

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

Full title: Integrated Single Cell and Spatial Transcriptomics Reveal Autoreactive Differentiated B Cells in Joints of Early Rheumatoid Arthritis

Authors: Uta Hardt^{1,2,#,*}, Konstantin Carlberg, PhD^{3,#}, Erik af Klint, MD, PhD¹, Peter Sahlström¹, Ludvig Larsson, PhD³, Annika van Vollenhoven¹, Susana Hernandez Machado¹, Lena Israelsson¹, Khaled Amara, PhD¹, Karine Chemin, PhD¹, Marina Korotkova, MD, PhD¹, Gunilla B Karlsson Hedestam, PhD², Anca I Catrina, MD, PhD¹, Sarah A Teichmann, PhD⁴, Patrik L Ståhl, PhD³, Vivianne Malmström, PhD^{1,*}

Author affiliations: ¹Division of Rheumatology, Department of Medicine Solna, Center for Molecular Medicine, Karolinska Institutet, Stockholm, Sweden and Karolinska University Hospital, Stockholm, Sweden. ²Department of Microbiology, Tumor and Cell Biology, Karolinska Institutet, Stockholm, Sweden. ³Department of Gene Technology, KTH Royal Institute of Technology, Science for Life Laboratory, Stockholm, Sweden. ⁴Wellcome Sanger Institute, Wellcome Genome Campus, Hinxton, Cambridge CB10 1SA, United Kingdom. [#]These authors contributed equally.

Corresponding authors: Uta Hardt, PhD student, Departments of Medicine, Solna, and Microbiology, Tumor and Cell Biology, Karolinska Institutet, uta.hardt@ki.se, Tel: +46 8 524 869 43, Dr. Vivianne Malmström, Professor, Department of Medicine, Solna, Center for Molecular Medicine, Karolinska Institutet, vivianne.malmstrom@ki.se, Tel: +46 8 517 756 09

This file includes: Figs. S1 to S17, Tables S1 to S3

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

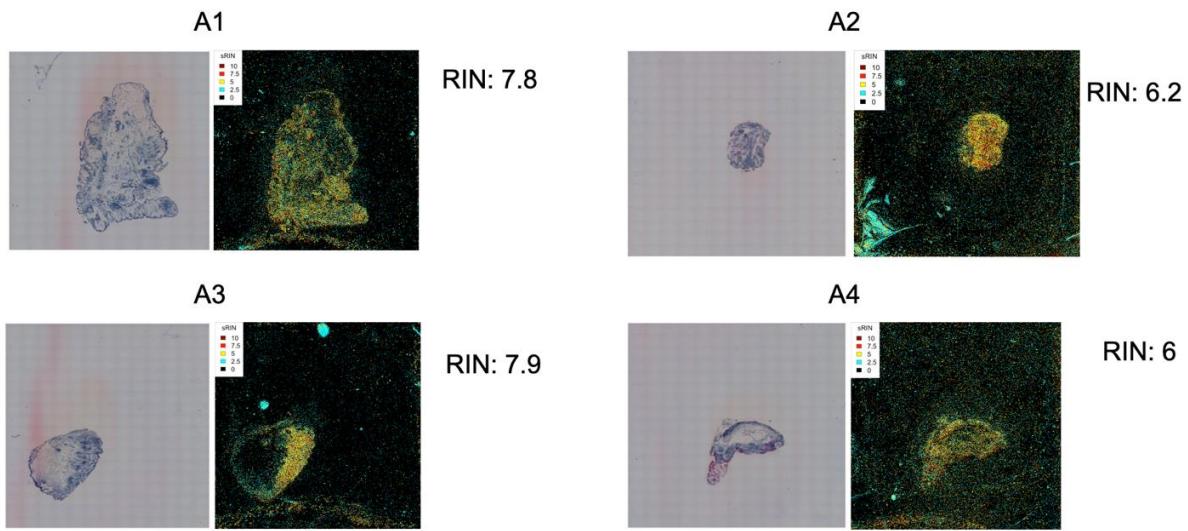


Fig. S1.

Spatial RIN. Heatmap over the spatial RIN scores with bulk RIN values from extracted material shown next to the heatmaps

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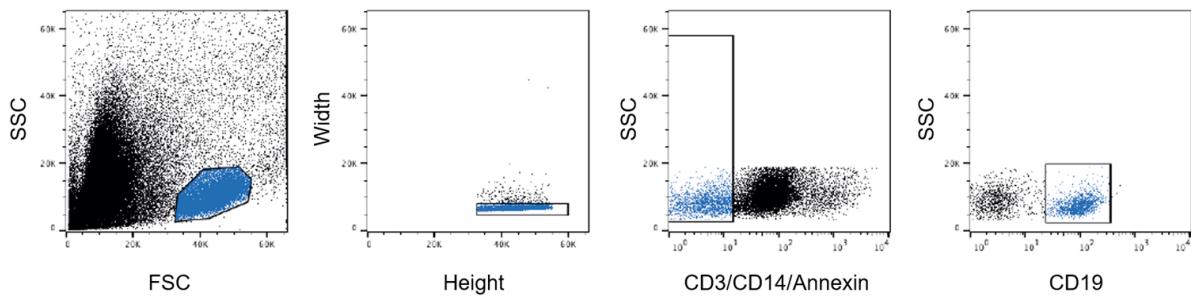


Fig. S2.

The gating strategy for the B cell sort is shown. We selected AnnexinV-CD3-CD14-CD19+ single lymphocytes.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

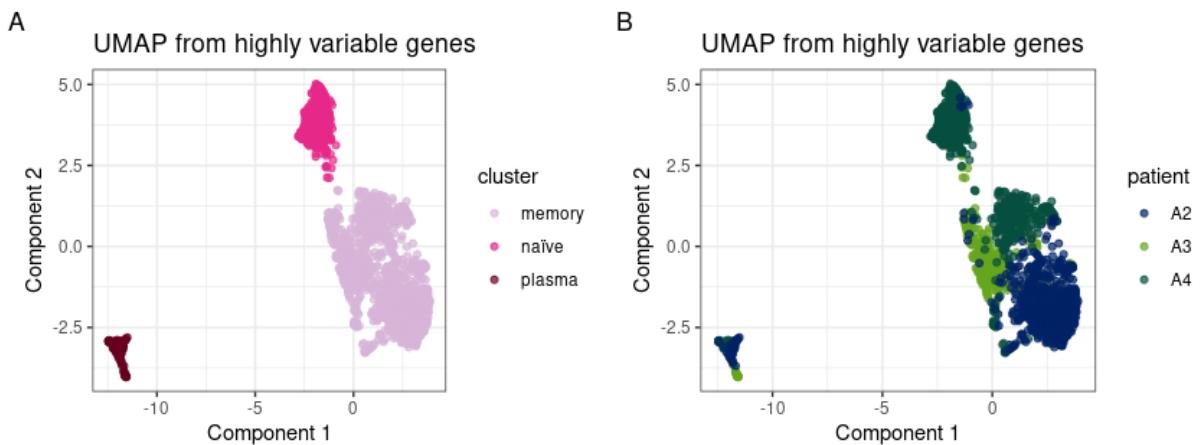


Fig. S3.

UMAP of scRNAseq data coloured by **(A)** cell cluster and **(B)** patient contribution.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

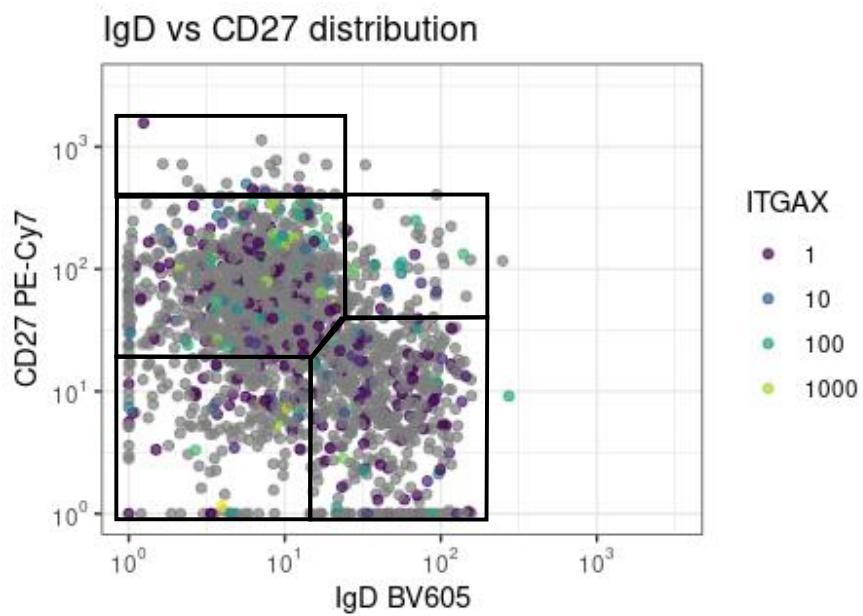


Fig. S4.

ITGAX (encoding CD11c) expression was projected onto the IgD vs CD27 distribution.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

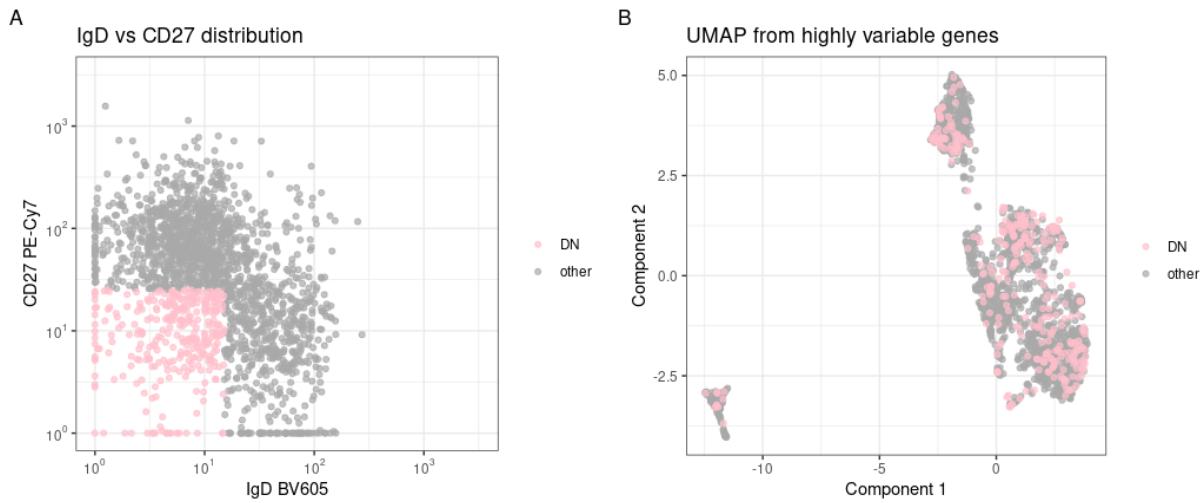


Fig. S5.

The double negative population in the scRNAseq data is shown.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

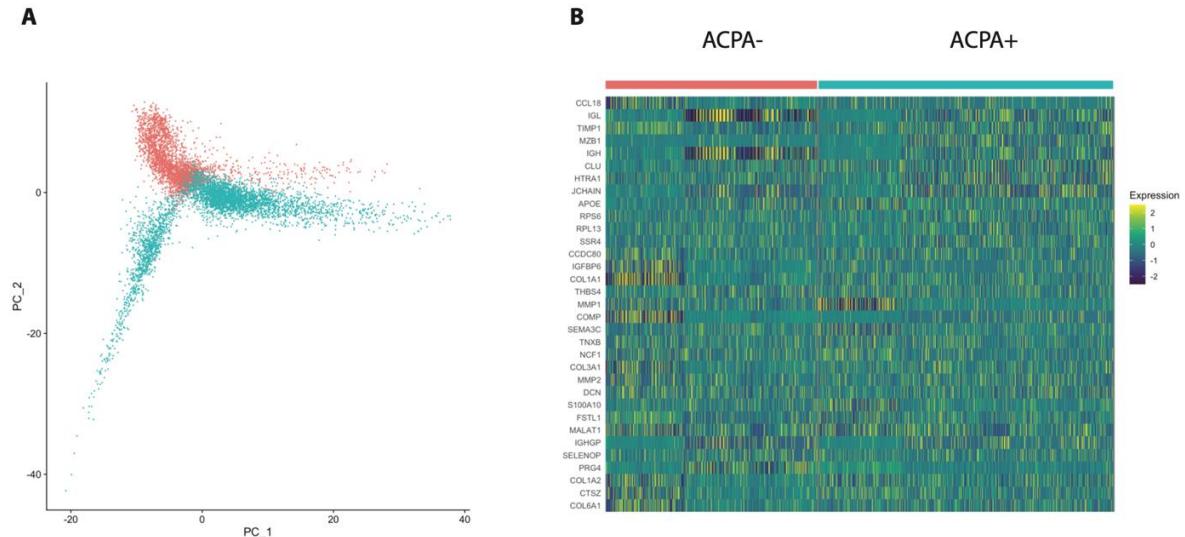


Fig. S6.

(A) PCA labelled by serotype (red ACPA-, turquoise ACPA+) showing overlaps and separation between data points for each group. (B) Heatmap over the marker genes for each group with little differences between ACPA+ and ACPA- RA.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

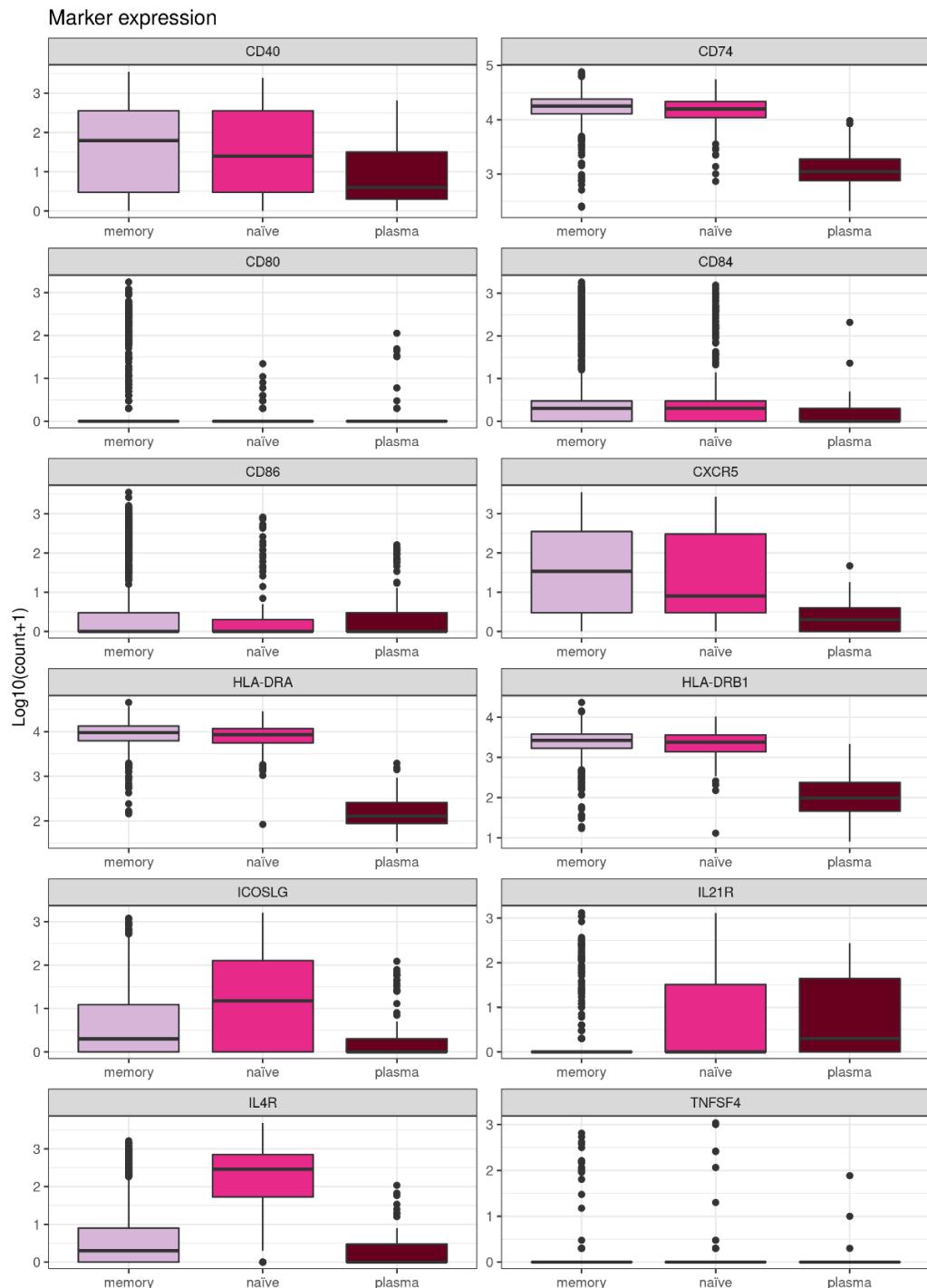
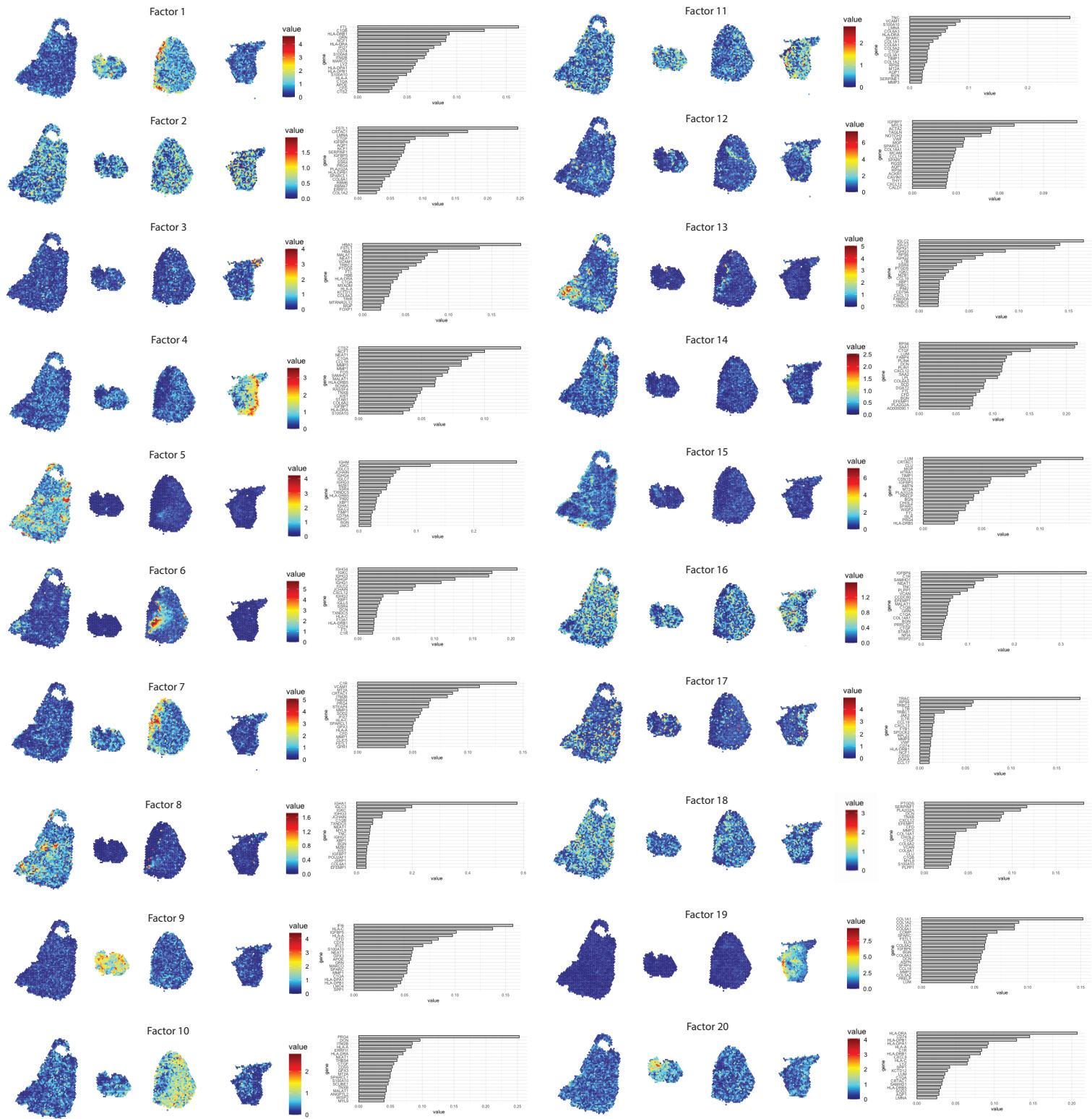


Fig. S7.

Logarithmic read count for B-T cell interaction markers are shown grouped by cell subset. Most markers are present in the naïve and the memory cell subset.



Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

Fig. S8.

The factor analysis for spatial transcriptomics is shown. Most factors (e.g. 2,11,16,18) are represented on all tissues. Several factors have expression activities overlapping with distinct morphological areas such as infiltrates, connective tissue as well as areas bordering the infiltrates. These could be regarded as baseline tissue signals with *CD55* being of particular interest as it marks the intimal lining fibroblast-like synoviocytes. Three factors (6,8,13) shows a strong activity towards the immune cell-rich areas in biopsy A1. These areas correlate with *IGH*, *IGL*, *JCHAIN*, *MZB1*, *XBP1* and *CD79A* expression suggesting plasma cell presence. Factor 20 has strong activity within cluster 4 for biopsy A2. Here the driver genes are the plasma cell markers mentioned before as well as *HLA-DRB1* and *CD74* further highlighting antigen presentation. Biopsy A3 present factor 6 with strong activity in lymphocyte aggregates. Factor 6 is among others associated with the HLA-associated *CD74* gene. For biopsy A4, two of the factors (4,19) have genes associated with connective tissue and several HLA genes.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

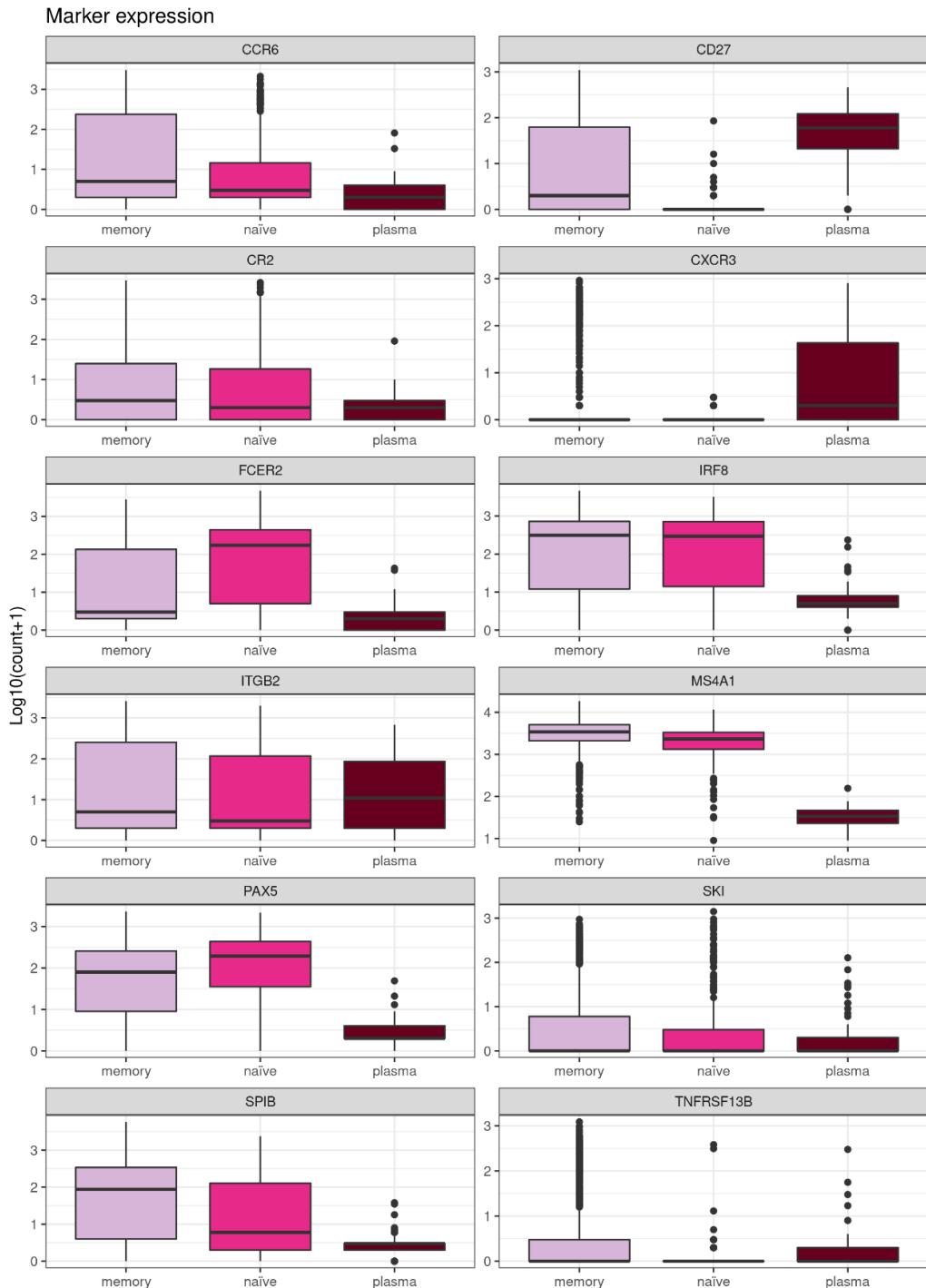


Fig. S9.

Logarithmic read count for memory B cell markers is shown grouped by cell subset. Most memory markers are either shared with the naïve or the plasma cell subset.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

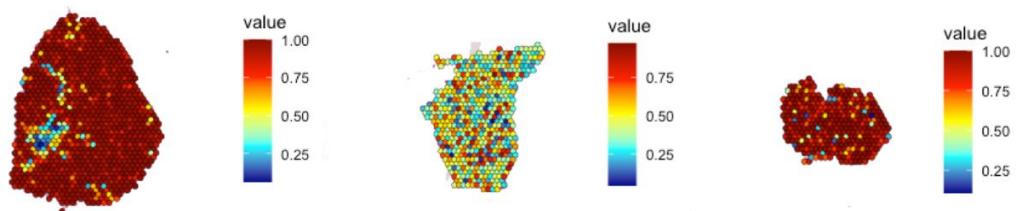


Fig. S10.

The memory B cell subset from paired data from the same individual was predicted on the tissue sections. The color bar represents the prediction score for memory B cells on the tissue sections.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

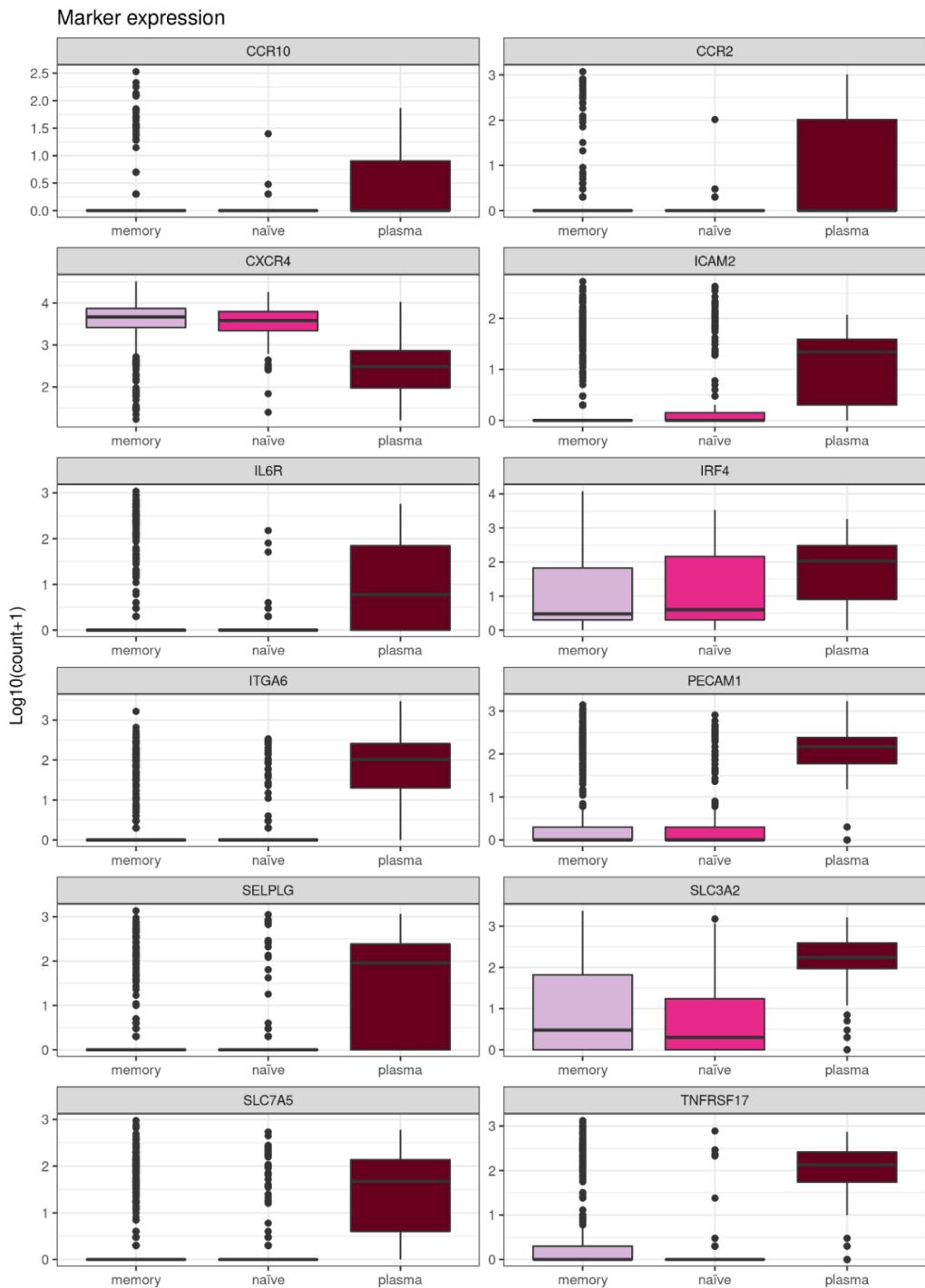


Fig. S11.

Logarithmic read count for plasma cell markers is shown grouped by cell subset. Most markers are clearly exclusive to plasma cells.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

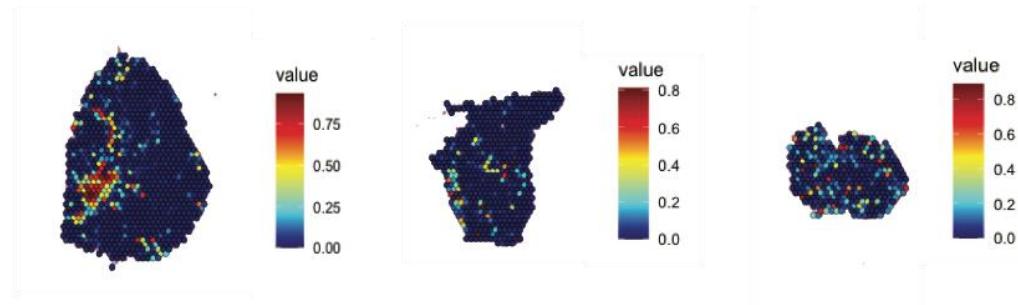
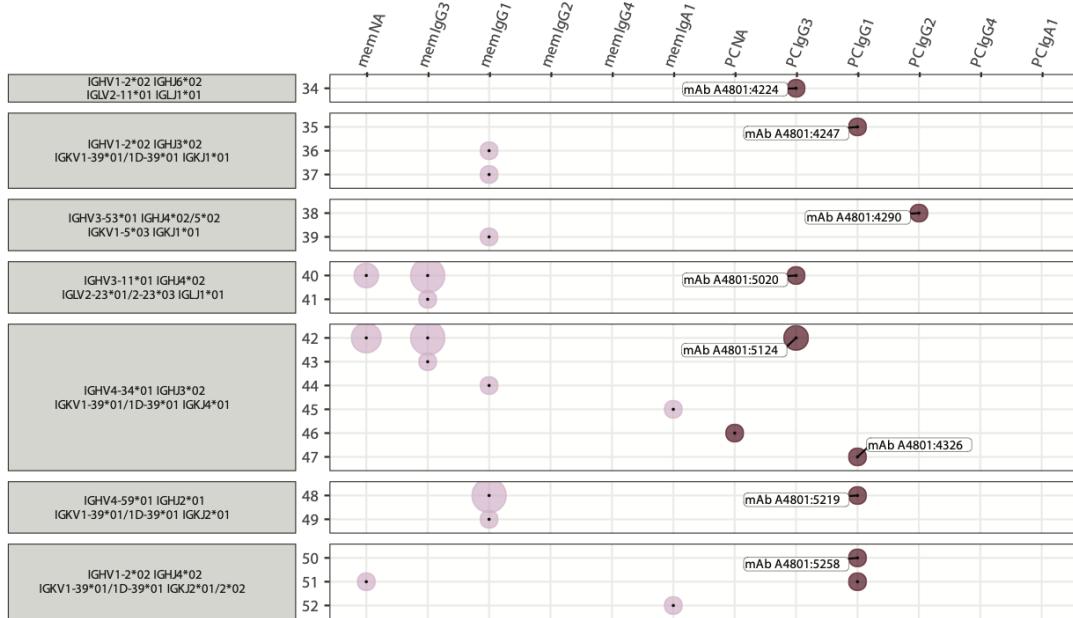


Fig. S12.

The plasma cell subset from paired data from one individual was predicted on the tissue sections.

The color bar represents the prediction score for plasma cells on the tissue sections. Integration showed a clear correlation with the Zhang et al.¹⁷ data.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis



Clone	HCDR3	VH SHM (%)	LCDR3	VL SHM (%)
34	<u>AGTNYDFWSGRPWYYYYGMDV</u>	1/294 (0.3)	CSYAGSYTYV	0/295 (0.0)
35	VRAARGSGWFHDVF DI	6/295 (2.0)	QQSYSTPRT	3/284 (1.1)
36	ARANFGGLRGYDAFDI	6/295 (2.0)	QQSYSTRT	3/283 (1.1)
37	<u>ARANFGGLRGYDAFDI</u>	8/295 (2.7)	QQSYSTRT	2/283 (0.7)
38	AKDGSSRSLVS	10/293 (3.4)	QQYNSFPWT	11/280 (3.9)
39	<u>AKDGSSRSLVS</u>	15/293 (5.1)	QQYNSYSPTWT	1/287 (0.3)
40	ARDKRRRH YDSSGYHFDY	6/296 (2.0)	CSYAGSSTS LYV	1/295 (0.3)
41	<u>ARDKRRRH YDSSGYHFDY</u>	6/295 (2.0)	CSYAGSSTS LYV	1/295 (0.3)
42	ARARSGISAIHGVFDI	3/293 (1.0)	QQSYSTPPT	4/287 (1.4)
43	<u>ARARSGISAIHGVFDI</u>	3/285 (1.1)	QQSYSTPPT	4/287 (1.4)
44	ARARSGLGATHGAFDI	7/292 (2.4)	QQSYSTPPT	1/287 (0.3)
45	ARGGGYGGNSVRRAFDI	28/293 (9.6)	QQTYITPRA	21/284 (7.4)
46	ARARSGLGATHGAFDI	4/292 (1.4)	QQSYSTPPT	4/287 (1.4)
47	<u>ARARSGLGATHGAFDI</u>	6/292 (2.1)	QQSFSTPPT	4/287 (1.4)
48	ARGKTGYSSRNHWYFDL	2/292 (0.7)	QQSSSTPMYT	2/284 (0.7)
49	<u>ARGKTGYSSRNHWYFDL</u>	2/292 (0.7)	QQSYSTPLST	2/284 (0.7)
50	ARGIMQFPPDY	2/293 (0.7)	QQSYSTPQT	4/286 (1.4)
51	<u>ARGIVGQFPPDH</u>	8/293 (2.7)	QQSYSTPQT	2/286 (0.7)
52	AREVGVGAILDY	30/296 (10.1)	QQSYSTPYT	15/284 (5.3)

Fig. S13.

The plot shows the equivalent clonotype information that is shown in figure 6A for the seven plasma cell derived monoclonal antibodies originating from A3.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

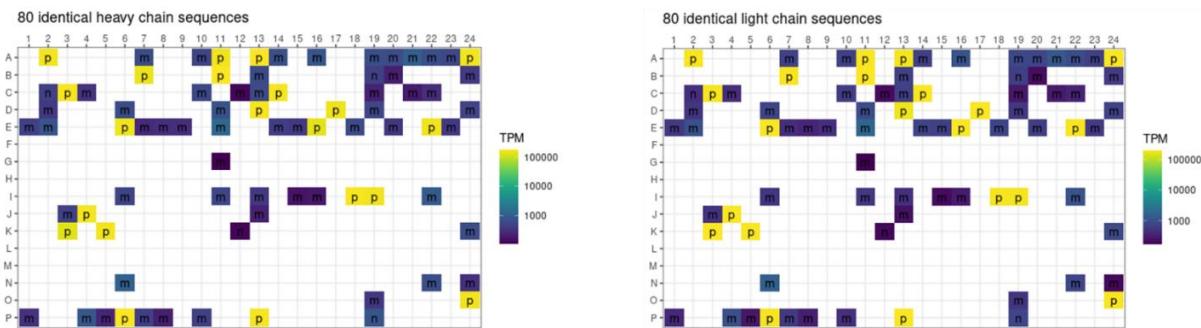


Fig. S14.

No index-hopping for monoclonal antibody A4797:2017. Cells expressing the A4797:2017 antibody were derived from plasma cells (p), memory B cells (m) and naïve B cells (n).

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

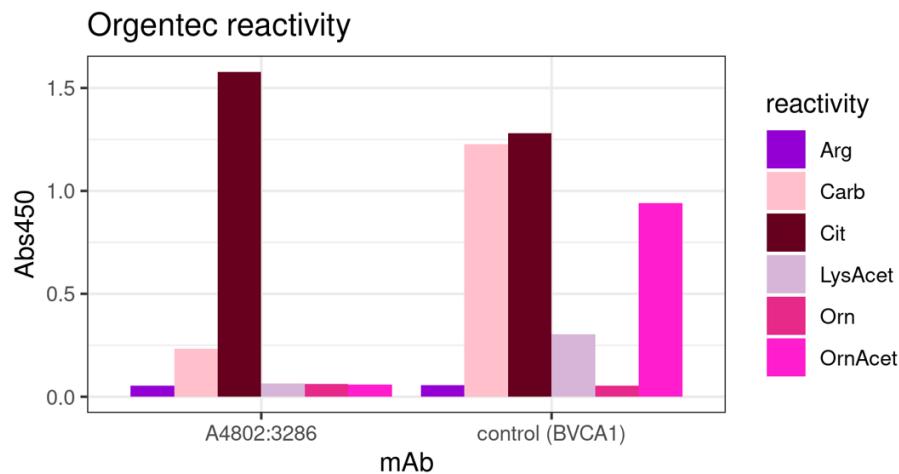


Fig. S15.

The Orgentec reactivity of the A4802:3286 clone compared to control (BVCA1²²) is shown. The ACPA clone was found to be negative for the acetylated peptide, weakly positive for the carbamylated peptide but strongly positive for the citrullinated peptide.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

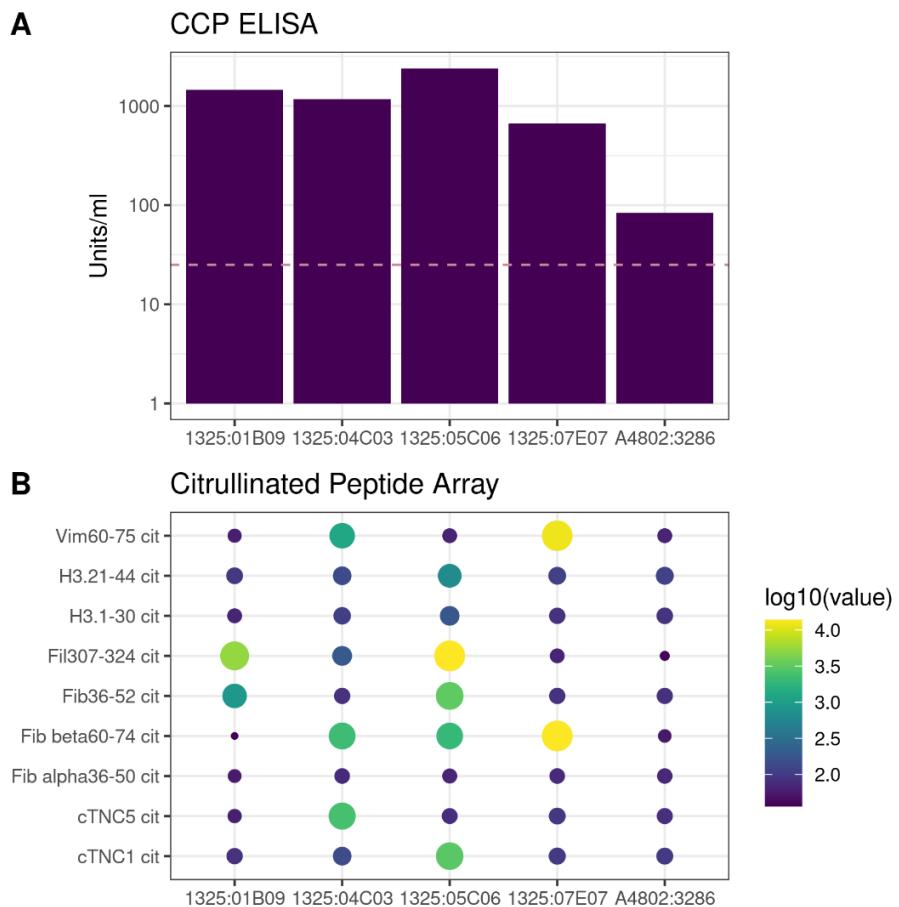


Fig. S16.

(A) CCP2 ELISA and (B) citrullinated peptide array²⁰ to test antigen specificity. We compared the identified A4802:3286 ACPA with previously described ACPAs generated from plasma cells from synovial fluid of RA patients with long lasting disease⁴⁰. The cut-off for the anti-CCP2 ELISA was 25 units/ml. On the citrullinated peptide array, we assessed reactivity towards citrullinated Vimentin (Vim), citrullinated Histone 3 (H3), citrullinated Filaggrin (Fil), citrullinated Fibrinogen (Fib) and citrullinated Tenascin-C (TNC). Numbers indicate amino acid residues.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

mAb	N-X-S/T glycosylation sites			
	germline VH	VH	germline VL	VL
A4797:2017	0	0	0	0
A4797:2216	0	0	0	0
A4801:4224	0	0	0	0
A4801:4247	0	1	0	0
A4801:4290	0	0	0	0
A4801:4326	2	2	0	0
A4801:5020	0	0	0	0
A4801:5124	1	1	0	0
A4801:5219	0	0	0	0
A4801:5258	0	0	0	0
A4802:3086	0	0	0	0
A4802:3286 (ACPA)	0	0	0	2
A4802:3314	0	0	0	0
A4802:6090	0	1	0	1
A4802:6117	0	0	0	0

Fig. S17.

Germline and germline-diverted glycosylation sites in monoclonal antibodies are shown.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

Table S1.

cluster	gene	p_val	avg_logFC	pct_1	pct_2	p_val_adj
0	<i>MALAT1</i>	6.57E-69	-12.05	0.55	0.74	1.20E-64
0	<i>CD74</i>	1.63E-56	-60.97	0.82	0.90	2.97E-52
0	<i>EEF1A1</i>	9.36E-55	-21.51	0.74	0.84	1.70E-50
0	<i>MMP3</i>	3.91E-51	-136.45	0.60	0.75	7.11E-47
0	<i>ACTB</i>	1.67E-49	-19.56	0.80	0.88	3.05E-45
0	<i>B2M</i>	3.88E-48	-70.56	0.94	0.96	7.05E-44
0	<i>TMSB4X</i>	3.90E-48	-14.42	0.69	0.80	7.10E-44
0	<i>COL3A1</i>	6.07E-48	-73.30	0.63	0.77	1.10E-43
0	<i>FTH1</i>	1.73E-47	-23.46	0.67	0.79	3.15E-43
0	<i>HLA-B</i>	7.41E-47	-70.56	0.88	0.93	1.35E-42
1	<i>IGF1</i>	1.24E-77	0.97	0.21	0.06	2.26E-73
1	<i>COL1A1</i>	8.88E-52	190.30	0.77	0.59	1.62E-47
1	<i>MMP2</i>	1.77E-30	25.89	0.56	0.40	3.22E-26
1	<i>COL3A1</i>	5.12E-24	61.33	0.82	0.72	9.31E-20
1	<i>ELN</i>	1.59E-22	7.58	0.23	0.13	2.90E-18
1	<i>FNDC1</i>	3.89E-19	0.31	0.11	0.05	7.08E-15
1	<i>ASPN</i>	4.95E-19	6.42	0.12	0.06	9.02E-15
1	<i>AEBP1</i>	8.85E-19	21.02	0.36	0.25	1.61E-14
1	<i>COMP</i>	3.25E-18	2.98	0.23	0.14	5.92E-14
1	<i>CCL18</i>	3.77E-16	5.23	0.33	0.23	6.86E-12
2	<i>MMP3</i>	2.67E-276	118.78	0.94	0.69	4.86E-272

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

2	<i>MMP1</i>	1.04E-210	36.51	0.72	0.28	1.89E-206
2	<i>FNI</i>	9.72E-146	269.70	0.99	0.98	1.77E-141
2	<i>HTRA1</i>	9.61E-138	-22.09	0.88	0.72	1.75E-133
2	<i>PRG4</i>	7.54E-126	Inf	0.99	0.97	1.37E-121
2	<i>CRTAC1</i>	8.49E-125	18.19	0.85	0.67	1.54E-120
2	<i>MT2A</i>	3.02E-110	25.90	0.85	0.69	5.49E-106
2	<i>CLU</i>	5.26E-110	-76.09	0.95	0.88	9.57E-106
2	<i>VCAM1</i>	9.61E-102	1.96	0.76	0.53	1.75E-97
2	<i>PLA2G2A</i>	5.94E-97	8.88	0.81	0.62	1.08E-92
3	<i>MALAT1</i>	1.21E-125	-4.57	1.00	0.67	2.21E-121
3	<i>ACTB</i>	8.96E-66	-34.59	0.77	0.88	1.63E-61
3	<i>EEF1A1</i>	6.41E-65	-23.05	0.71	0.83	1.17E-60
3	<i>HLA-B</i>	1.12E-58	-108.00	0.87	0.93	2.03E-54
3	<i>RPL13A</i>	4.02E-58	-12.76	0.46	0.68	7.31E-54
3	<i>TMSB4X</i>	3.17E-57	-27.81	0.65	0.79	5.77E-53
3	<i>MT-CO3</i>	5.39E-57	-53.99	0.86	0.92	9.81E-53
3	<i>TMSB10</i>	1.31E-56	-23.63	0.57	0.74	2.38E-52
3	<i>MT-ND1</i>	5.70E-56	-20.54	0.71	0.84	1.04E-51
3	<i>MT-ND4</i>	2.13E-55	-32.26	0.75	0.86	3.87E-51
4	<i>JCHAIN</i>	7.99E-184	42.19	0.78	0.35	1.45E-179
4	<i>IGH</i>	2.73E-141	Inf	0.97	0.84	4.97E-137
4	<i>MZB1</i>	6.11E-97	2.10	0.49	0.19	1.11E-92
4	<i>IGHGP</i>	2.00E-92	74.42	0.42	0.15	3.63E-88

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

4	<i>TXNDC5</i>	5.11E-77	9.17	0.57	0.29	9.30E-73
4	<i>DERL3</i>	4.95E-72	0.67	0.26	0.07	9.02E-68
4	<i>CD79A</i>	4.75E-58	0.57	0.24	0.07	8.64E-54
4	<i>XBP1</i>	4.74E-57	4.20	0.47	0.24	8.62E-53
4	<i>PIM2</i>	1.03E-52	0.93	0.25	0.08	1.88E-48
4	<i>SDC1</i>	1.82E-50	0.93	0.23	0.07	3.31E-46
5	<i>HLA-DQA1</i>	5.13E-72	1.70	0.62	0.34	9.34E-68
5	<i>LYZ</i>	3.39E-66	4.87	0.68	0.42	6.17E-62
5	<i>CD74</i>	3.22E-56	2.16	0.94	0.88	5.86E-52
5	<i>HLA-DRA</i>	6.95E-44	31.21	0.90	0.87	1.26E-39
5	<i>CXCL9</i>	3.52E-41	2.41	0.41	0.21	6.41E-37
5	<i>HLA-DRB1</i>	2.76E-38	5.17	0.78	0.67	5.02E-34
5	<i>FTH1</i>	3.77E-37	1.21	0.84	0.76	6.86E-33
5	<i>FTL</i>	1.95E-36	2.99	0.90	0.86	3.55E-32
5	<i>HLA-DPA1</i>	2.04E-36	6.94	0.75	0.60	3.71E-32
5	<i>TMSB10</i>	4.28E-36	11.04	0.82	0.71	7.79E-32
6	<i>IGFBP7</i>	1.01E-164	2.96	0.82	0.40	1.84E-160
6	<i>ACTA2</i>	1.87E-148	4.21	0.48	0.12	3.40E-144
6	<i>TAGLN</i>	1.90E-112	1.93	0.42	0.12	3.46E-108
6	<i>MCAM</i>	7.17E-101	0.81	0.33	0.08	1.30E-96
6	<i>VWF</i>	1.60E-86	2.41	0.35	0.10	2.91E-82
6	<i>MYL9</i>	1.06E-73	2.30	0.64	0.34	1.92E-69
6	<i>NR2F2</i>	5.89E-58	0.61	0.20	0.05	1.07E-53

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

6	<i>ACKR1</i>	4.38E-49	0.64	0.19	0.05	7.97E-45
6	<i>CCL19</i>	4.31E-48	17.11	0.29	0.11	7.85E-44
6	<i>NOTCH3</i>	1.16E-40	0.71	0.30	0.12	2.12E-36
7	<i>PREL</i>	9.81E-63	5.75	0.56	0.17	1.78E-58
7	<i>HTRA1</i>	2.30E-53	27.27	0.96	0.73	4.18E-49
7	<i>PRG4</i>	3.43E-26	-Inf	0.99	0.97	6.24E-22
7	<i>CSN1S1</i>	2.43E-22	5.88	0.25	0.08	4.43E-18
7	<i>FNI</i>	2.58E-22	-264.68	0.99	0.98	4.70E-18
7	<i>AMTN</i>	2.65E-22	10.23	0.21	0.06	4.82E-18
7	<i>CRTAC1</i>	1.36E-21	-13.01	0.80	0.69	2.48E-17
7	<i>CLU</i>	8.46E-20	81.27	0.90	0.89	1.54E-15
7	<i>IGFBP5</i>	1.34E-19	-2.67	0.67	0.46	2.44E-15
7	<i>SLC39A14</i>	1.70E-19	-0.47	0.47	0.24	3.10E-15
8	<i>FABP4</i>	1.51E-47	5.49	0.26	0.05	2.75E-43
8	<i>PLIN4</i>	1.03E-41	0.42	0.19	0.03	1.88E-37
8	<i>LPL</i>	2.04E-37	0.79	0.15	0.02	3.71E-33
8	<i>C3</i>	1.15E-33	-0.51	0.59	0.27	2.09E-29
8	<i>SCD</i>	1.48E-23	0.74	0.26	0.08	2.70E-19
8	<i>SAA1</i>	6.65E-19	0.86	0.18	0.05	1.21E-14
8	<i>B2M</i>	4.73E-12	-155.66	0.96	0.96	8.60E-08
8	<i>HLA-A</i>	8.23E-11	-65.66	0.69	0.80	1.50E-06
8	<i>HLA-B</i>	1.13E-10	-119.66	0.91	0.92	2.05E-06
8	<i>HLA-E</i>	6.72E-10	-4.90	0.56	0.68	1.22E-05

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

9	<i>HBA2</i>	0.00E+00	2.00	1.00	0.05	0.00E+00
9	<i>HBA1</i>	7.79E-33	1.84	0.14	0.02	1.42E-28
9	<i>MMP2</i>	1.82E-08	-26.12	0.24	0.43	3.32E-04
9	<i>CLU</i>	3.92E-08	-149.47	0.78	0.89	7.14E-04
9	<i>CD68</i>	8.59E-08	-4.06	0.19	0.36	1.56E-03
9	<i>ENO1</i>	1.02E-07	-4.21	0.26	0.44	1.86E-03
9	<i>PSAP</i>	1.39E-07	-24.50	0.72	0.81	2.53E-03
9	<i>SAT1</i>	1.73E-07	-4.67	0.29	0.46	3.16E-03
9	<i>CTSB</i>	2.22E-07	-19.17	0.67	0.77	4.04E-03
9	<i>HTRA1</i>	2.70E-07	-96.38	0.66	0.74	4.91E-03

Top 10 differentially expressed genes for 10 spatial transcriptomics clusters are shown.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

Table S2.

	OR	p-value
Naïve B cells (SC-B1)	20.3	<2.2e-16
Memory B cells (SC-B2)	99.6	<2.2e-16
Plasma cells (SC-B4)	3.27	<2.2e-16

Fisher's exact test for memory B cells and the naïve B cells, memory B cells and plasma cells clusters from previously published studies ¹⁷ is shown.

Supplementary Materials for Multimodal Synovial B Cell Sequencing in Arthritis

Table S3.

	OR	p-value
Naïve B cells (SC-B1)	0.438	1
Memory B cells (SC-B2)	0	1
Plasma cells (SC-B4)	45.1	<2.2e-16

Fisher's exact test for plasma cells and the naïve B cells, memory B cells and plasma cell clusters from previously published studies ¹⁷ is shown.