

Supplementary Materials for
**Cis-regulatory chromatin loops analysis identifies GRHL3 as a master
regulator of surface epithelium commitment**

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Other Supplementary Material for this manuscript includes the following:

Tables S2 to S5

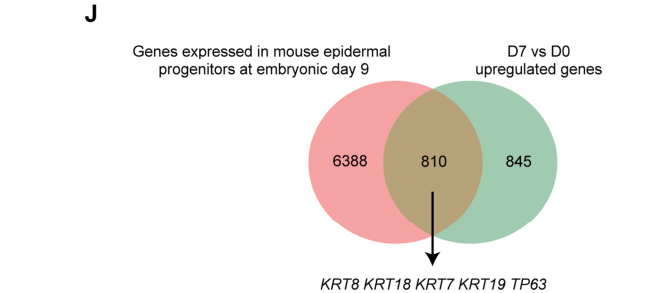
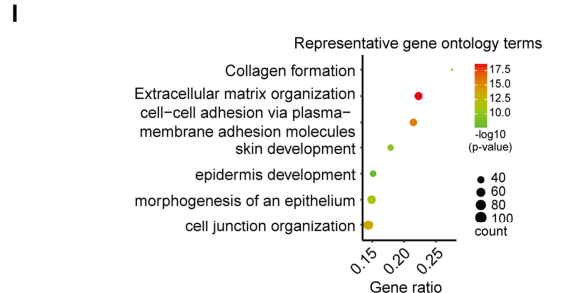
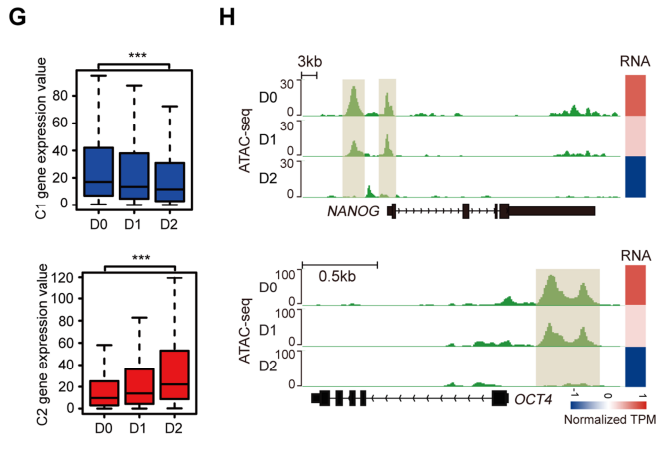
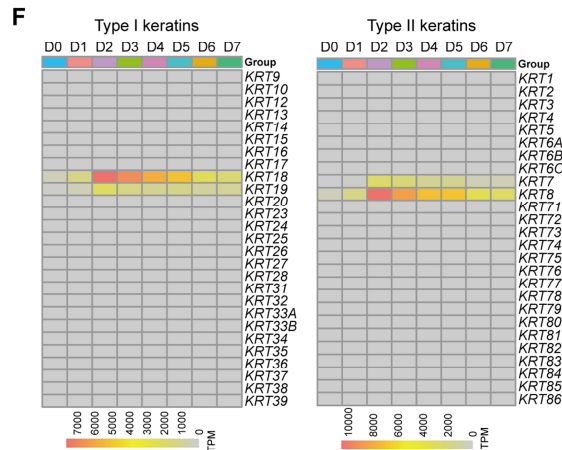
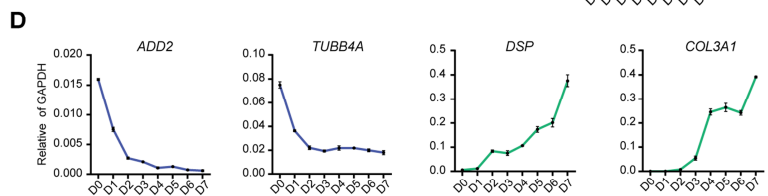
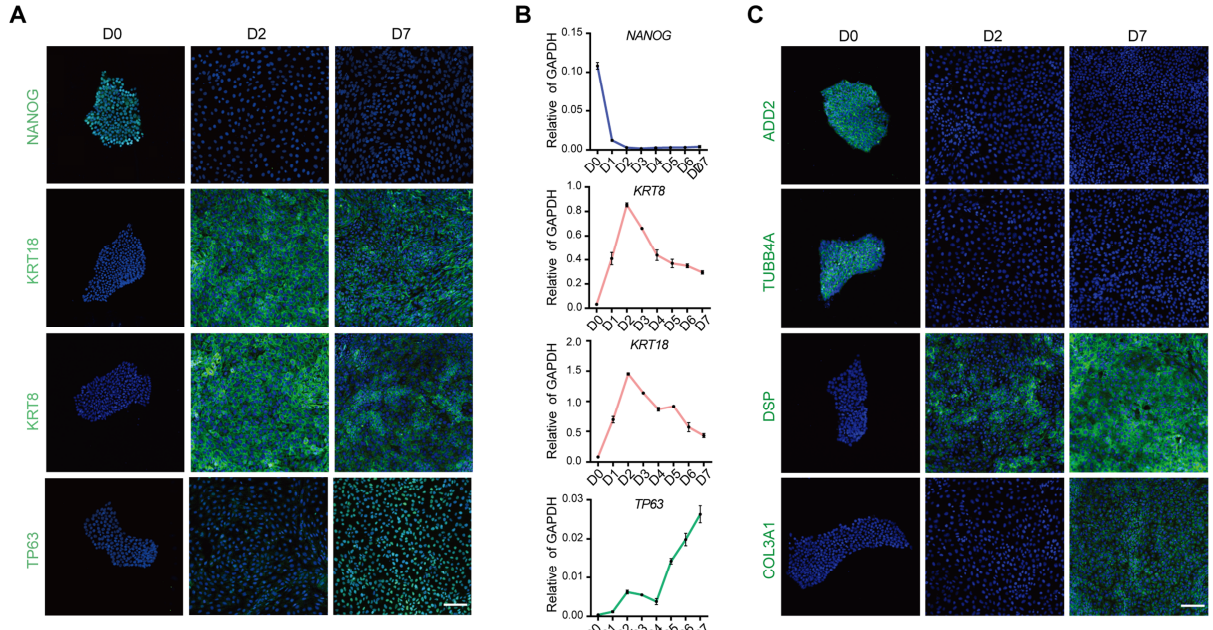


Fig. S1.

Gene expression profile during SE differentiation (A) Immunofluorescence staining of NANOG, KRT8, KRT18, and TP63 in the differentiated hESCs on D0, D2 and D7. Scale bar, 100um. (B) qRT-PCR validation of the expression levels of NANOG, KRT8, KRT18, and TP63 during SE differentiation. Values are shown as means \pm SD (n = 3 biological replicates). (C) Immunofluorescence staining of ADD2, TUBB4A, DSP and COL3A1 in the differentiated hESCs on D0, D2 and D7. Scale bar, 100um. (D) qRT-PCR validation of the expression levels of ADD2, TUBB4A, DSP and COL3A1 during SE differentiation. Values are shown as means \pm SD (n = 3 biological replicates). (E) qRT-PCR validation of the expression levels of KRT7 and KRT19 during SE differentiation. Values are shown as means \pm SD (n = 3 biological replicates). (F) Heatmap of type I and type II keratin expression changes during SE differentiation. The color bar shows the expression value (TPM) from the RNA-seq. (G) Box plots of the expression levels of genes closest to the C1 and C2 peaks in the differentiated hESCs on D0, D1, and D2. (H) Snapshots of genome browser showing chromatin accessibility at NANOG and OCT4 loci. Gene expression is also displayed in heatmaps (\log_2 TPM). The genome browser view scales were adjusted on the basis of the global data range. (I) Representative gene ontology terms identified from the genes highly expressed in D7-differentiated cells (compared to D2-differentiated cells). (J) Venn diagram showing overlap between genes expressed (TPM [transcripts per kilobase of exon model per million mapped reads] >10) in mouse epidermal progenitors at embryonic day 9 and genes (TPM > 10) highly expressed in SE cells (compared to hESCs).

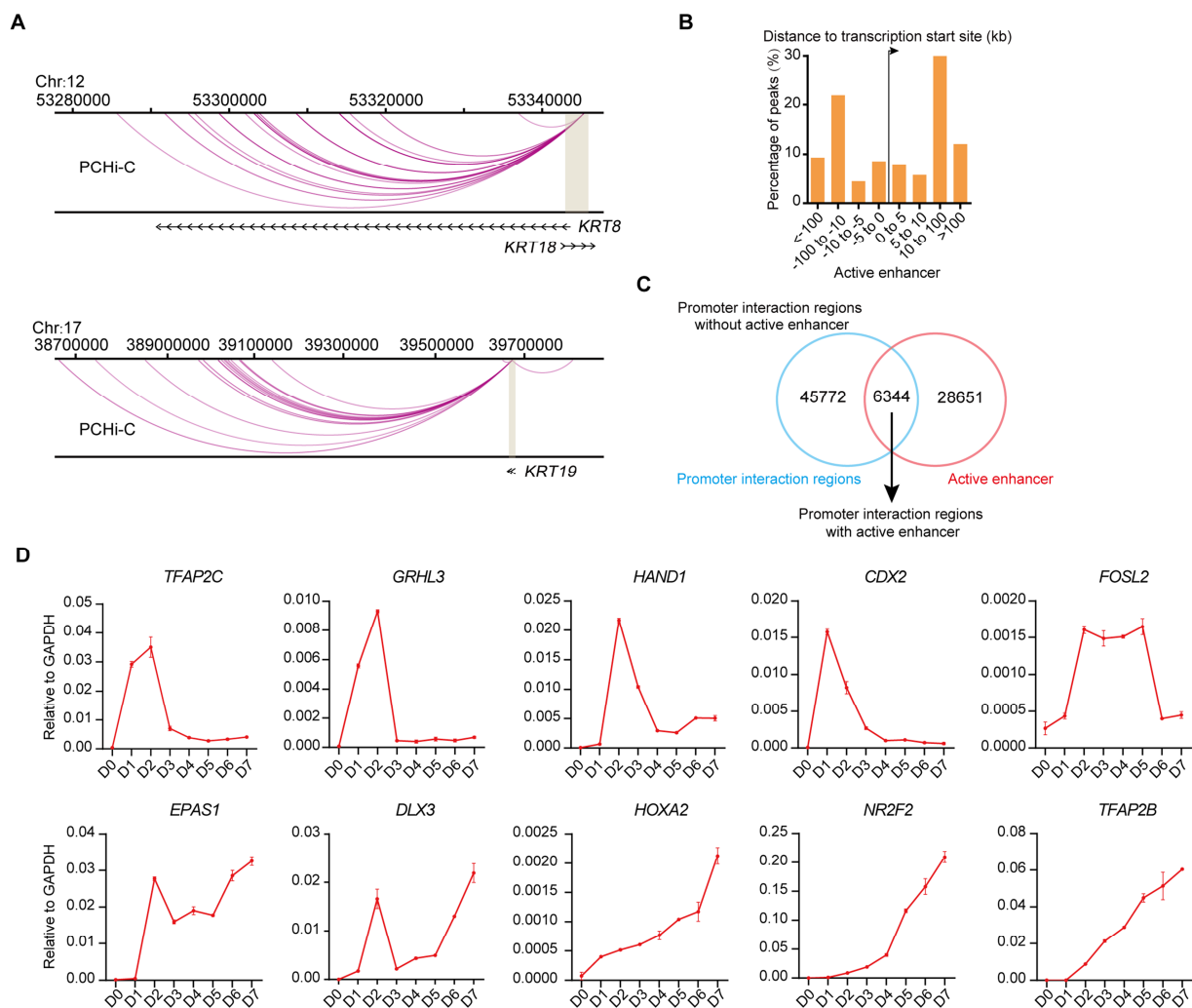


Fig. S2.

Promoter interactions and potential regulators of SE-initiating cells (A) Examples of promoter-anchored chromatin loops at *KRT8/18* and *KRT19* loci in SE-initiating cells. (B) Histogram showing the distribution of the distance between active enhancer and TSSs in SE-initiating cells. (C) Venn diagram showing the identification of promoter interaction regions with active enhancers in SE-initiating cells. (D) qRT-PCR analysis of top 10 candidate transcription factors during SE differentiation. Values are shown as means \pm SD ($n = 3$ biological replicates).

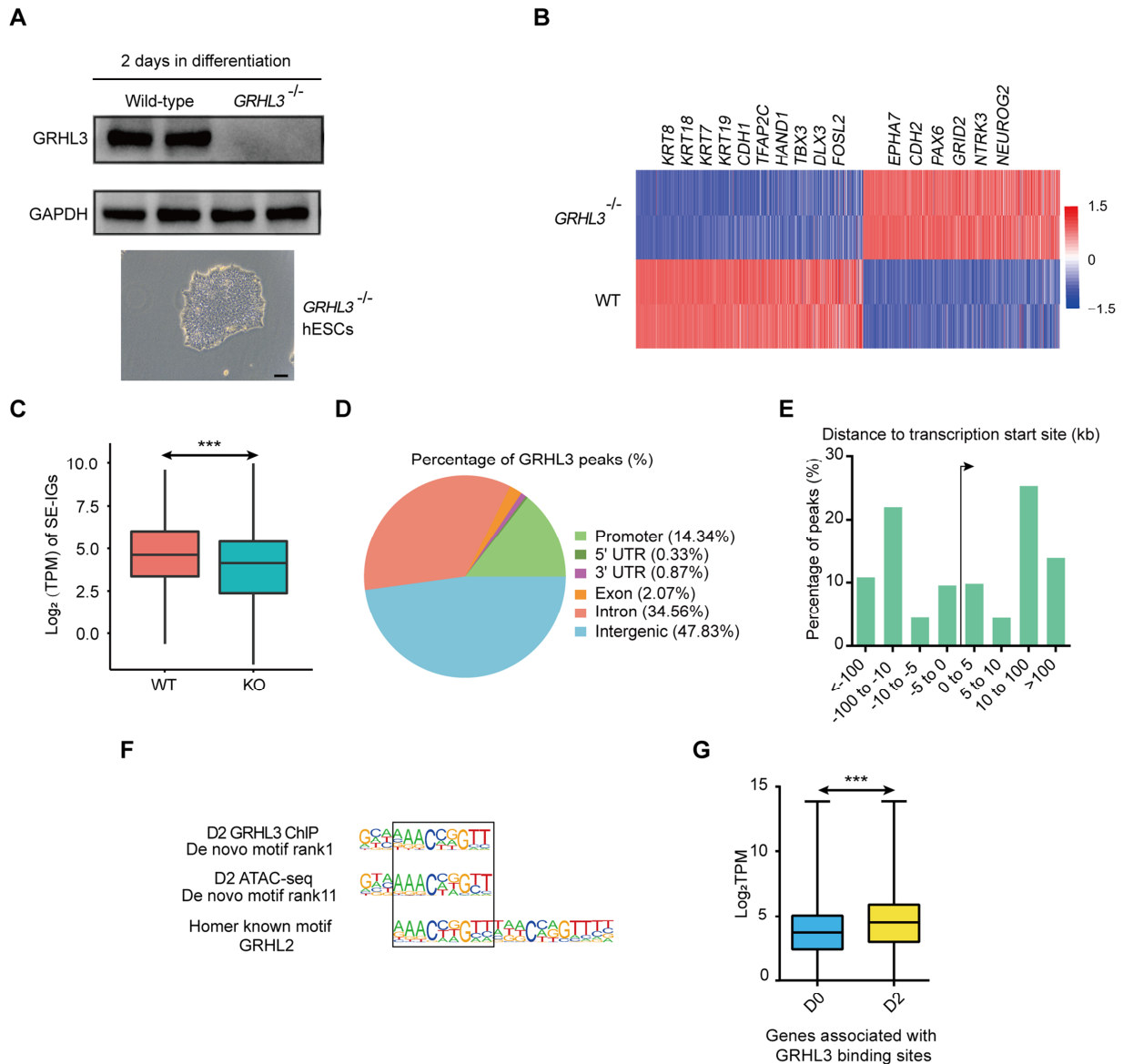


Fig. S3.

Loss of GRHL3 decreases chromatin accessibility in surface ectodermal genes (A) Top panel, western blot analysis of GRHL3 in wild-type and *GRHL3*-knockout hESCs after 2 days differentiation; Bottom panel, phase contrast image of *GRHL3*-knockout hESCs. Scale bar, 100µm. (B) Heatmap of differentially expressed genes in wild-type and *GRHL3*-knockout hESCs after 2 days differentiation. (C) Box plot representing the expression level of SE identity genes in wild-type and *GRHL3*-knockout hESCs after 2 days differentiation. (D) Pie chart showing the distribution patterns of GRHL3 peaks of SE-initiating cells. (E) Histogram showing the distribution of the distance between GRHL3 peaks and transcription start site. (F) *De novo* motif-enrichment analysis of GRHL3-binding sites (top). *De novo* motif-enrichment analysis of ATAC-seq peaks in SE-initiating cells (middle). The GRHL2 known motif identified by HOMER (bottom). (G) Box plot representing the expression level of genes associated with GRHL3 peaks in hESCs and SE-initiating cells.

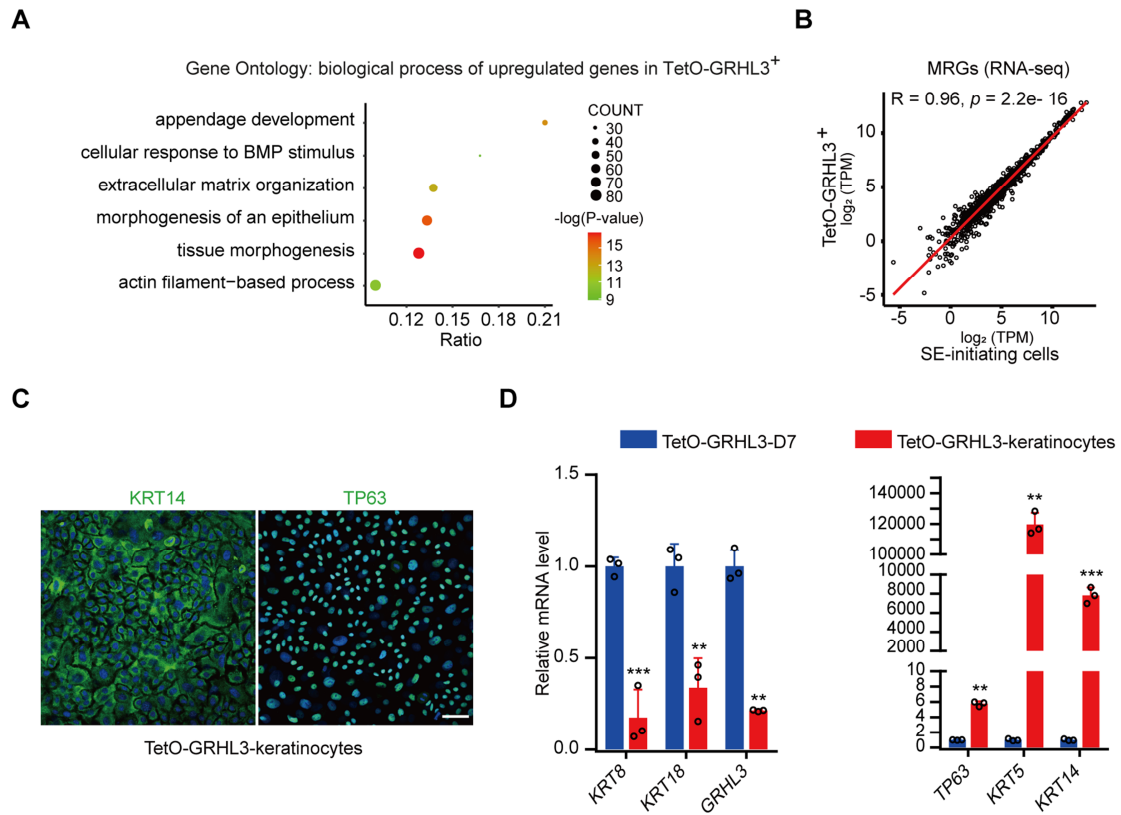


Fig. S4.

Differentiation of GRHL3-induced SE cells into keratinocytes (A) Gene ontology (biological process) analysis of the upregulated genes in TetO-GRHL3⁺ cells. (B) Scatterplot of gene expression from RNA-seq of MRGs in TetO-GRHL3⁺ cells and SE-initiating cells with Pearson correlation value. (C) Immunofluorescence staining of KRT14 and TP63 in keratinocytes derived from TetO-GRHL3⁺ cells (TetO-GRHL3-keratinocytes). Scale bar, 50um. (D) Comparison of gene expression level in TetO-GRHL3⁺ and TetO-GRHL3-keratinocytes. qRT-PCR values were normalized to the values in TetO-GRHL3⁺ cells. Values are shown as means \pm SD (n = 3 biological replicates; ** $P < 0.01$; *** $P < 0.001$ *t* test).

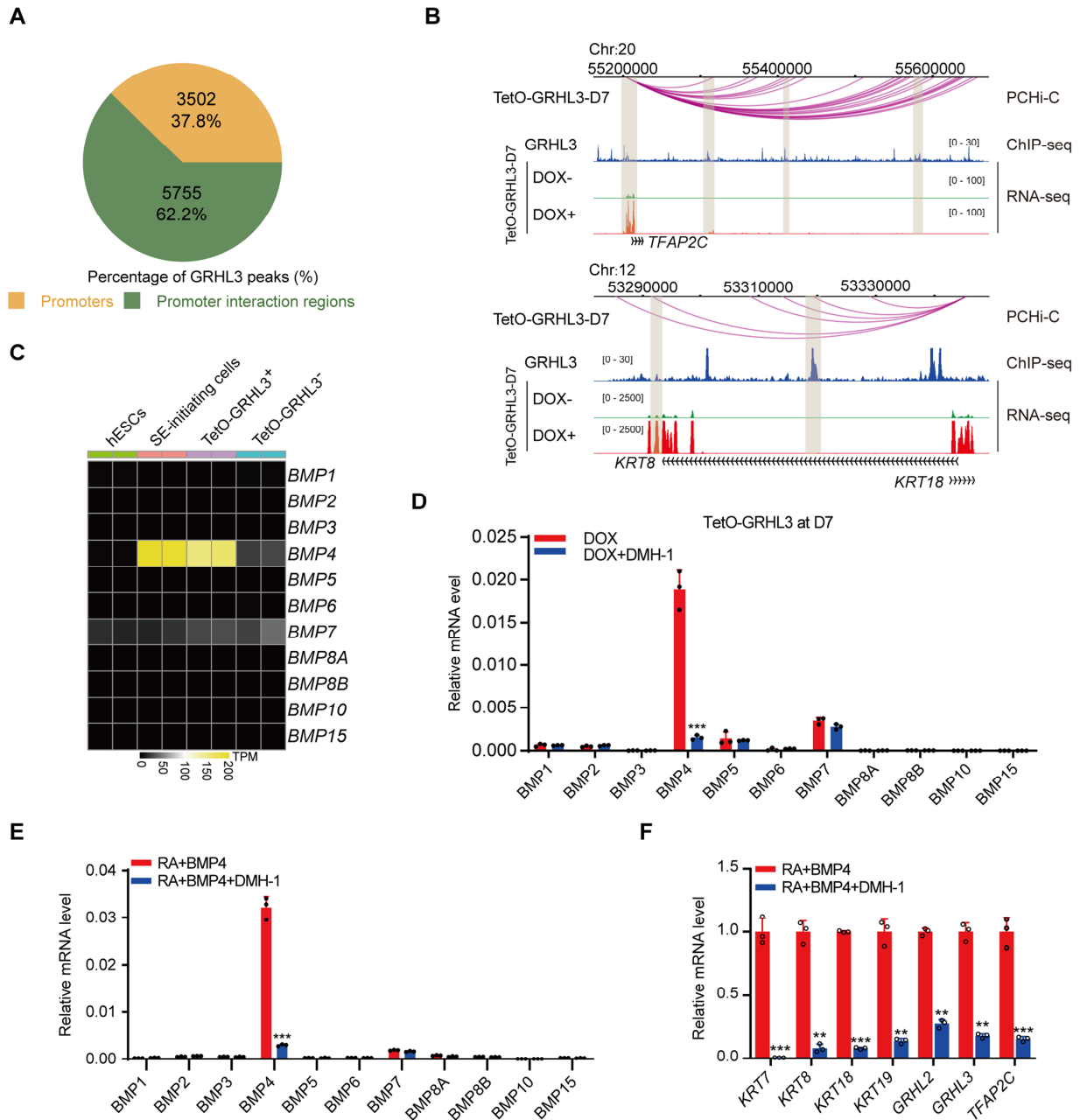


Fig. S5.

Characteristics of GRHL3-mediated promoter interactions (A) Pie chart showing the percentage of GRHL3 peaks in promoter interaction regions or promoters. (B) Genome browser view of promoter interactions, GRHL3, and RNA-seq signals at *TFAP2C* and *KRT8/18* loci. (C) Heatmap of the expression level of BMP family members in hESCs, SE-initiating cells, TetO-GRHL3⁺ and TetO-GRHL3⁻ cells. (D) qRT-PCR analysis of BMP family members in hESCs treated with or without DMH-1 upon SE differentiation for 2 days. qRT-PCR values were normalized to the values in RA/BMP4 group. Values are presented as means \pm SD (n = 3 biological replicates; ***P < 0.001 t test). (E) qRT-PCR analysis of BMP family members in TetO-GRHL3⁺ cells with or without DMH-1 for 7 days. qRT-PCR values were normalized to the

values in control cells. Values are shown as means \pm SD (n = 3 biological replicates; ***P < 0.001 t test). (F) Comparison of gene expression level in hESCs treated with or without DMH-1 upon SE differentiation for 2 days. qRT-PCR values were normalized to the values in SE-initiating cells. Values are shown as means \pm SD (n = 3 biological replicates; **P < 0.01; ***P < 0.001 t test).

table S1

List of primers used in this study.

Primers for RT-qPCR		
Gene Name	Forward	Reverse
<i>ADD2</i>	ATGAGCGAAGAGACGGTCC	TTGCGAAGGCGCATGTACT
<i>CDX2</i>	GACGTGAGCATGTACCCTAGC	GCGTAGCCATTCCAGTCCCT
<i>BMP4</i>	ATGATTCCTGGTAACCGAATGC	CCCCGTCTCAGGTATCAAACCT
<i>COL3A1</i>	GGAGCTGGCTACTTCTCGC	GGGAACATCCTCCTTCAACAG
<i>DLX3</i>	CTCGCCCAAGTCGGAATATAC	CTGGTAGCTGGAGTAGATCGT
<i>DSP</i>	GCAGGATGTAATACTCTCGGC	CCTGGATGGTGTCTGGTTCT
<i>EPAS1</i>	CGGAGGTGTTCTATGAGCTGG	AGCTTGTGTGTTTCGCAGGAA
<i>FOSL2</i>	CAGAAATTCGGGTAGATATGCC	GGTATGGGTTGGACATGGAGG
<i>GAPDH</i>	CTGAGAACGGGAAGCTTGT	GGGTGCTAAGCAGTTGGT
<i>GATA3</i>	GCGGGCTCTATCACAAAATGA	GCCTTCGCTTGGGCTTAAT
<i>GRHL2</i>	TCAATACCCGAAGAGCCTACA	CTTGGCTGTCACTTGCTTTGC
<i>GRHL3</i>	GCCAGTTCTACCCCGTCA	GTCAATGACCCGCTGCTT
<i>HAND1</i>	CCATGCTCCACGAACCCTTC	CCTGGCGTCAGGACCATAG
<i>KRT5</i>	ATCTCTGAGATGAACCGGATGATC	CAGATTGGCGCACTGTTTCTT
<i>KRT7</i>	TCCGCGAGGTCACCATTAAC	GCTCTGTCAACTCCGTCTCAT
<i>KRT8</i>	GATCGCCACCTACAGGAAGCT	ACTCATGTTCTGCATCCAGACT
<i>KRT14</i>	TGCCGAGGAATGGTTCTTCACC	GCAGCTCAATCTCCAGGTTCTG
<i>KRT18</i>	CCGTCTTGCTGCTGATGACT	GGCCTTTTACTTCTCTTCGTG
<i>KRT19</i>	AACGGCGAGCTAGAGGTGA	GGATGGTCGTGTAGTAGTGGC
<i>NANOG</i>	AAGGTCCCGGTCAAGAAACAG	CTTCTGCGTCACACCATTGC
<i>NR2F2</i>	TCATGGGTATCGAGAACATTTGC	TTCAACACAAACAGCTCGCTC
<i>TFAP2A</i>	CAGATATGCAAAGAGTTCACCGAC	TCAAGCAGCTCTGGATGCC
<i>TFAP2B</i>	CCATCCCGGAATGGAAGACG	TCACCGATTTGGGAGGAACTG
<i>TFAP2C</i>	AGATTGGGTTGAATCTTCCG	GGCTTCACAGACATAGGCAA
<i>TP63</i>	TTTCCCACCCGAGATGA	TGCGGCGAGCATCCAT
<i>TUBB4A</i>	CCGGACAACCTTCGTGTTTGG	TCGCGGATCTTACTGATGAGC
<i>BMP1</i>	GGGTCATCCCCTTTGTCAATTG	GCAAGGTCGATAGGTGAACACA
<i>BMP2</i>	ACCCGCTGTCTTCTAGCGT	TTTCAGGCCGAACATGCTGAG
<i>BMP3</i>	GCAGGGAGAGAGACCGAAG	TGGACCGTGCTGTACCTGT
<i>BMP5</i>	GCTGCTGGGTTCTAGTGGG	TTCGTGGTTCCGTAGTCTTCTA
<i>BMP6</i>	AGCGACACCACAAAGAGTTCA	GCTGATGCTCCTGTAAAGACTTGA
<i>BMP7</i>	GGAACGCTTCGACAATGAGAC	GCAGGAAGAGATCCGATTCCC
<i>BMP8A</i>	CACCTTCTCATCTGGATCG	CAGGAAGTAGGCACCGAGAG
<i>BMP8B</i>	AGGTGGCTTCCTTATCTGCG	ATGTGCCAACTCTGCTTCGT
<i>BMP10</i>	CCTCTGCCAACATCATTAGGAG	TTTTCGGAGCCCATTAATAACTGA
<i>BMP15</i>	TGTGAACTCGTGCTTTTCATG	CTCAATCAGGGGCAAAGTAGG

Supplemental Tables:

Table S2: Gene list of MRGs.

Table S3: Gene list of SE identity genes.

Table S4: The annotation of GRHL3 peaks.

Table S5: Gene list of TetO-GRHL3⁺ upregulated genes.