30

Days





vt MEF

prmt5^{-/-}MEF

24

Time (h)

48

Relative cell viability

0+



Ó

2

DOX (µg/ml)

3

1

Supplementary Figure 1. PRMT5 Decreases the Doxorubicin Sensitivity of Breast Cancer Cells

(A) Representative immunohistochemical (IHC) staining for γ H2AX and Ki-67 in breast cancer tissue samples from a PDX mouse model.

(B) Dot-blot assay of RNA m6A methylation in breast cancer tissue samples from a PDX mouse model treated with or without doxorubicin.

(C) Western blot analysis of γ H2AX in MDA-MB-231 and T47-D breast cancer cells treated with doxorubicin (DOX) for the duration and dose indicated.

(**D**) MDA-MB-231 and T47-D breast cancer cells with or without PRMT5 overexpression were treated with the indicated dose of DOX for 1 hour. The cells were then washed with PBS twice and cultured in fresh medium to recover (Re) for the indicated time, and γ H2AX expression was detected by Western blotting. MDA-MB-231: 0.4 µg/ml; T47-D: 0.5 µg/ml.

(E) MDA-MB-231 cells with or without PRMT5 overexpression were orthotopically implanted subcutaneously into female athymic nude mice. The mice were treated with DOX once a week via intravenous tail vein injection at 2 mg/kg·body weight after the tumor volume reached 100-150 mm³. Tumor volume was measured every 3 days, and the mice were sacrificed 27 days after injection. The tumor growth curves and harvested tumors are displayed.

(F) (G) Immunostaining for and relative quantification of Ki-67, PRMT5, RPA32 and γ H2AX in (E) were performed.

*p < 0.05, **p < 0.01, and ***p < 0.001.

(H) Dot-blot assay of RNA m6A methylation comparing wild-type and *prmt5*-knockout mouse embryonic fibroblasts (MEFs) without doxorubicin.

(I) (J) (K) Proliferation assay (MTT) comparing wild-type and *prmt*5-knockout mouse embryonic fibroblasts (MEFs) for 24 and 48 hours and comparing doxorubicin-treated with different doses for 24 or 48 hours, as indicated.

(L) Western blot analysis of χ H2AX and PRMT5 in wild-type and *prmt*5-knockout MEFs treated and untreated with 0.4 μ g/ml doxorubicin for 24 hours.













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BRCA1 mRNA sequence of 3'UTR	Score	Decision
149		
JAUGUACAUCAGCCUGAAAAGGACUUCUGGCUAUGCAAGGGUCC		
2	0.621	m6A site (High confidence)
1268		
ACUGUUUAUAGCUGUUGGAAGGACUAGGUCUUCCCUAGCCCCCC		
2	0.567	m6A site (Moderate confidence)
	149 IAUGUACAUCAGCCUGAAAAGGACUUCUGGCUAUGCAAGGGUCC 1268 CUGUUUAUAGCUGUUGGAAGGACUAGGUCUUCCCUAGCCCCCC	ARCA1 mRNA sequence of 3'UTR Score 149 149 IAUGUACAUCAGCCUGAAAAGGACUUCUGGCUAUGCAAGGGUCC 0.621 1268 1268 CUGUUUAUAGCUGUUGGAAGGACUAGGUCUUCCCUAGCCCCCC 0.567

Supplementary Figure 2. The GO Terms and KEGG Annotation Analysis of MeRIP-Seq Data and Doxorubicin Sensitivity Assay with BRCA1 Knockdown.

(A) Gene profile of MeRIP sequencing data for the different indicated MDA-MB-231 cells. m6A peak enrichment was analyzed in each group as indicated, with enrichment found mainly near start and stop codons.

(B) Quantification of the total m6A reads in each group.

(C) GO term enrichment analysis of biological processes with a significant difference in the m6A levels of overall RNA transcripts between MDA-MB-231 cells treated with or without 0.4 μ g/ml doxorubicin for 24 hours (left) or between MDA-MB-231-PRMT5 and MDA-MB-231 cells treated with 0.4 μ g/ml doxorubicin for 24 hours (right). The circle size represents the number of molecules and depth of color shows the relationship.

(**D**) Diagram of the homologous repair-related genes affected by m6A. The genes with differential expression between MDA-MB-231 cells treated with or without 0.4 μ g/ml doxorubicin for 24 hours (left) or between MDA-MB-231-PRMT5 and MDA-MB-231 cells treated with 0.4 μ g/ml doxorubicin for 24 hours (right) are marked in red. The diagram is based on KEGG annotations.

(E) BRCA1 mRNA expression in different groups determined by qPCR analysis.

(F) BRCA1 mRNA expression determined by qPCR analysis after transfection with BRCA1-specific siRNAs (siBRCA1). *p < 0.05, **p < 0.01, and ***p < 0.001.

(G) MTT assay of MDA-MB-231 cells with PRMT5 overexpression and BRCA1-specific siRNA transfection treated with different doses of doxorubicin for 24 hours. *p < 0.05, **p < 0.01, and ***p < 0.001.

(H) Schematic analysis of the potential m6A methylation sites in the BRCA1 mRNA 3'UTR.





Supplementary Figure 3. PRMT5 Regulates DNA Repair and RNA m6A Methylation Dependent on the Arginine Methylation of ALKBH7.

(A) Potential arginine methylation sites of the human and mouse ALKBH7 proteins, with arginine-rich motifs indicated in red.

(**B**) HEK293 cells transfected with HA-ALKBH7, FLAG-PRMT5 and FLAG-PRMT5^{R368A} (catalytically dead PRMT5) as indicated. HEK293 cell lysates were immunoprecipitated with an anti-HA antibody, and Western blot analysis was performed to evaluate the arginine methylation of ALKBH7 by PRMT5 detected with an anti-symmetric dimethylarginine (SDMA) antibody.

(C) Protein stability analysis by Western blotting of MDA-MB-231 cells stably expressing wild-type ALKBH7 or ALKBH7^{3RA} (three arginine site mutations to alanine) treated with 100 μ g/ml cycloheximide (CHX) for the indicated times.

(**D**) Western blot analysis of ALKBH7 expression in MDA-MB-231 cells stably expressing wild-type ALKBH7 or ALKBH7^{3RK} treated with MG132 (100 μ M), leupeptin (20 μ M) or MLN4924 (20 μ M) for 6 hours as indicated.

(E) Bioinformatic analysis of the E3 ligase responsible for ALKBH7 degradation performed at <u>http://ubibrowser.ncpsb.org/ubibrowser/</u>. (F) ALKBH7 expression detected by Western blotting of HEK293 cells transfected with FBW7 α , FBW7 α/Δ F-box and ALKBH7 for 48 hours as indicated.

(G) HEK293 cells transfected with FBW7 α , ALKBH7 and ALKBH7^{3RK} as indicated and treated with 100 μ g/ml CHX before harvest at the indicated times. ALKBH7 expression was detected by Western blotting.

(H) A comet assay was performed with MDA-MB-231 cells with or without PRMT5 overexpression and ALKBH7-specific siRNA knockdown treated with 0.4 μ g/ml doxorubicin for 24 hours. Right panel, the quantification of the comet tail that represents broken DNA strands (TDNA%). ^{##}p < 0.01 vs. MDA-MB-231-PRMT5-siNC, **p < 0.01 vs. MDA-MB-231-siNC, and ***p < 0.001 vs. MDA-MB-231-siNC.

(I) Western blotting was performed to analyze γ H2AX expression in MDA-MB-231 cells with or without PRMT5 overexpression and ALKBH7-specific siRNA knockdown treated with doxorubicin as indicated for 24 hours.

(J) Immunofluorescence (IF) analysis of RPA foci in MDA-MB-231 cells was performed with cells treated as in (H).

(K) A dot-blot assay of RNA m6A methylation in MDA-MB-231 and T47-D cells with ALKBH7 overexpression treated with doxorubicin for 24 hours was performed. The doxorubicin concentrations used to treat the MDA-MB-231 and T47-D cells were 0.4 μ g/ml and 0.5 μ g/ml, respectively.

(L) ELISA was performed to analyze the RNA m6A methylation levels in MDA-MB-231 cells overexpressing ALKBH7 treated with $0.4 \mu g/ml$ doxorubicin for 24 hours.

(M) RT-qPCR analysis of BRCA1 mRNA in T47-D cells overexpressing PRMT5 with ALKBH7-specific siRNA transfection treated with 0.5 μ g/ml doxorubicin for 24 hours was performed.

(N) T47-D cells overexpressing PRMT5 with ALKBH7-specific siRNA transfection were treated with 0.5 μ g/ml doxorubicin for 24 hours and then treated with vehicle or 3.2 μ M flavopiridol for 6 hours. BRCA1 mRNA levels were measured by RT-qPCR, and the F/V ratio of BRCA1 mRNA was determined (mean \pm SEM; n = 3). *p < 0.05, **p < 0.01, and ***p < 0.001.



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Methylene Blue









Supplementary Figure 4. PRMT5 and ALKBH7 Regulate RNA m6A Methylation Mediated by ALKBH5.

(A) Western blot analysis of ALKBH5 and FTO in PRMT5-overexpressing MDA-MB-231 cells treated with or without 0.4 μ g/ml doxorubicin for 24 hours.

(B) Western blot analysis of ALKBH5 and FTO in ALKBH7-overexpressing MDA-MB-231 cells treated with or without 0.4 μ g/ml doxorubicin for 24 hours.

(C) Immunofluorescence assay of ALKBH5 and ALKBH7 in MDA-MB-231 cells overexpressing ALKBH7 treated with or without 0.4 μ g/ml doxorubicin for 24 hours as indicated. The bars in each images represent for 10 μ m.

(D) Immunoprecipitation assay for MYC-ALKBH7 and ALKBH5 in MDA-MB-231 and T47-D cells. Cell lysates were immunoprecipitated with an anti-MYC tag antibody (MYC-ALKBH7), and endogenous ALKBH5 was detected with an anti-ALKBH5 antibody.

(E) Dot-blot assay with MDA-MB-231 cells overexpressing ALKBH7 with ALKBH5-specific siRNA transfection treated with 0.4 μ g/ml doxorubicin for 24 hours and then subjected to poly(a)+ RNA extraction. An anti-m6A antibody was used for the dot-blot assay with 200 ng of mRNA, and methylene blue staining was used as the loading control.

(F) BRCA1 mRNA levels measured by RT-qPCR and the F/V ratio of BRCA1 mRNA. MDA-MB-231 cells overexpressing PRMT5 or ALKBH7 with ALKBH5-specific siRNA transfection were treated with 0.4 μ g/ml doxorubicin for 24 hours and then treated with vehicle or 0.8 μ M flavopiridol for 3 hours (mean \pm SEM; n = 3; **P < 0.01 vs. Ctrl).

(G) RT-qPCR analysis of BRCA1 mRNA in MDA-MB-231 cells overexpressing PRMT5 or ALKBH7 with ALKBH5-specific siRNA transfection treated with 0.4 μ g/ml doxorubicin for 24 hours. *p < 0.05, **p < 0.01, and ***p < 0.001.

Supplementary Fig. 5



Supplementary Figure 5. Tadalafil is a Novel PRMT5 Inhibitor that Increases the Doxorubicin Sensitivity of Breast Cancer Cells.

(A) (B) Western blotting of two downstream targets of PRMT5, H4R3me2s and symmetric dimethylarginine (SDMA), in MDA-MB-231 and T47-D breast cancer cells treated with 100 μ M tadalafil for 24 hours as indicated.

(C) Proliferation analysis of MDA-MB-231 cells treated with different doses of doxorubicin with or without 100 μ M tadalafil for 24 hours as indicated.

(**D**) Proliferation analysis of HCC1937 cells treated with different doses of doxorubicin and 0, 25, 50, or 100 μ M tadalafil for 24 hours as indicated. *p<0.05, **p<0.01, and ***p<0.001.

(E) Proliferation analysis (MTT) of MDA-MB-231 cells transfected with scramble or PRMT5-specific siRNA and then treated with 100 μ M tadalafil and 0.4 μ g/ml doxorubicin for 24 hours as indicated.

(F) Proliferation analysis (MTT) of MDA-MB-231 cells transfected with scramble or PDE5-specific siRNA and then treated with 100 μ M tadalafil and different concentration of doxorubicin for 24 hours as indicated.

(G) Realtime RT-PCR analysis of PDE5 after siRNA transfection for 24 hours.

(H) Proliferation analysis of MDA-MB-231 cells treated with different doses of olaparib with or without 100 μ M tadalafil for 24 hours as indicated.

Supplementary Fig. 6



Supplementary Figure 6. Tadalafil Increases Doxorubicin Sensitivity in Breast Cancer Cells and a PDX Mouse Model.

(A) Immunohistochemical analysis (top panel) and quantification (bottom panel) of γ H2AX, Ki-67, PRMT5, and ALKBH7 in athymic nude mice with orthotopically implanted MDA-MB-231 cells treated with tadalafil and doxorubicin as indicated.

(B) Quantification (bottom panel) of the γ H2AX, Ki-67, PRMT5, and ALKBH7 staining shown in (A).

(C) Proliferation analysis of MDA-MB-231 cells with doxorubicin resistance (MDA-MB-231-ADR) treated with 100 μ M tadalafil and different doses of doxorubicin for 24, 48, or 72 hours. qPCR analysis of PRMT5 expression is shown in the top left corner.

(**D**) Tumor tissue display of MDA-MB-231-ADR breast cancer cells treated with tadalafil or doxorubicin as indicated (n=4-5/group).

(E) Tumor tissue display of patient-derived xenograft (PDX) models of breast cancer treated with tadalafil and doxorubicin as indicated (n=5-7/group). *p < 0.05, **p < 0.01, and ***p < 0.001.



Supplementary Figure 7. Tadalafil Increases RNA m6A Methylation by Reducing the Nuclear Translocation of ALKBH5.

(A) Immunofluorescence assay for ALKBH5 in MDA-MB-231 cells treated with tadalafil (100 μ M) and 0.4 μ g/ml doxorubicin for 24 hours as indicated.

(B) Nuclear, cytosolic and whole cell lysate (WCL) expression of ALKBH5 and ALKBH7 in MDA-MB-231 cells treated with 0.4 µg/ml doxorubicin with or without combination with tadalafil as indicated.

(C) Dot-blot assay of RNA m6A methylation in MDA-MB-231 cells treated with 0.4 μ g/ml doxorubicin with or without tadalafil (100 μ M) for 24 hours. An anti-m6A antibody was used for the dot-blot assay with 200 ng of mRNA, and methylene blue staining was used as the loading control.

(**D**) qPCR analysis of BRCA1 mRNA in MDA-MB-231 (left panel) and T47-D (right panel) cells treated with 0.4 μ g/ml and 0.5 μ g/ml doxorubicin, respectively, with or without 100 μ M tadalafil for 24 hours.

(E) BRCA1 mRNA levels measured by RT-qPCR and the F/V ratio of BRCA1 mRNA. MDA-MB-231 (left panel) and T47-D (right panel) cells were treated with doxorubicin and tadalafil as in (**D**) and treated with vehicle or 0.8 μ M flavopiridol for 3 hours (MDA-MB-231 cells) or vehicle or 3.2 μ M flavopiridol for 6 hours (T47-D cells) (mean \pm SEM; n = 3).

(F) Comet assay with MDA-MB-231 cells treated with doxorubicin and tadalafil as in (D). The right bottom column shows the quantification of the comet tail that represents broken DNA strands (TDNA%). **p < 0.01 and ***p < 0.001.

(G) Western blot analysis of γ H2AX and cleaved Caspase-3 expression in MDA-MB-231 cells treated with 100 μ M Tadalafil and 0.4 μ g/ml doxorubicin as indicated for 24 hours.

Table S1 Summarization of tumor-stage and degrade information of breast cancer patients

Neoadjuvant therapy		Yes	No
Stage	IIA	6	4
	IIB	4	2
	IIIA	2	6
	IIIC	3	2
	IV	0	1
	Not available	0	3
TNM grade	T1	0	1
	T2	15	13
	Т3	0	1
	Not available	0	3

 Table S2 Summarization of clinic-pathologic information of breast cancer patients

Gender	Female	n=174
	Male	n=0
	Invasive carcinoma	n=110
	Ductal carcinoma in situ	n=4
Туре	Invasive ductal carcinoma	n=5
	Ductal carcinoma in situ with microinvasion	n=4
	Invasive lobular carcinoma	n=2
	Complete response	n=2
Curativa offact	Partial response	n=124
curative effect	Stable disease	n=40
	Progressive disease	n=8