Supporting Information

Lysozyme is sterically trapped within the silica cage in Bioinspired Silicalysozyme Composites: a Multi-technique understanding of elusive Proteinmaterial Interactions

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NMR characterization

Solid-state NMR experiments were recorded on a Bruker Avance II spectrometer operating at 700 MHz ¹H Larmor frequency (16.4 T), corresponding to 139 MHz ²⁹Si Larmor frequency and 176 MHz ¹³C Larmor frequency. The spectrometer is equipped with a 3.2 mm BVT MAS probehead in double resonance mode.

The 90° pulse duration on ²⁹Si and ¹³C for all the analyzed samples were set to 7.5 ms and 3.5 ms respectively. The durations of 90° pulse on ¹H were different for the different samples (see Table S1a).

The 1D ²⁹Si spectra in direct excitation were acquired with CPMG echo train acquisition, and then the 24 echoes were separated. Only the first decay has been taken as quantitatively relevant, because the transverse relaxation times are different for the different sites. Interscan delays were set to 3 times the ²⁹Si T₁ of the slowest-relaxing species (Q⁴), in order to make the experiments quantitative.

For the 2D {¹H}-¹³C and {¹H}-²⁹Si HETCOR experiments, cross-polarization was achieved by matching the k = 1 Hartmann–Hahn condition. The spectral windows for the different nuclei were 60, 248 and 249 ppm for ¹H, ¹³C and ²⁹Si, respectively. During the ¹H magnetization evolution under the chemical shift in the indirect dimension of heteronuclear correlation experiments, the PMLG decoupling sequence was used to suppress ¹H–¹H dipolar couplings.

In these experiments, interscan delay was set to 1.3 T_1 of the ¹H relaxation time, shown in Table S1b.

CPMG echo train acquisition has been employed also for ${^{1}H}-{^{29}Si}$ HETCOR in the direct dimension, and the 24 echoes in each FID were coadded.

Freeze-dried sample	Rehydrated	Rehydrated	Washed	Washed
	with H₂O	with D₂O	with GnHCI	with NaCl
2.90	2.90	3.17	2.97	3.17

Table S1a. Duration of the ¹H 90° pulse in HETCOR experiments, in µs.

Freeze-dried sample	Rehydrated	Rehydrated	Washed	Washed
	with H₂O	with D₂O	with GnHCI	with NaCl
3.0	2.5	0.8	0.8	3.0

Table S1b. Interscan delay in HETCOR experiments, in seconds.

Small angle X-ray scattering (SAXS) measurements

SAXS measurements were performed on the as-prepared aqueous dispersions of the composite before and after treatment with GnHCl and DTT or washing with NaCl. Scattering profiles were collected using a Xeuss 3.0 HR apparatus (Xenocs, France), equipped with an EIGER2R (1M model) hybrid pixel photon counting detector (Dectris Ltd., Switzerland) consisting of 1,028 x 1,062 pixels with a size of 75 x 75 μ m², free to move on a rail in a chamber vacuum. The K α radiation (λ = 1.5406 Å) emitted by a Cu microfocus (30 W) sealed tube operated by the Genix 3D generator. Calibration of the sample to detector distance was performed against silver behenate (d = 58.376 Å).¹ Scattering curves were obtained in the 0.004 and 0.5 Å⁻¹ q-range by merging data collected with two different sample to detector distances (i.e. 0.3 and 1.8 m). All the samples were measured under vacuum in 1 mm thick quartz capillary tubes sealed with hot-melting glue. Scattering curves were converted in absolute intensity (mm⁻¹), by measuring the scattering from a 1.16 mm thick glassy carbon reference

specimen in the same experimental conditions.^{2,3} Finally the 1-D azimuthal averaged scattering patterns were reduced subtracting the scattering intensity from capillary + water and merging the curves obtained at the two sample-to-detector distances. Data reduction, normalisation and merging were performed by using XSACT (X-ray Scattering Analysis and Calculation Tool) software (Xenocs, France).

Scanning electron microscope (SEM) and energy-dispersive X-ray spectrometry microanalysis (EDS)

All the samples were investigated by a Σ IGMA high-resolution field-emission scanning electron microscope (FESEM, Carl Zeiss) based on the GEMINI column which features a high-brightness Schottky field emission source beam booster and in-lens secondary electron detector. Measurements were performed on uncoated samples mounted on graphite tape with an acceleration potential of 2 kV and at a working distance of about 4 mm.

Energy-dispersive X-ray microanalysis (EDS) analysis was performed using a silicon-drift detector (Oxford Instruments) coupled with SEM at a working distance of 8.5 mm and with an accelerating voltage of 10 kV.

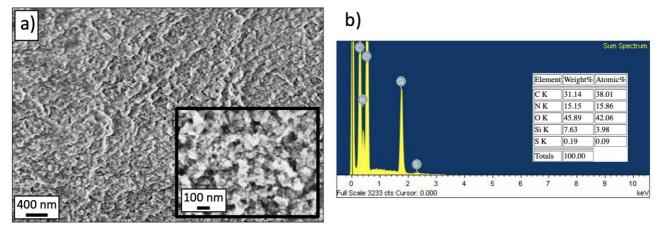


Figure S1. SEM micrographs of the composite after freeze drying at two different magnifications: 50 kX (a) and 200 kX (inset). EDS spectrum of the corresponding sample (b).

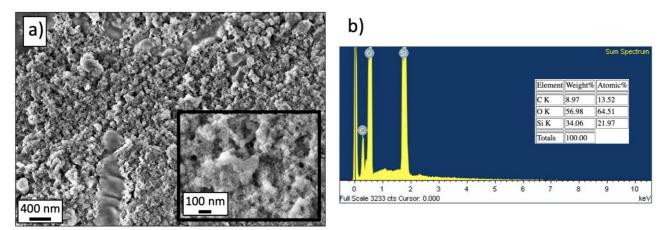


Figure S2. SEM micrographs of the composite after treatment with GnHCl and DTT at two different magnifications: 50 kX (a) and 200 kX (inset). EDS spectrum of the corresponding sample (b).

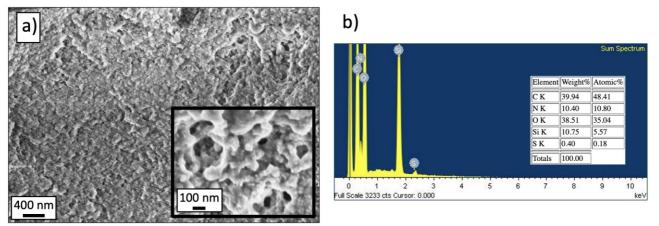
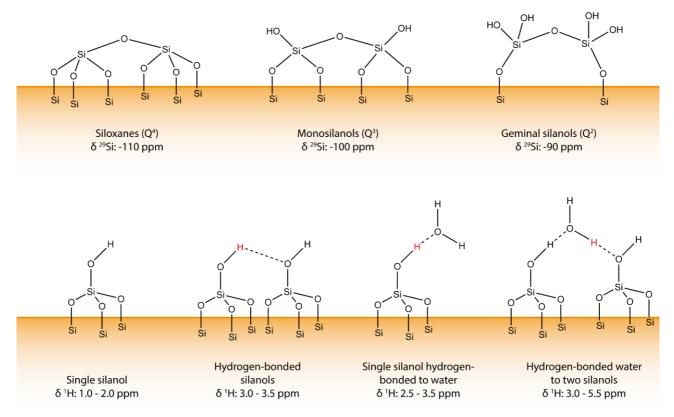


Figure S3. SEM micrographs of the composite after washing with NaCl (a) at two different magnifications: 50 kX and 200 kX (inset). EDS spectrum of the corresponding sample (b).



Scheme S1. Chemical shifts of ¹H and ²⁹Si species on silica.^{4–8}

Site	δ /ppm	σ/Hz	Relative Intensity	
Q^2	-93.5	3004	6.2%	
Q^3	-103.1	2216	37.5%	
Q^4	-112.0	2509	56.3%	

Table S2. Parameters from the Gaussian deconvolution of the direct excitation ²⁹Si spectrum of the composite.

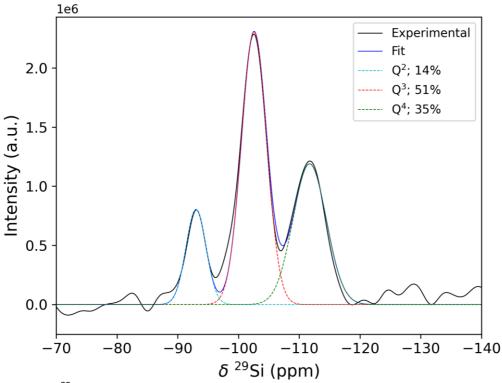


Figure S4. ²⁹Si direct excitation solid-state NMR spectra of silica obtained through condensation of the precursor without the protein. The signal has been deconvoluted using three Gaussian contributions corresponding to Q^2 , Q^3 and Q^4 sites.

δ /nnm	σ	Relative
0 /ppiii	/Hz	Intensity
-93.0	1155	14.1%
-102.6	1454	51.1%
-111.7	1921	34.8%
	-102.6	o /ppm /Hz -93.0 1155 -102.6 1454

Table S3. Parameters from the Gaussian deconvolution of the direct excitation ²⁹Si spectrum of silica obtained through condensation of the precursor without the protein (Figure S4).

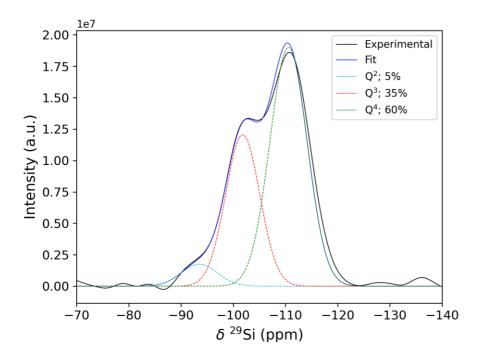


Figure S5. ²⁹Si direct excitation solid-state NMR spectrum of the composite treated with GnHCI and DTT.

Site	δ /ppm	σ /Hz	Relative Intensity
Q2	-93.5	2409	5.3%
Q3	-101.8	2297	34.8%
Q4	-110.6	2501	59.8%

Table S4. Parameters from the Gaussian deconvolution of the ²⁹Si direct excitation solid-state NMR spectrum of the composite treated with GnHCl and DTT (Figure S5).

SAXS analysis: Unified fit model

The Unified fit model was proposed by Beaucage in 1995.⁹ This approach describes scattering data as composed by multiple dimensional levels with different structural features. Each level is described by a Guinier and an associated Porod power-law regime. It is well known that the first derivation of the Beaucage model in which Guinier and Porod scale factors vary independently can introduce artefacts in the fitting and a detailed discussion on the limitations of this model are reported in the literature.¹⁰ However, this model can be used independently from the shape of the investigated system and for this reason has been widely used for the description of the scattering of randomly distributed structures as spheres, disks, rods and polymer coils.

The final fitting function then is composed by a series of structural levels. The scattering intensity of the level "i" is:

$$I_{i}(q) = G_{i}e^{-\frac{q^{2}R_{g}^{2}}{3}} + e^{-\frac{q^{2}R_{g(i-1)}^{2}}{3}}B_{i}\left\{\frac{\left[\operatorname{erf}\left(qR_{g}/\sqrt{6}\right]^{3}\right]}{q}\right\}^{P_{i}}$$
(S1)

where:

•
$$G = n^2 N_p I_e$$

" N_p " is the number of particles in the scattering volume; " I_e " is the scattering factor for a single electron; "n" is the number of electrons in a particle.

• $B = 2\pi N_p S_p \rho_e^2$

" ρ_e " is equal to n/V_p ; " S_p " is the surface area for the particle.

Data analysis was performed using the IRENA modelling package developed by Jan Ilavsky.¹¹

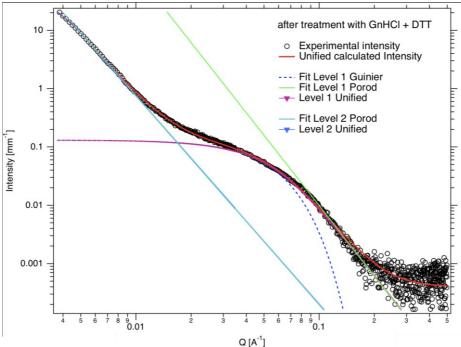


Figure S6. Experimental SAXS data of the sample after treatment with GnHCl + DTT fitted with equation (S1). The black markers are the experimental data, the red curve is the total unified fit, the blue dashed curve represents the Guinier component and green and cyan curves represent the power-law components corresponding to the first and second level, respectively.

	As-prepared	Treatment with GnHCI + DTT	Washing with NaCl
P (I level, fixed)	4	4	4
R _g (nm)	4.4 (1)	3.4 (1)	4.5 (1)
P (II level)	3.3 (1)	3.7 (1)	3.3 (1)
background	4.8·10 ⁻⁴	3.9.10-4	3.0.10-4
X ²	2690	1738	1531

Table S5. Parameters extracted from the modelling of SAXS pattern by Unified model. Values in parentheses are standard deviations on the last significant figures.

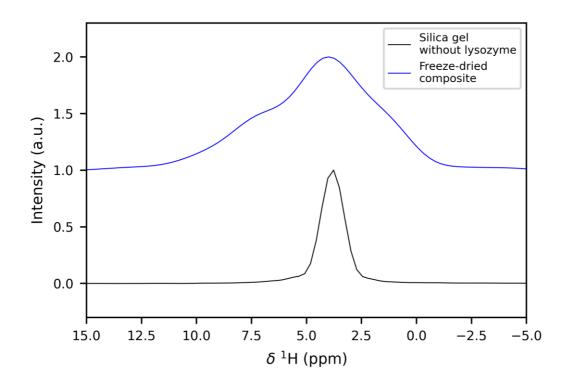


Figure S7. ¹H trace of the {¹H}-²⁹Si HETCOR acquired with 10 ms of CP contact time on the freezedried composite (blue) and on the freeze-dried silica gel obtained in the absence of the protein under otherwise similar conditions (black). The signals are integrated over all Si species.

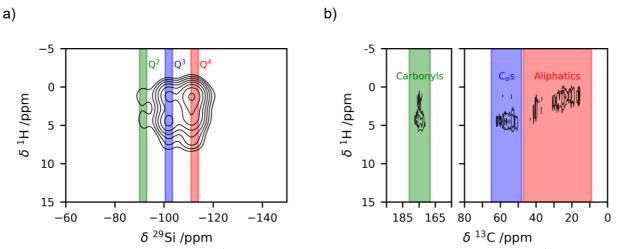


Figure S8. Integration range for ¹H traces over the heteronucleus dimension in {¹H}-X HETCOR spectra. X is ²⁹Si in the left panel and ¹³C in the right panel.

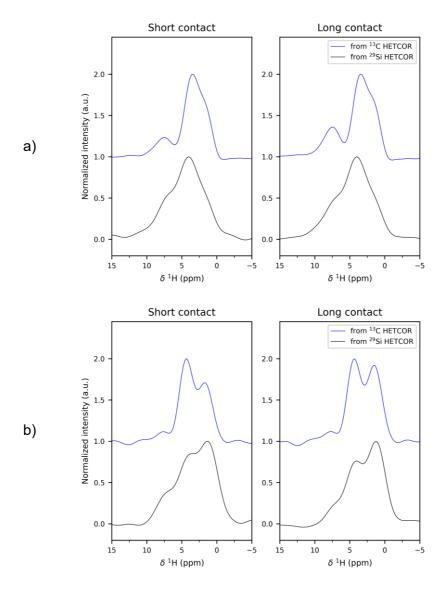


Figure S9. Comparison of the ¹H projection extracted from $\{^{1}H\}^{-13}C$ HETCOR (blue) and $\{^{1}H\}^{-29}Si$ HETCOR (black) spectra, at short (left panel) and long contact (right panel), in the dry sample (top) and in the rehydrated with D₂O sample (bottom).

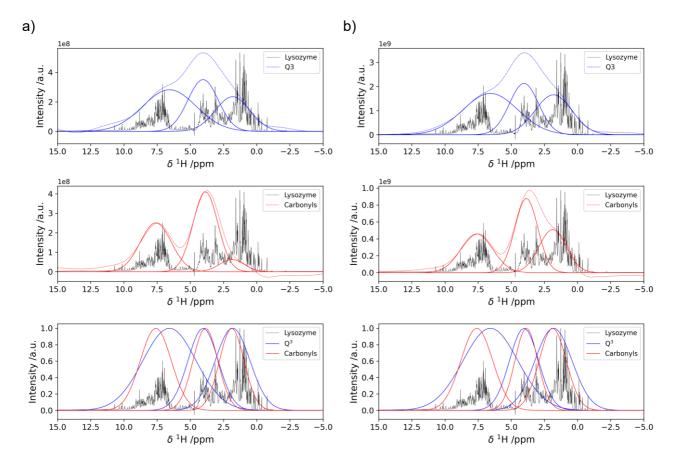


Figure S10. (top) ¹H trace of the {¹H }-²⁹Si HETCOR acquired with 500 μ s (a)/10 ms (b) of CP contact time on the freeze-dried sample. The signal is deconvoluted as a sum of three gaussian peaks. (middle) ¹H trace of the {¹H }-¹³C HETCOR acquired with 150 μ s (a)/1.5 ms (b) of CP contact time on the freeze-dried sample. The signal is deconvoluted as a sum of three gaussian peaks. (bottom) The two sets of gaussians fitting the {¹H }-²⁹Si and the {¹H }-¹³C ¹H HETCOR traces, all reported to the same height.

		Short To	CP		Long To	P
{ ¹ H }- ²⁹ Si site	δ /ppm	σ/Hz	Relative Intensity	δ /ppm	σ/Hz	Relative Intensity
1	6.6	1403	44%	6.6	1446	43%
2	4.0	797	32%	4.0	767	29%
3	1.8	914	24%	1.8	978	28%
{ ¹ H }- ¹³ C site	δ /ppm	σ/Hz	Relative Intensity	δ /ppm	σ/Hz	Relative Intensity
1	7.6	816	40%	7.6	842	30%
2	3.9	654	52%	3.9	603	41%
3	1.9	684	8%	1.9	718	29%

Table S6. Parameters of the gaussians that fit the spectra in Figure S9.

	Short TCP			
{ ¹ H }- ²⁹ Si site	δ /ppm	σ /Hz	Relative Intensity	
1	7.3	1013	22%	
2	4.5	787	33%	
3	1.3	1068	45%	
{ ¹ H }- ¹³ C site	δ /ppm	σ /Hz	Relative Intensity	
1	7.0	1019	21%	
2	4.5	574	64%	
3	1.6	523	15%	

Table S7. Parameters of the gaussians that fit the spectra in Figure 6.

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