Table S1: RNA EXTRACTION AND REAL TIME POLYMERASE CHAIN REACTION

RNA extraction was performed on liver sample snap frozen at -80° C in RNAlater® solution (Invitrogen, Thermofisher). An equal amount of each sample (200 mg) was suspended in 600 µl of TRIzol solution (Invitrogen, Thermofisher) together with 0.5 mm zirconium oxide beads (Next Advanced, NY). The homogenization of tissue was performed using a Bullet Blender (Next Advanced, NY) at a speed of 8 rpm for three minutes at 4°C. The homogenized tissue was centrifuged at 12,000 g for ten minutes at 4°C, and the collected supernatant was subjected to RNA extraction according to manufacturer's protocol. Briefly, 200 µl of chloroform was added to each sample and after three minutes recovery at RT, samples were centrifuged at 12,000 g for 15 minutes at 4° C. Chloroform-RNA phase was collected, and isopropanol was added. After ten minutes recovery at RT, samples were centrifuged at 12,000 g for ten minutes 4°C. The pelleted RNA was washed with absolute ethanol and centrifuged at 12,000 g for five minutes 4°C. The isolated RNA was suspended in RNAse free water and quantified by NanoDrop 2000 (Thermofisher); then, a total amount of 200 ng of RNA per sample was retro-transcribed in a 20 µl of total volume reaction using High Capacity cDNA Reverse Transcription Kit (Applied Biosystems, Thermofisher). The real time polymerase chain reaction (RT-PCR) was performed according to the chemistry of Power SYBR® Green PCR Master Mix (Applied Biosystems, Thermofisher), on 5 ng of cDNA and using specific oligonucleotide primers (listed in the following table) and ran by StepOne Real Time System (Applied Biosystems, Thermofisher). The ΔΔCt method was used to quantify the expression of the genes of interests. All the values were normalized towards the Actin-β (housekeeping gene) while livers from four healthy rats, named SHAM group, were used as reference samples. Data were expressed as Relative Quantification (RQ) mean ± SEM.

Gene	Forward primer (5'→3')	Reverse primer (3'→5')
ACTβ	ACCGTGAAAAGATGACCCAGAT	CACAGCCTGGATGGCTACGT
BAX	AGCAAACTGGTGCTCAAGGC	GCCACCCTGGTCTTGGATC
BCL-2	GAGGCTGGGATGCCTTTGT	AGGCTGAGCAGCGTCTTCAG
BECLIN-1	TATCTGGCACAGCGGACAATT	CCAGGCAGCATTGATTTCATT
CD14	CTACCGACCATGAAGCTTATGCT	CGTCCTGGTCCAGCTCACA
CXCL-10	CTGAGTCTGAGTGGGACTCAAGG	GACAGGATAGACTTGCAGGAATGA
e-NOS	AGTTACCAGCTGGCCAAAGTGA	ACGATCCAGGCCCAGTCA
E-SELECTIN	ACACAGCTTCCTGTACCAACACAT	CCTGTTCTTGGCAGGTCACA
ICAM1	ATCACTGTGTATTCGTTCCCAGAG	CACGGAGCAGCACTACTGAGAG
IL-6	AGAAAAGAGTTGTGCAATGGCAAT	CATCCATCATTTCTTTGTATTTCTGG
mTOR	ATTGCCAGCCTCATTGGAGT	TCTCCATGACAACTGGATCACTTG
NF-kB	CATCACCCATGGCACCATAA	CGGTCTTGGTGCTGGCC
P-SELECTIN	AACCTGCAAAGGTGTAACATCACTT	GTTCGGACCAAAGCTTTCCA
SOD1	CAATACACAAGGCTGTACCACTGC	AGGTCTCCAACATGCCTCTCTTC
TLR4	GAATCCCTGCATAGAGGTACTTCCTA	TTGAAGCTCAGATCTAGGTTCTTG
ULK-1	CCTACTAAGGCTGGACCCTCG	GCAGTGTACGGTTCCGAGGT
VCAM1	TGTCAACGTTGCTCCGAAAG	CTTTAGCTGTCTGCTCCACAGGA

Forward and reverse oligonucleotides sequences of RT-PCR primers.