

Chemistry–A European Journal

Supporting Information

Negishi Cross-Coupling Provides Alkylated Tryptophans and Tryptophan Regioisomers

Steffen Dachwitz, Bjarne Scharkowski, and Norbert Sewald*

Instruments

Analytical HPLC

Analytical HPLC was performed on a Shimadzu NexeraXR 20A System with autosampler, degasser, column oven, diode array detector and a Phenomenex Luna C18 column (2.9 μm , 50 \times 2.1 mm) with a gradient (in 5.5 min from 5 % B to 95 % B, 0.5 min 95 % B and back to 5 % B in 3 min, total run time 9 min) at a flow rate of 650 $\mu\text{L}/\text{min}$ and column oven temperature of 40 $^{\circ}\text{C}$. HPLC solvent A consists of 99.9% water with 0.1 % TFA, solvent B of 99.9 % acetonitrile with 0.1 % TFA.

Analytical LC-MS

Analytical LC-MS was performed on an Agilent 6220 TOF-MS with a Dual ESI-source, 1200 HPLC system with autosampler, degasser, binary pump, column oven, diode array detector and a Hypersil Gold C18 column (1.9 μm , 50 \times 2.1 mm) with a gradient (in 11 min from 0 % B to 98 % B, back to 0 % B in 0.5 min, total run time 15 min) at a flow rate of 300 $\mu\text{L}/\text{min}$ and column oven temperature of 40 $^{\circ}\text{C}$. HPLC solvent A consists of 94.9 % water, 5 % acetonitrile and 0.1 % formic acid, solvent B of 5 % water, 94.9 % acetonitrile with 0.1 % formic acid. ESI mass spectra were recorded after sample injection via 1200 HPLC system in extended dynamic range mode equipped with a Dual-ESI source, operating with a spray voltage of 2.5 kV.

NMR

NMR spectra were recorded on a Bruker Avance III 500 HD (^1H : 500 MHz, ^{13}C : 126 MHz) or Avance 600 (^1H : 600 MHz, ^{13}C : 151 MHz). Chemical shifts δ [ppm] are reported relative to residual solvent signal (CDCl_3 , ^1H : 7.26 ppm, ^{13}C : 77.1 ppm; $\text{DMSO}-d_6$, ^1H : 2.50 ppm, ^{13}C : 39.5 ppm; $\text{Methanol}-d_4$, ^1H : 3.31 ppm ^{13}C : 49.0 ppm). 2D spectra (COSY, HMQC, HMBC) were used for signal assignment.

General Procedures

GP1: Ni-catalysed reductive cross-coupling

Arylbromide (125 μmol), NiI_2 (4.7 mg, 13 μmol , 10 mol%) and 4,4'-Di-*tert*-butyl-2,2'-dipyridine (3.8 mg, 13 μmol , 10 mol%) were placed in a glass vial under argon atmosphere. The mixture was suspended in DMPU (0.5 mL) and purged with argon. Alkyl iodide (250 μmol , 2.0 eq.), pyridine (1.0 μL , 13 μmol , 10 mol%) and manganese (500 μmol , 4.0 eq.) were added. The green reaction mixture was heated to 60 $^{\circ}\text{C}$ and stirred for 21 h. The mixture was purified directly by column chromatography (Petrolether/EtOAc; 3:1).

GP2: Pd-catalysed Negishi cross-coupling (analytical scale)

Arylbromide (250 μmol , 1.0 eq.), zinc dust (1.5-4.0 eq.), Alkyl iodide (1.5-2.0 eq.) and $\text{Pd}(\text{amphos})_2\text{Cl}_2$ (5 mol%) were placed in a glass vial followed by DMF (0.5 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The crude

mixture was diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL) the organic phase was dried over MgSO₄ and the solvent removed in vacuum. The crude product was purified by automated column chromatography using a Büchi Reveleris X2 (Petroleum:EtOAc).

GP2b: Pd-catalysed Negishi cross-coupling (preparative scale)

Arylbromide (5.00 mmol, 1.0 eq.), zinc dust (1.5 eq.), Alkyl iodide (1.5 eq.) and Pd(amphos)₂Cl₂ (5 mol%) were placed in a glass vial followed by DMF (1-10 mL) The suspension was purged with argon and stirred at 37 °C for 24 h. The crude mixture was filtered over a short plug of silica and washed with Petroleum/EtOAc; 4:1, the solvent was evaporated in vacuum, giving a dark red oil. The crude product was purified by column chromatography (DCM/MeOH; 99:1)

GP3: Marfey's Test

The enantiopurities were determined using the chiral reagent FDAA (Marfey's reagent, 1-fluoro-2-4-dinitrophenyl-5-L-alanine amide). For this purpose, a 3.33 mM amino acid stock solution in a 1:2 mixture of aqueous 0.1 M NaHCO₃ and acetone (300 µL, 1.0 eq.) and a 15 mM FDAA stock solution in acetone (100 µL, 1.5 eq.) were mixed and incubated for 1 h at 40 °C. The resulting suspension was neutralized with aqueous 0.1 M HCl solution (150 µL) and diluted 1:1 with of acetonitrile. The obtained solution was analysed by LC-MS.

Analytical LC-MS was performed on an Agilent 6220 TOF-MS with a Dual ESI-source, 1200 HPLC system with autosampler, degasser, binary pump, column oven, diode array detector and a Hypersil Gold C18 column (1.9 µm, 50 × 2.1 mm) with a gradient (in 30 min from 5 % B to 55 % B, 3 min at 55 % B, back to 5 % B in 3 min, total run time 40 min) at a flow rate of 300 µL/min and column oven temperature of 40 °C. HPLC solvent A consists of 94.9 % water, 5 % acetonitrile and 0.1 % formic acid, solvent B of 5 % water, 94.9 % acetonitrile and 0.1 % formic acid. UV detection took place at 340 nm. ESI mass spectra were recorded after sample injection via 1200 HPLC system in extended dynamic range mode equipped with a Dual-ESI source, operating with a spray voltage of 2.5 kV. Nitrogen served both as the nebulizer gas and the dry gas. Nitrogen was generated by a nitrogen generator NGM 11.

GP4: Acid free deprotection of regioisomeric tryptophan surrogates

The protected tryptophan regioisomer (1.0 mmol) was suspended in Na₂HPO₄-Buffer (100 mM, pH = 7.4, 15 mL). The mixture was heated to reflux for 9 hours. The aqueous phase was washed with EtOAc (2x15 mL). The solvent of the aqueous phase was reduced in vacuum and the crude product was desalted using an acid-free reversed phase C18-column chromatography, washing with pure MPW (100 mL) and eluting the product with MeOH (100 mL). The enantiomeric excess was determined using Marfey's Test (see GP3).

GP5: Acidic deprotection of alkylated tryptophan derivatives

The protected alkylated tryptophan (25 µmol) was suspended in aqueous HCl (5 M, 2 mL) and heated to 90 °C for 2 hours. The crude reaction mixture was purified by reversed phase HPLC. The enantiomeric excess was determined using Marfey's Test (see GP3).

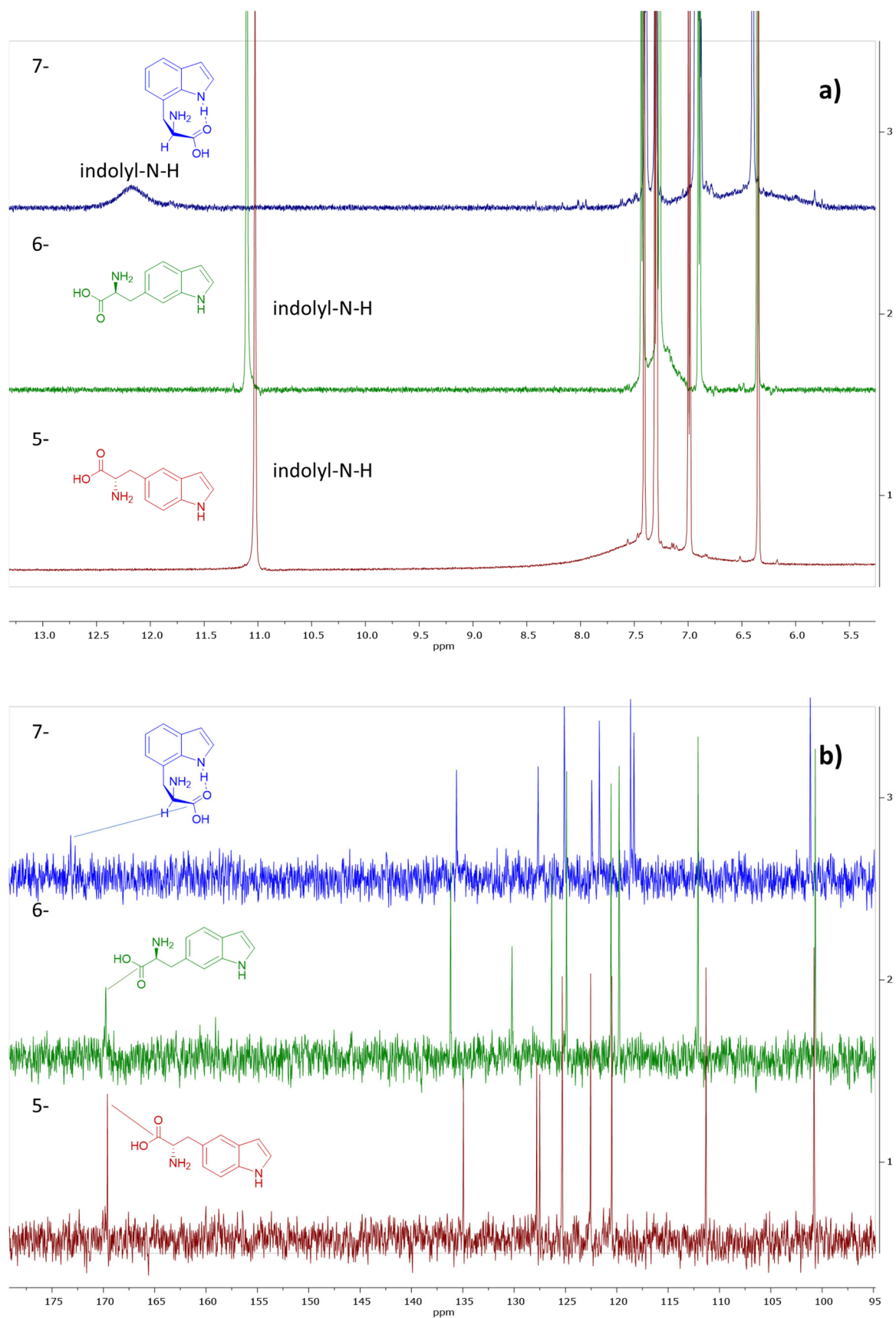
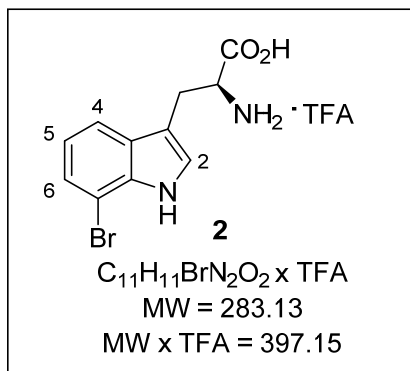


Figure S1: a) Comparison of ^1H NMR of the 3-(5-,6- and 7-indolyl)alanine **11b-d**; b) Comparison of ^{13}C NMR of the 3-(5-,6- and 7-indolyl)alanine **11b-d**.

Compounds

L-7-Bromotryptophan x TFA



L-7-bromotryptophan (**2**) was synthesized according to our previously reported procedure using RebH-PrnF-RR-ADH combiCLEAs.^[3] The biocatalyst was produced using lysed *E. coli* cells containing overexpressed tryptophan-7-halogenase RebH resulting from 1.5 L expression culture. The reaction buffer contained 1 mM L-tryptophan, 15 mM Na₂HPO₄, 30 mM NaBr, 0.1 mM NAD⁺, 1 μM FAD and 0.5 % (v/v) 2-propanol at pH = 7.4 in a total reaction volume of 1.250 L. Full conversion was usually observed after 2-4 days. The suspension was filtered and desalted.

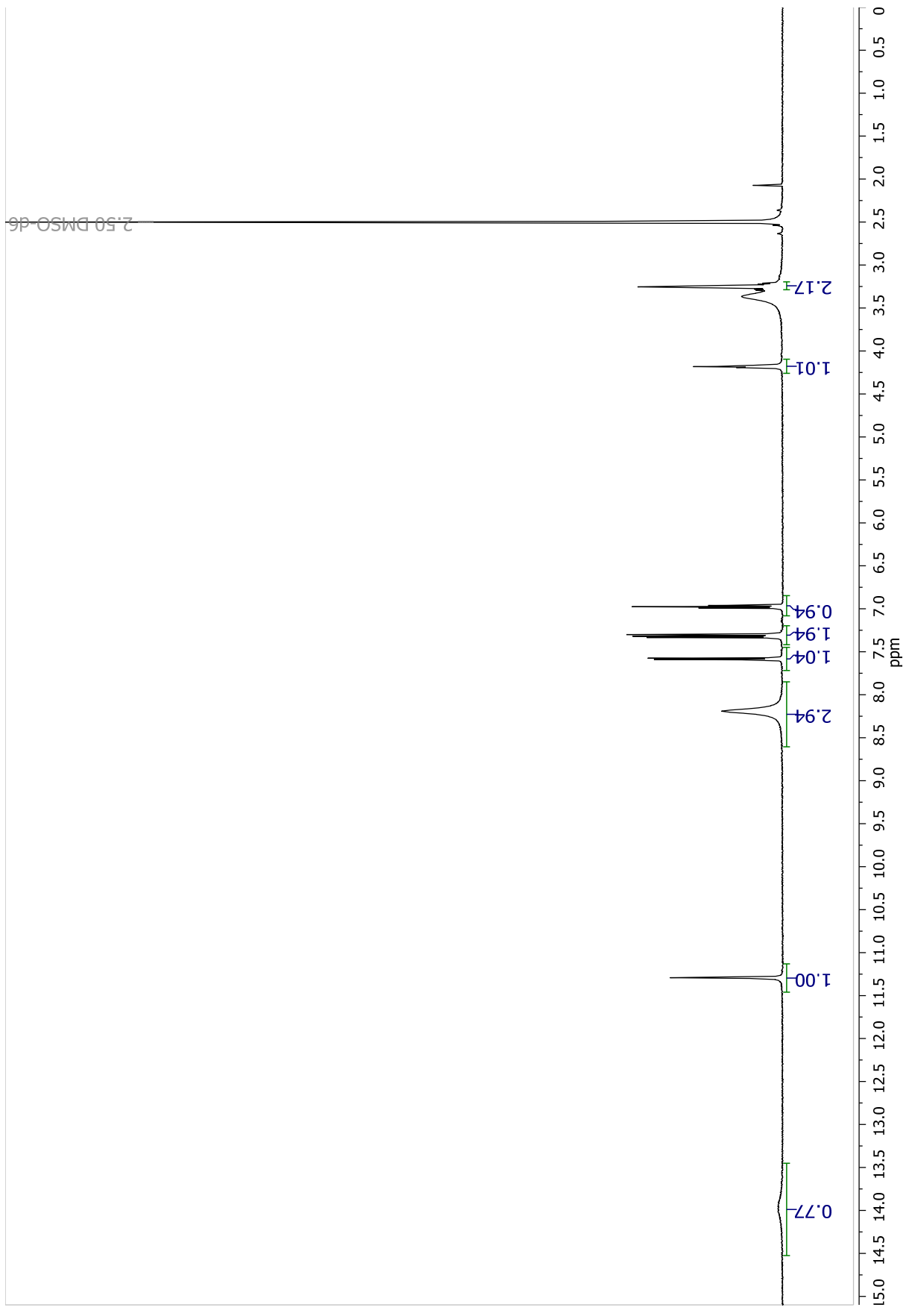
Therefore, the crude filtrate was concentrated up to a volume of about 100 mL and loaded on a 12 g RP-C₁₈-column and purified using an automated column chromatography using a Büchi Reveleris X2 with a binary pump and ELSD Detector. The gradient (4 min at 5% B, up to 25% B in 14 min, in 1 min up to 100 % B for 2 min and flushing with 80% B for 5 min, total run time 27 min) was used at a flow rate of 30 mL/min. Solvent A consisted of 99.9 % water and 0.1% TFA, solvent B of 99.9 % acetonitrile and 0.1 % TFA. Freeze drying gave L-7-bromotryptophan x TFA as a colorless to yellow solid.

Anal. RP-HPLC: $t_R = 3.3$ min;

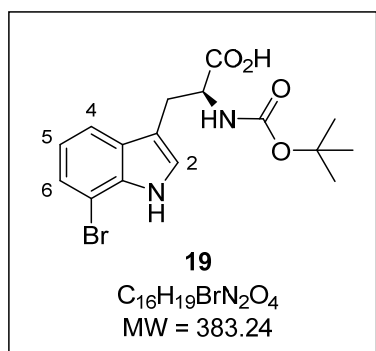
LC-MS: $t_R = 5.2$ min;

¹H NMR (500 MHz, DMSO-d₆) δ [ppm] = 13.96 (br s, 1H, COOH), 11.29 (d, ³J = 2.7 Hz, 1H, indole-NH), 8.19 (brs, 3H, NH₃), 7.58 (d, ³J = 7.9 Hz, 1H, C4-H), 7.33 (d, ³J = 7.5 Hz, 1H, C6-H), 7.30 (d, ³J = 2.7 Hz, 1H, C2-H), 6.98 (dd, ³J = 7.8 Hz, ³J = 7.8 Hz, 1H, C5-H), 4.18 (dd, ³J = 7.1 Hz, ³J = 6.2 Hz, 1H, C α -H), 3.27 (dd, ²J = 15.0 Hz, ³J = 5.7 Hz, 1H, C β -H), 3.23 (dd, ²J = 14.8 Hz, ³J = 6.9 Hz, 1H, C β -H);

MS (ESI): found $[m/z] = 265.98$ [M(⁷⁹Br)-NH₂]⁺, 267.98 [M(⁸¹Br)-NH₂]⁺, 283.01 [M(⁷⁹Br)+H]⁺, 285.01 [M(⁸¹Br)+H]⁺; calcd. $[m/z] = 265.98$ [M(⁷⁹Br)-NH₂]⁺, 267.98 [M(⁸¹Br)-NH₂]⁺, 283.01 [M(⁷⁹Br)+H]⁺, 285.01 [M(⁸¹Br)+H]⁺.



N^α-Boc-L-7-Bromotryptophan (**19**)



L-7-bromotryptophan x TFA (**2**, 159.7 mg, 402 μmol) was dissolved in 1,4-dioxane (5 mL) followed by addition of di-tert-butyl dicarbonate (105.8 mg, 484 μmol, 1.2 eq.) and aqueous NaOH (1.0 M, 800 μL, 800 μmol, 2.0 eq.). The reaction progress was monitored by analytical HPLC. After completion, the solvent was removed in vacuum and the crude residue was dissolved in water (25 mL) and adjusted to pH = 3 by addition of aqueous HCl (1.0 M). The aqueous layer was extracted with EtOAc (3 x 50 mL) and the combined organic layers were dried over MgSO₄.

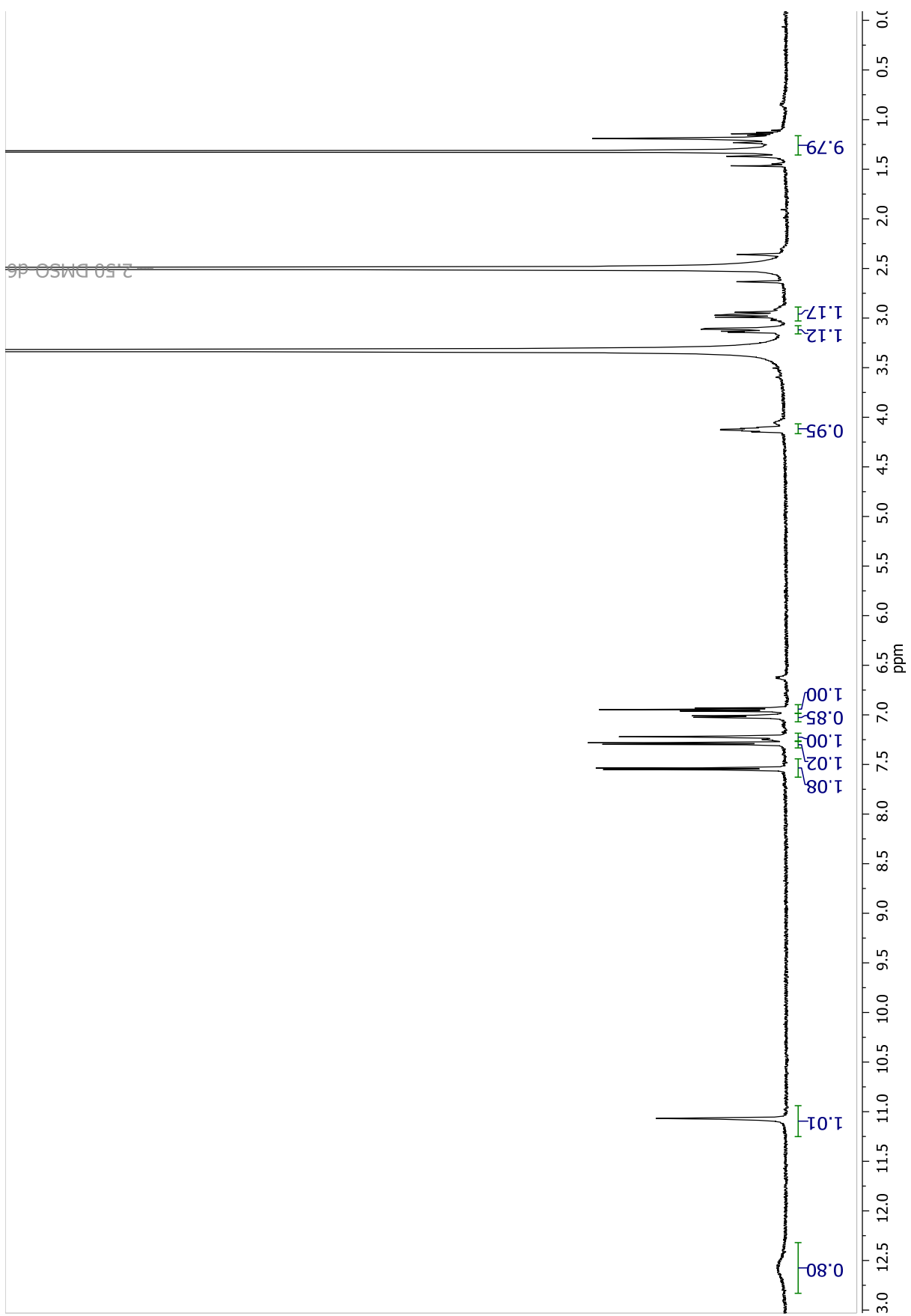
The organic solvent was removed in vacuum and the crude product was purified by RP-HPLC providing *N*^α-Boc-L-7-bromotryptophan (**19**) as a colorless solid (106.2 mg, 277 μmol, 69 %).

Anal. RP-HPLC: t_R = 5.0 min;

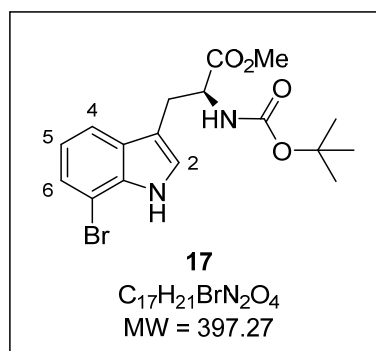
LC-MS: t_R = 8.7 min;

¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 12.58 (br s, 1H, COOH), 11.07 (d, ³*J* = 2.6 Hz, indole-NH), 7.54 (d, ³*J* = 7.8 Hz, 1H, C4-H), 7.29 (d, ³*J* = 7.5 Hz, 1H, C6-H), 7.22 (d, ³*J* = 2.6 Hz, 1H, C2-H), 7.02 (d, ³*J* = 7.9 Hz, 1H, OCONH), 6.95 (dd, ³*J* = 7.7 Hz, ³*J* = 7.7 Hz, 1H, C5-H), 4.13 (ddd, ³*J* = 9.3 Hz, ³*J* = 8.0 Hz, ³*J* = 4.7 Hz, 1H, C α -H), 3.12 (dd, ²*J* = 14.7, ³*J* = 4.7 Hz, 1H, C β -H), 2.97 (dd, ²*J* = 15.0 Hz, ³*J* = 9.6 Hz, 1H, C β -H), 1.32 (s, 9H, C(CH₃)₃; cis/trans ratio 5:1);

MS (ESI): found [*m/z*] = 381.05 [M(⁷⁹Br)-H]⁻, 383.05 [M(⁸¹Br)-H]⁻; calcd. [*m/z*] = 381.05 [M(⁷⁹Br)-H]⁻, 383.05 [M(⁸¹Br)-H]⁻.



N^α-Boc-L-7-Bromotryptophan methyl ester (**17**)



L-7-Bromotryptophan x TFA (**17**) (272.5 mg, 963 μmol) was dissolved in methanol (5 mL) and cooled to 0 °C followed by dropwise addition of thionyl chloride (150 μL, 2.0 mmol, 2.0 eq.). The mixture was heated to reflux and the reaction progress was monitored by analytical HPLC. After completion (1 h), the solvent was removed in vacuum and the crude residue was dissolved in ACN (10 mL) and adjusted to pH = 9 by addition of aqueous NaOH (1.0 M) followed by addition of di-*tert* butyl-dicarbonate (355.2 mg, 1.6 mmol, 1.7 eq.). The reaction progress was monitored by analytical HPLC and after

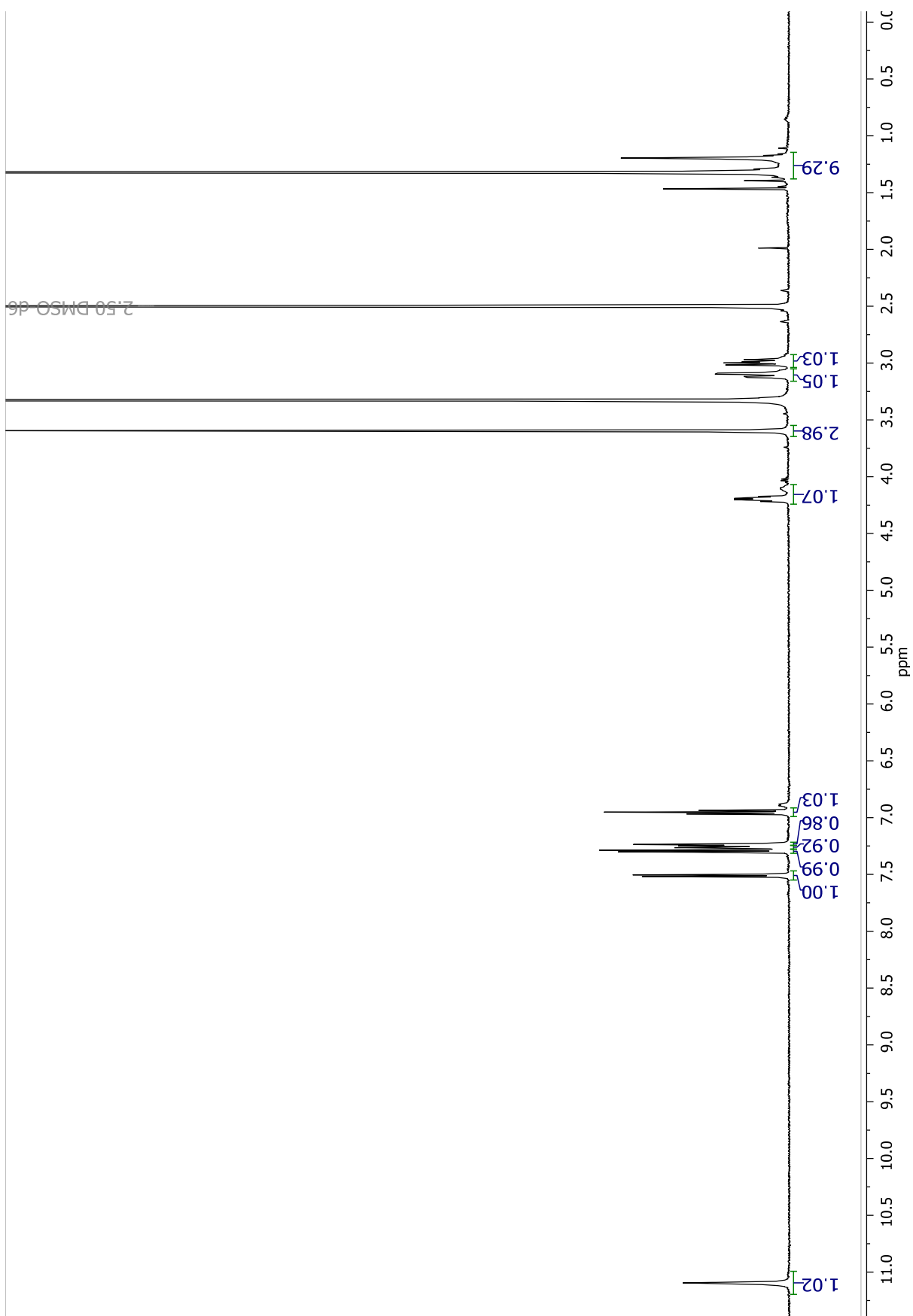
completion (20 h), the solvent was removed in vacuum and the crude residue was suspended in water (50 mL) and adjusted to pH = 4 by addition of aqueous HCl (1.0 M). The aqueous layer was extracted with EtOAc (3 x 70 mL) and the combined organic layers were dried over MgSO₄. The organic solvent was removed in vacuum and the crude product was purified by column chromatography (Petrolether/EtOAc; 4:1) providing *N*^α-Boc-L-7-bromotryptophan methyl ester (**17**) as a colorless solid (280.5 mg, 706 μmol, 73 %).

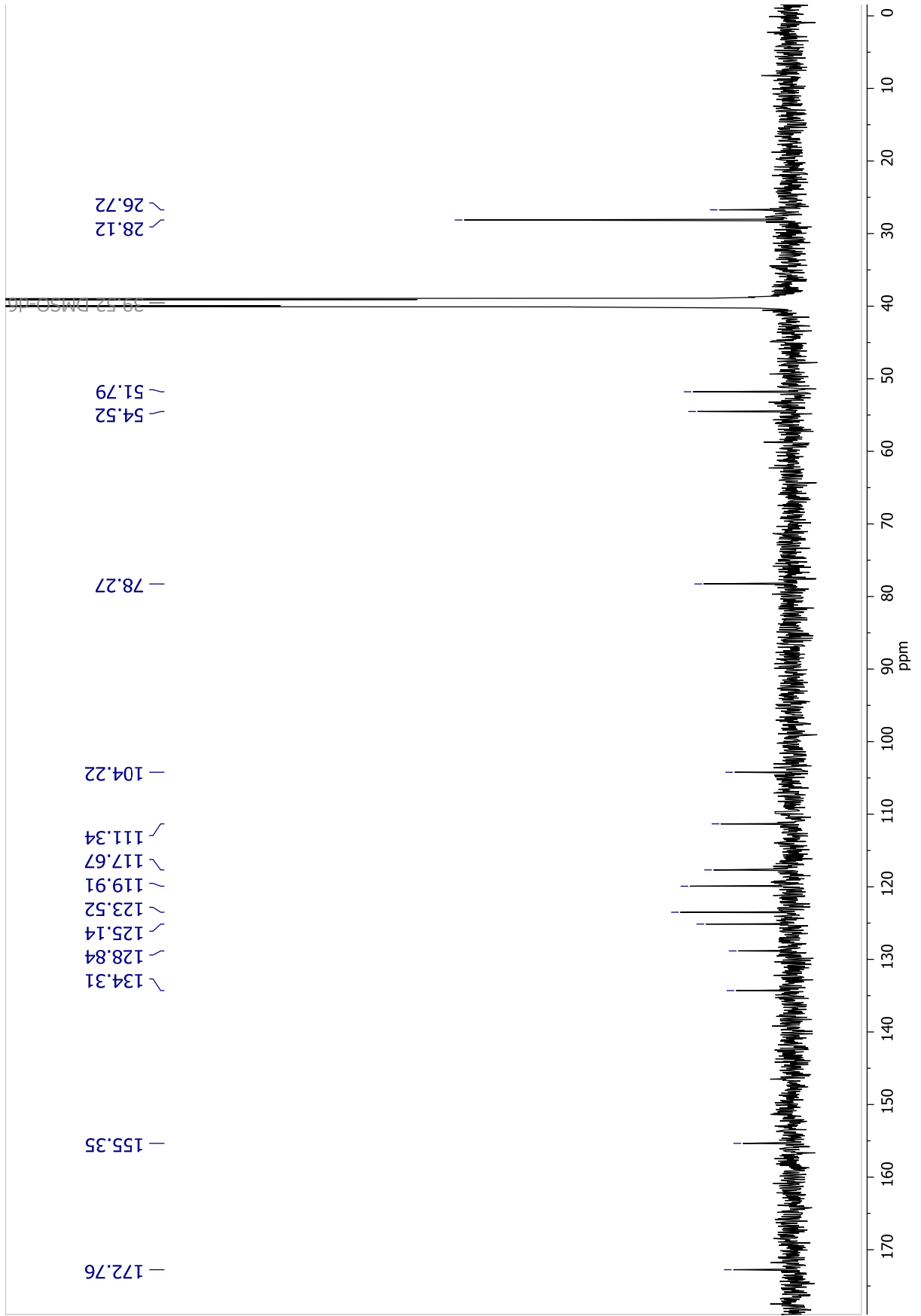
LC-MS: *t*_R = 9.7 min;

¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 11.09 (s, 1H, Indole-NH), 7.51 (d, ³*J* = 7.9 Hz, 1H, C4-H), 7.29 (d, ³*J* = 7.5 Hz, 1H, C6-H), 7.26 (d, ³*J* = 7.9 Hz, 1H, OCONH), 7.24 (d, ³*J* = 2.6 Hz, 1H, C2-H), 6.95 (dd, ³*J* = 7.8 Hz, 1H, C5-H), 4.20 (ddd, ³*J* = 9.4 Hz, ³*J* = 7.9 Hz, ³*J* = 5.3 Hz, 1H, Cα-H), 3.60 (s, 3H, CH₃), 3.11 (dd, ³*J* = 14.8 Hz, ³*J* = 5.0 Hz, 1H, Cβ-H), 2.99 (dd, ³*J* = 14.6 Hz, ³*J* = 9.5 Hz, 1H, Cβ-H), 1.32 (s, 9H, C(CH₃)₃; cis/trans ratio 5:1).

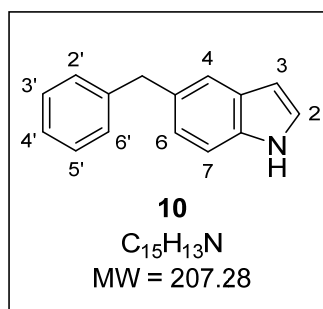
¹³C NMR (126 MHz, DMSO-*d*₆) δ [ppm] = 172.8 (COO(CH₃)), 155.4 (NHC=O), 134.3 (C7a), 128.8 (C4a), 125.1 (C2), 123.5 (C6), 119.9 (C5), 117.7 (C4), 111.3 (C3), 104.2 (C7), 78.2 (C(CH₃)₃), 54.5 (Cα), 51.8 (OCH₃), 28.1(C(CH₃)₃), 26.7 (Cβ).

MS (ESI): found [*m/z*] = 419.06 [M(⁷⁹Br)+Na]⁺, 421.05 [M(⁸¹Br)+Na]⁺, 341.01 [M(⁷⁹Br)-(*tert*Butyl)+H]⁺, 343.01 [M(⁸¹Br)-(*tert*Butyl)+H]⁺; 299.02 [M(⁷⁹Br)-Boc+H]⁺, 297.02 [M(⁸¹Br)-Boc+H]⁺, 279.99 [M(⁷⁹Br)-Boc-NH₂]⁺, 281.99 [M(⁸¹Br)-Boc-NH₂]⁺, calcd. [*m/z*] = 419.06 [M(⁷⁹Br)+Na]⁺, 421.05 [M(⁸¹Br)+Na]⁺, 341.01 [M(⁷⁹Br)-(*tert*Butyl)+H]⁺, 343.01 [M(⁸¹Br)-(*tert*Butyl)+H]⁺; 299.02 [M(⁷⁹Br)-Boc+H]⁺, 297.02 [M(⁸¹Br)-Boc+H]⁺, 279.99 [M(⁷⁹Br)-Boc-NH₂]⁺, 281.99 [M(⁸¹Br)-Boc-NH₂]⁺.





5-Benzylindole (**10**)



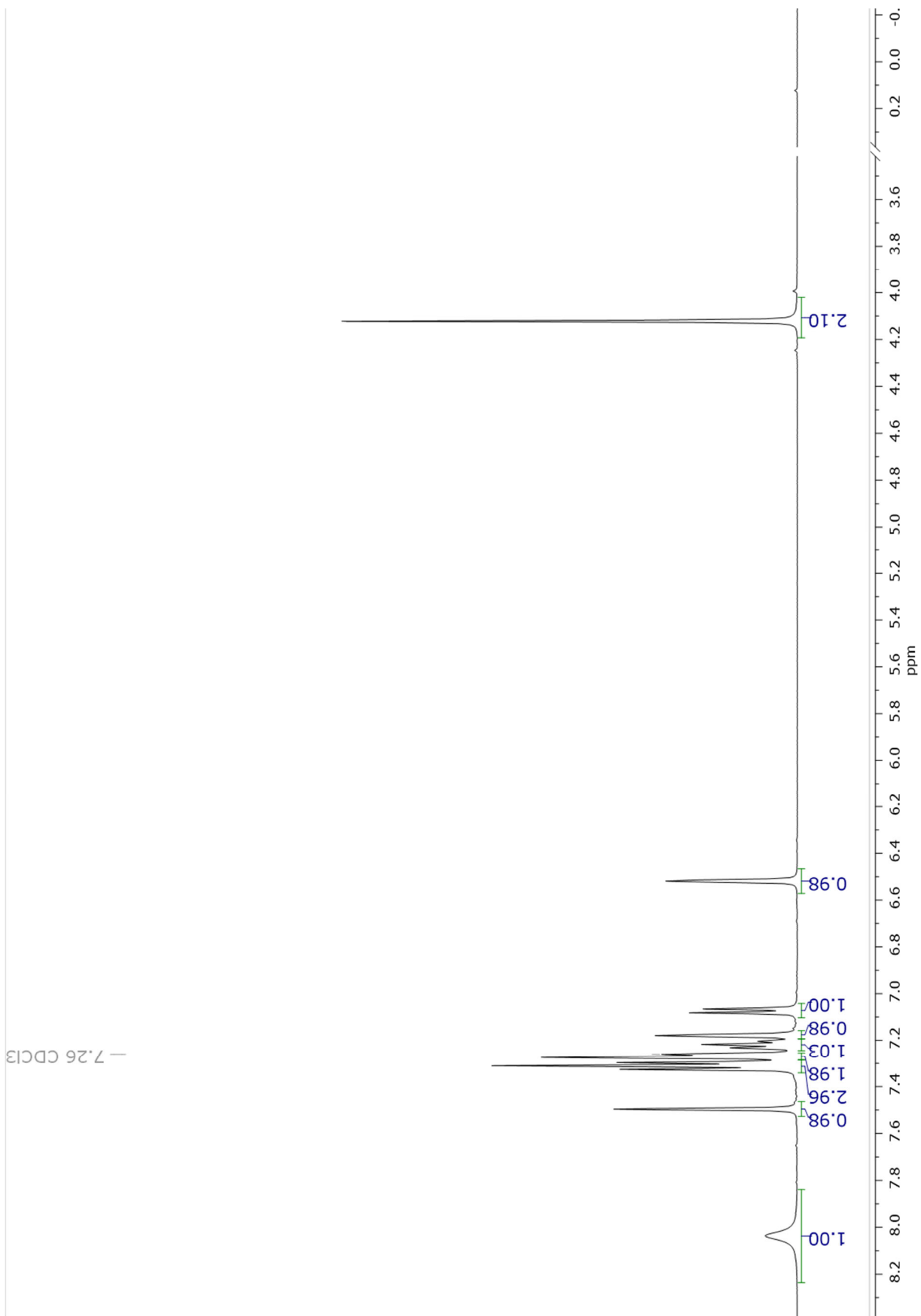
5-Benzylindole (**10**) was synthesized according to GP2. 5-Bromoindole (**1b**, 50.5 mg, 258 μ mol), zinc dust (69.9 mg, 1069 μ mol, 4 eq.) and Pd(amphos)₂Cl₂ (8.5 mg, 12.5 μ mol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL). The suspension was degassed by sparkling argon through it. Freshly distilled benzyl iodide (**9**, 103.5 μ L, 523 μ mol, 2 eq.) was added and the mixture was stirred at room temperature for 4 h. The solvent was evaporated in vacuum and the crude product was purified by column chromatography (Petrolether/EtOAc; 4:1) providing **10** as a pale brown solid (42.5 mg, 205 μ mol, 79 %).

Anal. RP-HPLC: t_R = 5.6 min;

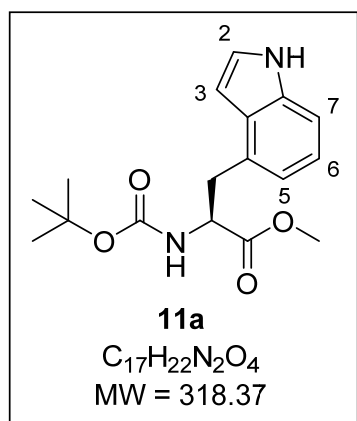
LC-MS: t_R = 9.7 min;

¹H NMR (500 MHz, Chloroform-*d*) δ [ppm] = 8.05 (br s, 1H, Indole-NH), 7.52 (s, 1H, C4-H), 7.38 – 7.31 (m, 3H, C2'-H/C6'-H/C7-H), 7.30 (m, 2H, C3'-H/C5'-H), 7.22 (m, 1H, C4'-H), 7.20 (dd, ³J = 3.5 Hz, ³J = 1.1 Hz, 1H, C2-H), 7.10 (dd, ³J = 8.4 Hz, ⁴J = 1.8 Hz, 1H, C6-H), 6.55 (dd, ³J = 3.5 Hz, ⁴J = 2.1 Hz, 1H, C3-H), 4.15 (s, 2H, CH₂).

MS (ESI): found $[m/z]$ = 208.11 [M+H]⁺, calcd. $[m/z]$ = 208.11 [M+H]⁺.



N^α-Boc-L-3-(1*H*-4-Indolyl)alanine methyl ester (**11a**)



Analytical scale:

N^α-Boc-L-3-(1*H*-4-Indolyl)alanine methyl ester (**11a**) was synthesized according to *GP2*. Therefore, 4-Bromoindole (**1a**, 32 μ L, 255 μ mol), zinc dust (24.9 mg, 382 μ mol, 1.5 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (164.4 mg, 399 μ mol, 1.5 eq.) and Pd(amphos)₂Cl₂ (9.2 mg, 13 μ mol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL) The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL) the organic phase was dried over MgSO₄ and

the solvent removed in vacuum. The crude product was purified by automated column chromatography using a Büchi Reveleris X2 (Petrolether:EtOAc) giving **11a** as a colorless solid (47.7 mg, 150 μ mol, 59 %).

Upscaling:

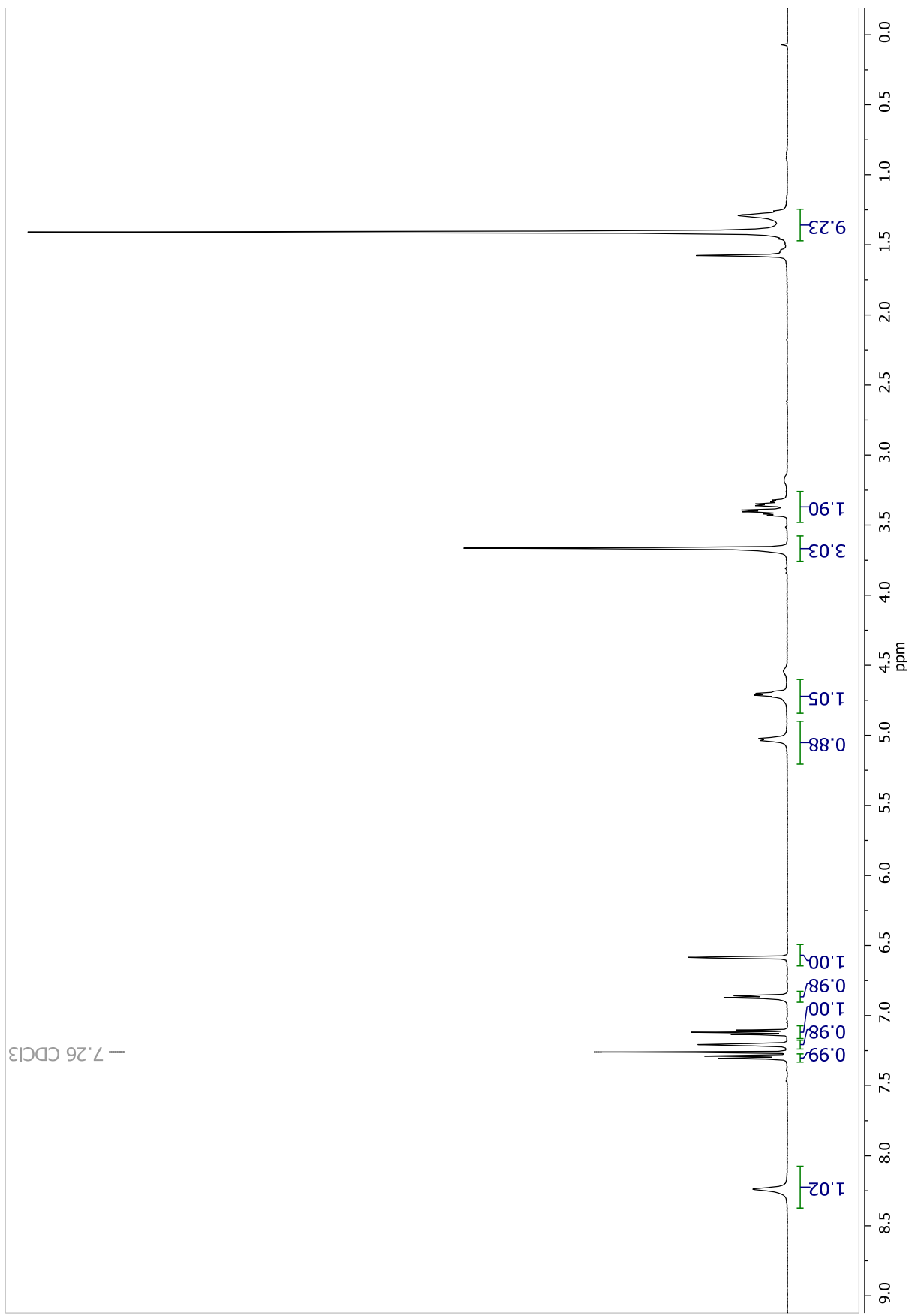
N^α-Boc-L-3-(1*H*-5-Indolyl)alanine methyl ester (**11a**) was synthesized according to *GP2b*. Therefore, 4-Bromoindole (**1a**, 0.627 mL, 5.00 mmol), zinc dust (0.496 g, 7.59 mmol, 1.5 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (**8**, 2.456 g, 7.46 mmol, 1.5 eq.) and Pd(amphos)₂Cl₂ (176 mg, 250 μ mol, 5 mol%) were placed in a glass vial followed by DMF (1 mL) The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was filtered over a short plug of silica and washed with Petrolether/EtOAc; 4:1, the solvent was evaporated in vacuum, giving a dark red oil. The crude product was purified by column chromatography (DCM/MeOH; 99:1) providing **11a** as a colorless solid (487.2 mg, 1.53 mmol, 31 %).

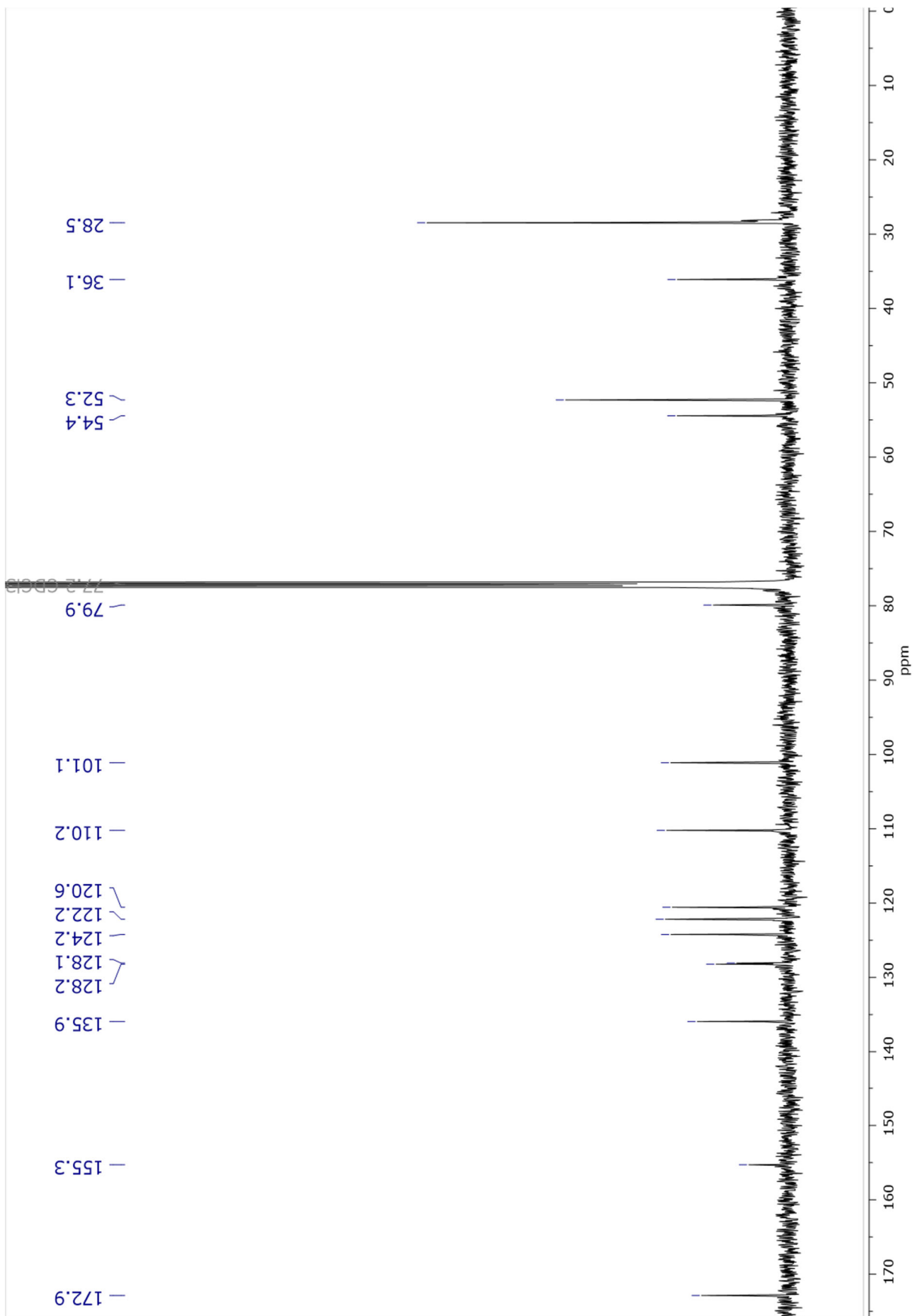
LC-MS: t_R = 8.7 min;

¹H NMR (500 MHz, CDCl₃) δ [ppm] = 8.24 (s, 1H, Indole-NH), 7.30 (d, ³*J* = 8.1 Hz, 1H, C7-H), 7.21 (dd, ³*J* = 4.8 Hz, ³*J* = 2.9 Hz, 1H, C2-H), 7.12 (dd, ³*J* = 7.6 Hz, ³*J* = 7.6 Hz, 1H, C6-H), 6.87 (d, ³*J* = 7.2 Hz, 1H, C5-H), 6.58 (dd, ⁴*J* = 2.8, ³*J* = 4.6 Hz, 1H, C3-H), 5.03 (d, ³*J* = 6.3 Hz, 1H, OCONH), 4.71 (ddd, ³*J* = 6.6 Hz ³*J* = 6.3 Hz, ³*J* = 6.1 Hz, 1H, C α -H), 3.66 (s, 3H, CH₃), 3.41 (dd, ³*J* = 13.9 Hz, ³*J* = 6.0 Hz, 1H, C β -H), 3.34 (dd, ³*J* = 14.1 Hz, ³*J* = 6.6 Hz, 1H, C β -H), 1.41 (s, 9H, C(CH₃)₃).

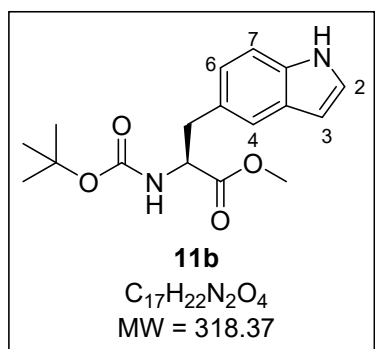
¹³C NMR (126 MHz, CDCl₃) δ [ppm] = 172.9 (COOMe), 155.3 (NHCOO), 135.9 (C7a), 128.2 (C4a), 128.1 (C4), 124.2 (C2), 122.2 (C6), 120.5 (C5), 110.2 (C7), 101.1 (C3), 79.9 (C(CH₃)₃), 54.4 (C α), 52.3 (COO(CH₃)), 36.1 (C β), 28.5 (C(CH₃)₃).

MS (ESI): found [*m/z*] = 319.16 [M+H]⁺, 263.10 [M-(*tert*Butyl)+H]⁺, 219.12[M-Boc+H]⁺, 202.09 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 319.17 [M+H]⁺, 263.10 [M-(*tert*Butyl)+H]⁺, 219.11 [M-Boc+H]⁺, 202.09 [M-Boc-NH₂]⁺.





N^α-Boc-L-3-(1*H*-5-Indolyl)alanine methyl ester (**11b**)



Analytical scale:

N^α-Boc-L-3-(1*H*-5-Indolyl)alanine methyl ester (**11b**) was synthesized according to *GP2*. Therefore, 5-Bromoindole (**1b**, 50.07 mg, 255 μmol), zinc dust (66.07 mg, 1.01 mmol, 4.0 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (**8**, 165.66 mg, 503 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (9.1 mg, 13 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The

crude mixture was purified by column chromatography (PE/EtOAc; 5:1) providing **11b** as a colorless solid (44.2 mg, 138.8 μmol, 54 %).

Upscaling:

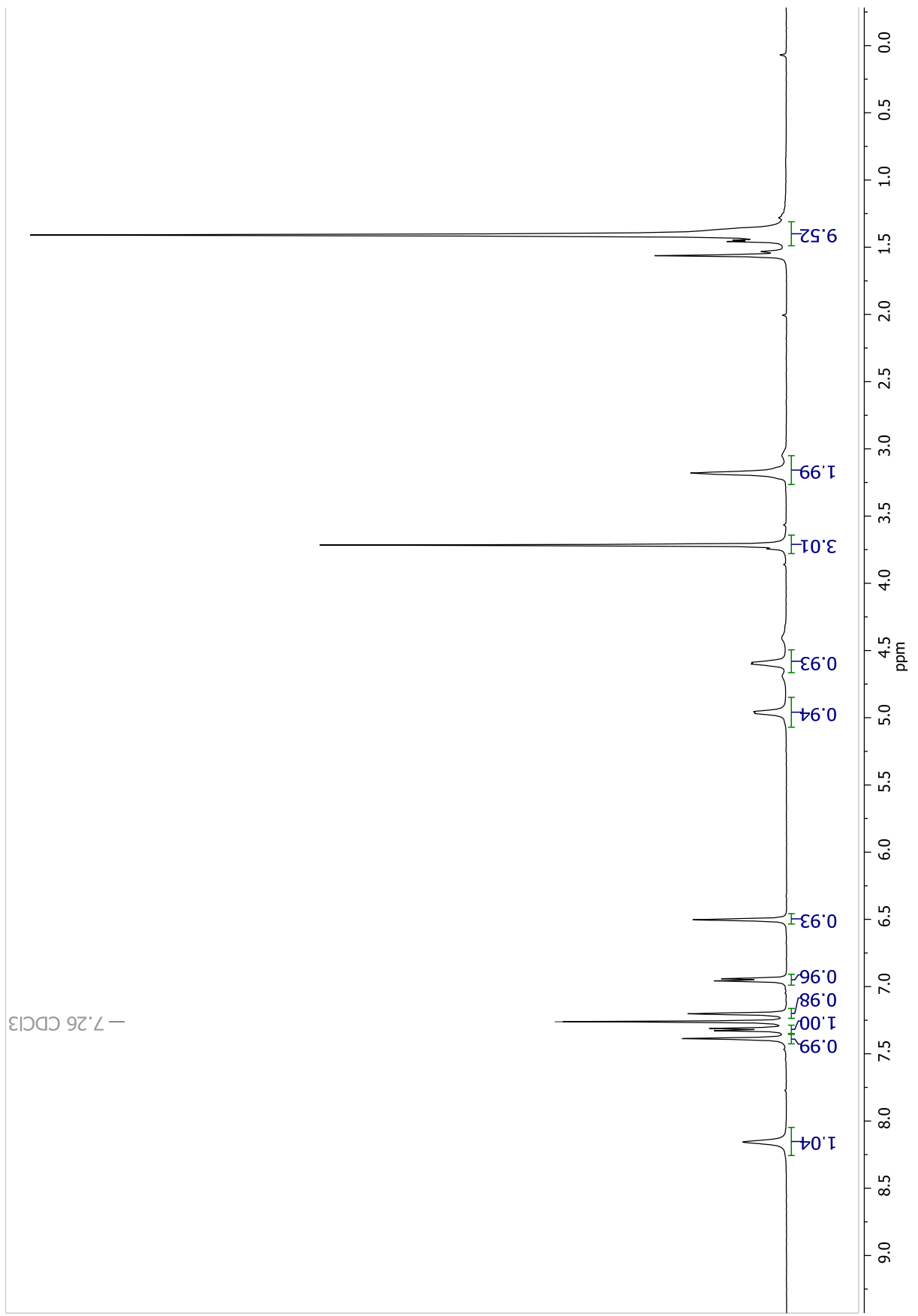
N^α-Boc-L-3-(1*H*-5-Indolyl)alanine methyl ester (**11b**) was synthesized according to *GP2b*. Therefore, 5-Bromoindole (**1b**, 0.981 g, 5.00 mmol), zinc dust (0.497 g, 7.60 mmol, 1.5 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (**8**, 1.816 g, 5.52 mmol, 1.1 eq.) and Pd(amphos)₂Cl₂ (179 mg, 250 μmol, 5 mol%) were placed in a glass vial followed by DMF (1 mL). The suspension was purged with argon and stirred at 37 °C for 24 h. The crude mixture was filtered over a short plug of silica and washed with Petrolether/EtOAc; 4:1, the solvent was evaporated in vacuum, giving a dark red oil. The crude product was purified by column chromatography (DCM/MeOH; 99:1) providing **11b** as a colourless solid (809.2 mg, 2.542 mmol, 51 %).

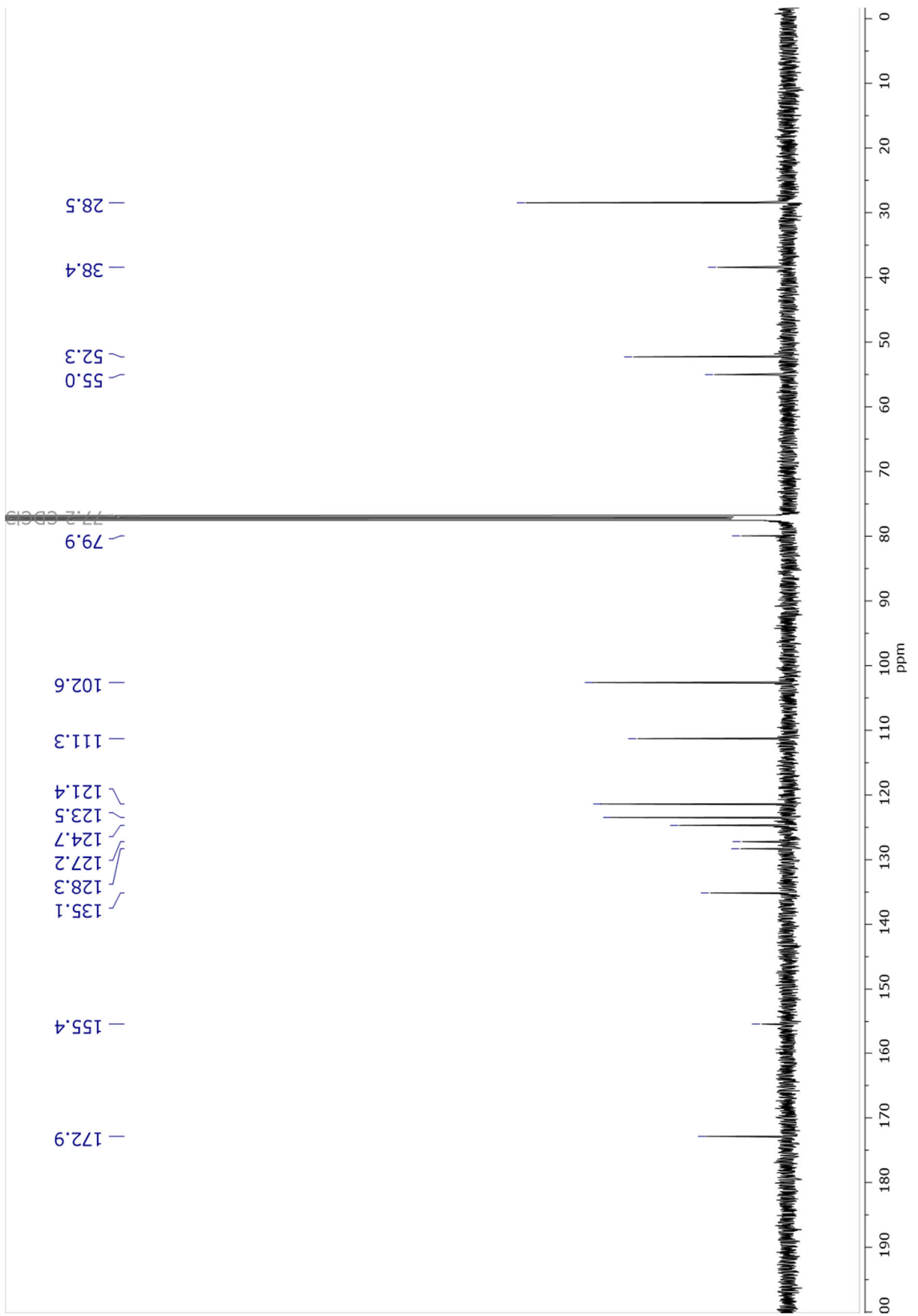
LC-MS: *t*_R = 8.9 min;

¹H NMR (500 MHz, Chloroform-*d*) δ [ppm] = 8.16 (s, 1H, Indole-NH), 7.39 (d, ⁴*J* = 1.8 Hz, 1H, C4-H), 7.32 (d, ³*J* = 8.4 Hz, 1H, C7-H), 7.20 (dd, ³*J* = 2.8 Hz, ³*J* = 2.7 Hz, 1H, C2-H), 6.95 (dd, ³*J* = 8.4, ⁴*J* = 1.7 Hz, 1H, C6-H), 6.50 (dd, ³*J* = 2.7 Hz, ⁴*J* = 1.6 Hz, 1H, C3-H), 4.96 (d, ³*J* = 7.6 Hz, 1H, OCON-H), 4.59 (ddd, ³*J* = 7.3 Hz, ³*J* = 6.6 Hz, ³*J* = 6.6 Hz, 1H, Cα-H), 3.72 (s, 3H, COOCH₃), 3.19 (dd, ³*J* = 11.5, ³*J* = 6.8 Hz, 1H, Cβ-H), 3.16 (dd, ³*J* = 12.0, ³*J* = 6.2 Hz, 1H, Cβ-H), 1.41 (s, 9H, C(CH₃)₃).

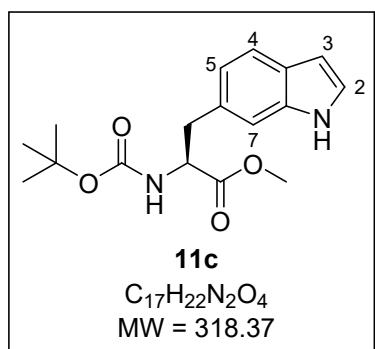
¹³C NMR (126 MHz, Chloroform-*d*) δ [ppm] = 172.8 (COOMe), 155.4 (OCONH), 135.1 (C7a), 128.3 (C3a), 127.2 (C5), 124.7 (C2), 123.4 (C6), 121.4 (C4), 111.3 (C7), 102.6 (C3), 79.9 (C(CH₃)₃), 55.0 (Cα), 52.3 (OCH₃), 38.4 (Cβ), 28.5 (C(CH₃)₃).

MS (ESI): found [*m/z*] = 319.16 [M+H]⁺, 263.10 [M-(*tert*Butyl)+H]⁺, 219.12 [M-Boc+H]⁺, 202.09 [M-Boc-NH₂]⁺; calcd. [*m/z*] = 319.17 [M+H]⁺, 263.10 [M-(*tert*Butyl)+H]⁺, 219.11 [M-Boc+H]⁺, 202.09 [M-Boc-NH₂]⁺.





N^α-Boc-L-3-(1*H*-6-Indolyl)alanine methyl ester (**11c**)



Analytical scale:

N^α-Boc-L-3-(1*H*-6-Indolyl)alanine methyl ester (**11c**) was synthesized according to *GP2*. Therefore, 6-Bromoindole (**1c**, 51.0 mg, 260 μmol), zinc dust (65.02 mg, 995 μmol, 3.8 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (**8**, 170.38 mg, 517 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (9.4 mg, 13 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The

crude mixture was diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL). The organic phase was dried over MgSO₄ and the solvent removed in vacuum. The crude product was purified by automated column chromatography using a Büchi Reveleris X2 (Petrolether:EtOAc) giving **11c** as a colourless solid (42.1 mg, 132 μmol, 51 %).

Upscaling:

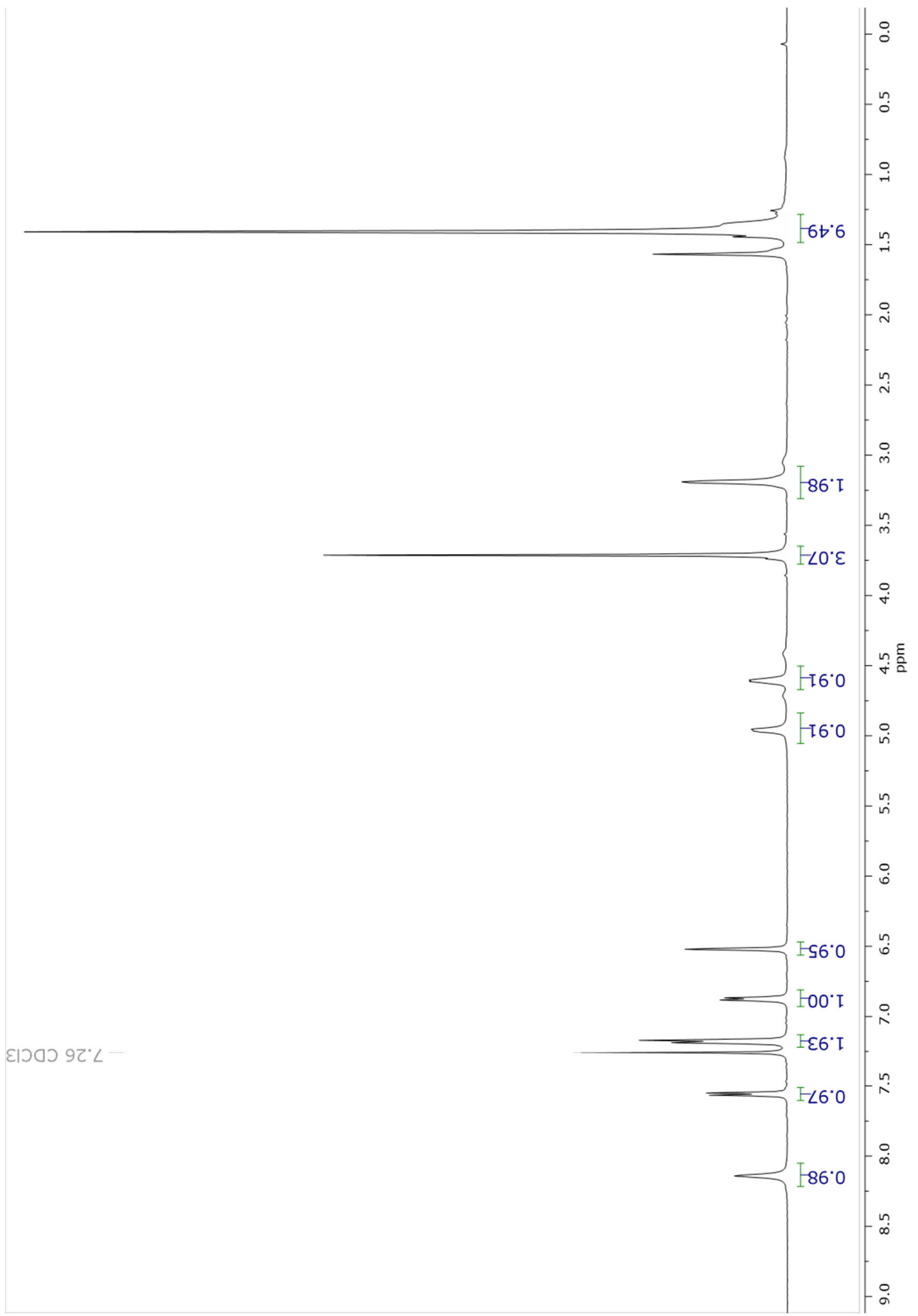
N^α-Boc-L-3-(1*H*-6-Indolyl)alanine methyl ester (**11c**) was synthesized according to *GP2b*. Therefore, 6-Bromoindole (**1c**, 980 mg, 5.00 mmol), zinc dust (490 mg, 7.50 mmol, 1.5 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (**8**, 1.833 g, 5.57 mmol, 1.1 eq.) and Pd(amphos)₂Cl₂ (178 mg, 249 μmol, 5 mol%) were placed in a glass vial followed by DMF (10 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was filtered over a short plug of silica and washed with Petrolether/EtOAc (4:1), the solvent was evaporated in vacuum, giving a dark red oil. The crude product was purified by column chromatography (DCM/MeOH; 99:1) providing **11c** as a colourless solid (482 mg, 1.51 mol, 30 %).

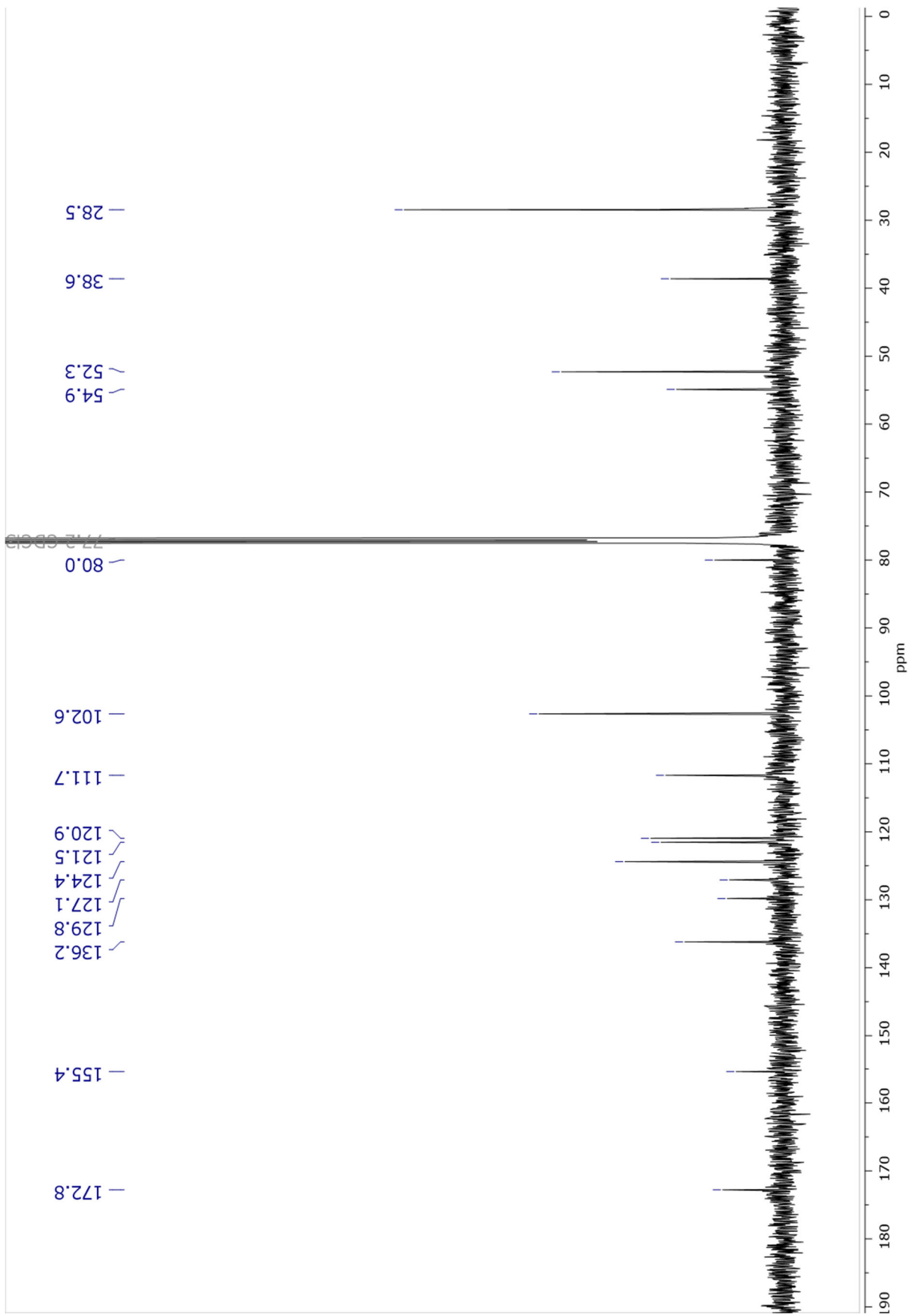
LC-MS: *t*_R = 8.6 min;

¹H NMR (500 MHz, Chloroform-*d*) δ [ppm] = 8.14 (s, 1H, Indole-NH), 7.56 (d, ³*J* = 8.4 Hz 1H, C4-H), 7.18 (m, 2H, C2-H/ C7-H), 6.88 (d, ³*J* = 8.1 Hz, 1H, C5-H), 6.52 (d, ³*J* = 2.0 Hz 1H, C3-H), 4.96 (d, ³*J* = 8.5 Hz, 1H, OCON-H), 4.61 (ddd, ³*J* = 9.2 Hz, ³*J* = 6.8 Hz, ³*J* = 6.8 Hz, 1H, C_α-H), 3.71 (s, 3H, COOCH₃), 3.19 (dd, ³*J* = 13.2 Hz, ³*J* = 6.8 Hz, 1H, C_β-H), 3.16 (dd, ³*J* = 13.0 Hz, ³*J* = 6.8 Hz, 1H, C_β-H), 1.41 (s, 9H, C(CH₃)₃).

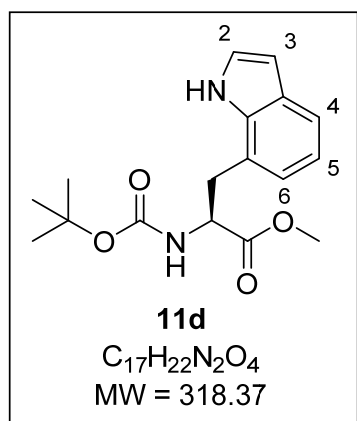
¹³C NMR (126 MHz, Chloroform-*d*) δ [ppm] = 172.8 (COOMe), 155.4 (OCONH), 136.2 (C7a), 129.8 (C3a), 127.1 (C6), 124.4 (C2), 121.5 (C4), 120.9 (C5), 111.6 (C7), 102.7 (C3), 80.0 (C(CH₃)₃), 54.9 (C_α), 52.3 (OCH₃), 38.6 (C_β), 28.5 (C(CH₃)₃).

MS (ESI): found [*m/z*] = 319.16 [M+H]⁺, 263.10 [M-(*tert*Butyl)+H]⁺, 219.12[M-Boc+H]⁺, 202.09 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 319.17 [M+H]⁺, 263.10 [M-(*tert*Butyl)+H]⁺, 219.11 [M-Boc+H]⁺, 202.09 [M-Boc-NH₂]⁺.





N^α-Boc-L-3-(1*H*-7-Indolyl)alanine methyl ester (**11d**)



Analytical scale:

N^α-Boc-L-3-(1*H*-7-Indolyl)alanine methyl ester (**11d**) was synthesized according to *GP2*. Therefore, 7-Bromoindole (**1d**, 50.1 mg, 255 μmol), zinc dust (65.1 mg, 995 μmol, 3.9 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (**8**, 164.1 mg, 499 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (9.1 mg, 13 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL). The organic phase was dried over

MgSO₄ and the solvent removed in vacuum. The crude product was purified by automated column chromatography using a Büchi Reveleris X2 (Petroleum ether:EtOAc) giving **11d** as a colourless solid (46.5 mg, 146 μmol, 58 %).

Upscaling:

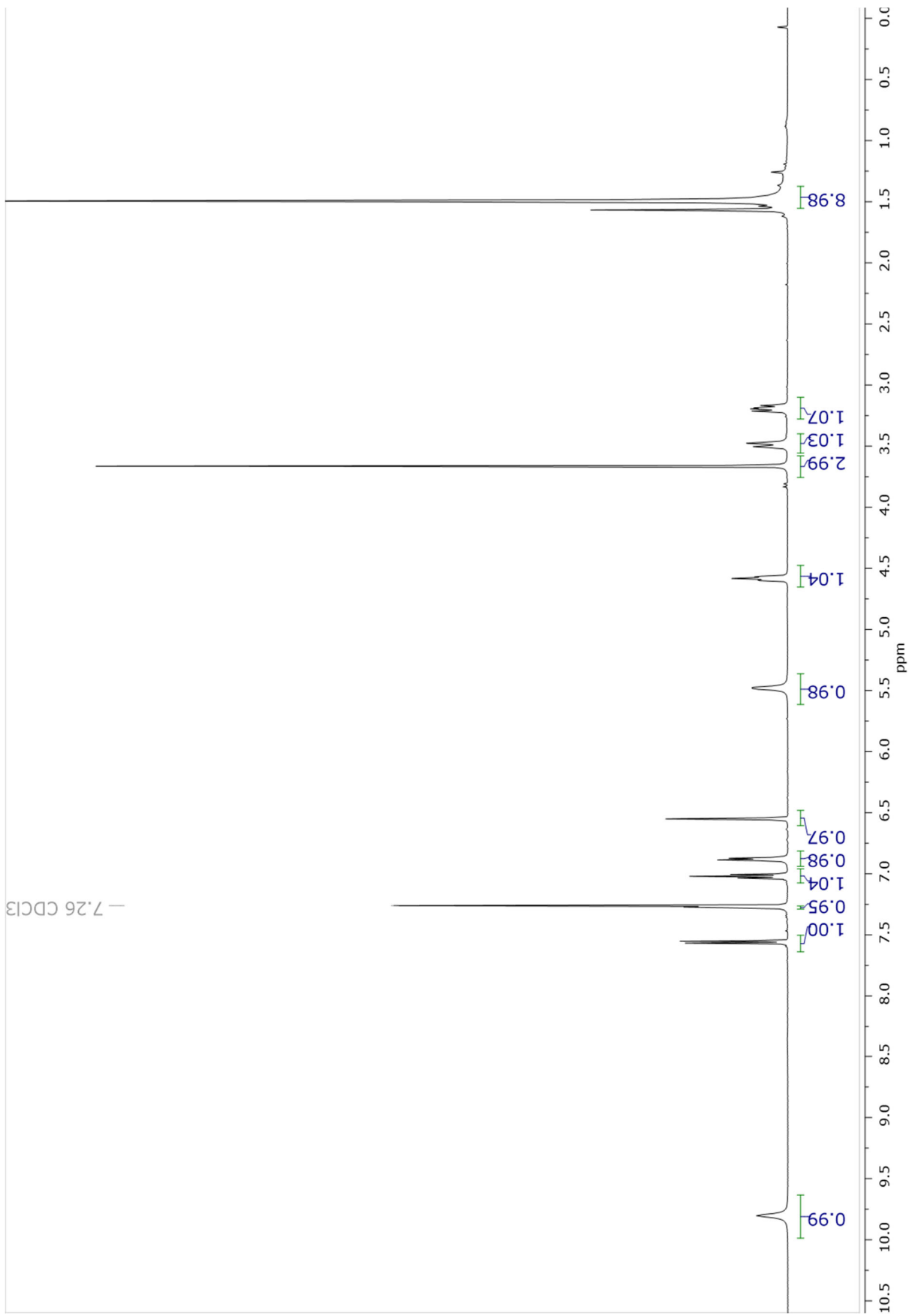
N^α-Boc-L-3-(1*H*-7-Indolyl)alanine methyl ester (**11d**) was synthesized according to *GP2b*. Therefore, 7-Bromoindole (**1d**, 980 mg, 5.00 mmol), zinc dust (490 mg, 7.50 mmol, 1.5 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (**8**, 2.501 g, 7.60 mmol, 1.5 eq.) and Pd(amphos)₂Cl₂ (178 mg, 249 μmol, 5 mol%) were placed in a glass vial followed by DMF (10 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was filtered over a short plug of silica and washed with Petroleum ether/EtOAc (4:1), the solvent was evaporated in vacuum, giving a dark red oil. The crude product was purified by column chromatography (DCM/MeOH; 99:1) providing **11d** as a colourless solid (586 mg, 1.84 mol, 37 %).

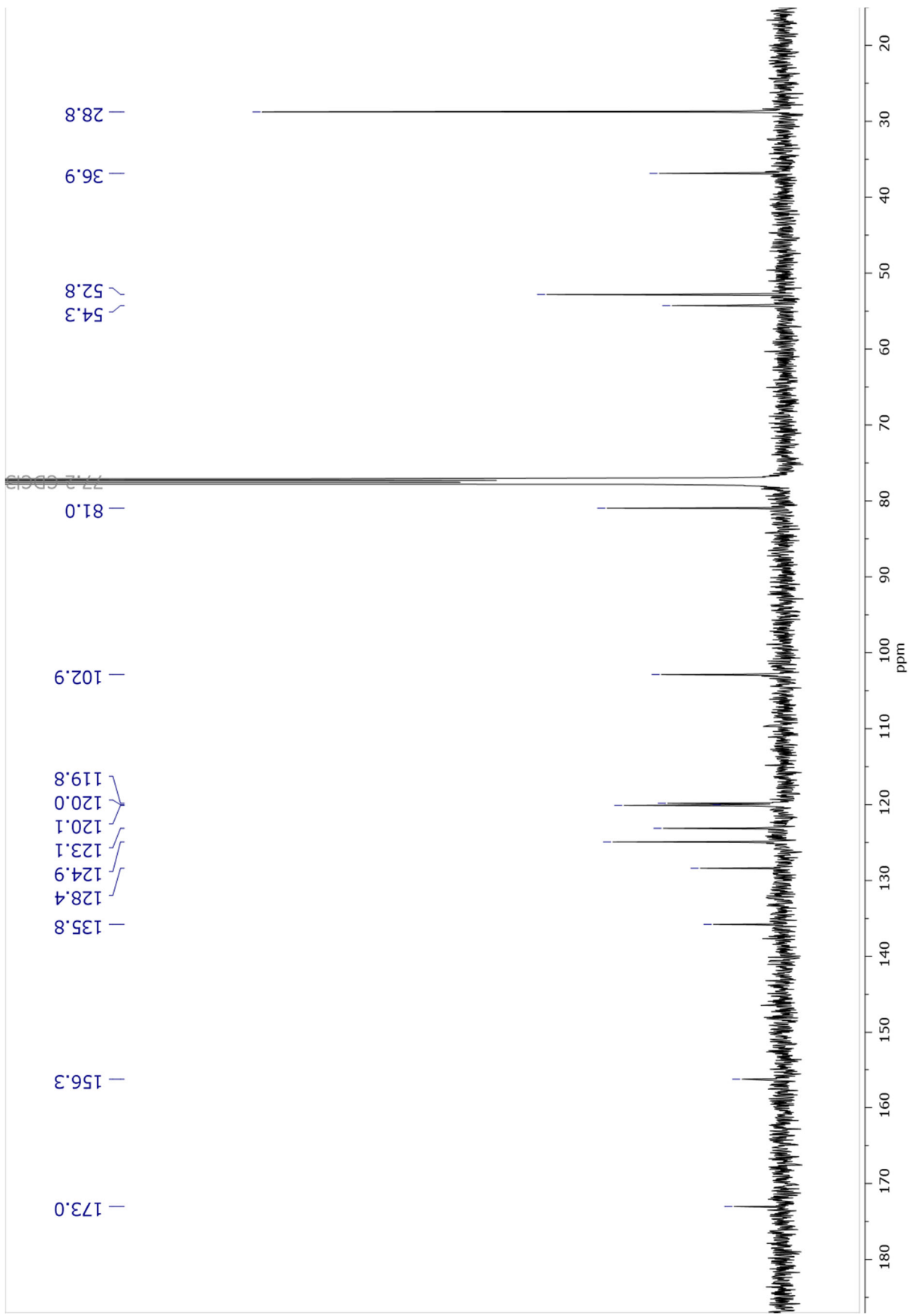
LC-MS: *t*_R = 9.4 min;

¹H NMR (500 MHz, CDCl₃) δ [ppm] = 9.80 (s, 1H, Indole-NH), 7.56 (d, ³*J* = 7.9 Hz, 1H, C4-H), 7.27 (dd, ³*J* = 3.8 Hz, ³*J* = 2.9 Hz, 1H, C2-H), 7.02 (dd, ³*J* = 7.7 Hz, ³*J* = 7.7 Hz, 1H, C5-H), 6.88 (d, ³*J* = 7.2 Hz, 1H, C6-H), 6.55 (dd, ⁴*J* = 2.8, ³*J* = 2.8 Hz, 1H, C3-H), 5.48 (d, ³*J* = 6.4 Hz, 1H, OCONH), 4.58 (ddd, ³*J* = 9.0 Hz, ³*J* = 6.8 Hz, ³*J* = 6.7 Hz, 1H, C_α-H), 3.66 (s, 3H, CH₃), 3.49 (dd, ³*J* = 13.0 Hz, ³*J* = 6.5 Hz, 1H, C_β-H), 3.34 (dd, ³*J* = 14.9 Hz, ³*J* = 9.0 Hz, 1H, C_β-H), 1.41 (s, 9H, C(CH₃)₃).

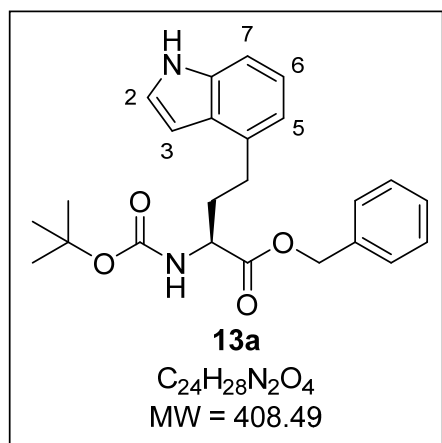
¹³C NMR (126 MHz, CDCl₃) δ [ppm] = 173.0 (COOMe), 156.3 (NHCOO), 135.8 (C7a), 128.4 (C4a), 124.9 (C7), 123.2 (C2), 120.2 (C4), 120.1 (C5), 120.0 (C6), 102.9 (C3), 80.9 (C(CH₃)₃), 54.3 (C_α), 52.8 (COO(CH₃)), 36.9 (C_β), 28.8 (C(CH₃)₃).

MS (ESI): found [*m/z*] = 319.08 [M+H]⁺, 263.02 [M-(*tert*Butyl)+H]⁺, 219.04 [M-Boc+H]⁺, 202.02 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 319.17 [M+H]⁺, 263.10 [M-(*tert*Butyl)+H]⁺, 219.11 [M-Boc+H]⁺, 202.09 [M-Boc-NH₂]⁺.





N^α-Boc-L-4-(1*H*-4-Indolyl)homoalanine benzyl ester (**13a**)



N^α-Boc-L-4-(1*H*-4-Indolyl)homoalanine benzyl ester (**13a**) was synthesized according to GP2. Therefore, 4-Bromoindole (**1a**, 32 μL, 255 μmol), zinc dust (65.9 mg, 1008 μmol, 4.0eq.), *N*^α-Boc-L-4-iodo homoalanine benzyl ester (**12**, 223.4 mg, 523 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (8.9 mg, 13 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL) The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL) the organic phase was dried over MgSO₄ and the solvent removed in vacuum.

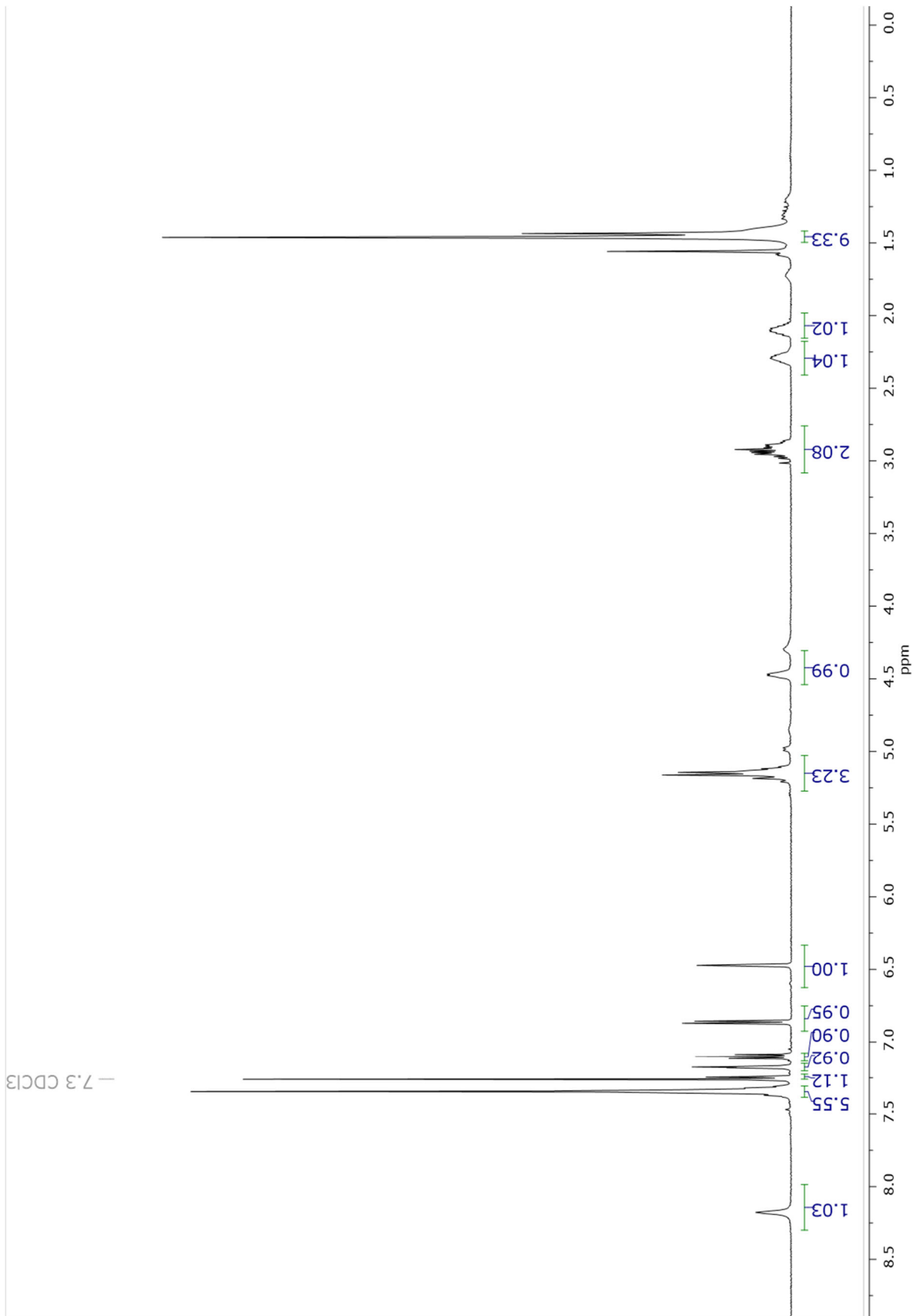
The crude product was purified by automated column chromatography using a Büchi Reveleris X2 (Petrolether:EtOAc) giving **13a** as a colourless solid (72.7 mg, 177 μmol, 70 %).

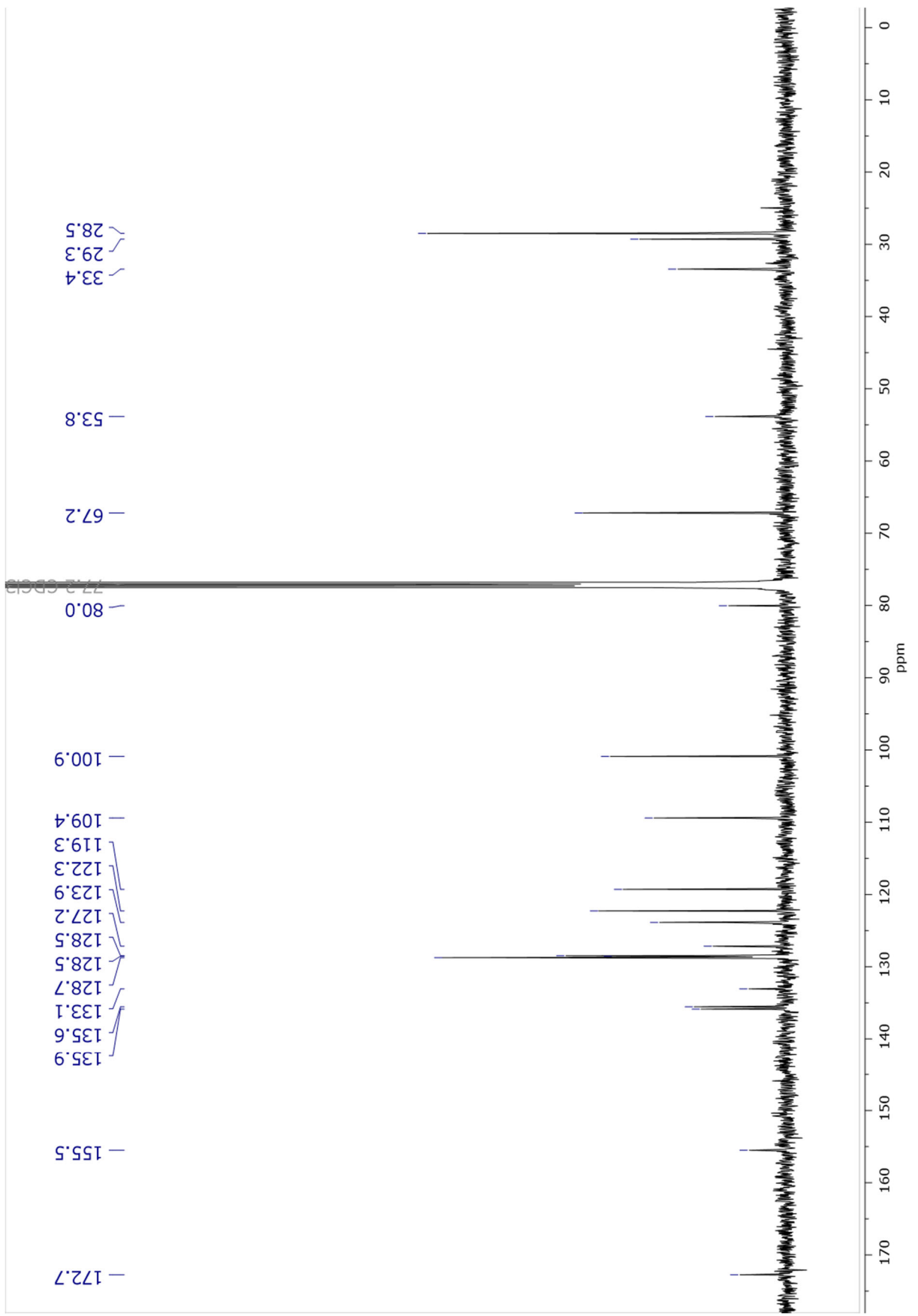
LC-MS: *t*_R = 10.4 min;

¹H NMR (500 MHz, CDCl₃) δ [ppm] = 8.18 (s, 1H, Indole-NH), 7.39 – 7.30 (m, 5H, C_{Phenyl}-H), 7.25 (d, ³*J* = 7.9 Hz, 1H, C7-H), 7.18 (dd, ³*J* = 2.9 Hz, ³*J* = 2.9 Hz, 1H, C2-H), 7.10 (dd, ³*J* = 7.6 Hz, ³*J* = 7.6 Hz, 1H, C6-H), 6.86 (d, ³*J* = 7.2 Hz, 1H, C5-H), 6.47 (dd, ⁴*J* = 3.0 Hz, ³*J* = 3.0 Hz, 1H, C3-H), 5.25 – 5.07 (m, 3H, OCONH/CH₂,Benzyl), 4.48 (ddd, ³*J* = 8.7 Hz ³*J* = 6.6 Hz, ³*J* = 5.5 Hz, 1H, C_α-H), 2.99 - 2.82 (m, 2H, C_μ-H₂), 2.30 (dd, ³*J* = 12.7 Hz, ³*J* = 5.5 Hz, 1H, C_β-H), 2.09 (dd, ³*J* = 13.0 Hz, ³*J* = 6.6 Hz, 1H, C_β-H), 1.45 (s, 9H, C(CH₃)₃).

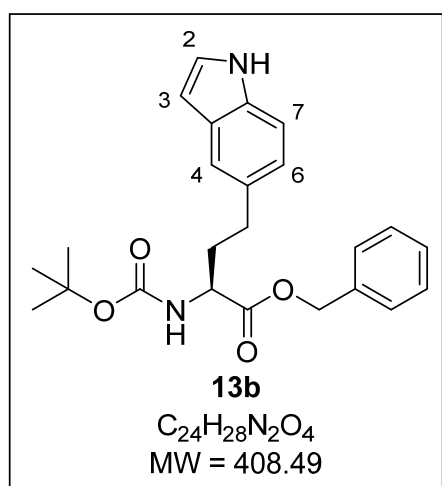
¹³C NMR (126 MHz, CDCl₃) δ [ppm] = 172.7 (COOBn), 155.5 (NHCOO), 135.8 (C7a), 135.5 (C_{Benzyl}), 133.1 (C4a), 128.8 (C_{Benzyl}), 128.6 (C_{Benzyl}), 126.6 (C_{Benzyl}), 127.2 (C4), 123.9 (C2), 122.3 (C5), 119.3 (C6), 109.4 (C7), 100.9 (C3), 80.0 (C(CH₃)₃), 67.2 ((CH₂)_{Benzyl}), 53.8 (C_α), 33.4 (C_β), 29.3 (C_μ), 28.5 (C(CH₃)₃).

MS (ESI): found [*m/z*] = 409.20 [M+H]⁺, 353.14 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.13 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 409.21 [M+H]⁺, 353.15 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.13 [M-Boc-NH₂]⁺.





N^α-Boc-L-4-(1*H*-5-Indolyl)homoalanine benzyl ester (**13b**)



N^α-Boc-L-4-(1*H*-5-Indolyl)homoalanine benzyl ester (**13b**) was synthesized according to *GP2*. Therefore, 5-Bromoindole (**1b**, 49.0 mg, 250 μmol), zinc dust (68.9 mg, 1.053 mmol, 4.0 eq.), *N*^α-Boc-L-μ-iodo homoalanine benzyl ester (**12**, 213.1 mg, 518 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (9.0 mg, 13 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL). The organic phase was dried over MgSO₄ and the solvent removed in vacuum. The crude product was purified by automated column chromatography using a Büchi

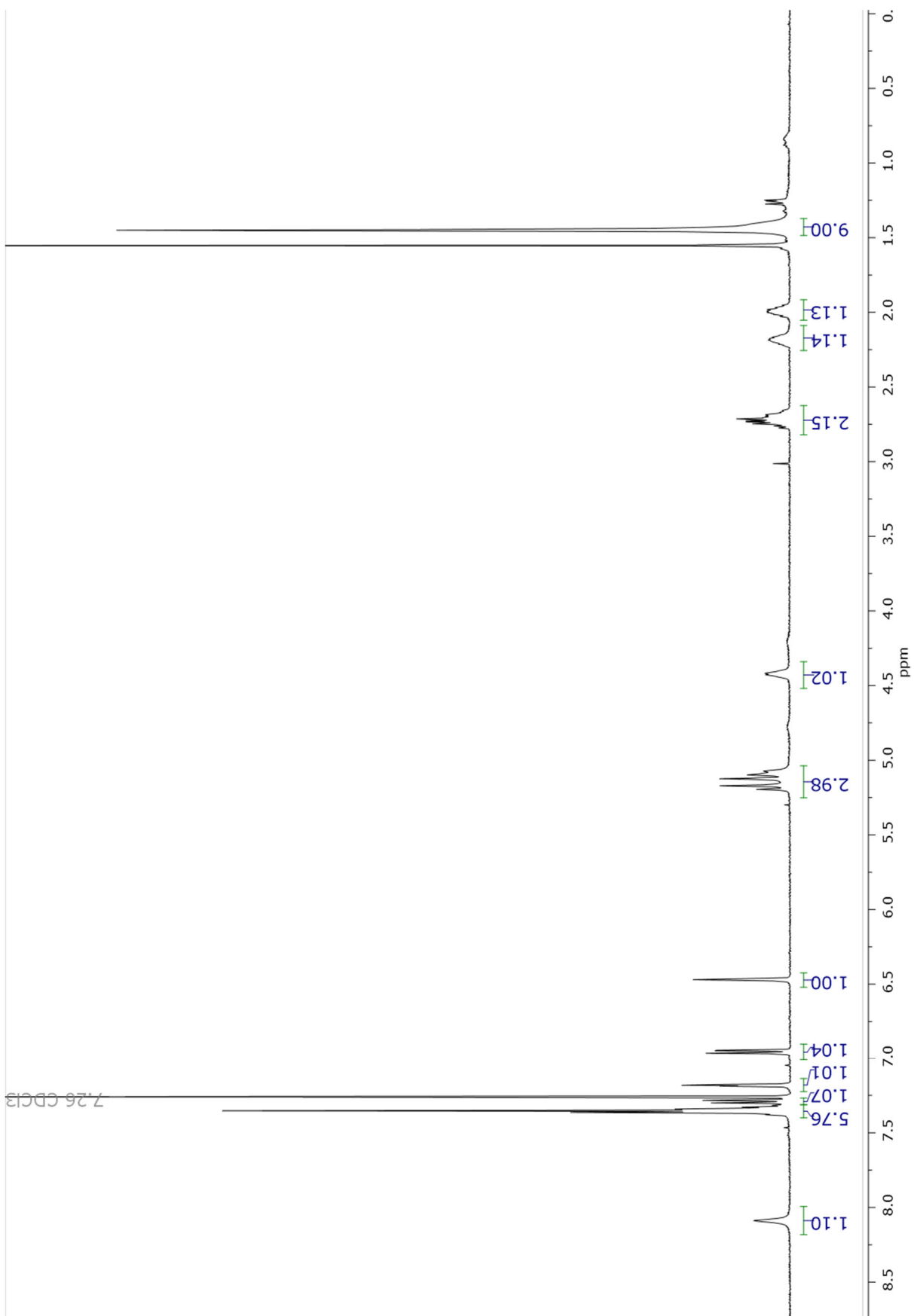
Reveleris X2 (Petrolether:EtOAc) giving *N*^α-Boc-L-4-(1*H*-5-indolyl)homoalanine benzyl ester (**13b**) as a colourless solid (67.4 mg, 165 μmol, 66 %).

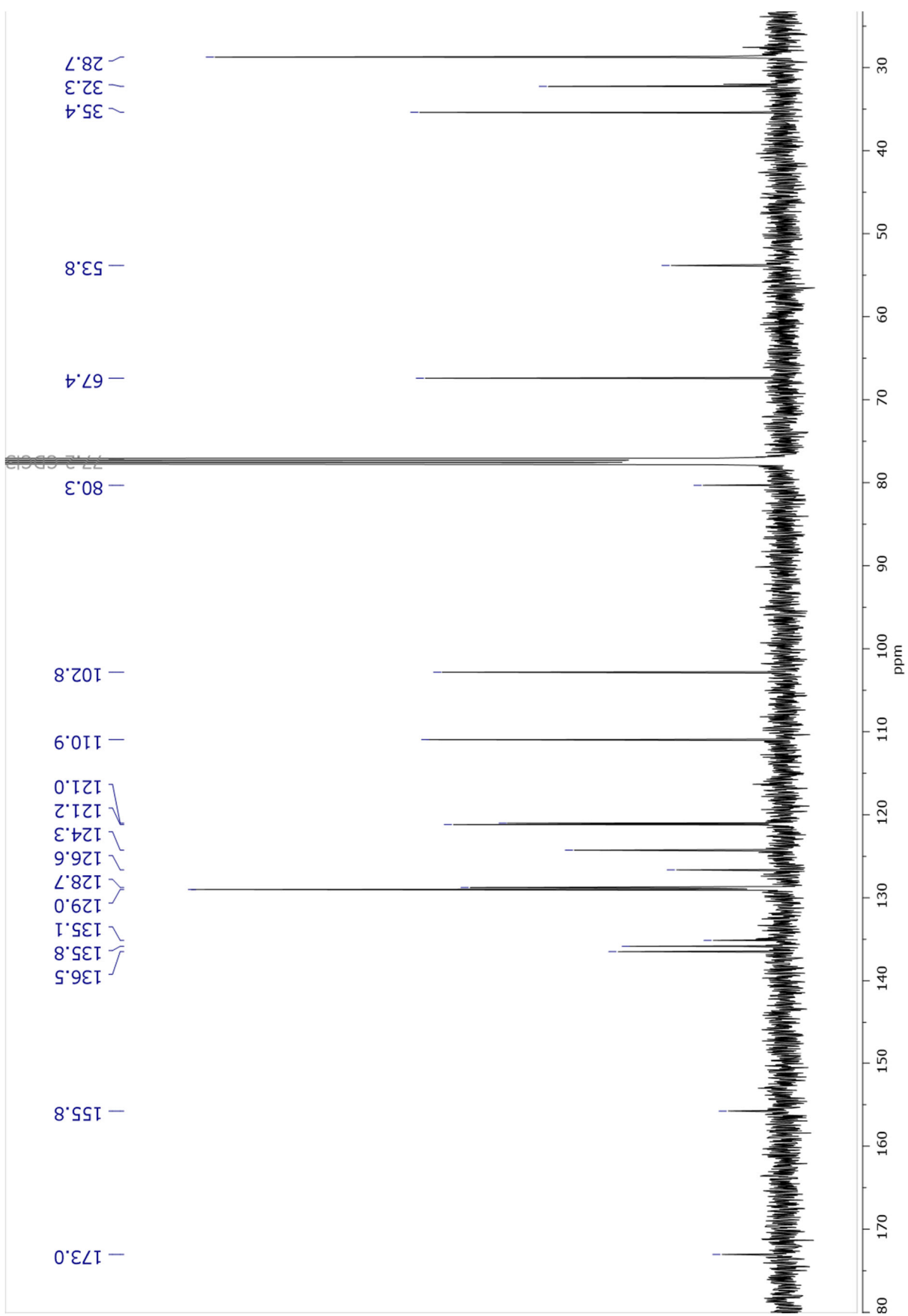
LC-MS: *t*_R = 10.3 min;

¹H NMR (500 MHz, CDCl₃) δ [ppm] = 8.09 (s, 1H, Indole-NH), 7.41-7.32 (m, 6H, C_{Phenyl}-H/C₄-H), 7.29 (d, ³*J* = 8.2 Hz, 1H, C₇-H), 7.18 (dd, ³*J* = 2.9 Hz, ³*J* = 2.9 Hz, 1H, C₂-H), 6.95 (dd, ³*J* = 8.3 Hz, ⁴*J* = 1.8 Hz, 1H, C₆-H), 6.47 (d, ³*J* = 3.1 Hz, 1H, C₃-H), 5.18 (d, ²*J* = 12.5, 1H, C_{Benzyl}-H), 5.11 (d, ²*J* = 13.0, 1H, C_{Benzyl}-H), 5.07 (d, ³*J* = 6.9 Hz, 1H, OCONH), 4.42 (ddd, ³*J* = 8.4 Hz, ³*J* = 7.9 Hz, ³*J* = 4.7 Hz, 1H, C_α-H), 2.80-2.61 (m, 2H, C_μ-H), 2.18 (m, 1H, C_β-H), 2.00 (m, 1H, C_β-H), 1.45 (s, 9H, C(CH₃)₃).

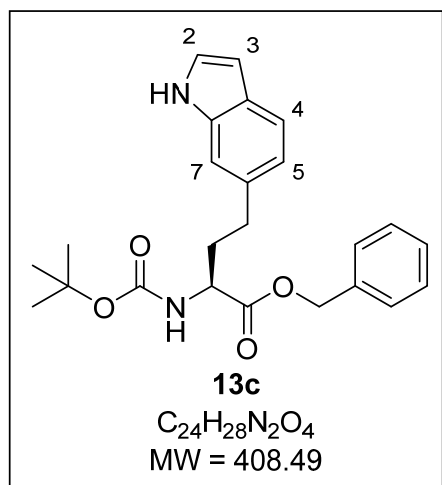
¹³C NMR (126 MHz, CDCl₃) δ [ppm] = 173.0 (COOBn), 155.8 (NHCOO), 136.5 (C_{7a}), 135.8 (C_{Benzyl}), 135.1 (C_{4a}), 129.0 (C_{Benzyl}), 128.7 (C_{Benzyl}), 128.6 (C₅), 126.6 (C_{Benzyl}), 124.3 (C₂), 121.2 (C₆), 121.0 (C₄), 111.0 (C₇), 102.8 (C₃), 80.3 (C(CH₃)₃), 67.4 ((CH₂)_{Benzyl}), 53.8 (C_α), 35.4 (C_β), 32.3 (C_μ), 28.7 (C(CH₃)₃).

MS (ESI): found [*m/z*] = 409.20 [M+H]⁺, 353.14 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.13 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 409.21 [M+H]⁺, 353.15 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.13 [M-Boc-NH₂]⁺.





N^α-Boc-L-4-(1*H*-6-Indolyl)homoalanine benzyl ester (**13c**)



N^α-Boc-L-4-(1*H*-6-Indolyl)homoalanine benzyl ester (**13c**) was synthesized according to *GP2*. Therefore, 6-Bromoindole (**1c**, 50.1, 255 μmol), zinc dust (66.54 mg, 1.018 mmol, 4.0 eq.), *N*^α-Boc-L- μ-iodo homoalanine benzyl ester (**12**, 213.5 mg, 509 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (8.9 mg, 13 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL) the organic phase was dried over MgSO₄ and the solvent removed in vacuum. The crude product was purified by automated column chromatography using a Büchi

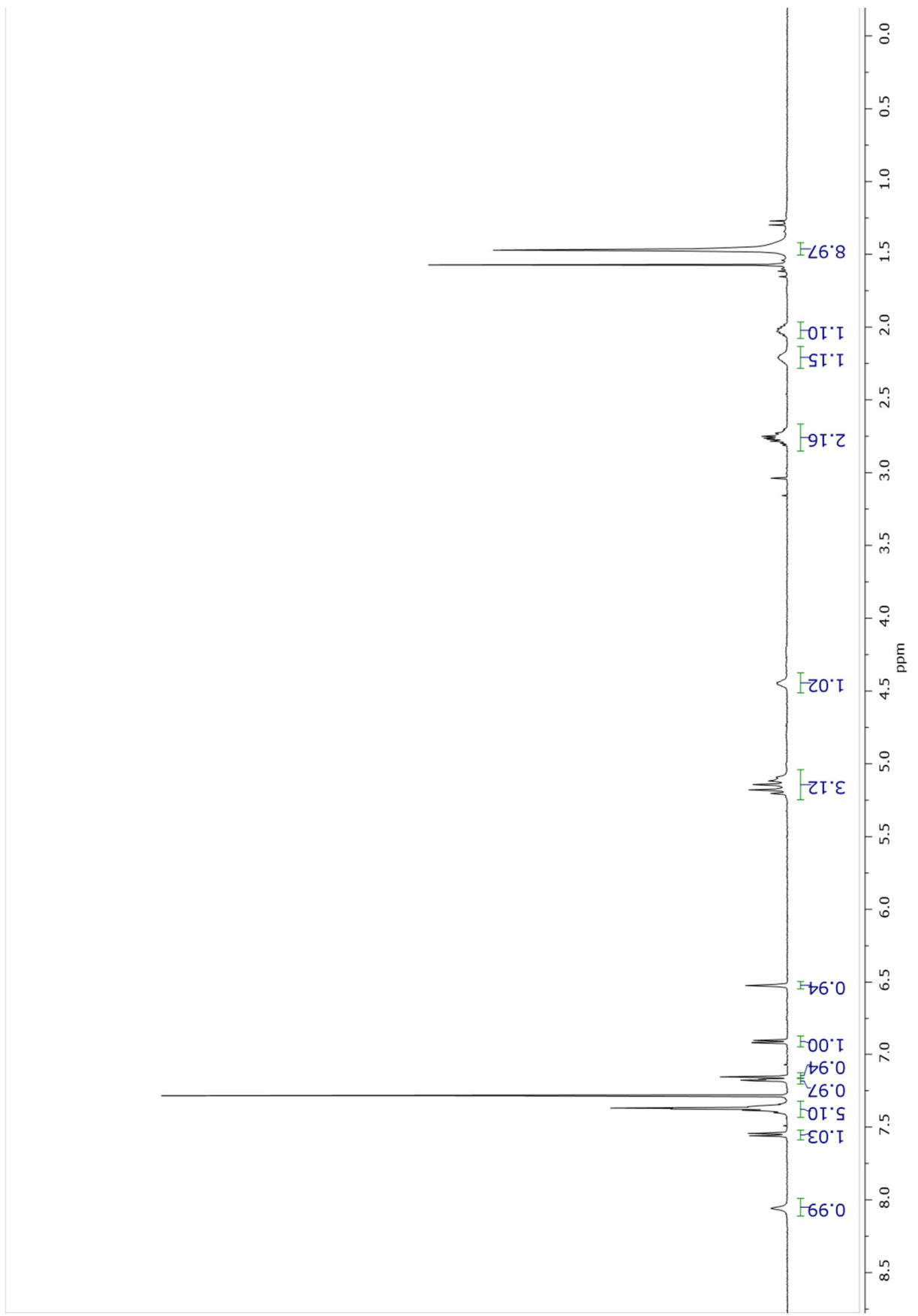
Reveleris X2 (Petroleum:EtOAc) giving **13c** as a colourless solid (60.5 mg, 148 μmol, 58 %).

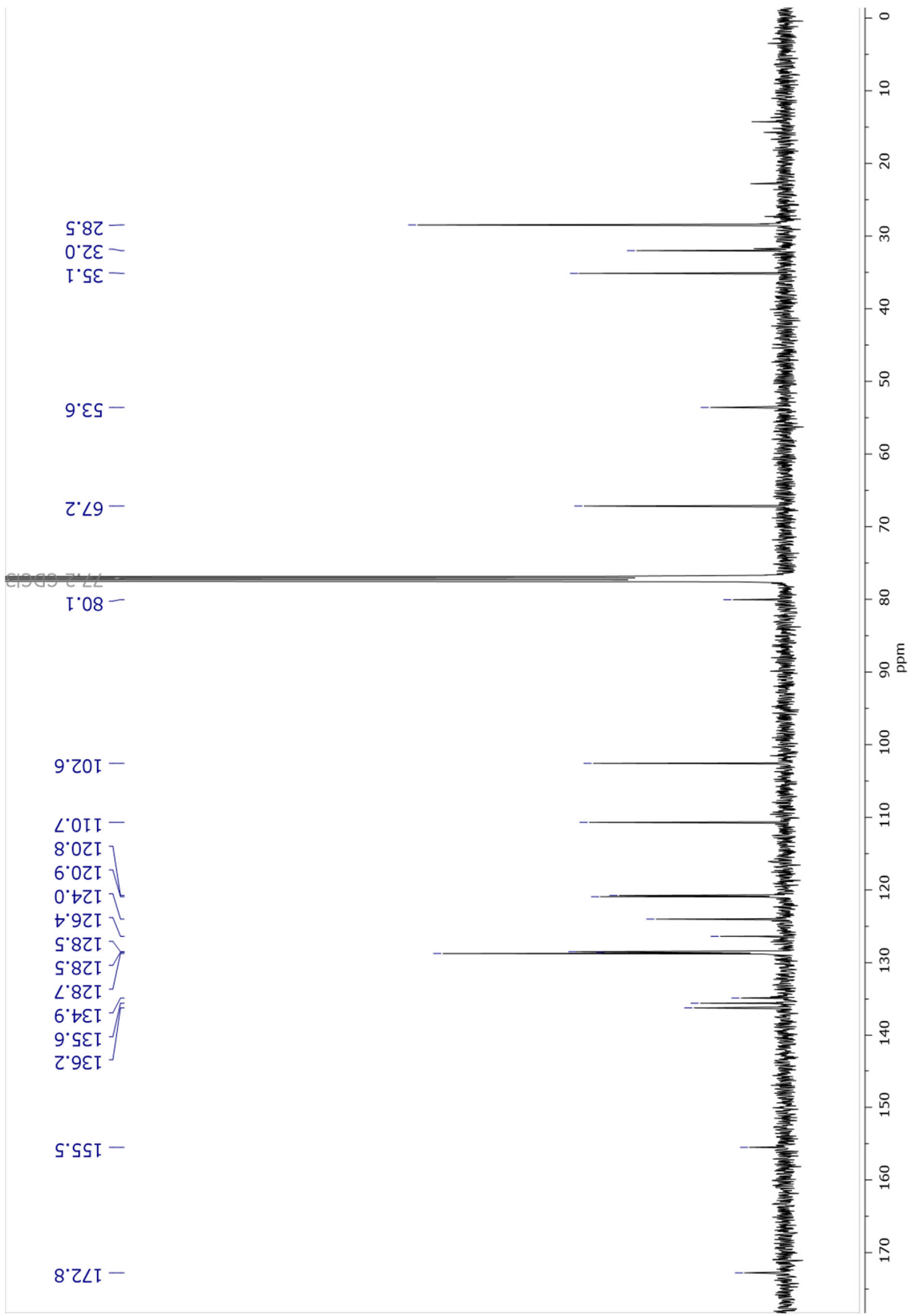
LC-MS: *t*_R = 10.5 min;

¹H NMR (500 MHz, CDCl₃) δ [ppm] = 8.06 (s, 1H, Indole-NH), 7.55 (d, ³*J* = 8.1 Hz, 1H, C4-H), 7.41-7.32 (m, 5H, C_{Phenyl}-H), 7.18 (dd, ³*J* = 3.0 Hz, ³*J* = 3.0 Hz, 1H, C2-H), 7.16 (s, 1H, C7-H), 6.91 (dd, ³*J* = 8.1 Hz, ⁴*J* = 1.6 Hz, 1H, C5-H), 6.52 (ddd, ³*J* = 4.2 Hz, ³*J* = 3.1 Hz, ³*J* = 2.9 Hz, 1H, C3-H), 5.19 (d, ²*J* = 12.4, 1H, C_{Benzyl}-H), 5.13 (d, ²*J* = 12.4, 1H, C_{Benzyl}-H), 5.09 (d, ³*J* = 8.3 Hz, 1H, OCONH), 4.44 (ddd, ³*J* = 8.4 Hz ³*J* = 7.9 Hz, ³*J* = 6.7 Hz, 1H, C_α-H), 2.84-2.64 (m, 2H, C_μ-H), 2.21 (m, 1H, C_β-H), 2.03 (m, 1H, C_β-H), 1.47 (s, 9H, C(CH₃)₃).

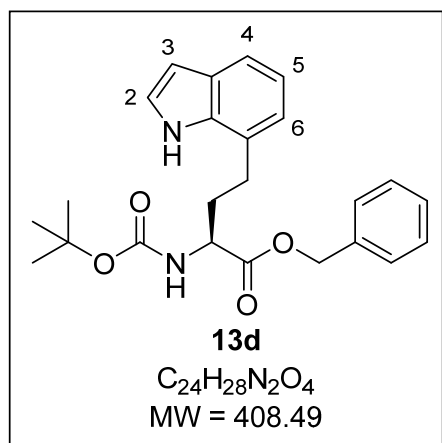
¹³C NMR (126 MHz, CDCl₃) δ [ppm] = 172.8 (COOBn), 155.5 (NHCOO), 136.2 (C7a), 135.6 (C_{Benzyl}), 134.9 (C4a), 128.8 (C_{Benzyl}), 128.6 (C_{Benzyl}), 128.5 (C_{Benzyl}), 126.4 (C6), 124.0 (C2), 120.9 (C5), 120.8 (C4), 110.7 (C7), 102.6 (C3), 80.1 (C(CH₃)₃), 67.2 ((CH₂)_{Benzyl}), 53.6 (C_α), 35.1 (C_β), 32.0 (C_μ), 28.5 (C(CH₃)₃).

MS (ESI): found [*m/z*] = 409.21 [M+H]⁺, 353.15 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.15 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 409.21 [M+H]⁺, 353.15 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.13 [M-Boc-NH₂]⁺.





N^α-Boc-L-4-(1*H*-7-Indolyl)homoalanine benzyl ester (**13d**)



N^α-Boc-L-4-(1*H*-7-Indolyl)homoalanine benzyl ester (**13d**) was synthesized according to *GP2*. Therefore, 7-Bromoindole (**1d**, 51.1 mg, 260 μmol), zinc dust (31.0 mg, 497 μmol, 1.9 eq.), *N*^α-Boc-L-4-iodo homoalanine benzyl ester (**12**, 200.4 mg, 595 μmol, 2.2 eq.) and Pd(amphos)₂Cl₂ (9.1 mg, 13 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.5 mL). The suspension was purged with argon and stirred at room temperature for 24 h. The crude mixture was diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL) the organic phase was dried over MgSO₄ and the solvent removed in vacuum.

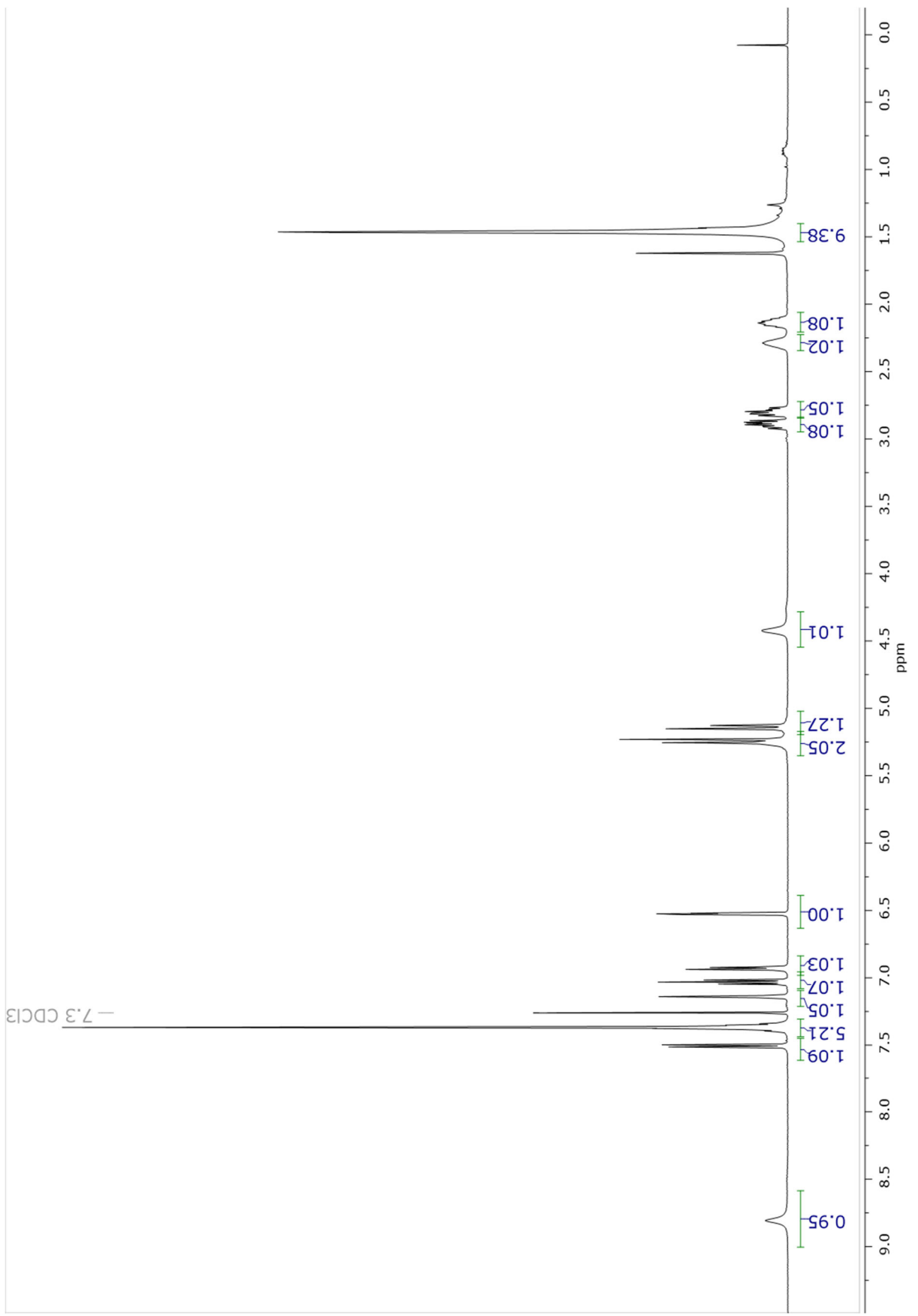
The crude product was purified by column chromatography using (Petrolether:EtOAc (5:1)) giving **13d** as a colourless solid (75.6 mg, 185 μmol, 71 %).

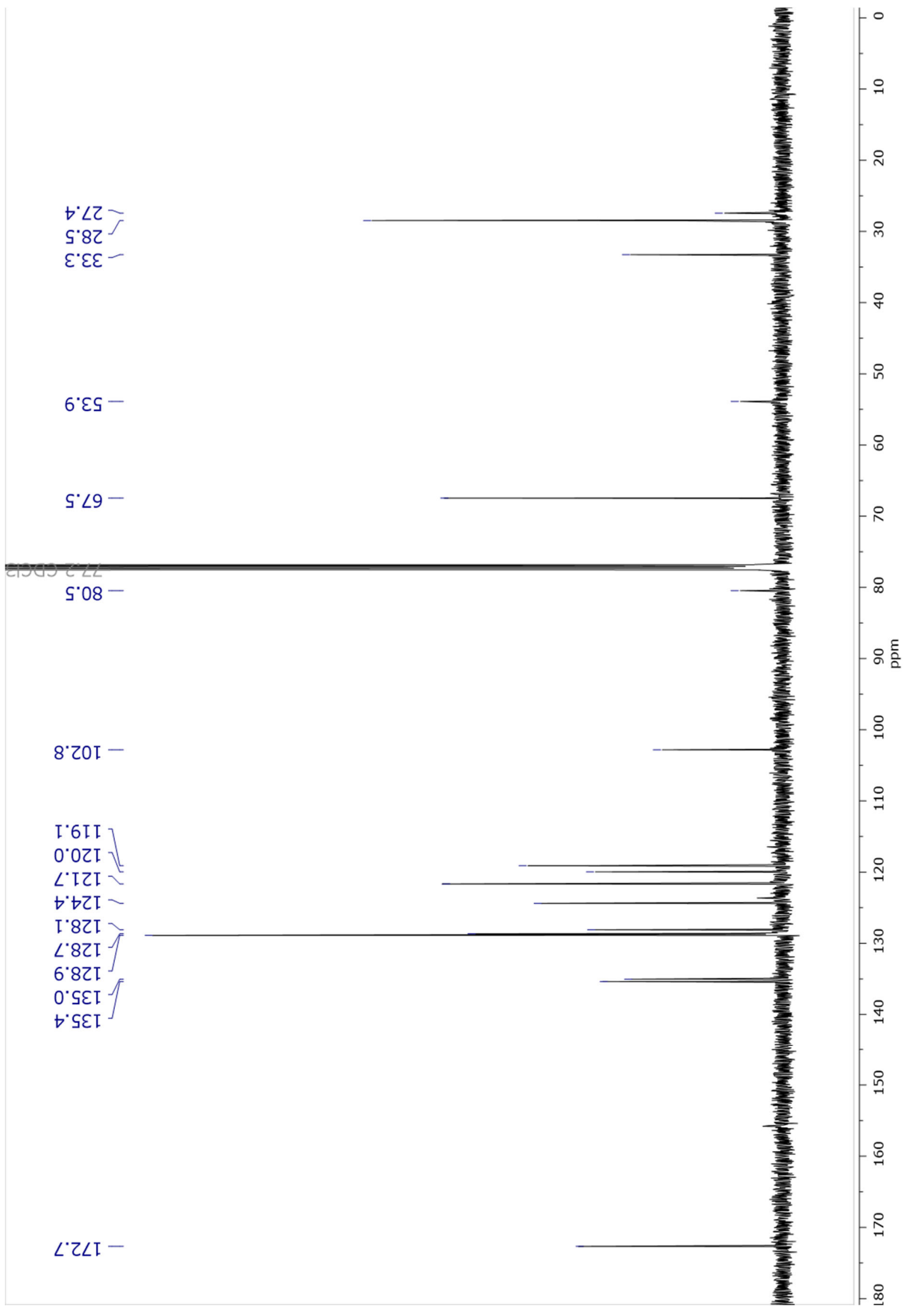
LC-MS: *t*_R = 10.9 min;

¹H NMR (500 MHz, CDCl₃) δ [ppm] = 8.81 (s, 1H, Indole-NH), 7.51 (d, ³*J* = 7.8 Hz, 1H, C4-H), 7.41 (m, 5H, C_{Phenyl}-H), 7.14 (dd, ³*J* = 2.7 Hz, ³*J* = 2.7 Hz, 1H, C2-H), 7.03 (dd, ³*J* = 7.6 Hz, ³*J* = 7.6 Hz, 1H, C5-H), 6.93 (d, ³*J* = 7.2 Hz, 1H, C6-H), 6.53 (dd, ⁴*J* = 2.0, ³*J* = 3.2 Hz, 1H, C3-H), 5.26 (d, ³*J* = 6.6 Hz, 1H, OCONH), 5.24 (d, ²*J* = 12.2 Hz, 1H, C_{Benzyl}-H), 5.14 (d, ²*J* = 12.2 Hz, 1H, C_{Benzyl}-H), 4.42 (m, 1H, C_α-H), 2.89 (ddd, ²*J* = 13.0 Hz, ³*J* = 9.6 Hz, ³*J* = 6.3 Hz, 1H, C_μ-H), 2.80 (ddd, ²*J* = 13.1 Hz, ³*J* = 9.6 Hz, ³*J* = 6.1 Hz, 1H, C_μ-H), 2.30 (m, 1H, C_β-H), 2.15 (m, 1H, C_β-H), 1.46 (s, 9H, C(CH₃)₃).

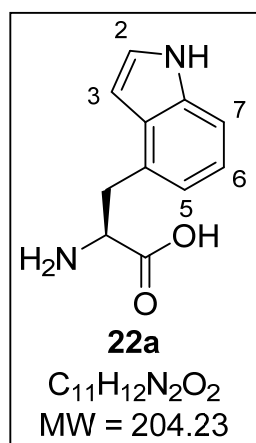
¹³C NMR (126 MHz, CDCl₃) δ [ppm] = 172.7 (COOBn), 155.7 (NHCOO), 135.4 (C7a), 135.1 (C_{Benzyl}), 135.0 (C4a), 128.9 (C_{Benzyl}), 128.8 (C_{Benzyl}), 128.7 (C_{Benzyl}), 128.1 (C7), 124.4 (C2), 121.7 (C6), 120.0 (C5), 119.1 (C4), 102.8 (C3), 80.5 (C(CH₃)₃), 67.5 ((CH₂)_{Benzyl}), 53.9 (C_α), 33.3 (C_β), 28.5 (C(CH₃)₃), 27.4 (C_μ).

MS (ESI): found [*m/z*] = 409.21 [M+H]⁺, 353.15 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.15 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 409.21 [M+H]⁺, 353.15 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.13 [M-Boc-NH₂]⁺.





L-3-(1*H*-4-Indolyl)alanine (**22a**)



N^α-Boc-L-3-(1*H*-4-Indolyl)alanine methyl ester (**11a**) (348.7 mg, 1.095 mmol) was suspended in Na₂HPO₄-Buffer (100 mM, pH = 7.4, 15 mL). The mixture was heated to reflux for 9 hours. The aqueous phase was washed with EtOAc (2x15 mL). The solvent of the aqueous phase was reduced in vacuum and the crude product was desalted using an acid-free reversed phase C18-column chromatography, washing with pure MPW (100 mL) and eluting the product with MeOH (100 mL). The solvent was removed in vacuum, giving L-3-(1*H*-4-indolyl)alanine (**22a**) as a slightly yellow solid (122.9 mg, 0.602 mmol, 55 %). The enantiomeric excess was determined using Marfey's Test (see GP3) giving an ee of 95 %.

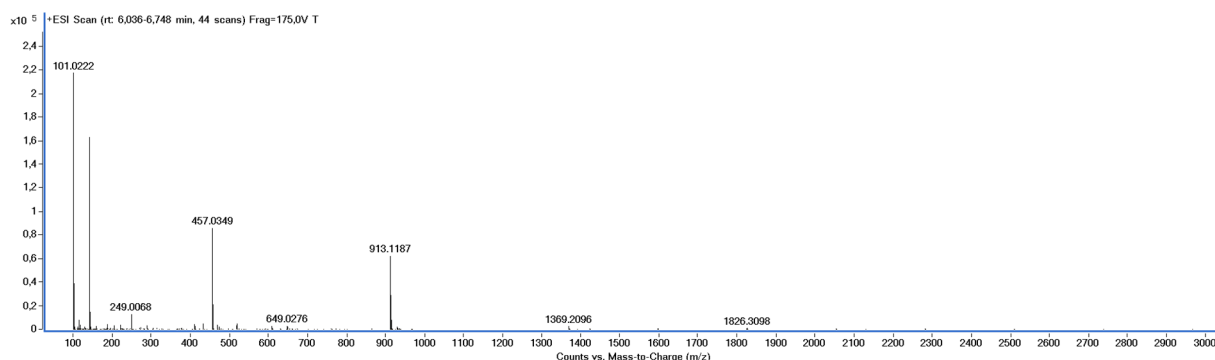
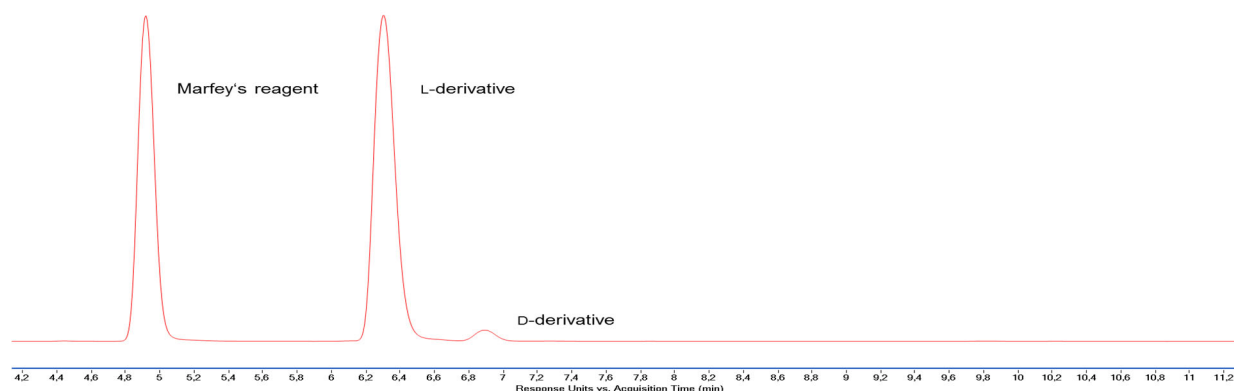
LC-MS: t_R = 2.8 min;

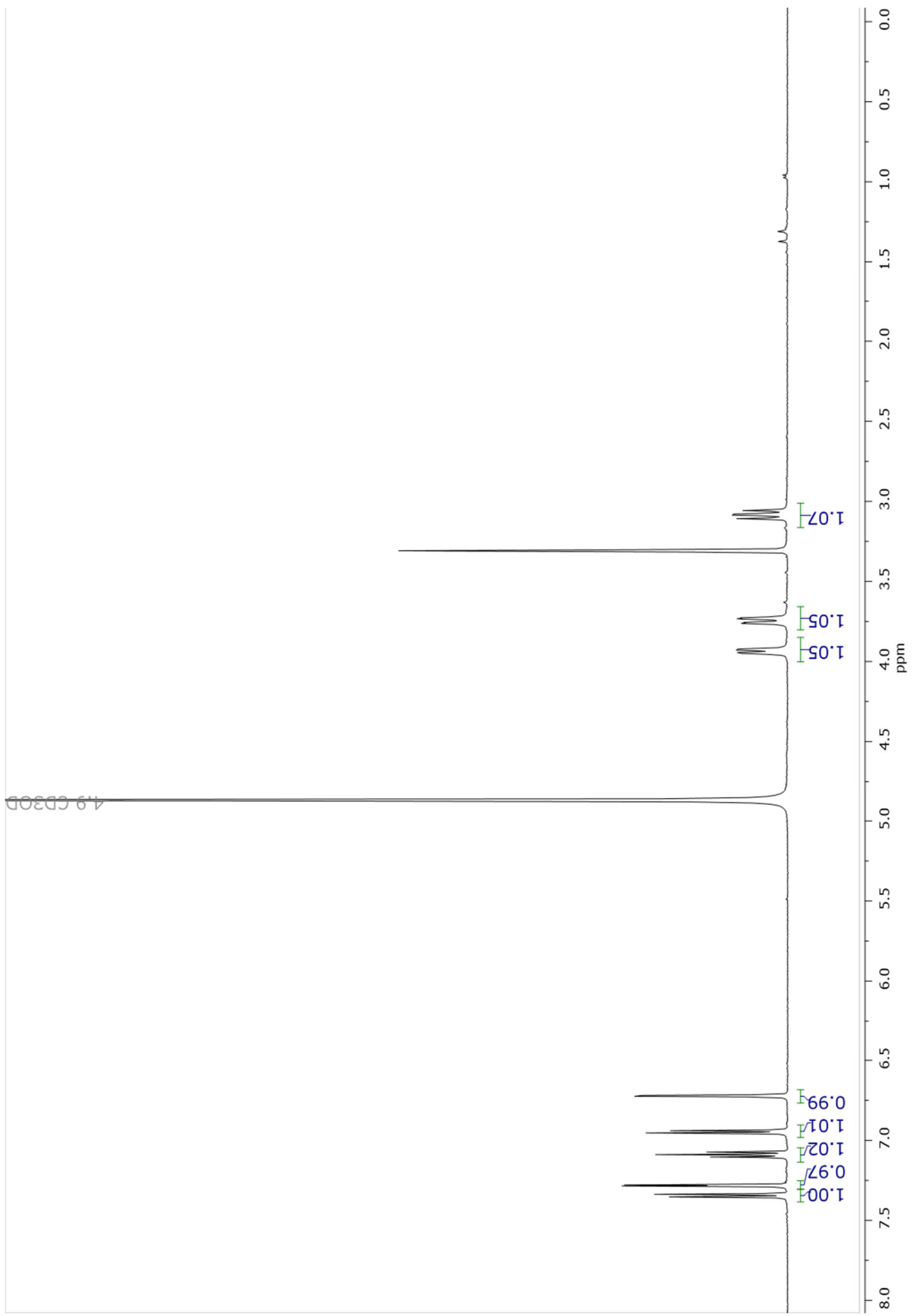
¹H NMR (500 MHz, MeOD) δ [ppm] = 7.35 (d, ³*J* = 8.2 Hz, 1H, C7-H), 7.28 (d, ³*J* = 3.1 Hz, 1H, C2-H), 7.09 (dd, ³*J* = 8.2 Hz, ³*J* = 7.1 Hz, 1H, C6-H), 6.95 (d, ³*J* = 7.2 Hz, 1H, C5-H), 6.72 (d, ³*J* = 3.1 Hz, 1H, C3-H), 3.94 (dd, ³*J* = 10.9 Hz ³*J* = 3.9 Hz, 1H, C α -H), 3.74 (dd, ³*J* = 14.3 Hz, ³*J* = 3.9 Hz, 1H, C β -H), 3.08 (dd, ³*J* = 14.4 Hz, ³*J* = 10.8 Hz, 1H, C β -H).

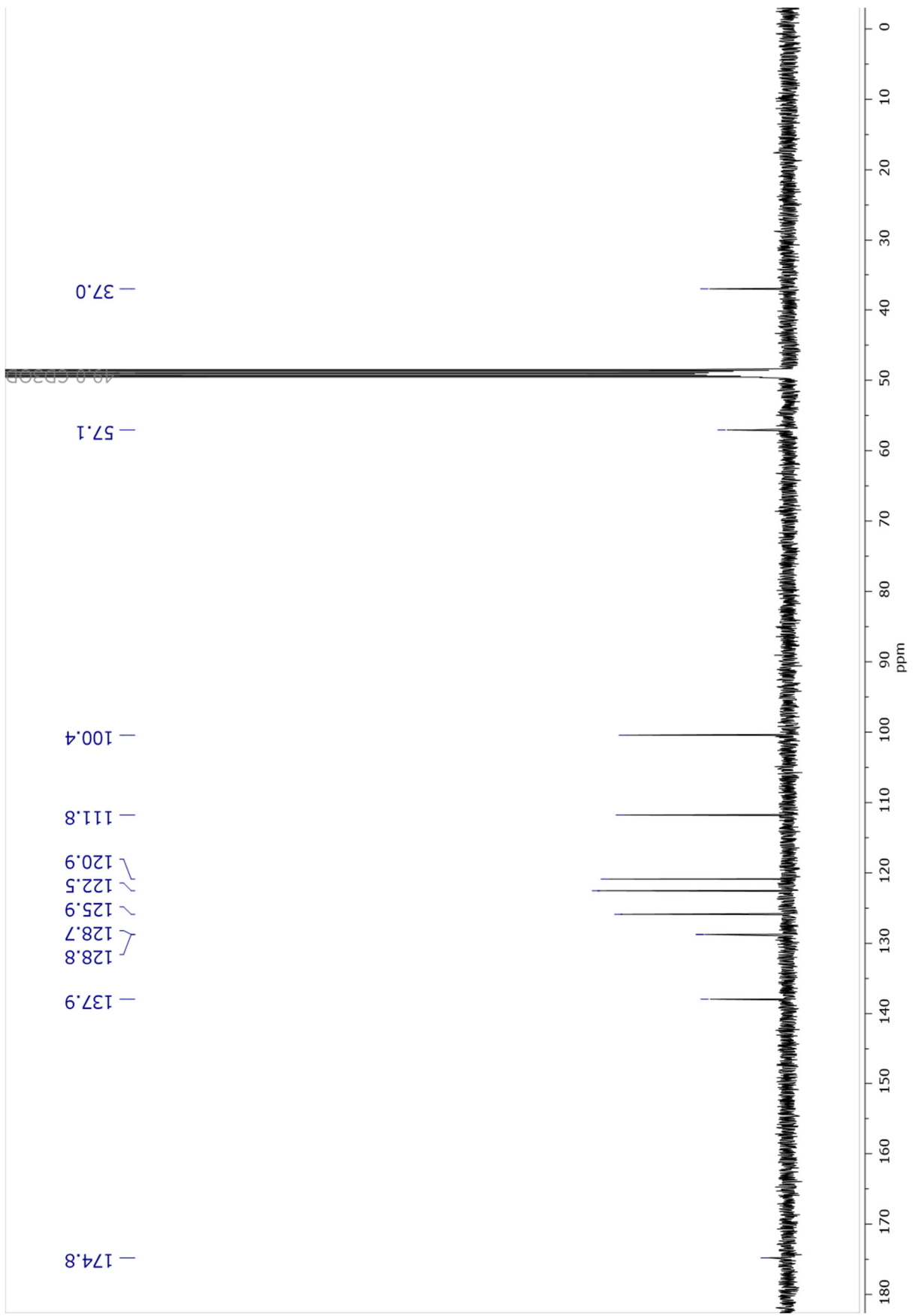
¹³C NMR (126 MHz, CDCl₃) δ [ppm] = 174.8 (COOH), 137.9 (C7a), 128.8 (C3a), 128.7 (C4), 125.8 (C2), 122.5 (C6), 120.9 (C5), 111.8 (C7), 100.4 (C3), 57.1 (C α), 37.0 (C β).

MS (ESI): found [*m/z*] = 205.03 [M+H]⁺, calcd. [*m/z*] = 205.10 [M+H]⁺.

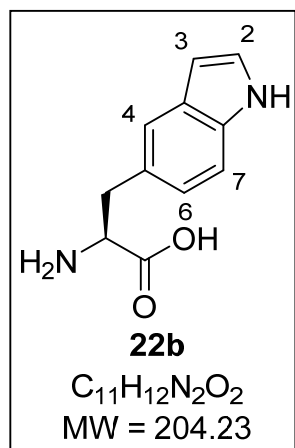
Marfey's Derivatization: t_R (L-deriv.) = 6.3 min (97.5 %); t_R (D-deriv.) = 6.9 min (2.5 %)
found [*m/z*] = 457.03 [M+H]⁺, calcd. [*m/z*] = 457.15 [M+H]⁺.







L-3-(1*H*-5-Indolyl)alanine (**22b**)



N^α -Boc-L-3-(1*H*-5-Indolyl)alanine methyl ester (**11b**) (300.0 mg, 0.942 mmol) was suspended in Na_2HPO_4 -Buffer (100 mM, pH = 7.4, 15 mL). The mixture was heated to reflux for 9 hours. The aqueous phase was washed with EtOAc (2x15 mL). The solvent of the aqueous phase was reduced in vacuum and the crude product was desalted using an acid-free reversed phase C18-column chromatography, washing with pure MPW (100 mL) and eluting the product with MeOH (100 mL). The solvent was removed in vacuum giving L-3-(1*H*-5-indolyl)alanine (**22b**) as a colourless solid (100.3 mg, 0.491 mmol, 52 %). The enantiomeric excess was determined using Marfey's Test (see GP3) giving an ee of 94 %.

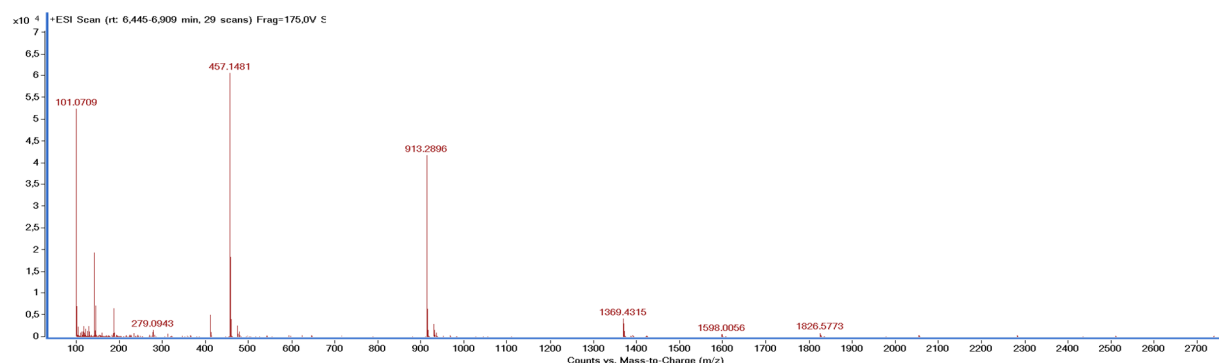
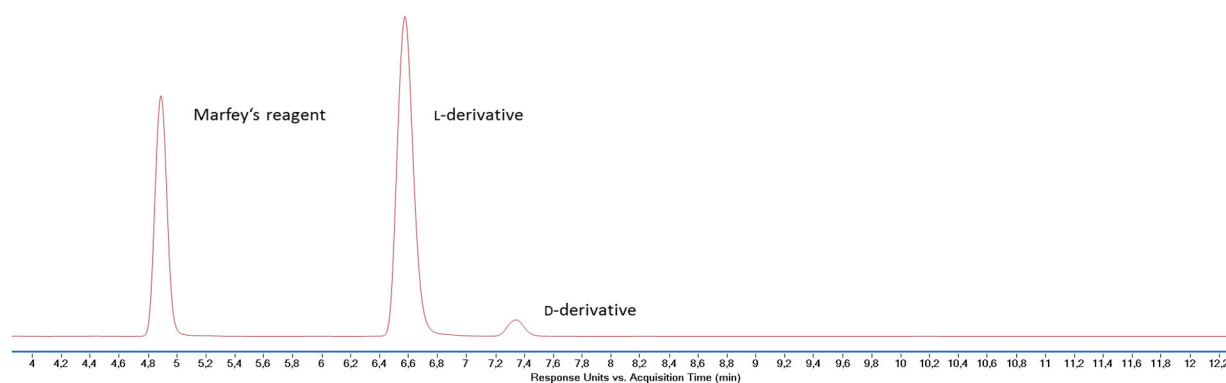
LC-MS: t_R = 2.7 min;

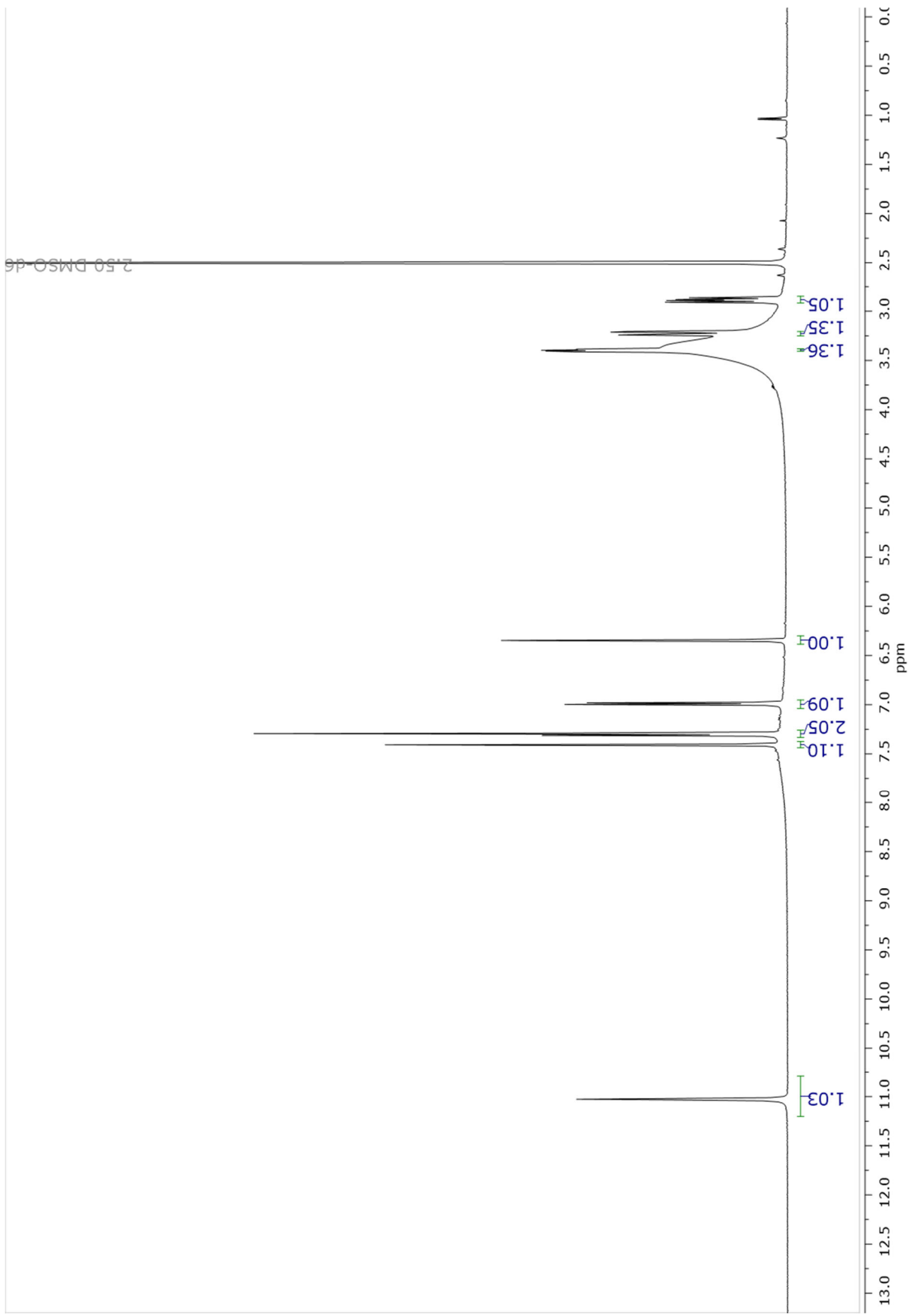
1H NMR (500 MHz, $DMSO-d_6$) δ [ppm] = 11.03 (s, 1H, Indole-NH), 7.41 (s, 1H, C4-H), 7.30 (d, 3J = 8.3 Hz, 1H, C7-H), 7.29 (s, 1H, C2-H), 6.99 (d, 3J = 8.4 Hz, 1H, C6-H), 6.35 (m, 1H, C3-H), 3.40 (dd, 3J = 8.7 Hz 3J = 4.3 Hz, 1H, C α -H), 3.22 (dd, 3J = 14.3 Hz, 3J = 4.3 Hz, 1H, C β -H), 2.89 (dd, 3J = 14.3 Hz, 3J = 8.7 Hz, 1H, C β -H).

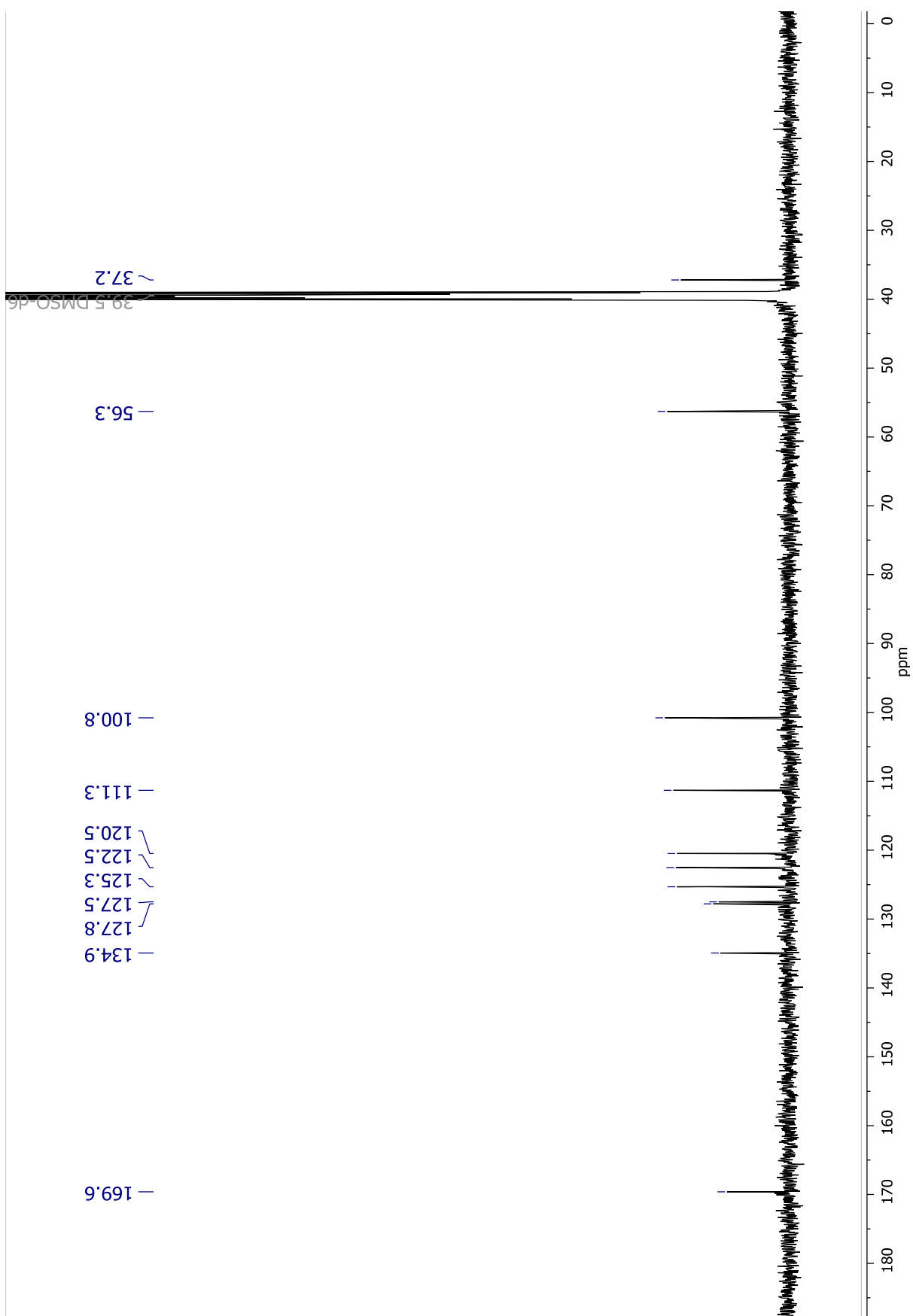
^{13}C NMR (126 MHz, $DMSO-d_6$) δ [ppm] = 169.9, 134.9 (C7a), 127.8 (C3a), 127.5 (C5), 125.3 (C2), 122.5 (C6), 120.5 (C4), 111.1 (C7), 100.8 (C3), 56.3 (C α), 37.2 (C β).

MS (ESI): found $[m/z]$ = 205.04 $[M+H]^+$, calcd. $[m/z]$ = 205.10 $[M+H]^+$.

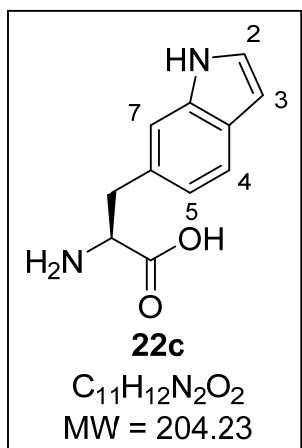
Marfey's Derivatization: t_R (L-deriv.) = 6.6 min (97 %); t_R (D-deriv.) = 7.3 min (3 %)
found $[m/z]$ = 457.15 $[M+H]^+$, calcd. $[m/z]$ = 457.15 $[M+H]^+$.







L-3-(1*H*-6-Indolyl)alanine (**22c**)



N^α-Boc-L-3-(1*H*-6-Indolyl)alanine methyl ester (**11c**) (279.2 mg, 0.877 mmol) was suspended in Na₂HPO₄-Buffer (100 mM, pH = 7.4, 15 mL). The mixture was heated to reflux for 9 hours. The aqueous phase was washed with EtOAc (2x15 mL). The solvent of the aqueous phase was reduced in vacuum and the crude product was desalted using an acid-free reversed phase C18-column chromatography, washing with pure MPW (100 mL) and eluting the product with MeOH (100 mL). The solvent was removed in vacuum giving L-3-(1*H*-6-indolyl)alanine (**22c**) as a colourless solid (176.6 mg, 0.864 mmol, 98 %). The enantiomeric excess was determined using Marfey's Test (see GP3) giving an ee of 94 %.

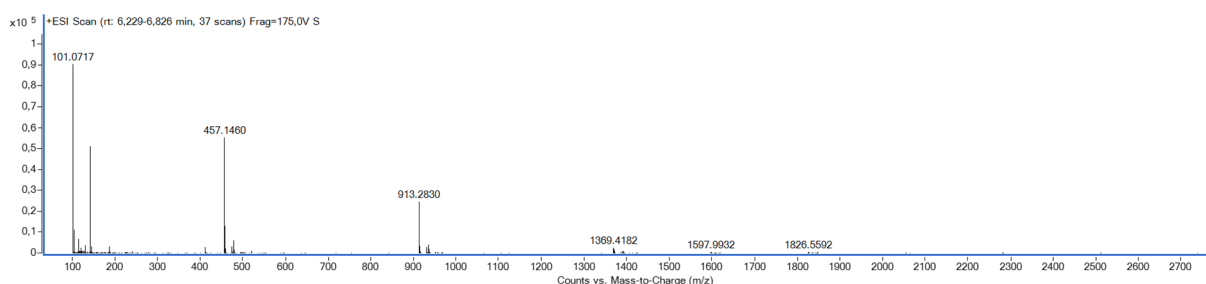
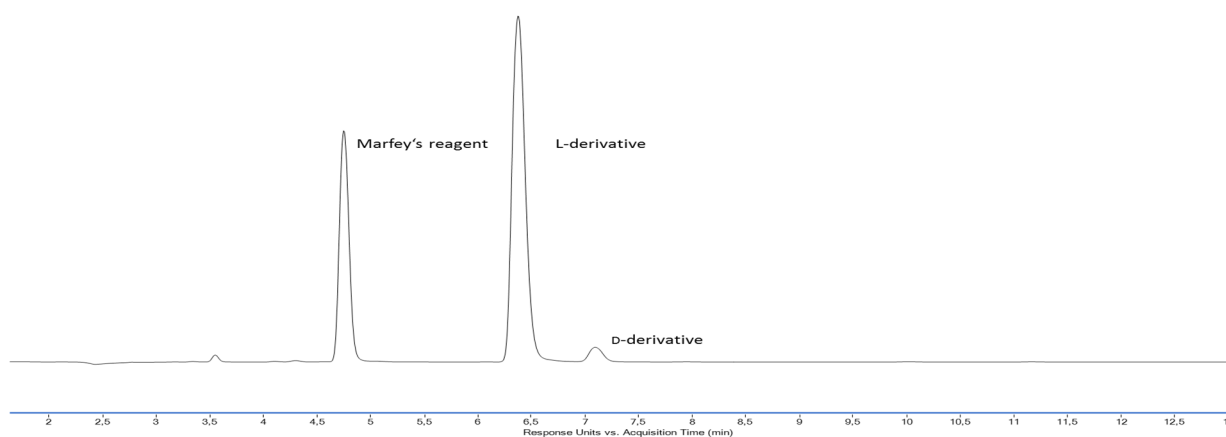
LC-MS: t_R = 3.6 min;

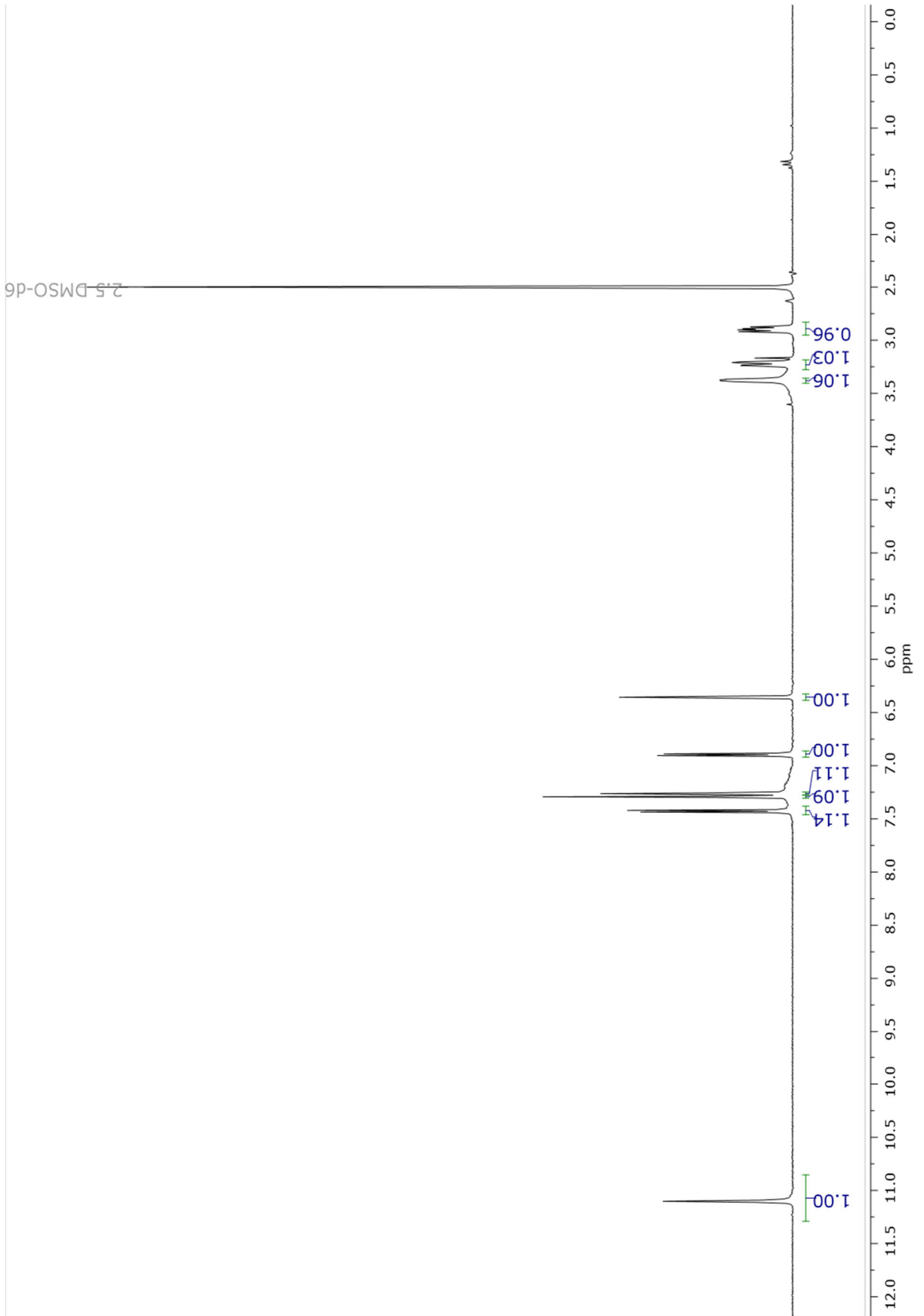
¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 11.10 (s, 1H, Indole-NH), 7.44 (d, ³*J* = 8.1 Hz 1H, C4-H), 7.29 (s, 1H, C7-H), 7.26 (dd, ³*J* = 2.8 Hz, ³*J* = 2.8 Hz, 1H, C2-H), 6.90 (d, ³*J* = 8.1 Hz, 1H, C5-H), 6.36 (dd, ³*J* = 2.8 Hz, ⁴*J* = 2.1 Hz 1H, C3-H), 3.38 (dd, ³*J* = 8.5 Hz ³*J* = 4.0 Hz, 1H, C α -H), 3.22 (dd, ³*J* = 14.2 Hz, ³*J* = 4.0 Hz, 1H, C β -H), 2.89 (dd, ³*J* = 14.2 Hz, ³*J* = 8.5 Hz, 1H, C β -H).

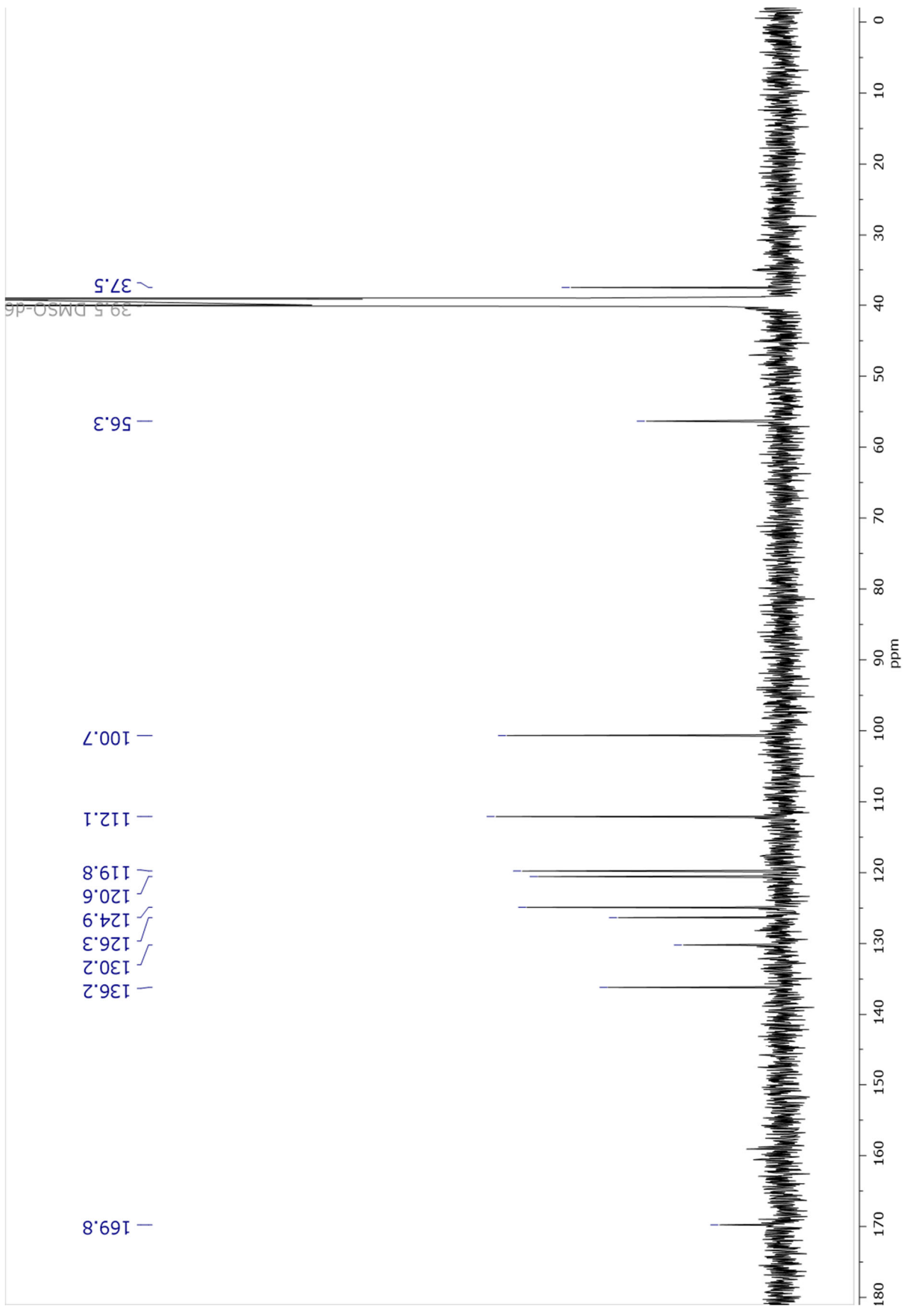
¹³C NMR (126 MHz, DMSO-*d*₆) δ = 169.8 (COOH), 136.2 (C7a), 130.2 (C3a), 126.3 (C6), 124.9 (C2), 120.6 (C4), 119.8 (C5), 112.1 (C7), 100.7 (C3), 56.3 (C α), 37.5 (C β)

MS (ESI): found [*m/z*] = 205.03 [M+H]⁺, calcd. [*m/z*] = 205.10 [M+H]⁺.

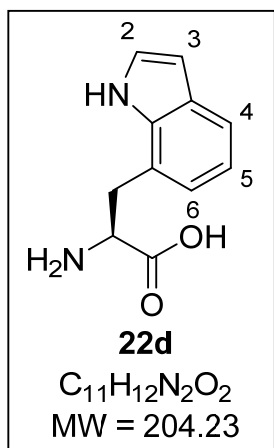
Marfey's Derivatization: t_R (L-deriv.) = 6.4 min (97 %); t_R (D-deriv.) = 7.2 min (3 %)
found [*m/z*] = 457.15 [M+H]⁺, calcd. [*m/z*] = 457.15 [M+H]⁺.







L-3-(1*H*-7-Indolyl)alanine (**22d**)



N^α-Boc-L-3-(1*H*-7-Indolyl)alanine methyl ester (**11d**) (253.4 mg, 0.796 mmol) was suspended in Na₂HPO₄-Buffer (100 mM, pH = 7.4, 15 mL). The mixture was heated to reflux for 9 hours. The aqueous phase was washed with EtOAc (2x15 mL). The solvent of the aqueous phase was reduced in vacuum and the crude product was desalted using an acid-free reversed phase C18-column chromatography, washing with pure MPW (100 mL) and eluting the product with MeOH (100 mL). The solvent was removed in vacuum giving L-3-(1*H*-7-indolyl)alanine (**22d**) as a colourless solid (160.5 mg, 0.786 mmol, 99 %). The enantiomeric excess was determined using Marfey's Test (see GP3) giving an ee of 38 %.

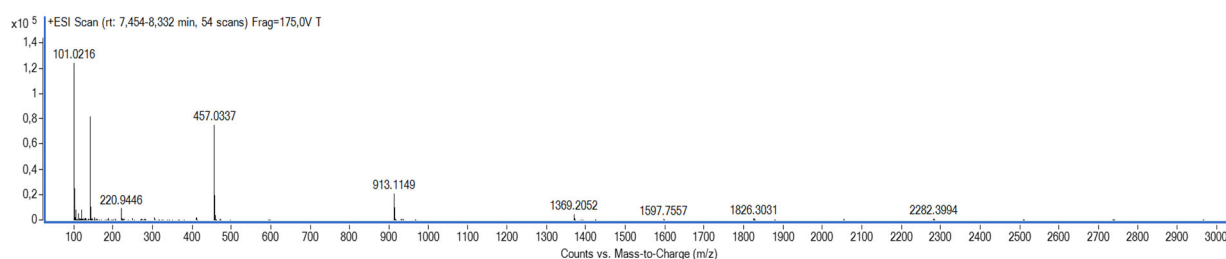
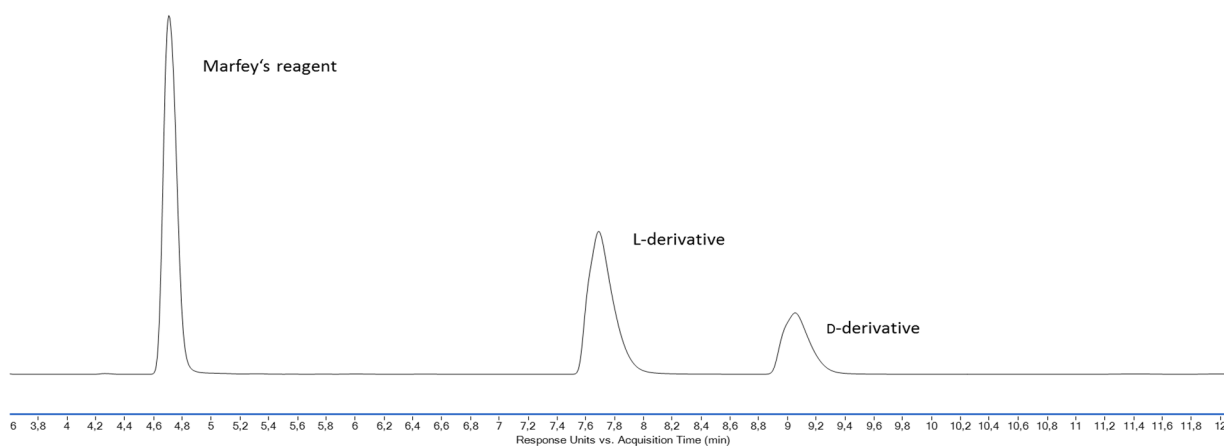
LC-MS: t_R = 4.1 min;

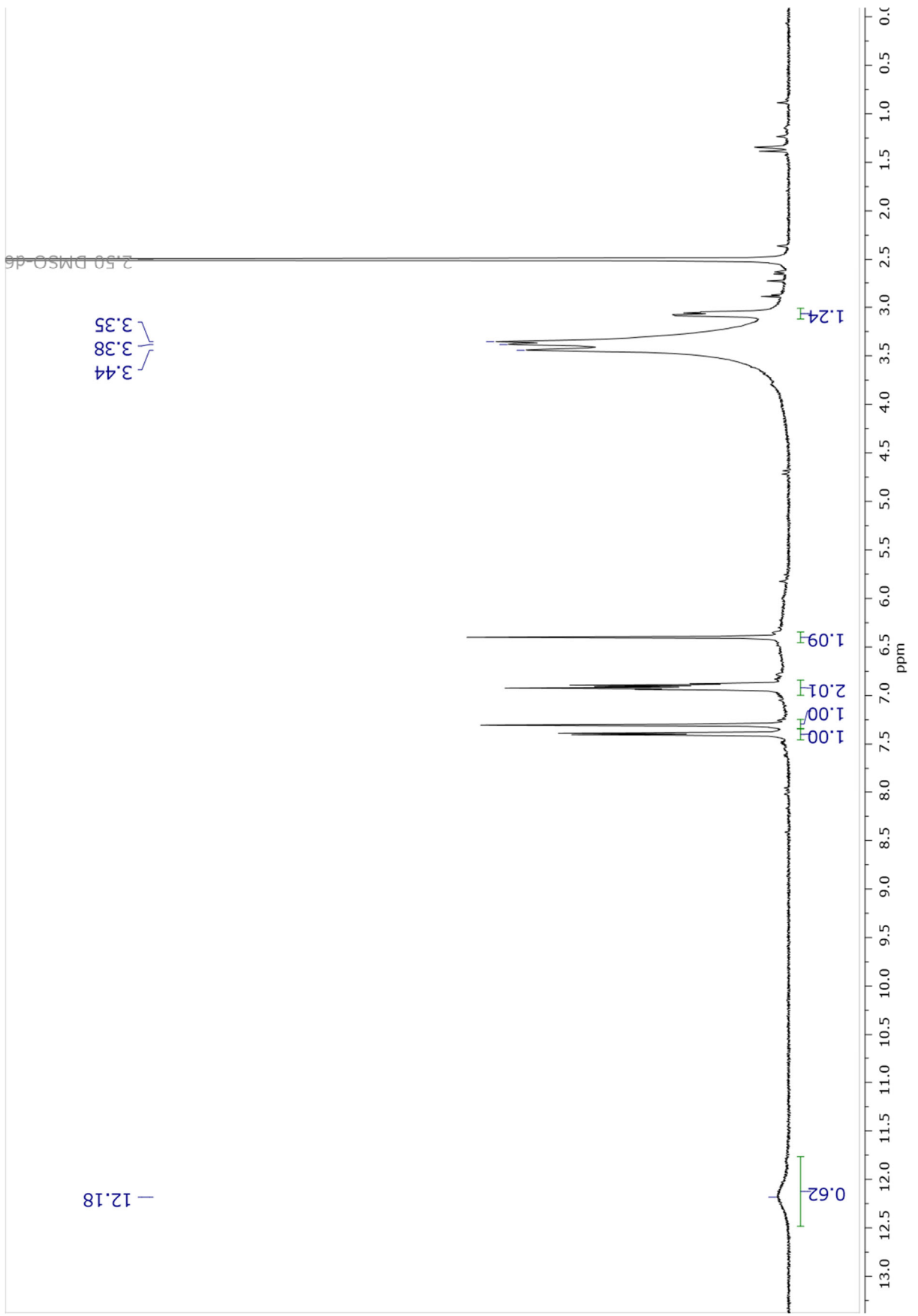
¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 12.18 (br s, 1H, Indole-NH), 7.40 (d, ³*J* = 7.8 Hz, 1H, C4-H), 7.30 (d, ³*J* = 3.2 Hz, 1H, C2-H), 6.95-6.86 (m, 2H, C5-H/C6-H), 6.40 (d, ³*J* = 3.1 Hz, 1H, C3-H), 3.44 (m, 1H, C α -H), 3.36 (dd, ³*J* = 14.3 Hz, ³*J* = 4.2 Hz, 1H, C β -H), 3.07 (dd, ³*J* = 14.2 Hz, ³*J* = 6.3 Hz, 1H, C β -H).

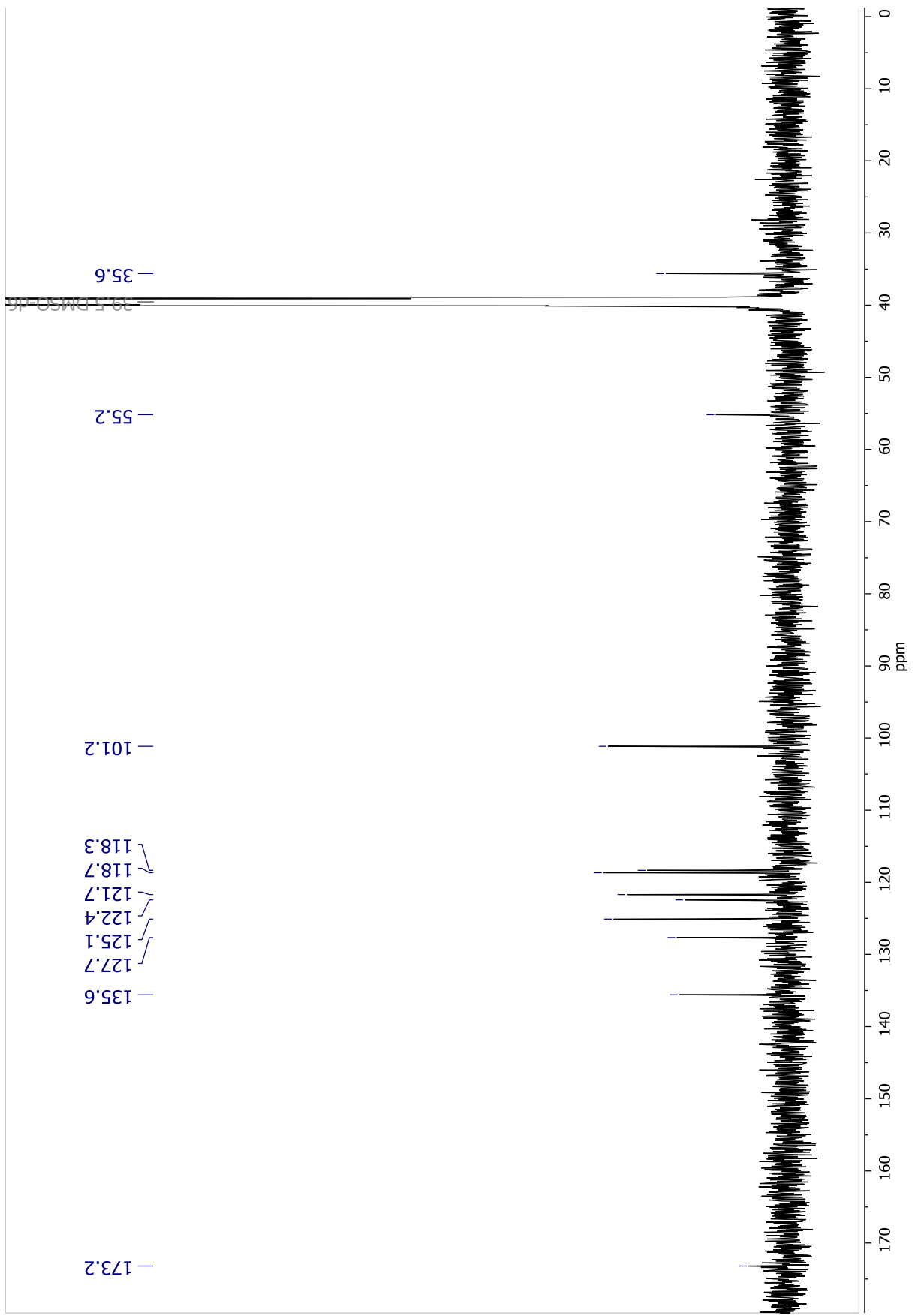
¹³C NMR (126 MHz, DMSO-*d*₆) δ = 173.2 (COOH), 135.6 (C7a), 127.7 (C3a), 125.1 (C7), 122.4 (C2), 121.7 (C4), 118.7 (C6), 118.3 (C5), 101.2 (C3), 55.2 (C α), 35.6 (C β).

MS (ESI): found [*m/z*] = 205.03 [M+H]⁺, calcd. [*m/z*] = 205.10 [M+H]⁺.

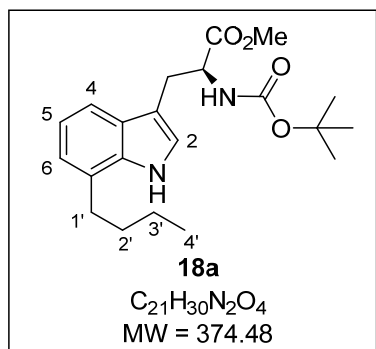
Marfey's Derivatization: t_R (L-deriv.) = 7.7 min (69 %); t_R (D-deriv.) = 9.1 min (31 %)
found [*m/z*] = 457.04 [M+H]⁺, calcd. [*m/z*] = 457.15 [M+H]⁺.







N^α-Boc-L-7-Butyltryptophan methyl ester (**18a**)



Method A: Nickel-catalysed reductive cross-coupling

N^α-Boc-L-7-Butyltryptophan methyl ester (**18a**) was synthesized according to GP1. *N*^α-Boc-L-7-bromotryptophan methyl ester (**17**) (49.2 mg, 124 μmol), NiI₂ (4.7 mg, 13 μmol, 10 mol%) and 4,4'-Di-*tert*-butyl-2,2'-dipyridine (3.8 mg, 13 μmol, 10 mol%) were placed in a glass vial under argon atmosphere. Everything was suspended in DMPU (0.5 mL) and purged with argon. 1-iodobutane (**3**, 31 μL, 252 μmol, 2.2 eq.), pyridine (1.0 μL, 13 μmol, 10 mol%) and manganese (27.4 mg, 499 μmol, 4.0 eq.)

were added. The green reaction mixture was heated to 60 °C and stirred for 21 h. The mixture was purified directly by column chromatography (Petroleum ether/EtOAc; 3:1) providing **18a** as a colorless solid (32.2 mg, 86 μmol, 70 %).

Method B: Negishi cross-coupling

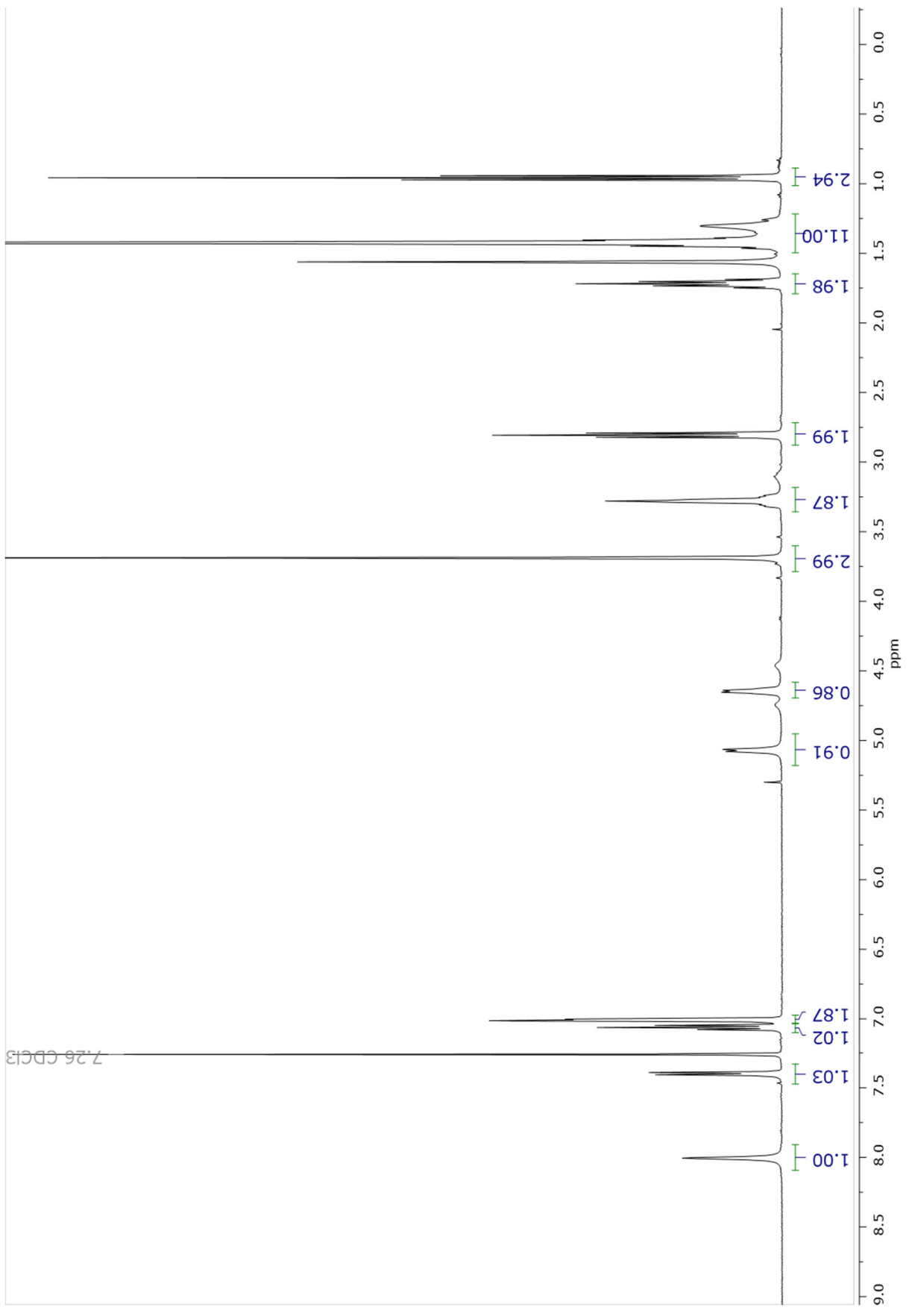
N^α-Boc-L-7-butyl tryptophan methyl ester (**18a**) was synthesized according to GP2. Therefore, *N*^α-Boc-L-7-bromotryptophan methyl ester (**17**, 39.1 mg, 98 μmol), zinc dust (13.4 mg, 207 μmol, 2.1 eq.), 1-iodobutane (**3**, 22.7 μL, 200 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (3.6 mg, 0.5 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.2 mL). The suspension was degassed by sparkling it with argon and stirred at 37 °C for 24 h. The mixture was purified by column chromatography (Petroleum ether/EtOAc; 4:1) providing **18a** as a colorless solid (30.5 mg, 81 μmol, 83 %).

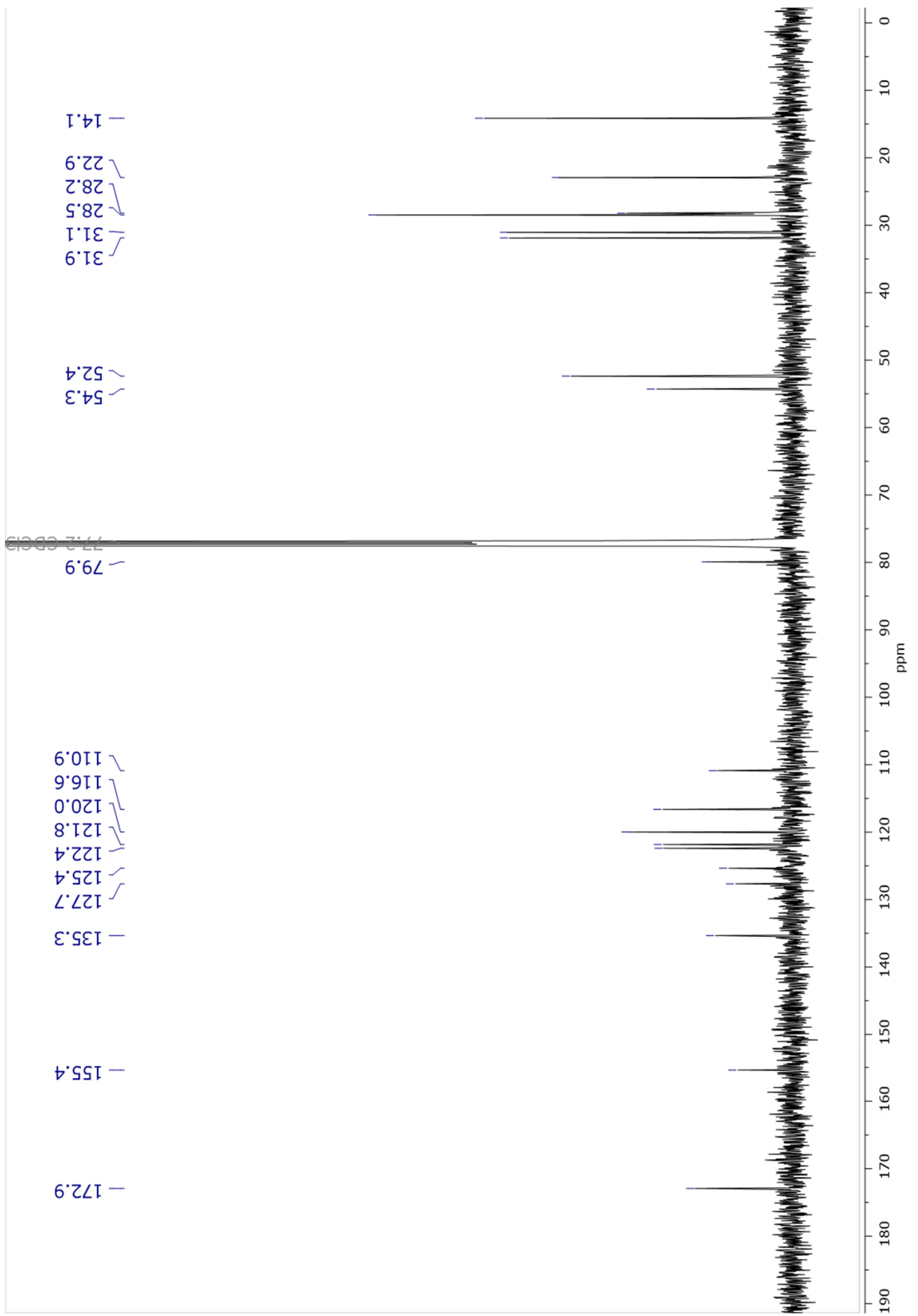
LC-MS: *t*_R = 11.0 min;

¹H NMR (500 MHz, CDCl₃) δ [ppm] = 8.01 (s, Indole-NH), 7.40 (d, ³*J* = 7.9 Hz, 1H, C4-H), 7.06 (dd, ³*J* = 7.5 Hz, ³*J* = 7.5 Hz, 1H, C5-H), 7.03-6.99 (m, 2H, C2-H/C6-H), 5.07 (d, ³*J* = 7.9 Hz, 1H, OCONH), 4.65 (ddd, ³*J* = 8.0 Hz, ³*J* = 5.2 Hz, ³*J* = 5.2 Hz, 1H, Cα-H), 3.69 (s, 3H, OCH₃), 3.31 (dd, ²*J* = 14.4 Hz, ³*J* = 5.0 Hz, 1H, Cβ-H), 3.21 (dd, ²*J* = 14.4 Hz, ³*J* = 5.2 Hz, 1H, Cβ-H), 2.81 (t, ³*J* = 7.7 Hz, 2H, C1'-H), 1.72 (tt, ³*J* = 7.7 Hz, ³*J* = 7.5 Hz, 2H, C2'-H), 1.50-1.25 (m, 11H, C(CH₃)₃/C3'-H), 0.96 (t, ³*J* = 7.3 Hz, 3H, C4'-H).

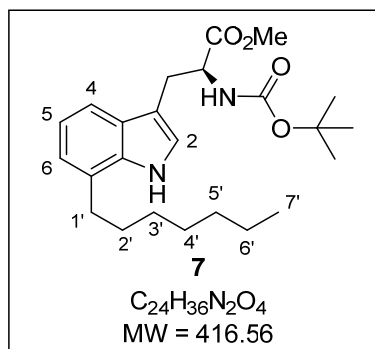
¹³C NMR (126 MHz, CDCl₃) δ [ppm] = 172.9 (COOMe), 155.4 (NHCOO), 135.3 (C7a), 127.7 (C4a), 125.4 (C2), 122.4 (C6), 121.8 (C7), 120.0 (C5), 116.6 (C4), 110.9 (C3), 79.9 (C(CH₃)₃), 54.3 (Cα), 52.4 (COO(CH₃)), 31.9 (C2'), 31.0 (C1'), 28.5 (C(CH₃)₃), 28.2 (Cβ), 22.9 (C3'), 14.1 (C4').

MS (ESI): found [*m/z*] = 375.23 [M+H]⁺, 319.17 [M-(*tert*Butyl)+H]⁺, 275.18 [M-Boc+H]⁺, 258.15 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 375.23 [M+H]⁺, 319.17 [M-(*tert*Butyl)+H]⁺, 275.18 [M-Boc+H]⁺, 258.15 [M-Boc-NH₂]⁺.





N^α-Boc-L-7-Heptyltryptophan methyl ester (**7**)



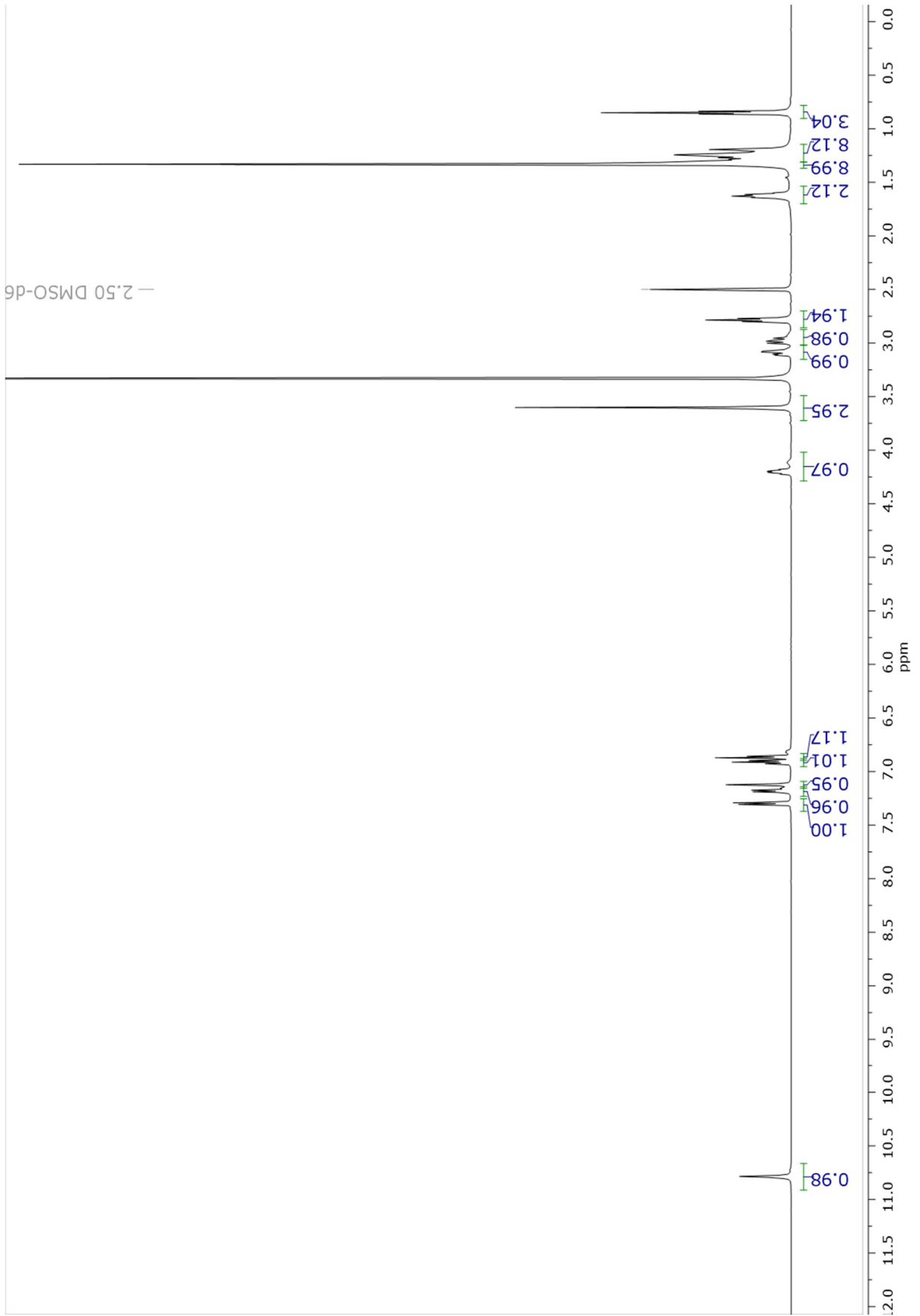
N^α-Boc-L-7-Heptyltryptophan methyl ester (**7**) was synthesized according to *GPKll*. *N*^α-Boc-L-7-bromo-tryptophan methyl ester (**17**) (20.0 mg, 50 μmol), NiI₂ (2.0 mg, 5 μmol, 10 mol%) and 4,4''-Di-*tert*-butyl-2,2'-dipyridine (1.4 mg, 5 μmol, 10 mol%) were placed in a glass vial under argon atmosphere. Everything was suspended in DMPU (0.2 mL) and purged with argon. 1-iodoheptane (**4**, 18 μL, 110 μmol, 2.2 eq.), pyridine (0.5 μL, 6 μmol, 12 mol%) and manganese (11.1 mg, 203 μmol, 4.0 eq.) were added. The green reaction mixture was heated to 60 °C and stirred for 21 h. The mixture was purified directly by column chromatography (Petroleum ether/EtOAc; 3:1) providing **7** as a colorless solid (17.2 mg, 41 μmol, 82 %).

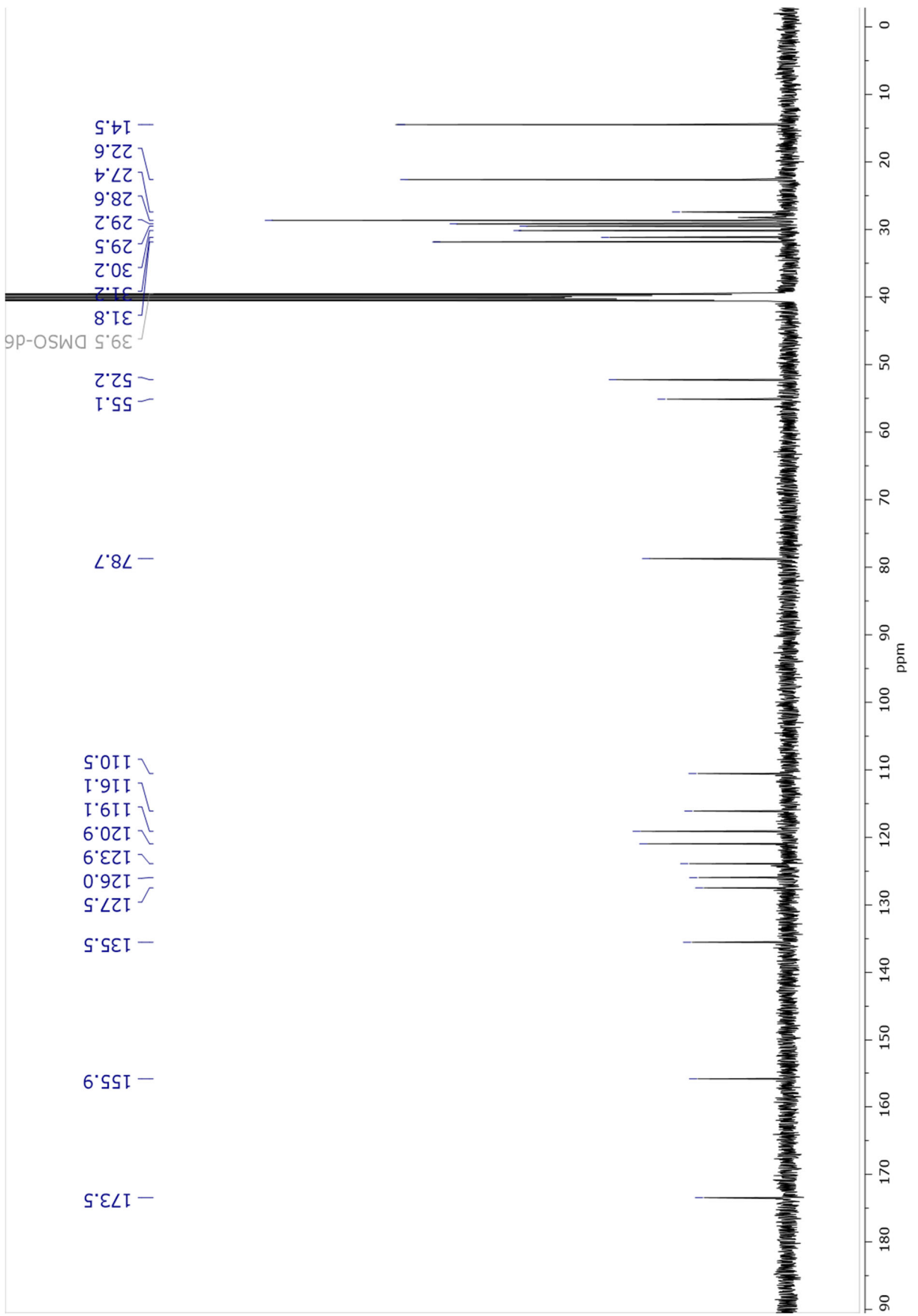
LC-MS: *t*_R = 11.7 min;

¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 10.79 (s, Indole-NH), 7.30 (d, ³*J* = 7.8 Hz, 1H, C4-H), 7.18 (d, ³*J* = 7.8 Hz, 1H, OCONH), 7.12 (d, ³*J* = 2.1 Hz, 1H, C2-H), 6.91 (dd, ³*J* = 7.5 Hz, ³*J* = 7.5 Hz, 1H, C5-H), 6.86 (d, ³*J* = 7.0 Hz, 1H, C6-H), 4.20 (ddd, ³*J* = 9.3 Hz, ³*J* = 8.0 Hz, ³*J* = 4.7 Hz, 1H, Cα-H), 3.09 (dd, ²*J* = 14.4, ³*J* = 4.8 Hz, 1H, Cβ-H), 2.97 (dd, ²*J* = 14.5 Hz, ³*J* = 9.4 Hz, 1H, Cβ-H), 2.79 (t, ³*J* = 7.6 Hz, 2H, C1'-H), 1.63 (tt, ³*J* = 7.5 Hz, ³*J* = 7.5 Hz, 2H, C2'-H), 1.32 (s, 9H, C(CH₃)₃), 1.29 – 1.15 (m, 8H, C3'-H/ C4'-H/ C5'-H/ C6'-H), 0.85 (t, ³*J* = 6.9 Hz, 3H, C7'-H).

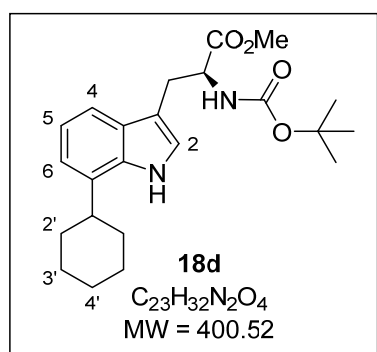
¹³C NMR (126 MHz, DMSO-*d*₆) δ [ppm] = 173.4 (COOMe), 155.8 (NHCOO), 135.4 (C7a), 127.4 (C3a), 125.9 (C7), 123.8 (C2), 120.9 (C6), 119.0 (C5), 116.0 (C4), 110.5 (C3), 78.7 (C(CH₃)₃), 55.1 (Cα), 52.2 (OCH₃), 31.8 (C1'), 31.1 (C2'), 30.1 (C3'), 29.5 (C4'), 29.1 (C5'), 28.6 (C6'), 27.4 (C(CH₃)₃), 22.6 (Cβ), 14.4 (C7').

MS (ESI): found [*m/z*] = 417.28 [M+H]⁺, 361.21 [M-(*tert*Butyl)+H]⁺, 317.27 [M-Boc+H]⁺, 300.20 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 417.27 [M+H]⁺, 361.21 [M-(*tert*Butyl)+H]⁺, 317.27 [M-Boc+H]⁺, 300.20 [M-Boc-NH₂]⁺.





N^α-Boc-L-7-Cyclohexyltryptophan methyl ester (**18d**)



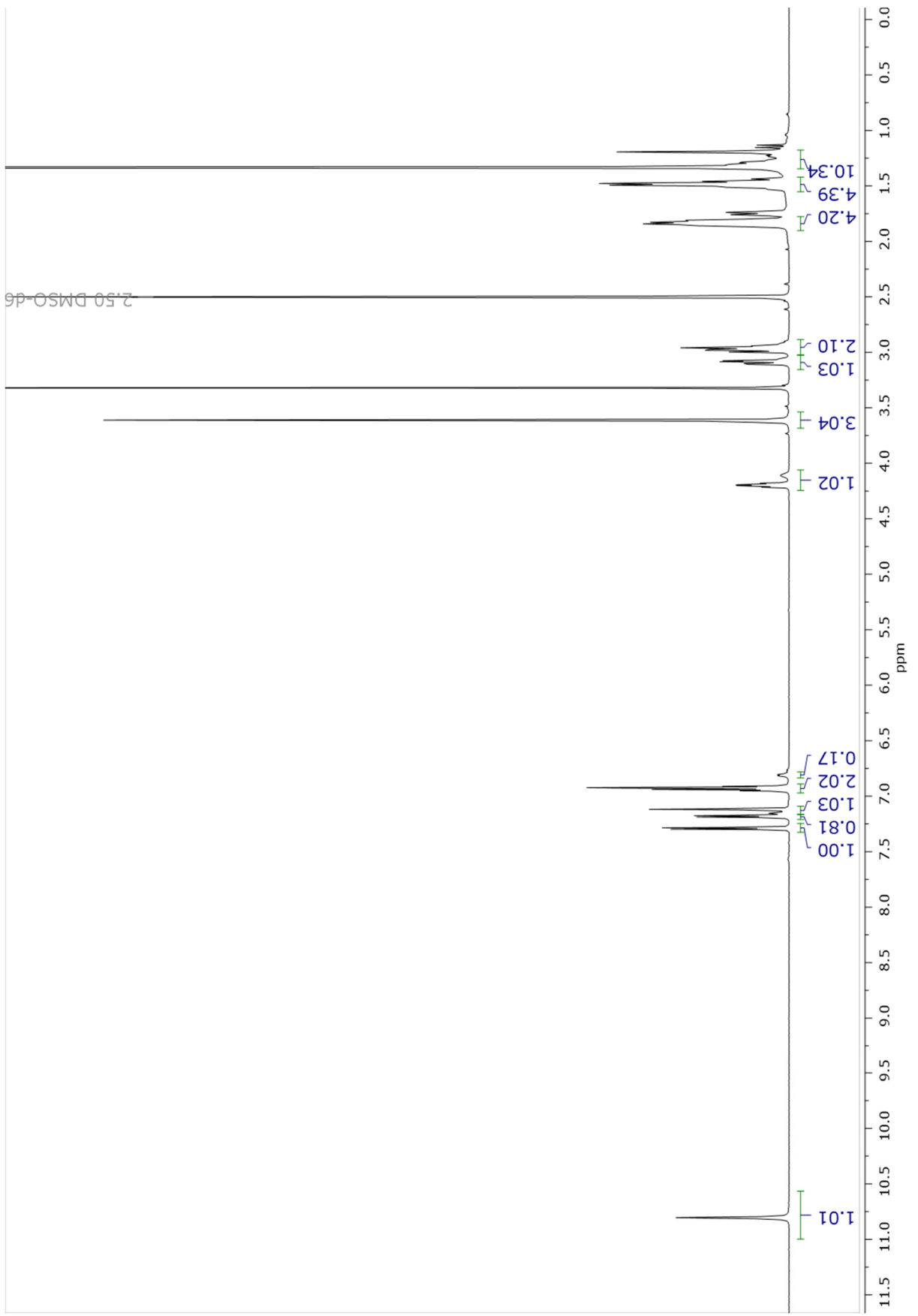
N^α-Boc-L-7-Cyclohexyltryptophan methyl ester (**18c**) was synthesized according to GP2. *N*^α-Boc-L-7-bromo-tryptophan methyl ester (**XX**) (40.0 mg, 100 μmol), zinc dust (13.2 mg, 202 μmol, 2.0 eq.), cyclohexyl iodide (**15**, 26 μL, 201 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (3.5 mg, 0.5 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.2 mL). The suspension was degassed by sparkling it with argon and stirred at 37°C for 24 h. The mixture was purified by column chromatography (Petrolether/EtOAc; 4:1) providing **18d** as a colorless solid (29.4 mg, 73 μmol, 73 %).

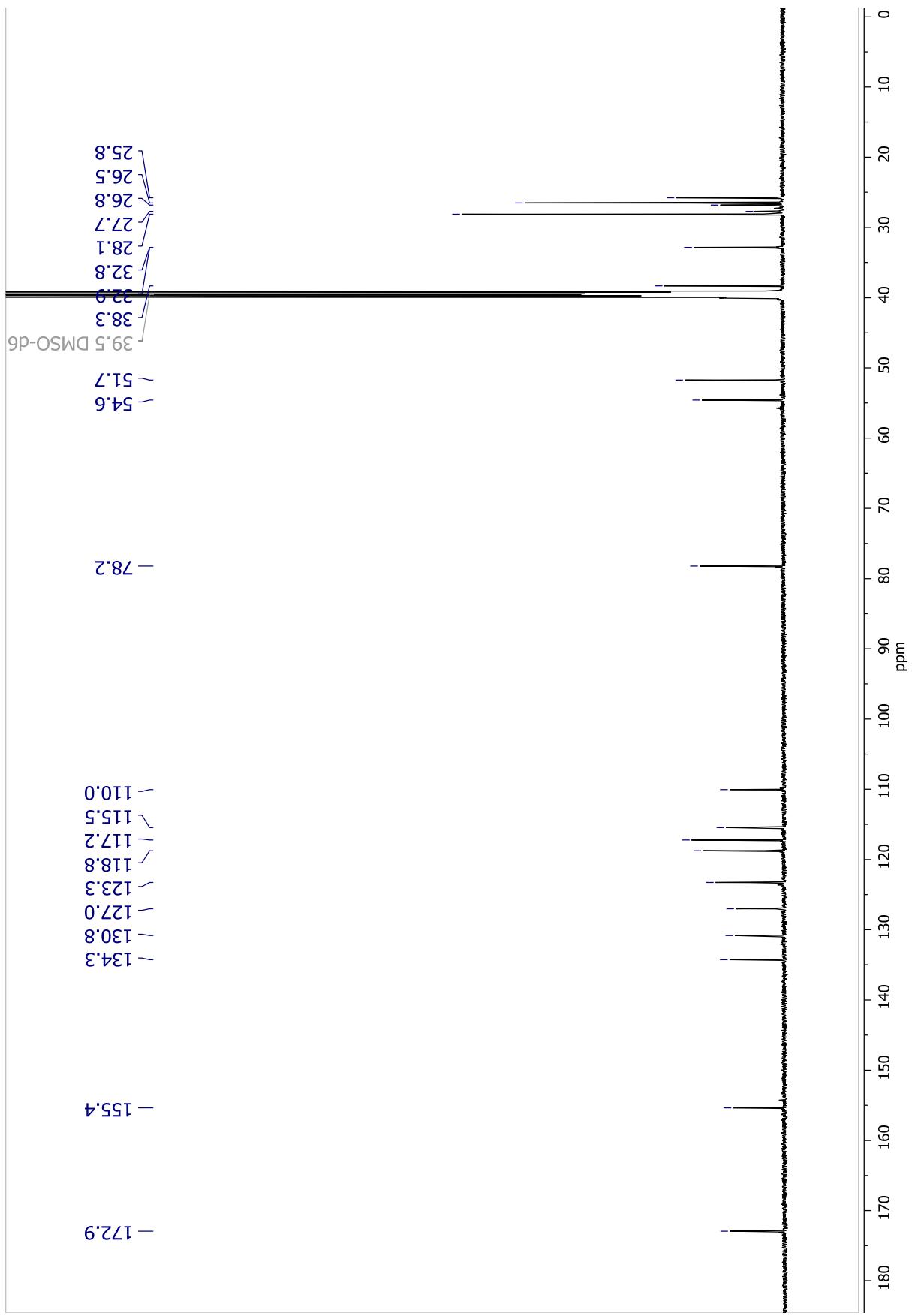
LC-MS: *t*_R = 11.2 min;

¹H NMR (600 MHz, DMSO-*d*₆) δ [ppm] = 10.80 (d, ³*J* = 2.1 Hz, indole-NH), 7.29 (dd, ³*J* = 7.4 Hz, ³*J* = 1.8 Hz, 1H, C4-H), 7.18 (d, ³*J* = 7.8 Hz, 0.8H, OCONH), 7.12 (d, ³*J* = 2.6 Hz, 1H, C2-H), 6.96-6.93 (m, 2H, C6-H/C5-H), 6.81 (d, ³*J* = 6.5 Hz, 0.2H, OCONH), 4.20 (ddd, ³*J* = 9.3 Hz, ³*J* = 8.0 Hz, ³*J* = 5.1 Hz, 1H, Cα-H), 3.62 (s, 3H, OCH₃), 3.09 (dd, ²*J* = 14.6, ³*J* = 5.2 Hz, 1H, Cβ-H), 3.03-2.90 (m, 2H, Cβ-H/C1'-H), 1.90-1.71 (m, 4H, C2'-H), 1.38-1.20 (m, 10H, (C(CH₃)₃; cis/trans ratio 4:1)/C4'-).

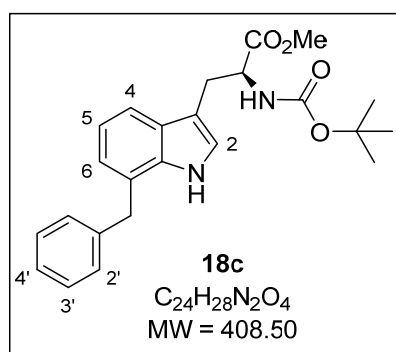
¹³C NMR (151 MHz, DMSO-*d*₆) δ [ppm] = 172.9 (COOMe), 155.4 (NHCOO), 135.3 (C7a), 130.8 (C3a), 127.0 (C7), 123.3 (C2), 118.8 (C6), 117.2 (C5), 115.5 (C4), 110.1 (C3), 78.2 (C(CH₃)₃), 54.6 (Cα), 51.7 (OCH₃), 38.3 (C1'), 32.9 (C2'), 32.8 (C6'), 28.1 (C(CH₃)₃), 27.7 (Cβ), 26.8 (C3'), 26.5 (C5'), 25.8 (C4').

MS (ESI): found [*m/z*] = 401.25 [M+H]⁺, 345.19 [M-(*tert*Butyl)+H]⁺, 301.19 [M-Boc+H]⁺, 284.17 [M-Boc-NH₂]⁺; calcd. [*m/z*] = 401.24 [M+H]⁺, 345.18 [M-(*tert*Butyl)+H]⁺, 301.19 [M-Boc+H]⁺, 284.16 [M-Boc-NH₂]⁺.





N^{α} -Boc-L-7-benzyltryptophan methyl ester (**18c**)



N^{α} -Boc-L-7-Benzyltryptophan methyl ester (**18c**) was synthesized according to GP2. Therefore, N^{α} -Boc-L-7-bromotryptophan methyl ester (**17**, 41.1 mg, 100 μ mol), zinc dust (13.1 mg, 200 μ mol, 2.0 eq.), freshly distilled benzyl iodide (**9**, 39.5 μ L, 200 μ mol, 2.0 eq.) and Pd(amphos)₂Cl₂ (3.5 mg, 0.5 μ mol, 5 mol%) were placed in a glass vial followed by DMF (0.2 mL). The suspension was degassed by sparkling it with argon and stirred at 37°C for 24 h. The mixture was purified by column chromatography (Petrolether/EtOAc; 4:1) providing **18c** as a colorless solid (33.0 mg,

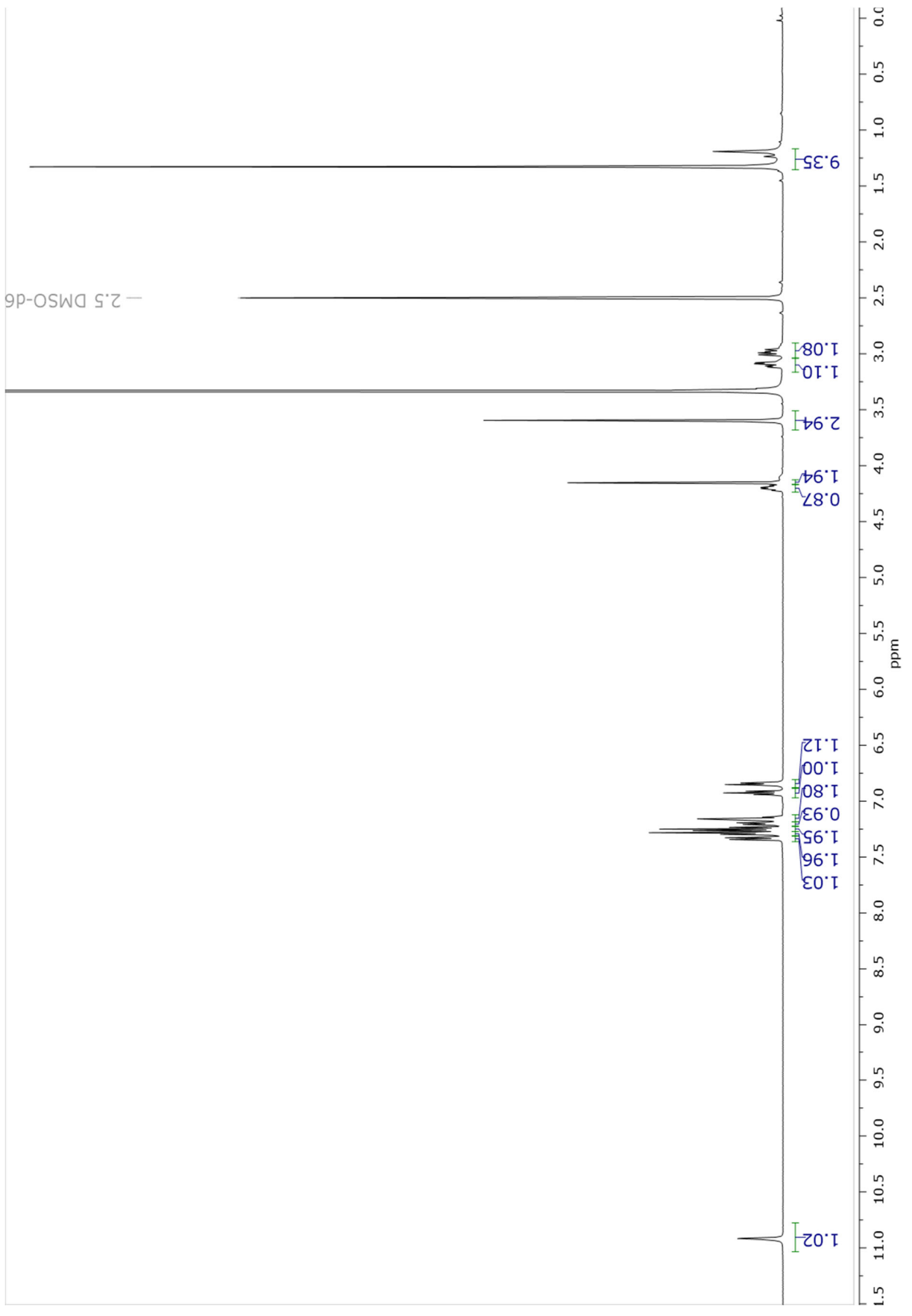
81 μ mol, 81 %)

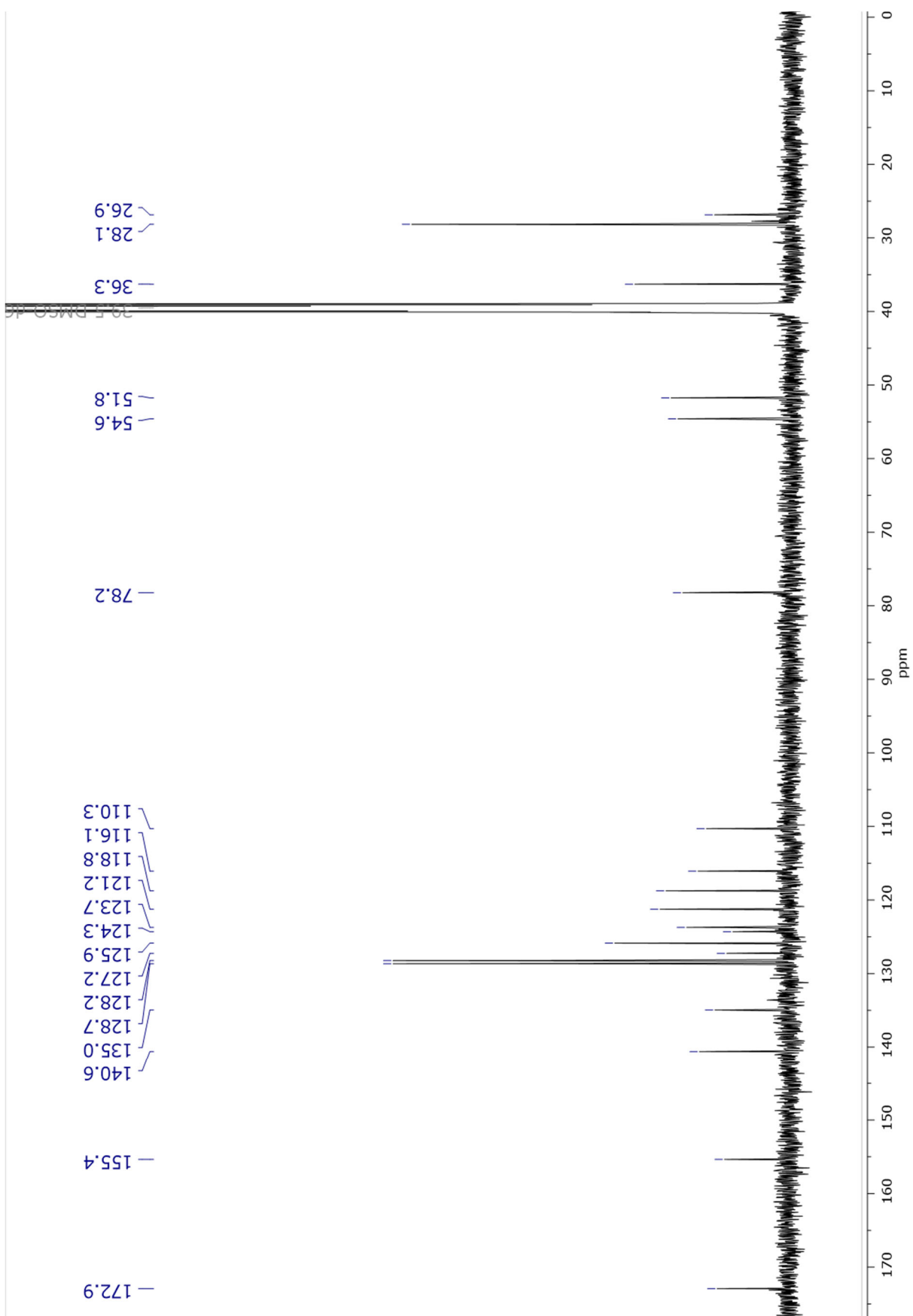
LC-MS: t_R = 10.0 min;

¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 10.92 (s, Indole-NH), 7.34 (d, ³*J* = 7.9 Hz, 1H, C4-H), 7.29 (m, 2H, C2'-H), 7.25 (m, 2H, C3'-H), 7.20 (d, ³*J* = 7.8 Hz, 1H, OCONH), 7.18-7.15 (m, 2H, C4'-H/C2-H), 6.93 (dd, ³*J* = 7.8 Hz, ³*J* = 7.0 Hz, 1H, C5-H), 6.85 (d, ³*J* = 7.0 Hz, 1H, C6-H), 4.20 (ddd, ³*J* = 9.2 Hz, ³*J* = 8.0 Hz, ³*J* = 5.3 Hz, 1H, C α -H), 4.15 (s, 2H, CH₂), 3.60 (s, 3H, OCH₃), 3.10 (dd, ²*J* = 14.4, ³*J* = 5.3 Hz, 1H, C β -H), 2.99 (dd, ²*J* = 14.6 Hz, ³*J* = 9.2 Hz, 1H, C β -H), 1.33 (s, 9H, C(CH₃)₃; cis/trans ratio 5:1);

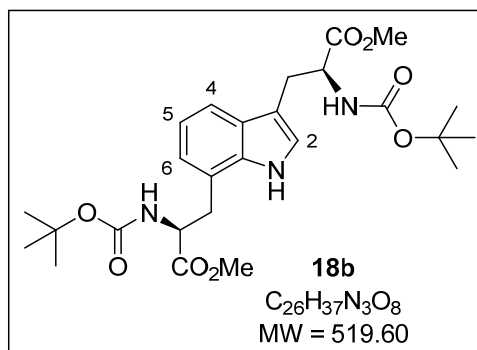
¹³C NMR (151 MHz, DMSO-*d*₆) δ [ppm] = 172.9 (COOMe), 155.4 (NHCOO), 140.6 (C1'), 134.9 (C7a), 128.7 (C2'), 128.2 (C3'), 127.2 (C3a), 125.9 (C4'), 124.3 (C7), 123.7 (C2), 121.2 (C4), 118.8 (C6), 116.1 (C5), 110.3 (C3), 78.2 (C(CH₃)₃), 54.6 (C α), 51.8 (OCH₃), 36.3 (CH₂), 28.2 (C(CH₃)₃), 26.9 (C β).

MS (ESI): found [*m/z*] = 409.20 [M+H]⁺, 353.15 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.13 [M-Boc-NH₂]⁺; calcd. [*m/z*] = 409.21 [M+H]⁺, 353.15 [M-(*tert*Butyl)+H]⁺, 309.16 [M-Boc+H]⁺, 292.13 [M-Boc-NH₂]⁺.





Dimethyl (2S,2'S)-3,3'-(1H-indole-3,7-diyl)bis(2-((tert-butoxycarbonyl)amino)propanoate) (**18b**)



Dimethyl (2S,2'S)-3,3'-(1H-indole-3,7-diyl)bis(2-((tert-butoxycarbonyl)amino)propanoate) (**18b**) was synthesized according to GP3. *N*^α-Boc-L-7-Bromotryptophan methyl ester (**17**, 39.1 mg, 98 μmol), zinc dust (13.0 mg, 199 μmol, 2.0 eq.), *N*^α-Boc-L-3-iodoalanine methyl ester (**8**, 65.9 mg, 200 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (3.6 mg, 0.5 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.2 mL). The suspension was degassed by sparkling it with argon and stirred at 37°C for 24 h. The mixture was purified by column

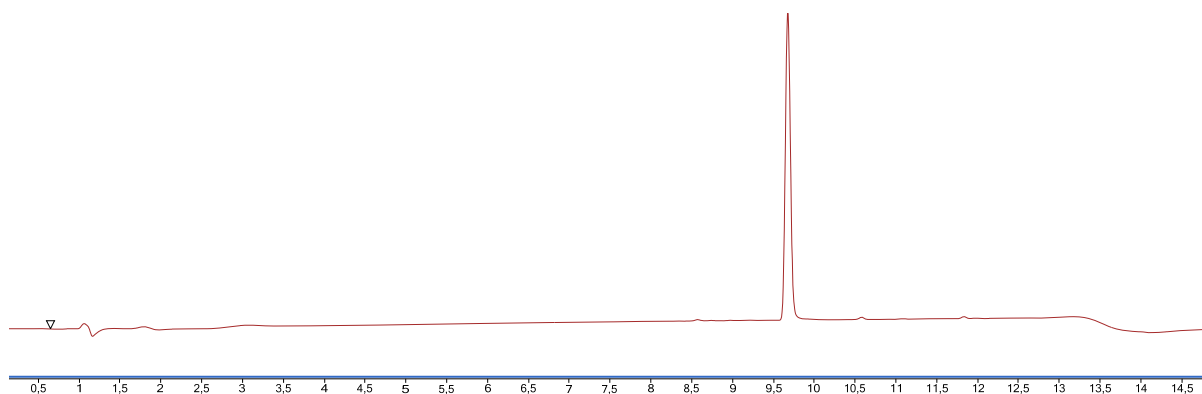
chromatography (Petrolether/EtOAc; 2:1) providing **18b** as a colorless solid (27.9 mg, 54 μmol, 55 %)

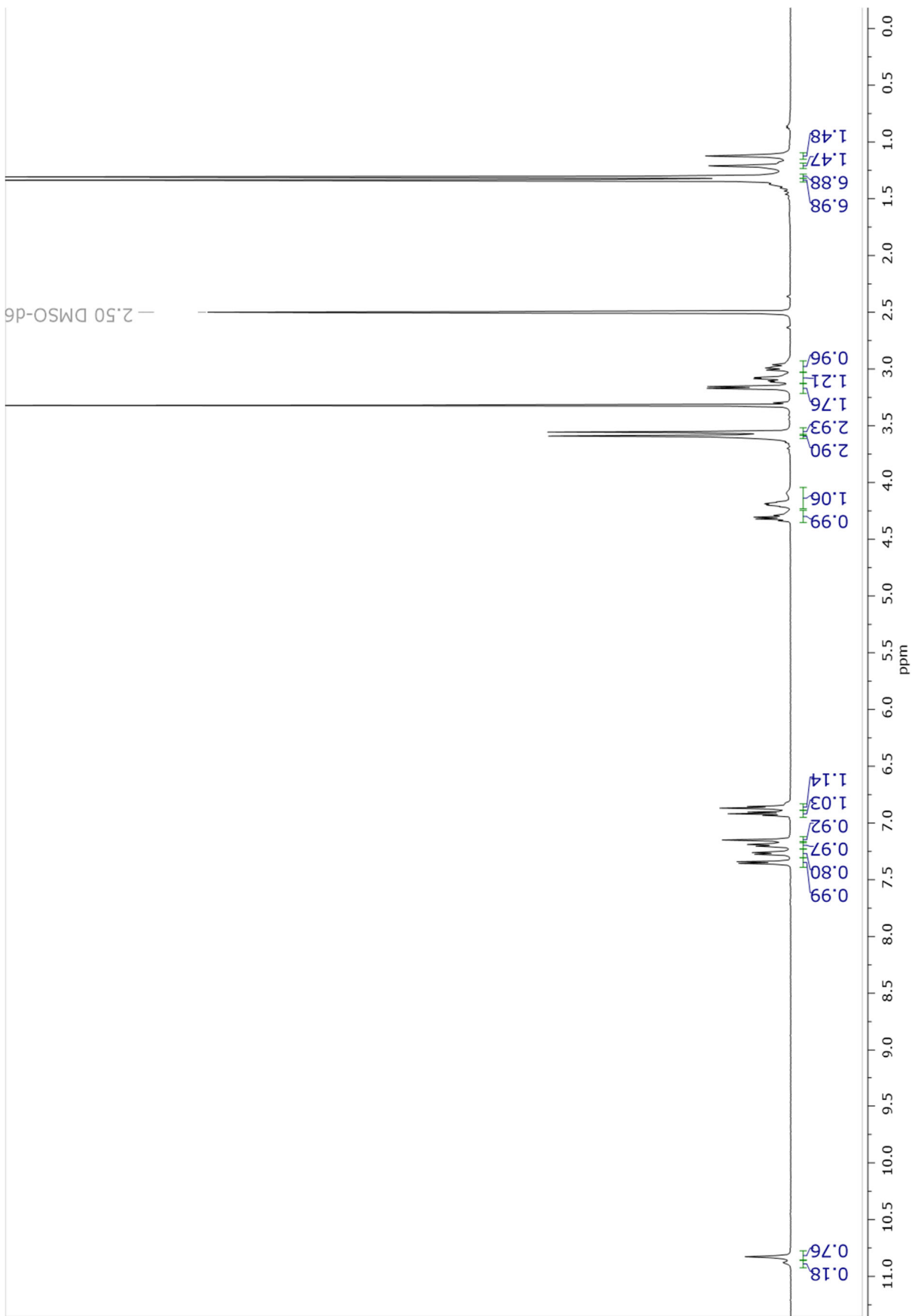
LC-MS: *t*_R = 9.6 min;

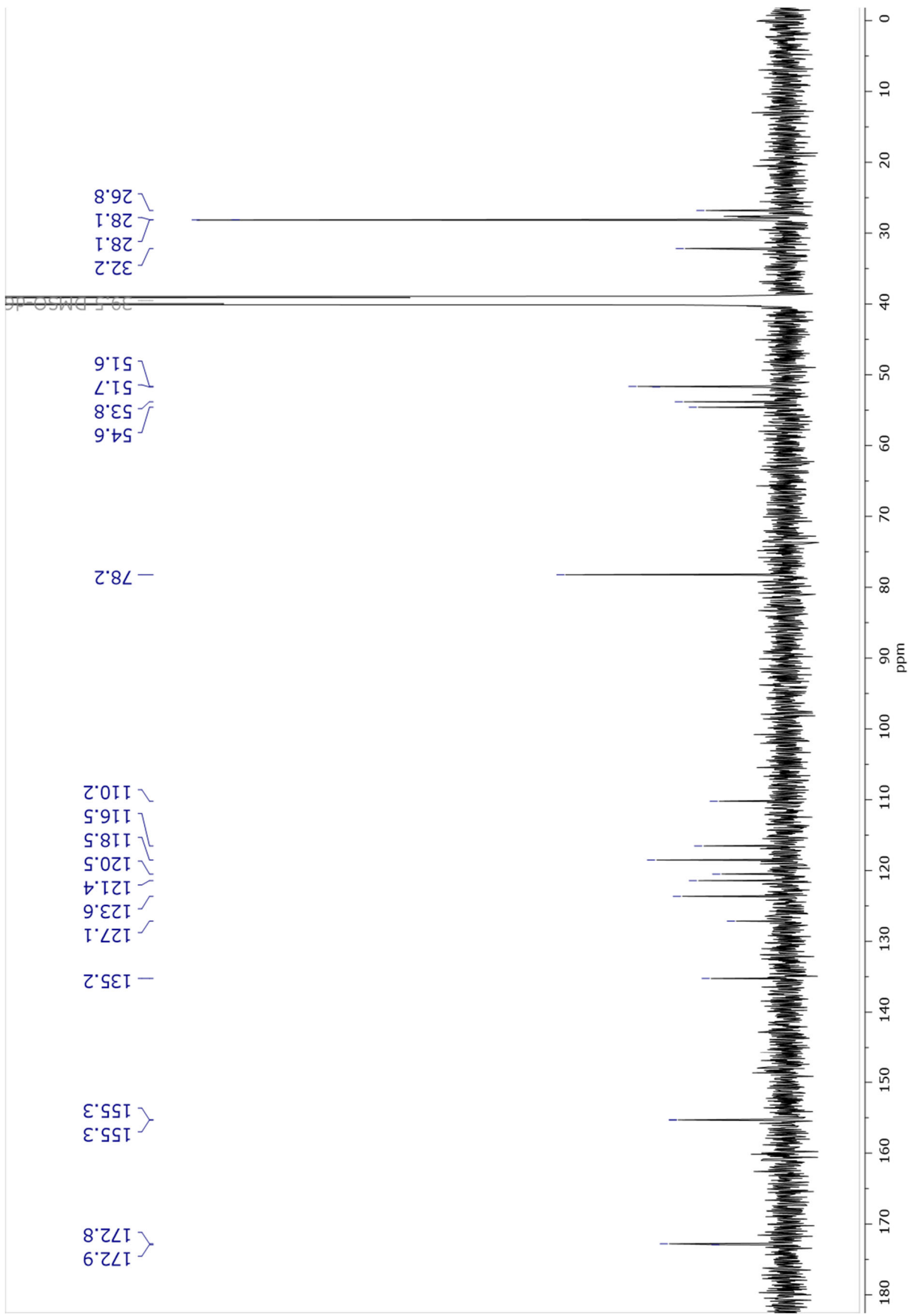
¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 10.86 (s, Indole-NH, cis/trans ratio 4:1), 7.36 (d, ³*J* = 7.8 Hz, 1H, C4-H), 7.27 (d, ³*J* = 7.5 Hz, 1H, OCONH^{Ala}), 7.20 (d, ³*J* = 7.5 Hz, 1H, OCONH^{Trp}), 7.15 (d, ³*J* = 2.6 Hz, 1H, C2-H), 6.92 (dd, ³*J* = 7.7 Hz, ³*J* = 7.7 Hz, 1H, C5-H), 6.86 (d, ³*J* = 7.5 Hz, 1H, C6-H), 4.31 (dd, ³*J* = 7.9 Hz, ³*J* = 7.9 Hz, 1H, Cα-H^{Ala}), 4.19 (ddd, ³*J* = 9.2 Hz, ³*J* = 7.5 Hz, ³*J* = 5.3 Hz 1H, Cα-H^{Trp}), 3.60 (s, 3H, OCH₃), 3.56 (s, 3H, OCH₃), 3.17 (d, ³*J* = 7.9 Hz, 2H, Cβ-H^{Ala}), 3.10 (dd, ²*J* = 14.4 Hz, ³*J* = 5.3 Hz, 1H, Cβ-H), 2.99 (dd, ²*J* = 14.9 Hz, ³*J* = 9.2 Hz, 1H, Cβ-H), 1.34 (s, 9H, C(CH₃)₃^{Ala}; cis/trans ratio 4:1), 1.32 (s, 9H, C(CH₃)₃^{Trp}; cis/trans ratio 4:1);

¹³C NMR (126 MHz, DMSO-*d*₆) δ [ppm] = 172.9 (COOMe), 172.8 (COOMe), 155.4 (NHCOO), 155.3 (NHCOO), 135.2 (C7a), 127.1 (C3a), 123.7 (C2), 121.4 (C4), 120.5 (C7), 118.5 (C6), 116.5 (C5), 110.2 (C3), 78.2 (C(CH₃)₃), 54.6 (Cα^{Trp}), 53.8 (Cα^{Ala}), 51.7 (OCH₃), 51.6 (OCH₃), 32.2 (CH₂^{Ala}), 28.2 (C(CH₃)₃), 28.1 (C(CH₃)₃), 26.8 (Cβ^{Trp}).

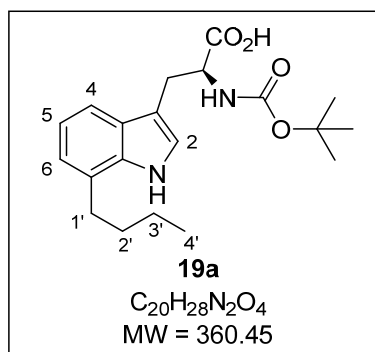
MS (ESI): found [*m/z*] = 520.26 [M+H]⁺, 464.21 [M-(*tert*Butyl)+H]⁺, 420.21 [M-Boc+H]⁺, 403.13 [M-Boc-NH₂]⁺, 364.15 [M-Boc-(*tert*Butyl)+H]⁺, 347.12 [M-Boc-(*tert*Butyl)-NH₂]⁺, 320.16 [M-2Boc+H]⁺, 303.13 [M-2Boc-NH₂]⁺; calcd. [*m/z*] = 520.27 [M+H]⁺, 464.20 [M-(*tert*Butyl)+H]⁺, 420.21 [M-Boc+H]⁺, 403.18 [M-Boc-NH₂]⁺, 364.15 [M-Boc-(*tert*Butyl)+H]⁺, 347.12 [M-Boc-(*tert*Butyl)-NH₂]⁺, 320.16 [M-2Boc+H]⁺, 303.13 [M-2Boc-NH₂]⁺.







N^α-Boc-L-7-Butyl tryptophan (**20a**)



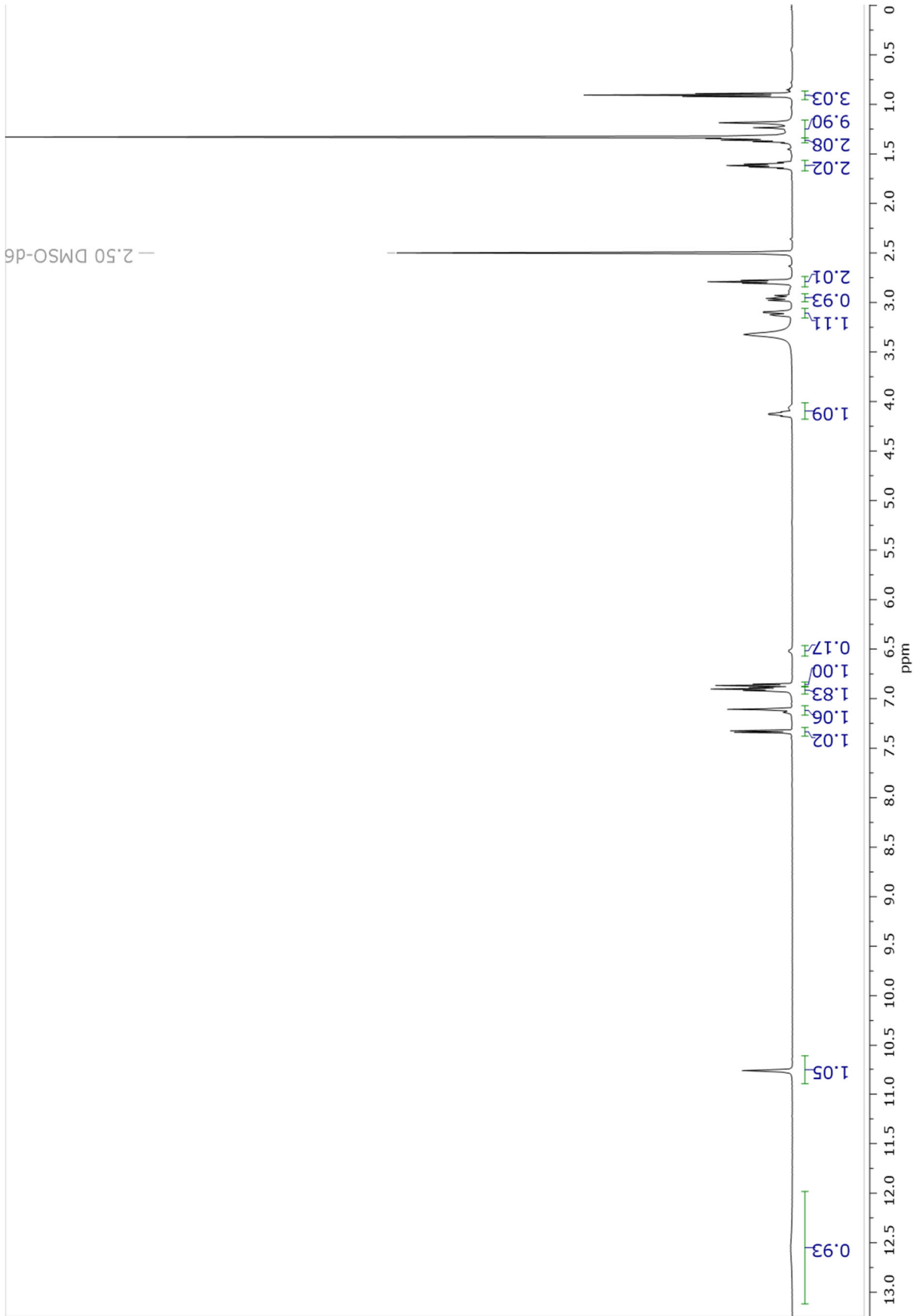
N^α-Boc-L-7-butyl tryptophan (**20a**) was synthesized according to GP2. Therefore, *N*^α-Boc-L-7-bromotryptophan (**19**, 38.3 mg, 97 μmol), zinc dust (13.2 mg, 202 μmol, 2.1 eq.), 1-iodobutane (**3**, 22.7 μL, 200 μmol, 2.0 eq.) and Pd(amphos)₂Cl₂ (3.5 mg, 0.5 μmol, 5 mol%) were placed in a glass vial followed by DMF (0.2 mL). The suspension was degassed by sparkling it with argon and stirred at 37°C for 24 h. The mixture was purified by preparative RP-HPLC providing **20a** as a colorless solid (16.0 mg, 44 μmol, 46 %).

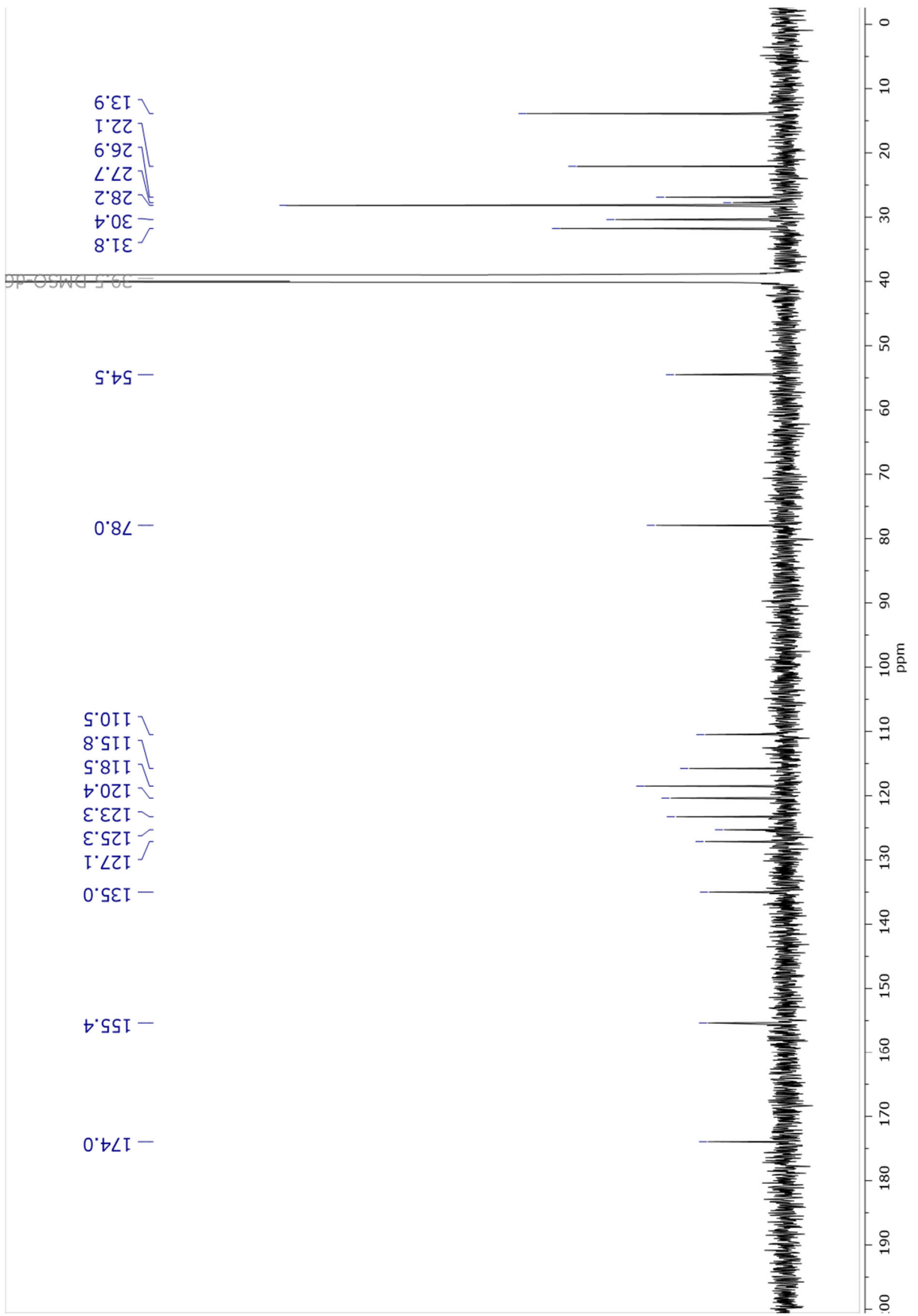
LC-MS: *t*_R = 9.6 min;

¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 12.58 (br s, 1H, COOH), 10.76 (s, 1H, Indole-NH), 7.33 (d, ³*J* = 7.8 Hz, 1H, C4-H), 7.11 (d, ³*J* = 2.4 Hz, 1H, C2-H), 6.90 (dd, ³*J* = 7.4 Hz, ³*J* = 7.4 Hz, 1H, C5-H), 6.86 (d, ³*J* = 7.0 Hz, 1H, C6-H), 4.13 (ddd, ³*J* = 9.3 Hz, ³*J* = 7.0 Hz, ³*J* = 4.7 Hz, 1H, C_α-H), 3.11 (dd, ²*J* = 14.5 Hz, ³*J* = 4.7 Hz, 1H, C_β-H), 2.95 (dd, ²*J* = 14.6 Hz, ³*J* = 9.3 Hz, 1H, C_β-H), 2.79 (t, ³*J* = 7.7 Hz, 2H, C1'-H), 1.62 (tt, ³*J* = 8.1 Hz, ³*J* = 7.6 Hz, 2H, C2'-H), 1.39-1.34 (m, 2H, C3'-H), 1.32 (s, 9H, C(CH₃)₃ (cis/trans ratio 5:1)), 0.91 (t, ³*J* = 7.3 Hz, 3H, C4'-H).

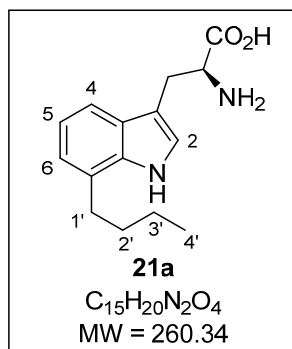
¹³C NMR (126 MHz, DMSO-*d*₆) δ [ppm] = 173.9 (COOH), 155.4 (NHCOO), 135.0 (C7a), 127.2 (C4a), 125.3 (C2), 123.3 (C6), 120.4 (C7), 118.5 (C5), 115.8 (C4), 110.5 (C3), 77.9 (C(CH₃)₃), 54.5 (C_α), 31.8 (C2'), 30.4 (C1'), 28.2 (C(CH₃)₃), 27.8 (C_β), 22.1 (C3') 13.9 (C4').

MS (ESI): found [*m/z*] = 361.20 [M+H]⁺, 305.14 [M-(*tert*Butyl)+H]⁺, 261.15 [M-Boc+H]⁺, 244.13 [M-Boc-NH₂]⁺, calcd. [*m/z*] = 361.21 [M+H]⁺, 305.15 [M-(*tert*Butyl)+H]⁺, 261.16 [M-Boc+H]⁺, 244.13 [M-Boc-NH₂]⁺.





L-7-Butyl tryptophan (**21a**)



L-7-butyl tryptophan (**21a**) was synthesized according to GP5. Therefore, *N*^α-Boc-L-7-butyl tryptophan methyl ester (**18a**) (9.5 mg, 25 μmol) was suspended in aqueous HCl (5 M, 2 mL) and heated to 90 °C for 2 hours. The crude reaction mixture was purified by reversed phase HPLC, providing **21a** as a colorless solid (8.0 mg, 21 μmol, 84 %). The enantiomeric excess was determined using Marfey's Test (see GP3) giving an ee higher 99 %.

LC-MS: *t*_R = 6.0 min;

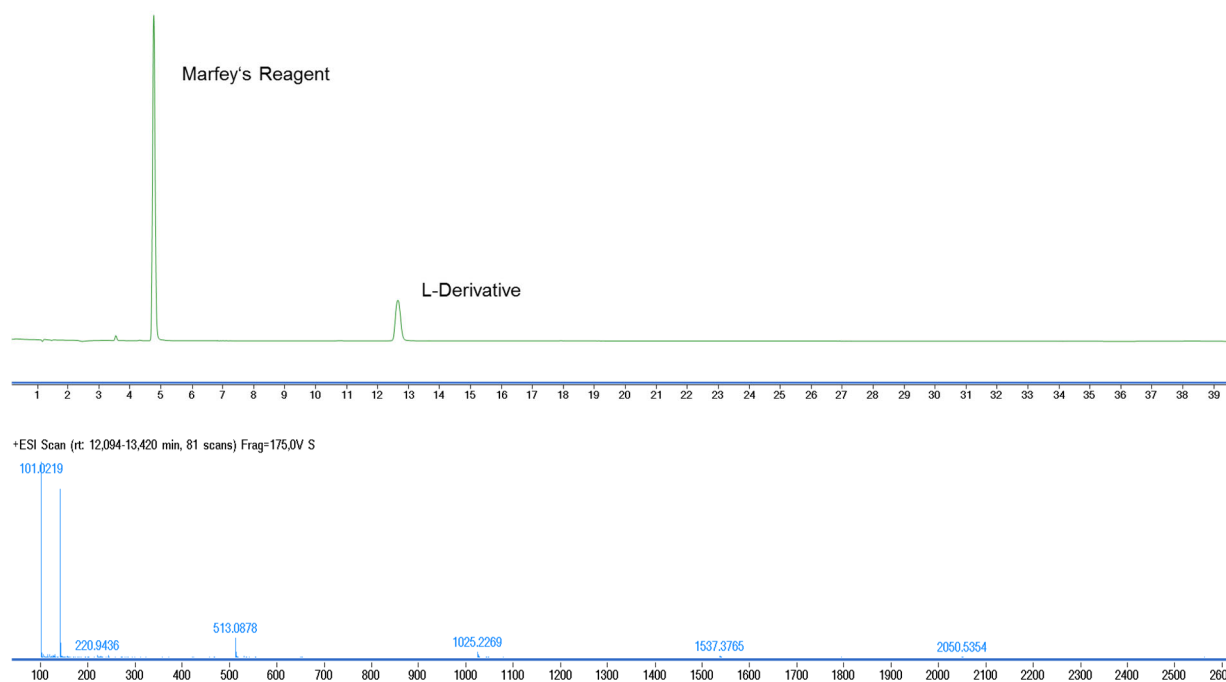
¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 10.83 (d, ³*J* = 2.6 Hz, 1H, Indole-NH), 7.52 (br s, 3H, NH₃⁺), 7.37 (d, ³*J* = 6.7 Hz, 1H, C4-H), 7.17 (d, ³*J* = 2.6 Hz, 1H, C2-H), 6.91 (dd, ³*J* = 6.7 Hz, ³*J* = 6.0 Hz, 1H, C5-H), 6.87 (d, ³*J* = 6.0 Hz, 1H, C6-H), 3.52 (dd, ³*J* = 8.8 Hz ³*J* = 4.2 Hz, 1H, Cα-H), 3.29 (dd, ²*J* = 15.2 Hz, ³*J* = 4.2 Hz, 1H, Cβ-H), 2.97 (dd, ²*J* = 15.2 Hz, ³*J* = 8.8 Hz, 1H, Cβ-H), 2.80 (t, ³*J* = 7.6 Hz, 2H, C1'-H), 1.62 (tt, ³*J* = 7.6 Hz, ³*J* = 6.5 Hz, 2H, C2'-H), 1.36 (h, ³*J* = 7.3 Hz, 2H, C3'-H) 0.91 (t, ³*J* = 7.4 Hz, 3H, C4'-H).

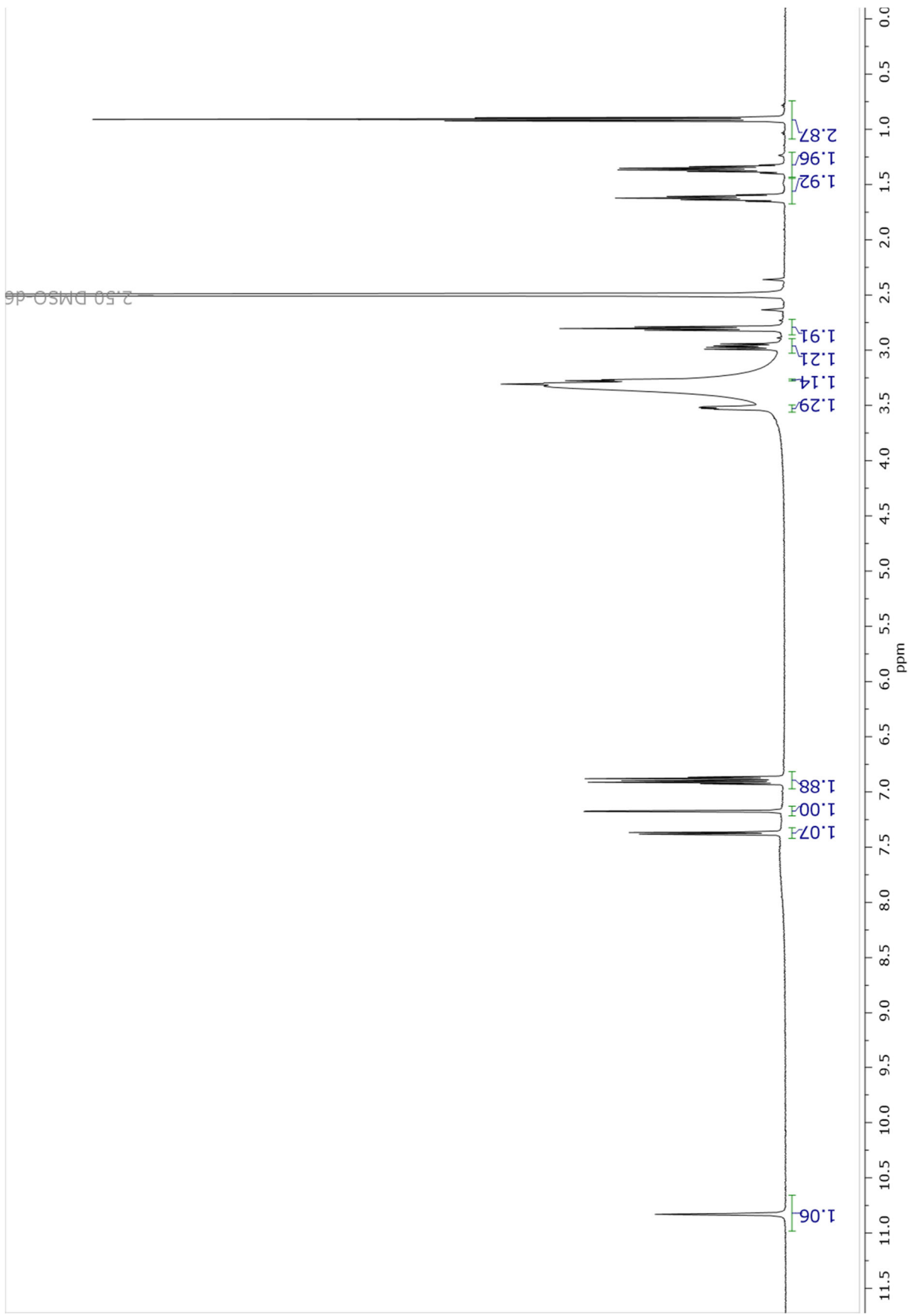
¹³C NMR (126 MHz, DMSO-*d*₆) δ [ppm] = 169.9 (COOH), 135.2 (C7a), 127.1 (C3a), 125.5 (C7), 123.7 (C2), 120.5 (C6), 118.5 (C5), 115.9 (C4), 109.6 (C3), 54.4 (Cα), 31.8 (C2'), 30.4 (C1'), 27.1 (Cβ), 22.1 (C3') 13.9 (C4')

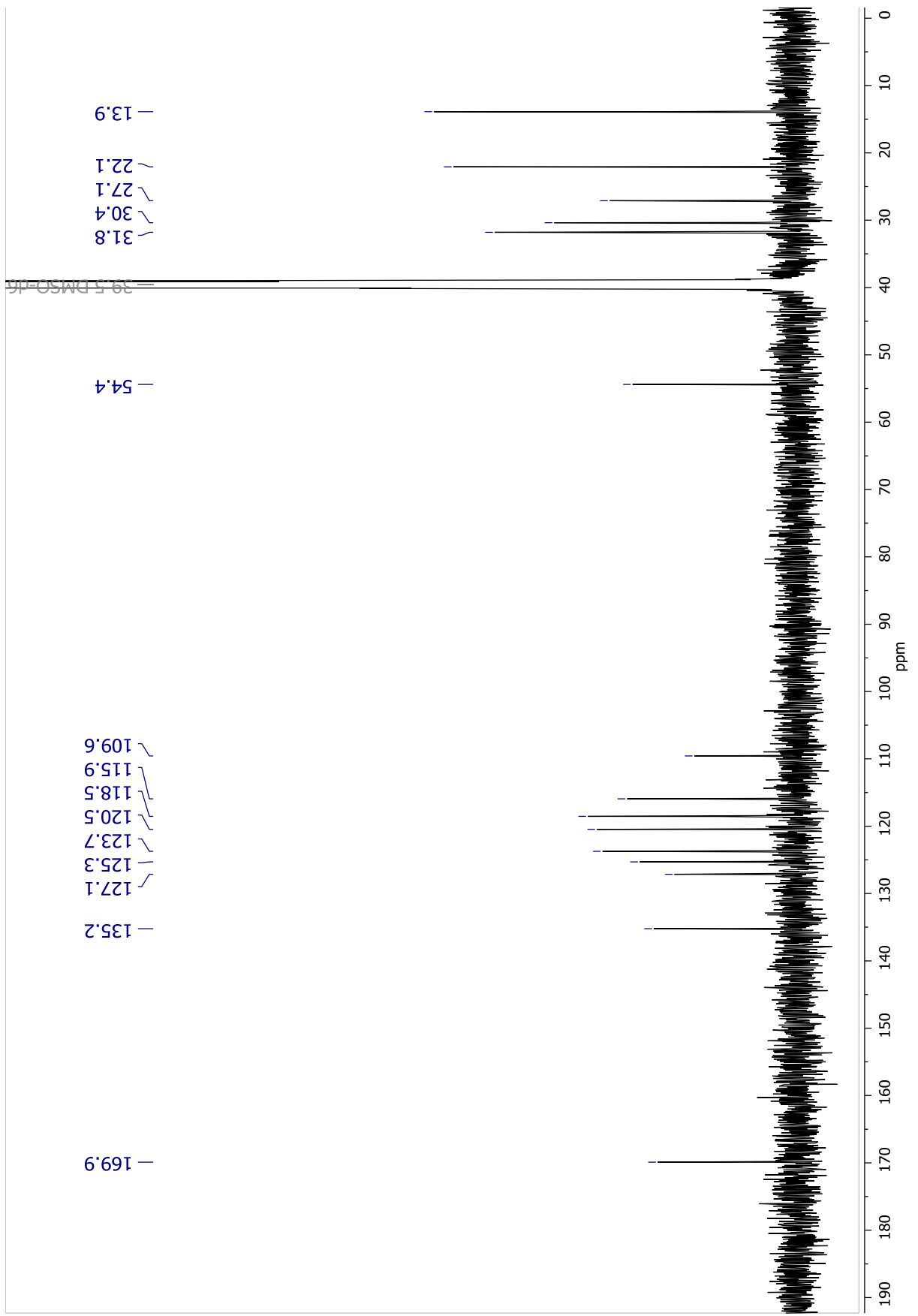
MS (ESI): found [*m/z*] = 261.09 [M+H]⁺, 244.06 [M-NH₂]⁺, calcd. [*m/z*] = 261.16 [M+H]⁺, 244.13 [M-NH₂]⁺.

Marfey's Derivatization: *t*_R (L-deriv.) = 12.7 min (>99 %)

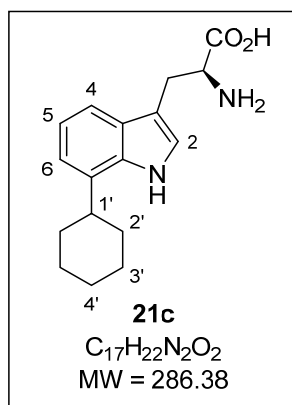
found [*m/z*] = 513.09 [M+H]⁺, calcd. [*m/z*] = 513.13 [M+H]⁺.







L-7-Cyclohexyl tryptophan (**21c**)



L-7-butyl tryptophan (**21c**) was synthesized according to GP5. Therefore, *N*^α-Boc-L-7-cyclohexyl tryptophan methyl ester (**18d**, 10.0 mg, 25 μmol) was suspended in aqueous HCl (5 M, 2 mL) and heated to 90 °C for 2 hours. The crude reaction mixture was purified by reversed phase HPLC, providing **21c** as a colorless solid (5.2 mg, 18 μmol, 73 %). The enantiomeric excess was determined using Marfey's Test (see GP3) giving an ee higher 99 %.

LC-MS: *t*_R = 6.3 min;

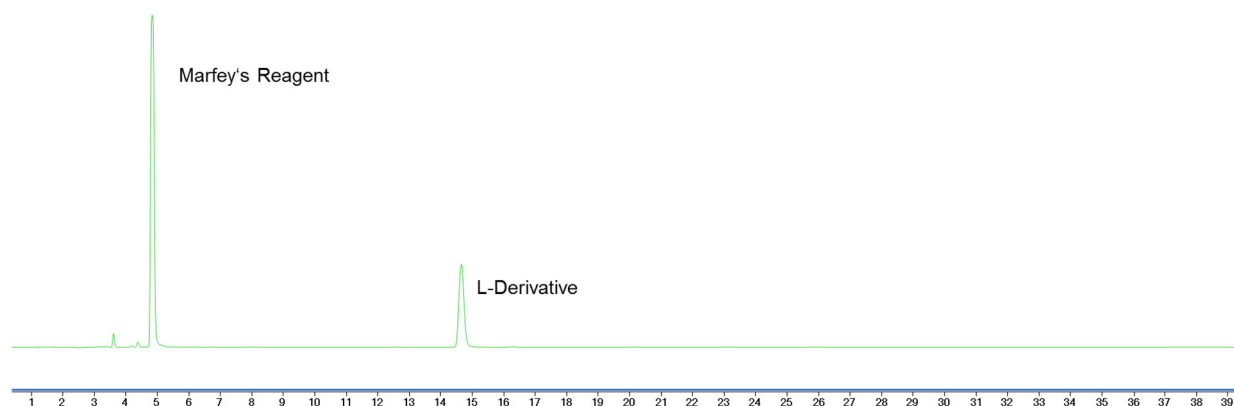
¹H NMR (500 MHz, DMSO-*d*₆) δ [ppm] = 10.95 (d, ³*J* = 2.3 Hz, 1H, Indole-NH), 7.95 (br s, 3H, NH₃⁺), 7.38 (d, ³*J* = 6.8 Hz, 1H, C4-H), 7.19 (d, ³*J* = 2.7 Hz, 1H, C2-H), 6.96 (dd, ³*J* = 6.9 Hz, ³*J* = 6.8 Hz, 1H, C5-H), 6.95 (d, ³*J* = 6.9 Hz, 1H, C6-H), 3.95 (dd, ³*J* = 7.4 Hz ³*J* = 5.1 Hz, 1H, Cα-H), 3.27 (dd, ²*J* = 15.2 Hz, ³*J* = 5.1 Hz, 1H, Cβ-H), 3.14 (dd, ²*J* = 15.0 Hz, ³*J* = 7.4 Hz, 1H, Cβ-H), 2.98 (m, 1H, C1'-H), 1.83 (m, 4H, C2'-H), 1.79 (m, 1H, C4'-H), 1.49 (m, 4H, C3'-H) 1.31 (m, 1H, C4'-H).

¹³C NMR (126 MHz, DMSO-*d*₆) δ [ppm] = 170.7 (COOH), 134.5 (C7a), 130.8 (C3a), 127.0 (C7), 124.2 (C2), 118.9 (C6), 117.4 (C5), 115.7 (C4), 107.8 (C3), 53.2 (Cα), 38.3 (C1'), 32.9 (C2'), 26.5 (C3'/C4'), 25.8 (Cβ).

MS (ESI): found [*m/z*] = 287.19 [M+H]⁺, 270.15 [M-NH₂]⁺, calcd. [*m/z*] = 287.18 [M+H]⁺, 270.15 [M-NH₂]⁺.

Marfey's Derivatization: *t*_R (L-deriv.) = 14.7 min (>99 %)

found [*m/z*] = 539.20 [M+H]⁺, calcd. [*m/z*] = 539.22 [M+H]⁺.



⁺ESI Scan (rt: 14.261-15.222 min, 59 scans) Frag=175.0V

