ChemBioChem

Supporting Information

Optochemical Control of Bacterial Gene Expression: Novel Photocaged Compounds for Different Promoter Systems

Fabian Hogenkamp⁺, Fabienne Hilgers⁺, Nora Lisa Bitzenhofer, Vera Ophoven, Mona Haase, Claus Bier, Dennis Binder, Karl-Erich Jaeger, Thomas Drepper,* and Jörg Pietruszka*

Table of Contents

| S1 Ba | cterial strains and plasmids | 1 |
|----------------|---|-----------|
| S2 Ge | eneral methods for chemical synthesis procedures | 2 |
| S3 Ex | perimental procedures for the preparation of compounds | 3 |
| S3.1 | Synthetic scheme for preparation of coumarin 7 | |
| S3.2 | Synthetic scheme for preparation of carbohydrates 8 and 11 | |
| S3.3 | Synthetic scheme for preparation of carbohydrates 10 and 12 | 12 |
| S3.4 | Synthetic scheme for preparation of photocaged arabinose 2b | |
| S3.5 | Synthetic scheme for preparation of photocaged IPTG 1c | 20 |
| S3.6 | Synthetic scheme for preparation of photocaged IPTG 1b and 1d | |
| S3.7 | Synthetic scheme for preparation of photocaged IPTG 1e and photocaged arabin | ose 2c 33 |
| S3.8 sodiui | Synthetic scheme for preparation of photocaged salicylic acid 22a and the corres m form 22b | |
| S4 Su | pporting data | 43 |
| S4.1 | UV-Vis spectra of compounds | 43 |
| S4.2 | Photon flux densities of light sources | 47 |
| S4.3 | Irradiation experiments | |
| S4.4 | HPLC-Traces | 53 |
| S4.5 | Stability measurements | 57 |
| S4.6 | ESI measurements | 59 |
| S4.7 | Toxicity of both the novel photocaged inducer variants and the light exposure | 61 |
| S4.8 | In vivo results of photocaged IPTG 1e | 62 |
| S4.9 | NMR spectra of compounds | 63 |

S1 Bacterial strains and plasmids

All bacterial strains, plasmids and oligonucleotides used in this study are listed in Table S1.

| Strains, plasmids, | Relevant features, description or sequences ^{a,b} | References |
|---|--|---------------------|
| oligonucleotides | | |
| | Strains | |
| E. coli DH5a | $F^{-}\Phi 80 lacZ\Delta M15 \Delta (lacZYA-argF) U169 recA1 endA1 hsdR17$ | [1] |
| | phoA supE44 thi-1 gyrA96 relA1 deoR | |
| E. coli Tuner (DE3) | F -ompT hsdS _B (r_{B} - m_{B} -) gal dcm lacY1(DE3) | Novagen, Merck KGaA |
| <i>E. coli</i> LMG194 | F– ΔlacX74 galE galK thi rpsL ΔphoA Δara714 leu::Tn10 | [2] |
| | Plasmids | |
| pRhotHi-2-lacI-eYFP | pBBR1-MCS-derivative, Km ^R , Cm ^R , pBBR22b-lacI, P _{T7} -lacO- | [3] |
| | MCS with NdeI XhoI inserted eyfp | |
| pM117-R45T-GFPmut3 | pMB1 replicon, xylS with R45T mutation, P _{m M1-17} with inserted | [4] |
| | gfpmut3 | |
| рМ-R45Т- | pMB1 replicon, xylS with R45T mutation, P _m with inserted | ^[5] and |
| GFPmut3 | gfpmut3 | this work |
| pBNTmcs(t)-Km | Km ^R , <i>nagR</i> , vector for <i>PnagAa</i> and <i>tac</i> RBS controlled | [6] |
| • • • • • | expression | |
| pBNTmcs-mCherry-Km | pBNTmcs(t)-Km derivative with <i>EcoRI/Xba</i> I inserted <i>mcherry</i> | This work |
| pBTBX-2 | pBBR1 replicon, Km ^R , <i>araC</i> , araBAD promoter | [7] |
| pBTBX-2-mCherry | pBBR1 replicon, Km ^R , <i>araC</i> , araBAD promoter with <i>tac</i> RBS | This work |
| r | and inserted <i>mcherry</i> | |
| | Oligonucleotides | |
| 1) XylS_SalI_fw | Binds upstream of SalI-site after xylS. | [4] |
| , | Sequence: 5'-GAGACACAACGTGGCTTTCC-3' | |
| 2) XylS_SacI_rev | Binds upstream of <i>Sac</i> I-site in front of <i>xylS</i> . | [4] |
| _, _, _, _, _, _, _, _, _, | Sequence: 5'- ATCGACTTGGCGCCTTTCTAC-3' | |
| 3) XylS_R45T_rev | Binds within <i>xylS</i> and inserts R45T point mutation. Sequence: | [4] |
| •) 11j10_101_101 | 5'- CAGGCACGCTGCACCACAGAATC-3' | |
| 4) XlyS_R45T_fw | Binds within <i>xylS</i> and inserts R45T point mutation. Sequence: | [4] |
| -,,~,~, | 5'- GATTCTGTGGTGCAGC <u>G</u> TGCCTG-3' | |
| 5) pBTBX_for | Binds at the 3' end of the pBTBX-2 plasmid. | This work |
| -, <u>F21211_</u> 101 | 5'- GTTCTAGAAAATTCGTCAACG -3' | THIS WORK |
| 6) pBTBX_rev | Binds at the 5' end of the araBAD promoter on the pBTBX-2 | This work |
| | plasmid. 5'- CATACCCGTTTTTTTGGGCTAG -3' | |
| 7) mCherry_for | Binds at the 5' end of the <i>mcherry</i> gene, inserts overhangs for | This work |
| // meneny_101 | In-Fusion® cloning. | THIS WOLK |
| | Sequence: 5'- GTTTTTTGGGCTAGCAGGAA | |
| | ACAGGAGGTACC-3' | |
| 8) mCherry_rev | Binds at the 3' end of mcherry gene, inserts overhangs for In- | This work |
| - | Fusion® cloning. | |
| | Sequence: 5'-GTTGACGAATTTTCTAGAACT TACTTGTACAGCTCG-3' | |

Table S1: Bacterial strains, plasmids and oligonucleotides used in this study

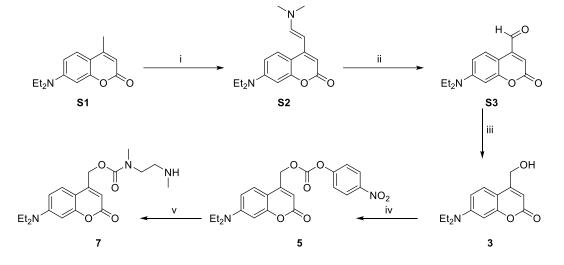
^a Underlined nucleotides indicate the point mutation used for XylS mutagenesis (AGG \rightarrow ACG).

^b Bold nucleotides indicate the inserted overhangs for In-Fusion® cloning.

S2 General methods for chemical synthesis procedures

All chemicals for synthesis were obtained from commercial suppliers and used without further purification unless stated otherwise. Solvents were reagent grade and were dried as well as purified by common methods. Thin-layer chromatography (TLC) was performed using pre-coated silica gel plates (Polygram[®] SIL G/UV, Macherey-Nagel) and components were visualized via oxidative staining or UV-light. Flash chromatography was performed on silica gel (Merck silica gel 60 (0.063-0.200 µm) and solvents for flash chromatography (petroleum ether/ethyl acetate/dichloromethane/n-pentane) were distilled prior to use. Optical rotation was determined at 20 °C on a Perkin Elmer Polarimeter 241 MC against sodium D-line and melting points were recorded using a Büchi melting point B-545 apparatus. The NMR spectra (¹H and ¹³C) were measured at 20 °C on a Bruker Avance/DRX 600 spectrometer in deuterated solvents (CDCl₃, DMSO- d_6 , acetone- d_6 , D₂O). The chemical shifts are given in ppm relative to the solvent (¹H: CDCl₃ = 7.26 ppm, ¹H: DMSO- d_6 = 3.31 ppm, ¹H: acetone- d_6 = 2.05 ppm, ¹H: $D_2O = 4.79 \text{ ppm} / {}^{13}C: CDCl_3 = 77.16 \text{ ppm}, {}^{13}C: DMSO-d_6 = 39.52 \text{ ppm}, {}^{13}C: acetone-d_6 = 29.84 \text{ ppm}).$ Signals were assigned by means of H-COSY-, HSQC- and HMBC-experiments and splitting patterns are reported as singlet (s), doublet (d), triplet (t), multiplet (m), and broad singlet (brs). The IR spectra were recorded with a Perkin Elmer SpectrumOne IR-spectrometer ATR (Waltham, USA). HRMS (ESI) spectra were recorded by the centrum of analytics of the Heinrich Heine University. UV-Vis absorption spectra were recorded on a Genesys 10S UV/VIS Spectrophotometer (Thermo Scientific) and uncaging experiments were performed in a quartz cuvette with the LUMOS 43® from Atlas Photonics at 375 nm, 405 nm and 430 nm. Light intensity was quantified using a Thermal Power Sensor (S302C, Thorlabs Inc, USA) and the decay was detected by a Jasco HPLC system [column: Hyperclone 5 μ ODS (C18) 120 (Phenomenex)] combined with an UV/Vis-detector.

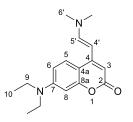
S3 Experimental procedures for the preparation of compounds



S3.1 Synthetic scheme for preparation of coumarin 7

Scheme S1: Synthetic scheme for preparation of coumarin 7. Reagents and conditions: i) DMF-DMA, DMF, reflux, 23 h; ii) NaIO₄, THF/H₂O (1:1), RT, 2 h; iii) NaBH₄, EtOH, 0 °C \rightarrow RT, 4 h; iv) 4-nitrophenyl chloroformate, DIPEA, CH₂Cl₂, RT, 19 h; v) *N*,*N*'-dimethylethylenediamine, CH₂Cl₂, 0 °C, 30 min.

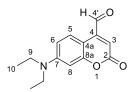
Synthesis of (*E*)-7-(Diethylamino)-4-[2-(dimethylamino)vinyl]-2*H*-chromen-2-one (S2)



Coumarin **S2** was synthesized using a procedure of Weinrich *et al.*^[8] Coumarin **S1** (15.0 g, 64.9 mmol) was dissolved in DMF (150 mL). After the addition of DMF-DMA (17.2 mL, 130 mmol) the reaction mixture was heated to reflux for 23 h. The reaction mixture was cooled to room temperature, diluted with CH₂Cl₂ and saturated NaHCO₃ solution was added. The organic phase was separated, and the aqueous phase was extracted with CH₂Cl₂. The combined organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure to yield a brown solid (18.4 g, 64.3 mmol, 99%). The compound **S2** was used in the following reactions without further purification. The spectroscopic data are consistent with previously reported literature values.^{[8] 1}H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.18 (t, ³J_{10.9} = 7.2 Hz, 6 H, 10-H), 2.98 (s, 6 H, 6'-H), 3.38 (q, ³J_{9,10} = 7.2 Hz, 4 H, 9-H), 5.21 (d, ³J_{4',5'} = 13 Hz, 1 H, 4'-H), 5.84 (s, 1 H, 3-H), 6.47 (d, ⁴J_{8,6} = 2.6 Hz, 1 H, 8-H), 6.54 (dd, ³J_{6,5} = 9.0 Hz, ⁴J_{6,8} = 2.6 Hz, 1 H, 6-H), 7.20 (d, ³J_{5',4'} = 13.0 Hz, 1 H, 5'-H), 7.51 (d, ³J_{5,6} = 9.0 Hz, 1 H, 5-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.6 (C-10), 41.0 (C-6'), 44.7 (C-9), 87.5 (C-4'), 93.5 (C-3), 98.2 (C-8), 108.0 (C-6), 108.2 (C-4a), 124.9 (C-5), 146.7 (C-5'), 150.2 (C-7), 152.4 (C-4), 156.5 (C-8a), 163.5

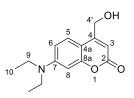
(C-2); $R_f = 0.33$ (CH₂Cl₂/EtOAc 70:30); IR (atr-film): \tilde{v} [cm⁻¹] = 1676, 1606, 1566, 1375, 1234, 1114, 1054, 974, 772, 625; MS (ESI, positive ion): m/z (%) = 287.3 (100) [M+H]⁺; m.p.: 183 °C.

Synthesis of 7-(Diethylamino)-2-oxo-2H-chromene-4-carbaldehyde (S3)



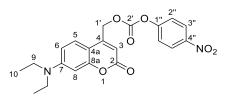
Coumarin **S3** was synthesized using a procedure of Weinrich *et al.*^[8] Coumarin **S2** (18.4 g, 64.3 mmol) was dissolved in THF/H₂O (1:1, 110 mL). After the addition of NaIO₄ (41.3 g, 193 mmol) the reaction mixture was stirred for 2 h at room temperature. The precipitate was filtered off, washed with ethyl acetate and volatile solvents were removed under reduced pressure and washed with saturated NaHCO₃ solution. The organic phase was separated and the aqueous layer was extracted with CH₂Cl₂. The combined organic phase was dried with anhydrous MgSO4 and concentrated under reduced pressure to yield a brown solid (15.7 g, 64.1 mmol, quant.). The compound S3 was used in the following reactions without further purification. The spectroscopic data are consistent with previously reported literature values.^[8] ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.22 (t, ³J_{10.9} = 7.1 Hz, 6 H, 10-H), 3.43 (q, ³J_{9.10} = 7.1 Hz, 4 H, 9-H), 6.45 (s, 1 H, 3-H), 6.53 (d, ${}^{4}J_{8,6} = 2.6$ Hz, 1 H, 8-H), 6.63 (dd, ${}^{3}J_{6,5} = 9.2$ Hz, ${}^{4}J_{6,8} =$ 2.6 Hz, 1 H, 6-H), 8.31 (d, ³*J*_{5,6} = 9.2 Hz, 1 H, 5-H), 10.03 (s, 1 H, 4'-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 45.0 (C-9), 97.9 (C-8), 104.0 (C-4a), 109.8 (C-6), 117.6 (C-3), 127.2 (C-5), 144.0 (C-8a), 151.0 (C-7), 157.5 (C-4), 162.0 (C-2), 192.6 (C-4'); $R_f = 0.29$ (PE/EtOAc 50:50); IR (atr-film): \tilde{v} [cm⁻¹] = 2972, 1703, 1607, 1582, 1518, 1424, 1376, 1354, 1267, 1228, 1196, 1142, 1111, 1077, 1053, 901, 822, 780, 732, 640, 475; MS (ESI, positive ion): m/z (%) = 278.3 (100) [M+CH₃OH+H]⁺; m.p.: 77.7 °C.

Synthesis of 7-(Diethylamino)-4-(hydroxymethyl)-2H-chromen-2-one (3)



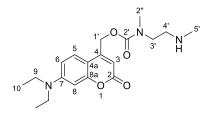
Coumarin **3** was synthesized using a modified procedure of Weinrich *et al.*^[8] Coumarin **S3** (5.00 g, 20.4 mmol) was dissolved in ethanol (405 mL) and cooled to 0 °C. After the addition of NaBH₄ at 0 °C the reaction mixture was stirred at room temperature for 4 h. The reaction was quenched by addition of 1 M HCl (150 mL) and diluted with H₂O (90.0 mL). The solution was extracted with CH₂Cl₂ (3×100 mL) and the combined organic phase was washed with H₂O (90 mL). Subsequently it was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 50:50) to yield a yellow solid (2.07 g, 8.37 mmol, 41%). The spectroscopic data are consistent with previously reported literature values.^{[8] 1}H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.20 (t, ³J_{10.9} = 7.1 Hz, 6 H, 10-H), 1.93 (t, ³J_{OH,4'} = 5.8 Hz, 1 H, 4'-OH), 3.41 (q, ³J_{9.10} = 7.1 Hz, 4 H, 9-H), 4.83 (dd, ³J_{4',OH} = 5.8 Hz, ⁴J_{4',3} = 1.3 Hz, 2 H, 4'-H), 6.51 (d, ⁴J_{8.6} = 2.6 Hz, 1 H, 8-H), 6.56 (dd, ³J_{6.5} = 9.0 Hz, ⁴J_{6.8} = 2.6 Hz, 1 H, 6-H), 7.32 (d, ³J_{5.5} = 9.0 Hz, 1 H, 5-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 44.8 (C-9), 60.8 (C-4'), 97.7 (C-8), 105.2 (C-3), 106.4 (C-4a), 108.7 (C-6), 124.5 (C-5), 150.6 (C-7), 155.6 (C-4), 156.1 (C-8a), 163.2 (C-2); R_f = 0.36 (PE/EtOAc 50:50); IR (atr-film): \hat{v} [cm⁻¹] = 2013, 2001, 1000; MS (ESI, positive ion): m/z (%) = 254.3 (100) [M+Li]⁺; m.p.: 139.9 °C.

Synthesis of [7-(Diethylamino)-2-oxo-2*H*-chromen-4-yl]methyl (4-nitrophenyl) carbonate (5)



Coumarin 5 was synthesized using modified procedures of Gao et al.^[9] and Fomina et al.^[10] Coumarin 3 (2.00 g, 8.09 mmol) was dissolved in dry CH₂Cl₂ (10.0 mL) under nitrogen atmosphere. N,N-Diisopropylethylamine (DIPEA) (2.82 mL, 16.2 mmol) was added and the reaction mixture was stirred for 15 min before 4-nitrophenyl chloroformate (3.26 g, 16.2 mmol) dissolved in dry CH₂Cl₂ (10 mL) was added over 2 h via a syringe pump. The reaction mixture was stirred for 19 h and diluted with CH₂Cl₂. It was washed with 1 M HCl (20 mL) and saturated NaHCO₃ solution (3×20 mL). The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (CH₂Cl₂/n-pentane 98:2) to yield a yellow solid (1.48 g, 3.60 mmol, 44%). The spectroscopic data are consistent with previously reported literature values.^{[9] 1}H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.22 (t, ${}^{3}J_{10.9}$ = 7.1 Hz, 6 H, 10-H), 3.43 (q, ${}^{3}J_{9.10}$ = 7.1 Hz, 4 H, 9-H), 5.40 (s, 2 H, 1'-H) 6.22 (s, 1 H, 3-H), 6.54 (d, ${}^{4}J_{8,6} = 2.6$ Hz, 1 H, 8-H), 6.61 (dd, ${}^{3}J_{6,5} = 3.6$ Hz, 1 H, 8-H), 6.61 (dd, {}^{3}J_{6,5} = 3.6 Hz, 1 H, 8-H), 6.61 (dd, {}^{3}J_{6,5} = 3.6 Hz, 1 H, 8-H), 6.61 (dd, {}^{3}J_{6,5} = 3.6 Hz, 1 H, 8-H), 6.61 (dd, {}^{3}J_{6,5} = 3.6 Hz, 1 H, 8-H), 6.61 (dd, {}^{3}J_{6,5} = 3.6 Hz, 1 H, 8-H), 6.61 (dd, {}^{3}J_{6,5} = 3.6 Hz, 1 H, 8-H), 6.61 (dd, {}^{3}J_{ = 9.0 Hz, ${}^{4}J_{6,8}$ = 2.6 Hz, 1 H, 6-H), 7.31 (d, ${}^{3}J_{5,6}$ = 9.0 Hz, 1 H, 5-H), 7.42 (d, ${}^{3}J_{2'',3''}$ = 8.6 Hz, 2 H, 2"-H), 8.30 (d, ${}^{3}J_{3'',2''} = 8.6$ Hz, 2 H, 3"-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 44.9 (C-9), 65.9 (C-1'), 98.0 (C-8), 105.7 (C-4a), 107.0 (C-3), 108.9 (C-6), 121.8 (C-2"), 124.4 (C-5), 125.5 (C-3"), 145.7 (C-4"), 147.9 (C-4), 151.0 (C-7), 152.3 (C-2'), 155.3 (C-1"), 156.5 (C-8a), 161.7 (C-2); R_f = 0.10 (CH₂Cl₂/*n*-pentane 98:2); IR (atr-film): \tilde{v} [cm⁻¹] = 2968, 1772, 1708, 1591, 1520, 1489, 1446, 1423, 1335, 1268, 1218, 1194, 1142, 1109, 1090, 1040, 986, 956, 856, 838, 816, 792, 750, 704, 679, 567, 528, 493, 467; HRMS (ESI): m/z calculated for $C_{21}H_{21}N_2O_7^+$ [M+H]⁺: 413.1343; found: 413.1340; m.p.: 159.9 °C.

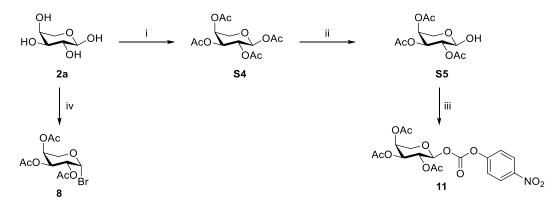
Synthesis of [7-(Diethylamino)-2-oxo-2*H*-chromen-4-yl]methyl methyl[2-(methylamino)ethyl]carbamate (7)



Coumarin 7 was synthesized using a modified procedure of Fomina *et al.*^[10] Coumarin 5 (50.0 mg, 0.12 mmol) was dissolved in CH₂Cl₂ (1.30 mL). It was added dropwise over 30 min to a solution of N,N'-dimethylethylenediamine (0.14 mL, 1.21 mmol) in CH₂Cl₂ (2.6 mL) at 0 °C. The reaction mixture was stirred for 30 min at 0 °C. The solvent was removed under reduced pressure, the residue was

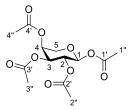
dissolved in ethyl acetate and washed subsequently with saturated NaHCO₃ and saturated NaCl solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (ethyl acetate/methanol/triethylamine 87:10:3) to yield a viscous dark yellow oil (27.0 mg, 74.7 µmol, 62%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.19 (t, ³*J*_{10.9} = 7.1 Hz, 6 H, 10-H), 2.42–2.51 (m, 4 H, 5'-H, NH), 2.80 (m_c, 2 H, 4'-H), 2.95–3.04 (m, 3 H, 2"-H), 3.39 (q, ³*J*_{9.10} = 7.1 Hz, 4 H, 9-H), 3.44–3.49 (m, 2 H, 3'-H), 5.23 (s, 2 H, 1'-H), 6.10 (s, 1 H, 3-H), 6.49 (d, ⁴*J*_{8.6} = 2.6 Hz, 1 H, 8-H), 6.55 (dd, ³*J*_{6.5} = 9.0 Hz, ⁴*J*_{6.8} = 2.6 Hz, 1 H, 6-H), 7.27–7.31 (m, 1 H, 5-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 34.8, 35.5 (C-2"), 36.1, 36.4 (C-5'), 44.8 (C-9), 48.6, 48.9 (C-3'), 49.3, 49.7 (C-4'), 62.5, 62.6 (C-1'), 97.9 (C-8), 105.8 (C-4a), 106.1, 106.2 (C-3), 108.7 (C-6), 124.4, 124.5 (C-5), 150.6, 150.7 (C-4, C-7), 155.6, 155.9 (C-2'), 156.3 (C-8a), 162.2 (C-2); R*f* = 0.15 (EtOAc/MeOH/NEt₃ 87:10:3); IR (atr-film): $\tilde{\nu}$ [cm⁻¹] = 2960, 2926, 2859, 1709, 1605, 1526, 1422, 1358, 1275, 1197, 1143, 1078, 821, 767; HRMS (ESI): m/z calculated for C₁₉H₂₈N₃O₄⁺ [M+H]⁺: 362.2074; found: 362.2077.

S3.2 Synthetic scheme for preparation of carbohydrates 8 and 11

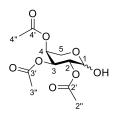


Scheme S2: Synthetic scheme for preparation of carbohydrates 8 and 11. Reagents and conditions: i) Ac₂O, DMAP, pyridine, 0 °C \rightarrow RT, 18 h; ii) AcOH, ethylenediamine, THF, RT, 24 h; iii) 4-nitrophenyl chloroformate, 2,6-lutidine, MeCN, RT, 18 h; iv) Ac₂O, HBr (33 wt%) in AcOH, RT, 3 h.

Synthesis of L-Arabinopyranose tetraacetate (S4)

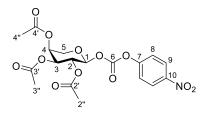


 α -L-Arabinopyranose tetraacetate (S4) was synthesized using a procedure of Wahler *et al.*^[11] α -L-Arabinopyranose (2a) (1.00 g, 6.66 mmol) was dissolved in dry pyridine (5.00 mL) and cooled to 0 °C. After the addition of acetic anhydride (5.04 mL, 53.3 mmol) and 4-dimethylaminopyridine (DMAP) (325 mg, 2.66 mmol) the reaction mixture was stirred for 18 h with the temperature slowly rising to room temperature. The reaction was quenched and diluted by addition of water and ethyl acetate. The organic phase was separated and washed with water as well as saturated NaCl solution. Subsequently it was dried with anhydrous MgSO₄ and concentrated under reduced pressure. Repeated coevaporation with toluene under reduced pressure yielded a colourless solid (1.77 g, 5.58 mmol, 84%). The spectroscopic data are consistent with previously reported literature values.^[11] ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 2.01, 2.01, 2.13, 2.14 (s, 12 H, 1"-H, 2"-H, 3"-H, 4"-H), 3.81 (dd, ² $J_{5a,5b}$ = 13.2 Hz, ${}^{3}J_{5a,4} = 2.0$ Hz, 1 H, 5-H_a), 4.05 (dd, ${}^{2}J_{5b,5a} = 13.2$ Hz, ${}^{3}J_{5a,4} = 1.5$ Hz, 1 H, 5-H_b), 5.28–5.39 (m, 3 H, 2-H, 3-H, 4-H), 6.33 (d, ${}^{3}J_{1,2} = 3.1$ Hz, 1 H, 1-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 20.7, 20.8, 21.0, 21.0 (C-1", C-2", C-3", C-4"), 62.9 (C-5), 66.8 (C-3 or C-4), 67.1 (C-2), 68.5 (C-3 or C-4), 90.3 (C-1), 169.2 (C-1'), 170.0, 170.2, 170.4 (C-2', C-3', C-4'); $R_f = 0.63$ (PE/EtOAc 1:1); IR (atr-film): \tilde{v} [cm⁻¹] = 1736, 1372, 1212, 1137, 1113, 1065, 1010, 942, 894, 755, 602, 552, 471; MS (ESI, positive ion): m/z $(\%) = 341.0 (100) [M+Na]^+, 357.0 (15) [M+K]^+; m.p.: 94.4 \text{ °C}; [\alpha]_D^{20} = 151.8 (c = 0.5, CHCl_3).$

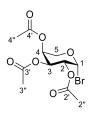


 $(2,3,4-\text{tri-}O-\text{acetyl})-\alpha,\beta-L-\text{arabinopyranose}$ (S5) was synthesized using a procedure of Duléry *et al.*^[12] Glacial acetic acid (1.01 mL, 17.6 mmol) was added dropwise to a solution of ethylenediamine (1.01 mL, 15.1 mmol) in THF (250 mL), which immediately lead to the formation of a precipitate. Larabinopyranose tetraacetate (S4) (4.00 g, 12.6 mmol) was added and the reaction mixture was stirred at room temperature for 24 h. After the addition of water, the reaction mixture was extracted with CH₂Cl₂. The organic phase was subsequently washed with 1 M HCl, saturated NaHCO₃ solution and water. Following this, the organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO_2 (petroleum ether/ethyl acetate 55:45) to yield (2,3,4-tri-O-acetyl)-α,β-L-arabinopyranose (S5) as a colorless oil (2.28 g, 8.26 mmol, 66%) in α : β ratio of 1:3. The spectroscopic data are consistent with previously reported literature values.^[13] α/β -Anomers: $R_f = 0.18$ (PE/EtOAc 1:1); IR (atr-film): \tilde{v} [cm⁻¹] = 3455, 1739, 1371, 1216, 1139, 1057, 1007, 936, 889, 765, 736, 603, 465; MS (ESI, positive ion): m/z (%) = 299.0 (100) $[M+Na]^+$; α -Anomer: ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 2.00 (s, 3 H, CH₃), 2.08 (s, 3 H, CH₃), 2.13 (s, 3 H, CH₃), 3.66 (dd, ${}^{2}J_{5a,5b} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 H, 5-H_a), 4.01 (dd, ${}^{2}J_{5b,5a} = 13.4$ Hz, ${}^{3}J_{5a,4} = 1.1$ Hz, 1 Hz, 13.4 Hz, ${}^{3}J_{5a,4} = 2.5$ Hz, 1 H, 5-H_b), 4.60 (d, ${}^{3}J_{1,2} = 6.7$ Hz, 1 H, 1-H), 5.03–5.09 (m, 2 H, 2-H and 3-H), 5.24–5.27 (m, 1 H, 4-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 20.7, 20.9, 21.0 (C-2", C-3", C-4"), 64.2 (C-5), 68.1 (C-4), 70.2 (C-2), 71.3 (C-3), 96.2 (C-1), 170.2, 170.5, 171.2 (C-2', C-3', C-4'); β-Anomer: ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 2.00 (s, 3 H, 4"-H), 2.08 (s, 3 H, 2"-H or 3"-H), 2.12 (s, 3 H, 2"-H or 3"-H), 3.59 (brs, 1 H, 1-OH), 3.68 (dd, ${}^{2}J_{5a,5b} = 13.1$ Hz, ${}^{3}J_{5a,4} = 2.3$ Hz, 1 H, 5-H_a), 4.18 $(dd, {}^{2}J_{5b,5a} = 13.1 \text{ Hz}, {}^{3}J_{5a,4} = 1.5 \text{ Hz}, 1 \text{ H}, 5 \text{-H}_{b}), 5.16 (dd, {}^{3}J_{3,2} = 10.5 \text{ Hz}, {}^{3}J_{3,4} = 3.4 \text{ Hz}, 1 \text{ H}, 3 \text{-H}), 5.33 \text{-}$ 5.36 (m, 1 H, 4-H), 5.38 (dd, ${}^{3}J_{2,3} = 10.5$ Hz, ${}^{3}J_{2,1} = 3.5$ Hz, 1 H, 2-H), 5.45 (d, ${}^{3}J_{1,2} = 3.5$ Hz, 1 H, 1-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 20.8 (C-4"), 20.9 (C-2" or C-3"), 21.0 (C-2" or C-3"), 60.4 (C-5), 67.0 (C-2), 68.8 (C-3), 69.2 (C-4), 91.0 (C-1), 170.3 (C-4'), 170.6 (C-2'), 170.6 (C-3').

Synthesis of 2,3,4-Tri-O-acetyl-1-O-(4-nitrophenyloxycarbonyl)- α -L-arabinopyranoside (11)

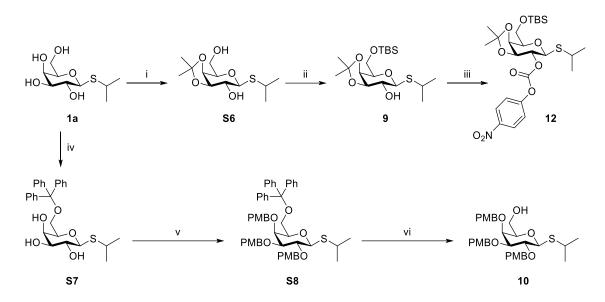


Carbohydrate 11 was synthesized using a modified procedure of André et al.^[14] The anomeric mixture of carbohydrate S5 (408 mg, 1.48 mmol) was dissolved in MeCN (49.0 mL) and cooled to 0 °C. After the addition of 4-nitrophenyl chloroformate (316 mg, 1.57 mmol) and 2,6-lutidine (0.18 mL, 1.57 mmol) the reaction mixture was stirred for 18 h at room temperature. The reaction mixture was quenched by addition of water and diluted with CH₂Cl₂. The organic phase was washed with water and saturated NaCl solution. Following this, the organic phase was dried with anhydrous MgSO4 and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 60:40) to mainly yield the α -anomer as a colorless solid (133 mg, 302 μ mol, 20%). Unreacted substrate was reisolated as an anomeric mixture with α : β ratio of 1:1.8. ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 2.10 (s, 3 H, 3"-H), 2.14 (s, 3 H, 4"-H), 2.14 (s, 3 H, 2"-H), 3.83 $(dd, {}^{2}J_{5a,5b} = 12.5 \text{ Hz}, {}^{3}J_{5a,4} = 2.8 \text{ Hz}, 1 \text{ H}, 5 \text{-H}_{a}), 4.18 (dd, {}^{2}J_{5b,5a} = 12.5 \text{ Hz}, {}^{3}J_{5a,4} = 5.1 \text{ Hz}, 1 \text{ H}, 5 \text{-H}_{b}),$ 5.19 (dd, ${}^{3}J_{3,2} = 7.8$ Hz, ${}^{3}J_{3,4} = 3.4$ Hz, 1 H, 3-H), 5.31–5.36 (m, 2 H, 2-H, 4-H), 5.69 (d, ${}^{3}J_{1,2} = 5.6$ Hz, 1 H, 1-H), 7.41 (d, ${}^{3}J_{8,9} = 9.1$ Hz, 2 H, 8-H), 8.29 (d, ${}^{3}J_{9,8} = 9.1$ Hz, 2 H, 9-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 20.8, 20.9, 21.0 (C-2", C-3", C-4"), 62.8 (C-5), 66.3 (C-4), 67.9 (C-2), 69.0 (C-3), 96.1 (C-1), 121.8 (C-8), 125.5 (C-9), 145.8 (C-10), 151.1 (C-6), 155.1 (C-7), 169.3 (C-2'), 170.0 (C-3'), 170.1 (C-4'); $R_f = 0.28$ (PE/EtOAc 70:30); IR (atr-film): \tilde{v} [cm⁻¹] = 1746, 1594, 1527, 1492, 1370, 1346, 1217, 1173, 1087, 1055, 965, 913, 860, 729, 599, 504; HRMS (ESI): m/z calculated for $C_{18}H_{23}N_2O_{12}^+$ $[M+NH_4]^+$: 459.1246; found: 459.1247; m.p.: 52–58 °C; $[\alpha]_D^{20} = -3$ (c = 1.0, CHCl₃).



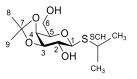
The carbohydrate **8** was synthesized using a procedure of Kartha *et al.*^[15] α -L-Arabinopyranose (**2a**) (1.00 g, 6.66 mmol) was dissolved in acetic anhydride (5.04 mL, 53.3 mmol) and stirred at room temperature. HBr solution 33 wt% in AcOH (1.50 mL, 8.57 mmol) was added to the suspension. After the solid was completely dissolved (1 h) additional HBr solution 33 wt% in AcOH (7.5 mL, 42.9 mmol) was added and the reaction mixture was stirred for additional 2 h. Subsequently the reaction mixture was concentrated under reduced pressure. Toluene (3×20 ml) was added and removed under reduced pressure. The crude product was recrystallized from Et₂O to yield a colorless solid (926 mg, 2.73 mmol, 41%). The spectroscopic data are consistent with previously reported literature values.^[16] ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 2.03 (s, 3 H, CH₃), 2.11 (s, 3 H, CH₃), 2.15 (s, 3 H, CH₃), 3.93 (dd, ²*J*_{5a,5b} = 13.3 Hz, ³*J*_{5a,4} = 1.7 Hz, 1 H, 5-Ha), 4.21 (d, ²*J*_{5b,5a} = 13.3 Hz, 1 H, 5-Hb), 5.09 (ddd, ³*J*_{2,3} = 11.8 Hz, ³*J*_{2,1} = 3.9 Hz, ³*J*_{2,4} = 1.6 Hz, 1 H, 2-H), 5.37–5.43 (m, 2 H, 3-H, 4-H), 6.70 (d, ³*J*_{1,2} = 3.9 Hz, 1 H, 1-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 20.8, 20.9, 21.0 (C-2", C-3", C-4"), 64.9 (C-5), 67.8, 68.0 (C-3, C-4), 68.1 (C-2), 89.8 (C-1), 169.9, 170.2, 170.2 (C-2', C-3', C-4').; R_f = 0.70 (PE/EtOAc 1:1); IR (atr-film): $\tilde{\nu}$ [cm⁻¹] = 1734, 1375, 1211, 1098, 1069, 1043, 992, 929, 891, 685, 577, 601, 539, 473; m.p.: 115.6 °C; [α]²_D² = 153.3 (c = 1.0, CHCl₃)

S3.3 Synthetic scheme for preparation of carbohydrates 10 and 12



Scheme S3: Synthetic scheme for preparation of carbohydrates 10 and 12. Reagents and conditions: i) 2,2-dimethoxypropane, CSA, acetone, RT, 8 h; ii) TBS-Cl, pyridine, RT, 20 h; iii) 4-nitrophenyl chloroformate, pyridine, RT, 20 h; iv) TrCl, DMAP, pyridine, RT, 18 h; v) PMB-Cl, NaH, DMF, RT, 18 h; vi) CSA, MeOH, CH₂Cl₂, 0 °C \rightarrow RT, 48 h.

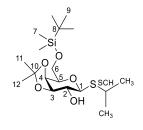
Synthesis of Isopropyl 3,4-O-(1-methylethylidene)-1-thio-β-D-galactopyranoside (S6)



Carbohydrate **S6** was synthesized using a modified procedure of Du *et al.*^[17] Isopropyl β -D-1-thiogalactopyranoside (**1a**) (5.00 g, 19.9 mmol) was dissolved in acetone (147 mL). After the addition of camphorsulfonic acid (CSA) (945 mg, 3.99 mmol) and 2,2-dimethoxypropane (3.66 mL, 29.9 mmol) the reaction mixture was stirred for 8 h at room temperature. The reaction mixture was quenched by addition of NaHCO₃ and the solvent was removed under reduced pressure. The residue was dissolved in CH₂Cl₂ and subsequently washed with NaHCO₃ solution. The aqueous phase was extracted with CH₂Cl₂ (3×) and the combined organic phases were washed with saturated NaCl solution. Following this, the organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 50:50) to yield a white solid (3.90 g, 14.0 mmol, 70%). ¹H-NMR (600 MHz, DMSO-*d*₆): δ [ppm] = 1.22 (d, ³*J*_{CH3,SCH} = 6.9 Hz, 3 H, CH₃), 1.23 (d, ³*J*_{CH3,SCH} = 6.9 Hz, 3 H, CH₃), 1.25 (s, 3 H, 7-H or 8-H), 1.38 (s, 3 H, 7-H or 8-H), 3.14 (septet, ³*J*_{SCH,CH3} = 6.9 Hz, 1 H, SCH), 3.23 (ddd, ³*J*_{2,1} = 9.9 Hz, ³*J*_{2,3} = 6.6 Hz, ³*J*_{2,2-OH} = 6.1 Hz, 1 H, 2-H), 3.51 (m, 2 H, 6-H), 3.73 (td, ³*J*_{5,6} = 6.3 Hz, ³*J*_{5,4} = 2.0 Hz, 1 H, 5-H), 3.92 (dd, ³*J*_{3,4} = 5.6 Hz, 1 H, 3-H), 4.14 (dd, ³*J*_{4,3} = 5.6 Hz, ³*J*_{4,5} = 2.0 Hz, 1 H, 4-H), 4.35 (d, ³*J*_{1,2} = 9.9 Hz, 1 H, 1-H), 4.73 (t, ³*J*_{6-OH,6} = 5.6 Hz, 1 H, 6-OH), 5.29 (d, ³*J*_{2-OH,2} = 6.1 Hz, 1 H, 2-OH);

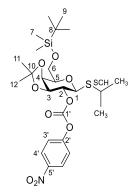
¹³C-NMR (151 MHz, DMSO-*d*₆): δ [ppm] = 23.7 (CH₃), 23.8 (CH₃), 26.4, 28.2 (C-7, C-8), 33.4 (SCH), 60.6 (C-6), 71.8 (C-2), 73.4 (C-4), 76.5 (C-5), 79.5 (C-3), 83.9 (C-1), 108.4 (C-7); R_f = 0.23 (PE/EtOAc 50:50); IR (atr-film): \tilde{v} [cm⁻¹] = 3301, 2990, 2924, 2864, 1460, 1369, 1239, 1215, 1141, 1073, 1020, 962, 872, 840, 725, 641, 570, 536, 504; HRMS (ESI): m/z calculated for C₁₂H₂₆NO₅S⁺ [M+NH₄]⁺: 296.1526; found: 296.1527; m.p.: 89.8 °C; [α]_D²⁰ = 3.4 (c = 1.0, CHCl₃)

Synthesis of Isopropyl 6-*O*-(*tert*-butyldimethylsilyl)-3,4-*O*-(1-methylethylidene)-1-thio- β -D-galactopyranoside (9)



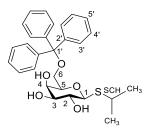
Carbohydrate 9 was synthesized using a modified procedure of Du et al.^[17] Carbohydrate S6 (1.00 g, 3.59 mmol) was dissolved in dry pyridine (20.0 mL) under nitrogen atmosphere. Tert-butyldimethylsilyl chloride (TBS-Cl) (1.14 g, 7.54 mmol) was added portion wise and the reaction mixture was stirred for 20 h at room temperature. After completion, the solvent was removed under reduced pressure. The residue was dissolved in CH₂Cl₂ and subsequently washed with water. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 85:15) to yield a white solid (1.05 g, 2.67 mmol, 74%). ¹H-NMR (600 MHz, DMSO- d_6): δ [ppm] = 0.04 (s, 3 H, 7-H), 0.05 (s, 3 H, 7-H), 0.86 (s, 9 H, 9-H), 1.22 (d, ${}^{3}J_{CH3,SCH} = 6.8$ Hz, 3 H, CH₃), 1.23 (d, ${}^{3}J_{CH3,SCH} = 6.8$ Hz, 3 H, CH₃), 1.24 (s, 3 H, 11-H or 12-H), 1.38 (s, 3 H, 11-H or 12-H), 3.13 (septet, ${}^{3}J_{SCH,CH3} = 6.8$ Hz, 1 H, SCH), 3.23 (dd, ${}^{3}J_{2,1} = 9.9$ Hz, ${}^{3}J_{2,3} = 7.0$ Hz, 1 H, 2-H), 3.66 (dd, ${}^{2}J_{6a,6b} = 10.4$ Hz, ${}^{3}J_{6a,5} = 7.2$ Hz, 1 H, 6-H_a), 3.71 (dd, ${}^{2}J_{6b,6a} = 10.4$ Hz, ${}^{3}J_{6b,5} = 5.4$ Hz, 1 H, 6-H_b), 3.81 (ddd, ${}^{3}J_{5,6a} = 7.2$ Hz, ${}^{3}J_{5,6b} = 5.4$ Hz, ${}^{3}J_{5,4} = 2.0$ Hz, 1 H, 5-H), 3.93 (dd, ${}^{3}J_{3,2} = 7.0$ Hz, ${}^{3}J_{3,4} = 5.4$ Hz, 1 H, 3-H), 4.14 (dd, ${}^{3}J_{4,3} = 5.4$ Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, ${}^{3}J_{1,2} = 5.4$ Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, ${}^{3}J_{1,2} = 5.4$ Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, ${}^{3}J_{1,2} = 5.4$ Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, ${}^{3}J_{1,2} = 5.4$ Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, ${}^{3}J_{1,2} = 5.4$ Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, ${}^{3}J_{1,2} = 5.4$ Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,2} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{1,5} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{4,5} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{4,5} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{4,5} = 5.4 Hz, ${}^{3}J_{4,5} = 2.0$ Hz, 1 H, 4-H), 4.37 (d, {}^{3}J_{4,5} = 5.4 9.9 Hz, 1 H, 1-H), 5.28 (brs, 1 H, 2-OH); ¹³C-NMR (151 MHz, DMSO- d_6): δ [ppm] = -5.6 (C-7), -5.4 (C-7), 17.9 (C-8), 23.7 (CH₃), 23.8 (CH₃), 25.6 (C-9), 26.3, 28.1 (C-11, C-12), 33.5 (SCH), 62.3 (C-6), 71.7 (C-2), 73.2 (C-4), 76.0 (C-5), 79.5 (C-3), 83.8 (C-1), 108.5 (C-10); $R_f = 0.62$ (PE/EtOAc 60:40); IR (atr-film): \tilde{v} [cm⁻¹] = 3407, 2954, 2929, 2857, 1472, 1385, 1358, 1223, 1241, 1141, 1110, 1075, 1025, 962, 876, 836, 772, 714, 582, 532, 477; HRMS (ESI): m/z calculated for C₁₈H₃₇O₅SSi⁺ [M+H]⁺: 393.2125; found: 393.2121; m.p.: 45.3 °C; $[\alpha]_D^{20} = -15.5$ (c = 1.0, CHCl₃)

Synthesis of Isopropyl 6-*O*-(*tert*-butyldimethylsilyl)-3,4-*O*-(1-methylethylidene)-2-*O*-(4-nitrophenyloxycarbonyl)-1-thio- β -D-galactopyranoside (12)



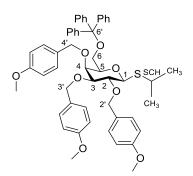
Carbohydrate 9 (1.00 g, 2.55 mmol) was dissolved in dry pyridine (20.0 mL) under nitrogen atmosphere. 4-nitrophenyl chloroformate (2.26 g, 11.2 mmol) was added portion wise and the reaction mixture was stirred for 20 h. The reaction mixture was diluted with CH₂Cl₂ and washed with water. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 85:15) to yield a white solid (1.23 g, 2.21 mmol, 87%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 0.08 (s, 6 H, 7-H), 0.89 (s, 9 H, 9-H), 1.33 (d, ${}^{3}J_{CH3,SCH} = 6.8$ Hz, 3 H, CH₃), 1.33 (d, ${}^{3}J_{CH3,SCH} = 6.8$ Hz, 3 H, CH₃), 1.36 (s, 3 H, 11-H or 12-H), 1.56 (s, 3 H, 11-H or 12-H), 3.21 (septet, ${}^{3}J_{SCH,CH3} = 6.8$ Hz, 1 H, SCH), 3.81–3.93 (m, 3 H, 5-H, 6-H), 4.27 (dd, ${}^{3}J_{3,2} = 7.3$ Hz, ${}^{3}J_{3,4} = 5.2$ Hz, 1 H, 3-H), 4.32 (dd, ${}^{3}J_{4,3} = 5.2$ Hz, ${}^{3}J_{4,5} = 1.4$ Hz, 1 H, 4-H), 4.53 (d, ${}^{3}J_{1,2} = 10.4$ Hz, 1 H, 1-H), 4.85 (dd, ${}^{3}J_{2,1} = 10.4$ Hz, ${}^{3}J_{2,3} = 7.3$ Hz, 1 H, 2-H), 7.37– 7.45 (m, 2 H, 2'-H), 8.21–8.31 (m, 2 H, 3'-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = -5.4 (C-7), -5.2 (C-7), 18.4 (C-8), 24.1 (CH₃), 24.2 (CH₃), 25.9 (C-9), 26.5, 28.0 (C-11, C-12), 35.8 (SCH), 62.1 (C-6), 73.6 (C-4), 76.8 (C-3), 77.3 (C-5), 77.8 (C-2), 82.4 (C-1), 110.8 (C-10), 122.0 (C-2'), 125.4 (C-3'), 145.6 (C-4'), 152.1 (C-2'), 155.7 (C-1'); $R_f = 0.73$ (PE/EtOAc 70:30); IR (atr-film): \tilde{v} [cm⁻¹] = 2954, 2929, 2853, 1768, 1617, 1594, 1519, 1492, 1464, 1374, 1345, 1307, 1247, 1215, 1164, 1117, 1072, 989, 957, 878, 836, 774, 751, 707, 674, 641, 573, 536, 499; HRMS (ESI): m/z calculated for C₂₅H₄₀NO₉SSi⁺ $[M+H]^+$: 558.2188; found: 558.2186; m.p.: 135.0 °C; $[\alpha]_D^{20} = 48.8$ (c = 1.0, CHCl₃)

Synthesis of Isopropyl 6-O-trityl-1-thio-β-D-galactopyranoside (S7)



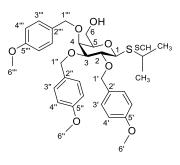
Carbohydrate S7 was synthesized using a modified procedure of Du et al.^[18] Isopropyl β-D-1thiogalactopyranoside (1a) (5.00 g, 21.0 mmol) was dissolved in dry pyridine (5 mL) under nitrogen atmosphere. Trityl chloride (TrCl) (11.7 g, 42.0 mmol) and 4-dimethylaminopyridine (DMAP) (256 mg, 2.10 mmol) were added and the reaction mixture was stirred for 18 h at room temperature. The reaction mixture was diluted with ethyl acetate and subsequently washed with water $(3\times)$, saturated NaHCO₃ solution $(1\times)$ and saturated NaCl solution $(1\times)$. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 50:50) to yield a white solid (7.68 g, 16.0 mmol, 76%). ¹H-NMR (600 MHz, DMSO- d_6): δ [ppm] = 1.28 (d, ³ $J_{CH3,SCH}$ = 6.8 Hz, 3 H, CH₃), 1.31 (d, ${}^{3}J_{CH3.SCH} = 6.8$ Hz, 3 H, CH₃), 2.93 (dd, ${}^{2}J_{6a,6b} = 9.7$ Hz, ${}^{3}J_{6a,5} = 3.7$ Hz, 1 H, 6-H_a), 3.20–3.28 (m, 2 H, SCH, 6-H_b), 3.28–3.32 (m, 2 H, 2-H, 3-H), 3.56 (brs, 1 H, 4-H), 3.61 (dd, ${}^{3}J = 7.7$ Hz, ${}^{3}J_{5.6a} = 3.7$ Hz, 1 H, 5-H), 4.36–4.39 (m_c, 1 H, 1-H), 4.42 (d, ${}^{3}J_{4-OH,4} = 4.5$ Hz, 1 H, 4-OH), 4.82 (d, ${}^{3}J_{3-OH,3} = 4.6$ Hz, 1 H, 3-OH), 4.92 (d, ${}^{3}J_{2-OH,2} = 5.2$ Hz, 1 H, 2-OH), 7.25 (t, ${}^{3}J_{5',4'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 3 H, 5'-H), 7.32 (t, ${}^{3}J_{4',5'} = 7.2$ Hz, 7 Hz, 7.6 Hz, 6 H, 4'-H), 7.41 (d, ${}^{3}J_{3',4'}$ = 7.4 Hz, 6 H, 3'-H); 13 C-NMR (151 MHz, DMSO- d_6): δ [ppm] = 23.7 (CH₃), 23.8 (CH₃), 33.6 (SCH), 64.0 (C-6), 69.3 (C-4), 69.8 (C-2), 74.5 (C-3), 77.6 (C-5), 84.8 (C-1), 85.7 (C-1'), 126.9 (C-5'), 127.8 (C-4'), 128.3 (C-3'), 143.9 (C-2'); R_f = 0.11 (PE/EtOAc 50:50); IR (atrfilm): \tilde{v} [cm⁻¹] = 3400, 3062, 2925, 2870, 1736, 1594, 1490, 1448, 1368, 1240, 1152, 1060, 1030, 899, 872, 833, 766, 746, 701, 650, 632, 584; HRMS (ESI): m/z calculated for C₂₈H₃₆NO₅S⁺ [M+NH₄]⁺: 498.2309; found: 498.2311; m.p.: 68.8 °C; $[\alpha]_D^{20} = -23.5$ (c = 1.0, CHCl₃)

SynthesisofIsopropylgalactopyranoside (S8)



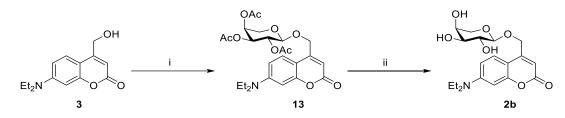
Carbohydrate S8 was synthesized using modified procedures of Ruda et al.^[19] Carbohydrate S7 (100 mg, 208 µmol) and 4-methoxybenzyl chloride (PMB-Cl) (126 µL, 936 mmol) was dissolved in dry DMF (2.00 mL) under nitrogen atmosphere. The mixture was added dropwise over 30 min via a syringe pump to NaH (60%, 37.5 mg, 936 µmol) in dry DMF (2.00 mL) and was stirred for 18 h at room temperature. After completion, the reaction mixture was cooled to 0 °C and quenched by the addition of MeOH. Ethyl acetate was added, and the mixture was washed with water $(3\times)$ and saturated NaCl solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 90:10 to ethyl acetate/methanol 90:10) to yield a white solid (129 mg, 153 µmol, 74%). ¹H-NMR (600 MHz, DMSO d_6): δ [ppm] = 1.27 (d, ${}^{3}J_{CH3,SCH} = 6.8$ Hz, 3 H, CH₃), 1.30 (d, ${}^{3}J_{CH3,SCH} = 6.8$ Hz, 3 H, CH₃), 2.82 (dd, ${}^{2}J_{6a,6b} = 9.5$ Hz, ${}^{3}J_{6a,5} = 4.5$ Hz, 1 H, 6-H_a), 3.17–3.28 (m, 2 H, 6-H_b, SCH), 3.40 (t, ${}^{3}J_{2,3} = 9.4$ Hz, 1 H, 2-H), 3.63 (dd, ${}^{3}J_{3,2} = 9.4$ Hz, ${}^{3}J_{3,4} = 2.9$ Hz, 1 H, 3-H), 3.70–3.74 (m, 1 H, 5-H), 3.72 (s, 3 H, OCH₃), 3.73 (s, 3 H, OCH₃), 3.74 (s, 3 H, OCH₃), 3.88 (d, ${}^{3}J_{4,3} = 2.9$ Hz, 1 H, 4-H), 4.23 (d, ${}^{2}J_{4'a,4'b} = 11.0$ Hz, 1 H, 4-H_a), 4.53–4.62 (m, 5 H, 1-H, 2'-H, 3'-H_a, 4'-H_b), 4.67 (d, ${}^{2}J_{3^{+}b,3^{+}a} = 11.4$ Hz, 1 H, 3'-H_b), 6.72–6.79 (m, 2 H, arom. H), 6.85–6.92 (m, 6 H, arom. H), 7.20–7.25 (m, 2 H, arom. H), 7.25–7.41 (m, 17 H, arom. H); ¹³C-NMR (151 MHz, DMSO- d_6): δ [ppm] = 23.7 (CH₃), 23.7 (CH₃), 34.3 (SCH), 55.0 (OCH₃), 63.6 (C-6), 71.2 (C-3'), 73.3 (C-4'), 73.9 (C-4), 74.1 (C-2'), 76.7 (C-5), 77.8 (C-2), 82.8 (C-3), 83.3 (C-1), 85.9 (C-1'), 113.4 (arom. C), 113.6 (arom. C), 127.0 (arom. C), 127.9 (arom. C), 128.2 (arom. C), 129.3 (arom. C), 129.3 (arom. C), 129.4 (arom. C), 130.4 (arom. C), 130.5 (arom. C), 130.6 (arom. C), 143.7 (arom. C), 158.6 (arom. C), 158.7 (arom. C), 158.7 (arom. C); $R_f = 0.05$ (PE/EtOAc 90:10); IR (atr-film): \tilde{v} [cm⁻¹] = 3038, 2931, 2835, 1612, 1586, 1512, 1449, 1360, 1302, 1245, 1173, 1152, 1078, 1031, 899, 820, 747, 705, 650, 632, 600, 515; HRMS (ESI): m/z calculated for C₅₂H₆₀NO₈S⁺ $[M+NH_4]^+$: 858.4034; found: 858.4031; m.p.: 53.4 °C; $[\alpha]_D^{20} = 6.4$ (c = 1.0, CHCl₃)

Synthesis of Isopropyl 2,3,4-tri-O-(4-methoxybenzyl)-1-thio-β-D-galactopyranoside (10)



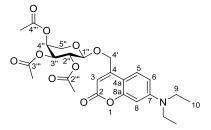
Carbohydrate 10 was synthesized using modified procedures of Ruda et al.^[19] Carbohydrate S8 (1.28 g, 1.52 mmol) was dissolved in dry CH₂Cl₂ (63 mL) under nitrogen atmosphere and cooled to 0 °C. Camphorsulfonic acid (CSA) (37.6 mg, 167 µmol) in methanol (6.3 mL) was added and the reaction mixture was allowed to warm to room temperature. The reaction mixture was stirred for 48 h. After completion, the solvent was evaporated under reduced pressure. The residue was purified by flashcolumn chromatography on SiO₂ (petroleum ether/ethyl acetate 70:30 to petroleum ether/ethyl acetate 30:70) to yield a white solid (662 mg, 1.11 mmol, 73%). The spectroscopic data are consistent with previously reported literature values.^[20] ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.32 (d, ³J_{CH3,SCH} = 6.8 Hz, 3 H, CH₃), 1.33 (d, ${}^{3}J_{CH3,SCH} = 6.8$ Hz, 3 H, CH₃), 3.21 (septet, ${}^{3}J_{SCH,CH3} = 6.8$ Hz, 1 H, SCH), 3.34–3.38 (m, 1 H, 5-H), 3.40 (ddd, ${}^{2}J_{6a,6b} = 11.1$ Hz, ${}^{3}J_{6a,5} = 8.2$ Hz, ${}^{3}J_{6a,6-OH} = 5.1$ Hz, 1 H, 6-H_a), 3.53 (dd, ${}^{3}J_{3,2} = 9.3$ Hz, ${}^{3}J_{3,4} = 2.8$ Hz, 1 H, 3-H), 3.70–3.79 (m, 3 H, 2-H, 4-H, 6-H_b), 3.80 (s, 6 H, OCH₃), 3.82 (s, 3 H, OCH₃), 4.46 (d, ${}^{3}J_{1,2} = 9.7$ Hz, 1 H, 1-H), 4.59 (d, ${}^{2}J_{1''a,1''b} = 11.7$ Hz, 1 H, 1'''-H_a), 4.68 (d, ${}^{2}J_{1''a,1''b} = 11.4$ Hz, 1 H, 1"-H_a), 4.71 (d, ${}^{2}J_{1'a,1'b} = 9.8$ Hz, 1 H, 1'-H_a), 4.72 (d, ${}^{2}J_{1''b,1''a} = 11.4$ Hz, 1 H, 1"-H_b), 4.82 (d, ${}^{2}J_{1'b,1'a} = 9.8$ Hz, 1 H, 1'-H_b), 4.87 (d, ${}^{2}J_{1''b,1''a} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1"'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1'-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1''-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1''-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1''-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1''-H_b), 6.86 (d, ${}^{3}J_{4',3'} = 11.7$ Hz, 1 H, 1''-H_b), 6.86 (d, {}^{3}J_{4',3'} = 11.7 Hz, 1 H, 1''-H_b), 6.86 (d, {}^{3}J_{4',3'} = 11.7 Hz, 1 H, 1''-H_b), 6.86 (d, {}^{3}J_{4',3'} = 11.7 Hz, 1 H, 1''-H_b), 6.86 (d, {}^{3}J_{4',3'} = 11.7 Hz, 1 H, 1''-H_b), 6.86 (d, {}^{3}J_{4',3'} = 11.7 Hz, 1 H, 1''-H_b), 6.86 (d, {}^{3}J_{4',3'} = 11.7 Hz, 1 H, 1''-H_b), 6.86 (d, {}^{3}J_{4',3'} = 11.7 8.5 Hz, 2 H, 4'-H), 6.86 (d, ${}^{3}J_{4'',3''}$ = 8.5 Hz, 2 H, 4^{''}-H), 6.89 (d, ${}^{3}J_{4'',3''}$ = 8.5 Hz, 2 H, 4^{''}-H), 7.25 (d, ${}^{3}J_{3'',4''}$ = 8.5 Hz, 2 H, 3^{*m*}-H), 7.31 (d, ${}^{3}J_{3^{n},4^{n}}$ = 8.5 Hz, 2 H, 3^{*n*}-H), 7.33 (d, ${}^{3}J_{3^{\prime},4^{\prime}}$ = 8.5 Hz, 2 H, 3^{*i*}-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 24.1 (CH₃), 35.4 (CH₃), 55.4 (SCH), 55.4, 55.4 (C-6', C-6'', C-6''), 62.4 (C-6), 72.6 (C-4), 73.0 (C-1"), 73.6 (C-1"), 75.6 (C-1'), 78.6 (C-2), 78.7 (C-5), 84.1 (C-3), 85.2 (C-1), 113.8, 113.9, 114.0 (C-4', C-4", C-4"), 129.4 (C-3"), 130.1 (C-3"'), 130.2 (C-3'), 130.5, 130.6, 130.7 (C-2', C-2'', C-2'''), 159.4, 159.4, 159.5 (C-5', C-5'', C-5''').; $R_f = 0.09$ (PE/EtOAc 70:30); IR (atr-film): \tilde{v} [cm⁻¹] = 3355, 2948, 2910, 2864, 2841, 1614, 1585, 1513, 1462, 1360, 1302, 1249, 1169, 1136, 1097, 1081, 1050, 1028, 996, 875, 819, 776, 700, 636, 603, 568, 515; HRMS (ESI): m/z calculated for $C_{33}H_{46}NO_8S^+[M+NH_4]^+: 616.2939;$ found: 616.2945; m.p.: 137.2 °C; $[\alpha]_D^{20} = -12.7$ (c = 1.0, CHCl₃).

S3.4 Synthetic scheme for preparation of photocaged arabinose 2b



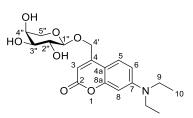
Scheme S4: Synthetic scheme for preparation of photocaged arabinose **2b**. Reagents and conditions: i) **8**, AgOTf, CH₂Cl₂, RT, 22 h; ii) NH₃ in MeOH (7 M), MeOH, RT.

Synthesis of 2,3,4-Tri-*O*-acetyl-1-*O*-{[7-(diethylamino)-2-oxo-2*H*-chromen-4-yl]methyl}- α -L-arabinopyranose (13)



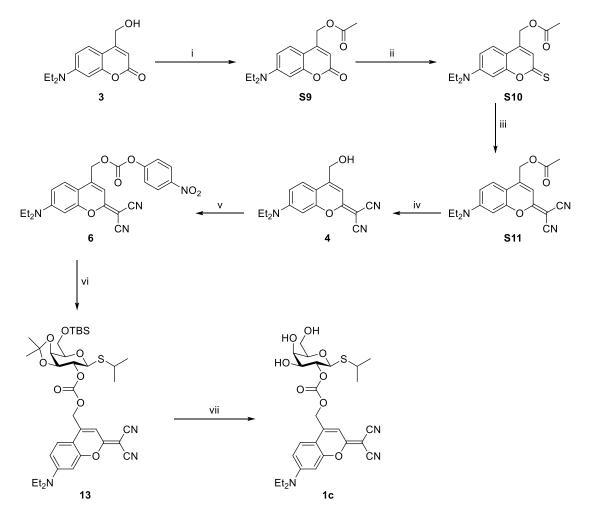
Coumarin 13 was synthesized using a modified procedure of Binder et al.^[16] A Schlenk tube was charged with 500 mg molecular sieve (5 Å) and carbohydrate 8 (200 mg, 0.59 mmol) dissolved in dry CH₂Cl₂ (10 mL) under nitrogen atmosphere. A second Schlenk tube was charged with 500 mg molecular sieve (5 Å) and coumarin **3** (438 mg, 1.77 mmol) dissolved in dry CH₂Cl₂ (10 mL) under nitrogen atmosphere. After stirring for 1 h the dissolved carbohydrate 8 was added to the coumarin solution. Silver triflate (182 mg, 708 µmol) was added and the reaction was stirred for 21 h at room temperature in the dark. The reaction mixture was filtered over celite to remove the molecular sieve. The solvent was removed under reduced pressure and the residue was purified by flash-column chromatography on aluminium oxide (neutral) (petroleum ether/ethyl acetate 50:50) to yield a yellow solid (176 mg, 348 µmol, 59%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.18 (t, ³J_{10,9} = 7.1 Hz, 6 H, 10-H), 2.00 (s, 3 H, CH₃), 2.04 (s, 3 H, CH₃), 2.13 (s, 3 H, CH₃), 3.39 (q, ${}^{3}J_{9,10} = 7.1$ Hz, 4 H, 9-H), 3.65 (dd, ${}^{2}J_{5''a,5''b} = 12.8$ Hz, ${}^{3}J_{5''a,4''} = 12.8$ 2.1 Hz, 1 H, 5"-H_a), 4.04 (dd, ${}^{2}J_{5"b,5"a} = 12.8$ Hz, ${}^{3}J_{5"b,4"} = 4.0$ Hz, 1 H, 5"-H_b), 4.58 (d, ${}^{3}J_{1",2"} = 6.4$ Hz, 1 H, 1"-H), 4.67 (dd, ${}^{2}J_{4'a,4'b} = 14.7$ Hz, ${}^{4}J_{4'a,3} = 1.3$ Hz, 1 H, 4'-H_a), 4.93 (dd, ${}^{2}J_{4'b,4'a} = 14.7$ Hz, ${}^{4}J_{4'b,3} = 14.7$ 1.3 Hz, 1 H, 4'-H_b), 5.07 (dd, ${}^{3}J_{3'',2''} = 8.8$ Hz, ${}^{3}J_{3'',4''} = 3.5$ Hz, 1 H, 3"-H), 5.24 (dd, ${}^{3}J_{2'',3''} = 8.8$ Hz, ${}^{3}J_{2'',1''}$ = 6.4 Hz, 1 H, 2"-H), 5.27 (ddd, ${}^{3}J_{4",5"b}$ = 4.0 Hz, ${}^{3}J_{4",3"}$ = 3.5 Hz, ${}^{3}J_{4",5"a}$ = 2.1 Hz, 1 H, 4"-H), 6.17 (t, ${}^{4}J_{3,4'} = 1.3$ Hz, 1 H, 3-H), 6.51 (d, ${}^{4}J_{8,6} = 2.6$ Hz, 1 H, 8-H), 6.56 (dd, ${}^{3}J_{6,5} = 8.9$ Hz, ${}^{4}J_{6,8} = 2.6$ Hz, 1 H, 6-H), 7.27 (d, ${}^{3}J_{5.6} = 8.9$ Hz, 1 H, 5-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 20.8, 20.8, 21.0 (CH₃), 45.0 (C-9), 62.8 (C-5"), 65.9 (C-4'), 67.3 (C-4"), 69.1 (C-2"), 69.8 (C-3"), 98.1 (C-8), 99.6 (C-1"), 106.5 (C-4a), 107.1 (C-3), 108.8 (C-6), 124.8 (C-5), 150.5 (C-4), 150.6 (C-7), 156.4 (C-8a), 162.0 (C-2), 169.5 (C-2"), 170.2 (C-3"), 170.4 (C-4"); $R_f = 0.51$ (PE/EtOAc 50:50); IR (atr-film): \tilde{v} [cm⁻ ¹] = 2972, 1744, 1710, 1602, 1527, 1419, 1358, 1217, 1138, 1062, 1022, 601, 530; HRMS (ESI): m/z calculated for $C_{25}H_{32}NO_{10}^+$ [M+NH₄]⁺: 506.2021; found: 506.2028; m.p.: 113.6 °C; $[\alpha]_D^{20} = 36.4$ (c = 0.25, CHCl₃); UV-Vis (MeOH): λ_{max} (ϵ) = 246 nm (11206 dm³ mol⁻¹ cm⁻¹), 379 (16536).

Synthesis of {[7-(Diethylamino)-2-oxo-2*H*-chromen-4-yl]methyl}- α -L-arabinopyranose (2b)



Photocaged arabinose 2b was synthesized using modified procedures of Bier et al.^[21] Coumarin 13 (135 mg, 267 µmol) was dissolved in MeOH (0.60 mL) and stirred at room temperature in the dark. Ammonia in MeOH (7 M, 230 µL, 1.61 mmol) was added and the reaction mixture was stirred until complete conversion. The solvent was evaporated under reduced pressure to yield a yellow solid (100 mg, 264 μ mol, quant.). ¹H-NMR (600 MHz, Aceton- d_6): δ [ppm] = 1.20 (t, ³ $J_{10,9}$ = 7.1 Hz, 6 H, 10-H), 3.50 (q, ${}^{3}J_{9,10} = 7.1$ Hz, 4 H, 9-H), 3.59 (dd, ${}^{2}J_{5''a,5''b} = 12.4$ Hz, ${}^{3}J_{5''a,4''} = 1.9$ Hz, 1 H, 5''-H_a), 3.59-3.64 (m, 1 H, 3"-H), 3.68 (d, ${}^{3}J_{4"-OH,4"} = 4.3$ Hz, 1 H, 4"-OH), 3.71 (ddd, ${}^{3}J_{2",3"} = 8.4$ Hz, ${}^{3}J_{2",1"} = 6.7$ Hz, ${}^{3}J_{2'',2''-OH} = 4.1$ Hz, 1 H, 2''-H), 3.82–3.86 (m, 1 H, 4''-H), 3.91 (dd, ${}^{2}J_{5''b,5''a} = 12.4$ Hz, ${}^{3}J_{5''b,4''} = 3.3$ Hz, 1 H, 5"-H_b), 3.98 (d, ${}^{3}J_{3"-OH,3"} = 5.5$ Hz, 1 H, 3"-OH), 4.41 (d, ${}^{3}J_{1",2"} = 6.7$ Hz, 1 H, 1"-H), 4.47 (d, ${}^{3}J_{2''-OH,2''} = 4.1$ Hz, 1 H, 2"-OH), 4.76 (dd, ${}^{2}J_{4'a,4'b} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, ${}^{2}J_{4'b,4'a} = 1.4$ Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 Hz, 1 H, 4'-H_a), 4.98 (dd, {}^{2}J_{4'b,4'a} = 1.4 = 15.2 Hz, ${}^{4}J_{4^{+}b,3}$ = 1.4 Hz, 1 H, 4'-H_b), 6.26 (t, ${}^{4}J_{3,4'}$ = 1.4 Hz, 1 H, 3-H), 6.49 (d, ${}^{4}J_{8,6}$ = 2.6 Hz, 1 H, 8-H), 6.69 (dd, ${}^{3}J_{6,5} = 9.0$ Hz, ${}^{4}J_{6,8} = 2.6$ Hz, 1 H, 6-H), 7.47 (d, ${}^{3}J_{5,6} = 9.0$ Hz, 1 H, 5-H); 13 C-NMR $(151 \text{ MHz, Aceton-}d_6): \delta \text{ [ppm]} = 12.7 \text{ (C-10)}, 45.1 \text{ (C-9)}, 66.3 \text{ (C-5'')}, 66.4 \text{ (C-4')}, 68.9 \text{ (C-4'')}, 72.3 \text{ (C-10)}, 66.3 \text{ (C-5'')}, 66.4 \text{ (C-4')}, 68.9 \text{ (C-4'')}, 72.3 \text{ (C-10)}, 66.3 \text{ (C-5'')}, 66.4 \text{ (C-4')}, 68.9 \text{ (C-4'')}, 72.3 \text{ (C-10)}, 72.3 \text{$ (C-2"), 73.9 (C-3"), 98.0 (C-8), 104.1 (C-1"), 106.9 (C-3), 107.1 (C-4a), 109.4 (C-6), 126.0 (C-5), 151.5 (C-7), 152.9 (C-4), 157.2 (C-8a), 161.8 (C-2); $R_f = 0.33$ (PE/EtOAc 50:50); IR (atr-film): \tilde{v} [cm⁻¹] = 3443, 2954, 2921, 2847, 1706, 1623, 1527, 1441, 1360, 1328, 1138, 1088, 1011, 855, 824, 794, 763, 610, 511; HRMS (ESI): m/z calculated for C₁₉H₂₆NO₇⁺ [M+H]⁺: 380.1704; found: 380.1697; m.p.: 129– 130 °C; $[\alpha]_D^{20} = -18.6$ (c = 0.5, MeOH); UV-Vis (MeOH): λ_{max} (ϵ) = 272 nm (60914 dm³ mol⁻¹ cm⁻¹), 388 (17236).

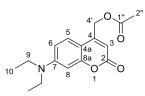
reversed-phase HPLC: $t_R = 4.5$ min; column: *HyperClone* 5 μ ODS (C18) 120 (*Phenomenex*); detection (UV): 388 nm; eluent: H₂O/MeOH 40:60; flowrate: 0.5 mL/min; column temperature: 25 °C; sample solvent: H₂O.



S3.5 Synthetic scheme for preparation of photocaged IPTG 1c

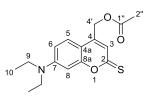
Scheme S5: Synthetic scheme for preparation of photocaged IPTG 1c. Reagents and conditions: i) AcOH, DCC, DMAP, 0 °C \rightarrow RT, 20 h; ii) Lawesson's reagent, toluene, reflux, 12 h; iii) malononitrile, NEt₃, AgNO₃, RT, 4 h; iv) HCl in EtOH (1.25 M), EtOH, reflux, 15 h; v) 4-nitrophenyl chloroformate, DIPEA, CH₂Cl₂, 22 h; vi) 9, DMAP, CH₂Cl₂, RT, 20 h; vii) TFA, H₂O, CH₂Cl₂, 0 °C, 10 min.

Synthesis of [7-(Diethylamino)-2-oxo-2H-chromen-4-yl]methyl acetate (S9)



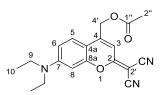
Coumarin S9 was synthesized using a modified procedure of Gandioso et al.^[22] Coumarin 3 (500 mg. 2.02 mmol) was dissolved in dry CH₂Cl₂ (35 mL) under nitrogen atmosphere. 4-Dimethylaminopyridine (DMAP) (299 mg, 2.43 mmol) and acetic acid (139 µL, 2.43 mmol) were added, and the reaction mixture was cooled to 0 °C. N,N'-Dicyclohexylcarbodiimide (DCC) (501 mg, 2.43 mmol) was added at 0 °C and the reaction mixture was stirred for 20 h in the dark. After filtration, the organic filtrate was washed with 1 M HCl and saturated NaHCO₃ solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (CH₂Cl₂) to yield a red solid (518 mg, 1.79 mmol, 89%). The spectroscopic data are consistent with previously reported literature values.^[22] ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.19 (t, ${}^{3}J_{10,9}$ = 7.1 Hz, 6 H, 10-H), 2.18 (s, 3 H, 2"-H), 3.40 (q, ${}^{3}J_{9,10}$ = 7.1 Hz, 4 H, 9-H), 5.20 (d, ${}^{4}J_{4',3}$ = 1.3 Hz, 2 H, 4'-H) 6.11 (t, ${}^{4}J_{3,4'}$ = 1.3 Hz, 1 H, 3-H), 6.50 (d, ${}^{4}J_{8,6}$ = 2.6 Hz, 1 H, 8-H), 6.56 (dd, ${}^{3}J_{6,5}$ = 9.0 Hz, ${}^{4}J_{6,8} = 2.6$ Hz, 1 H, 6-H), 7.27 (d, ${}^{3}J_{5,6} = 9.0$ Hz, 1 H, 5-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 20.9 (C-2"), 44.9 (C-9), 61.4 (C-4'), 97.9 (C-8), 106.1 (C-4a), 106.5 (C-3), 108.8 (C-6), 124.5 (C-5), 149.5 (C-4), 150.8 (C-7), 156.4 (C-8a), 162.0 (C-2), 170.3 (C-1"); $R_f = 0.57$ (PE/EtOAc 50:50); IR (atr-film): \tilde{v} [cm⁻¹] = 2974, 1748, 1706, 1597, 1527, 1440, 1415, 1376, 1337, 1272, 1240, 1196, 1140, 1074, 1013, 933, 841, 823, 811, 666, 598, 560; HRMS (ESI): m/z calculated for C₁₆H₂₀NO₄⁺ [M+H]⁺: 290.1387; found: 290.1389; m.p.: 108.3 °C.

Synthesis of [7-(Diethylamino)-2-thioxo-2H-chromen-4-yl]methyl acetate (S10)



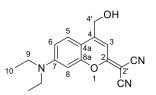
Coumarin **S10** was synthesized using a modified procedure of Gandioso *et al.*^[22] Coumarin **S9** (2.00 g, 6.91 mmol) was dissolved in dry toluene (237 mL) under nitrogen atmosphere. Lawesson's reagent (1.82 g, 4.49 mmol) was added, and the reaction mixture was heated to reflux for 12 h in the dark. The solution was concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (CH₂Cl₂) to yield an orange solid (1.78 g, 5.83 mmol, 84%). The spectroscopic data are consistent with previously reported literature values.^[22] ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.22 (t, ³*J*_{10.9} = 7.1 Hz, 6 H, 10-H), 2.19 (s, 3 H, 2"-H), 3.43 (q, ³*J*_{9.10} = 7.1 Hz, 4 H, 9-H), 5.18 (d, ⁴*J*_{4',3} = 1.3 Hz, 2 H, 4'-H), 6.66 (dd, ³*J*_{6.5} = 9.0 Hz, ⁴*J*_{6.8} = 2.6 Hz, 1 H, 6-H), 6.68 (d, ³*J*_{8.6} = 2.6 Hz, 1 H, 8-H), 7.06 (t, ⁴*J*_{3.4'} = 1.3 Hz, 1 H, 3-H), 7.34 (d, ³*J*_{5.6} = 9.0 Hz, 1 H, 5-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 20.9 (C-2") 45.1 (C-9), 61.1 (C-4'), 97.6 (C-8), 108.3 (C-4a), 110.4 (C-6), 120.7 (C-3), 124.6 (C-5), 142.0 (C-4), 151.2 (C-7), 159.1 (C-8a), 170.4 (C-1"), 197.3 (C-2); R_{*f*} = 0.60 (CH₂Cl₂); IR (atr-film): \hat{v} [cm⁻¹] = 2971, 1743, 1625, 1574, 1516, 1432, 1399, 1376, 1354, 1290, 1250, 1217, 1197, 1180, 1147, 1071, 1029, 967, 855, 822, 795, 652; HRMS (ESI): m/z calculated for C₁₆H₂₀NO₃S⁺ [M+H]⁺: 306.1158; found: 306.1160; m.p.: 137.9 °C

Synthesis of [2-(Dicyanomethylene)-7-(diethylamino)-2*H*-chromen-4-yl]methyl acetate (S11)



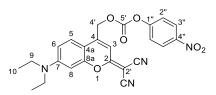
Coumarin S11 was synthesized using a procedure of Gandioso et al.^[22] Coumarin S10 (1.00 g, 3.27 mmol) was dissolved in dry MeCN (100 mL) under nitrogen atmosphere. After addition of malononitrile (1.09 g, 16.5 mmol) and triethylamine (NEt₃) (9.13 mL, 65.5 mmol), the reaction mixture was stirred for 20 min at room temperature in the dark. The reaction mixture became intensely red. Silver nitrate (1.12 g, 6.58 mmol) was added and the reaction mixture was stirred for additional 4 h at room temperature in the dark. The reaction mixture was filtered and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO_2 (CH₂Cl₂) to yield an orange-red solid (616 mg, 1.83 mmol, 56%). The spectroscopic data are consistent with previously reported literature values.^{[22] 1}H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.24 (t, ³J_{10.9} = 7.1 Hz, 6 H, 10-H), 2.21 (s, 3 H, 2"-H), 3.45 (q, ${}^{3}J_{9,10} = 7.1$ Hz, 4 H, 9-H), 5.24 (d, ${}^{4}J_{4',3} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, ${}^{3}J_{8,6} = 1.2$ Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 2 H, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 Hz, 4'-H), 6.62 (d, {}^{3}J_{8,6} = 1.2 2.6 Hz, 1 H, 8-H), 6.70 (dd, ${}^{3}J_{6,5} = 9.0$ Hz, ${}^{4}J_{6,8} = 2.6$ Hz, 1 H, 6-H), 6.75 (t, ${}^{4}J_{3,4'} = 1.2$ Hz, 1 H, 3-H), 7.34 (d, ${}^{3}J_{5.6} = 9.0$ Hz, 1 H, 5-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 20.9 (C-2"), 45.3 (C-9), 55.9 (C-2'), 61.2 (C-4'), 97.9 (C-8), 106.2 (C-3), 107.4 (C-4a), 111.1 (C-6), 113.9 (CN), 114.6 (CN), 125.0 (C-5), 146.1 (C-4), 151.6 (C-7), 155.1 (C-8a), 170.3 (C-1"), 171.9 (C-2); R_f = 0.48 (PE/EtOAc 70:30); IR (atr-film): \tilde{v} [cm⁻¹] = 2969, 2924, 2215, 1750, 1638, 1586, 1524, 1430, 1358, 1320, 1259, 1224, 1148, 1079, 816, 687; HRMS (ESI): m/z calculated for $C_{19}H_{20}N_3O_3^+$ [M+H]⁺: 338.1499; found: 338.1495; m.p.: 202-203 °C.

Synthesis of 2-(Dicyanomethylene)-7-(diethylamino)-4-(hydroxymethyl)-2H-chromen (4)



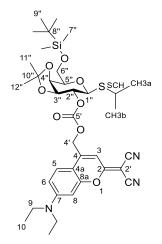
Coumarin **4** was synthesized using a procedure of Fournier *et al.*^[23] Coumarin **S11** (500 mg, 1.48 mmol) was dissolved in dry EtOH (316 mL) under nitrogen atmosphere. After addition of HCl in EtOH (1.25 M, 2.96 mL, 3.71 mmol), the reaction mixture was heated to reflux for 15 h in the dark. Then, the reaction mixture was concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (dichloromethane/acetone 95:5) to yield an orange-red solid (403 mg, 1.36 mmol, 92%). The spectroscopic data are consistent with previously reported literature values.^[23] ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.24 (t, ³*J*_{10.9} = 7.1 Hz, 6 H, 10-H), 3.45 (q, ³*J*_{9.10} = 7.1 Hz, 4 H, 9-H), 4.89 (s, 2 H, 4'-H), 6.63 (d, ³*J*_{8.6} = 2.6 Hz, 1 H, 8-H), 6.71 (dd, ³*J*_{6.5} = 9.0 Hz, ⁴*J*_{6.8} = 2.6 Hz, 1 H, 6-H), 6.98 (s, 1 H, 3-H), 7.36 (d, ³*J*_{5.6} = 9.0 Hz, 1 H, 5-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.6 (C-10), 45.4 (C-9), 54.9 (C-2'), 60.8 (C-4'), 97.8 (C-8), 105.4 (C-3), 107.7 (C-4a), 111.2 (C-6), 114.2 (CN), 115.0 (CN), 125.0 (C-5), 151.4 (C-4), 151.6 (C-7), 154.9 (C-8a), 172.3 (C-2); R_{*f*} = 0.44 (CH₂Cl₂/acetone 95:5); IR (atr-film): \hat{v} [cm⁻¹] = 3431, 2972, 2918, 2199, 1741, 1634, 1575, 1504, 1418, 1355, 1319, 1254, 1189, 1144, 1085, 821, 686, 508; MS (ESI, positive ion): m/z (%) = 296.3 (100) [M+H]⁺; m.p.: 179–180 °C

Synthesis of [2-(Dicyanomethylene)-7-(diethylamino)-2*H*-chromen-4-yl]methyl (4nitrophenyl) carbonate (6)



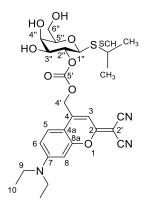
Coumarin 6 was synthesized using modified procedures of Gao et al.^[9] and Fomina et al.^[10] Coumarin 4 (250 mg, 846 umol) was dissolved in dry CH₂Cl₂ (15.0 mL) under nitrogen atmosphere. N.N-Diisopropylethylamine (DIPEA) (295 µL, 1.69 mmol) was added and the reaction mixture was stirred for 15 min before 4-nitrophenyl chloroformate (341 mg, 1.69 mmol) was added portion wise. The reaction mixture was stirred for 22 h and diluted with CH₂Cl₂. It was washed with 1 M HCl (1×) and saturated NaHCO₃ solution (3×). The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO_2 (petroleum ether/ethyl acetate 70:30) to yield a red solid (179 mg, 389 µmol, 46%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.25 (t, ${}^{3}J_{10.9}$ = 7.1 Hz, 6 H, 10-H), 3.46 (q, ${}^{3}J_{9.10}$ = 7.1 Hz, 4 H, 9-H), 5.42 (s, 2 H, 4'-H), 6.61 (d, ${}^{3}J_{8,6} = 2.5$ Hz, 1 H, 8-H), 6.69 (dd, ${}^{3}J_{6,5} = 9.0$ Hz, ${}^{4}J_{6,8} = 2.5$ Hz, 1 H, 6-H), 6.80 (s, 1 H, 3-H), 7.35 (d, ${}^{3}J_{5,6} = 9.0$ Hz, 1 H, 5-H), 7.43 (m_c, 2 H, 2"-H), 8.31 (m_c, 2 H, 3"-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.6 (C-10), 45.2 (C-9), 56.5 (C-2'), 65.5 (C-4'), 97.6 (C-8), 106.3 (C-3), 106.7 (C-4a), 110.9 (C-6), 113.7 (CN), 114.5 (CN), 121.9 (C-2"), 124.9 (C-5), 125.6 (C-3"), 144.2 (C-4), 145.9 (C-4"), 152.0 (C-7), 152.2 (C-5'), 155.2, 155.2 (C-8a, C-1"), 171.7 (C-2); R_f = 0.28 (PE/EtOAc 60:40); IR (atrfilm): \tilde{v} [cm⁻¹] = 2215, 1774, 1638, 1584, 1548, 1523, 1489, 1432, 1350, 1321, 1259, 1216, 1148, 1083, 860; HRMS (ESI): m/z calculated for C₂₄H₂₁N₄O₆⁺ [M+H]⁺: 461.1456; found: 461.1457; m.p.: 209– 211 °C

Synthesis of Isopropyl 6-*O*-(*tert*-butyldimethylsilyl)-2-*O*-{[(dicyanomethylene)-7-(diethylamino)-2*H*-chromen-4-yl]methyloxycarbonyl}-3,4-*O*-(1-methylethylidene)-1-thio- β -D-galactopyranoside (13)



Coumarin 13 was synthesized using modified procedures of Suzuki et al.^[24] Coumarin 6 (0.12 g, 0.25 mmol) was dissolved in dry CH₂Cl₂ (4.0 mL) under nitrogen atmosphere. After the addition of 4dimethylaminopyridine (DMAP) (31 mg, 0.26 mmol) and carbohydrate 9 (90 mg, 0.23 mmol) the reaction mixture was stirred for 20 h at room temperature in the dark. Then, it was diluted with CH₂Cl₂, washed with saturated NaHCO₃ and saturated NaCl solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 80:20) to yield a red solid (0.15 g, 0.22 mmol, 96%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 0.07 (s, 3 H, 7"-H), 0.07 (s, 3 H, 7"-H), 0.89 (s, 9 H, 9"-H), 1.23 (t, ${}^{3}J_{10,9} = 7.1$ Hz, 6 H, 10-H), 1.29 (d, ${}^{3}J_{CH3,SCH} = 6.8$ Hz, 3 H, CH₃), 1.30 (d, ${}^{$ 6.8 Hz, 3 H, CH₃), 1.35 (s, 3 H, 11"-H or 12"-H), 1.55 (s, 3 H, 11"-H or 12"-H), 3.19 (septet, ³J_{SCH,CH3}) = 6.8 Hz, 1 H, SCH), 3.44 (q, ${}^{3}J_{9,10} = 7.1$ Hz, 4 H, 9-H), 3.80-3.89 (m, ${}^{3}J_{5'',4''} = 5.6$ Hz, 3 H, 5"-H, 6"-H), 4.23 (dd, ${}^{3}J_{3'',2''} = 7.3$ Hz, ${}^{3}J_{3'',4''} = 5.3$ Hz, 1 H, 3"-H), 4.29 (dd, ${}^{3}J_{4'',3''} = 5.3$ Hz, ${}^{3}J_{4'',5''} = 1.9$ Hz, 1 H, 4"-H), 4.49 (d, ${}^{3}J_{1",2"} = 10.4$ Hz, 1 H, 1"-H), 4.79 (dd, ${}^{3}J_{2",1"} = 10.4$ Hz, ${}^{3}J_{2",3"} = 7.3$ Hz, 1 H, 2"-H), 5.25 (dd, ${}^{3}J_{4'a, 4'b} = 15.5$ Hz, ${}^{4}J_{4'a,3} = 1.2$ Hz, 1 H, 4'-H_a), 5.39 (dd, ${}^{3}J_{4'b, 4'a} = 15.5$ Hz, ${}^{4}J_{4'b,3} = 1.2$ Hz, 1 H, 4'-H_b), 6.58 (d, ${}^{4}J_{8,6}$ = 2.6 Hz, 1 H, 8-H), 6.64 (dd, ${}^{3}J_{6,5}$ = 9.1 Hz, ${}^{4}J_{6,8}$ = 2.6 Hz, 1 H, 6-H), 6.81 (m_c, 1 H, 3-H), 7.29 (d, ${}^{3}J_{5.6} = 9.1$ Hz, 1 H, 5-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = -5.4 (C-7"), -5.3 (C-7"), 12.6 (C-10), 18.3 (C-8"), 24.0 (CH₃), 24.1 (CH₃), 25.9 (C-9"), 26.5, 28.0 (C-11", C-12"), 35.8 (SCH), 45.1 (C-9), 55.9 (C-2'), 62.1 (C-6"), 64.7 (C-4'), 73.6 (C-4"), 77.0 (C-3"), 77.2 (C-5"), 77.3 (C-2"), 82.4 (C-1"), 97.5 (C-8), 106.0 (C-3), 106.8 (C-4a), 110.6 (C-10"), 110.8 (C-6), 114.0 (CN), 114.5 (CN), 124.9 (C-5), 145.4 (C-4), 151.8 (C-7), 154.2 (C-5'), 155.0 (C-8a), 171.9 (C-2); R_f = 0.24 (PE/EtOAc 80:20); IR (atr-film): \tilde{v} [cm⁻¹] = 2955, 2929, 2859, 2216, 1756, 1639, 1586, 1548, 1525, 1432, 1383, 1356, 1319, 1257, 1223, 1196, 1148, 1111, 1078, 1047, 983, 871, 839, 760; HRMS (ESI): m/z calculated for $C_{36}H_{52}N_3O_8SSi^+$ [M+H]⁺: 714.3239; found: 714.3237; m.p.: 79–81 °C; $[\alpha]_D^{20} = -18.0$ $(c = 0.1, CHCl_3).$

Synthesis of Isopropyl 2-O-{[(dicyanomethylene)-7-(diethylamino)-2*H*-chromen-4-yl]methyloxycarbonyl}-1-thio- β -D-galactopyranoside (1c)



Photocaged IPTG 1c was synthesized using modified procedures of Suzuki et al.^[24] Coumarin 13 (0.10 g, 0.14 mmol) was dissolved in CH₂Cl₂ (1.0 mL) and cooled to 0 °C. After the addition of trifluoroacetic acid (TFA) (1.0 mL, 13 mmol) and water (40 µL, 2.2 mmol), the reaction mixture was stirred for 10 min at 0 °C in the dark. Then the reaction mixture was concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO_2 (ethyl acetate) to yield a red solid (72 mg, 0.13 mmol, 92%). ¹H-NMR (600 MHz, Aceton- d_6): δ [ppm] = 1.18–1.28 (m, 12 H, 10-H, CH₃), 3.20 (septet, ${}^{3}J_{SCH,CH3} = 6.8$ Hz, 1 H, SCH), 3.59 (q, ${}^{3}J_{9,10} = 7.1$ Hz, 4 H, 9-H), 3.67 (t, ${}^{3}J_{5'',4''}$ = 5.6 Hz, 1 H, 5"-H), 3.77 (m, 2 H, 6"-H), 3.86 (dd, ${}^{3}J_{3",2"}$ = 9.3 Hz, ${}^{3}J_{3",4"}$ = 3.4 Hz, 1 H, 3"-H), 4.10 (d, ${}^{3}J_{4'',3''} = 3.4$ Hz, 1 H, 4"-H), 4.65 (d, ${}^{3}J_{1'',2''} = 10.1$ Hz, 1 H, 1"-H), 4.88 (dd, ${}^{3}J_{2'',1''} = 10.1$ Hz, ${}^{3}J_{2'',3''} = 10.1$ Hz, ${}^{3}J_{2'',3'''} = 10.1$ Hz, ${}^{3}J_{2'',3'''$ 9.3 Hz, 1 H, 2"-H), 5.43 (d, ${}^{3}J_{4'a, 4'b} = 15.5$ Hz, 1 H, 4'-H_a), 5.53 (d, ${}^{3}J_{4'b, 4'a} = 15.5$ Hz, 1 H, 4'-H_b), 6.74– 6.79 (m, 2 H, 3-H, 8-H), 6.91 (dd, ${}^{3}J_{6.5} = 9.1$ Hz, ${}^{4}J_{6.8} = 2.6$ Hz, 1 H, 6-H), 7.63 (d, ${}^{3}J_{5.6} = 9.1$ Hz, 1 H, 5-H); ¹³C-NMR (151 MHz, Aceton- d_6): δ [ppm] = 12.7 (C-10), 24.2 (CH₃), 24.6 (CH₃), 35.5 (SCH), 45.5 (C-9), 55.1 (C-2'), 62.3 (C-6"), 65.4 (C-4'), 70.4 (C-4"), 73.4 (C-3"), 77.3 (C-2"), 79.8 (C-5"), 83.5 (C-1"), 97.6 (C-8), 106.0 (C-3), 107.7 (C-4a), 111.9 (C-6), 114.3 (CN), 115.2 (CN), 126.8 (C-5), 147.9 (C-4), 153.0 (C-7), 155.1 (C-5'), 155.9 (C-8a), 172.6 (C-2); $R_f = 0.23$ (EtOAc); IR (atr-film): \tilde{v} [cm⁻¹] = 3359, 2954, 2924, 2855, 2216, 1748, 1672, 1638, 1584, 1522, 1433, 1381, 1320, 1258, 1193, 1139, 1076, 1053, 984, 800, 725; HRMS (ESI): m/z calculated for $C_{27}H_{34}N_3O_8S^+$ [M+NH₄]⁺: 560.2061; found: 560.2055; m.p.: 153.8 °C; $[\alpha]_{D}^{20} = 74$ (c = 0.1, CHCl₃); UV-Vis [Tris buffer (20 mM, pH 7.5)/MeCN 1:1]: $\lambda_{\text{max}}(\epsilon) = 252 \text{ nm} (18618 \text{ dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}), 276 (14338), 488 (18938).$

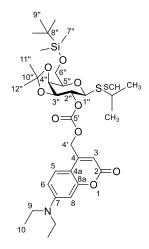
reversed-phase HPLC: $t_R = 14.6$ min; column: *HyperClone* 5 μ ODS (C18) 120 (*Phenomenex*); detection (UV): 488 nm; eluent: H₂O/MeOH 40:60; flowrate: 0.5 mL/min; column temperature: 25 °C; sample solvent: Tris buffer (20 mM, pH 7.5)/MeCN 1:1.

OTBS но HO-NO/ \cap ii Et₂N 5 iii Et₂N Et₂N 14 1b iv РМВО HO Et₂N Et₂N РМВО но PMBÒ ЮH 16 1d

S3.6 Synthetic scheme for preparation of photocaged IPTG 1b and 1d

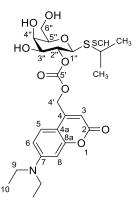
Scheme S6: Synthetic scheme for preparation of photocaged IPTG 1b and 1d. Reagents and conditions: i) 9, DMAP, CH₂Cl₂, RT, 20 h; ii) TFA, H₂O, CH₂Cl₂, 0 °C, 10 min; iii) 10, DMAP, CH₂Cl₂, RT, 20 h; iv) TFA, H₂O, CH₂Cl₂, 0 °C \rightarrow RT, 1 h.

Synthesis of Isopropyl 6-*O*-(*tert*-butyldimethylsilyl)-2-*O*-{[7-(diethylamino)-2-oxo-2*H*chromen-4-yl]methyloxycarbonyl}-3,4-*O*-(1-methylethylidene)-1-thio- β -Dgalactopyranoside (14)



Coumarin 14 was synthesized using modified procedures of Suzuki *et al.*^[24] Coumarin 5 (98 mg, 0.24 mmol) was dissolved in dry CH_2Cl_2 (2 mL) under nitrogen atmosphere. After the addition of 4dimethylaminopyridine (DMAP) (30 mg, 0.24 mmol) and carbohydrate 9 (85 mg, 0.22 mmol) the reaction mixture was stirred for 20 h at room temperature in the dark. Then, it was diluted with CH_2Cl_2 , washed with saturated NaHCO₃ and saturated NaCl solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 80:20) to yield a yellow solid (0.11 g, 0.17 mmol, 77%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 0.06 (s, 3 H, 7"-H), 0.07 (s, 3 H, 7"-H), 0.89 $(s, 9 H, 9''-H), 1.19 (t, {}^{3}J_{10,9} = 7.1 Hz, 6 H, 10-H), 1.29 (d, {}^{3}J_{CH3,SCH} = 6.8 Hz, 3 H, CH_{3}), 1.30 (d, {}^{3}J_{CH3,SCH} = 6.8 Hz, 3 Hz, 2 H$ = 6.8 Hz, 3 H, CH₃), 1.35 (s, 3 H, 11"-H or 12"-H), 1.56 (s, 3 H, 11"-H or 12"-H), 3.17 (septet, ³J_{SCH,CH3}) = 6.8 Hz, 1 H, SCH), 3.40 (q, ${}^{3}J_{9,10}$ = 7.1 Hz, 4 H, 9-H), 3.79–3.91 (m, 3 H, 5"-H, 6"-H), 4.22 (dd, ${}^{3}J_{3",2"}$ = 7.3 Hz, ${}^{3}J_{3'',4''}$ = 5.3 Hz, 1 H, 3"-H), 4.29 (d, ${}^{3}J_{4'',3''}$ = 5.2 Hz, ${}^{3}J_{4'',5''}$ = 1.9 Hz, 1 H, 4"-H), 4.48 (d, ${}^{3}J_{1'',2''}$ = 10.4 Hz, 1 H, 1"-H), 4.80 (dd, ${}^{3}J_{2",1"}$ = 10.4 Hz, ${}^{3}J_{2",3"}$ = 7.3 Hz, 1 H, 2"-H), 5.23 (d, ${}^{3}J_{4'a,4'b}$ = 14.9 Hz, 1 H, 4'-H_a), 5.38 (d, ${}^{3}J_{4'b, 4'a}$ = 14.9 Hz, 1 H, 4'-H_b), 6.21 (s, 1 H, 3-H), 6.50 (d, ${}^{4}J_{8,6}$ = 2.6 Hz, 1 H, 8-H), $6.56 (dd, {}^{3}J_{6,5} = 9.0 Hz, {}^{4}J_{6,8} = 2.6 Hz, 1 H, 6-H), 7.25 (d, {}^{3}J_{5,6} = 9.0 Hz, 1 H, 5-H); {}^{13}C-NMR (151 MHz, 1.5)$ $CDCl_3$: δ [ppm] = -5.4 (C-7"), -5.3 (C-7"), 12.5 (C-10), 18.3 (C-8"), 24.0 (CH₃), 24.1 (CH₃), 25.9 (C-9"), 26.5, 28.0 (C-11", C-12"), 35.7 (SCH), 44.9 (C-9), 62.1 (C-6"), 65.0 (C-4'), 73.5 (C-4"), 76.9 (C-3"), 77.1 (C-5"), 77.1 (C-2"), 82.5 (C-1"), 98.0 (C-8), 106.0 (C-4a), 106.7 (C-3), 108.8 (C-6), 110.6 (C-10''), 124.4 (C-5), 148.7 (C-4), 150.7 (C-7), 154.3 (C-5'), 156.4 (C-8a), 162.0 (C-2); $R_f = 0.24$ (PE/EtOAc 80:20); IR (atr-film): \tilde{v} [cm⁻¹] = 2969, 2931, 2870, 1745, 1713, 1603, 1527, 1420, 1356, 1336, 1217, 1139, 1094, 1064, 1023, 841, 751, 667, 601, 510; HRMS (ESI): m/z calculated for $C_{33}H_{52}NO_9SSi^+$ [M+H]⁺: 666.3127; found: 666.3124; m.p.: 69 °C; $[\alpha]_D^{20} = 16$ (c = 0.1, CHCl₃).

SynthesisofIsopropyl2-O-{[7-(diethylamino)-2-oxo-2H-chromen-4-yl]methyloxycarbonyl}-1-thio-β-D-galactopyranoside (1b)

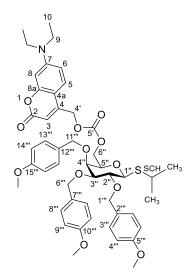


Photocaged IPTG **1b** was synthesized using modified procedures of Suzuki *et al.*^[24] Coumarin **14** (159 mg, 238 µmol) was dissolved in CH₂Cl₂ (1 mL) and cooled to 0 °C. After the addition of trifluoroacetic acid (TFA) (1.70 mL, 22.2 mmol) and water (68.8 µL, 3.82 mmol), the reaction mixture was stirred for 10 min at 0 °C in the dark. Then the reaction mixture was concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (ethyl acetate) to yield a yellow solid (117 mg, 229 µmol, 96%). ¹H-NMR (600 MHz, Aceton-*d*₆): δ [ppm] = 1.21 (t, ³*J*_{10,9} = 7.1 Hz, 6 H, 10-H), 1.25 (d, ³*J*_{CH3,SCH} = 6.7 Hz, 3 H, CH₃), 1.29 (d, ³*J*_{CH3,SCH} = 6.7 Hz, 3 H, CH₃), 3.21 (septet, ³*J*_{SCH,CH3} = 6.7 Hz, 1 H, SCH), 3.51 (q, ³*J*_{9,10} = 7.1 Hz, 4 H, 9-H), 3.67 (t, ³*J*_{5",4"} = 5.6 Hz, 1 H, 5"-H), 3.77 (m, 2 H, 6"-H), 3.84 (dd, ³*J*_{3",2"} = 9.3 Hz, ³*J*_{3",4"} = 3.4 Hz, 1 H, 3"-H), 4.08 (d, ³*J*_{4",3"} = 3.4 Hz,

1 H, 4"-H), 4.65 (d, ${}^{3}J_{1",2"} = 10.1$ Hz, 1 H, 1"-H), 4.87 (dd, ${}^{3}J_{2",1"} = 10.1$ Hz, ${}^{3}J_{2",3"} = 9.3$ Hz, 1 H, 2"-H), 5.33 (d, ${}^{3}J_{4'a,4'b} = 15.2$ Hz, 1 H, 4'-H_a), 5.45 (d, ${}^{3}J_{4'b,4'a} = 15.2$ Hz, 1 H, 4'-H_b), 6.09 (s, 1 H, 3-H), 6.52 (d, ${}^{4}J_{8,6} = 2.6$ Hz, 1 H, 8-H), 6.72 (dd, ${}^{3}J_{6,5} = 9.0$ Hz, ${}^{4}J_{6,8} = 2.6$ Hz, 1 H, 6-H), 7.48 (d, ${}^{3}J_{5,6} = 9.0$ Hz, 1 H, 5-H); 13 C-NMR (151 MHz, Aceton- d_6): δ [ppm] = 12.7 (C-10), 24.2 (CH₃), 24.6 (CH₃), 35.6 (SCH), 45.2 (C-9), 62.3 (C-6"), 65.4 (C-4'), 70.4 (C-4"), 73.5 (C-3"), 77.0 (C-2"), 79.9 (C-5"), 83.8 (C-1"), 98.1 (C-8), 106.5 (C-4a), 106.7 (C-3), 109.5 (C-6), 125.9 (C-5), 150.5 (C-4), 151.8 (C-7), 155.4 (C-5'), 157.3 (C-8a), 161.4 (C-2); R_f = 0.24 (EtOAc); IR (atr-film): \tilde{v} [cm⁻¹] = 3405, 2962, 2925, 2862, 1755, 1679, 1603, 1528, 1424, 1357, 1255, 1200, 1139, 1054, 984, 800, 725; HRMS (ESI): m/z calculated for C₂₄H₃₄NO₉S⁺ [M+H]⁺: 512.1949; found: 512.1952; m.p.: 96.0 °C; [α]²⁰ = 28 (c = 0.1, CHCl₃); UV-Vis [Tris buffer (20 mM, pH 7.5)/MeCN 1:1]: λ_{max} (ϵ) = 246 nm (21496 dm³ mol⁻¹ cm⁻¹), 386 (24984).

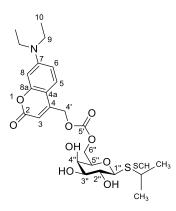
reversed-phase HPLC: $t_R = 9.8$ min; column: *HyperClone* 5 μ ODS (C18) 120 (*Phenomenex*); detection (UV): 392 nm; eluent: H₂O/MeOH 40:60; flowrate: 0.5 mL/min; column temperature: 25 °C; sample solvent: Tris buffer (20 mM, pH 7.5)/MeCN 1:1.

SynthesisofIsopropyl6-O-{[7-(diethylamino)-2-oxo-2H-chromen-4-yl]methyloxycarbonyl}-2,3,4-tri-O-(4-methoxybenzyl)-1-thio-β-D-galactopyranoside (16)



Coumarin **16** was synthesized using modified procedures of Suzuki *et al.*^[24] Coumarin **5** (60 mg, 0.15 mmol) was dissolved in dry CH₂Cl₂ (2.0 mL) under nitrogen atmosphere. After the addition of 4dimethylaminopyridine (DMAP) (20 mg, 0.16 mmol) and carbohydrate **10** (96 mg, 0.16 mmol) the reaction mixture was stirred for 20 h at room temperature in the dark. Then, it was diluted with CH₂Cl₂, washed with saturated NaHCO₃ and saturated NaCl solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 60:40) to yield a yellow solid (84 mg, 0.10 mmol, 66%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.20 (t, ³J_{10,9} = 7.1 Hz, 6 H, 10-H), 1.31 (d, ³J_{CH3,SCH} = 6.8 Hz, 6 H, CH₃), 3.20 (septet, ³J_{SCH,CH3} = 6.8 Hz, 1 H, SCH), 3.41 (q, ³J_{9,10} = 7.1 Hz, 4 H, 9-H), 3.52–3.58 (m, 2 H, 3"-H, 5"-H), 3.74–3.84 (m, 11 H, 2"-H, 4"-H, OCH₃), 4.03 (dd, ${}^{2}J_{6"a,6"b} =$ 10.9 Hz, ${}^{3}J_{6''a,5''} = 5.9$ Hz, 1 H, 6"-H_a), 4.29 (dd, ${}^{2}J_{6''b,6''a} = 10.9$ Hz, ${}^{3}J_{6''b,5''} = 6.6$ Hz, 1 H, 6"-H_b), 4.47 (d, ${}^{3}J_{1,2} = 9.7$ Hz, 1 H, 1-H), 4.58 (d, ${}^{2}J_{11''a,11''b} = 11.5$ Hz, 1 H, 11'''-H_a), 4.67–4.74 (m, 3 H, 1'''-H_a, 6'''-H), $4.81 \text{ (d, } {}^{2}J_{1'''_{b},1'''_{a}} = 9.8 \text{ Hz}, 1 \text{ H}, 1'''-\text{H}_{b}), 4.90 \text{ (d, } {}^{2}J_{11'''_{b},11'''_{a}} = 11.5 \text{ Hz}, 1 \text{ H}, 11'''-\text{H}_{b}), 5.21 \text{ (m}_{c}, 2 \text{ H}, 4'-\text{H}), 5.21 \text$ 6.12 (t, ${}^{4}J_{3,4'}$ = 1.3 Hz, 1 H, 3-H), 6.51 (d, ${}^{4}J_{8,6}$ = 2.6 Hz, 1 H, 8-H), 6.57 (dd, ${}^{3}J_{6,5}$ = 9.0 Hz, ${}^{4}J_{6,8}$ = 2.6 Hz, 1 H, 6-H), 6.84 (d, ${}^{3}J_{14'',13''} = 8.5$ Hz, 2 H, 14'''-H), 6.86 (d, ${}^{3}J_{4'',3''} = 8.5$ Hz, 2 H, 4'''-H), 6.89 (d, ${}^{3}J_{9'',8''} = 8.5$ 8.5 Hz, 2 H, 9"-H), 7.24 (d, ${}^{3}J_{13",14"}$ = 8.5 Hz, 2 H, 13"-H), 7.26 (d, ${}^{3}J_{5,6}$ = 9.0 Hz, 1 H, 5-H), 7.31 (d, ${}^{3}J_{8'',9''} = 8.5$ Hz, 2 H, 8'''-H), 7.32 (d, ${}^{3}J_{3'',4''} = 8.5$ Hz, 2 H, 3'''-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.6 (C-10), 24.0 (CH₃), 35.6 (SCH), 44.9 (C-9), 55.4, 55.4, 55.4 (OCH₃), 64.8 (C-4'), 67.2 (C-6"), 72.3 (C-4"), 73.0 (C-6"), 73.8 (C-11"), 75.6 (C-1"), 75.6 (C-5"), 78.3 (C-2"), 83.9 (C-3"), 85.2 (C-1"), 98.0 (C-8), 105.9 (C-4a), 106.7 (C-3), 108.8 (C-6), 113.8, 113.9, 114.0 (C-4", C-9", C-14"), 124.4 (C-5), 129.4 (C-8"), 130.1 (C-3"), 130.2 (C-13"), 130.4, 130.5, 130.7 (C-2", C-7", C-12"), 148.7 (C-4), 150.9 (C-7), 154.3 (C-5'), 156.4 (C-8a), 159.4, 159.4, 159.4 (C-5''', C-10''', C-15'''), 161.8 (C-2); $R_f = 0.31$ (PE/EtOAc 60:40); IR (atr-film): \tilde{v} [cm⁻¹] = 2966, 2930, 2906, 2864, 2835, 1753, 1715, 1604, 1513, 1422, 1357, 1244, 1173, 1078, 1032, 822, 570, 519; HRMS (ESI): m/z calculated for C₄₈H₆₁N₂O₁₂S⁺ $[M+NH_4]^+$: 889.3940; found: 889.3929; m.p.: 93.6 °C; $[\alpha]_D^{20} = -18.8$ (c = 1.0, CHCl₃).

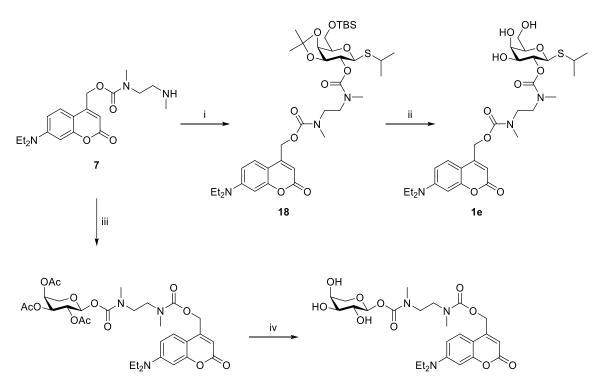
Synthesis of Isopropyl $6-O-\{[7-(diethylamino)-2-oxo-2H-chromen-4-y]$ methyloxycarbonyl}-1-thio- β -D-galactopyranoside (1d)



Photocaged IPTG **1d** was synthesized using modified procedures of Suzuki *et al.*^[24] Coumarin **16** (40 mg, 46 µmol) was dissolved in CH₂Cl₂ (1.0 mL) and cooled to 0 °C. After the addition of trifluoroacetic acid (TFA) (0.33 mL, 4.3 mmol) and water (13 µL, 0.73 mmol), the reaction mixture was stirred in the dark for 10 min at 0 °C and 1 h at room temperature. Then the reaction mixture was concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 50:50 to 10:90) to yield a yellow solid (20 mg, 39 µmol, 78%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.19 (t, ³J_{10.9} = 7.1 Hz, 6 H, 10-H), 1.31 (d, ³J_{CH3,SCH} = 6.7 Hz, 3 H, CH₃), 1.32 (d, ³J_{CH3,SCH} = 6.7 Hz, 3 H, CH₃), 3.15–3.25 (m, 2 H, SCH, 2"-OH), 3.31 (s, 1 H, 4"-OH),

3.40 (q, ${}^{3}J_{9,10} = 7.1$ Hz, 4 H, 9-H), 3.62–3.71 (m, 2 H, 2"-H, 3"-H), 3.74 (s, 1 H, 3"-OH), 3.80 (m_c, 1 H, 5"-H), 4.04 (brs, 1 H, 4"-H), 4.40 (dd, ${}^{3}J_{6a'',6b''} = 11.4$ Hz, ${}^{3}J_{6a'',5''} = 5.0$ Hz, 1 H, 6"-H_a), 4.42 (m_c, 1 H, 1"-H), 4.48 (dd, ${}^{3}J_{6b'',6a''} = 11.4$ Hz, ${}^{3}J_{6b'',5''} = 7.2$ Hz, 1 H, 6"-H_b), 5.23 (dd, ${}^{3}J_{4'a,4'b} = 15.2$ Hz, ${}^{4}J_{4'a,3} = 1.3$ Hz, 1 H, 4'-H_a), 5.27 (dd, ${}^{3}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'b,3} = 1.3$ Hz, 1 H, 4'-H_b), 6.12 (dd, ${}^{4}J_{3,4'a} = 1.3$ Hz, 4J, 4, ${}^{4}J_{3,4'b} = 1.3$ Hz, 1 H, 4'-H_a), 5.27 (dd, ${}^{3}J_{4'b,4'a} = 15.2$ Hz, ${}^{4}J_{4'b,3} = 1.3$ Hz, 1 H, 4'-H_b), 6.12 (dd, ${}^{4}J_{3,4'a} = 1.3$ Hz, 4J, 4, ${}^{4}J_{3,4'b} = 1.3$ Hz, 1 H, 3-H), 6.49 (d, ${}^{4}J_{8,6} = 2.6$ Hz, 1 H, 8-H), 6.56 (dd, ${}^{3}J_{6.5} = 9.0$ Hz, 4J, 4, a = 2.6 Hz, 1 H, 6-H), 7.26 (d, ${}^{3}J_{5.6} = 9.0$ Hz, 1 H, 5-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.6 (C-10), 24.1 (CH₃), 24.3 (CH₃), 36.1 (SCH), 44.9 (C-9), 64.9 (C-4'), 67.4 (C-6''), 68.8 (C-4''), 70.4 (C-2''), 74.6 (C-3''), 75.9 (C-5''), 86.1 (C-1''), 97.9 (C-8), 105.9 (C-4a), 106.5 (C-3), 108.9 (C-6), 124.5 (C-5), 148.9 (C-4), 150.9 (C-7), 154.7 (C-5'), 156.4 (C-8a), 162.0 (C-2); R_f = 0.35 (EtOAc); IR (atr-film): $\tilde{\nu}$ [cm⁻¹] = 3424, 2971, 2923, 2870, 1751, 1720, 1605, 1528, 1423, 1356, 1266, 1196, 1143, 1100, 1080, 1052, 1031, 967, 871, 828, 791, 743; HRMS (ESI): m/z calculated for C₂₄H₃₄NO₉S⁺ [M+H]⁺: 512.1949; found: 512.1953; m.p.: 85–87 °C; [α]²⁰_D = 22.0 (c = 0.1, CHCl₃); UV-Vis [Tris buffer (20 mM, pH 7.5)/MeCN 1:1]: λ_{max} (ϵ) = 273 nm (28210 dm³ mol⁻¹ cm⁻¹), 386 (12004).

reversed-phase HPLC: $t_R = 8.0$ min; column: *HyperClone* 5 μ ODS (C18) 120 (*Phenomenex*); detection (UV): 386 nm; eluent: H₂O/MeOH 40:60; flowrate: 0.5 mL/min; column temperature: 25 °C; sample solvent: Tris buffer (20 mM, pH 7.5)/MeCN 1:1.



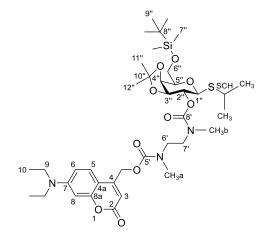
S3.7 Synthetic scheme for preparation of photocaged IPTG 1e and photocaged arabinose 2c

Scheme S7: Synthetic scheme for preparation of photocaged IPTG 1e and photocaged arabinose 2c. Reagents and conditions: i) 12, DIPEA, DMAP, CH₂Cl₂, RT, 24 h; ii) TFA, H₂O, 0 °C, 10 min; iii) 11, DIPEA, DMAP, CH₂Cl₂, RT, 24 h; iv) NH₃ in MeOH (7 M), MeOH, RT.

2c

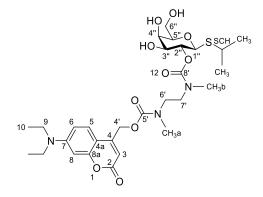
17

Synthesis of IsopropyI-6-*O*-(*tert*-butyIdimethyIsiIyI)-2-*O*-{[2-({[7-(diethylamino)-2-oxo-2*H*-chromen-4-yI]methoxycarbonyI}{methyI}amino)ethyI][methyI]carbamoyI}-3,4-*O*-(1-methylethylidene)-1-thio- β -D-galactopyranoside (18)



Coumarin 18 was synthesized using modified procedures of Wang et al.^[25] Coumarin 7 (227 mg, 628 µmol) was dissolved in dry CH₂Cl₂ (11 mL) under nitrogen atmosphere. After the addition of 4dimethylaminopyridine (DMAP) (6 mg, 0.06 mmol), carbohydrate 12 (200 mg, 359 µmol) and N.Ndiisopropylethylamine (DIPEA) (1.10 mL, 6.29 mmol) the reaction mixture was stirred for 24 h at room temperature in the dark. Then, it was diluted with CH₂Cl₂, washed with saturated NaHCO₃ and saturated NaCl solution. The organic phase was dried with anhydrous MgSO4 and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 50:50) to yield a slightly yellow foam (270 mg, 346 µmol, 97%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 0.06 (s, 6 H, 7"-H), 0.88 (s, 9 H, 9"-H), 1.20 (t, ${}^{3}J_{10,9}$ = 7.1 Hz, 6 H, 10-H), 1.23–1.33 (m, 9 H, CH₃, 11-H or 12-H), 1.55 (s, 3 H, 11-H or 12-H), 2.86–3.07 (m, 6 H, CH₃a, CH₃b), 3.17 (septet, ³J_{SCH,CH3}) = 6.8 Hz, 1 H, SCH), 3.28–3.62 (m, 4 H, 6'-H, 7'-H), 3.41 (q, ${}^{3}J_{9,10}$ = 7.1 Hz, 4 H, 9-H), 3.75–3.90 (m, 3 H, 5"-H, 6"-H), 4.13–4.22 (m, 1 H, 3"-H), 4.22–4.28 (m, 1 H, 4"-H), 4.44–4.55 (m, 1 H, 1"-H), 4.79– 4.91 (m, 1 H, 2"-H), 5.20–5.30 (m, 2 H, 4'-H), 6.03–6.13 (m, 1 H, 3-H), 6.50 (d, ${}^{3}J_{8,6} = 2.6$ Hz, 1 H, 8-H), 6.57 (d, ${}^{3}J_{6.5} = 9.0$ Hz, 1 H, 6-H), 7.27–7.36 (m, 1 H, 5-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = -5.4 (C-7"), -5.3 (C-7"), 12.6 (C-10), 18.4 (C-8"), 23.9 (CH₃), 24.0 (CH₃), 25.9 (C-9"), 26.5, 28.0 (C-11", C-12"), 34.9 (SCH), 35.0 (CH₃a/b), 35.3 (CH₃a/b), 35.6 (CH₃a/b), 35.9 (CH₃a/b), 36.0 (CH₃a/b), 36.1 (CH₃a/b), 44.9 (C-9), 46.7, 46.8, 47.2, 47.3, 47.5, 48.0, 48.1 (C-6', C-7'), 62.2 (C-4'), 62.5 (C-6''), 62.7 (C-6"), 73.6, 73.6, 73.7 (C-2", C-4"), 77.1 (C-5"), 77.5 (C-3"), 77.6 (C-3"), 77.6 (C-3"), 82.6 (C-1"), 82.7 (C-1"), 82.8 (C-1"), 82.9 (C-1"), 97.9 (C-8), 105.9, 106.1, 106.4 (C-3, C-4a), 108.8 (C-6), 110.4 (C-10"), 124.4 (C-5), 124.5 (C-5), 124.6 (C-5), 124.7 (C-5), 150.5, 150.6, 150.8 (C-4, C-7), 155.1, 155.3, 155.5, 155.6 (C-5', C-8'), 156.3 (C-8a), 156.4 (C-8a), 162.1 (C-2); $R_f = 0.31$ (PE/EtOAc 50:50); IR (atr-film): \tilde{v} [cm⁻¹] = 2960, 2930, 2859, 1710, 1606, 1529, 1466, 1422, 1358, 1219, 1125, 1081, 874, 839, 779; HRMS (ESI): m/z calculated for $C_{38}H_{62}N_3O_{10}SSi^+$ [M+H]⁺: 780.3920; found: 780.3935; m.p.: 68.0 °C; $[\alpha]_{D}^{20} = 0.4$ (c = 1.0, CHCl₃).

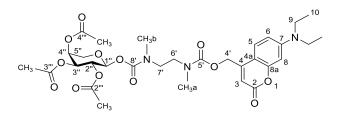
Synthesis of Isopropyl 2-O-{[2-({[7-(diethylamino)-2-oxo-2*H*-chromen-4-yl]methoxycarbonyl}{methyl}amino)ethyl][methyl]carbamoyl}-1-thio- β -D-galactopyranoside (1e)



Photocaged IPTG 1e was synthesized using modified procedures of Suzuki et al.^[24] Coumarin 18 (0.11 mg, 0.14 mmol) was dissolved in trifluoroacetic acid (TFA) (1.0 mL) and cooled to 0 °C. After the addition of water (41 µL, 2.3 mmol) the reaction mixture was stirred for 10 min at 0 °C in the dark. Then the reaction mixture was concentrated under reduced pressure. The residue was purified by flashcolumn chromatography on SiO₂ (ethyl acetate/methanol 95:5) to yield a yellow solid (89 mg, 0.14 mmol, quant.). The product was a mixture of cis and trans isomers on carbamate bonds. ¹H-NMR (600 MHz, DMSO- d_6 , 60 °C): δ [ppm] = 1.15 (t, ${}^{3}J_{10.9}$ = 7.0 Hz, 6 H, 10-H), 1.19 (d, ${}^{3}J_{CH3,SCH}$ = 6.8 Hz, 3 H, CH₃), 1.22 (d, ³J_{CH3,SCH} = 6.8 Hz, 3 H, CH₃), 2.85 (s, 3 H, CH₃a or CH₃b), 2.95 (brs, 3 H, CH₃a or CH₃b), 3.07–3.12 (m, 1 H, SCH), 3.34–3.47 (m, 9 H, 9-H, 6'-H, 7'-H, 5"-H), 3.50 (dd, ²*J*_{6"a,6"b} = 10.9 Hz, ${}^{3}J_{6"a,5"} = 6.0$ Hz, 1 H, 6"-H_a), 3.54 (dd, ${}^{2}J_{6"b,6"a} = 10.9$ Hz, ${}^{3}J_{6"b,5"} = 6.0$ Hz, 1 H, 6"-H_b), 3.56 (brs, 1 H, 3"-H), 3.78 (d, ${}^{3}J_{4",3"} = 3.3$ Hz, 1 H, 4"-H), 4.48 (brs, 1 H, 1"-H), 4.72 (dd, ${}^{3}J_{2",1"} = 10.1$ Hz, ${}^{3}J_{2",3"} = 10.1$ Hz, ${}^{3}J_{2",$ 9.3 Hz, 1 H, 2"-H), 5.25 (s, 2 H, 4'-H), 5.95 (s, 1 H, 3-H), 6.53 (d, ${}^{4}J_{8,6} = 2.6$ Hz, 1 H, 8-H), 6.71 (dd, ${}^{3}J_{6,5} = 9.0$ Hz, ${}^{4}J_{6,8} = 2.6$ Hz, 1 H, 6-H), 7.48 (d, ${}^{3}J_{5,6} = 9.0$ Hz, 1 H, 5-H); 13 C-NMR (151 MHz, DMSO- d_6 , 60 °C): δ [ppm] = 12.0 (C-10), 23.4 (CH₃), 23.8 (CH₃), 33.7 (SCH), 34.7, 34.9 (CH₃a/b), 43.7 (C-9), 45.9, 46.1 (C-6', C-7'), 60.3 (C-6"), 61.9 (C-4'), 68.4 (C-4"), 72.1 (C-2"), 72.6 (C-3"), 78.9 (C-5"), 82.7 (C-1"), 96.8 (C-8), 104.6 (C-3), 105.3 (C-4a), 108.7 (C-6), 125.0 (C-5), 150.4 (C-7), 151.0 (C-4), 154.5 (C-5'), 155.1 (C-8'), 155.6 (C-8a), 160.3 (C-2); $R_f = 0.15$ (EtOAc/MeOH 95:5); IR (atrfilm): \tilde{v} [cm⁻¹] = 3406, 2966, 2928, 2870, 1697, 1603, 1526, 1484, 1423, 1357, 1274, 1200, 1133, 1078, 862, 825, 803, 759; HRMS (ESI): m/z calculated for C₂₉H₄₄N₃O₁₀S⁺ [M+H]⁺: 626.2742; found: 626.2753; m.p.: 95.5 °C; $[\alpha]_D^{20} = -22.6$ (c = 1.0, CHCl₃); UV-Vis [Tris buffer (20 mM, pH 7.5)/MeCN 1:1]: $\lambda_{\text{max}}(\varepsilon) = 272 \text{ nm} (17662 \text{ dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}), 305 (4142), 386 (17184).$

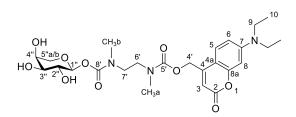
reversed-phase HPLC: $t_R = 8.2 \text{ min}$; column: *HyperClone* 5 μ ODS (C18) 120 (*Phenomenex*); detection (UV): 386 nm; eluent: H₂O/MeOH 40:60; flowrate: 0.5 mL/min; column temperature: 25 °C; sample solvent: Tris buffer (20 mM, pH 7.5)/MeCN 1:1.

Synthesis of 2,3,4-Tri-O-acetyl-1-O-{[2-({[7-(diethylamino)-2-oxo-2*H*-chromen-4-yl]methoxycarbonyl}{methyl}amino)ethyl][methyl]carbamoyl}- α -L-arabinopyranoside (17)



Coumarin 17 was synthesized using modified procedures of Wang et al.^[25] Coumarin 7 (57 mg, 0.16 mmol) was dissolved in dry CH₂Cl₂ (2.9 mL) under nitrogen atmosphere. After the addition of 4dimethylaminopyridine (DMAP) (2.0 mg, 16 µmol), carbohydrate 11 (40 mg, 91 µmol) and N,Ndiisopropylethylamine (DIPEA) (0.28 mL, 1.6 mmol) the reaction mixture was stirred for 24 h at room temperature in the dark. Then, it was diluted with CH₂Cl₂, washed with saturated NaHCO₃ and saturated NaCl solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 30:70) to yield a slightly yellow foam (51 mg, 77 µmol, 85%). The product was a mixture of cis and trans isomers on carbamate bonds. ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.19 (t, ³J_{10.9} = 7.1 Hz, 6 H, 10-H), 1.99–2.07 (m, 6 H, CH₃), 2.09–2.16 (m, 3 H, CH₃), 2.88–2.98 (m, 3 H, CH₃b), 2.98–3.07 (m, 3 H, CH₃a), 3.18–3.68 (m, 4 H, 6'-H, 7'-H), 3.40 (q, ${}^{3}J_{9,10} = 7.1$ Hz, 4 H, 9-H), 3.77 (m_c, 1 H, 5"-H_a), 3.98 (dd, ²*J*_{5b,5a} = 12.9 Hz, ³*J*_{5b,4} = 3.7 Hz, 1 H, 5"-H_b), 5.06–5.33 (m, 5 H, 2"-H, 3"-H, 4"-H, 4'-H), 5.52– 5.61 (m, 1 H, 1"-H), 6.01–6.13 (m, 1 H, 3-H), 6.50 (s, 1 H, 8-H), 6.57 (d, ${}^{3}J_{6,5} = 9.0$ Hz, 1 H, 6-H), 7.26– 7.34 (m, 1 H, 5-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 20.7 (CH₃), 20.7 (CH₃), 20.8 (CH₃), 20.9 (CH₃), 21.0 (CH₃), 34.7, 34.7, 34.8, 35.2, 35.5, 35.5, 35.6, 35.8 (CH₃a, CH₃b), 44.9 (C-9), 46.3, 46.4, 46.9, 46.9, 47.1, 47.3, 47.6 (C-6', C-7'), 62.4 (C-4'), 62.5 (C-4'), 62.7 (C-4'), 63.5 (C-5"), 63.6 (C-5"), 64.4 (C-5"), 64.5 (C-5"), 67.2, 67.6, 67.7, 68.2, 68.3 (C-2", C-4"), 69.7 (C-3"), 69.8 (C-3"), 70.3 (C-3"), 70.4 (C-3"), 93.5 (C-1"), 93.6 (C-1"), 93.9 (C-1"), 94.1 (C-1"), 98.0 (C-8), 105.8, 105.9, 106.2, 106.4 (C-3, C-4a), 108.8 (C-6), 124.4 (C-5), 124.5 (C-5), 124.6 (C-5), 150.3, 150.4, 150.5, 150.7 (C-4, C-7), 153.9 (C-8'), 154.1 (C-8'), 154.4 (C-8'), 155.2 (C-5'), 155.3 (C-5'), 155.6 (C-5'), 155.7 (C-5'), 156.3 (C-8a), 162.0 (C-2), 162.1 (C-2), 169.5, 169.7, 169.8, 169.9, 169.9, 169.9, 170.0, 170.2, 170.3, 170.3 (C-2^{*iii*}, C-3^{*iii*}, C-4^{*iii*}); $R_f = 0.25$ (PE/EtOAc 30:70); IR (atr-film): \tilde{v} [cm⁻¹] = 2972, 2930, 1746, 1713, 1605, 1528, 1422, 1371, 1221, 1137, 1088, 1055, 760; m.p.: 82.7 °C; $[\alpha]_{D}^{20} = 10.6$ (c = 1.0, CHCl₃)

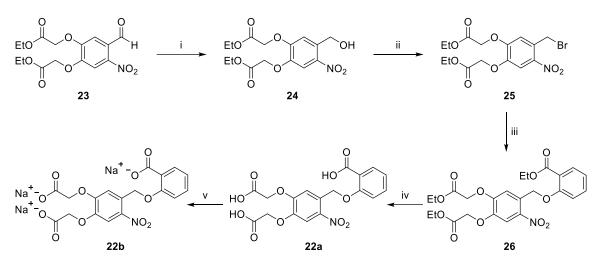
Synthesisof1-O-{[2-({[7-(diethylamino)-2-oxo-2H-chromen-4-yl]methoxycarbonyl}{methyl}amino)ethyl][methyl]carbamoyl}-α-L-arabinopyranoside(2c)



Photocaged arabinose 2c was synthesized using modified procedures of Binder et al.^[16] Coumarin 17 (40 mg, 60 µmol) was dissolved in MeOH (0.20 mL) and stirred at room temperature in the dark. Ammonia in MeOH (7 M, 54 µL, 0.38 mmol) was added and the reaction mixture was stirred until complete conversion. The solvent was evaporated under reduced pressure to yield a yellow solid (28 mg, 52μ mol, 86%). The product was a mixture of cis and trans isomers on carbamate bonds. ¹H-NMR $(600 \text{ MHz}, \text{CDCl}_3): \delta \text{[ppm]} = 1.20 \text{ (t, } {}^{3}J_{10.9} = 7.1 \text{ Hz}, 6 \text{ H}, 10\text{-H}), 2.92\text{--}2.98 \text{ (m, 3 H, CH}_3\text{b}), 2.99\text{--}3.05$ (m, 3 H, CH₃a), 3.05-3.99 (m, 4 H, 6'-H, 7'-H), 3.41 (q, ${}^{3}J_{9,10} = 7.1$ Hz, 4 H, 9-H), 3.63-3.70 (m, 1 H, 5"-Ha), 3.70–3.78 (m, 1 H, 3"-H), 3.78–3.92 (m, 1 H, 2"-H), 3.92–3.99 (m, 1 H, 4"-H), 3.99–4.09 (m, 1 H, 5"-H_b), 5.14–5.32 (m, 2 H, 4'-H), 5.32–5.46 (m, 1 H, 1"-H), 6.03–6.17 (m, 1 H, 3-H), 6.52 (s, 1 H, 8-H), 6.57–6.68 (m, 1 H, 6-H), 7.27–7.36 (m, 1 H, 5-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 12.5 (C-10), 34.5, 34.6, 34.9, 35.4, 35.8, 36.2 (CH₃a, CH₃b), 45.0 (C-9), 46.5, 46.7, 47.1, 47.4, 47.5, 47.5 (C-6', C-7'), 62.4 (C-4'), 62.5 (C-4'), 62.9 (C-4'), 63.1 (C-4'), 66.6 (C-5"), 66.8 (C-5"), 66.8 (C-5"), 68.2 (C-4"), 70.6 (C-2"), 70.8 (C-2"), 71.0 (C-2"), 73.3 (C-3"), 73.4 (C-3"), 73.6 (C-3"), 96.4 (C-1"), 96.5 (C-1"), 96.6 (C-1"), 98.0 (C-8), 105.9, 106.0 (C-3, C-4a), 109.1 (C-6), 124.5 (C-5), 124.6 (C-5), 150.4, 150.5, 151.4 (C-4, C-7), 154.6 (C-8'), 154.7 (C-8'), 156.0, 156.0, 156.2, 156.3, 156.6 (C-5', C-8a), 162.6 (C-2), 162.7 (C-2); $R_f = 0.08$ (EtOAc/MeOH 95:5); IR (atr-film): \tilde{v} [cm⁻¹] = 3423, 2966, 2927, 1703, 1603, 1528, 1490, 1423, 1357, 1275, 1216, 1131, 1080, 827, 761; HRMS (ESI): m/z calculated for $C_{25}H_{36}N_3O_{10}^+$ [M+H]⁺: 538.2395; found: 538.2397; m.p.: 67–73 °C; $[\alpha]_D^{20} = 5.0$ (c = 1.0, CHCl₃); UV-Vis [Tris buffer (20 mM, pH 7.5)/MeCN 1:1]: λ_{max} (ϵ) = 273 nm (28656 dm³ mol⁻¹ cm⁻¹), 385 (14070).

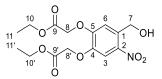
reversed-phase HPLC: $t_R = 18.3$ min; column: *HyperClone* 5 μ ODS (C18) 120 (*Phenomenex*); detection (UV): 385 nm; eluent: H₂O/MeOH 55:45; flowrate: 0.5 mL/min; column temperature: 25 °C; sample solvent: Tris buffer (20 mM, pH 7.5)/MeCN 1:1.

S3.8 Synthetic scheme for preparation of photocaged salicylic acid 22a and the corresponding sodium form 22b



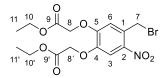
Scheme S8: Synthetic scheme for preparation of photocaged salicylic acid 22a and the corresponding sodium form 22b. Reagents and conditions: i) NaBH₄, CH₂Cl₂, EtOH, AcOH, 0 °C, 3 h; ii) CBr₄, PPh₃, CH₂Cl₂, 0 °C \rightarrow RT, 6 h; iii) ethyl salicylate, K₂CO₃, acetone, RT, 2 d; iv) KOH (0.2 M), MeOH, 60 °C, 4 h; v) NaOH (0.2 M), MeOH, RT, 5 min.

Synthesis of 4,5-Bis(ethoxycarbonylmethoxy)-2-nitrobenzylalcohol (24)



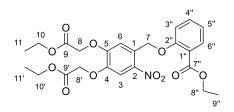
Alcohol **24** was synthesized using a procedure of Ni *et al.*^[26] 4,5-Bis(ethoxycarbonylmethoxy)-2nitrobenzaldehyde (**23**) (881 mg, 2.48 mmol) was dissolved in a mixture of CH₂Cl₂, EtOH and acetic acid (35:5:1, 10.0 mL). After the solution was cooled to 0 °C NaBH₄ (188 mg, 4.96 mmol, 2.00 Äq.) was added and the reaction mixture was stirred for 3 h at 0 °C. The reaction was quenched by addition of 1 M HCl (2 mL), diluted with ethyl acetate and washed with saturated NaCl solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (toluene/ethyl acetate 80:20) to yield a yellow solid (649 mg, 1.82 mmol, 73%). The spectroscopic data are consistent with previously reported literature values.^[26] ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.30 (t, ³J_{11,10 or 11',10'} = 7.2 Hz, 3 H, 11-H or 11'-H), 1.31 (t, ³J_{11,10 or 11',10'} = 7.2 Hz, 3 H, 11-H or 11'-H), 2.13 (br, 1 H, OH), 4.28 (q, ³J_{10,11 or 10',11'} = 7.2 Hz, 2 H, 10-H or 10'-H), 4.28 (q, ³J_{10,11 or 10',11'} = 7.2 Hz, 2 H, 10-H or 10'-H), 4.77 (s, 2 H, 8-H or 8'-H), 4.83 (s, 2 H, 8-H or 8'-H), 4.94 (s, 2 H, 7-H), 7.17 (s, 1 H, 3-H or 6-H), 7.70 (s, 1 H, 3-H or 6-H); ¹³C-NMR (151 MHz, CDCl₃): δ [ppm] = 14.3, 14.3 (C-11, C-11'), 61.8, 61.9 (C-10, C-10'), 62.5 (C-7), 66.2 (C-8'), 66.6 (C-8), 112.0, 113.5 (C-3, C-6), 133.6, 140.4 (C-1, C-2), 146.5 (C-5), 152.7 (C-4), 167.8 (C-9'), 168.1 (C-9); R_f = 0.32 (Toluol/EtOAc 80:20); IR (atr-film): $\tilde{\nu}$ [cm⁻¹] = 2992, 1742, 1580, 1507, 1282, 1193, 1072, 1019, 792; MS (ESI, positive ion): m/z (%) = 380.2 (100) [M+Na]⁺, 737.3 (5) [2M+Na]⁺; m.p.: 76 °C.

Synthesis of 4,5-Bis(ethoxycarbonylmethoxy)-2-nitrobenzyl bromide (25)



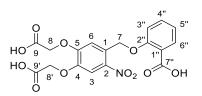
Bromide 25 was synthesized using a procedure of Tietze et al.^[27] A Schlenk tube was charged with Alcohol 24 (1.00 g, 2.80 mmol) and tetrabromomethane (CBr₄) (1.16 g, 3.50 mmol) dissolved in dry CH₂Cl₂ (14.0 mL) under nitrogen atmosphere in the dark. A second Schlenk tube was charged with triphenylphosphane (PPh₃) (918 mg, 3.50 mmol) dissolved in dry CH₂Cl₂ under nitrogen atmosphere. Both solutions were cooled to 0 $^{\circ}$ C and the cooled PPh₃ solution was added dropwise to the dissolved alcohol 24. The reaction mixture was stirred for 10 min at 0 °C and 6 h at room temperature. SiO₂ was added to the reaction mixture and the solvent was removed under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 65:35) to yield a white solid (1.13 g, 2.69 mmol, 96%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.31 (t, ³J_{11.10} = 7.1 Hz, 3 H, 11-H), 1.31 (t, ${}^{3}J_{11',10'} = 7.1$ Hz, 3 H, 11'-H), 4.29 (q, ${}^{3}J_{10,11;10',11'} = 7.1$ Hz, 4 H, 10-H, 10'-H), 4.78 (2, 2 H, 8'-H), 4.81 (s, 2 H, 7-H), 4.83 (s, 2 H, 8-H), 6.94 (s, 1 H, 6-H), 7.65 (s, 1 H, 3-H); ¹³C-NMR (151 MHz, $CDCl_3$): δ [ppm] = 14.3 (C-11), 14.3 (C-11'), 29.7 (C-7), 61.9 (C-10), 62.0 (C-10'), 66.5 (C-8), 66.5 (C-8'), 112.1 (C-3), 117.2 (C-6), 128.5 (C-2), 141.2 (C-1), 147.6 (C-5), 151.9 (C-4), 167.7 (C-9), 167.8 (C-9'); $R_f = 0.62$ (PE/EtOAc 60:40); IR (atr-film): \tilde{v} [cm⁻¹] = 2992, 1739, 1616, 1581, 1522, 1479, 1449, 1409, 1380, 1356, 1339, 1289, 1264, 1202, 1184, 1120, 1080, 1043, 1025, 930, 881, 798, 758, 732, 672; HRMS (ESI): m/z calculated for C₁₅H₁₈O₈NBrNa⁺ [M+Na]⁺: 442.0108; found: 442.0108; m.p.: 118°C.

Synthesis of Ethyl 2-O-[4,5-bis(ethoxycarbonylmethoxy)-2-nitrobenzyl]salicylate (26)



Ethyl salicylate (0.11 mL, 0.71 mmol) was dissolved in dry acetone (1.0 mL) and dry K₂CO₃ (56 mg, 0.40 mmol) was added. After stirring for 10 min bromide 25 (0.10 g, 0.24 mmol) was added and the reaction mixture was stirred for 2 d at room temperature in the dark. After completion, the solvent was removed under reduced pressure. The residue was dissolved in ethyl acetate and subsequently washed with water and saturated NaHCO₃ solution. The organic phase was dried with anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash-column chromatography on SiO₂ (petroleum ether/ethyl acetate 80:20) to yield a white solid (0.11 g, 0.22 mmol, 92%). ¹H-NMR (600 MHz, CDCl₃): δ [ppm] = 1.25 (t, ${}^{3}J_{11,10}$ = 7.1 Hz, 3 H, 11-H), 1.31 (t, ${}^{3}J_{11',10'}$ = 7.1 Hz, 3 H, 11'-H), 1.40 (t, ${}^{3}J_{9'',8''}$ = 7.1 Hz, 3 H, 9"-H), 4.23 (q, ${}^{3}J_{10,11}$ = 7.1 Hz, 2 H, 10-H), 4.29 (q, ${}^{3}J_{10',11'}$ = 7.1 Hz, 2 H, 10'-H), 4.38 (q, ${}^{3}J_{8'',9''} = 7.1$ Hz, 2 H, 8"-H), 4.81 (s, 2 H, 8'-H), 5.05 (s, 2 H, 8-H), 5.52 (s, 2 H, 7-H), 7.03 (dd, ${}^{3}J_{5'',4''} = 7.5$ Hz, ${}^{3}J_{5'',6''} = 7.5$ Hz, 1 H, 5"-H), 7.14 (d, ${}^{3}J_{3'',4''} = 8.4$ Hz, 1 H, 3"-H), 7.53 (dd, ${}^{3}J_{4'',3''}$ = 8.4 Hz, ${}^{3}J_{4'',5''}$ = 7.5 Hz, 1 H, 4"-H), 7.82 (s, 1 H, 3-H), 7.93 (d, ${}^{3}J_{6'',5''}$ = 7.5 Hz, 1 H, 6"-H), 8.23 (s, 1 H, 6-H); 13 C-NMR (151 MHz, CDCl₃): δ [ppm] = 14.2 (C-11), 14.3 (C-11'), 14.5 (C-9''), 60.7 (C-8''), 61.6 (C-10), 61.7 (C-10'), 65.8 (C-8), 66.8 (C-8'), 67.3 (C-7), 111.9 (C-3), 112.8 (C-6), 113.2 (C-3"), 119.8 (C-1"), 120.8 (C-5"), 130.8 (C-1), 132.1 (C-6"), 134.2 (C-4"), 138.9 (C-2), 146.1 (C-4), 153.4 (C-5), 158.0 (C-2"), 165.2 (C-7"), 168.0 (C-9), 168.3 (C-9'); $R_f = 0.23$ (PE/EtOAc 70:30); IR (atr-film): \tilde{v} [cm⁻¹] = 3104, 2992, 1771, 1743, 1713, 1582, 1524, 1488, 1449, 1425, 1377, 1329, 1291, 1242, 1195, 1112, 1080, 1020, 895, 859, 826, 803, 753, 700, 683, 661; HRMS (ESI): m/z calculated for C₂₄H₂₇O₁₁NNa⁺ [M+Na]⁺: 528.1476; found: 528.1476; m.p.: 133 °C

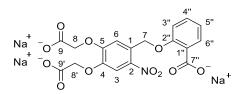
Synthesis of 2-O-[4,5-Bis(carboxymethoxy)-2-nitrobenzyl]salicylic acid (BC-cSal) (22a)



To a solution of salicylate **26** (200 mg, 396 µmol) in MeOH (11.9 mL) a 0.2 M solution of KOH (11.9 mL, 2.37 mmol) was added. The reaction mixture was heated to 60 °C and stirred until complete conversion (4 h). After the reaction was completed as indicated by TLC, 1 M HCl was added and the precipitate was filtered off and washed. The precipitate was dried to yield a white solid (153 mg, 363 µmol, 92%). ¹H-NMR (600 MHz, DMSO-*d*₆): δ [ppm] = 4.89 (s, 2 H, 8'-H), 4.91 (s, 2 H, 8-H), 5.49 (s, 2 H, 7-H), 7.06 (dd, ³*J*_{5",6"} = 7.0 Hz, ³*J*_{5",4"} = 6.9 Hz 1 H, 5"-H), 7.22 (d, ³*J*_{3",4"} = 7.9 Hz, 1 H, 3"-H), 7.56 (dd, ³*J*_{4",3"} = 7.9 Hz, ³*J*_{4",5"} = 6.9 Hz, 1 H, 4"-H), 7.72 (s, 1 H, 3-H), 7.79 (d, ³*J*_{6",5"} = 7.0 Hz, 1 H, 6"-H), 7.80 (s, 1 H, 6-H); ¹³C-NMR (151 MHz, DMSO-*d*₆): δ [ppm] = 65.0 (C-8), 65.5 (C-8'), 66.7 (C-7), 110.4 (C-3), 112.2 (C-6), 113.7 (C-3"), 120.7 (C-1"), 120.7 (C-5"), 128.7 (C-1), 131.6 (C-6"), 133.7 (C-4"), 139.0 (C-2), 145.9 (C-4), 152.2 (C-5), 157.1 (C-2"), 166.9 (C-7"), 169.1 (C-9), 169.8 (C-9'); IR (atr-film): $\tilde{\nu}$ [cm⁻¹] = 2923, 1708, 1801, 1585, 1525, 1490, 1428, 1380, 1331, 1285, 1245, 1214, 1079, 1027, 900, 849, 821, 750, 698, 671; HRMS (ESI): m/z calculated for C₁₈H₁₅O₁₁NK⁺ [M+K]⁺: 460.0276; found: 460.0276; m.p.: 264 °C (decay); UV-Vis [sodium phosphate buffer (100 mM, pH 7.4)]: λ_{max} (ε) = 290 nm (5027 dm³ mol⁻¹ cm⁻¹), 346 (5852).

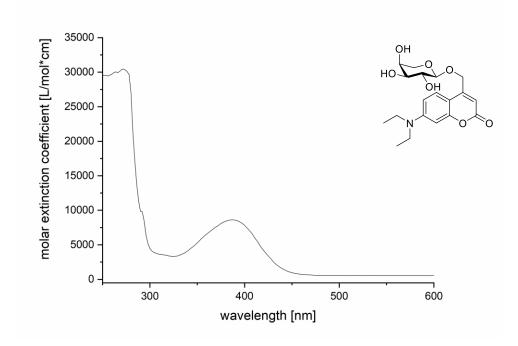
reversed-phase HPLC: $t_R = 9.6$ min; column: *HyperClone* 5 μ ODS (C18) 120 (*Phenomenex*); detection (UV): 346 nm; eluent: sodium phosphate buffer (100 mM, pH 7.4)/MeOH 15:85; flowrate: 0.5 mL/min; column temperature: 25 °C; sample solvent: sodium phosphate buffer (100 mM, pH 7.4).

Synthesis of Trisodium 2-O-[4,5-bis(carboxymethoxy)-2-nitrobenzyl]salicylate (22b)



To a solution of salicylic acid **22a** (60 mg, 0.14 mmol) in MeOH (2.5 mL) a 0.2 M solution of NaOH (2.1 mL, 0.43 mmol) was added. The reaction mixture was stirred for 5 min before it was lyophilised overnight to yield a solid (69 mg, 0.14 mmol, quant.). ¹H-NMR (600 MHz, D₂O): δ [ppm] = 4.56 (s, 2 H, 8'-H), 4.62 (s, 2 H, 8-H), 5.43 (s, 2 H, 7-H), 7.03 (d, ${}^{3}J_{3",4"} = 8.2$ Hz, 1 H, 3"-H), 7.06 (ddd, ${}^{3}J_{5",6"} = 7.5$ Hz, ${}^{3}J_{5",4"} = 7.4$ Hz, ${}^{4}J_{5",3"} = 1.0$ Hz, 1 H, 5"-H), 7.36 (ddd, ${}^{3}J_{4",3"} = 8.2$ Hz, ${}^{3}J_{4",5"} = 7.4$ Hz, ${}^{4}J_{4",6"} = 1.8$ Hz, 1 H, 4"-H), 7.41 (s, 1 H, 3-H), 7.46 (dd, ${}^{3}J_{6",5"} = 7.5$ Hz, ${}^{3}J_{6",4"} = 1.8$ Hz, 1 H, 6"-H), 7.74 (s, 1 H, 6-H); 13 C-NMR (151 MHz, D₂O): δ [ppm] = 67.3, 67.3 (C-8, C-8'), 67.6 (C-7), 109.4 (C-3), 111.4 (C-6), 113.8 (C-3"), 121.4 (C-5"), 128.3 (C-6"), 129.5 (C-1), 129.7 (C-4"), 130.2 (C-1"), 139.0 (C-2), 145.7 (C-4), 152.1 (C-5), 154.1 (C-2"), 174.9 (C-9), 175.4 (C-9'), 176.0 (C-7"); IR (atr-film): $\tilde{\nu}$ [cm⁻¹] = 3216, 1603, 1553, 1521, 1425, 1382, 1329, 1273, 1214, 1100, 1070, 1020, 855, 818, 753, 695, 663; m.p.: 245 °C (decay).

S4 Supporting data



S4.1 UV-Vis spectra of compounds

Figure S1: UV-Vis spectrum of compound 2b (0.10 mM in MeOH, 25 °C).

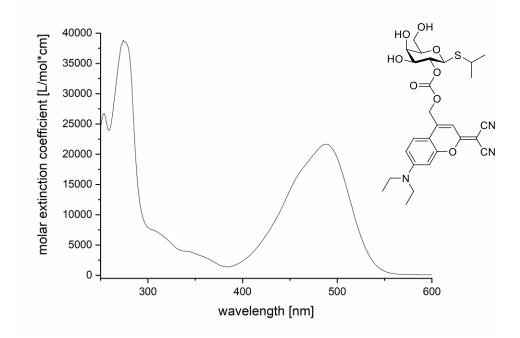


Figure S2: UV-Vis spectrum of compound 1c [0.05 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50, 25 °C].

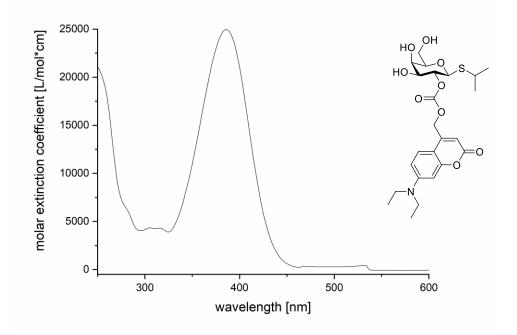


Figure S3: UV-Vis spectrum of compound 1b [25.0 µM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50, 25 °C].

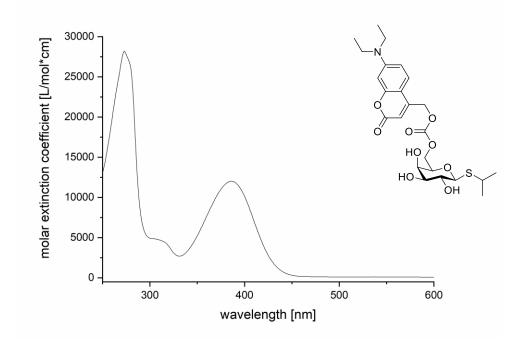


Figure S4: UV-Vis spectrum of compound 1d [0.05 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50, 25 °C].

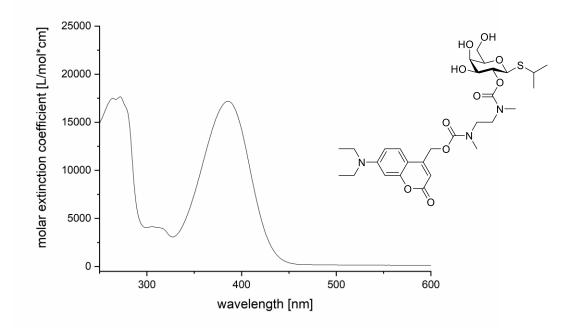


Figure S5: UV-Vis spectrum of compound 1e [0.05 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50, 25 °C].

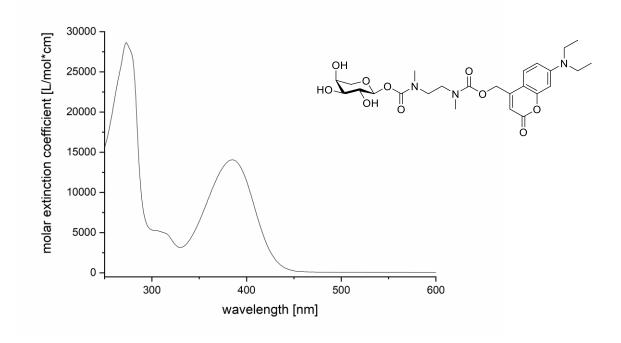


Figure S6: UV-Vis spectrum of compound 2c [0.05 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50, 25 °C].

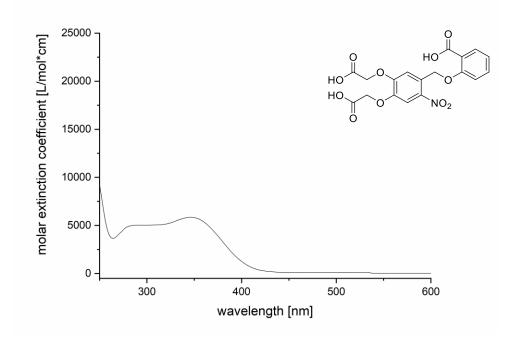


Figure S7: UV-Vis spectrum of compound 22a [125 µM in sodium phosphate buffer (100 mM, pH 7.4), 25 °C].

S4.2 Photon flux densities of light sources

| wavelength | q _{n,p} (mol s⁻¹) | | |
|------------|----------------------------|--|--|
| 365 nm | 6.49 × 10 ⁻⁸ | | |
| 405 nm | 1.91 × 10 ⁻⁷ | | |
| 430 nm | 1.22 × 10 ⁻⁷ | | |

Table S3: Determined photon flux densities $(q_{n,p})$

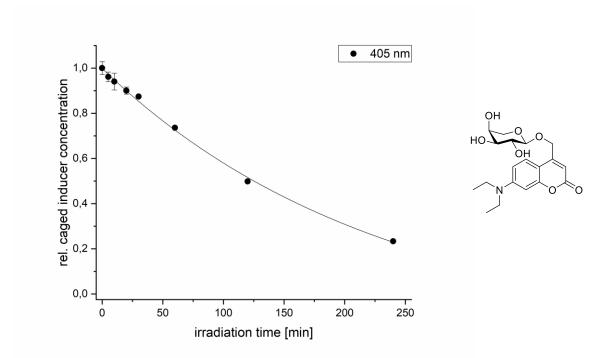


Figure S8: *In vitro* decay of **2b** (1 mM in H₂O/DMSO 99:1) via reversed-phase HPLC after irradiation with 405 nm (44.6 mW cm⁻², room temperature).

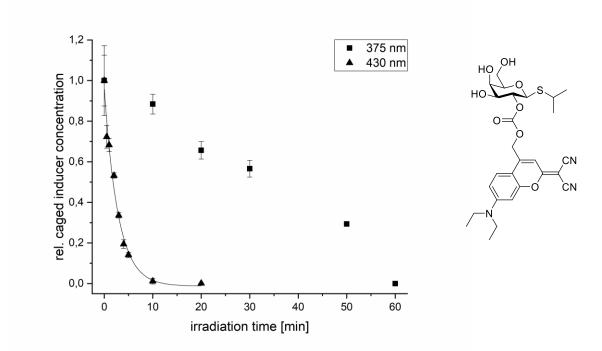


Figure S9: *In vitro* decay of **1c** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 1:1] via reversed-phase HPLC after irradiation with 375 nm (6.4 mW cm⁻², room temperature) and 430 nm (45.6 mW cm⁻², room temperature).

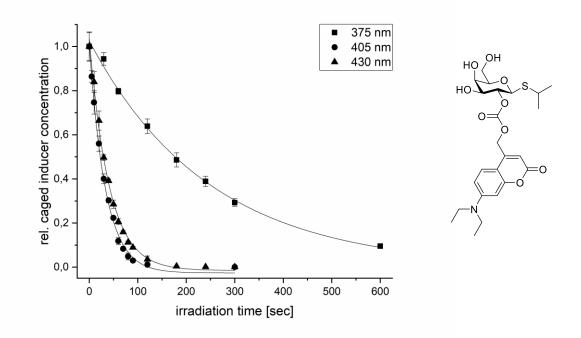


Figure S10: *In vitro* decay of **1b** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 1:1] via reversed-phase HPLC after irradiation with 375 nm (6.4 mW cm⁻², room temperature), 405 nm (44.6 mW cm⁻², room temperature) and 430 nm (45.6 mW cm⁻², room temperature).

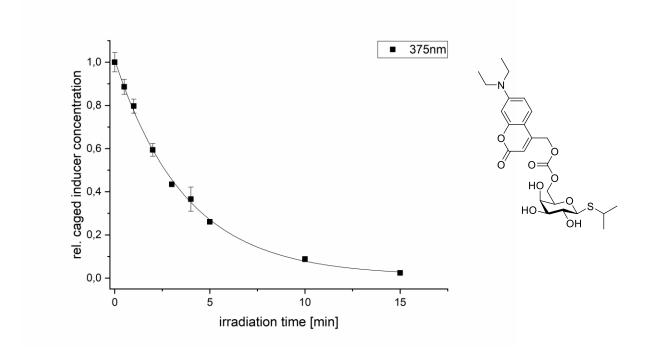


Figure S11: *In vitro* decay of **1d** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 1:1] via reversed-phase HPLC after irradiation with 375 nm (6.4 mW cm⁻², room temperature).

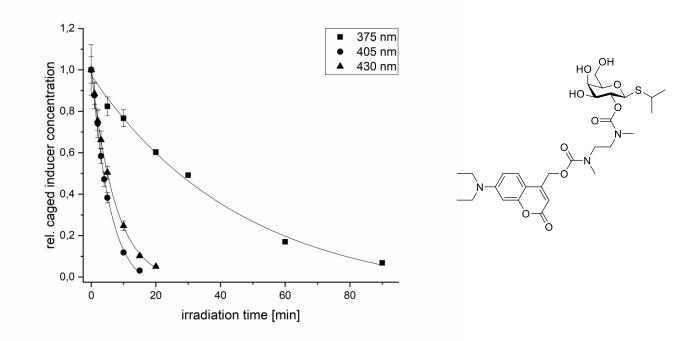


Figure S12: *In vitro* decay of **1e** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 1:1] via reversed-phase HPLC after irradiation with 375 nm (6.4 mW cm⁻², room temperature), 405 nm (44.6 mW cm⁻², room temperature) and 430 nm (45.6 mW cm⁻², room temperature).

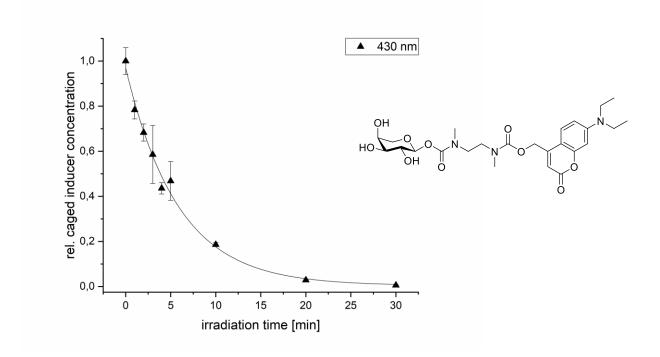


Figure S13: *In vitro* decay of **2c** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 1:1] via reversed-phase HPLC after irradiation with 430 nm (45.6 mW cm⁻², room temperature).

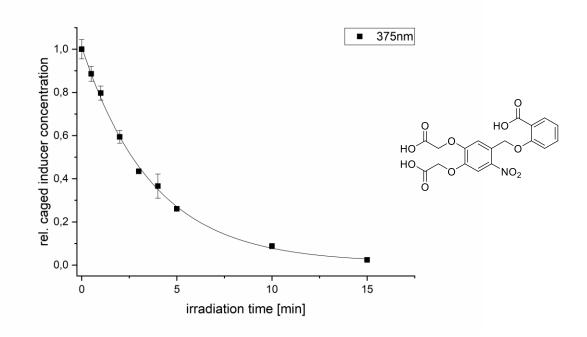


Figure S14: *In vitro* decay of **22a** [0.5 mM in sodium phosphate buffer (100 mM, pH 7.4)] via reversed-phase HPLC after irradiation with 375 nm (6.4 mW cm⁻², room temperature).

| photocaged inducer | λ [nm] | Уo | A ₁ | t ₁ | k | T |
|--------------------|--------|-----------|-----------------------|----------------|---------|-----------|
| 2b | 375 | -0,19104 | 1,18966 | 230,51535 | 0,00434 | 159,78106 |
| 1c | 430 | -0,01254 | 0,97483 | 2,83993 | 0,35212 | 1,96849 |
| 1b | 375 | 0,0004873 | 1,02624 | 246,92187 | 0,00405 | 171,1532 |
| 1b | 405 | -0,02632 | 1,03705 | 33,42057 | 0,02992 | 23,16538 |
| 1b | 430 | -0,01504 | 1,05356 | 40,75615 | 0,02454 | 28,25001 |
| 1d | 375 | 0,01084 | 1,00002 | 3,7118 | 0,26941 | 2,57283 |
| 1e | 375 | -0,09535 | 1,06979 | 46,27316 | 0,02161 | 32,07411 |
| 1e | 405 | -0,06395 | 1,09129 | 5,79684 | 0,17251 | 4,01806 |
| 1e | 430 | -0,02656 | 1,03055 | 7,48242 | 0,13365 | 5,18642 |
| 2c | 430 | 0,00417 | 0,96575 | 5,79819 | 0,17247 | 4,019 |
| 22a | 375 | -0,00435 | 0,9935 | 3,52342 | 0,28382 | 2,44225 |

Table S3: Fitting parameters for 1b-e, 2b-c and 22a at different wavelength.

S4.4 HPLC-Traces

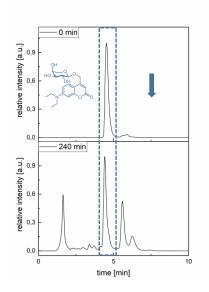


Figure S15: UV traces at 388 nm of the reversed-phase HPLC analysis of **2b** (1 mM in H₂O/DMSO 99:1) before irradiation and after 240 min of irradiation at 405 nm (44.6 mW cm⁻², room temperature).

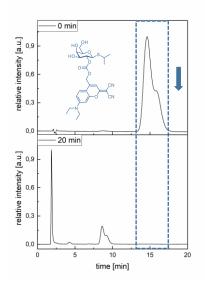


Figure S16: UV traces at 488 nm of the reversed-phase HPLC analysis of **1c** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation and after 20 min of irradiation at 430 nm (45.6 mW cm⁻², room temperature).

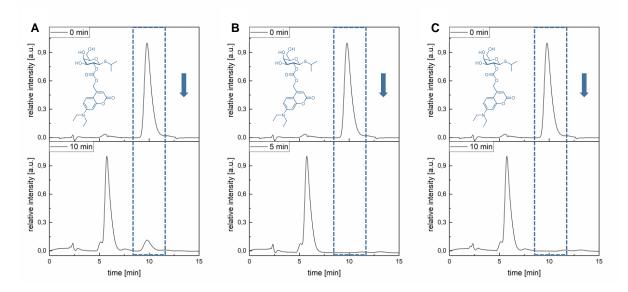


Figure S17: **A**) UV traces at 392 nm of the reversed-phase HPLC analysis of **1b** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation and after 10 min of irradiation at 375 nm (6.4 mW cm⁻², room temperature); **B**) UV traces at 392 nm of the reversed-phase HPLC analysis of **1b** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation and after 5 min of irradiation at 405 nm (44.6 mW cm⁻², room temperature); **C**) UV traces at 392 nm of the reversed-phase HPLC analysis of **1b** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation at 430 nm (45.6 mW cm⁻², room temperature).

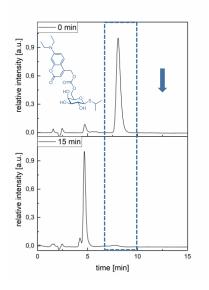


Figure S18: UV traces at 386 nm of the reversed-phase HPLC analysis of **1d** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation and after 15 min of irradiation at 375 nm (6.4 mW cm⁻², room temperature).

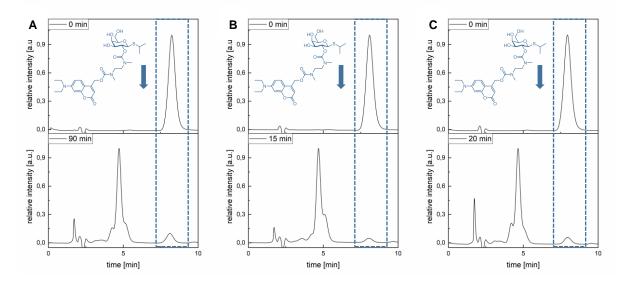


Figure S19: **A**) UV traces at 386 nm of the reversed-phase HPLC analysis of **1e** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation and after 10 min of irradiation at 375 nm (6.4 mW cm⁻², room temperature); **B**) UV traces at 386 nm of the reversed-phase HPLC analysis of **1e** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation and after 5 min of irradiation at 405 nm (44.6 mW cm⁻², room temperature); **C**) UV traces at 386 nm of the reversed-phase HPLC analysis of **1e** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation at 430 nm (45.6 mW cm⁻², room temperature).

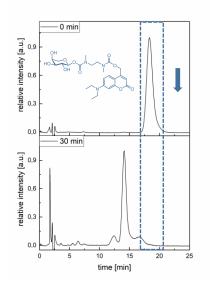


Figure S20: UV traces at 385 nm of the reversed-phase HPLC analysis of **2c** [0.5 mM in Tris buffer (20 mM, pH 7.5)/MeCN 50:50] before irradiation and after 10 min of irradiation at 430 nm (45.6 mW cm⁻², room temperature).

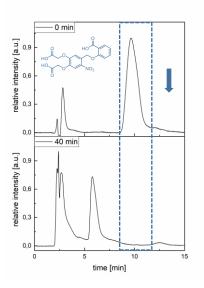


Figure S21: UV traces at 346 nm of the reversed-phase HPLC analysis of **22a** [0.5 mM in sodium phosphate buffer (100 mM, pH 7.4)] before irradiation and after 15 min of irradiation at 375 nm (6.4 mW cm⁻², room temperature).

S4.5 Stability measurements

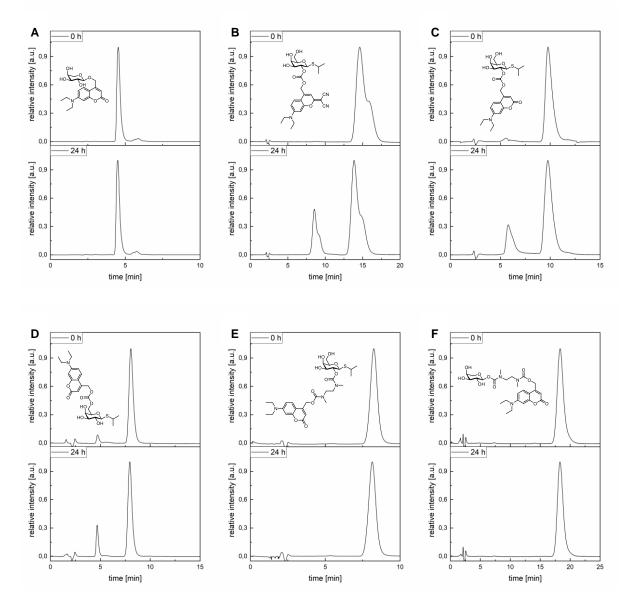


Figure S22: UV traces at absorption maxima of the reversed-phase HPLC analysis of 2b (A), 1c (B), 1b (C), 1d (D), 1e (E) and 2c (F) for determination of stability. The reported values in Table 1 are means of triplicate measurements.

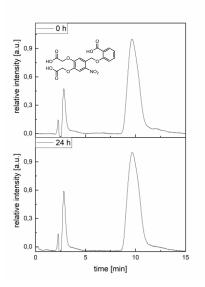


Figure S23: UV traces at absorption maxima of the reversed-phase HPLC analysis of **22a** for determination of stability. The reported value is the mean of triplicate measurements.

S4.6 ESI measurements

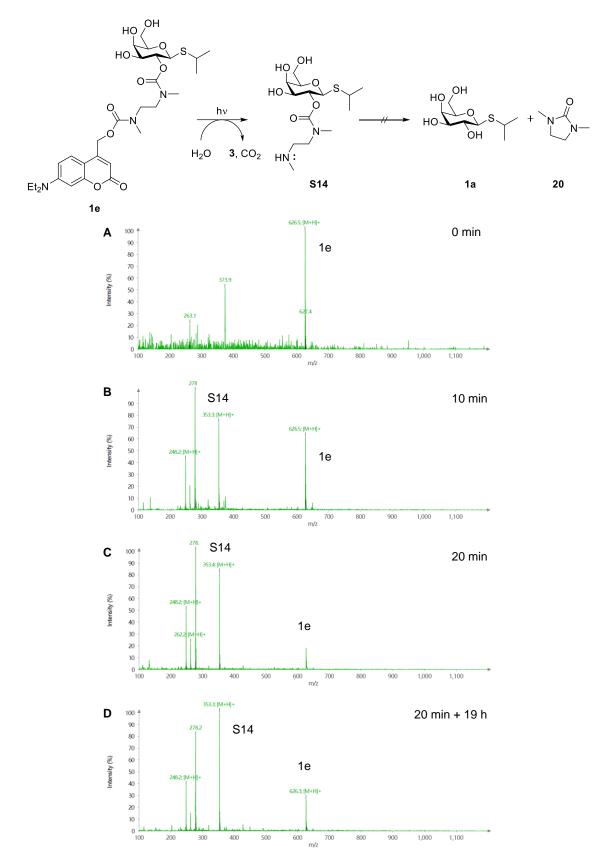


Figure S24: ESI-MS measurements of **1e** (1 mM in MeOH/H₂O 1:1) for detection of the intermediate **S14** after irradiation with 430 nm (45.6 mW cm⁻², room temperature) for **A**) 0 min; **B**) 10 min; **C**) 20 min; **D**) 20 min plus allowing it to stand for 19 h at room temperature.

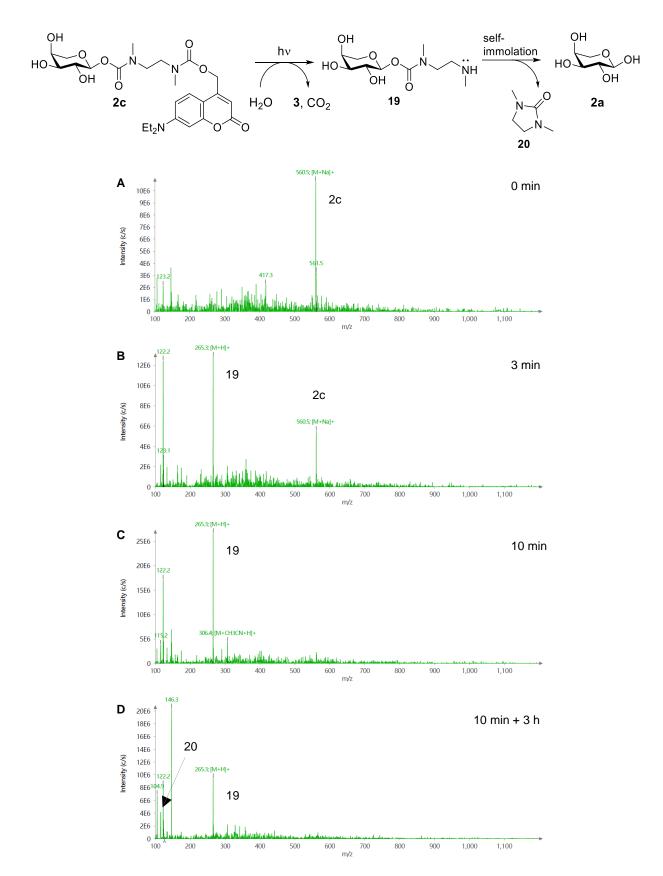
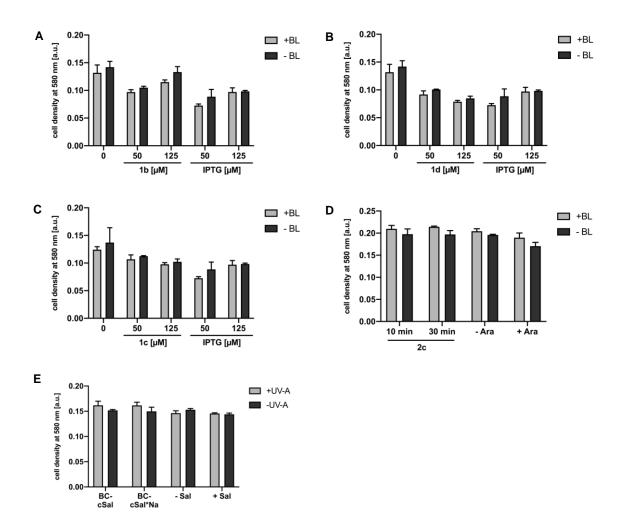


Figure S25: ESI-MS measurements of **2c** (1 mM in MeOH/H₂O 1:1) for monitoring of the intermediate **19** after irradiation with 430 nm (45.6 mW cm⁻², room temperature) for **A**) 0 min; **B**) 3 min; **C**) 10 min; **D**) 10 min plus allowing it to stand for 3 h at room temperature.



S4.7 Toxicity of both the novel photocaged inducer variants and the light exposure

Figure S26: A-C: Growth of E. coli Tuner (DE3)/pRhotHi-2-lacI-eYFP expression cultures in the presence of the novel photocaged IPTG variants 1b (A), 1d (B) or 1c (C) compared to uninduced (0 µM) and induced (50 or 125 µM IPTG) cultures. Cells were grown in LB medium at 30 °C and 1200 rpm over 20 h using a ThermoMixer C (Eppendorf, Germany). Cell growth was analysed by determining the optical density at 580 nm. After 2.5 h, formation of photoproducts was induced in cultures via blue light exposure at 447 nm for 10 min (BL; ~10 mW cm⁻²) and conventional IPTG (1a) was added manually. D: Growth of E. coli LMG194/pBTBX-2-mCherry expression cultures in the presence of the novel photocaged arabinose variant 2c (50 μM) compared to uninduced (-Ara; 0 μM) and induced (+Ara; 50 μM) cultures. Cells were grown in LB medium at 37 °C and 1200 rpm over 20 h using a ThermoMixer C (Eppendorf, Germany). Cell growth was analysed by determining the optical density at 580 nm. After 2.5 h, formation of photoproducts was induced in cultures via blue light exposure at 447 nm for 10 min or 30 min (BL; ~10 mW cm⁻²) and conventional arabinose (2a) was added manually. E: Growth of E. coli Tuner(DE3)/pBNTmcs-mCherry expression cultures in the presence of the novel photocaged salicylic acid (Sal) variants BC-cSal (22a) and BC-cSal*Na (22b) compared to uninduced (-Sal; 0 µM) and induced (+ Sal; 1000 µM Sal) cultures. Cells were grown in LB medium at 30 °C and 1200 rpm over 20 h using a ThermoMixer C (Eppendorf, Germany). Cell growth was analysed by determining the optical density at 580 nm. After 2.5 h formation of photoproducts was induced in cultures via light exposure at 365 nm for 30 min (~1 mW cm⁻²) and conventional Sal was added manually. Values are means of biological triplicate measurements and error bars indicate the respective standard deviation.

S4.8 In vivo results of photocaged IPTG 1e

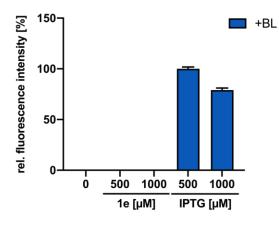


Figure S27: Normalized *in vivo* eYFP fluorescence intensity of *E. coli* Tuner (DE3)/pRhotHi-2-lacI-eYFP expression cultures supplemented with 500 μ M or 1000 μ M of the photocaged compound **1e**. All cultures were incubated in the dark for 20 h in LB medium at 30 °C and light-mediated induction of reporter gene expression was performed after 2.5 h by blue light exposure at 447 nm (BL; ~90 mW cm²) for 10 min or the addition of respective amounts of conventional IPTG (**1a**). *In vivo* fluorescence intensities were determined by using a BioLector system ($\lambda_{ex} = 508$ nm, $\lambda_{em} = 532$ nm), normalized to cell densities and are shown in relation to the respective fluorescence intensities of a culture induced with conventional IPTG (**1a**). Values are means of triplicate measurements. Error bars indicate the respective standard deviations.

S4.9 NMR spectra of compounds

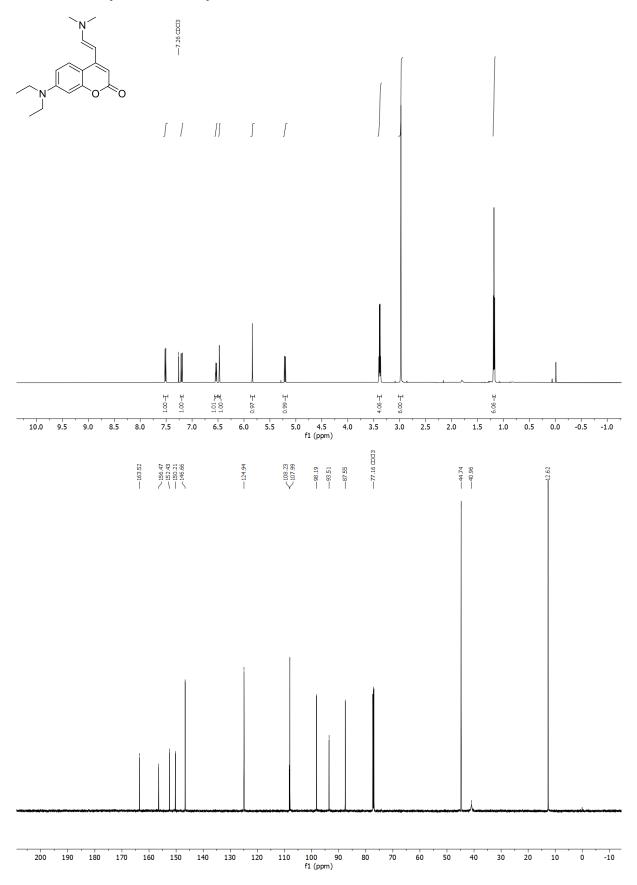


Figure S29: ¹H- and ¹³C-NMR spectra of S2 in CDCl₃ (600 MHz/151 MHz).

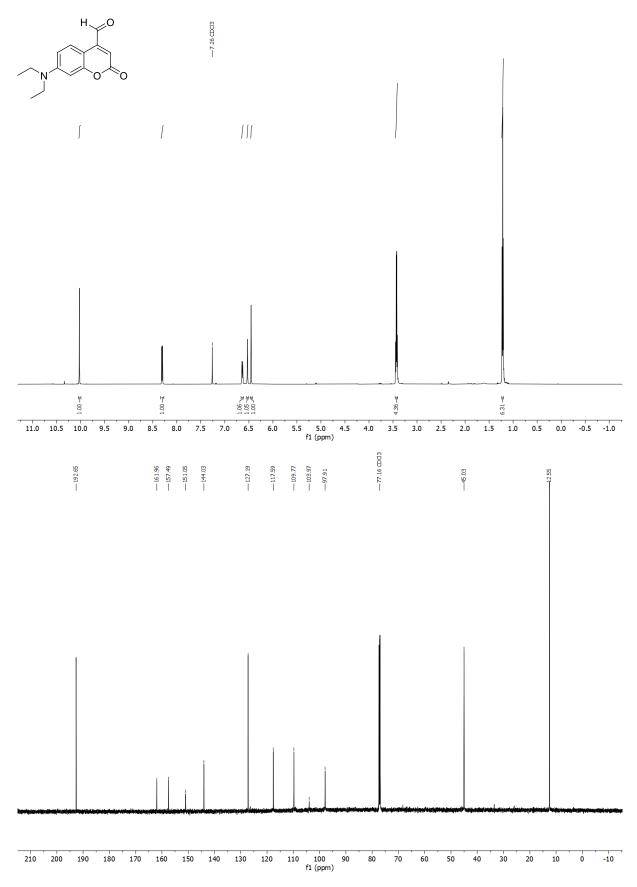


Figure S30: ¹H- and ¹³C-NMR spectra of S3 in CDCl₃ (600 MHz/151 MHz).

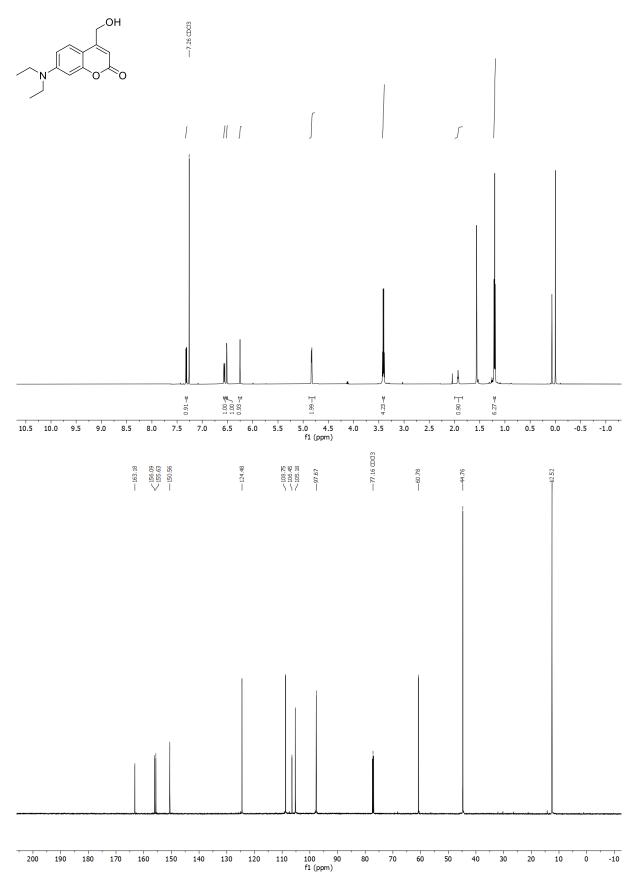


Figure S31: ¹H- and ¹³C-NMR spectra of 3 in CDCl₃ (600 MHz/151 MHz).

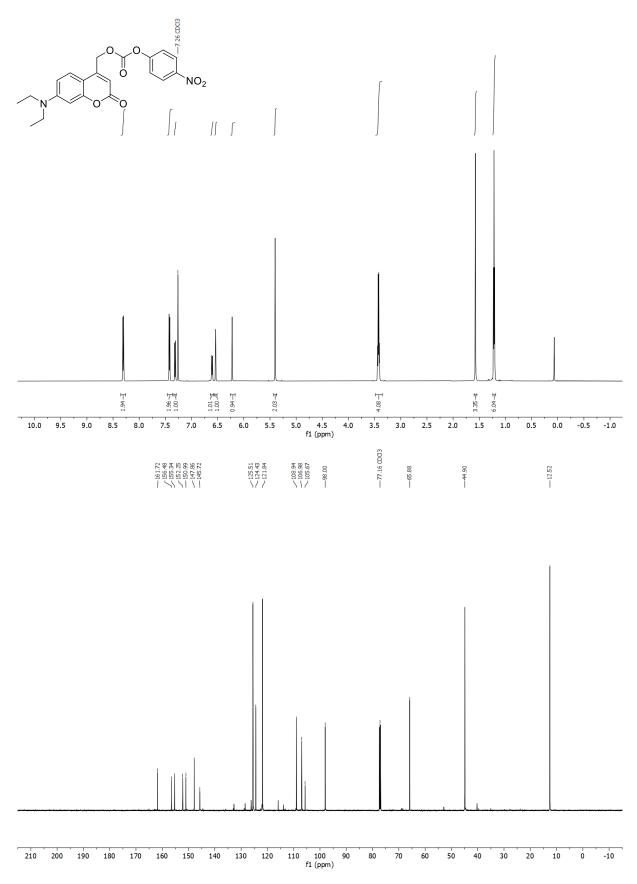


Figure S32: ¹H- and ¹³C-NMR spectra of 5 in CDCl₃ (600 MHz/151 MHz).

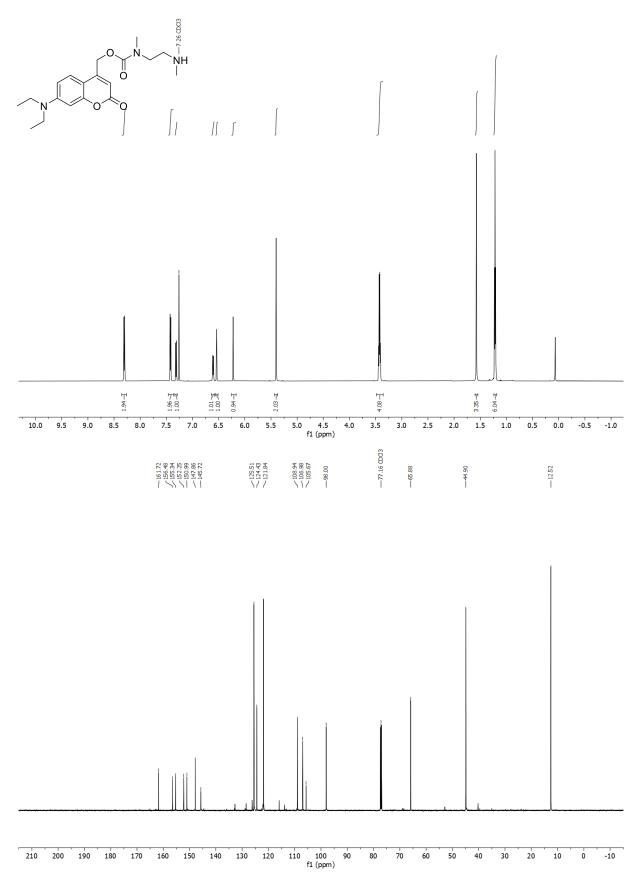


Figure S33: ¹H- and ¹³C-NMR spectra of 7 in CDCl₃ (600 MHz/151 MHz).

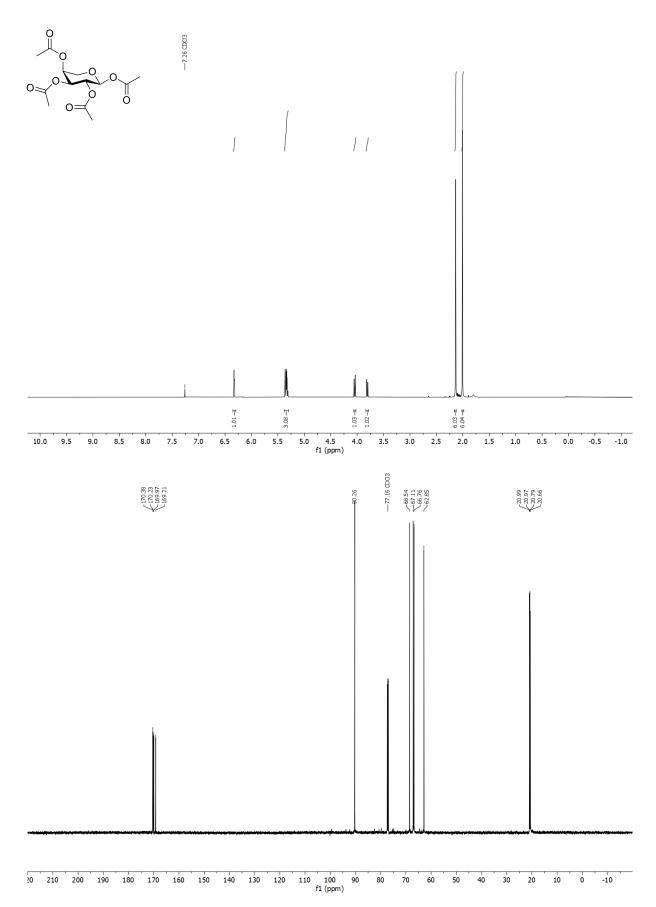


Figure S34: ¹H- and ¹³C-NMR spectra of S4 in CDCl₃ (600 MHz/151 MHz).

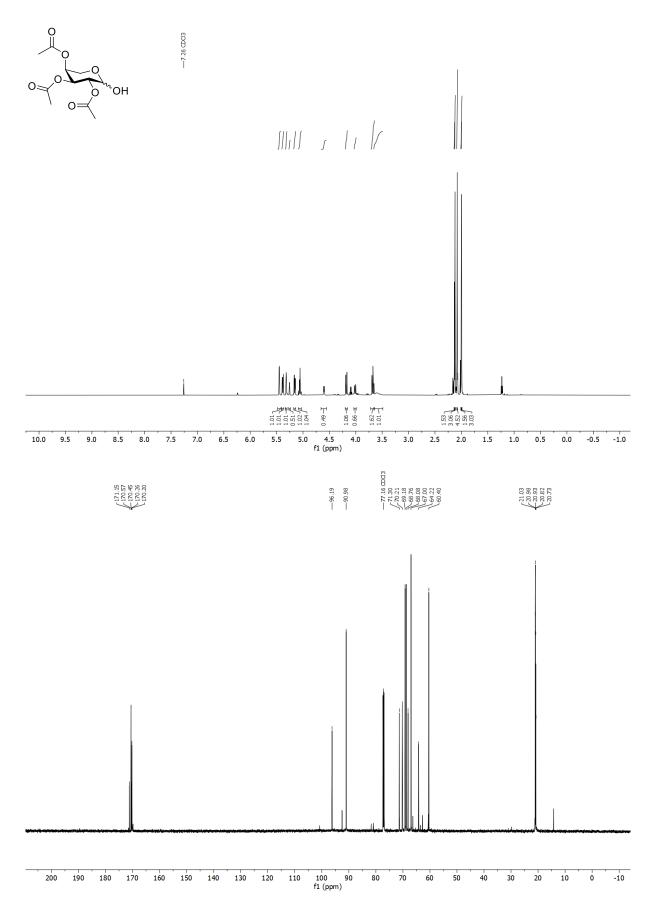


Figure S35: ¹H- and ¹³C-NMR spectra of S5 in CDCl₃ (600 MHz/151 MHz).

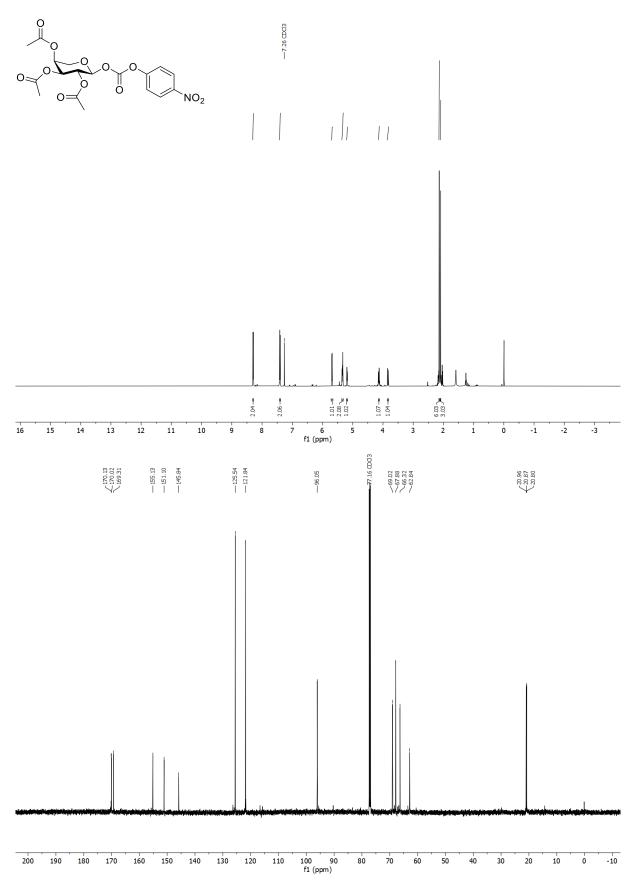


Figure S36: ¹H- and ¹³C-NMR spectra of 11 in CDCl₃ (600 MHz/151 MHz).

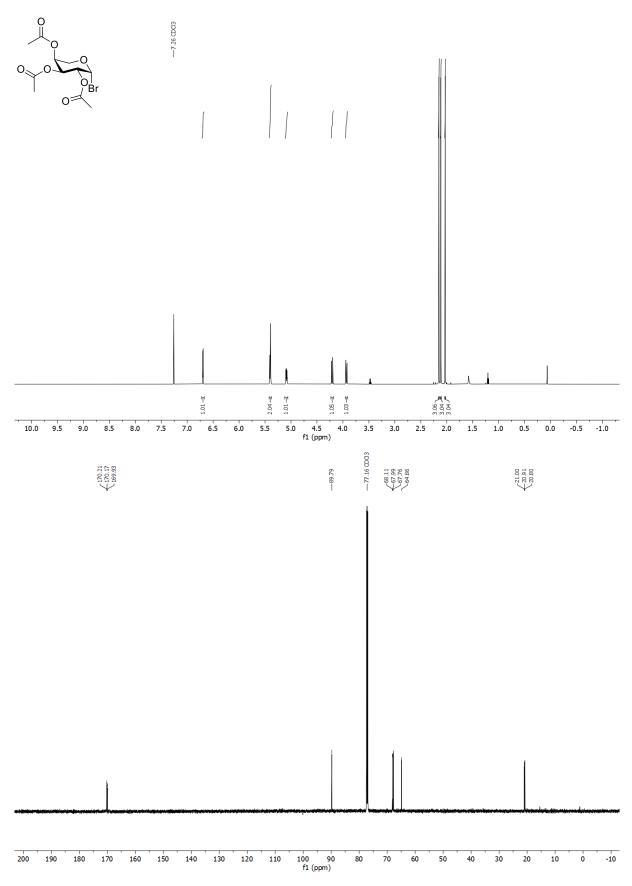


Figure S37: ¹H- and ¹³C-NMR spectra of 8 in CDCl₃ (600 MHz/151 MHz).

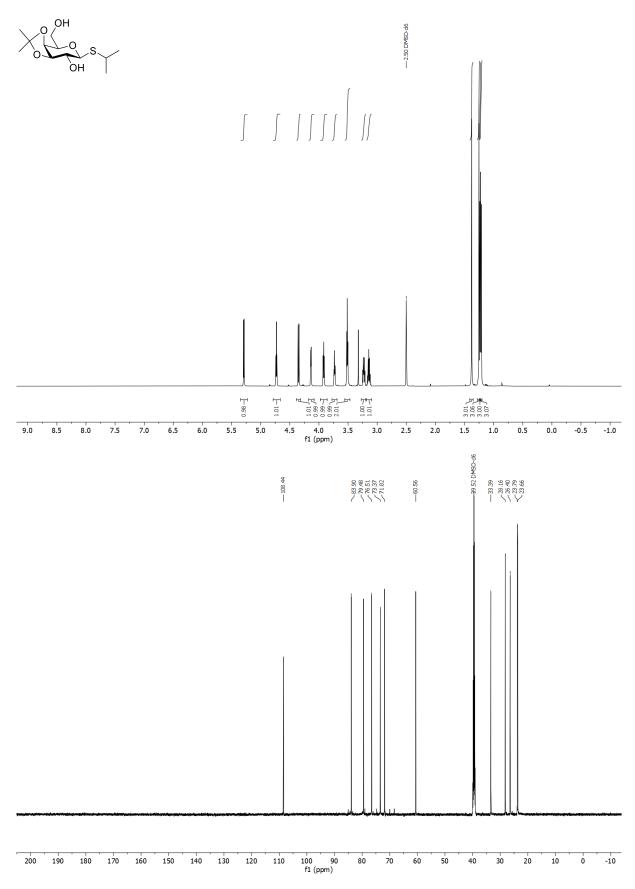


Figure S38: ¹H- and ¹³C-NMR spectra of S6 in DMSO- d_6 (600 MHz/151 MHz).

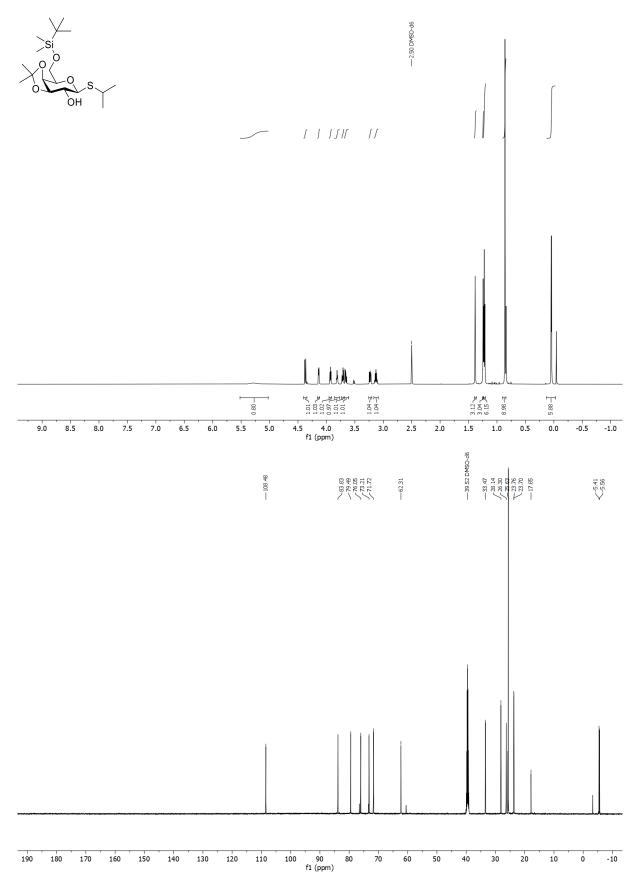


Figure S39: ¹H- and ¹³C-NMR spectra of 9 in DMSO- d_6 (600 MHz/151 MHz).

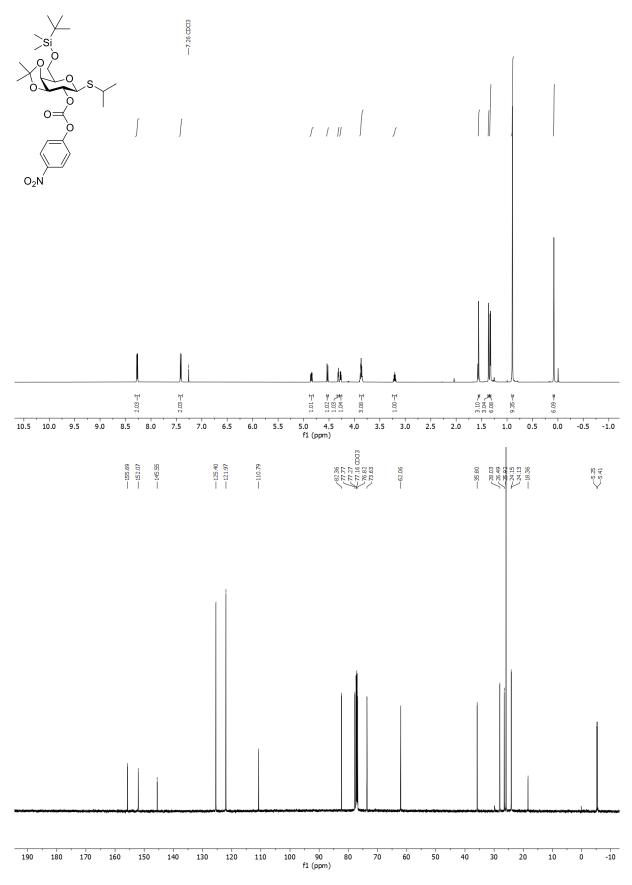


Figure S40: ¹H- and ¹³C-NMR spectra of 12 in CDCl₃ (600 MHz/151 MHz).

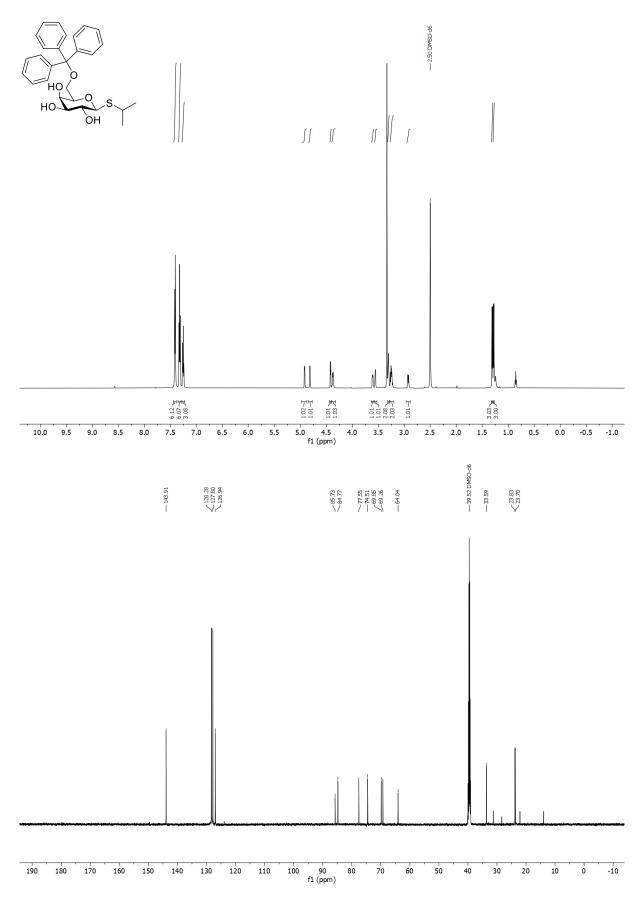


Figure S41: ¹H- and ¹³C-NMR spectra of S7 in DMSO- d_6 (600 MHz/151 MHz).

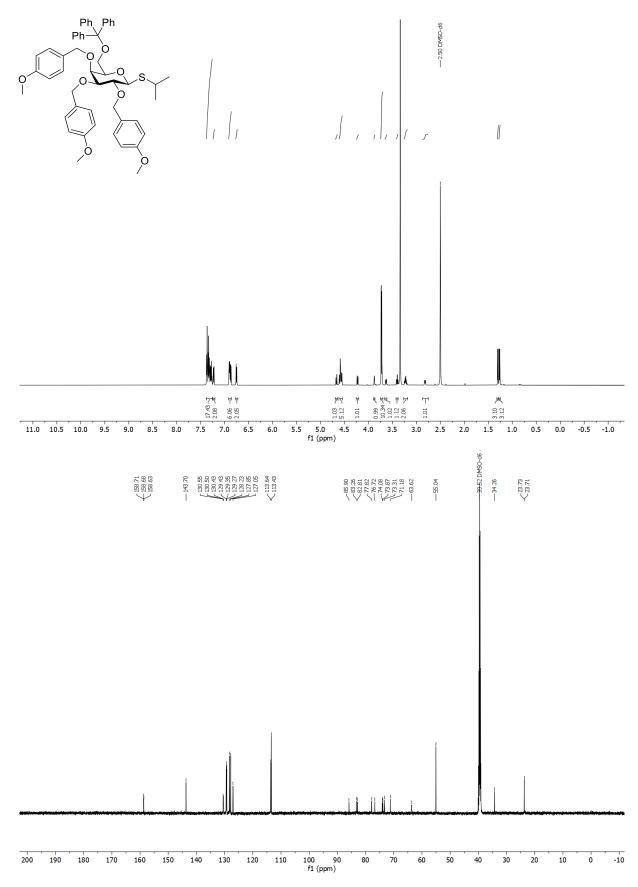


Figure S42: ¹H- and ¹³C-NMR spectra of S8 in DMSO- d_6 (600 MHz/151 MHz).

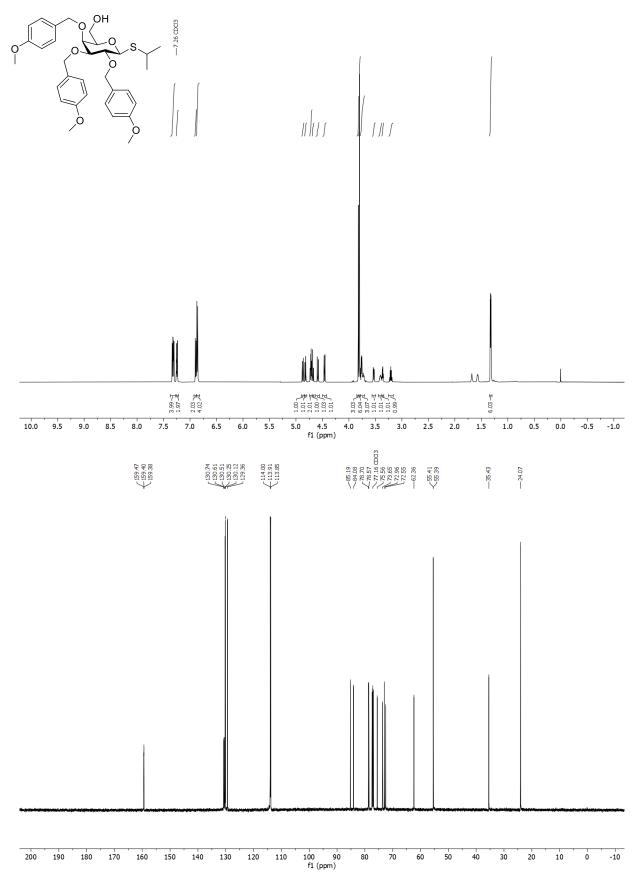


Figure S43: ¹H- and ¹³C-NMR spectra of 10 in CDCl₃ (600 MHz/151 MHz).

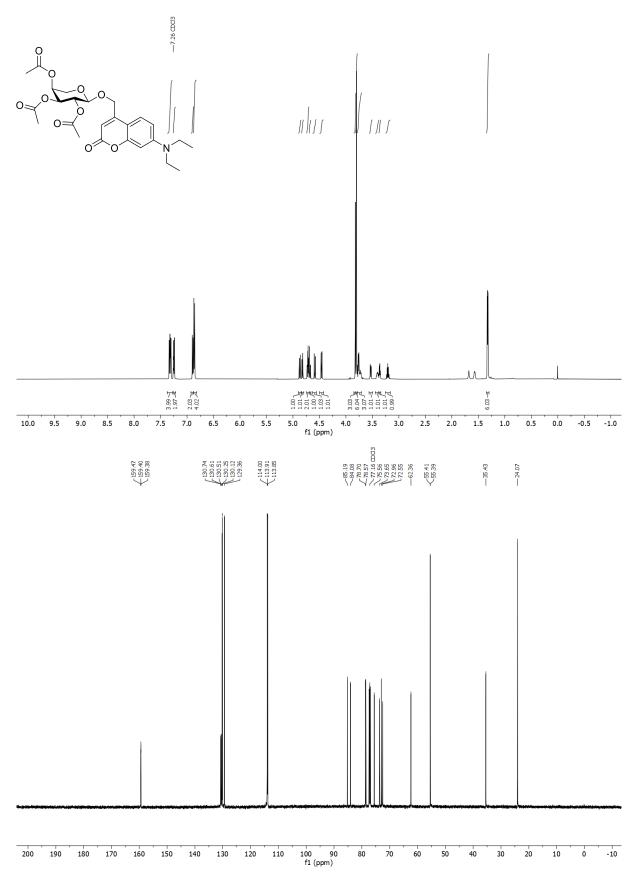


Figure S44: ¹H- and ¹³C-NMR spectra of 13 in CDCl₃ (600 MHz/151 MHz).

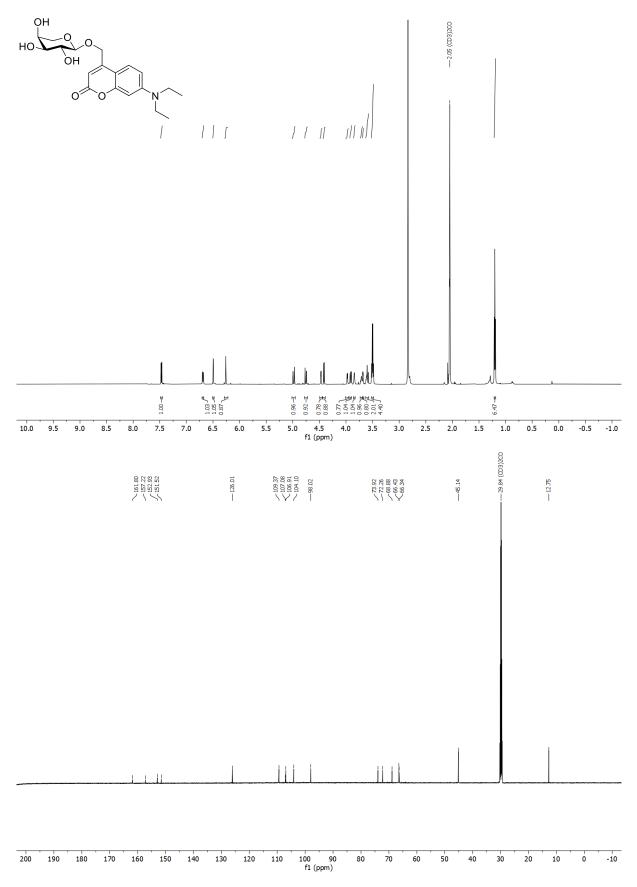


Figure S45: ¹H- and ¹³C-NMR spectra of 2b in acetone- d_6 (600 MHz/151 MHz).

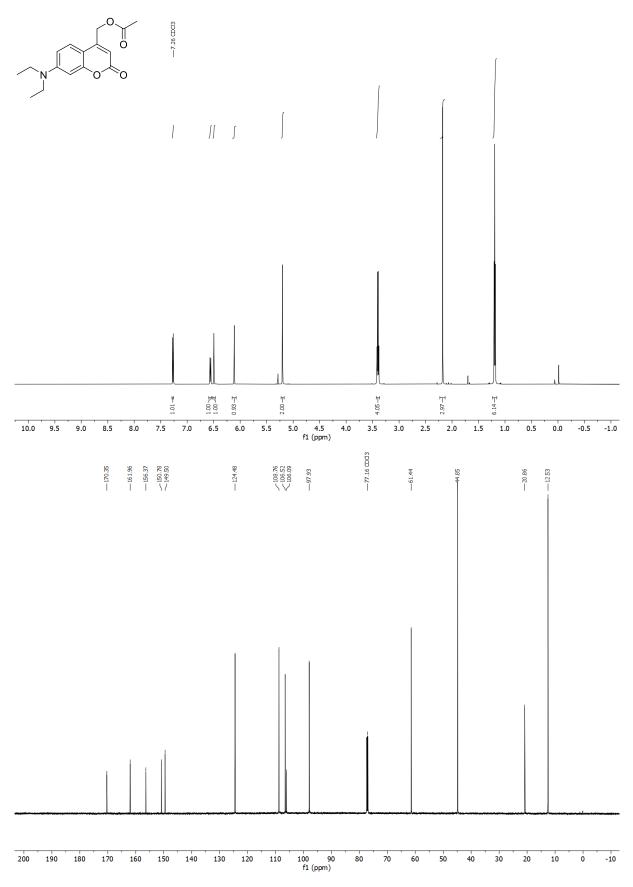


Figure S46: ¹H- and ¹³C-NMR spectra of S9 in CDCl₃ (600 MHz/151 MHz).

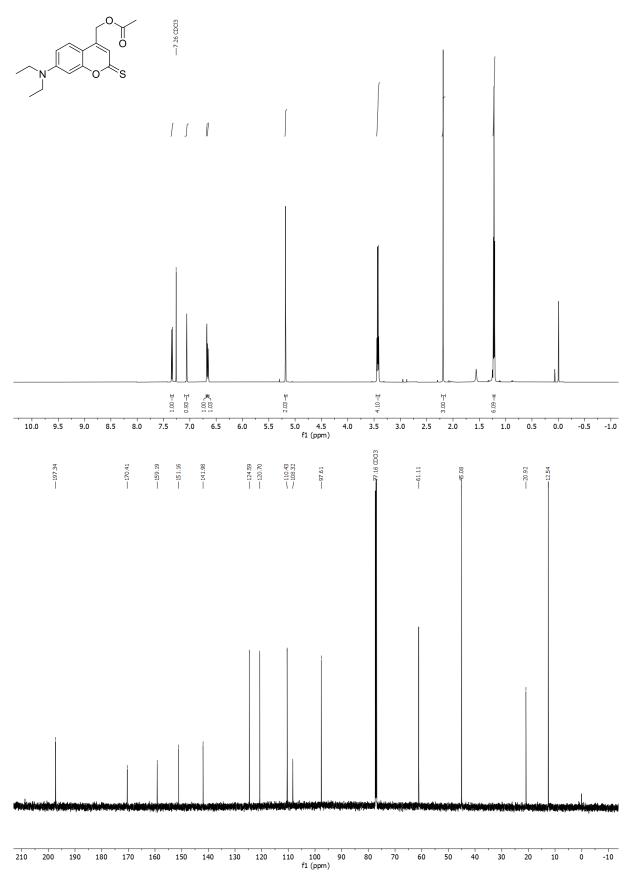


Figure S47: ¹H- and ¹³C-NMR spectra of S10 in CDCl₃ (600 MHz/151 MHz).

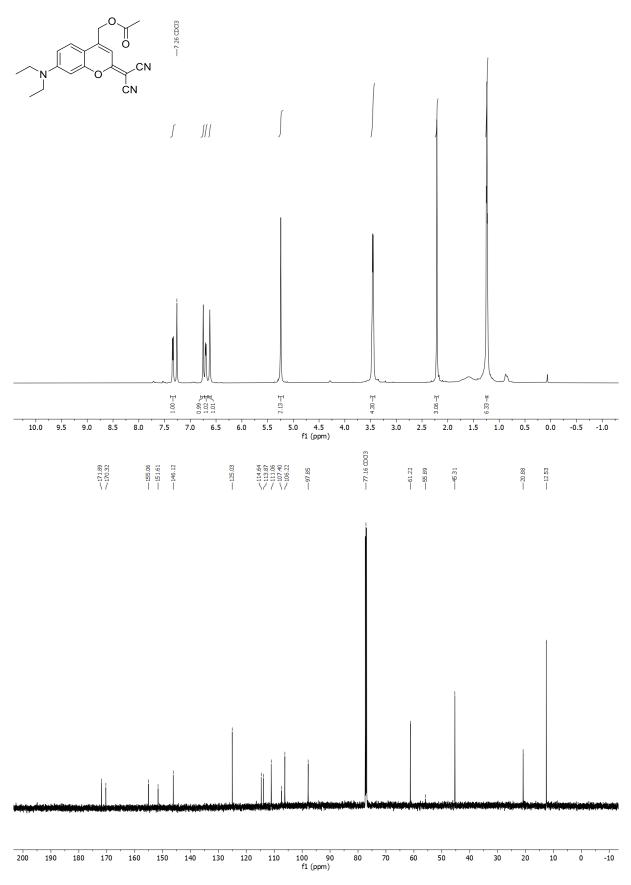


Figure S48: ¹H- and ¹³C-NMR spectra of S11 in CDCl₃ (600 MHz/151 MHz).

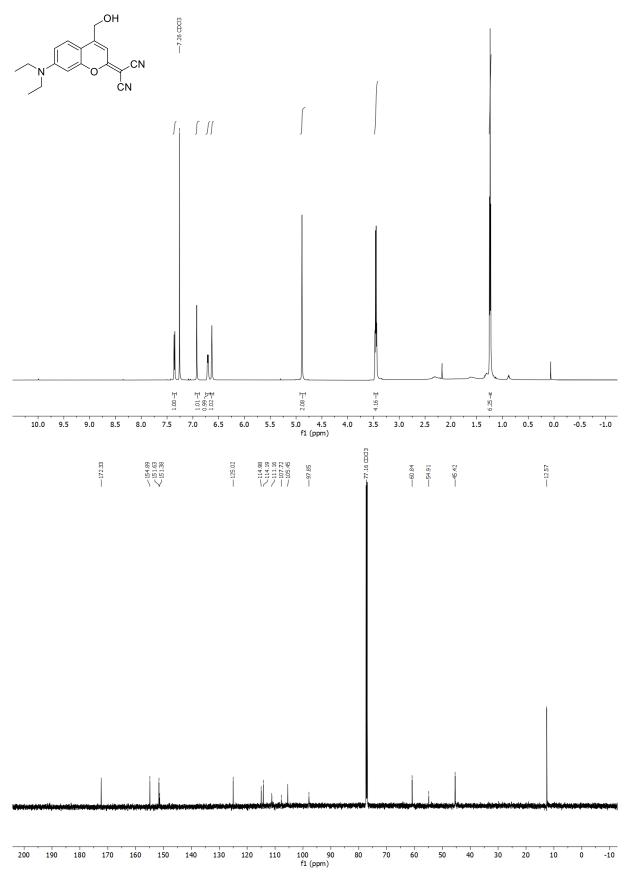


Figure S49: ¹H- and ¹³C-NMR spectra of 4 in CDCl₃ (600 MHz/151 MHz).

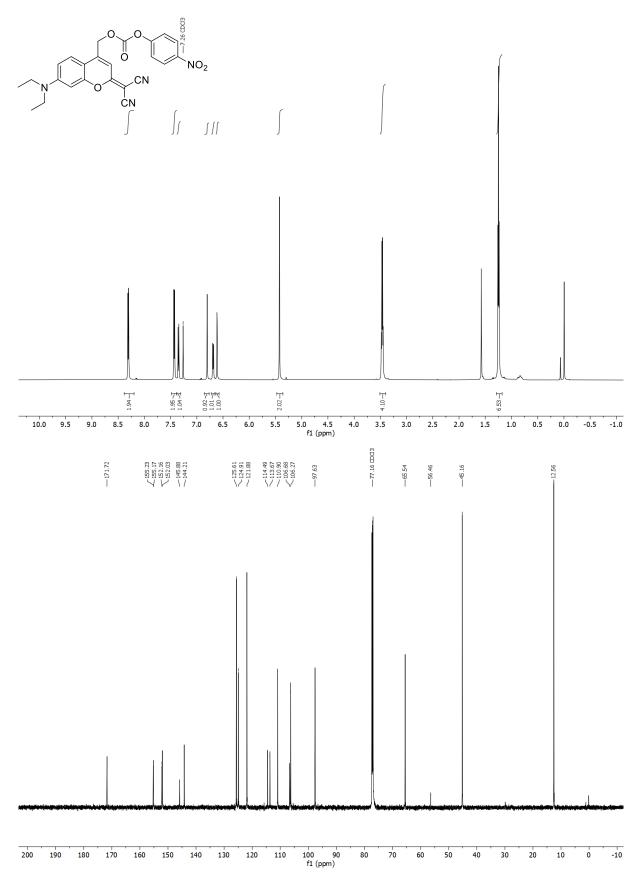


Figure S50: ¹H- and ¹³C-NMR spectra of 6 in CDCl₃ (600 MHz/151 MHz).

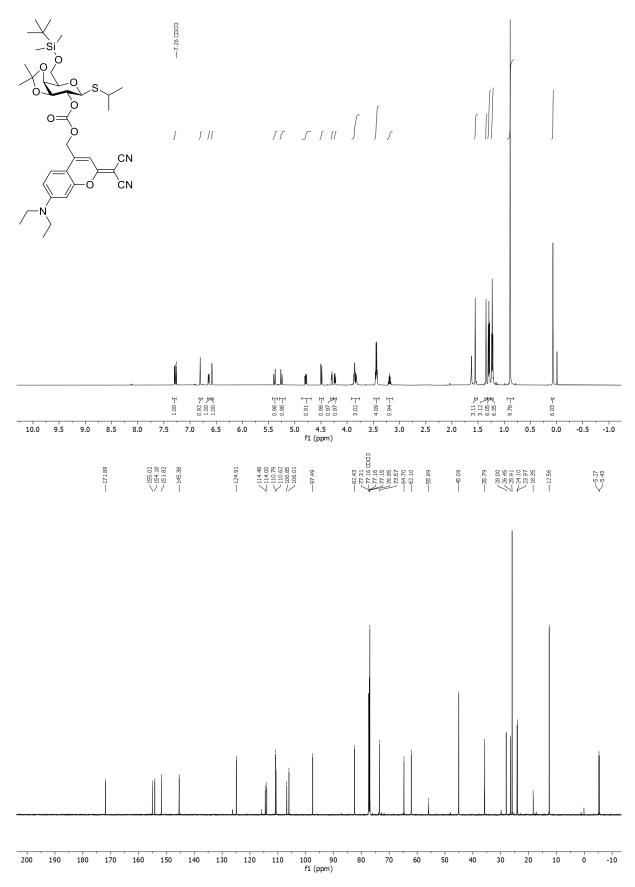


Figure S51: ¹H- and ¹³C-NMR spectra of 13 in CDCl₃ (600 MHz/151 MHz).

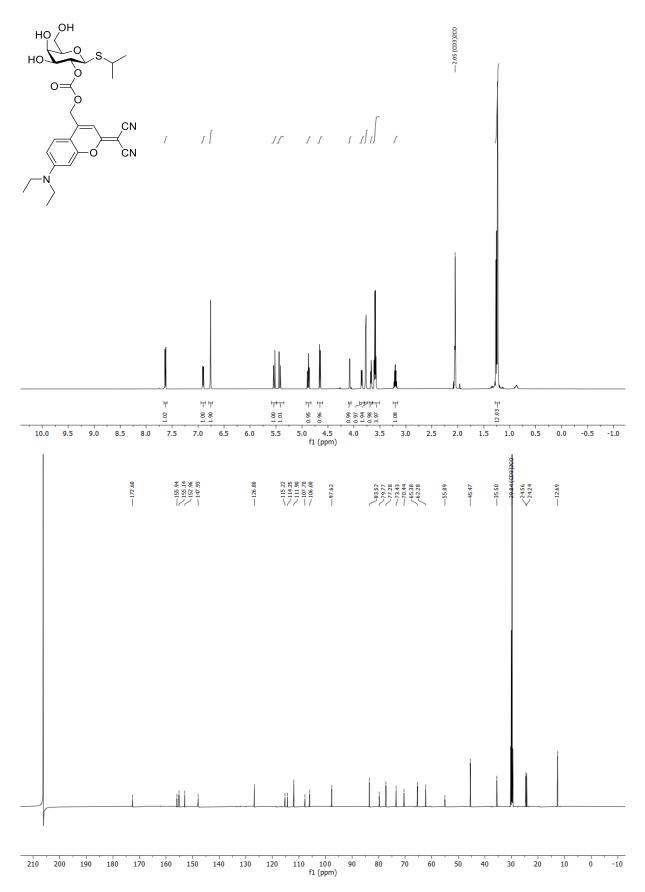


Figure S52: ¹H- and ¹³C-NMR spectra of 1c in acetone- d_6 (600 MHz/151 MHz).

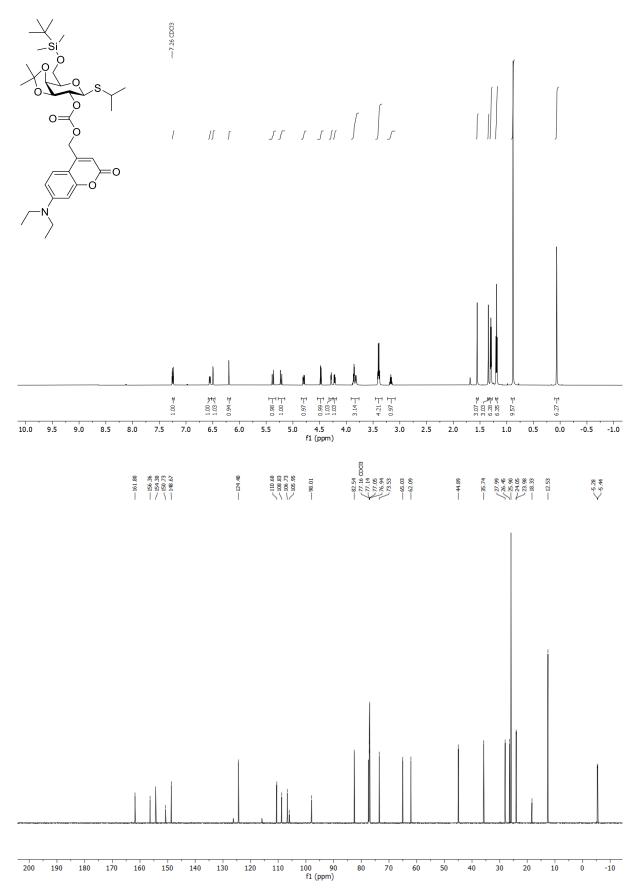


Figure S53: ¹H- and ¹³C-NMR spectra of 14 in CDCl₃ (600 MHz/151 MHz).

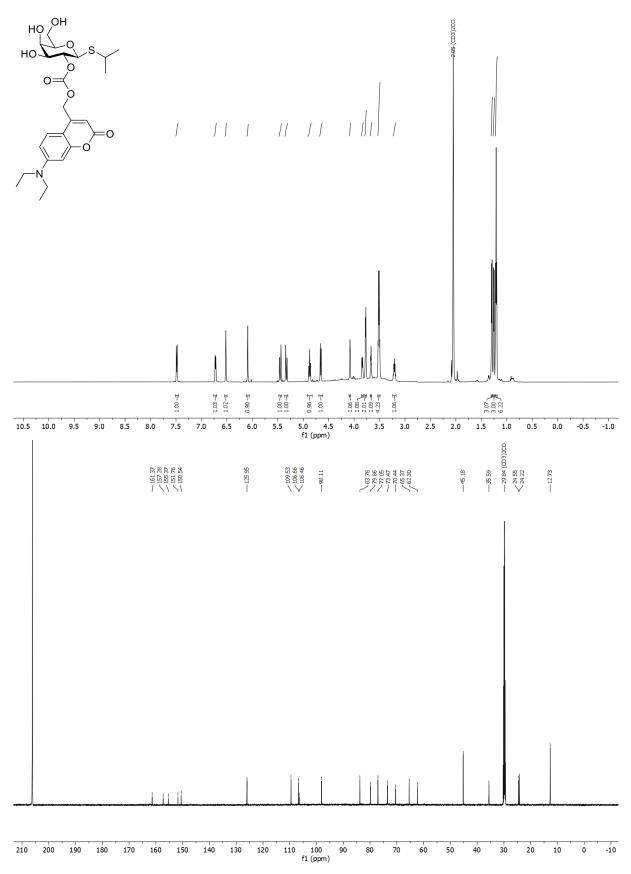


Figure S54: ¹H- and ¹³C-NMR spectra of 1b in acetone-*d*₆ (600 MHz/151 MHz).

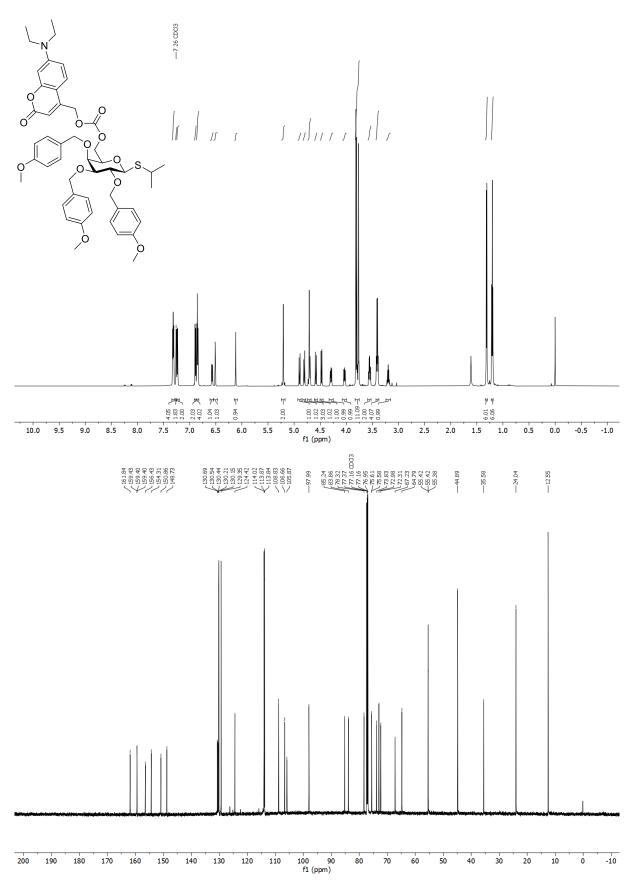


Figure S55: ¹H- and ¹³C-NMR spectra of 16 in CDCl₃ (600 MHz/151 MHz).

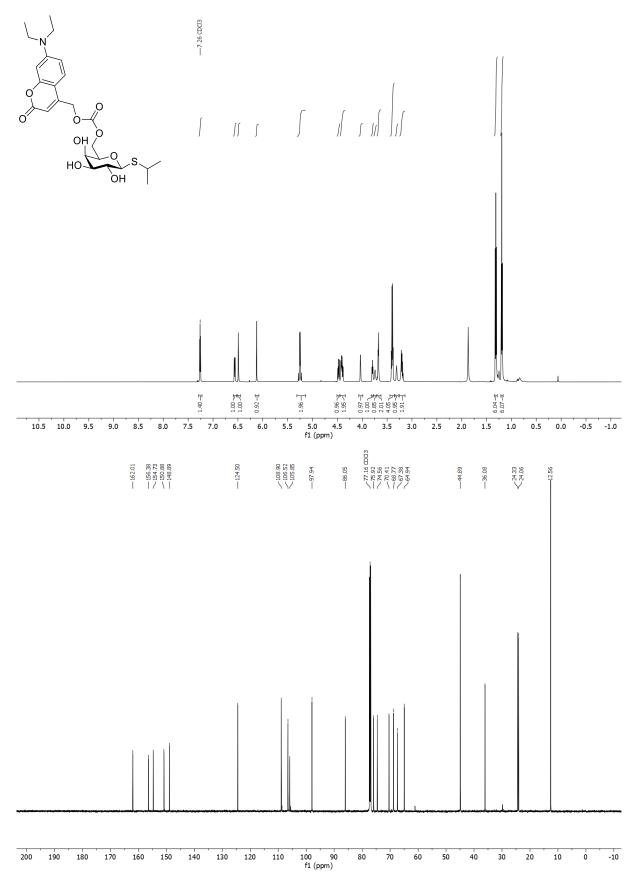


Figure S56: ¹H- and ¹³C-NMR spectra of 1d in CDCl₃ (600 MHz/151 MHz).

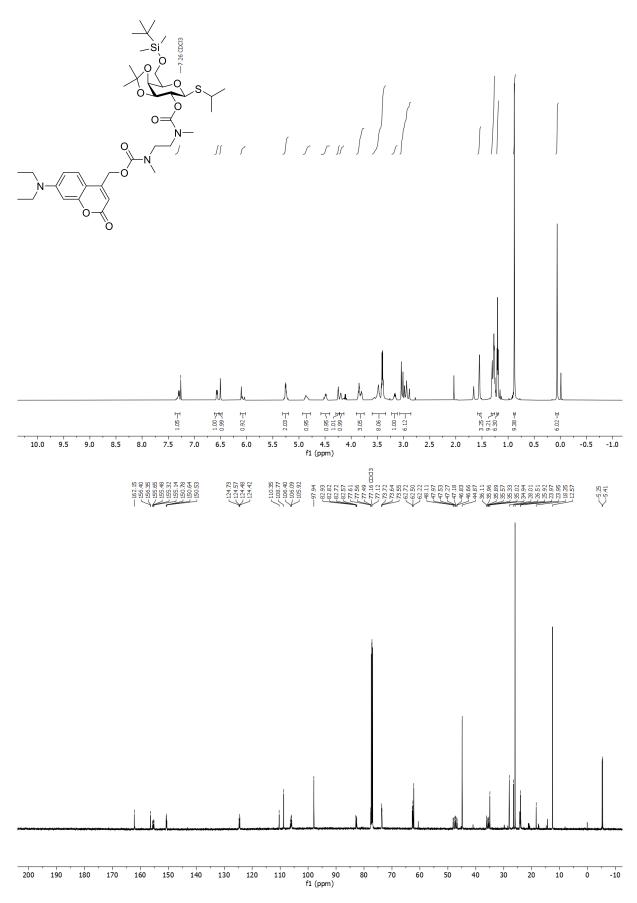


Figure S57: ¹H- and ¹³C-NMR spectra of 18 in CDCl₃ (600 MHz/151 MHz).

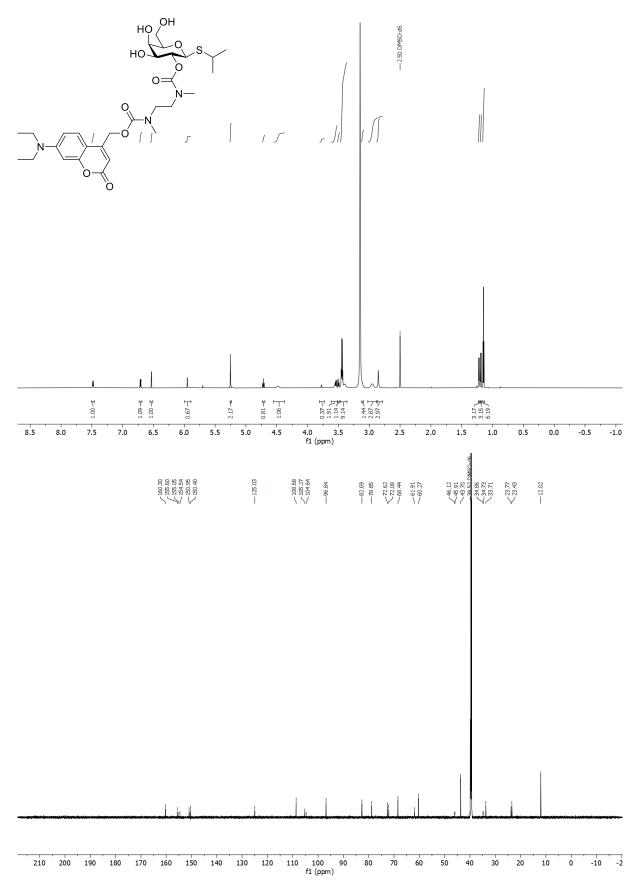


Figure S58: ¹H- and ¹³C-NMR spectra of 1e in DMSO- d_6 (600 MHz/151 MHz) at 60 °C.

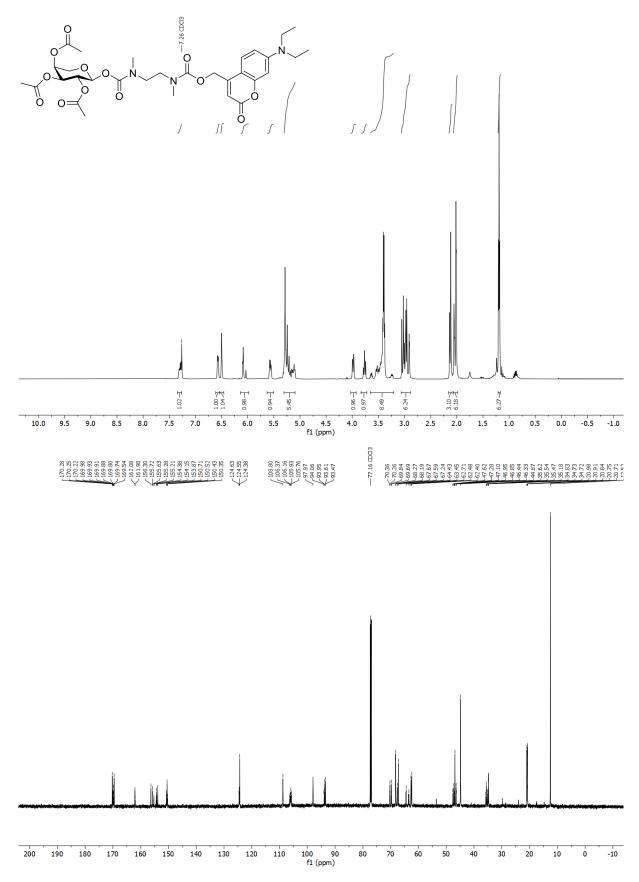


Figure S59: ¹H- and ¹³C-NMR spectra of 17 in CDCl₃ (600 MHz/151 MHz).

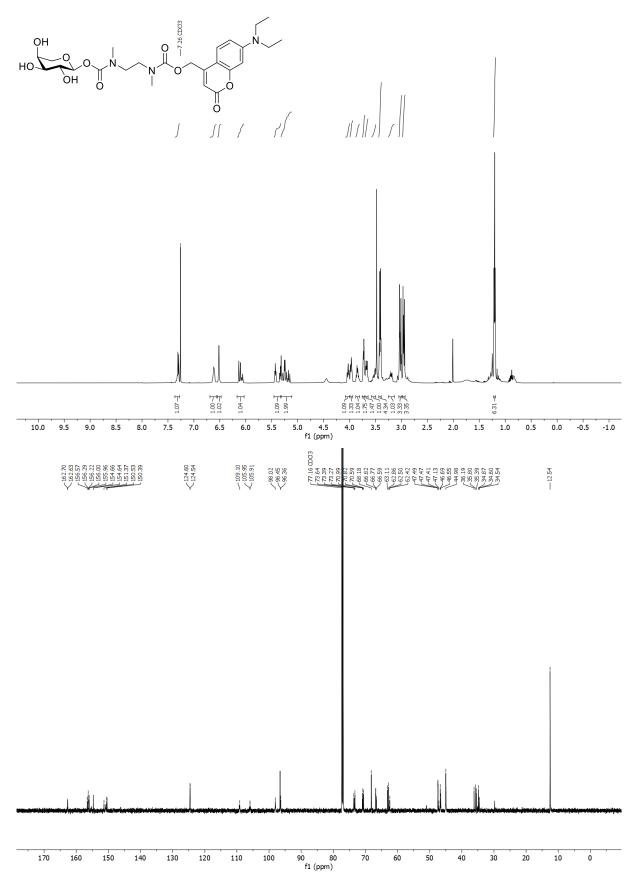


Figure S60: ¹H- and ¹³C-NMR spectra of 2c in CDCl₃ (600 MHz/151 MHz).

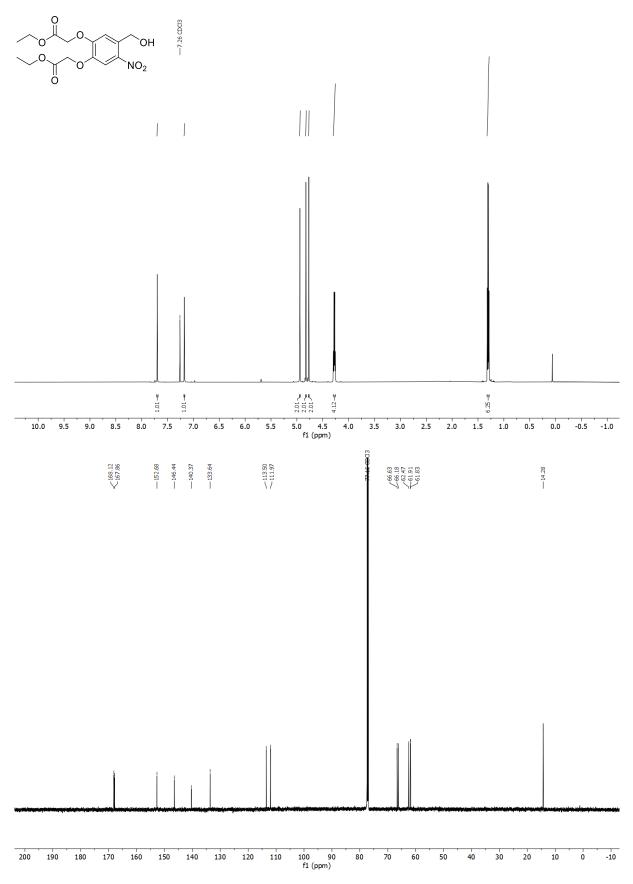


Figure S61: ¹H- and ¹³C-NMR spectra of 22 in CDCl₃ (600 MHz/151 MHz).

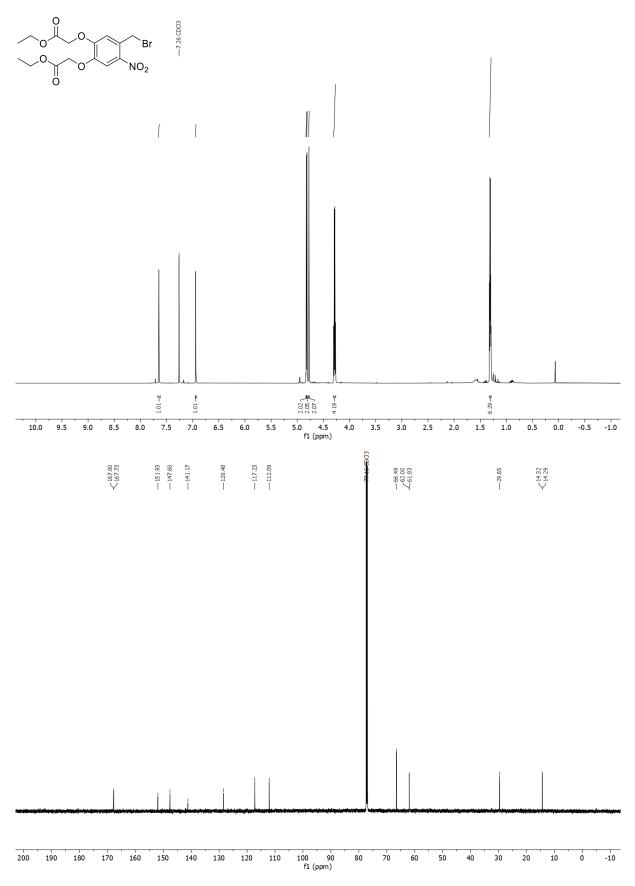


Figure 62: ¹H- and ¹³C-NMR spectra of 23 in CDCl₃ (600 MHz/151 MHz).

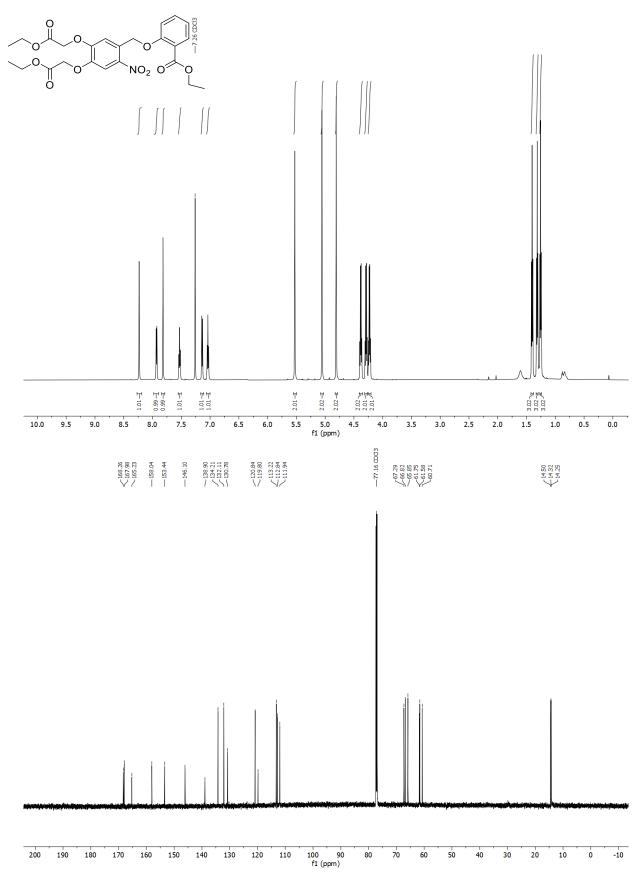


Figure S63: ¹H- and ¹³C-NMR spectra of 24 in CDCl₃ (600 MHz/151 MHz).

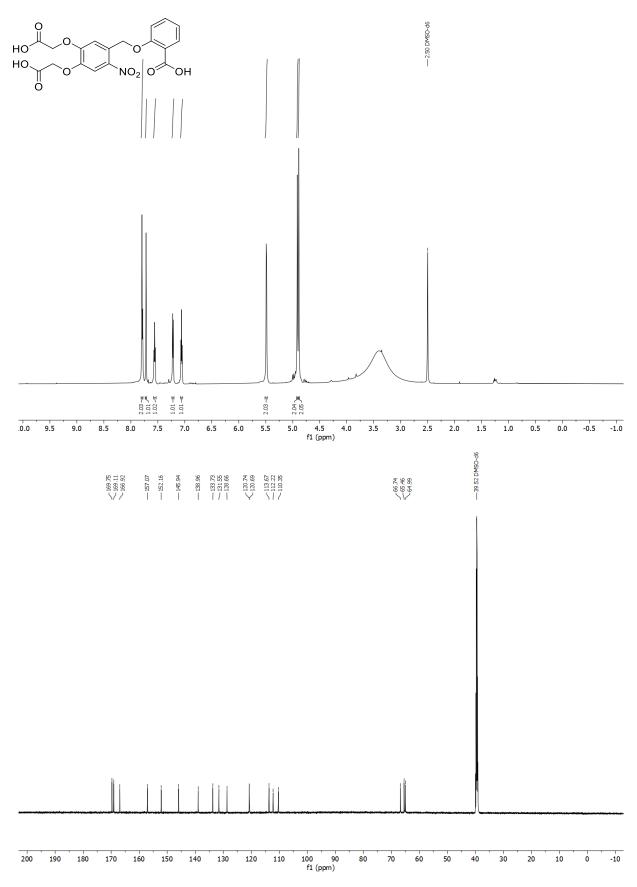


Figure S64: ¹H- and ¹³C-NMR spectra of 22a in DMSO-*d*₆ (600 MHz/151 MHz).

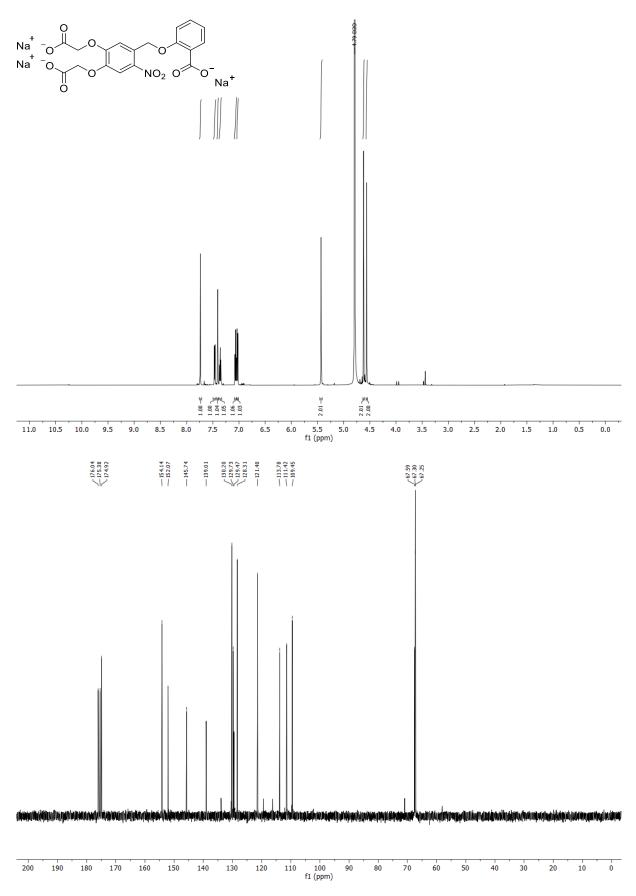


Figure S65: ¹H- and ¹³C-NMR spectra of 22b in D₂O (600 MHz/151 MHz).

- [1] D. Hanahan, J. Mol. Biol. 1983, 166, 557–580.
- [2] L. M. Guzman, D. Belin, M. J. Carson, J. Beckwith, J. Bacteriol. 1995, 177, 4121.
- [3] D. Binder, A. Grünberger, A. Loeschcke, C. Probst, C. Bier, J. Pietruszka, W. Wiechert, D. Kohlheyer, K.-E. Jaeger, T. Drepper, *Integr. Biol.* 2014, 6, 755–765.
- [4] D. Binder, C. Probst, A. Grünberger, F. Hilgers, A. Loeschcke, K.-E. Jaeger, D. Kohlheyer, T. Drepper, *PLoS One* **2016**, *11*, e0160711.
- [5] S. Balzer, V. Kucharova, J. Megerle, R. Lale, T. Brautaset, S. Valla, Microb. Cell Fact. 2013, 12, 26.
- [6] S. Verhoef, H. Ballerstedt, R. J. M. Volkers, J. H. de Winde, H. J. Ruijssenaars, Appl. Microbiol. Biotechnol. 2010, 87, 679–690.
- [7] a) J. E. Prior, M. D. Lynch, R. T. Gill, *Biotechnol. Bioeng.* 2010, 106, 326–332; b) N. Oberleitner, A. K. Ressmann, K. Bica, P. Gärtner, M. W. Fraaije, U. T. Bornscheuer, F. Rudroff, M. D. Mihovilovic, *Green Chem.* 2017, 19, 367–371.
- [8] T. Weinrich, M. Gränz, C. Grünewald, T. Prisner, M. Göbel, Eur. J. Org. Chem. 2017, 491–496.
- [9] Z. Gao, P. Yuan, D. Wang, Z. Xu, Z. Li, X. Shao, *Bioorg. Med. Chem. Lett.* 2017, 27, 2528–2535.
- [10] N. Fomina, C. L. McFearin, M. Sermsakdi, J. M. Morachis, A. Almutairi, *Macromolecules* 2011, 44, 8590– 8597.
- [11] D. Wahler, O. Boujard, F. Lefèvre, J.-L. Reymond, Tetrahedron 2004, 60, 703-710.
- [12] V. Duléry, O. Renaudet, C. Philouze, P. Dumy, Carbohydr. Res. 2007, 342, 894–900.
- [13] Annaleise R. Grummitt, Margaret M. Harding, Pia I. Anderberg, A. Rodger, Eur. J. Org. Chem. 2003, 63–71.
- [14] S. André, D. Specker, N. V. Bovin, M. Lensch, H. Kaltner, H.-J. Gabius, V. Wittmann, *Bioconjugate Chem.* 2009, 20, 1716–1728.
- [15] K. P. R. Kartha, H. J. Jennings, J. Carbohydr. Chem. 1990, 9, 777–781.
- [16] D. Binder, C. Bier, A. Grünberger, D. Drobietz, J. Hage-Hülsmann, G. Wandrey, J. Büchs, D. Kohlheyer, A. Loeschcke, W. Wiechert, K.-E. Jaeger, J. Pietruszka, T. Drepper, *ChemBioChem* **2016**, *17*, 296–299.
- [17] Y. Du, G. Gu, G. Wei, Y. Hua, R. J. Linhardt, Org. Lett. 2003, 5, 3627–3630.
- [18] Y. Du, M. Zhang, F. Yang, G. Gu, J. Chem. Soc., Perkin Trans. 1 2001, 3122–3127.
- [19] K. Ruda, J. Lindberg, P. J. Garegg, S. Oscarson, P. Konradsson, J. Am. Chem. Soc. 2000, 122, 11067–11072.
- [20] H. Wang, W.-G. Li, K. Zeng, Y.-J. Wu, Y. Zhang, T.-L. Xu, Y. Chen, Angew. Chem. Int. Ed. 2019, 58, 561– 565.
- [21] C. Bier, D. Binder, D. Drobietz, A. Loeschcke, T. Drepper, K.-E. Jaeger, J. Pietruszka, Synthesis 2017, 49, 42–52.
- [22] A. Gandioso, M. Palau, A. Nin-Hill, I. Melnyk, C. Rovira, S. Nonell, D. Velasco, J. García-Amorós, V. Marchán, ChemistryOpen 2017, 6, 375–384.
- [23] L. Fournier, I. Aujard, T. Le Saux, S. Maurin, S. Beaupierre, J. B. Baudin, L. Jullien, Chem. Eur. J. 2013, 19, 17494–17507.
- [24] A. Z. Suzuki, T. Watanabe, M. Kawamoto, K. Nishiyama, H. Yamashita, M. Ishii, M. Iwamura, T. Furuta, Org. Lett. 2003, 5, 4867–4870.
- [25] R. Wang, K. Cai, H. Wang, C. Yin, J. Cheng, Chem. Commun. 2018, 54, 4878–4881.
- [26] J. Ni, D. A. Auston, D. A. Freilich, S. Muralidharan, E. A. Sobie, J. P. Y. Kao, J. Am. Chem. Soc. 2007, 129, 5316–5317.
- [27] L. Tietze, M. Müller, S.-C. Duefert, K. Schmuck, I. Schuberth, Chem. Eur. J. 2013, 19, 1726–1731.