Supporting Information

Backbone distortions in lactam-bridged helical peptides

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Materials

Chemical reagents and solvents for the peptide syntheses were of peptide-synthesis grade; solvents for HPLC and spectroscopy were of HPLC or spectroscopy grade. Fmoc-protected amino acids, Rink-amide MBHA resin (100-200 mesh, loading 0.57 mmol/g), N,Ndiisopropylethylamine (DIPEA), piperidine, N,N-dimethylformamide (DMF), N-methyl-2pyrrolidone (NMP), dichloromethane (DCM), diethylether and trifluoroacetic acid (TFA) were purchased from Iris Biotech (Germany). Thioanisole (TIA), acetic anhydride, PhSiH₃, Pd(PPh₃)₄, acetonitrile, α-cyano-4-hydroxycinnamic acid, triisopropylsilane (TIS) and 1,2ethanedithiol (EDT) were purchased from Sigma Aldrich (Germany). 2-(1H-benzotriazole-1yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HBTU) and N-hydroxybenzotriazole (HOBt) were purchased from Biosolve (The Netherland). D₂O was from Armar GmbH (Germany). 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), fetal bovine serum, penicillin-streptomycin and L-glutamine were purchased from Sigma-Aldrich (Austria). Dulbecco's modified Eagle's medium (DMEM)-high glucose and Roswell Park Memorial Institute (RPMI) 1640 medium were purchased from Szabo-Scandic (Austria). Dimethyl sulfoxide (DMSO) was from VWR (Austria). The primary human lung fibroblasts were a gift from Prof. Dr. Jutta Horejs-Höck, University of Salzburg (Austria). The cell line MCF-7 was a gift from Prof. Dr. Barbara Krammer, University of Salzburg (Austria). The cell lines HT1975, A427 and SKLU-1 were a gift from Prof. Dr. Emilio Casanova, Medical University of Vienna (Austria). The cell lines A549 (ATCC: CLL-185), H460 (ATCC: HTB-177), H520 (ATCC: HTB-182) and HCC827 (ATCC: CRL-2868) were purchased from ATCC.

Methods

Solid-phase peptide synthesis was carried out on an automatic peptide synthesizer (Syro I, Biotage). The analytical HPLC equipment was from Thermo Fisher Scientific (Ultimate 3000). The analytical column was from Thermo Fisher Scientific (Syncronis C₁₈, 4.6x250 mm). The gradient used for analytical HPLC was the following: 3% B for 8 min, up to 60% B over 35 min (A = H₂O with 0.06% TFA; B = CH₃CN with 0.05% TFA). MALDI-TOF mass spectra were recorded on an Autoflex mass spectrometer from Bruker Daltonics using α -cyano-4-hydroxycinnamic acid as matrix. The CD measurements were recorded on a Chirascan Plus CD spectrometer from Applied Photophysics. UV measurements were carried out on a Varian Cary UV-visible spectrophotometer. For the determination of the cell viability with the MTT assay, a GloMax® Multimode Microplate Reader was used.

Cell-viability assay

The primary human lung fibroblast and the MCF-7 cell line were cultured in DMEM-high glucose supplemented with 10% fetal bovine serum, 1% penicillin–streptomycin and 1% L-glutamine. All other cell lines were maintained in RPMI 1640 medium supplemented with 10% fetal bovine serum, 1% penicillin–streptomycin and 1% L-glutamine. Cells were grown in a humidified atmosphere at 37 °C in 5% CO₂. For all experiments, 70–80% confluent cells were used. Cells (10000 cells/well) in complete growth medium were seeded into 96-well culture plates. The day after, the medium was changed to a serum free medium with peptides at a concentration of 100 μ M. After an additional incubation period of 24 h, the cell viability was determined by adding 10 μ l MTT solution (5 mg/ml in PBS) to the treated and untreated control cells for 2 h at 37 °C in the dark. Then, the medium was aspirated, the cells were lysed with 100 μ l DMSO and the absorbance of the resulting product formazan in viable cells was measured at 550 nm. Three independent experiments (with each sample in triplicate) were performed and cell viability was normalized to the untreated control.

Figures



Figure S1. Analytical HPLC of the synthetic lactam-bridged peptides used in this work (see Table S1 for t_R values and gradient).



Figure S2. MALDI-TOF-MS of the synthetic lactam-bridged peptides used in this work (see Table S1 for M_{found}).



Figure S3. CD spectra of the lactam-bridged peptides in (a, b) phosphate buffer (50 mM, pH 7.3) and (c, d) water (pH range 3-4). The dashed boxes show the maximal CD contribution below 200 nm, which is higher than 6000 deg cm² dmol⁻¹ for **1Y**, **2-5**, **7** (a, c), and smaller than 4000 deg cm² dmol⁻¹ for **6**, **8-10** (b, d). Difference spectra (peptide – **1Y**) in (e) phosphate buffer and (f) water.



Figure S4. Comparison of the CD spectra of the lactam-bridged peptides in phosphate buffer (50 mM, pH 7.3) and water (pH range 3-4).



Figure S5. Comparison of the CD spectra of the lactam-bridged peptides **2** and **5** in phosphate buffer (50 mM, pH 7.3), acetate buffer (10 mM, pH 4.5) and water (pH range 3-4).



Figure S6. Chemical shifts differences (ppm) between the measured and random coil¹ H^N for each residue of the peptides in water (pH range 3-4).



Figure S7. NOE pattern of the lactam-bridged peptides in water (pH range 3-4).



Figure S8. 1D-NMR spectra of the lactam-bridged peptides in water (pH range 3-4).



Figure S8. continued.



Figure S9. Effect of the lactam-bridged peptides **1Y** and **2-10** (100 μ M) on cancer-cell viability. MCF-7: human breast adenocarcinoma; A549, SKLU-1, H1975 und A427: human lung non-small cell adenocarcinoma; HCC827: human lung non-small cell adenocarcinoma with an EGFR mutation; H520: human lung squamous cell carcinoma; H460: human lung large cell adenocarcinoma. Human primary lung fibroblasts were used as control. p-Values (referred to **1Y**): ***: <0.001; **: <0.01; *: <0.05. In previous reports we showed that **1Y** reduces cancer-cell viability in the low micromolar concentration range²⁻⁴. Here, we show that none of the new analogs was more effective than **1Y**, displaying similar or reduced efficiency depending on the cell line. About the three analogs with the highest sequence similarity (**2-4**), the Tyr-4-containing analog **3** was less effective than **1Y** in all tested cell lines, whereas the Nle-4-containing analog **2** and the Ile-7-containing analog **3** were comparable to **1Y** only in four and three cell lines, respectively.

Tables

Number	Peptide sequence	M _{theor.} a (Da)	M _{found} ^b (Da)	t _R e (min)	μH ^f
1Y	cyclo-(2,6)-(Ac-VKRLQDLQY-NH2)	1185.40	1186.936°	25.7	0.466
2	cyclo-(2,6)-(Ac-VKRXQDLQY-NH2)	1185.40	1186.612 ^c	25.8	0.466
3	cyclo-(2,6)-(Ac-VKRYQDLQY-NH2)	1235.41	1234.537 ^d	23.7	0.385
4	cyclo-(2,6)-(Ac-VKRLQDIQY-NH ₂)	1185.40	1184.557 ^d	25.4	0.470
5	cyclo-(2,6)-(Ac-YKRLQDVQY-NH2)	1235.41	1236.679°	24.3	0.430
6	cyclo-(2,6)-(Ac-XKRIQDVQY-NH ₂)	1185.40	1186.888°	25.8	0.502
7	cyclo-(2,6)-(Ac-XKRXQDXQY-NH ₂)	1199.42	1198.571 ^d	27.9	0.502
8	cyclo-(2,6)-(Ac-XKRYQDLQV-NH2)	1185.40	1186.756 ^c	25.2	0.409
9	cyclo-(2,6)-(Ac-LKRXQDYQL-NH ₂)	1199.42	1200.490°	27.3	0.458
10	cyclo-(2,6)-(Ac-IKRVQDYQL-NH2)	1185.40	1186.852 ^c	25.2	0.425

Table S1. Analytical characterization of the synthetic lactam-bridged peptides used in this work (X = Nle).

a. Averaged mass

b. Measured by MALDI-TOF-MS

c. Positive mode (M+H)*

d. Negative mode $(M-H)^{-}$

e. HPLC gradient: 3% B for 8 min, 3-60% in 35 min, with A = 0.06% TFA in water and B = 0.05% TFA in acetonitrile

f. The hydrophobic moment μ H was calculated for the <u>linear</u> sequence (Leu was used for Nle) by using the web server HeliQuest (https://heliquest.ipmc.cnrs.fr/)⁵

Peptide no.	1Y	2	3	4	5	6	7	8	9	10
NMR distance and	d dihedra	I restrain	ts							
NOE distance res	traints									
Total	119	168	176	163	166	161	113	186	141	147
Intraresidue	61	82	97	87	93	99	65	101	74	85
Interresidue										
Sequential	36	46	55	11	10	46	32	60	40	30
(<i>i-j</i> = 1)	50	40	- 55	44	43	40	52	00	40	- 55
Medium-range	22	40	24	32	24	16	16	25	27	23
(<i>i-j</i> ≤ 5)	22	-10	27	52	27	10	10	20	21	20
Long-range				_	_					_
(<i>i-j</i> ≥ 5)										
Dihedral restraint	s					1				
ф	5	5	4	7	7	3	7	3 ^a	7	3ª
Ψ	5	5	4	7	7	3	7	3 ^a	7	3 ^a
Structural statisti	cs									
Violations (mean	and SD)	-					-	-	-	
No. of violated										
distance	0.0	0.0	0.0	0.0	0.3	0.0	0.0	0.0	0.0	0.6
constraints	± 0.0	± 0.0	± 0.0	± 0.0	± 0.4	± 0.0	± 0.0	± 0.0	± 0.0	± 0.5
> 0.2 Å										
No. of violated	11	10	10	10	09	0.0	10	10	10	0.0
dihedral angle	+0.2	+ 0 0	+ 0 0	+ 0 0	+03	+ 0 0	+0.0	+0.0	+ 0 0	+ 0 0
constraints > 5°	<u> </u>	<u> </u>	± 0.0	± 0.0	± 0.0	± 0.0	± 0.0	± 0.0	<u> </u>	± 0.0
Max. distance	0.02	0 16	0.06	0.08	0 19	0 15	0.06	0.12	0.04	0.20
constraint	+0.02	+0.00	+0.00	+0.02	+0.05	+0.05	+0.00	+0.02	+0.01	+0.02
violation (A)	± 0.02	± 0.00	10.01	± 0.02	± 0.00	10.00	± 0.00	± 0.02	10.01	± 0.02
Max. dihedral	12.9	671	9 48	5 87	7 99	0.05	5 48	6 95	7 37	1 35
angle violation	+0.1	+0.40	+0.17	+0.45	+1.58	+0.08	+0.47	+0.56	+0.70	+0.16
(°)	- 0.1		_ 0.11	_ 0.10	- 1.00	- 0.00	± 0.11	_ 0.00	_ 0.10	_ 0.10
Average pairwise	rms dev	iation (A)								
backbone	0.50	0.29	0.74	0.26	0.65	0.77	0.29	0.32	0.24	0.67
	± 0.23	± 0.22	± 0.34	± 0.10	± 0.24	± 0.30	± 0.13	± 0.16	± 0.14	± 0.29
heavy	1.56	1.39	2.08	1.32	1.96	1.82	1.40	1.31	1.23	1.64
nouvy	± 0.28	± 0.37	± 0.55	± 0.25	± 0.49	± 0.47	± 0.25	± 0.34	± 0.25	± 0.32
a TALOS detects dynamics in the terminal residues										

Table S2. Statistics of the NMR-derived structures of the lactam-bridged peptides.

^b Pairwise rms deviation was calculated among 20 structures of the peptide considering residues 2-9

Table S3. Random coil chemical shifts of norleucine in the reference peptide *Ac*-GGXGG-*NH*₂ measured in 7 M urea in D₂O at pH 2.3 or 7.4 (the peptide was synthesized by using the protocol reported above for the linear precursors of the lactam-bridged peptides **1Y**, **2-10**. Purity based on analytical HPLC: 95%. MS_{theor}.: 400.44, MS_{found} for M+Na⁺: 423.602 Da).

Resonance	δ (ppm)						
	pH 2.3	pH 7.4					
С	177.8	177.7					
Cα	57.0	57.0					
Сβ	33.3	33.4					
Сү	30.1	30.1					
Сδ	24.4	24.5					
Cε	15.9	15.9					
Н	8.24	8.25					
Ηα	4.30	4.30					
Ηβ2	1.82	1.82					
НβЗ	1.72	1.72					
Hγ	1.31	1.31					
Ηδ	1.31	1.31					
Ηε	0.88	0.88					

	1	Y		2			3			4	
	δH ^N	${}^{3}J_{HN\alpha}$		δH ^N	${}^{3}J_{HN\alpha}$		δH ^N	³ J _{HNα}		δH ^N	³ <i>J</i> _{ΗΝα}
	(ppm)	(Hz)		(ppm)	(Hz)		(ppm)	(Hz)		(ppm)	(Hz)
V	8.15	6.4	V	8.14	6.3	V	8.15	6.7	V	8.135	(6.4)
Κ	8.35	3.8	Κ	8.33	(4.8)	Κ	8.33	(4.4)	К	8.35	4.1
R	8.20	n.e.	R	8.25	3.8	R	8.26	4.5	R	8.24	4.2
L	7.39	n.e.	Х	7.46	5.8	Y	7.47	7.4	L	7.44	n.e.
Q	8.20	n.e.	Q	8.22	5.1	Q	8.39	5.3	Q	8.14	(6.0)
D	8.27	6.5	D	8.33	(4.8)	D	8.33	(6.7)	D	8.15	(6.2)
L	7.67	6.0	L	7.78	5.6	L	7.86	6.4	Ι	7.68	6.6
Q	7.99	6.5	Q	8.03	(6.5)	Q	8.09	6.6	Q	8.15	(6.8)
Y	7.92	7.8	Y	7.95	7.8	Y	7.96	7.7	Y	8.09	7.9

Table S4. Chemical shifts of the backbone amide protons (H^N) and ${}^{3}J_{HN\alpha}$ coupling constants of the lactambridged peptides in water (pH range 3-4). Values extrapolated from partially overlapped signals are in brackets (n.e.: not extractable. X = norleucine).

	ļ	5	6 7		7		8				
	δαΝΗ	³ J _{ΗΝα}		δαΝΗ	³ J _{ΗΝα}		δαΝΗ	${}^{3}J_{HN\alpha}$		δαΝΗ	³ J _{ΗΝα}
	(ppm)	(Hz)		(ppm)	(Hz)		(ppm)	(Hz)		(ppm)	(Hz)
Y	8.15	5.9	Х	8.17	(5.7)	Х	8.21	(5.6)	X	8.21	5.6
K	8.19	4.9	К	8.33	4.7	К	8.30	(3.6)	K	8.30	(4.8)
R	8.09	(4.5)	R	8.32	4.6	R	8.17	3.8	R	8.22	4.5
L	7.49	7.0	Ι	7.29	8.0	Х	7.42	6.6	Y	7.45	7.5
Q	8.06	5.7	Q	8.18	(5.5)	Q	8.20	(5.6)	Q	8.36	5.3
D	8.14	7.3	D	8.26	6.9	D	8.30	(6.7)	D	8.30	(6.4)
۷	7.75	6.7	v	7.82	6.8	Х	7.68	5.8	L	7.84	6.2
Q	8.21	7.00	Q	8.22	6.9	Q	7.98	6.6	Q	8.16	6.8
Y	8.10	(7.7)	Y	8.11	8.0	Y	7.94	7.7	V	8.00	(8.0)

	ę	Ð		10		
	δαΝΗ	³ J _{ΗΝα}		δαΝΗ	³ J _{HNα}	
	(ppm)	(Hz)		(ppm)	(Hz)	
L	8.21	(5.5)	Ι	8.13	6.7	
K	8.27	4.3	Κ	8.38	4.8	
R	8.18	(4.4)	R	8.42	4.5	
Χ	7.48	6.4	۷	7.26	7.7	
Q	8.20	(5.8)	Q	8.24	5.1	
D	8.31	6.9	D	8.33	7.2	
Y	7.87	6.2	Y	7.93	6.5	
Q	8.12	7.0	Q	8.21	7.1	
L	7.97	7.1	L	8.04	6.8	





Table S5. Continued.



Table S6. Tilt angles θ and residues per turn calculated for the lactam-bridged cyclized motifs from the crystal structures of Ac-(cyclo-2,6)-F(p-NO₂)KLLLDF(p-NO₂)-NH₂ (CCDC deposition number 1941068⁶), Ac-(cyclo-6,10)-HKILHKLLQDS-NH₂ (PDB ID: 5WGD⁷), and Ac-(bicyclo-3,7+6,10)-HKS₅LHKS₅LQDS -NH₂ with S₅ = (S)-2-(4-pentenyl)Ala (PDB ID: 5WGQ⁷).

Ac-(cyclo-2,6)-F(p-NO₂)KLLLDF(p-NO₂)-NH₂ (CCDC deposition number 1941068⁶)

Rho (residues/turn):	4.2
Theta(PPN2@C,O):	7
Theta(LYS3@C,O):ª	21
Theta(LEU4@C,O):	20
Theta(LEU5@C,O):	6
Theta(LEU6@C,O):	20
Theta(ASP7@C,O): ^a	29

^a lactam bridge

Ac-(cyclo-6,10)-HKILHKLLQDS-NH₂ (PDB ID: 5WGD⁷)

Chain	F	E
Rho (residues/turn):	4.0	4.0
Theta(ILE3@C,O):	14	11
Theta(LEU4@C,O):	10	22
Theta(HIS5@C,O):	5	15
Theta(LYS6@C,O):ª	12	-6
Theta(LEU7@C,O):	26	38
Theta(LEU8@C,O):	15	
Theta(GLN9@C,O):	11	
Theta(ASP10@C,O):ª	48	

^a lactam bridge

Ac-(bicyclo-3,7+6,10)-HKS₅LHKS₅LQDS-NH₂ with $S_5 = (S)-2-(4-pentenyl)Ala$ (PDB ID: 5WGQ⁷)

Chain	E	F
Rho (residues/turn):	4.0	3.9
Theta(LYS3@C,O):	23	24
Theta(MK84@C,O):ª	5	21
Theta(LEU5@C,O):	4	10
Theta(HIS6@C,O):	25	18
Theta(LYS7@C,O): ^b	1	14
Theta(MK88@C,O):ª	24	33
Theta(LEU9@C,O):	26	20
Theta(GLN10@C,O):	16	26
Theta(ASP11@C,O): ^b	42	-1
Theta(SER12@C,O):	58	

^a hydrocarbon bridge. ^b lactam bridge

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