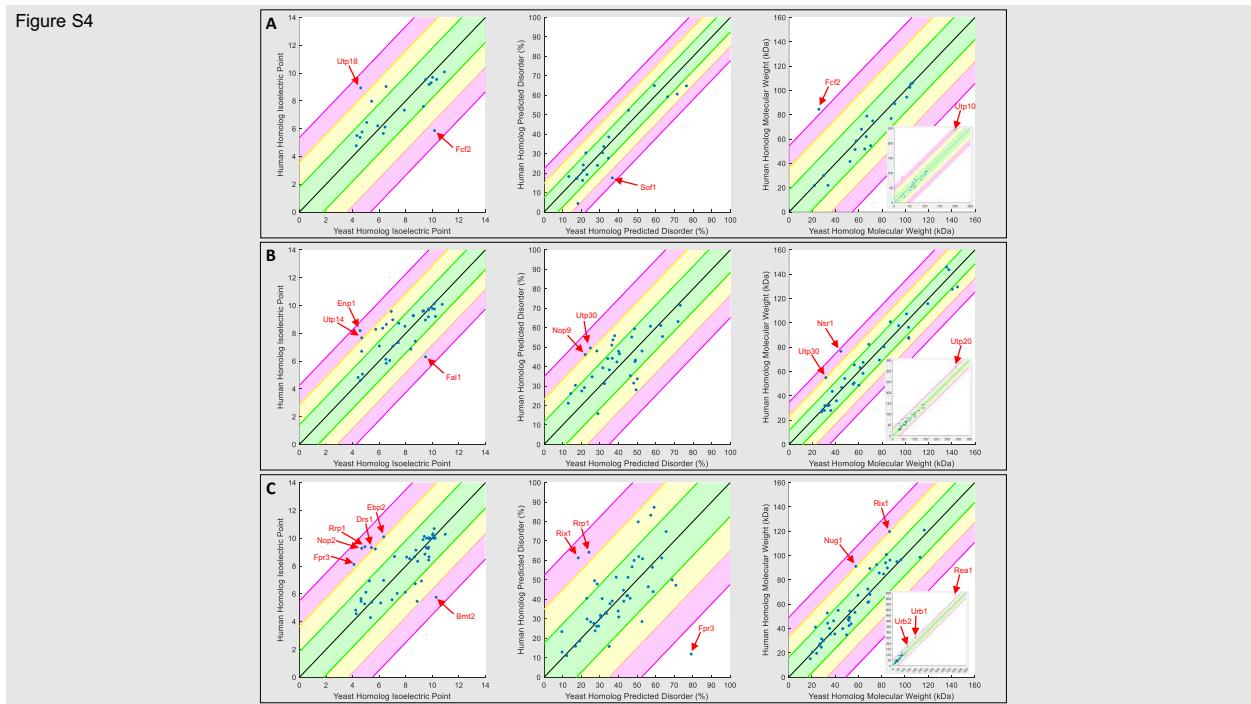


Figure S4 Yeast vs Human Comparison of Physical Properties of Assembly Factors - related to Figure 3

The comparative diagonal plots show isoelectric points, predicted disorder and molecular weights of **(A)** 5'-ETS, **(B)** pre-40S and **(C)** pre-LSU AFs. Colored margins indicate how far away yeast and human values are from each other, with green being 1 standard deviation, yellow being 2 standard deviations, and pink being 3 standard deviations. AF pairs were considered outliers if the difference between their yeast and human values (when compared to all AFs in their subset) was more than 2 standard deviations away from 0. These are indicated with red arrows using yeast names.

Figure S5

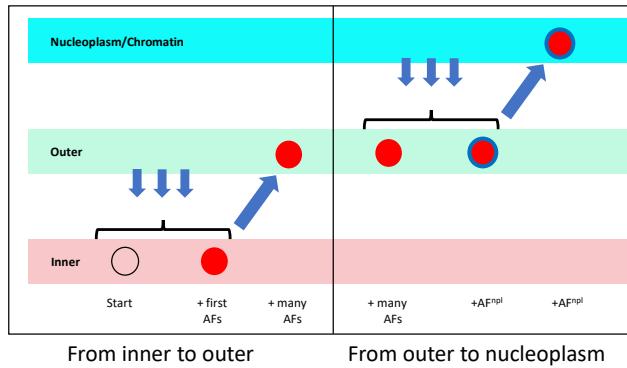


Figure S5 Vectorial 2-Phase Partitioning

On the left, in a typical T-diagram, we illustrate the progressive loading of AFs onto nascent rRNA within the inner compartment. At "start," no AFs have been added so the circle is empty. Due to arrival of latent AFs from the outer layer (vertical blue arrows) the nascent RNPs progressively acquire AFs (red filling) and becomes phase compatible with the outer layer/volume. The nascent particles therefore shift in an energetically downhill fashion to the outer layer/volume. At the right, the further maturation of precursor particles that reside in the outer layer. Since multiple AFs that are conspicuous in the nucleoplasm are also found in the outer compartment, we propose that when they arrive in the outer layer (blue arrows) they bind to the surface of immature particles (represented by the blue surface on the red particles). Those particles therefore can diffuse in energetically neutral fashion into the nucleoplasmic volume. Our previous article [1] introduces the use of T-diagrams.

Literature Cited

1. Tartakoff, A.M., et al., *The nucleolus as a polarized coaxial cable in which the rDNA axis is surrounded by dynamic subunit-specific phases*. Curr Biol, 2021. **31**(12): p. 2507-2519 e4.
2. Giaever, G., et al., *Functional profiling of the *Saccharomyces cerevisiae* genome*. Nature, 2002. **418**(6896): p. 387-91.
3. Bohnsack, K.E. and M.T. Bohnsack, *Uncovering the assembly pathway of human ribosomes and its emerging links to disease*. EMBO J, 2019. **38**(13): p. e100278.
4. Farley-Barnes, K.I., L.M. Ogawa, and S.J. Baserga, *Ribosomopathies: Old Concepts, New Controversies*. Trends Genet, 2019. **35**(10): p. 754-767.
5. Ho, B., A. Baryshnikova, and G.W. Brown, *Unification of Protein Abundance Datasets Yields a Quantitative *Saccharomyces cerevisiae* Proteome*. Cell Syst, 2018. **6**(2): p. 192-205 e3.
6. Zhang, L., et al., *Stepwise and dynamic assembly of the earliest precursors of small ribosomal subunits in yeast*. Genes Dev, 2016. **30**(6): p. 718-32.
7. Chaker-Margot, M., et al., *Stage-specific assembly events of the 6-MDa small-subunit processome initiate eukaryotic ribosome biogenesis*. Nat Struct Mol Biol, 2015. **22**(11): p. 920-3.
8. Espinar-Marchena, F.J., R. Babiano, and J. Cruz, *Placeholder factors in ribosome biogenesis: please, pave my way*. Microb Cell, 2017. **4**(5): p. 144-168.
9. Chen, W., et al., *Stepwise assembly of the earliest precursors of large ribosomal subunits in yeast*. Nucleic Acids Res, 2017.
10. Hunziker, M., et al., *Conformational switches control early maturation of the eukaryotic small ribosomal subunit*. Elife, 2019. **8**.
11. Ebersberger, I., et al., *The evolution of the ribosome biogenesis pathway from a yeast perspective*. Nucleic Acids Res, 2014. **42**(3): p. 1509-23.