

**Supplementary Information for**

Methanol biotransformation toward high-level production of fatty acid derivatives by engineering the industrial yeast *Pichia pastoris*.

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**This PDF file includes:**

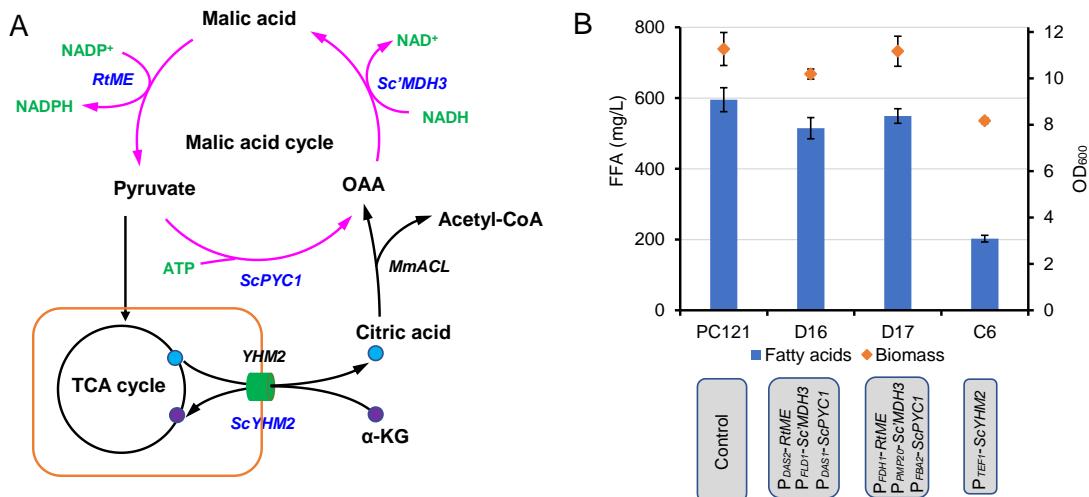
Supplementary text

Figures S1 to S6

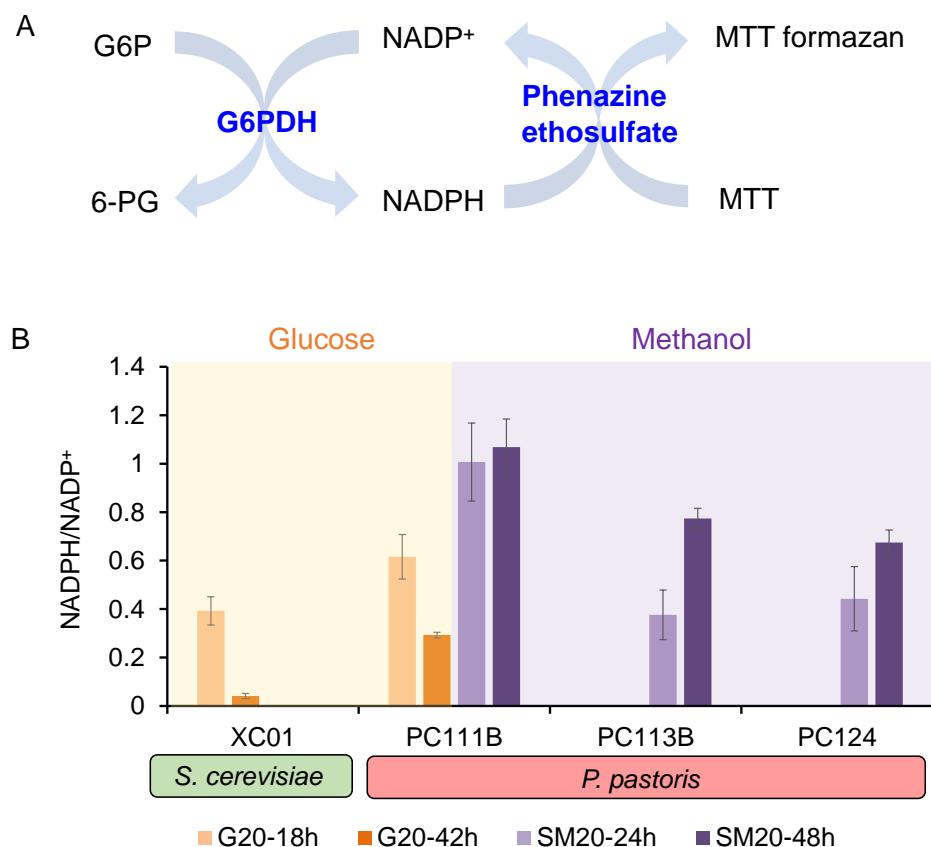
Tables S1 to S3

SI References

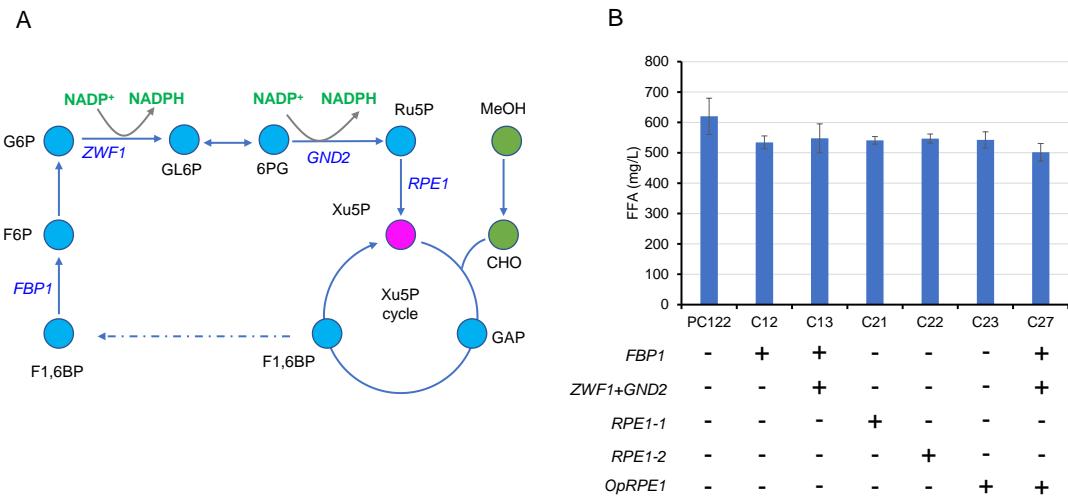
## Supplementary Figures



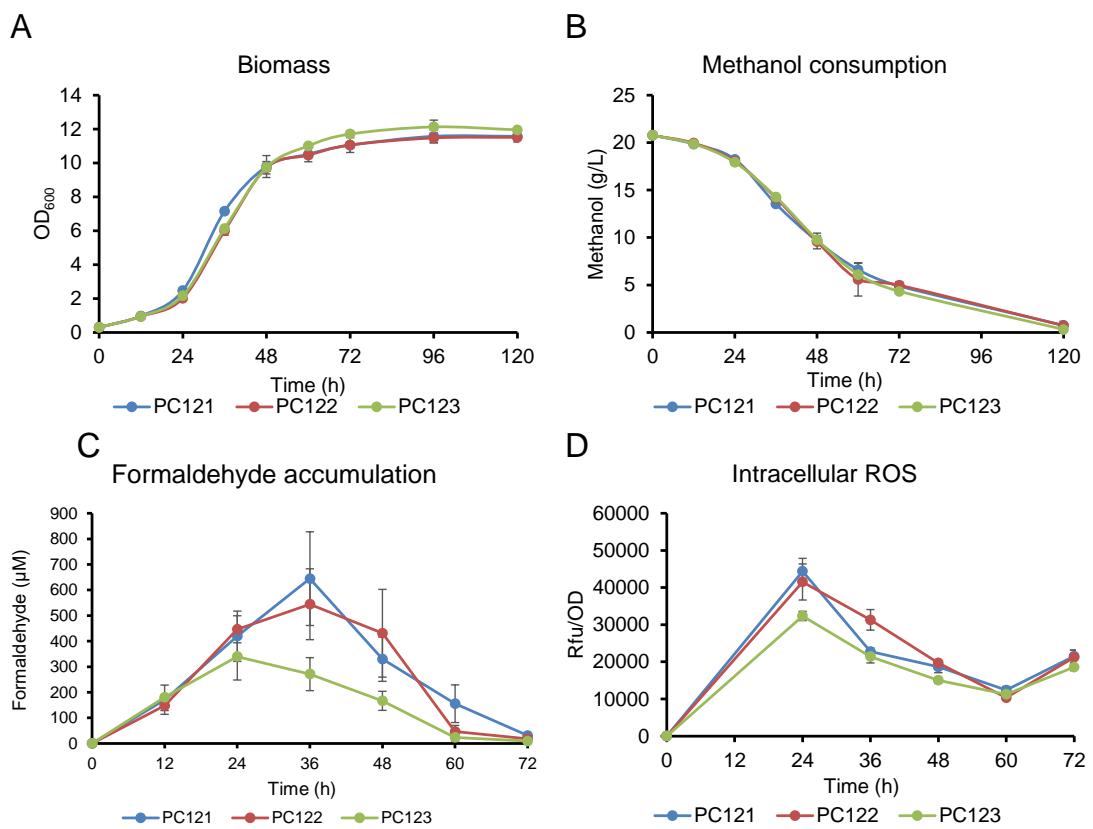
**Figure S1. Construction of malate cycle (transhydrogenase cycle) in cytoplasm. (A)**  
 Schematic diagram of malate cycle. Overexpressed genes are marked in light blue. **(B)**  
 FFA production in engineered strains. Error bars correspond to the SD of the mean ( $n = 3$ , corresponding to three biological replicates)



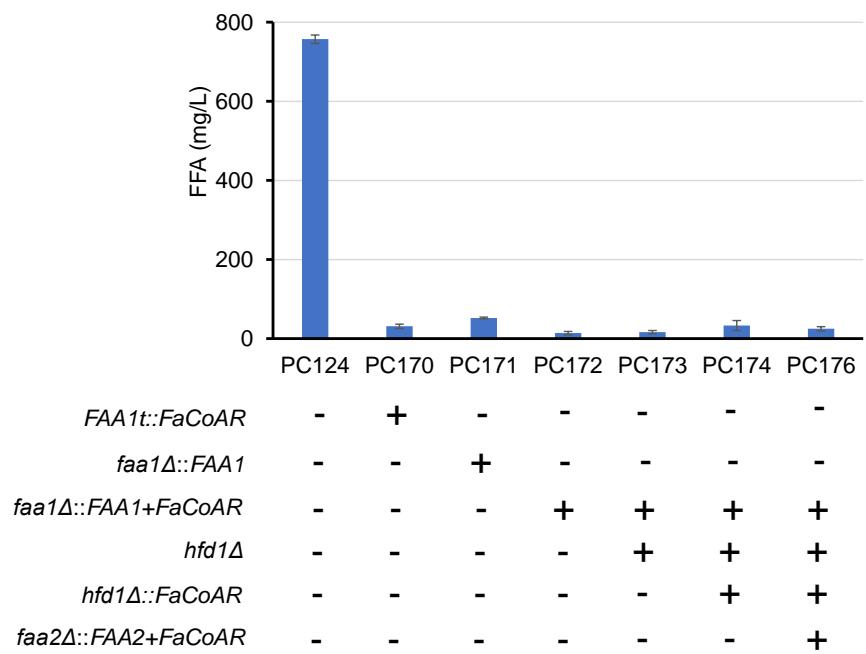
**Figure S2. Quantification of NADPH/NADP<sup>+</sup> in *S. cerevisiae* and *P. pastoris*.** (A) Schematic diagram of cofactor recycling assay. (B) NADPH/NADP<sup>+</sup> of *S. cerevisiae* and *P. pastoris* (PC111B) in glucose medium and methanol medium. PC111B, wild type; PC113B, FFA-producing strain with blocked FFA activation; PC124, FFA over-producing strain with engineered central metabolism. \*Due to the slow growth of PC113B, the sampling time was delayed by 24 h. Error bars correspond to the SD of the mean ( $n = 3$ , corresponding to three biological replicates).



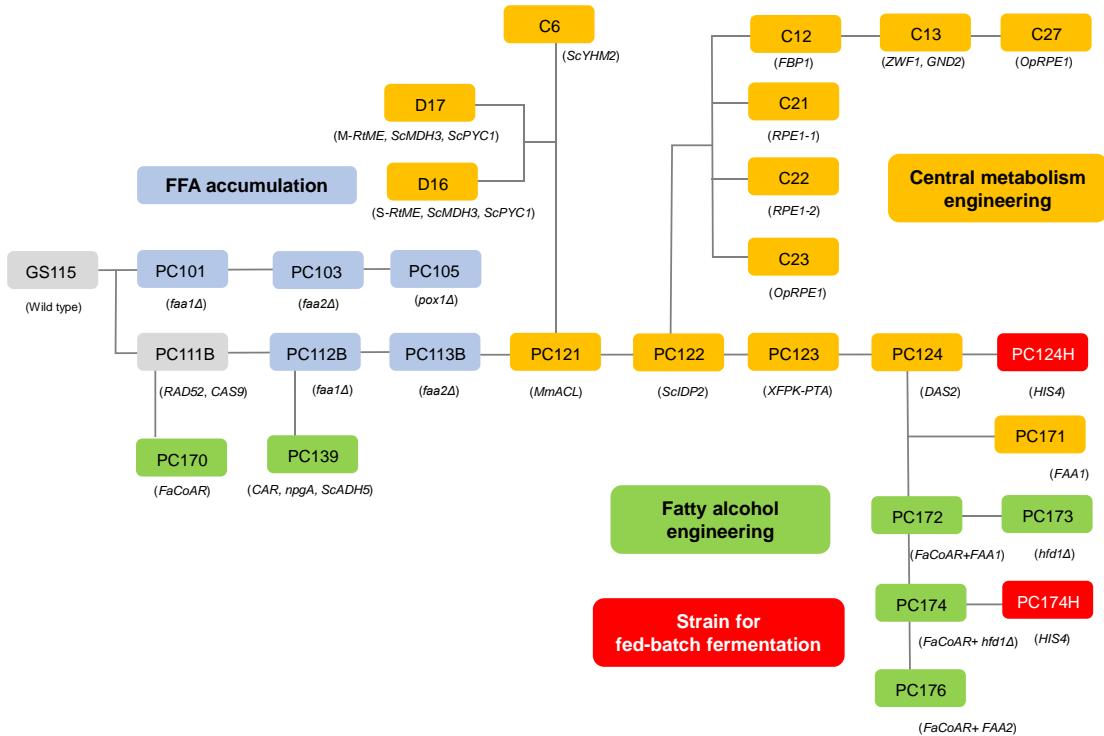
**Figure S3. Enhancing gluconeogenesis pathway to increase Xu5p supply. (A)** Schematic diagram of gluconeogenesis pathway engineering. Overexpressed genes are marked in light blue. **(B)** FFA production in engineered strains with rewired gluconeogenesis pathway. Error bars correspond to the SD of the mean ( $n = 3$ , corresponding to three biological replicates).



**Figure S4. Rewiring central metabolism for improving FFA production from methanol.** (A-D) The growth curves, methanol consumption, formaldehyde accumulation and intracellular ROS level. Error bars represent SD of triplicate samples.



**Figure S5. FFA accumulation of the fatty alcohols producing strains.** Error bars correspond to the SD of the mean ( $n = 3$ , corresponding to three biological replicates)



**Figure S6. Flowchart of yeast strain construction in this study.**

## Supplementary Tables

**Table S1.** Plasmids used in this study.

Plasmid name	Genotype or characteristic	Resource or Reference
pPICZ-Cas9-gFAA1	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gFAA1-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gFAA2	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gFAA2-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gPOX1	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPOX1-T <sub>AOX1</sub>	This study
pPICZ-Cas9-gHFD1	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gHFD1-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gHIS4	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gHIS4-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gPNSI2	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPNSI2-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gPNSI3	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPNSI3-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gPNSI4	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPNSI4-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gPNSI5	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPNSI5-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gPNSI6	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPNSI6-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gPNSI8	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPNSI8-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-gPNSI10	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPNSI10-T <sub>AOX1</sub>	(1)
pPICZ-Cas9-	panARS, <i>Ble</i> <sup>R</sup> , P <sub>HTX1</sub> -Cas9-T <sub>DAS1</sub> , P <sub>HTX1</sub> -gPNSI12-	(1)

gPNSI12	$T_{AOX1}$	
pPICZ-Cas9-	panARS, <i>Ble</i> <sup>R</sup> , $P_{HTX1}$ -Cas9- $T_{DAS1}$ , $P_{HTX1}$ -gPNSI13-	(1)
gPNSI13	$T_{AOX1}$	
pCAI-gPNSII-4	panARS, <i>KanMX</i> , $P_{HTX1}$ -gPNSII-4- $T_{AOX1}$	(1)
pCAI-gPNSIII-5	panARS, <i>KanMX</i> , $P_{HTX1}$ -gPNSIII-5- $T_{AOX1}$	(1)
pCAI-gFAA1t	panARS, <i>KanMX</i> , $P_{HTX1}$ -gFAA1t- $T_{AOX1}$	This study
pCAI-gFAA2t	panARS, <i>KanMX</i> , $P_{HTX1}$ -gFAA2t- $T_{AOX1}$	This study

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**Table S2. Strains used in this study**

Strain name	Genotype	Resource or Reference
GS115	Mut+, <i>his4</i> , AOX1, AOX2	From Pro Cai
XC01	<i>MATa; MAL2-8c; SUC2; his3Δ1; ura3-52; XI-5::P<sub>TEF1</sub>-Cas9-T<sub>CYC1</sub></i>	Lab reserved
PC110	GS115, <i>HIS4::P<sub>GAP</sub>-PpRAD52-T<sub>AOX1</sub></i>	This study
PC111B	PC110, <i>PNSI-2::P<sub>GAP</sub>-hCas9-T<sub>DAS1</sub></i>	This study
<b>Engineered strains for FFA production</b>		
PC113B	Mut+, <i>his</i> <sup>-</sup> , AOX1, AOX2, <i>his4Δ::P<sub>GAP</sub>-PpRAD52-T<sub>AOX1</sub>, PNSI-3::P<sub>GAP</sub>-hCas9-T<sub>DAS1</sub>, faa1Δ,faa2Δ</i> =[PC111B]+(faa1Δ,faa2Δ)	This study
PC121	Mut+, <i>his</i> <sup>-</sup> , AOX1, AOX2, <i>his4Δ::P<sub>GAP</sub>-PpRAD52-T<sub>AOX1</sub>, PNSI-3::P<sub>GAP</sub>-hCas9-T<sub>DAS1</sub>, faa1Δ,faa2Δ,</i> <i>PNSI2::P<sub>AOX1</sub>-MmACL-KpOpt1-T<sub>FAA1</sub></i> =[PC131B]+( <i>PNSI2::P<sub>AOX1</sub>-MmACL-KpOpt1-T<sub>FAA1</sub></i> )	This study
PC122	Mut+, <i>his</i> <sup>-</sup> , AOX1, AOX2, <i>his4Δ::P<sub>GAP</sub>-PpRAD52-T<sub>AOX1</sub>, PNSI-3::P<sub>GAP</sub>-hCas9-T<sub>DAS1</sub>, faa1Δ,faa2Δ, PNSI-</i> <i>2::P<sub>AOX1</sub>-MmACL-KpOpt1-T<sub>FAA1</sub>, PNSI-4::P<sub>FLD1</sub>-ScIDP2o-T<sub>DAS2</sub></i> =[PC121]+( <i>PNSI-4::P<sub>FLD1</sub>-ScIDP2o-T<sub>DAS2</sub></i> )	This study
PC123	Mut+, <i>his</i> <sup>-</sup> , AOX1, AOX2, <i>his4Δ::P<sub>GAP</sub>-PpRAD52-T<sub>AOX1</sub>, PNSI-3::P<sub>GAP</sub>-hCas9-T<sub>DAS1</sub>, faa1Δ,faa2Δ, PNSI-</i> <i>2::P<sub>AOX1</sub>-MmACL-KpOpt1-T<sub>FAA1</sub>, PNSI-4::P<sub>FLD1</sub>-ScIDP2o-T<sub>DAS2</sub>,PNSIII-5::BbXFPK-P<sub>HTX1</sub>-CkPTA</i> =[PC122]+( <i>PNSIII-5::BbXFPK-P<sub>HTX1</sub>-CkPTA</i> )	This study

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	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::P <sub>GAP</sub> -PpRAD52-T <sub>AOX1</sub> , PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , faa1Δ,faa2Δ, PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> =[PC123]+(PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> )	This study
PC124	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , faa1Δ,faa2Δ, PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> , his4Δ::HIS4 =[PC124]+(his4Δ::HIS4)	This study
PC124H	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , faa1Δ,faa2Δ, PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> , his4Δ::HIS4 =[PC124]+(his4Δ::HIS4)	This study
<b>Engineered strains for fatty alcohol production</b>		
PC170	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::P <sub>GAP</sub> -PpRAD52-T <sub>AOX1</sub> , PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , FAA1t::P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1t</sub> =[PC111B]+(FAA1t::P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> )	This study
PC171	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::P <sub>GAP</sub> -PpRAD52-T <sub>AOX1</sub> , PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , faa2Δ, PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> , faa1Δ::FAA1 =[PC124]+(faa1Δ::FAA1)	This study
PC172	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::P <sub>GAP</sub> -PpRAD52-T <sub>AOX1</sub> , PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , faa2Δ, PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> , faa1Δ::FAA1+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> =[PC124]+(faa1Δ::FAA1+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> )	This study
PC173	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::P <sub>GAP</sub> -PpRAD52-T <sub>AOX1</sub> , PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , faa2Δ, PNSI-2::P <sub>AOX1</sub> -	This study

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	<i>MmACL</i> -KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2-</sub> DAS2-T <sub>GAP</sub> , faa1Δ::FAA1+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> , hfd1Δ =[PC172]+(hfd1Δ)	
PC174	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::P <sub>GAP</sub> -PpRAD52-T <sub>AOX1</sub> , PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , faa2Δ, PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2-</sub> DAS2-T <sub>GAP</sub> , faa1Δ::FAA1+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> , hfd1Δ::P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> =[PC173]+(hfd1Δ::P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> )	This study
PC175	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::P <sub>GAP</sub> -PpRAD52-T <sub>AOX1</sub> , PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> , faa1Δ::FAA1+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> , hfd1Δ::P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> , faa2Δ::FAA2=[PC174]+(faa2Δ::FAA2)	This study
PC176	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::P <sub>GAP</sub> -PpRAD52-T <sub>AOX1</sub> , PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> , faa1Δ::FAA1+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> , hfd1Δ::P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> , faa2Δ::FAA2+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> =[PC175]+(faa2Δ::FAA2+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> )	This study
PC174H	Mut <sup>+</sup> , his <sup>-</sup> , AOX1, AOX2, his4Δ::HIS4, PNSI-3::P <sub>GAP</sub> -hCas9-T <sub>DAS1</sub> , PNSI-2::P <sub>AOX1</sub> -MmACL-KpOpt1-T <sub>FAA1</sub> , PNSI-4::P <sub>FLD1</sub> -ScIDP2o-T <sub>DAS2</sub> , PNSIII-5::BbXFPK-P <sub>HTX1</sub> -CkPTA, PNSII-4::P <sub>DAS2</sub> -DAS2-T <sub>GAP</sub> , faa1Δ::FAA1+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> , hfd1Δ::P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> , faa2Δ::FAA2+P <sub>TEF1</sub> -FaCoAR-KpOpt1-T <sub>FBP1</sub> =[PC174]+(his4Δ::HIS4)	This study

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**Table S3. Primers used in this study.**

Primer No.	Name	Sequence (5' - 3')
<b>Primers for gRNA plasmid construction</b>		
P1	AOX1t-Kpn-ARS-R	AAACGTCAAATCATAATCAGCACTAGGTACCGCACAAACGAACGTCTCACTTAATCTTC
P2	HTX1-Cas9-F	GGAGTACTTCTTGTCCATCGTTGACTAGTTGTTAGTTAATATAGTTGAGTATGAGATGGAA CTC
P3	gPOX1-F	GATTTCTGATGAGTCCGTGAGGACGAAACGAGTAAGCTCGTAAAATCTGGGACTTCCGGGTT TTAGAGCTAGAAATAGCAAGTTAAAATAAGGCT
P4	gPOX1-R	GTTCGTCCTCACGGACTCATCAGAAAATCTTGATTGTTAGGTAACTGAACTGGATGTATTAGT TTGG
P5	gFAA1t-F	GAAGCTATGAATGAAAAGCAGTTTAGAGCTAGAAATAGCAAGTTAAAATAAGGCTAG
P6	gFAA1t-R	TGCTTTCATTAGCTCGACGAGCTTACTCGTTCGTCC
P7	gFAA2t-F	AAAAATGAAATAAAAACAGGTTTAGAGCTAGAAATAGCAAGTTAAAATAAGGCTAG
P8	gFAA2t-R	CTGTTTTATTCATTGACGAGCTTACTCGTTCGTCC
<b>Primers for seamless gene deletion of POX1</b>		

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P9	POX1-HRUP-F	GTCAAGTTGTATGCATAGAACAGAACAGAATGGAAG
P10	POX1-HRUP-R1	CGTAATCAATAACACCGTTGTATTTCAGTTAGACTGGTTAAAAAAG
P11	POX1-HRDN-F1	ACTGAAAATCACAAACGGTGTATTGATTACGTAGTAATGCTATCACAAAGTA
P12	POX1-HRDN-R	GAAGCTTCTCGTAGCTCTCATCTTC

**Primers for engineering FFA production**

P13	MmACL-KpOpt1-AOX1p-F	AGATCAAAAAACAACTAATTATTGAAACGATGTCTGCCAAGGCCATCAG
P14	MmACL-KpOpt1-FAA1t-R	GTCCGTAGAAAACCTCAATCGGCTGCTCGCTCACATACTCATATGTTCTGGAAGGACAT
P15	ScIDP2o-FLD1p-F	TGCTTGTTCATACAATTCTTGATATTACAATGACAAAGATAAAGGTTGCAAACCCTA
P16	ScIDP2o-DAS2t-R	AACTACTAACCGTTAGTGGCCAAATCTACTCAAAGTGCTGCTGCTCGAAC
P17	ScYHM2o-TEF1p-F	TCACTACATACATTAGTTATTGCCAACATGCCATCTACTACTAACCGCCG
P18	ScYHM2o-AOX1t-R	CAGGCAAATGGCATTCTGACATCCTCTTGATCAATGTTGGCGACTGGAGTCTC
P19	RtME-DAS2p-F	TCACTCTTATCAAACATCAAACATCAAAATGCCTGCTCATTTGCCCG
P20	RtME-AOX1t-R	CAGGCAAATGGCATTCTGACATCCTCTTGATCATTGTGCTGTTCTGCTCTAATA

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P21	RtME-FDH1p-F	CAAATACCTCCAACATCACCCACTAAACAATGCCTGCTCATTTGCC
P22	ScMDH3-FLD1p-F	TGCTTGTTCATACAATTCTGATATTACAATGGTCAAAGTCGCAATTCTGGC
P23	ScMDH3-DAS1t-R	CTCCTAACTAAAAGTAAAGACTTCCC GTTCAAGAGTCTAGGATGAAACTCTGCCT
P24	ScMDH3-PMP20p-F	AATCCCACCAAGCAAAAAAAATCTAAGATGGTCAAAGTCGCAATTCTGGC
P25	ScPYC1-DAS1p-F	TTGATTTATCTGGAGAATAATCGAACAAATGTCGCAAAGAAAATTGCCG
P26	ScPYC1-AOX1t-R	CAGGCAAATGGCATTCTGACATCCTCTTGATCATGCCTAGTTAACAGGAAC TTGG
P27	ScPYC1-FBA2p-F	TTGATAAGGTAATTGATTAATTCTAAATATGTCGCAAAGAAAATTGCCG
P28	KpZWF1-DAK1p-F	ACAGGAAACAAAGGAATTTATACACTTAAATGACCGATACGAAAGCCGTAGAA
P29	KpZWF1t-PI10DN-R	AAGCAAAAGTCCGAAGAAATCTGAAAACACAGAAAATCAGGAGTAGAGTTGTGAAAG
P30	KpGND2-PEX5p-F	ATTCTCAACCCAACCCTTAACTAATCGTAATGGTTGAAGCAACAGGAGATATTGG
P31	KpRPE1-2-R	TTTTCGAATAGCTAGGTGATATGAAGGAAAGGTA
P32	KpGND2t-RPE2t-R	CTTCCCTTCATATCACCTAGCTATTGAAAAACACAGCAAATATGGCTGTGC
P33	KpRPE1-2-F	ATGGTTAAAACAATTATTGCTCCTCAATCCTG
P34	KpRPE1-1-R	CAAGGCAACGGCCCTAGTGA
P35	KpRPE1-1-FLD1p-F	CTTGCTTGTTCATACAATTCTGATATTACAATGGTCAAACCTGTTATCGCTCC

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P36	OpRPE-ADH2t-R	CATTACATAAGACGTATACAAACTATTCGGCTCACTCAAGAAGTCCGCGGG
P37	OpRPE-FLD1p-F	TGCTTGTTCATACAATTCTGATATTACAATGGTAAACCAATTATTGCTCCCTC
P38	BbXFPKo-FDH1t-R	TTACTTAATATCAAATTAAATACATTCAATTATTCAATTACACCAGCAGTAGCAGCAG
P39	BbXFPKo-HTX1p-F	TCATACTCAAACATATTAAAACACTACAACAATGACTTCTCCTGTTATTGGTACTCCATG
P40	CkPTAo-HTX1p-F	ATCCAGTTCAAGTTACCTAAACAAATCAAATGAAATTGATGGAAAATATTTGGTTGGC
P41	CkPTAo-FBP1t-R	AAATCTCGAACAGTGCCAATCGAACGCATTAACCTTGAGCTTGAGCTTGAACAGC
P42	DAS2-GAPt-R	CGAATTTCAGCTATTCACATACAAATCGATTACAACCTGTCATGCTTGGTTCCC
P43	DAS2p-F	ATTACTGTTTGGCAATCCTGTTGATAAG

**Primers for constructing pathways for fatty alcohol production**

P44	FAA1-CAT1t-R	AATCTTAATTAATAATAATAGTTAGCTCACTGTTGCCTATATACTTCATCGACAC
P45	CAT1t-F	GCTAACTATTTATTAAATTAAAGATTCTTAACCTCGG
P46	CAT1t-FBP1t-R	ATAGGGATAATAGAAAAGTAAGGTTCCCGCGTTAGTTCTGATGATGTTGATCATCG
P47	FaCoAR-KpO-FBP1t-R	CAAATCTCGAACAGTGCCAATCGAACGCATCACCAAGTAAATTCTCATGATGGC
P48	FaCoAR-R1-TEF1p-F	CTCACTACATACATTAGTTATTGCCAACATGAACACTTCCTACCGGAGGTA
P49	FAA1DN-TEF1p-F	AGTGCAGATAAAAGAGGCGACAGTTACACGTACTACGTAGTTCAATTGTACTTTTC

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P50	HFD1UP-TEF1p-R	AGTGCAGATAAAAGAGGCGACAGTTATCAATGAAAAAGATGAATTGCCAAATAGAAGAC
P51	HFD1DN-FBP1t-F	ATAGGGATAATAGAAAAGTAAGGTTCCGCGACTTTATTGTTCGTCTGTATCATCTGTTAAGAG
P52	FAA2-GAPt-R	GAATTCAGCTATTCACATACAAATCGATCTACATCTTAGTCTCCCTCAGAAGACTC
P53	FBP1t-GAPt-F	CTTCCGATCAAATTGGAATGGAAAATTGCCGGAACCTTACTTTCTATTATCCCTA
P54	FAA2DN-TEF1p-F	AGTGCAGATAAAAGAGGCGACAGTTATGGTATGCTCAATGTGTCAATTACAGG

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#### SI References

1. P. Cai et al., Recombination machinery engineering facilitates metabolic engineering of the industrial yeast *Pichia pastoris*. *Nucleic Acids Res* **49**, 7791-7805 (2021).