

Supplementary Information

Loss of immunity-related GTPase GM4951 leads to nonalcoholic fatty liver disease without obesity

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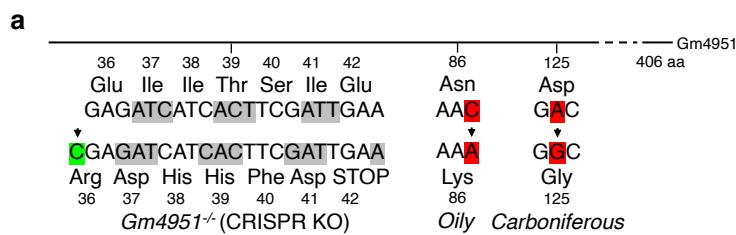
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Supplementary Table 1

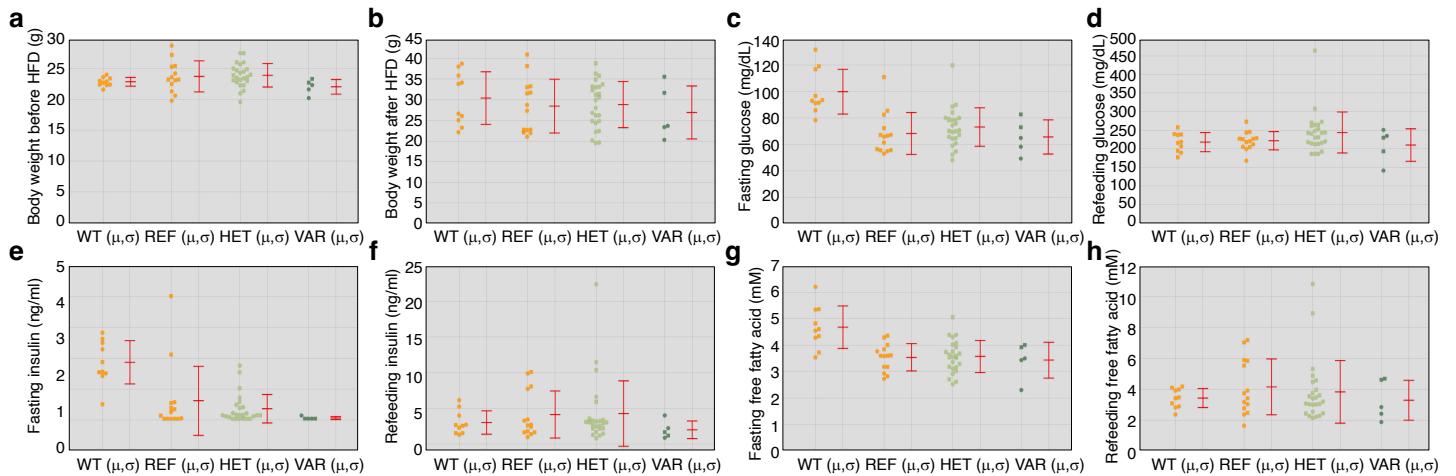


b

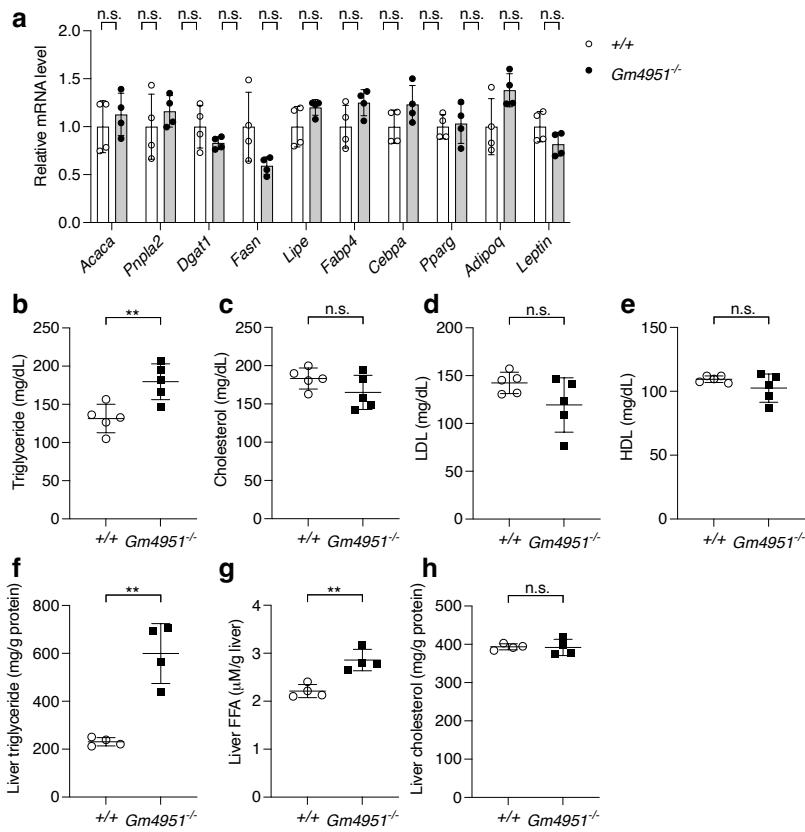
Birth ratio of *Gm4951^{+/+}* breedings

Genotype	Pup numbers	Percentage
<i>+/-</i>	47	23.2%
<i>Gm4951^{+/+}</i>	105	51.7%
<i>Gm4951^{-/-}</i>	51	25.1%

Supplementary Fig. 1 *Gm4951* mutations and birth ratios of knockout mice. **a** Illustration of nucleotide and amino acid changes in *Oily*, *Carboniferous*, and *Gm4951* knockout mice. Green indicates an insertion; red indicates a substitution. **b** Number of pups with different genotypes generated from *Gm4951^{+/+}* breedings.

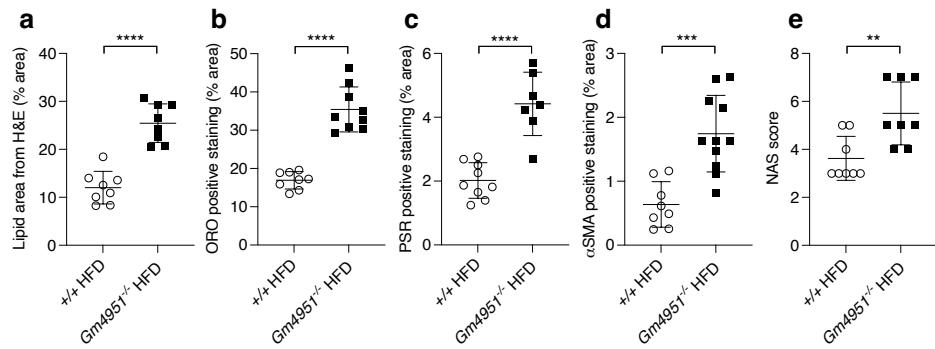


Supplementary Fig. 2 The Oily phenotype. **a-h** Body weight before HFD (**a**), body weight after 4 weeks on a HFD (**b**), overnight fasting glucose (**c**), glucose 2 h after refeeding (**d**), overnight fasting insulin (**e**), insulin 2 h after refeeding (**f**), overnight fasting free fatty acid (**g**), and free fatty acid 2 h after refeeding (**h**) in G3 mice were plotted vs. genotype at the *Oily* mutation site of *Gm4951* (n=10 mice in WT; n=14 mice in REF; n=25 mice in HET; n=5 mice in VAR). Data in **c-h** were measured after 2 weeks on a HFD. WT, C57BL/6J mice age-matched with G3 mice; REF, G3 mice homozygous for the reference allele of *Gm4951*; HET, G3 mice heterozygous for the reference allele and for the *Oily* allele; VAR, G3 mice homozygous for the *Oily* allele. Each data point represents one mouse. Mean (μ) and SD (σ) are indicated. Data are from one experiment.

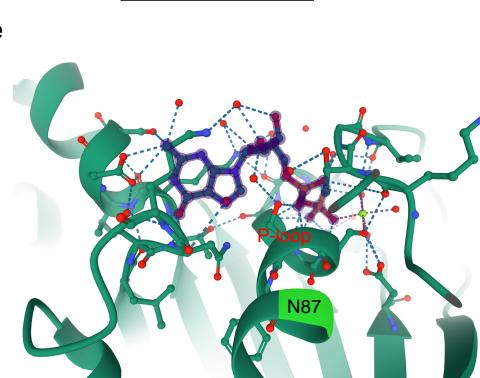
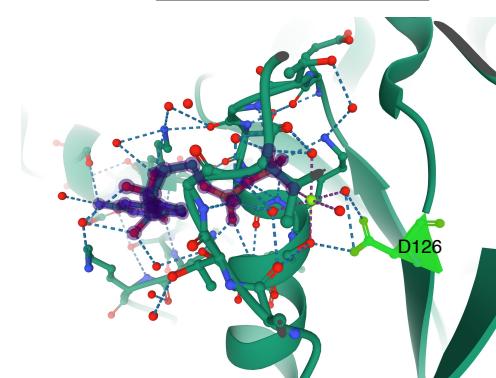
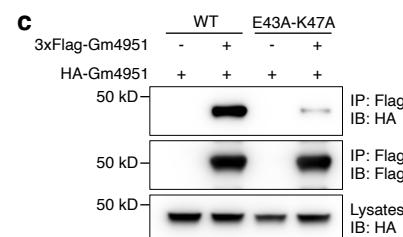
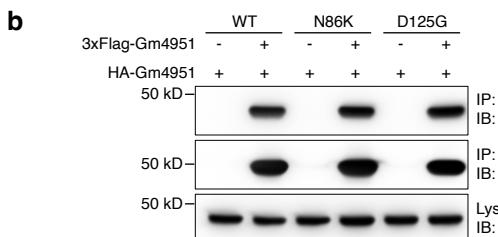
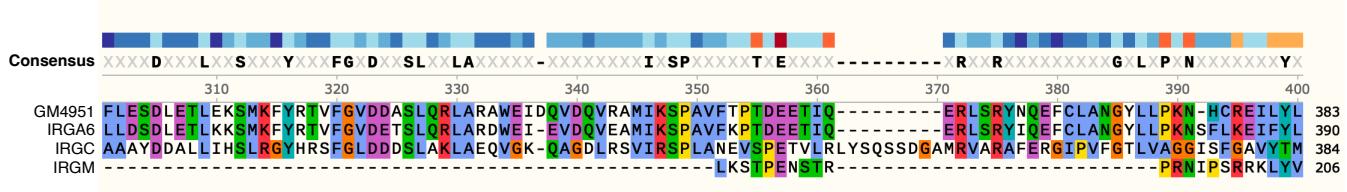


Supplementary Fig. 3 Lipid metabolism of adipose tissue, liver, and serum in WT and Gm4951^{-/-} mice.

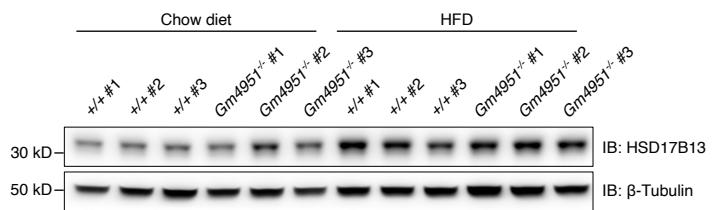
a Relative mRNA level of different genes in the iWAT from WT and Gm4951^{-/-} mice on HFD for 4 weeks (n=4 mice per genotype). Levels were normalized to Polr2a mRNA and then to levels in WT mice. **b-e** Fasting serum triglyceride (**b**), whole cholesterol (**c**), LDL cholesterol (**d**), and HDL cholesterol (**e**) of WT and Gm4951^{-/-} mice on HFD for 4 weeks (n=5 mice per genotype). **f-h** Liver triglyceride (**f**), FFA (**g**), and cholesterol (**h**) of WT and Gm4951^{-/-} mice on HFD for 4 weeks (n=4 mice per genotype). Each data point represents one mouse. Data are presented as means ± SD. P values were determined by two-tailed Student's t test. P values are denoted by ** P < 0.01; ns, not significant with P > 0.05. The exact P values of statistically significant groups are: 0.0069 (**b**), 0.0011 (**f**), 0.0026 (**g**). Data are representative of two independent experiments.



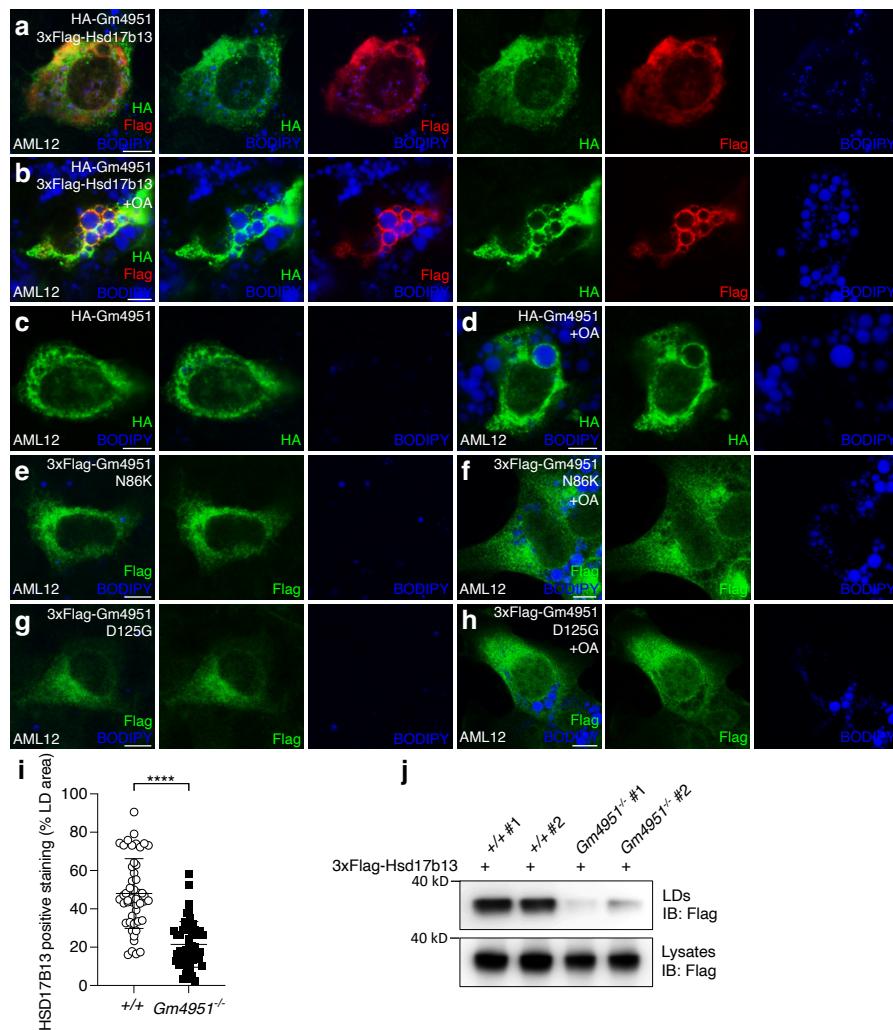
Supplementary Fig. 4 Quantification of histological stains in liver sections. **a**, Quantification of lipid area in liver sections stained with H&E (n=8 images per genotype). **b-d**, Quantification of stained area in liver sections stained with ORO (n=8 images in +/-; n=9 images in *Gm4951*^{-/-}) (**b**), PSR (n=9 images in +/-; n=7 images in *Gm4951*^{-/-}) (**c**), and α SMA (n=8 images in +/-; n=11 images in *Gm4951*^{-/-}) (**d**). Data in **a-d** are expressed as a percentage of total area per image. **e**, NAS score. Data points represent individual images. Data are presented as means \pm SD. *P* values were determined by two-tailed Student's *t* test. *P* values are denoted by ** *P* < 0.01; *** *P* < 0.001; **** *P* < 0.0001; ns, not significant with *P* > 0.05. The exact *P* values of statistically significant groups are: 4.29×10^{-6} (**a**), 5.094×10^{-7} (**b**), 2.505×10^{-5} (**c**), 0.0002 (**d**), 0.0051 (**e**). Data are from one experiment.



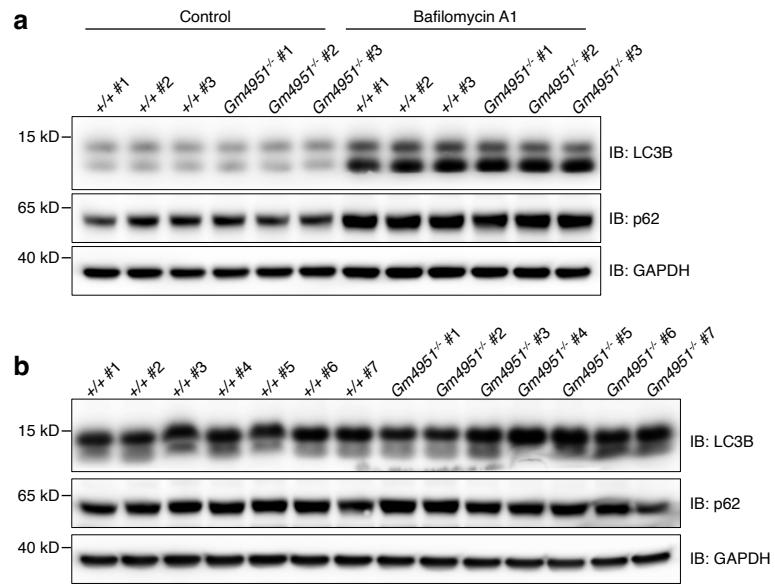
Supplementary Fig. 5 N86 and D125 are conserved between GM4951 and other IRG proteins. a Protein sequence alignment between mouse GM4951, mouse IRGA6, human IRGC, and human IRGM. According to IRGA6 structure⁹, residues involved in dimer formation are marked with “**”, and nucleotide interacting residues are marked with “+”. b-c Immunoblots of immunoprecipitates (top and middle) or lysates (bottom) of 293T cells. d-e Crystal structure of IRGA6 (PDB# 1TPZ) and GDP visualized with Mol* to show the location of D126 (d), N87 and P-loop (e), Mg²⁺ (green ball), and GDP (highlighted in dark color). Data are representative of two independent experiments (b-c).



Supplementary Fig. 6 No change in total protein level of HSD17B13 in *Gm4951*^{+/} liver.
Immunoblots of liver lysates from +/+ and *Gm4951*^{+/} mice on chow diet or HFD for 4 weeks. Data are representative of two independent experiments.



Supplementary Fig. 7 GM4951 specifically regulates the LD translocation of HSD17B13. **a-b** AML12 hepatocytes expressing HA-tagged Gm4951 and 3xFlag-tagged Hsd17b13 were immunostained with HA antibody (green), Flag antibody (red), and BODIPY (blue) to visualize LDs. 0.5 mM oleic acid for 24 h was used in +OA groups. **c-d** AML12 hepatocytes expressing HA-tagged Gm4951 without or with OA treatment were immunostained with HA antibody (green) and BODIPY (blue) to visualize LDs. **e-h** AML12 hepatocytes expressing 3xFlag-tagged Gm4951 mutants without (**e, g**) or with OA treatment (**f, h**) were immunostained with Flag antibody (green) and BODIPY (blue) to visualize LDs. **i** Quantification of the area with 3xFlag-HSD17B13 positive signal as a percentage of LD area in OA-treated primary hepatocytes of the indicated genotypes. LD area was defined by BODIPY positive signal. Each data point represents one LD (n=50 LDs in +/-; n=53 LDs in Gm4951^{-/-}). Data are presented as means \pm SD. P value was determined by two-tailed Student's t test. P values are denoted by **** P < 0.0001. The exact P value is 3.927×10^{-14} . **j** Immunoblots of isolated LDs from 24h OA treated +/- and Gm4951^{-/-} primary hepatocytes expressing 3xFlag-HSD17B13. Hepatocytes were isolated from mice on chow diet. Data are representative of two independent experiments.



Supplementary Fig. 8 Autophagic flux in *Gm4951*^{-/-} primary hepatocytes and liver. **a** Immunoblots of lysates of primary hepatocytes isolated from +/+ and *Gm4951*^{-/-} mice maintained on chow diet and treated *in vitro* with or without bafilomycin A1. **b** Immunoblots of liver lysates from +/+ and *Gm4951*^{-/-} mice. Data are representative of two independent experiments.

Supplementary Table 1 Primers used for quantitative PCR.

Target name	Direction	Forward primer (5'-3')	Reverse primer (5'-3')
<i>Gm4951</i>	Forward	GTAGACTCAAGCCAAGAGCA	AGCAGAAACTTAAACAGTGGG
<i>Polr2a</i>	Forward	CAAGATGCAAGAGGAGGAAGAG	TGTTGTCTGTCGAGGTAAGTG
<i>Slc27a1</i>	Forward	CAGGCAAGGGCATGGATGAT	ATGCGGTAGTACCTGCTGTG
<i>Cd36</i>	Forward	ATTAATGGCACAGACGCAGC	TTCAGATCCAACACAGCGT
<i>Acaca</i>	Forward	AGACTCACAAACGAGATTAC	CATTTATAAGACCACCGAC
<i>Fasn</i>	Forward	GATTCAAGGAGTGGATATTGTC	GAATGTTACACCTGCTCCT
<i>Elov16</i>	Forward	ACCGCAAGGCATTCAATTCC	AGTCGCTACGTGTTCTCTGC
<i>Scd1</i>	Forward	GTTCCGCCACTCGCCTAC	TGTAAGAACTGGAGATCTCTGG
<i>Apob</i>	Forward	TACTTCCACCCACAGTCCCCT	TGCTTTTAGGGAGCCTAGCA
<i>Mtp</i>	Forward	AGCCCCACTCAGGCAATT	GTTCACATCCGGCCACTAGG
<i>Ppara</i>	Forward	ATTGCTGTGGAGATCGGC	GCTTGGGAAGAGGAAGGTGT
<i>Acot1</i>	Forward	GGCTTGCACATGCGGTG	GAAAGGGTCCAGGTTCTGGG
<i>Acox1</i>	Forward	TCCAATCATGCGATAGCCTGG	TCCCCAACAGTGATGCCTGG
<i>Acadl</i>	Forward	TGCACACATACAGACGGTGC	CATGGAAGCAGAACCGGAGT
<i>3xFlag-Gm4951</i>	Forward	TCTGCTATAACCTGCTGCGG	CCCATCTTGTATCGTCATCCT
<i>Pnpla2</i>	Forward	TCTACTAAAGACCCCTGCCTG	CAGACATTGGCCTGGATGAG
<i>Dgat1</i>	Forward	GAGGACGAGGTGCGAGAC	CAGACGATGGCACCTCAGAT
<i>Lipe</i>	Forward	CTATTCAAGGGACAGAGGCAG	TAGTTCCAGGAAGGAGTTGAG
<i>Fabp4</i>	Forward	ACACCGAGATTCCCTCAAATG	CCATCTAGGGTTATGATGCTCTC
<i>Cebpa</i>	Forward	CTCTGATTCTGCCAAACTGAG	GACCCACTACTACATACACCC
<i>Pparg</i>	Forward	CTGAGGAGAAGTCACACTCTG	CGCTTCTTCAAATCTGTCTG
<i>Adipoq</i>	Forward	GCACTGGCAAGTTCTACTGCAA	GTAGGTGAAGAGAACGGCCTGT
<i>Leptin</i>	Forward	TCATCCAAGTAGAACCCCTGTC	TGCTTCCATCAAGTGTCT
<i>ACTB</i>	Forward	AAGATCAAGATCATTGCTCCTC	ATACTCCTGCTTGCTGATCC
<i>IRGC</i>	Forward	TTCTGGACAGCTGGAAGAACAA	AGACTTAGTTGTGGGTGAGGC
<i>PPARA</i>	Forward	GGATGTCACACAACGCGATTC	AGGCCTCGTAGATTCTCTGG
<i>ACOT1</i>	Forward	GCTCAGTCATCCTGAGGTAA	GGAAAGAGGCCATGGAAAG
<i>ACOX1</i>	Forward	CTCCGTGCAGCCAGATTAGT	ATGTGCCTCACTGCTCGAA
<i>ACADL</i>	Forward	CCCCTGTGTTCGGACATGG	GAATGAGAACATCGCGCGC