

SUPPLEMENTAL FIGURE TITLES & LEGENDS

Figure S1. *Allobaculum* sp. 128 does not bloom during inflammation, and a second UC patient *Allobaculum* isolate is colitogenic in gnotobiotic mice, Related to Figure 1. (A) Time course of fecal microbiota profiling of MC+Allo colonized mice across 7 days of 2% DSS administration (n=4 mice/group). (B) Time course of fecal lipocalin (LCN-2) overlaid with *Allobaculum* sp. 128 abundance. (C) H&E-stained colon sections from *Rag1*^{-/-} mice on d7 of DSS administration. Scale bars, 200µm. (D-E) Colon lamina propria CD4⁺IL-17A⁺ T cells from *Il10*^{-/-} mice after 8 weeks colonization (n=5 mice/group). (F-G) GF mice were monocolonized with either *Ery47* (control bacteria) or *Allobaculum* sp. 128 for 7 days before the induction of acute colitis via 2% DSS administration. Colonic inflammation was assessed by fecal lipocalin on d3 (F) and colon length on d7 (G) (n=5 mice/group). (H) Phylogenetic tree constructed using 16S rRNA gene sequences from members of family *Erysipelotrichaceae*, using maximum likelihood estimation, bootstrapped (BS) to 1,000 replicates. BS values are shown along branches. (I) Experimental schematic. (J) Second *Allobaculum* isolate (Allo2) was used for acute colitis model in WT gnotobiotic mice as in Figure 1C-1H. Three of six mice colonized with Allo2 were found dead (3 F.D.) prior to endpoint. (K) Fecal lipocalin, LCN2, was measured longitudinally by ELISA. Error bars show Mean ± SEM. Data shown in (A-B) from N=1 experiment. Data in (D-E) from one of N=2 experiments. Data in (F-G) from N=1 experiment. Data shown in (J-K) from one of N=2 independent experiments, n=5-6 mice per group.

Figure S2. Unremarkable histopathology and total serum Ig of untreated *Allobaculum* sp. 128-colonized WT mice, Related to Figure 1. (A) Bouin's-fixed H&E-stained colon sections from WT mice colonized with MC bacteria or MC+*Allobaculum* sp. 128, euthanized 12 weeks later. Scale bars, 200µm. (B) Blinded scoring for colitis. One experiment shown is representative of N=2 independent experiments. (C-D) Total serum IgA and IgG content at 7 weeks (n=3-4 mice/group). (E) FITC-dextran concentration in serum 1h after oral gavage (n=4-8 mice/group). (F) *Allobaculum* sp. 128-specific IgG in fecal water at 7 weeks post-colonization with MC or MC+Allo (n=5 mice/group). (G) Thickness of the colonic inner mucus layer after 2 weeks of colonization (tissues fixed in Carnoy's solution to preserved the inner mucus layer; n=10-12 mice/group). Welch's t-test was used to compare microbiota groups at each time point. ** $P < 0.01$, * $P < 0.05$. Data shown in (A-B) are from one representative of N=3 experiments. Data shown in (C-D) show one of N=2 experiments. Data in (E) are from N=1 experiments. Data in (F) show one of N=2 experiments. Data in (G) are compiled from N=2 experiments.

Figure S3. Microbial diversity cannot explain the relationship between *Allobaculum* and *A. muciniphila*, *A. muciniphila* and *Allobaculum* co-colonize the ileal mucosa, and co-colonization has minimal impacts on *A. muciniphila* and *Allobaculum* gene expression, Related to Figures 3 & 4. (A) Genus-level richness, Simpson's diversity index, Shannon's diversity index, and evenness of each microbiome that contained *Allobaculum* (n=14) or lacked *Allobaculum* sp. 128 (n=5). (B) Legend accompanying Fig. 4B. Data shown from N=1 experiment. (C) GF WT mice were mono- or bi-colonized as shown for 10 days. Terminal ilea were fixed and sections stained with bacterial FISH probes EUB338 (to stain *Allobaculum*) and VP403 (to stain *A. muciniphila*). Scale bars, 10µm. (D) *In vivo* bacterial transcriptomes from the

ileum and colon were compared for differential expression of ORFs across single colonization or co-colonization conditions (MC+Allo vs MC+Both, and MC+Akk vs. MC+Both). (E) *Allobaculum* sp. 128 and (F) *A. muciniphila* ORFs that were differentially expressed in the ileum, along with their fold changes, P-values, and annotations (Prokka). Data are from N=1 experiment.

Figure S4. Type strain *A. muciniphila* attenuates *Allobaculum* sp. 128-mediated colitis and *Allobaculum* sp. 128 blunts *A. muciniphila*-induced dendritic cell responses in MLN, Related to Figure 5. (A) Experimental schematic for acute DSS colitis in WT gnotobiotic mice colonized with MC, MC+*Allobaculum* sp. 128, MC+*A. muciniphila*^T (type strain ATCC BAA-835), or MC+*Allobaculum* sp. 128+*A. muciniphila*^T (ATCC BAA-835) (n=4-6 mice/group). (B) Fecal microbiota profiling, (C) colon length, (D) d2 fecal lipocalin (LCN2), and (E) gross colon pathology. (F) Representative gating strategy for analysis of MLN cells performed in FlowJo after ≥100,000 events per sample were collected. (G) Immunophenotyping of MLN cell populations (% of viable cells) (n=4-6 mice/group). (H) Quantification of DCs (Live B220⁻TCRb⁻CD11b⁺CD11c⁺MHCII⁺). Welch's t-test was used to compare microbiota groups. **P*<0.05. Data shown in (A-E) are from N=1 experiment. Data shown in (F-H) are from one of N=2 independent experiments.

Figure S5. Approach for profiling microbiota-dependent mucosal immune landscape using single cell RNAseq, Related to Figure 7. (A) Schematic depicting single cell RNAseq (scRNAseq) experiment. (B) Quality control metrics used for filtration of scRNAseq data before proceeding to clustering and differential expression analyses. Only cells with 500-5000 RNA features were retained (between blue dashed lines), as well as cells with <8% genes of mitochondrial (mt) origin. Welch's t-test was used to compare each cell lineage across microbiota groups; * *P*<0.05, ** *P*<0.01, *** *P*<0.001. N=1 experiment.

Figure S6. Expression of marker genes mapped to MLN cell clusters, Related to Figure 7. (A) Violin plots showing expression of marker genes across MLN cell clusters, numbered to match clusters shown in Figure S7.

Figure S7. Epistatic reversal of *A. muciniphila*-induced MLN immune cell clusters by co-colonization with *Allobaculum* sp. 128, and direct assessment of MLN DC function in co-colonized gnotobiotic mice, Related to Figure 7 and Figure S6. (A-E) Graph-based probabilistic analysis of MLN scRNAseq data, comparing two microbiota groups at a time. (A-B) MC+Allo relative to MC+Both or relative to MC. (C-E) MC+Akk relative to MC+Both reveals strong reversal (high MELD score) of cell clusters induced by *A. muciniphila* after co-colonization with *Allobaculum* sp. 128. The marker genes that define each cluster are displayed in Figure S6. (F-G) *Ex vivo* co-cultures of purified MLN DCs from gnotobiotic mice were examined for their capacity to prime CellTrace Violet (CTV)-labeled naïve OT-II T cells, by proliferation and dilution of CTV. Data shown are from N=1 experiment.

SUPPLEMENTAL TABLES

Table S1. *Allobaculum* and *A. muciniphila* abundance in two human cohorts broadly reflects an inverse relationship, Related to Figure 3. (Sheet 1. American Gut Project) Enumeration of sequencing data from American Gut Project (McDonald, et al. 2018; QIITA study IDs: 48742, 51570, 52698, 53379, & 54454). (Sheet 2. Schirmer_UC) Relative abundance data from pediatric ulcerative colitis (Schirmer, et al. 2018).

Table S2. Human microbiota-associated gnotobiotic mouse screen reveals inverse correlation between *Allobaculum sp. 128* and *A. muciniphila*, Related to Figure 3. (Sheet 1. Relative Abundance) fecal microbiota profiles across n=19 gnotobiotic mice. (Sheet 2. Correlation Metrics, Sorted) Compiled statistics evaluating correlation and logistic regression between each pair of microbiota OTUs.

Figure S1.

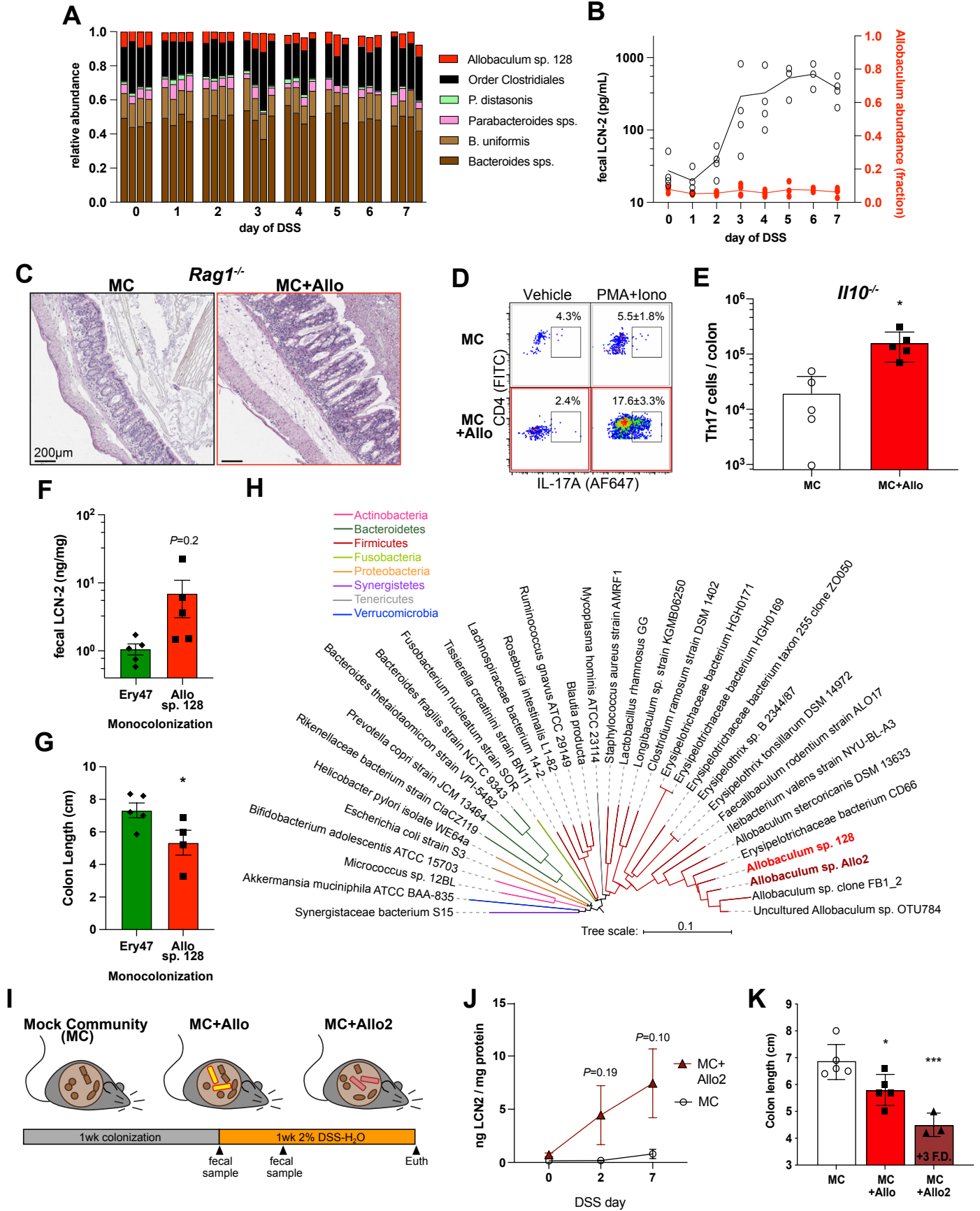


Figure S2.

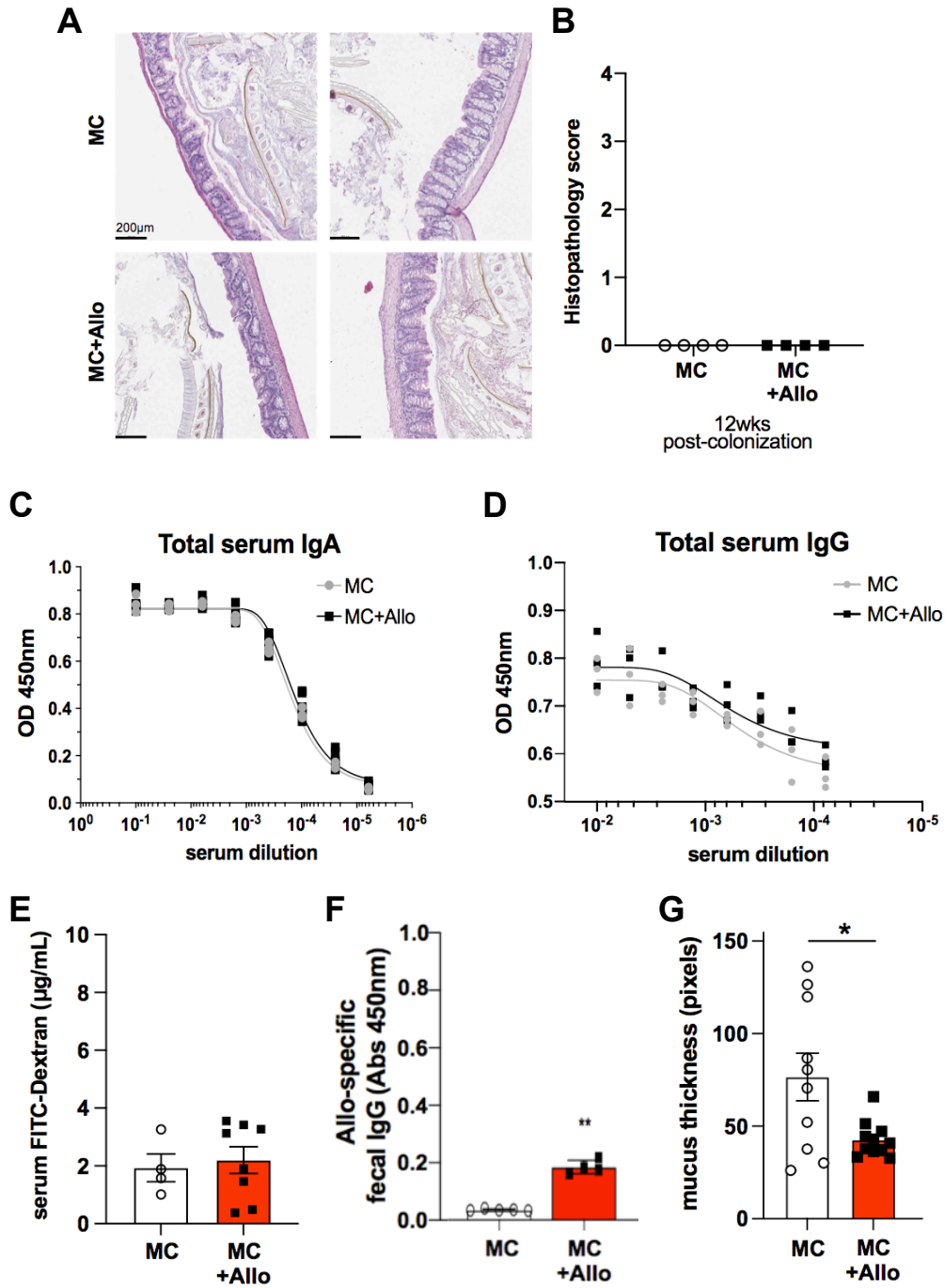


Figure S3.

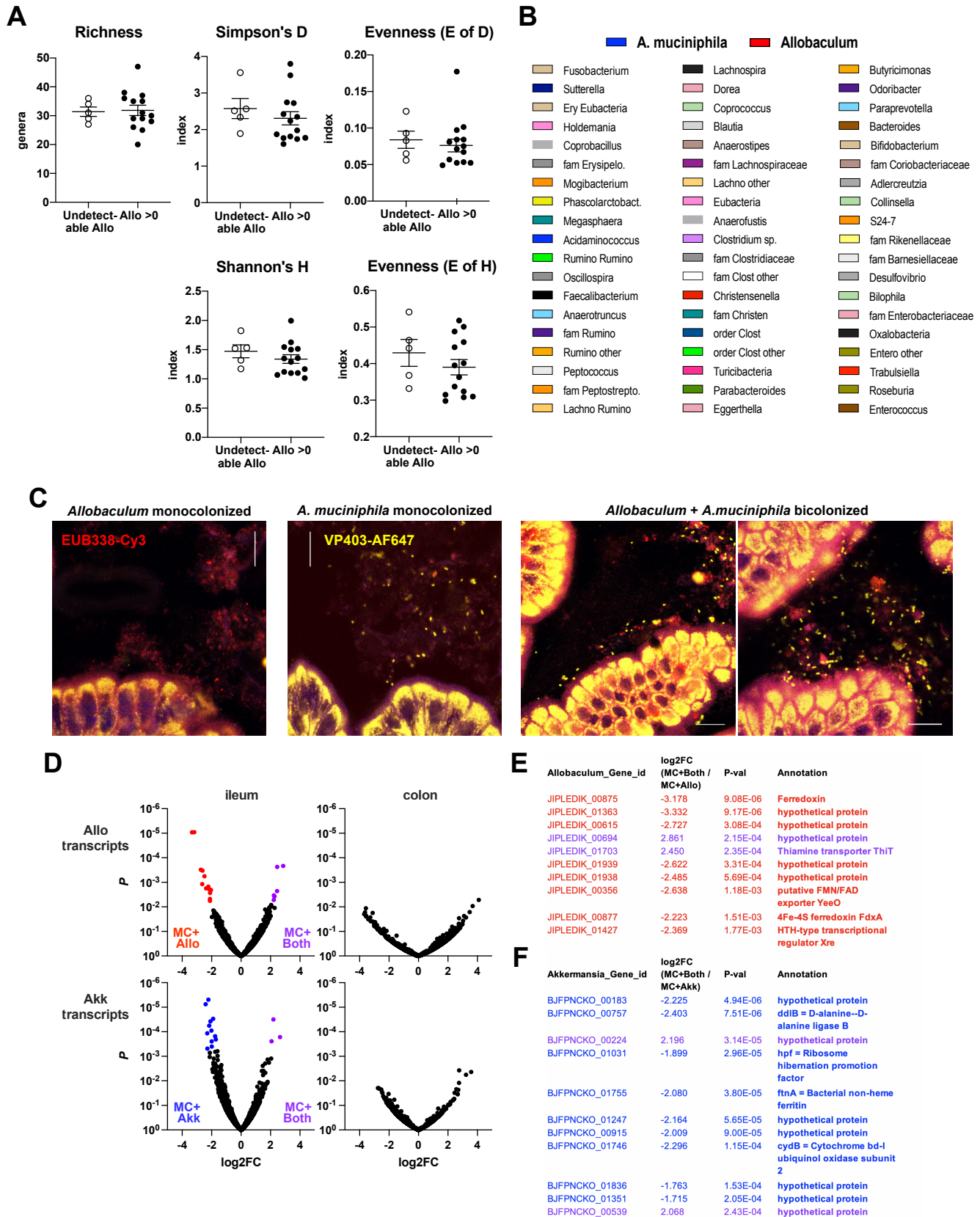


Figure S4.

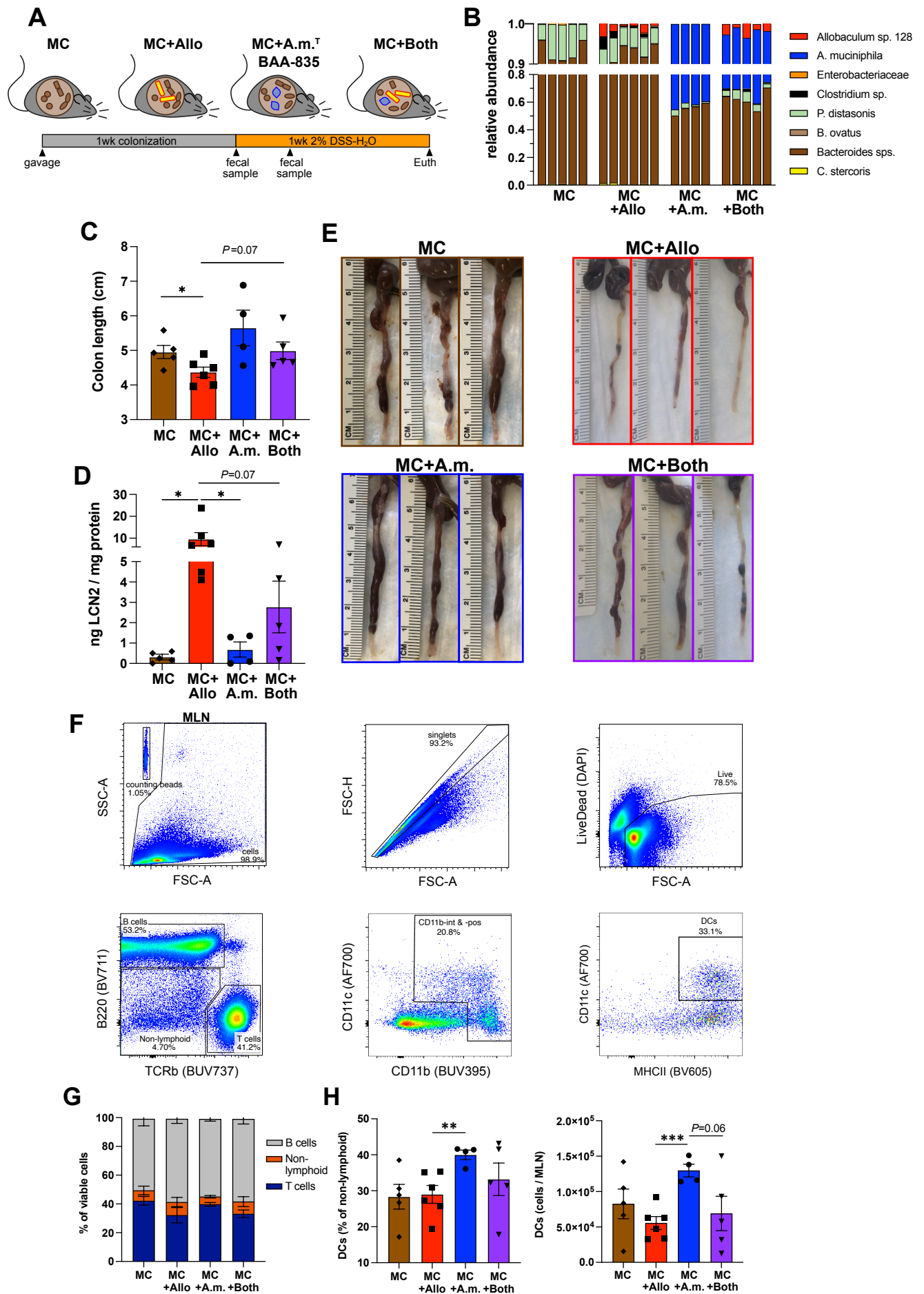


Figure S5.

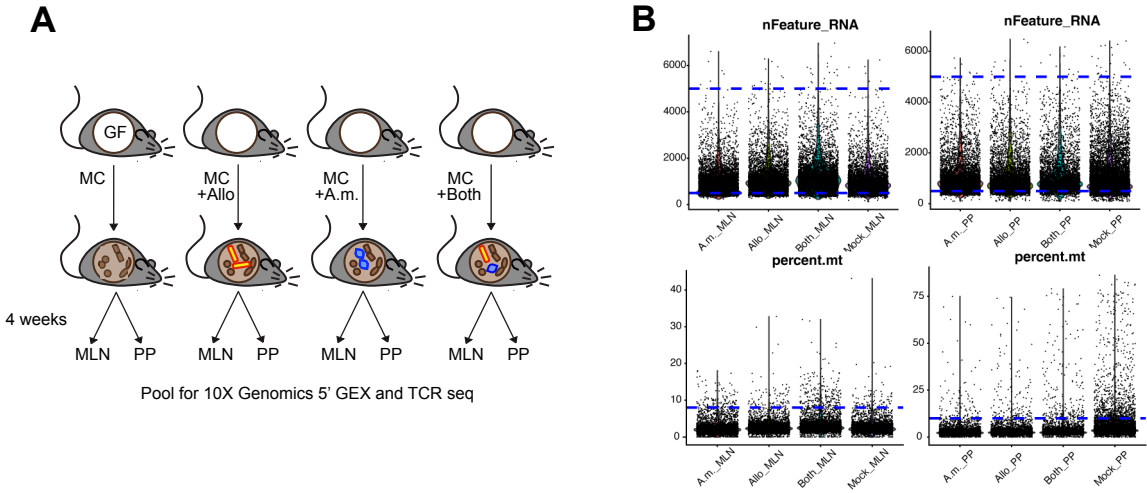


Figure S6.

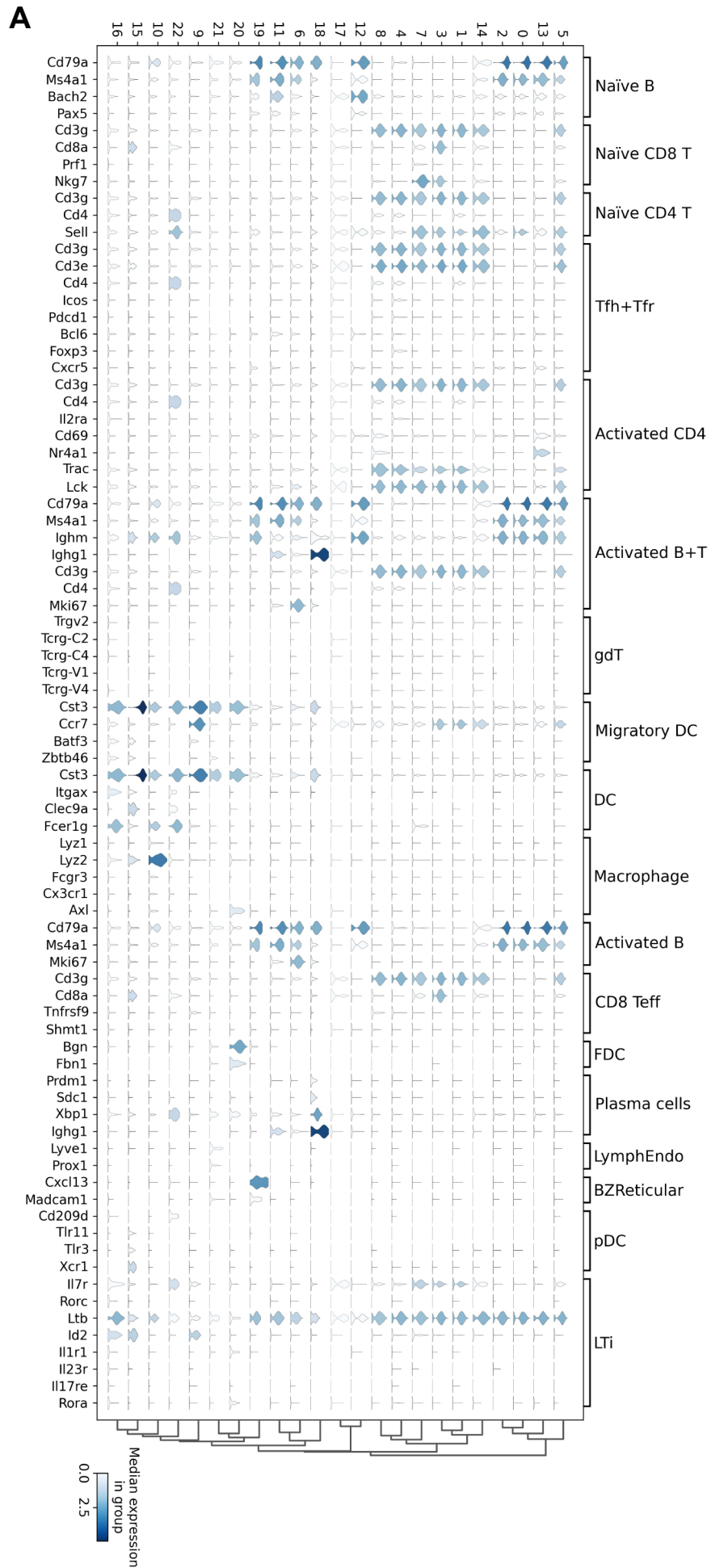


Figure S7.

