

## **Supporting Information for:**



**Microscopy Provides Insights into Nitrous Acid Speciation at the** 

## **Kaolin Mineral-Air Interface**

- 7 Cheng Zhu<sup>1</sup>, Gargi Jagdale<sup>1</sup>, Adrien Gandolfo<sup>2</sup>, Kristen Alanis<sup>1</sup>, Rebecca Abney,<sup>2,3</sup>
- 8 Lushan Zhou<sup>1</sup>, David L. Bish,<sup>1</sup> Jonathan D. Raff<sup>1,2,\*</sup>, and Lane A. Baker<sup>1,\*</sup>
- *1Department of Chemistry*
- *Indiana University*
- *Bloomington, IN 47401*
- 
- *2Paul H. O'Neill School of Public & Environmental Affairs*
- *Indiana University*
- *Bloomington, IN 47405*

- *3Warnell School of Forestry and Natural Resources*
- *University of Georgia*
- *Athens, GA 30602*

- \*Authors to whom correspondence should be addressed.
- e-mail: lanbaker@indiana.edu; phone 812-856-1873 ; fax 812-855-8300
- e-mail: jdraff@indiana.edu; phone 812-855-6525



# **SI-1 Election Micrographs of Nanopipettes**

<span id="page-2-0"></span>

- **Figure S1.1** Side view (**a**) and the end-on view (**b**) of a typical nanopipette used in these
- experiments.
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## <span id="page-3-0"></span>**SI-2 Experimental Details of Flow Reactor Studies**

## **Flow Reactor Configuration**

 **Figure S2.1** displays a schematic of the experimental system used to measure HONO released from nitrite reactions on clay mineral surfaces. Panel A provides an overview, and Panel B and C provide details of the sample holder located in the dashed box in Panel A.





 **Figure S2.1** 3D representation of the experimental setup used to study HONO(g) emission from nitrite reaction on mineral surfaces. Panel A shows the laminar flow tube reactor, gas in- and outlets, and the nitrite injection system (septum port and syringe). Gas exiting the flow reactor flows to the chemical ionization mass spectrometer. Panels B and C show details of the Teflon holder used to hold pure water or clay mineral samples placed underneath the injection system.

#### **Calibration of the Chemical Ionization Mass Spectrometer (CIMS)**

 The CIMS response to HONO was calibrated in the range of 2 to 10 ppb following 62 a method described previously.<sup>1</sup> Briefly, HONO is produced from the acid displacement of gaseous hydrochloric acid (HCl) flowing across a stirred bed of sodium nitrate powder at 50% relative humidity (RH). The HONO generation system, including the permeation device used to supply HCl is housed in an oven held at 55 °C. The HONO concentration in the generator outflow is monitored continuously by cavity-enhanced absorption 67 spectroscopy (CEAS)<sup>2</sup> to assess the absolute mixing ratio. The generator output was 50  $\pm$  5 ppb HONO and was subsequently diluted with humid (30% RH) high purity air to obtain the desired concentration prior to being measured by CIMS.

 Following the calibration procedure described above, the CIMS sensitivity is 5442  $\pm$  178 cps ppb<sup>-1</sup>. A limit of detection (LOD) of 60 ppt is calculated according to the following equation:

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$$
[HONO]_{LOD} = [HONO]_0 + (3 \times SD_{[HONO]_0})
$$
 Eqn S2.1

 where [HONO]0 is the background HONO concentration measured under a flow of pure nitrogen (30% RH) for 15 min following 1 h of purging with the same carrier gas mixture; SD<sub>IHONOI</sub> is the standard deviation calculated from the same data set.

#### <span id="page-4-0"></span>**SI-3 Finite Element Method (FEM) Simulations**

 To quantify the surface charge density on dickite crystals, finite-element method (FEM) simulation models were created with COMSOL Multiphysics (COMSOL, Inc.). **Figure S3.1** shows the schematic of the model. Typical dimensions of the pipette were

83 acquired from the electron micrographs. The inner radius  $(r_i)$  of the pipette tip opening 84 was set to 37 nm and the outer radius of the pipette  $(r_0)$  was set to  $r_1/0.7$ . The tip-to-85 sample distance is defined as  $D_{\text{ps}}$ . Other parameters such as ion concentrations and diffusion coefficients (taken here at infinite dilution) are shown in **Table 3.1**. The potential was applied inside the pipette (B1). In contrast, the boundary on the side of the model 88 was set as ground ( $V = 0$ , B3). The boundaries highlighted in green represent the pipette 89 wall that carries a surface charge density of  $\sigma_p$ . It should be noted that the radius of the pipette, surface charge density of the pipette, and the concentration of the solution were adjusted when measuring the pH-dependent surface charge to compensate for the change in the surface charge, geometry of the pipette, and the concentration change resulting from the addition of acid.

 $\sigma$  Of note, work by Thatenhorst et. al,<sup>3</sup> convincingly demonstrated that steeply angled surfaces result in an overestimation of the tip-sample distance, and values for the maximum overestimation can be found from the product of the inner radius of the pipette 97 (ri) and the slope (maximum slopes imaged here were  $\sim$ 2.7). For conditions here, this results in a maximum of ~100 nm overestimation of the tip-sample distance, although we note that this does not include possible deviations in the angle of approach of the pipette. For models here, attempts to include both surface charge and sample tilt reduced model symmetry and resulted in nonconvergent results. For models used here, as a first approximation, no height correction for was applied.

 After collecting the *ICD* images, the average I-V curve for basal planes was calculated by averaging all the I-V curves at negative sites. Similarly, the average I-V curve for edge surfaces was collected by averaging all the I-V curves at positive sites.

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106 An exact value of  $\sigma_p$  was first calculated by fixing the D<sub>ps</sub> at 2.34  $\mu$ m (the retracted 107 distance used in the experiment, where the I-V is independent from the surface charge 108 on sample surfaces) and then changing the  $\sigma_{p}$  until the resultant I-V curve matched the 109 retracted I-V curve measured in the experiment.



110 **Figure S3.1** (**a**) Schematic of the model created with COMSOL Multiphysics. The 111 potential was applied inside the pipette (B1) and the boundary on the side of the model<br>112 was set as ground (B3). The green lines highlight the boundaries that carry a surface was set as ground (B3). The green lines highlight the boundaries that carry a surface 113 charge on the pipette (B2). The sample surface charge is highlighted in blue (B4). (**b**) At 114 the highlighted areas near the tip opening, the inner and outer radius of the pipette was 115 set as  $r_i$  and  $r_o$ , respectively.

116 After resolving the surface charge of the pipette,  $D_{ps}$  was then reduced to a small 117 value which is close to the outer radius of the pipette used in the experiment. Both the 118 D<sub>ps</sub> and the surface charge on samples ( $\sigma$ <sub>s</sub>) were then adjusted in simulations until the 119 resultant I-V curve matched the extended I-V curve measured in the experiment. 120 Reducing D<sub>ps</sub> typically reduces the current magnitude of I-V at both positive potentials 121 and negative potentials. In contrast, increasing  $\sigma_s$  results in an increase of current at

122 positive potentials and a decrease in current at negative potentials. Only one of the  $D_{ps}$ 123 and σs combinations will result in an I-V that exactly matches that collected in the 124 experiment.

### 125 **Table S3.1** Common Parameters used in the FEM Simulation



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### 127 **Table S3.2** Simulation Parameters and Results for Figure 4



**Table S3.3** Simulation Parameters and Results for Figure 3

pH of bulk solution	n	$\sigma_{p}$	$\mathsf{ID}_{\mathsf{ps}}$	$\sigma_{\rm s}$
	25.55 nm	$-14.25$ mC/m <sup>2</sup>	$24.5$ nm	$-132$ mC/m <sup>2</sup>
	26.43 nm	$-8.33$ mC/m <sup>2</sup>	$26.1 \text{ nm}$	$-133 \text{ mC/m}^2$
	27.31 nm	$-3.92$ mC/m <sup>2</sup>	30.5 nm	$-135$ mC/m <sup>2</sup>

### <span id="page-8-0"></span>**SI-4 Characterization of Dickite Samples**

#### **Structure of Kaolin Group Minerals**

134 Kaolin group minerals,  $Al_2Si_2O_5(OH)_4$ , are among the most abundant clay minerals in soil. The structure of the kaolin minerals, kaolinite, dickite, and nacrite have already been well investigated previously with multiple techniques, including X-ray and neutron 137 diffraction,<sup>4</sup> X-ray absorption spectroscopy,<sup>5, 6</sup> Fourier-transform infrared spectroscopy 138 (FTIR),<sup>4, 7-9</sup> atomic force microscopy (AFM),<sup>10-12</sup> nuclear magnetic resonance (NMR)<sup>13</sup> 139 and electron microscopy (EM).<sup>14</sup> All of these techniques demonstrate that kaolin group minerals are composed of a periodic layered structure consisting of a layer of SiO4 tetrahedra (T, siloxane) linked to a sheet of aluminum (hydr)oxide octahedra (O, gibbsite) via shared oxygen and hydroxide groups (**Figure S4.1)**. The basic TO structure is repeated in space with hydrogen bonds holding together neighboring TO layers. Depending on the stacking of individual TO layers the kaolin group minerals are further 145 classified into kaolinite, nacrite, and dickite.<sup>15</sup> Dickite was used in these studies as a proxy for all of the kaolin group minerals because it is readily available in well-ordered macroscopic crystals. Dickite samples used for this study were characterized by scanning

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 electron microscopy (FEI Quanta-FEG, Hillsboro, OR). Selected micrographs of dickite are shown in **Figure S4.2** and **S4.3**.



 **Figure S4.1** The typical structure of kaolin group minerals. Each silicon is surrounded by four oxygens, which forms an extended tetrahedral silica sheet. Four hydroxide groups and two oxygens surround each aluminum ion to form an extended octahedral gibbsite sheet. Silica and gibbsite sheets are bonded together by shared oxygens and hydroxide 156 groups. Repeated silica-gibbsite units held together by interlayer hydrogen bonds to yield<br>157 a layered structure. Color code: Cyan = Si: Red = O. White = H: Blue = Al.  $\alpha$  layered structure. Color code: Cyan = Si; Red = O, White = H; Blue = Al.



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- 160 **Figure S4.2** Scanning electron micrographs of dickite crystals in various orientations. In<br>161 (a) and (c) the basal plane is in contact with the substrate whereas in (b) and (d) edge
- 161 (**a)** and (**c**) the basal plane is in contact with the substrate whereas in (**b)** and (**d**) edge
- surfaces contact the substrate.
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165 **Figure S4.3** Scanning electron micrographs of dickite crystals. Yellow circles highlight numerous small dickite flakes adhered to the dickite crystals.

#### **Characterizing and Orienting Individual Dickite Crystals for SICM Analysis**

 Single crystals of dickite were selected under a polarizing microscope and mounted without adhesives or oil to avoid surface contamination. They were placed on MiTeGenTM Kapton micro loops or micro grippers and mounted on a Bruker Venture D8 diffractometer equipped with a PhotonIII detector for data collection at 297(2) K.

173 X-ray diffraction data collection was carried out using an I<sub>u</sub>S 3.0, Incoatec source 174 and Mo K $\alpha$  radiation (graphite monochromator) with a frame time of 0.5, 0.75, or 1 second and a detector distance of 40.00 mm. Data were measured to a resolution of 0.71 Å with 176 a redundancy of 4. Three or five major sections of frames were collected with 1<sup>o</sup>  $\omega$  and  $\phi$  scans. An additional fast scan was collected to obtain accurate intensities for a few high- intensity reflections. The frames were integrated with the Bruker SAINT software 179 package<sup>16</sup> using a narrow-frame algorithm, and data were scaled and corrected for 180 absorption effects using the Multi-Scan method  $(SADABS<sup>17</sup>)$ .

 Space group Cc was determined based on intensity statistics and systematic absences, consistent with the published structure. The structure was solved and refined 183 using the SHELX suite of programs.<sup>18, 19</sup> An intrinsic-methods solution was calculated, which provided all non-hydrogen atoms from the E-map. Full-matrix least squares / difference Fourier cycles were performed, by which hydrogen atoms were located. All non-hydrogen atoms were refined with anisotropic displacement parameters. The hydrogen atoms were refined freely with isotropic displacement parameters. The final 188 anisotropic full-matrix least-squares refinement on  $F<sup>2</sup>$  resulted in low R values and a near-featureless difference map. Flack parameters were calculated from anomalous dispersion.

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 The face index of the crystals was determined to relate Miller planes (0 0 1) and (0 0 -1) to the orientation of the TO layers of dickite, comprised of Si-O tetrahedra on one side and Al-O,OH octahedra on the other side. Subsequently, the crystals were transferred with known orientation onto the PDMS substrate for further investigation.

 Additional measurement parameters and refinement results are summarized below. Images of the crystal, the asymmetric unit, and the unit cell are shown as well. CIFs are included as supplemental information.



**Table S4.1** Crystallographic measurement parameters and refinement results for oriented dickite crystals and their corresponding charge maps.





**Figure S4.4 Left:** Packing and unit cell of crystal 21061, representative of 21063 and 21069. **Right:** Packing and unit cell of crystal 21062, representative of 21068 and 21070. View along the crystallographic *a* axis.



**Figure S4.5** Asymmetric unit of crystal 21061, which is representative of all other crystals.

Displacement ellipsoids are at 50% probability.

## <span id="page-17-0"></span>**SI-5 Topography and Surface Charge Images of the Basal Plane of a**

# **Dickite Crystal**



**Figure S5.1** Surface charge images of the basal plane of a dickite crystal at pH 5 (**a**), 4 (**b**), and 3 (**c**) and the topography of the scan area (**d**). These images show a defect (pit) on the surface exposed a positively charged surface underneath the surface. Highresolution surface charge (**e**) and topography (**f**) images of the area highlighted by the red square in (**d**).

<span id="page-18-0"></span>**SI-6 Secondary Ion Mass Spectrometry Images of Dickite Crystals**



**Figure S6.1** Secondary ion mass spectrometry images of dickite crystals. As indicated by the similar intensity shown in (**b**) and (**d**), dickite crystals are composed of an almost equal amount of silicon and aluminum; magnesium appears as a minor component of the surface in (**c**).

## <span id="page-19-0"></span>**SI-7 Additional SICM Results of the Interactions between Malonate**

## **and Dickite Crystals**



**Figure S7.1** Electron micrographs of dickite particles before (top) after (bottom) immersion in 100 mM KCl and 10 mM potassium malonate overnight. After immersion, adsorbates appear and step edges are less well defined.



**Figure S7.2** (a) Combined topography and surface charge image of a dickite crystal in 100 mM KCl solution. (b) The addition of malonate turned the negative surface charge into positive on basal planes. Image (b) is recorded 105 minutes after image (a), solution  $pH = 5$ .



**Figure S7.3** Combined topography and surface charge images of a dickite crystal before (**a**) and after (**b**) the addition of malonate. Upon lowering the pH (**c, d**), the addition of acid does not change the apparent surface charge further. Images recorded in succession, with image acquisition time ca. 135 minutes. Image (**a**) recorded prior to addition of malonate.

#### <span id="page-22-0"></span>**SI-8 HONO Emission Flux Estimation**

Chemical ionization mass spectrometry was used to quantify the amount of HONO(q) emitted from the surface of clay mineral substrates upon addition of 1 nmol NO $_2$ <sup>-</sup>. A dilution step was applied to compensate for the CIMS inlet flow (3500 ml min<sup>-1</sup>) compared to the flow tube outflow (1000 ml min-1). Before each experiment, 5 to 10 consecutive measurements of both flow, reactor outflow, and dilution flow was performed to estimate the dilution correction factor. The emission flux was calculated from **Eqn S8.1**:

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F_{HONO} = \frac{[HONO]_{corr} \times c \times Q}{A \times m \times 10000}
$$
Eqn S8.1

where [HONO]<sub>corr</sub> is the HONO mixing ratio corrected for dilution in ppt, c is a conversion factor from ppt to molecule cm-3, *Q*, the sheath flow running in the flow tube (cm3∙s-1), *A* is the specific surface area (as presented in **Table S9.3**) in g∙m-2 and *m* is the mass sample in g. The specific surface area of the clay minerals was determined using a gravimetric method that measures the adsorption of vapor-phase ethylene glycol monoethyl ether (EGME), which probes both the external surface and sub-nanometer pores of phyllosilicates.20 Emission flux is also presented as a function of the geometric surface area in **Figure S8.2**. In this case, the numerator in **Eqn S2** is replaced by the geometric surface area of exposed sample surface,  $1.327 \text{ cm}^2$ , in our experimental setup. HONO flux calculations presented in this study are corrected for background levels of HONO present in the system, as defined by the average HONO mixing ratio determined five minutes before nitrite injection.

Before the nitrite solution injection (**Figure S2.1b**), the HONObkg signal recorded was stable at low levels, as presented in **Figure S8.1** (light grey area). In this figure, the mean HONO signal is 801  $\pm$  181 cps, which is typical of the background level observed in all experiments. Following nitrite injection (**Figure S2.1c**), the CIMS measured the sum of the HONO<sub>bkg</sub> and the HONO released into the gas phase from the surface (HONO<sub>surf</sub>) (**Figure S8.1**, grey area).



**Figure S8.1** Experimental profile of the nitrite reactivity on the mineral surface forming HONO. Raw HONO signal is express in counts per second (cps) corrected for dilution as recorded by the CIMS. The light grey and grey areas represent the HONO measurements before and after nitrite injection, respectively. The light red area represents the HONO integrated area surface area presented in titration figures.



**Figure S8.2** The pH profile for the integrated HONO emission flux stemming from the addition of 1 nmol of nitrite on clay mineral surfaces. The amount of HONO emitted is relative to the geometric surface area of the sample holder, rather than the specific surface area used to display the data in Figure 6. Dotted lines are guides for the eye, and the error bars represent the 95% confidence interval of the mean of three replicate measurements. Note that amount of HONO released from kaolinite has been multiplied by 10 for clarity.



**Figure S8.3** Time series of the HONO emitted from pH adjusted systems (normalized to the geometric surface area of the substrate). The traces represent the means of triplicate experiments. Statistical standard deviation is not shown for clarity purposes. The mean statistical standard deviations are  $17-33$  %,  $12-39$  %,  $18-49$  %, and  $16-26$  % for the water, kaolinite, dickite, and dickite + malonate systems, respectively.

#### <span id="page-26-0"></span>**SI-9 Dissolution Studies and ICP-MS analysis of the bulk solution**

Soluble reaction products stemming from surface complexation of malonate on dickite surfaces were analyzed as follows. Dickite crystals were immersed in the malonate-containing solutions overnight, and the bulk solutions were subsequently analyzed with inductively coupled plasma-mass spectrometry (ICP-MS). Details of the experiment conditions are listed in **Table S9.1**. The samples are classified into two groups. The first group consists of samples 1 and 2, which are similar to the SICM experiment condition, in which several tens of dickite crystals were transferred onto a PDMS coated Petri dish and immersed in the same solution used in SICM measurements. The second group consists of samples 3 and 4, which are prepared by immersing dickite crystals in a more concentrated malonate solution.

To prepare samples for ICPMS analysis, each sample vial containing the malonate extract solution was submerged in a water bath at 75 °C and dried under a gentle flow of ultrapure  $N_2$  (Airgas, 5.0), delivered by a needle above the solution. The resulting dry samples were then diluted in 2% nitric acid. Nitric acid was vacuum distilled to ensure highest purity, and then diluted with DI water to obtain the desired concentration. Samples were then analyzed on an Agilent 7700 inductively coupled plasma mass spectrometer (ICP-MS). Results of ICP-MS analyses are summarized in **Table S9.2**.

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## **Table S9.1 Experiment Conditions for ICP-MS Analysis**

## **Table S9.2 Element Concentration in Solutions**



### **Table S9.3 Specific Surface Area of Clay Minerals Measured using EGME Method**



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