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**Pharmacological properties of TRPM3 isoforms are determined by the length of the pore
loop**

SUPPLEMENTARY INFORMATION

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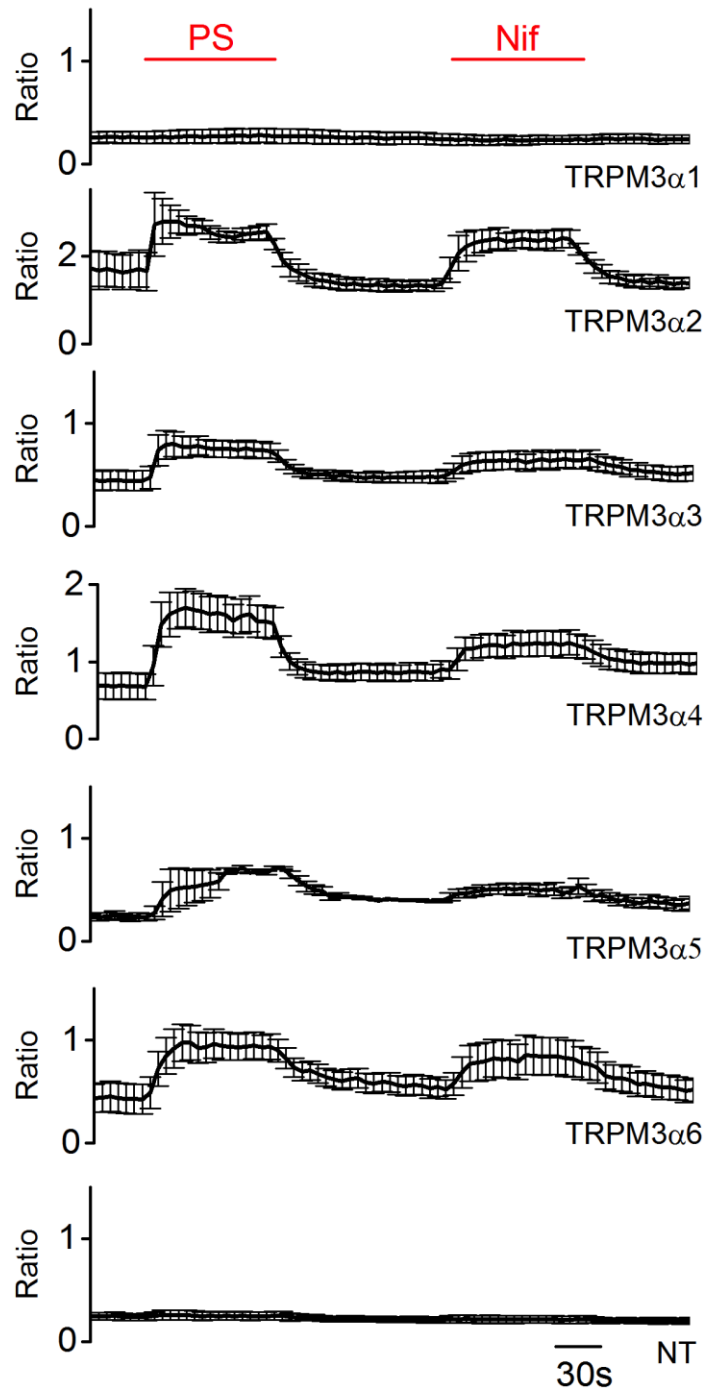
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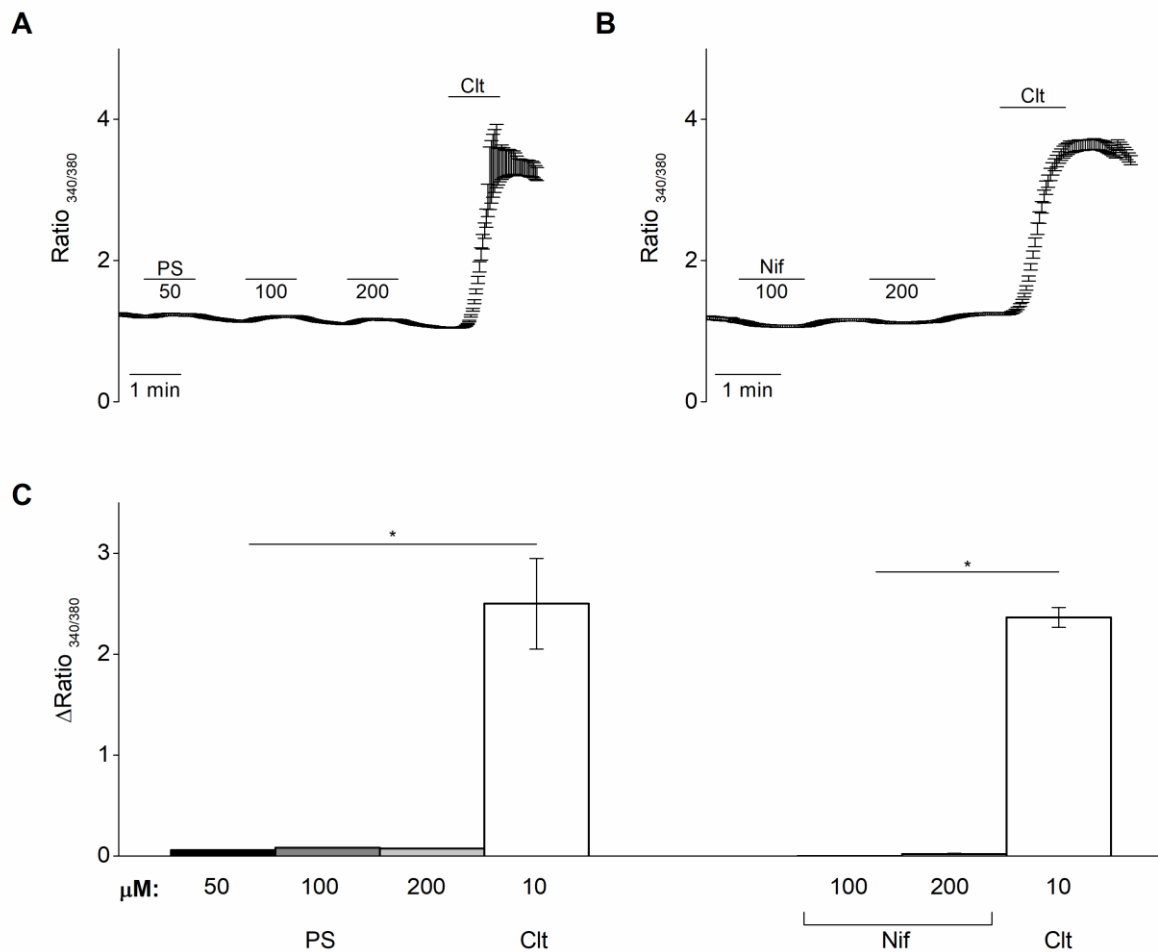
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21

22 **FIGURE S1**

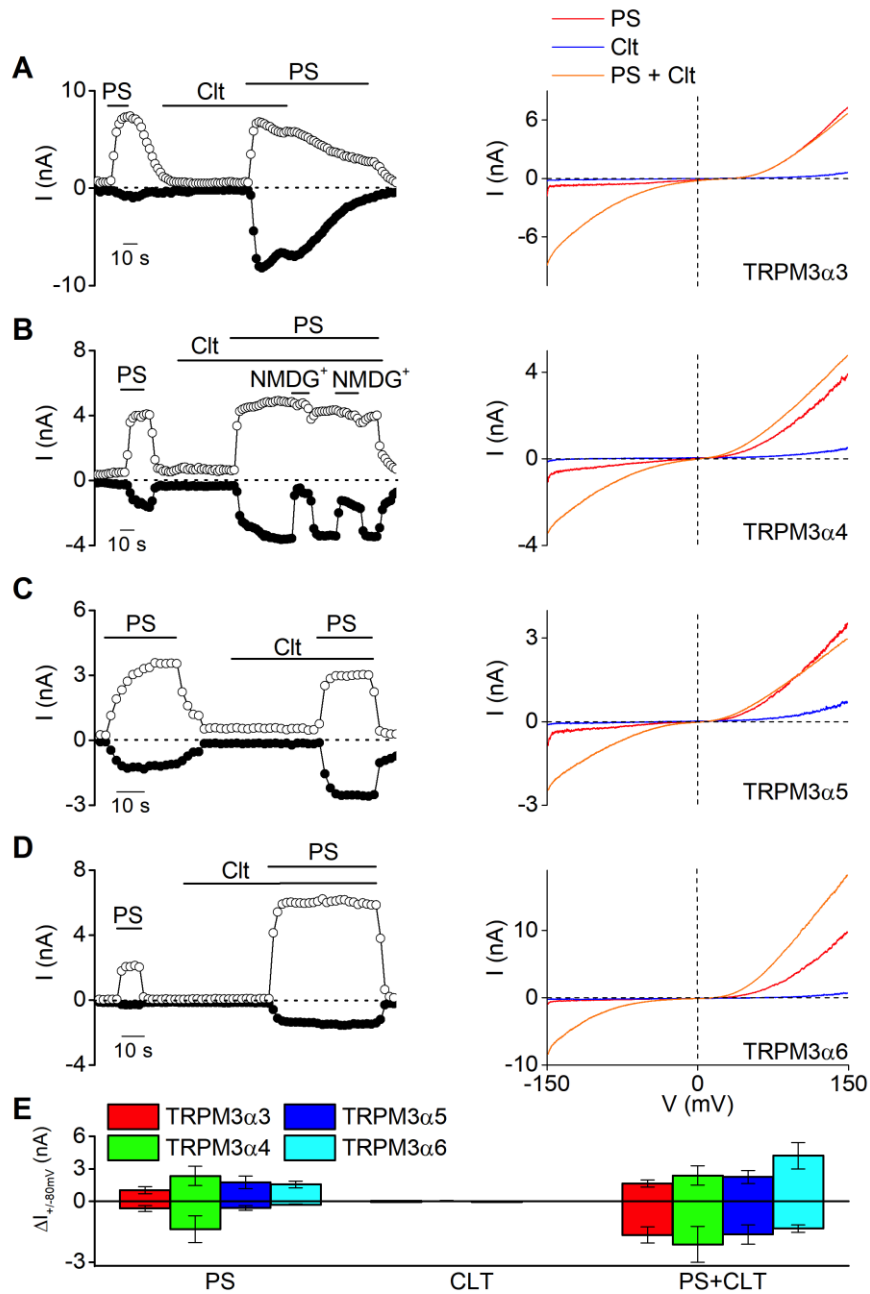
23 Mean fluorescence ratio (Fura₃₄₀/Fura₃₈₀) of HEK293T cells transfected with TRPM3α1 - α6 or
 24 of non-transfected cells during treatment with PS (40 μM) and Nif (50 μM) (N = 3 cover slips
 25 for each experiment).



26

27 **FIGURE S2**

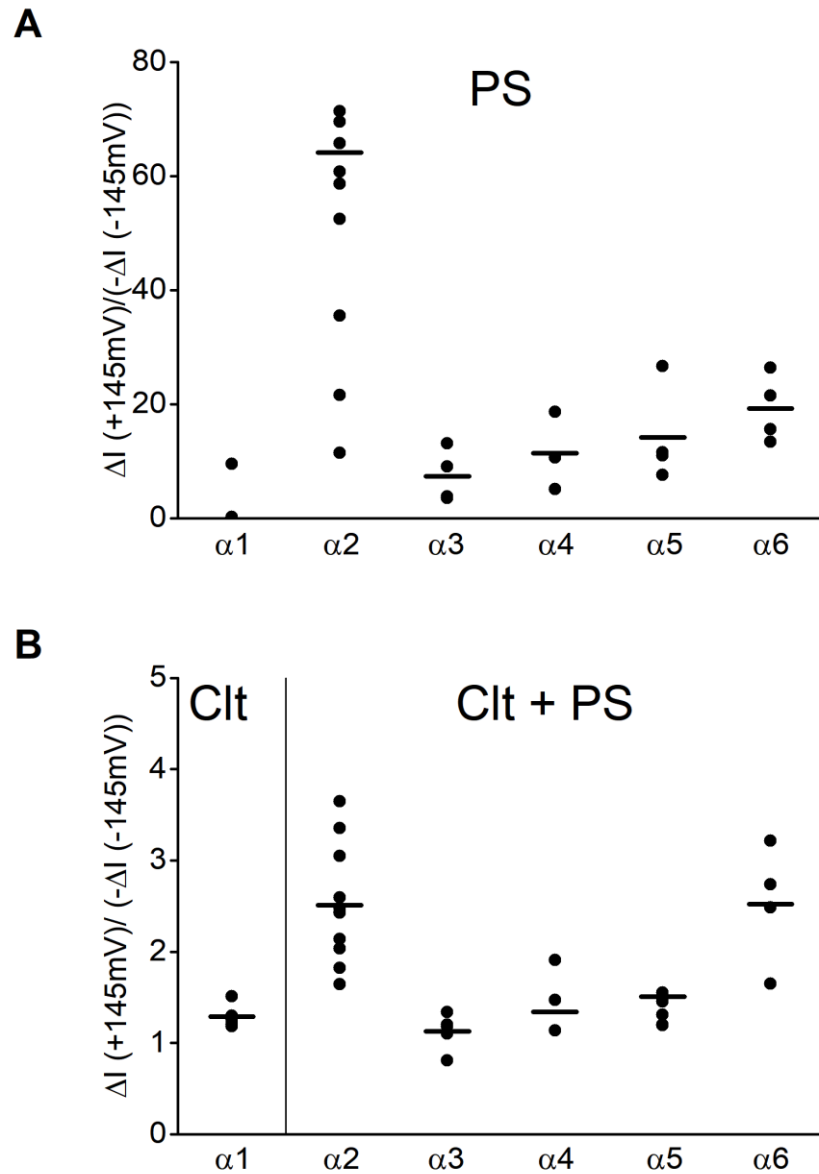
28 **A)** Time course of calcium fluorimetric measurements in HEK293T cells transiently expressing
 29 TRPM3 α 1 upon application of different concentrations of PS (50, 100 and 200 μ M) and Clt
 30 (10 μ M). **B)** As in *A*) during application of different concentrations of Nif (100 and 200 μ M)
 31 and Clt (10 μ M). **C)** Statistics of calcium increases that occur during the different treatments
 32 in *A*) and *B*) with $n = 366$ for PS treatment and $n = 393$ for Nif treatment with $N = 3$ cover slips
 33 for both experiments. * < 0.05 with Kruskal-Wallis ANOVA.



34

35 **FIGURE S3**

36 **A - D)** Time courses and corresponding *I-V* curves of whole-cell currents recorded in
 37 HEK293T cells transiently transfected with A) TRPM α 3, B) TRPM α 4, C) TRPM α 5, D) TRPM α 6
 38 during treatment with PS and Clt. **E)** Current increases of the different TRPM3 α variants at \pm
 39 80 mV upon chemical treatment as shown in B) – E) (n = 5 for TRPM3 α 3, n = 3 for TRPM3 α 4
 40 and n = 5 for TRPM3 α 5 and n = 4 for TRPM3 α 6).

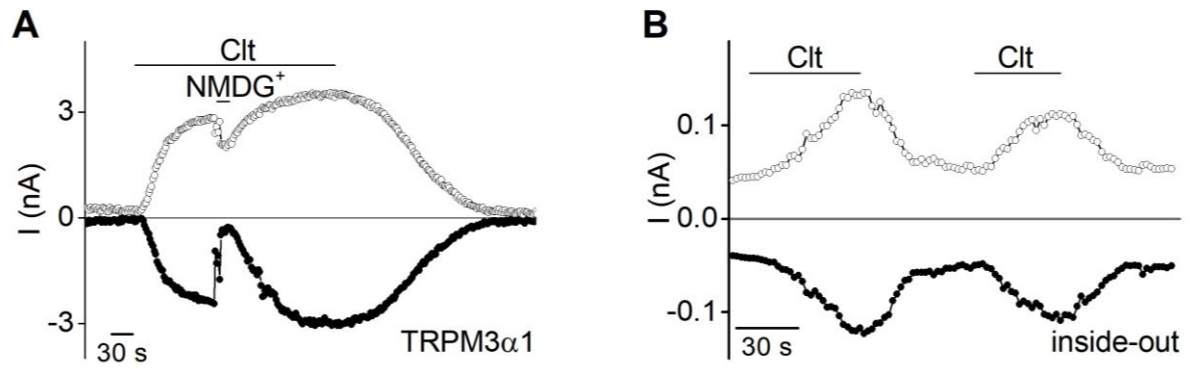


41

42 **FIGURE S4**

43 **A)** Current ratios for HEK293 cells expressing the different TRPM3 α isoforms calculated for
 44 values at ± 145 mV during application of PS (40 μ M). **B)** Current ratios at ± 145 mV for
 45 TRPM3 $\alpha 1$ and TRPM3 $\alpha 2$ during Clt (10 μ M) application and for TRPM3 $\alpha 2$ - $\alpha 6$ during PS (40
 46 μ M) + Clt (10 μ M) co-application. For both graphs: n = 5 for TRPM3 $\alpha 1$, n = 11 for TRPM3 $\alpha 2$,
 47 n = 5 for TRPM3 $\alpha 3$, n = 3 for TRPM3 $\alpha 4$ and n = 5 for TRPM3 $\alpha 5$ and n = 4 for TRPM3 $\alpha 6$)

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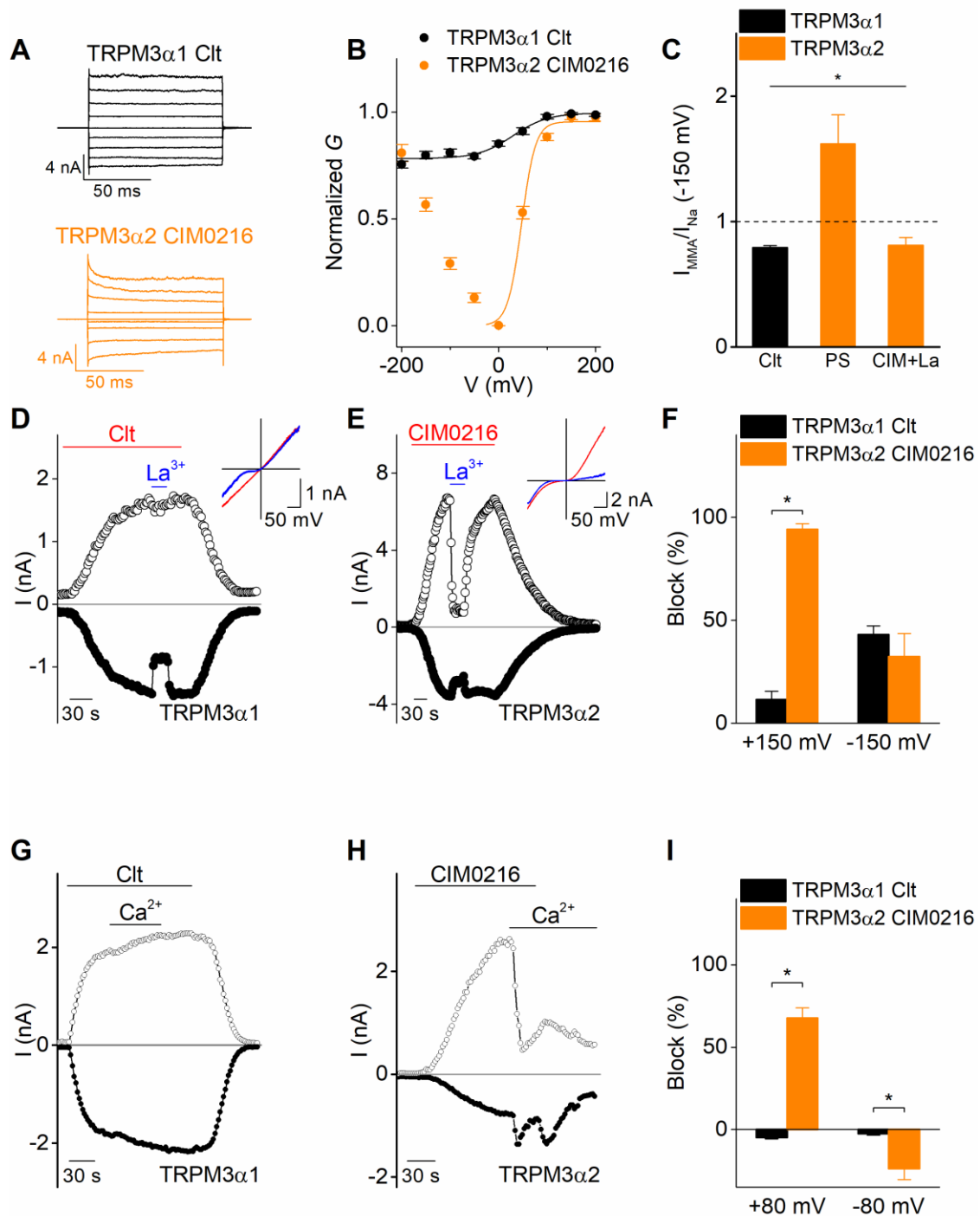


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50 **FIGURE S5**

51 **A)** Time course of whole-cell patch clamp recording in HEK293T cells transiently transfected
 52 with TRPM3 α 1 upon application of Clt (10 μ M). At the indicated time points all monovalent
 53 ions in the extracellular solution were replaced by NMDG $^+$. **B)** Time course of inside-out patch
 54 clamp recording in HEK293T expressing TRPM3 α 1 in presence of Clt (10 μ M). Mean current
 55 increase in TRPM3 α 1 after stimulation by Clt ($\Delta I_{+80\text{mV}}$: 67.3 ± 23.7 pA; $\Delta I_{-80\text{mV}}$: -58.9 ± 20.3
 56 pA) (n=8).

57

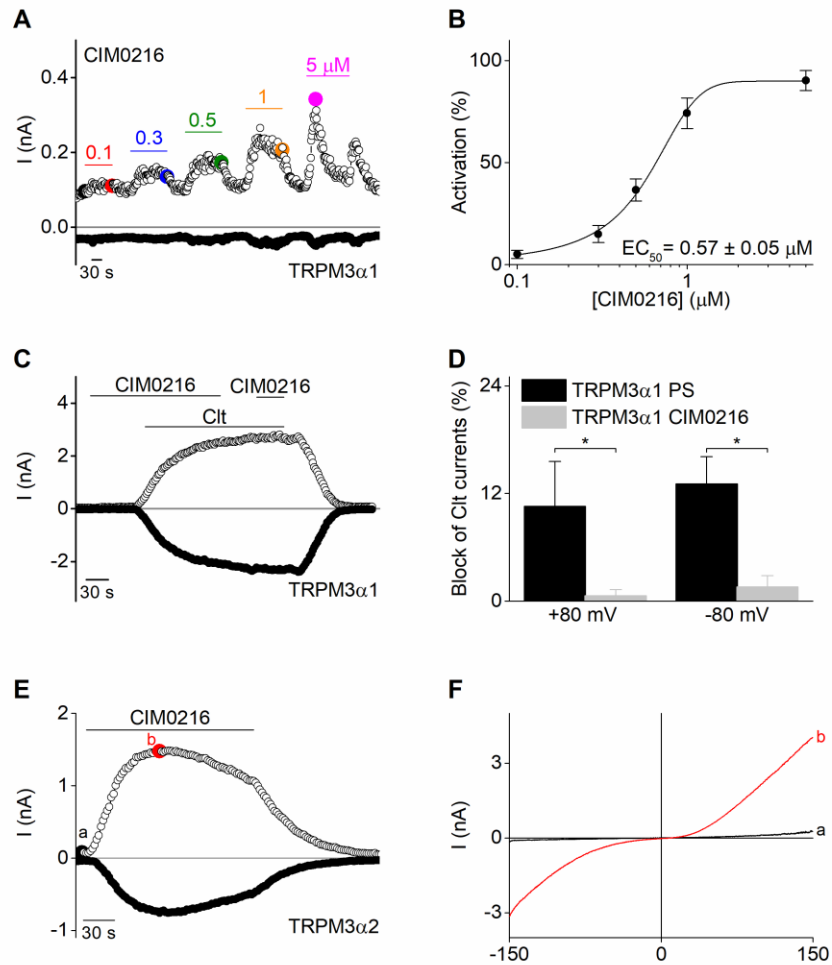


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59 **FIGURE S6**

60 **A)** Current traces obtained with a step protocol ranging from -200 mV to +200 mV in +50 mV
 61 steps for HEK293T cells transiently transfected with TRPM3 α 1 (black, upper panel) or stably
 62 transfected with TRPM3 α 2 (orange, lower panel). **B)** Normalized G-V relation for TRPM3 α 1
 63 CIt-induced and TRPM3 α 2 CIM0216-induced currents obtained from current traces as in A),

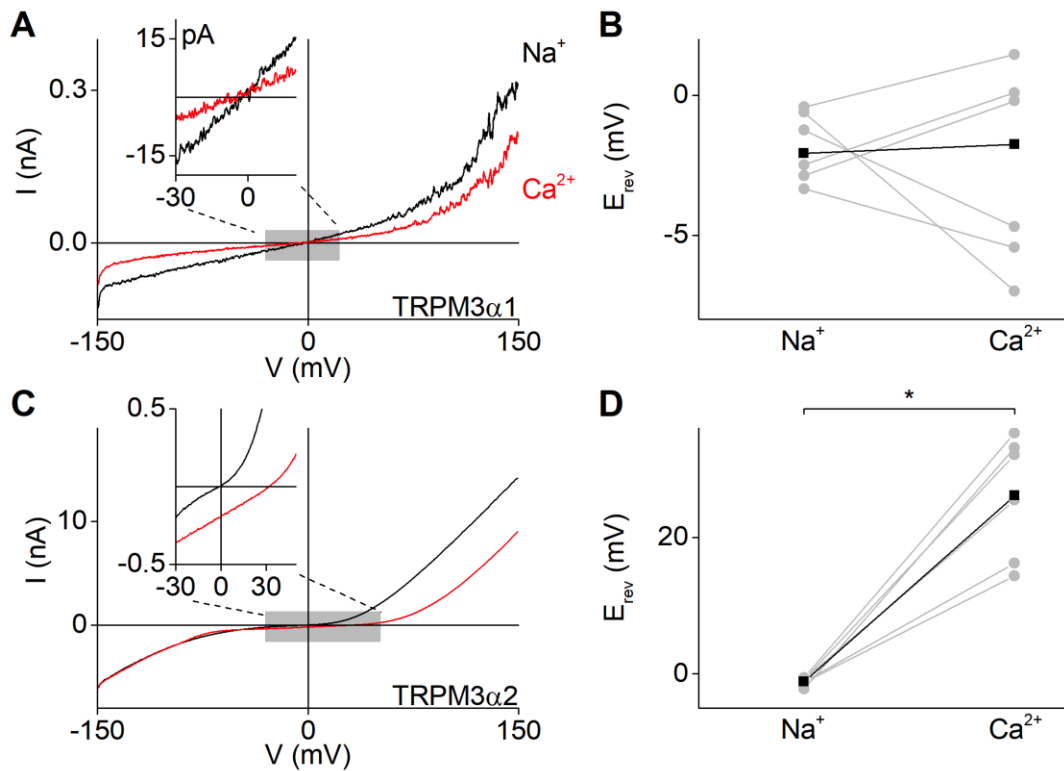
64 n = 6 for both conditions. **C)** Relative currents at -150 mV carried by monomethylammonium
65 (MMA⁺) for TRPM3 α 1 Clt-induced currents (black) and TRPM3 α 2 PS-induced currents and
66 CIM0216-induced currents in the presence of Lanthanum (La³⁺, orange). MMA⁺ currents were
67 normalized to the currents carried by Na⁺; PS (40 μ M), Clt (10 μ M), CIM0216 (1 μ M) and La³⁺
68 (100 μ M), n = 5 for all conditions. * < 0.05, One-way ANOVA. **D and E)** Time course of whole-
69 cell patch clamp recordings showing the effect of La³⁺ on TRPM3 α 1 Clt-induced currents *D)*
70 and TRPM3 α 2 CIM0216-induced currents *E)*. **F)** Percentage of La³⁺-induced block for
71 experiments as in *D)* and *E)* for n = 6 cells for each experiment. * < 0.05 with students two-
72 tailed unpaired t-test. **G and H)** Time course of whole-cell patch clamp recordings showing
73 the effect of calcium on TRPM3 α 1 Clt-induced currents *G)* and TRPM3 α 2 CIM0216-induced
74 currents *H)*. **I)** Percentage (%) of calcium-induced block for experiments as in *G)* and *H)* for n
75 = 5 cells. * < 0.05 with Mann-Whitney test.
76



77

78 **FIGURE S7**

79 **A)** Time course of whole-cell patch clamp recording at ± 80 mV in HEK293T cells transiently
80 expressing TRPM3 α 1 upon application of different concentrations of CIM0216 (in μ M). **B)**
81 Concentration-response curve for CIM0216 on TRPM3 α 1 extracted from experiments as in A)
82 for $n = 9$ cells. **C)** Time course of whole-cell patch clamp recording at ± 80 mV in HEK293T cells
83 transiently expressing TRPM3 α 1 upon co-application of CIM0216 and Clt. **D)** Statistics of the
84 inhibiting effects of PS and CIM0216 on Clt-induced TRPM3 α 1 currents for $n = 5$ cells per
85 treatment. **E)** Time course of whole-cell patch clamp recording at ± 80 mV in HEK293T cells
86 stably expressing TRPM3 α 2 upon application of CIM0216 (1 μ M). **F)** *I-V* traces for time points
87 indicated in E).

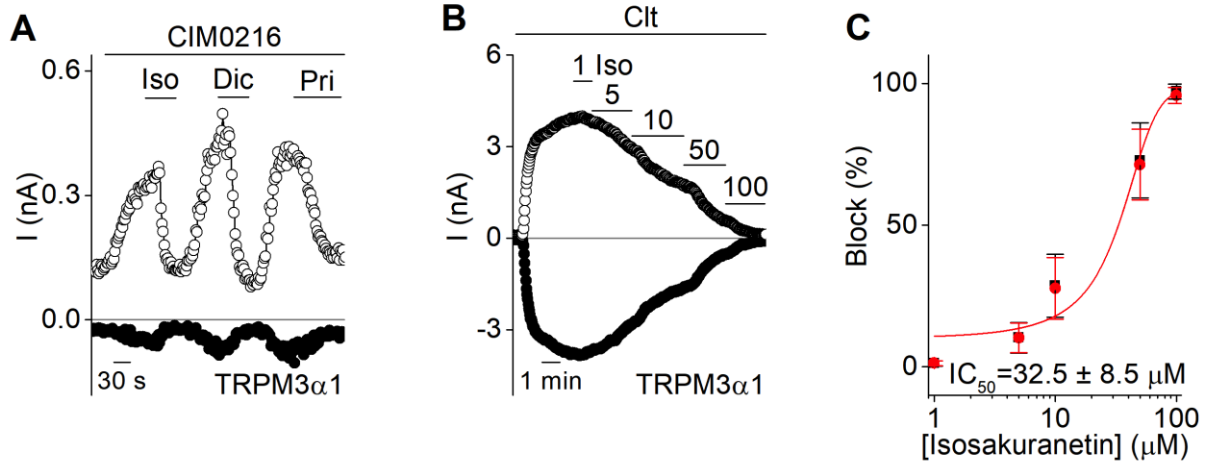


88

89 **FIGURE S8**

90 **A)** *I-V* Traces of whole-cell patch clamp recording in HEK293 cells transiently expressing
 91 TRPM3α1 upon application of CIM0216 (1 μM) in standard extracellular solution (black) and
 92 in 100 mM extracellular Ca²⁺ (red trace). **B)** Reversal potentials of n = 6 individual cells
 93 measured in A) in 150 mM extracellular Na⁺ and 100 mM extracellular Ca²⁺, mean values
 94 represented in black squares. **C)** As in panel A) but for TRPM3α2. **D)** Reversal potentials of n
 95 = 6 individual cells (grey circles) measured in C) in 150 mM extracellular Na⁺ and 100 mM
 96 extracellular Ca²⁺; mean values represented in black squares. * < 0.05 with student two-tailed
 97 unpaired t-test.

98

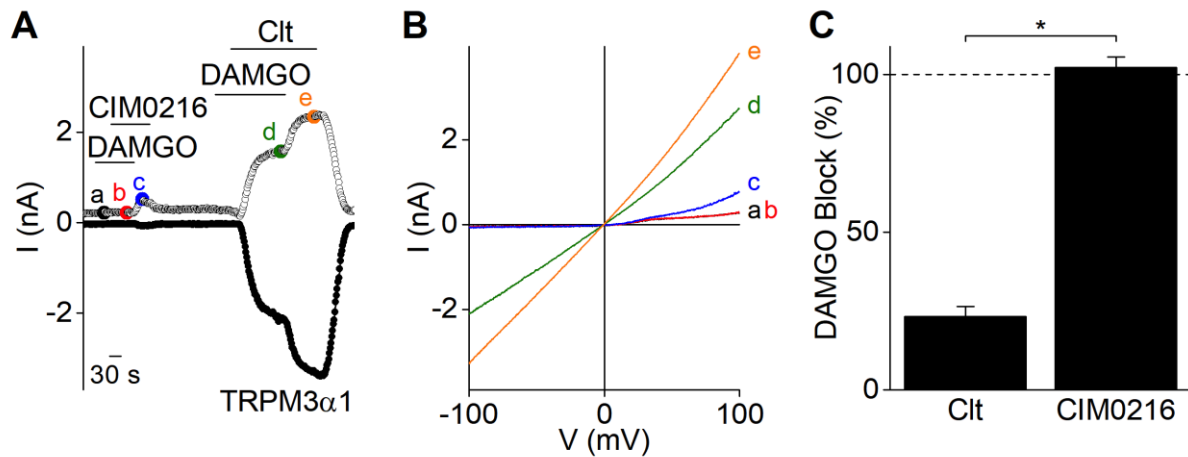


99

100 **FIGURE S9**

101 **A)** Time course of whole-cell patch clamp recording at \pm 80 mV in HEK293T cells transiently
 102 expressing TRPM3 α 1 upon application of CIM0216 (1 μ M) and the TRPM3 antagonists
 103 isosakuranetin (Iso; 5 μ M), diclofenac (Dic; 100 μ M) and primidone (Pri; 25 μ M). **B)** Time
 104 course of whole-cell patch clamp recording at \pm 80 mV in HEK293T cells transiently expressing
 105 TRPM3 α 1 upon application of different concentrations of Isosakuranetin (1 - 100 μ M) during
 106 CIt-treatment (10 μ M). **C)** Concentration-response curve for Isosakuranetin on TRPM3 α 1 at
 107 +80 mV (red) and -80 mV (black trace) extracted from experiments shown in *B*) (n = 4).

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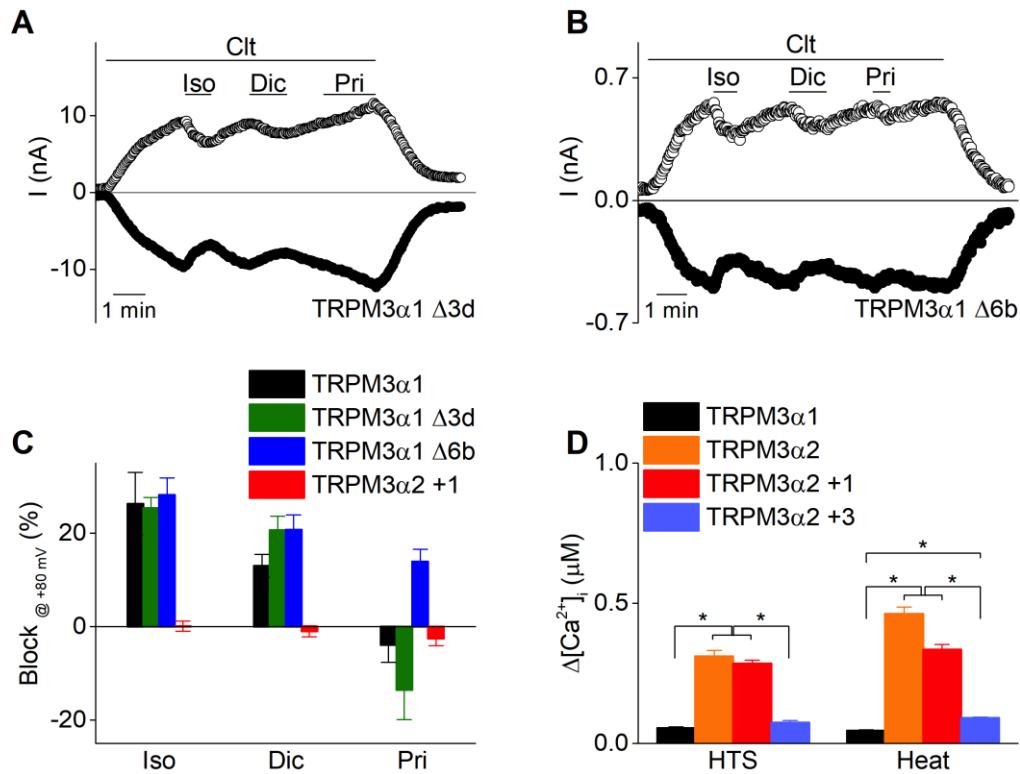


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111 **FIGURE S10**

112 **A)** Time course of whole-cell patch clamp recording at ± 80 mV in HEK293T cells co-expressing
 113 TRPM3 α 1 and μ -opioid receptors upon application of DAMGO (1 μ M), CIM0216 (5 μ M) and
 114 Clt (10 μ M). **B)** *I-V* Traces for time points indicated in **A**). **C)** Percentage of block induced by
 115 DAMGO in experimental conditions of panel **A**) ($n = 8$). * < 0.05 with student two-tailed
 116 unpaired t-test.

117



118

119 **FIGURE S11**

120 **A)** Time course of a whole-cell patch clamp recording at ± 80 mV in HEK293T cells transiently
 121 expressing deletion mutant TRPM3 α 1 Δ 3d upon application of Clt (10 μ M) and different
 122 TRPM3 antagonists (isosakuranetin (Iso; 5 μ M), diclofenac (Dic; 100 μ M), primidone (Pri; 25
 123 μ M)). **B)** As in A) but for the deletion mutant TRPM3 α 1 Δ 6b. **C)** Percentage of block calculated
 124 from experiments in A) and B) and compared to results of TRPM3 α 1 and insertion mutant
 125 TRPM3 α 2 +1; n = 5 for TRPM3 α 1, n = 3 for TRPM3 α 1 Δ 3d, n = 4 for TRPM3 α 1 Δ 6b and n = 6
 126 for TRPM3 α 2 +1. **D)** Statistics of calcium increases induced by hypotonicity (HTS) and heat in
 127 HEK293T cells transfected with TRPM3 α 1, TRPM3 α 2 or the mutant channels TRPM3 α 2 +1
 128 and TRPM3 α 2 +3. For HTS: n = 678 for TRPM3 α 1, n = 547 for TRPM3 α 2, n = 539 for TRPM3 α 2
 129 +1 and n = 788 for TRPM3 α 2 +3. For heat: n = 484 for TRPM3 α 1, n = 651 for TRPM3 α 2, n =
 130 751 for TRPM3 α 2 +1, n = 487 for TRPM3 α 2 +3. N = 3 independent recordings for each
 131 experiment. * < 0.05, Kruskal-Wallis ANOVA with Dunn's posthoc test.