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Supplementary Information for the article:

Reduced physiological plasticity in a fish adapted to stable temperatures

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Supplementary text

Estimation of the number of generations AB fish held in lab

According to the ZFIN website (https://zfin.org/action/genotype/view/ZDB-GENO-960809-7), the AB line was brought into the lab in the 1970s. By 1991, the AB fish were 70 generations removed from the AB-wt fish used to establish the line. From this we can assume that 3.4 generations were produced per year (since 21 years = 70 generations). Since we do not have an exact number of generations from then, as each lab has different protocols, we extrapolated these numbers to give us an estimate of the number of generations the AB-wt (lab) fish had been in a laboratory environment at the start of this experiment. Extrapolating from 1991-2017 (26 years \times 3.4 generations) gives 88.4 generations and a total of 158 generations from establishing the line. We therefore used the estimate of over 150 generations in this paper.

Supplementary Data

Link to all raw data: https://figshare.com/s/91f03a69303257e477f1

Supplementary figures

Figure S1: Thermal profiles showing the raw traces for how acclimation temperatures (10-38°C; (a) wild fish, 15 aquaria & (b) lab fish, 15 aquaria) were reached and maintained over the entire experimental period (38°C was terminated before the end of the experiment due to high mortality)

Figure S2: Growth (length), condition and maximum swim speed (individuals) of lab and wild fish acclimated from 10-36(38)°**C.** b - Specific growth rate for length for each individual fish. c - Condition of fish after 35 days of acclimation (calculated as the Fulton´s condition index). d - Maximum swim speed of individual fish measured in a swim respirometer. (see Table S4 & S6)

Figure S3: Heat map showing the difference in liver (a) and brain (b) gene expression between wild and lab fish. If there are no differences between wild and lab fish then the colour is white, this then proceeds along a colour gradient with the strongest green showing a higher expression in wild fish and the darkest blue a higher expression in lab fish. Genes are grouped according to their function and colours represent relative expression within these groups. Fish were

Figure S4: Metabolic gene expression using qPCR in wild and lab zebrafish acclimated from 10-36°**C**. a - Relative quantity of AMP-activated protein kinase subunit alpha-1 (*prkaa1*); b - Relative quantity of succinate dehydrogenase subunit A (*sdha*); c- Relative quantity of thyroid hormone receptor alpha-A (*thraa*). Wild fish are illustrated by green circles and lab fish by blue triangles.

Figure S5: Behaviours corresponding to Fig. 2e & f, pre (a) and post (b) alarm cue time spent bottom dwelling and post-cue activity (c) in lab and wild zebrafish acclimated from 10-36°**C**. Wild fish: green circles, lab fish: blue circles. Statistically significant differences indicated on each panel: Temp –significant effect of temperature on trait, Pop – significant difference between wild and lab fish (at 23°C), Pop x Temp – significant interaction (see Table S4- S6)

ACCLIMATION TEMPERATURE (°C)

Figure S6: Additional behaviours: time spent freezing, time at the surface, and distance from the surface, in lab and wild zebrafish acclimated from 10-36°**C**. All behaviours were analysed pre-alarm cue (baseline) and post-alarm cue. The alarm cue response was quantified as the change in behaviour. Wild fish: green circles, lab fish: blue circles. Statistically significant differences indicated on each panel: Temp –significant effect of temperature on trait, Pop – significant difference between wild and lab fish (at 23°C), Pop x Temp – significant interaction (see Table S4-S6)

Figure S7: Coefficient of variation for weight of the fish at the (a) start and (b) end of the experiment. During phenotyping not all individuals from each tank were measured but the measured ones were assumed to be representative for that temperature. The coefficient of variation in weight at the start and end of the experiment shows how much variation there was at each temperature and how representative the measured individuals are. At the start of the experiment the two populations (wild: green circles; lab: blue triangles) show consistent variation within population but overall there is less variation in the lab than wild. By the end of the experiment the variation at the colder temperatures was similar as the start but at the warmer temperatures both wild and lab fish had converged and show similar variation.

Figure S8: Design of the behavioural arena. The behavioural tests were conducted in eight parallel arenas (a) with one fish in each (b). The tanks had white back and sides, and clear front where the video recording was taken from. A pipette tip introduced chemical cues at the centre top of the arenas.

Supplementary Tables

Table S1: Statistical output from ANOVA´s comparing whether acclimation temperature (T), population (P; wild or lab) and the interaction between them (TxP) differs in a range of traits.

Table S2: Model estimates and standard errors. Population estimates are related to the intercept (lab population at 23°C)

Table S3: Primers used for gene expression analysis by qPCR

* Used as housekeeping gene in muscle; + used as house keeping gene in liver; ¥ used as housekeeping gene in brain

Table S4: Statistical output from ANOVA´s comparing whether acclimation temperature (T), population (P; wild or lab) and the interaction between them $(T\times P)$ differs in a range of traits (supplementary data traits only)

Table S5: Statistical output from Chi-squared tests for one physiological and six behavioural traits. The test sequentially adds acclimation temperature (T), population (P; wild or lab) and the interaction between them $(T\times P)$ into the model to compare whether there is a significant difference from the NULL model.

Table S6: Model estimates and standard errors for extended data phenotypes. Population estimates are related to the intercept (lab population at 23°C) Estimates
from proportion data are derived from glm models with a binom

Table S7: Optimal temperatures (T_{ork}) and thermal performance breadths (TPB; 80% performance) of lab and wild populations of zebrafish for five phenotypes. The
difference in thermal performance breadth is calculated as:

Supplementary References

1. J. Tian, K. Wang, X. Wang, H. Wen, H. Zhou, C. Liu, K. Mai, G. He, Soybean saponin modulates nutrient sensing pathways and metabolism in zebrafish. Gen. Comp. Endocrinol. 257, 246–254 (2018).

2. C. Best, M. M. Vijayan, Cortisol elevation post-hatch affects behavioural performance in zebrafish larvae. Gen. Comp. Endocrinol. 257, 220–226 (2018).

3. V. Stancová, A. Ziková, Z. Svobodová, W. Kloas, Effects of the non-steroidal antiinflammatory drug(NSAID) naproxen on gene expression of antioxidant enzymes in zebrafish (Danio rerio). Environ. Toxicol. Pharmacol. 40, 343–348 (2015).

4. Y. Yang, W. Liu, D. Li, L. Qian, B. Fu, C. Wang, Altered glycometabolism in zebrafish exposed to thifluzamide. Chemosphere. 183, 89–96 (2017).

5. O. Monroig, J. Rotllant, E. Sánchez, J. M. Cerdá-Reverter, D. R. Tocher, Expression of longchain polyunsaturated fatty acid (LC-PUFA) biosynthesis genes during zebrafish Danio rerio early embryogenesis. Biochim. Biophys. Acta 1791, 1093–1101 (2009).

6. G. M. Her, C.-C. Hsu, J.-R. Hong, C.-Y. Lai, M.-C. Hsu, H.-W. Pang, S.-K. Chan, W.-Y. Pai, Overexpression of gankyrin induces liver steatosis in zebrafish (Danio rerio). Biochim. Biophys. Acta. 1811, 536–548 (2011).

7. Y.-S. Shieh, Y.-S. Chang, J.-R. Hong, L.-J. Chen, L.-K. Jou, C.-C. Hsu, G. M. Her, Increase of hepatic fat accumulation by liver specific expression of Hepatitis B virus X protein in zebrafish. Biochim. Biophys. Acta. 1801, 721–730 (2010).

8. C. C. Lazado, S. Nayak, I. Khozin-Goldberg, D. Zilberg, The gut mucosal barrier of zebrafish (Danio rerio) responds to the time-restricted delivery of Lobosphaera incisa-enriched diets. Fish Shellfish Immunol. 89, 368–377 (2019).

9. Y. Yun, Y. Zhang, G. Li, S. Chen, N. Sang, Embryonic exposure to oxy-polycyclic aromatic hydrocarbon interfere with pancreatic β-cell development in zebrafish via altering DNA methylation and gene expression. Sci. Total Environ. 660, 1602–1609 (2019).

10. D. Boyle, G. G. Goss, Effects of silver nanoparticles in early life-stage zebrafish are associated with particle dissolution and the toxicity of soluble silver. NanoImpact. 12, 1–8 (2018).

11. Y. Yang, F. Dong, X. Liu, J. Xu, X. Wu, W. Liu, Y. Zheng, Crosstalk of oxidative damage, apoptosis, and autophagy under endoplasmic reticulum (ER) stress involved in thifluzamideinduced liver damage in zebrafish (Danio rerio). Environ. Pollut. 243, 1904–1911 (2018).

12. M. Qi, Y. Dang, Q. Xu, L. Yu, C. Liu, Y. Yuan, J. Wang, Microcystin-LR induced developmental toxicity and apoptosis in zebrafish (Danio rerio) larvae by activation of ER stress response. Chemosphere. 157, 166–173 (2016).

13. V. Christen, M. Capelle, K. Fent, Silver nanoparticles induce endoplasmatic reticulum stress response in zebrafish. Toxicol. Appl. Pharmacol. 272, 519–528 (2013).