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Supporting Information

Stereoselective Synthesis, Configurational Assignment and Biological Evaluations of the Lipid Mediator RvD2_{n-3 DPA}

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General Information

Unless otherwise stated, all commercially available reagents and solvents were used in the form they were supplied without any further purification. The stated yields are based on the isolated material. All sensitive reactions were performed under an argon atmosphere using Schlenk techniques. Reaction flasks were covered with aluminum foil during sensitive reactions and storage to minimize exposure to light. Thin layer chromatography was performed on silica gel 60 F₂₅₄ aluminum-backed plates fabricated by Merck. Flash column chromatography was performed on silica gel 60 (40 - 63 μ m) fabricated by Merck. NMR spectra were recorded on a Bruker AVII 400 spectrometer at 400 MHz for ¹H NMR and at 101 MHz for ¹³C NMR. Coupling constants (*J*) are reported in hertz and chemical shifts are reported in parts per million (δ) relative to the central residual protium solvent resonance in ¹H NMR (CDCl₃ = δ 7.27, CD₃OD = δ 3.31, and C₆D₆ = δ 7.16) and the central carbon solvent resonance in ¹³C NMR (CDCl₃ = δ 77.0 ppm, CD₃OD = δ 49.0, and C₆D₆ = δ 7.28). Highresolution mass spectra were recorded at 70 eV on a Micromass Prospec Q or Micromass QTOF 2W spectrometer using ESI as the method of ionization. Optical rotations were measured using a 0.7 mL cell with a 1.0 dm path length on a PerkinElmer 341 polarimeter. HPLC-analyses were performed using an AD-H stationary phase (CHIRALPAK, 4.6 x 250 mm, particle size 5 μ m, from Daicel Corporation) or a C₁₈ stationary phase (Eclipse XDBC₁₈, 4.6 x 250 mm, particle size 5 μ m, from Agilent Technologies), applying the conditions stated. The UV-Vis spectrum was recorded using an Agilent Technologies Cary 8485 UV-Vis spectrophotometer using quartz cuvettes.

Experimental Details

(2R,3S)-5,5-Bis(ethylthio)pentane-1,2,3-triol (18)



Compound **8** (5.01 g, 37.4 mmol, 1.00 equiv.) was dissolved in concentrated hydrochloric acid (6.50 mL), followed by dropwise addition of ethanethiol (6.50 mL, 89.5 mmol, 2.40 equiv.). As this was added, the reaction mixture changed to a dark brown solution. The reaction mixture was stirred for 3 h at room temperature before it was neutralized by the addition of sat. aq. K₂CO₃ (30 mL). Then, the phases were separated, and the aq. phase was extracted with CH₂Cl₂ (3 × 50 mL). The combined organic phase was dried (MgSO₄), filtrated and the solvent removed in vacuo. The crude product was purified by flash chromatography (SiO₂, gradient elution, 2.5 – 5% MeOH in CH₂Cl₂) to afford the desired thioacetal **18** (6.29 g, 26.2 mmol, 70%) as a clear yellow oil. All spectroscopic and physical data were in agreement with those reported in the literature.¹ *R*_f (10% MeOH in CH₂Cl₂, visualized by KMnO₄ stain) = 0.57; $[\alpha]_D^{20}$ = -19.0 (*c* 1.0, MeOH) [Lit.¹ $[\alpha]_D^{20}$ = -19.2 (*c* 1.0, MeOH)]; ¹H NMR (400 MHz, CDCl₃): δ 4.09 – 4.04 (m, 2H), 3.83 – 3.70 (m, 2H), 3.61 (td, *J* = 5.2, 3.8 Hz, 1H), 2.78 – 2.56 (m, 4H), 2.29 (s, 2H), 2.22 (s, 1H), 2.03 – 1.99 (m, 2H), 1.30 – 1.24 (m, 6H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 73.8, 72.2, 63.6, 48.6, 38.9, 24.5, 24.2, 14.6 (2C).

(5*S*,6*R*)-5-(2,2-*Bis*(ethylthio)ethyl)-6-((*tert*-butyldimethylsilyl)oxy)-2,2,3,3,9,9,10,10-octamethyl-4,8-dioxa-3,9-disilaundecane (19)



Triol **18** (5.09 g, 21.2 mmol, 1.00 equiv.) was dissolved in CH₂Cl₂ (96 mL) and cooled to -78 °C. The solution was stirred for 10 min before 2,6-lutidine (19.6 mL, 169 mmol, 8.00 equiv.) was added, followed by TBSOTf (19.4 mL, 84.5 mmol, 4.00 equiv.). The reaction mixture was allowed to slowly warm up to room temperature and stirred for 18 h. Sat. aq. NH₄Cl (65 mL) was added, the phases were separated, and the aqueous phase was extracted with CH₂Cl₂ (3 × 40 mL). The combined organic layer was washed with brine (30 mL), dried (MgSO₄), filtrated, and the solvent removed in vacuo. TBS-triol **19** (11.7 g, 20.8 mmol, 95%) was obtained after purification by flash chromatography (SiO₂, 1% EtOAc in heptane) as a clear, colorless oil. All spectroscopic and physical data were in agreement with those reported in the literature.¹ *R*_f (2.5% EtOAc in heptane, visualized by KMnO₄ stain) = 0.61; $[\alpha]_D^{20} = -9.4$ (*c* 1.0, MeOH) [Lit.¹ $[\alpha]_D^{20} = -8.9$ (*c* = 1.0, MeOH)]; ¹H NMR (400 MHz, CDCl₃): δ 4.17 (ddd, *J* = 9.5, 2.6, 1.4 Hz, 1H), 3.93 (dd, *J* = 11.0, 3.7 Hz, 1H), 3.69 (ddd, *J* = 7.2, 5.8, 1.4 Hz, 1H), 3.53 – 3.42 (m, 2H), 2.72 – 2.54 (m, 4H), 2.06 (ddd, *J* = 14.6, 9.4, 3.7 Hz, 1H), 1.79 (ddd, *J* = 14.6, 11.0, 2.6 Hz, 1H), 1.24 (td, *J* = 7.5, 6.3 Hz, 6H), 0.90 – 0.89 (m, 27H), 0.11 – 0.05 (m, 18H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 77.8, 72.2, 64.5, 48.2, 39.0, 26.2 (3C), 26.1 (3C), 26.1 (3C), 24.6, 23.2, 18.4, 18.4, 18.3, 14.8, 14.5, -2.8, -3.5, -4.4, -4.7, -5.3, -5.3.

(3S,4R)-3,4,5-Tris((tert-butyldimethylsilyl)oxy)pentanal (6)



Compound **19** (5.48 g, 9.40 mmol, 1.00 equiv.) was dissolved in a mixture of acetone/water (64 mL/22 mL) and cooled to 0 °C, followed by addition of 2,6-lutidine (8.70 mL, 75.2 mmol, 8.00 equiv.) and NBS (13.4 g, 75.2 mmol, 8.00 equiv.). The reaction was stirred at 0 °C until completion as deemed by thin-layer chromatography. Then, the reaction mixture was added sat. aq. Na₂S₂O₃ (50 mL) and diluted with Et₂O (50 mL). The layers were separated, and the aq. phase was extracted with Et₂O (3 × 50 mL). The combined organic phase was washed with HCl (1.0 M, 20 mL), sat. aq. NaHCO₃ (20 mL) and brine (20 mL), dried (MgSO₄), filtrated and the solvent removed in vacuo. Aldehyde **6** (3.36 g, 7.05 mmol, 75%) was obtained after purification by flash chromatography (SiO₂, 1% EtOAc in hexane) as a clear, colorless oil. All spectroscopic and physical data were in agreement with those reported in the literature.¹ *R*₁ (1% EtOAc in hexane, visualized by KMnO₄ stain) = 0.20; [*a*]_D²⁰ = -6.0 (*c* 1.0, MeOH) [Lit.¹ [*a*]_D²⁰ = -5.9 (*c* 1.0, MeOH)]; ¹H NMR (400 MHz, CDCl₃): δ 9.85 (dd, *J* = 3.4, 1.7 Hz, 1H), 4.35 (td, *J* = 5.2, 2.2 Hz, 1H), 3.77 (ddd, *J* = 7.7, 5.2, 2.2 Hz, 1H), 3.51 – 3.47 (m, 1H), 3.43 – 3.39 (m, 1H), 2.59 (ddd, *J* = 16.3, 5.5, 3.4 Hz, 1H), 2.46 (ddd, *J* = 16.3, 4.9, 1.7 Hz, 1H), 0.89 (m, 18H), 0.87 (s, 9H), 0.09 (s, 6H), 0.07 (s, 6H), 0.05 (s, 6H); ¹³C{¹H</sup> NMR (100 MHz, CDCl₃): δ 202.8, 77.3, 69.7, 64.5, 45.8, 26.0 (6C), 26.0 (3C), 18.4, 18.3, 18.2, -4.2, -4.4, -4.5, -4.8, -5.3, -5.4.

(5*S*,6*R*)-6-((*Tert*-butyldimethylsilyl)oxy)-2,2,3,3,9,9,10,10-octamethyl-5-((*Z*)-pent-2-en-1-yl)-4,8-dioxa-3,9-disilaundecane (21)



Wittig salt **20** (1.14 g, 2.95 mmol, 1.20 equiv.) was dissolved in dry CH₂Cl₂ (18 mL) and cooled to -78 °C in a dry ice/acetone bath. NaHMDS (0.6 M in toluene, 4.92 mL, 2.95 mmol, 1.20 equiv.) was added in a dropwise manner. Aldehyde **6** (1.17 g, 2.46 mmol, 1.00 equiv.) was dissolved in CH₂Cl₂ (3 mL) and added dropwise to the orange solution. As this was added, the reaction mixture turned yellow. The reaction was stirred for 6 h at -78 °C, and then allowed to slowly warm up to room temperature overnight. After completion, the reaction was added phosphate buffer (25 mL, pH = 7) and diluted with Et₂O (30 mL). The layers were separated, and the aqueous phase was extracted with Et₂O (3 × 30 mL). The combined organic phase was dried (Na₂SO₄), filtrated and the solvent removed in vacuo. The desired *Z*-alkene **21** (1.02 g, 2.09 mmol, 82%, *Z/E* >98:2) was obtained after purification by flash chromatography (SiO₂, hexane) as a clear, colorless oil. All spectroscopic and physical data were in agreement with those reported in the literature.² *R*₁ (hexane, visualized by KMnO₄ stain) = 0.42; $[\alpha]_D^{25} = -5.3$ (*c* 0.2, CHCl₃) [Lit.² $[\alpha]_D^{20} = -5.3$ (*c* 0.2, CHCl₃)]; ¹H NMR (400 MHz, CDCl₃): δ 5.47 – 5.35 (m, 2H), 3.77 (ddd, *J* = 7.0, 5.5, 2.2 Hz, 1H), 3.69 (dt, *J* = 6.0, 2.2 Hz, 1H), 3.62 (dd, *J* = 10.1, 6.2 Hz, 1H), 3.47 (dd, *J* = 10.1, 5.9 Hz, 1H), 2.32 – 2.17 (m, 2H), 2.08 – 1.99 (m, 2H), 0.95 (t, *J* = 7.5 Hz, 3H), 0.89 (s, 9H), 0.89 (s, 9H), 0.88 (s, 9H), 0.07 (s, 3H), 0.06 (s, 3H), 0.05 (s, 3H), 0.04 (s, 3H), 0.04 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 133.1, 126.3, 77.3, 74.7, 64.9, 30.8, 26.1 (6C), 26.1 (3C), 20.8, 18.5, 18.4, 18.3, 14.3, -4.1, -4.3, -4.4, -5.2, -5.3.

(2R,3S,Z)-2,3-Bis((tert-butyldimethylsilyl)oxy)oct-5-en-1-ol (22)



TBS-triol **21** (2.27 g, 4.52 mmol, 1.00 equiv.) was dissolved in a mixture of MeOH/EtOH (52 mL, 1:1) and cooled to -18 °C. *p*-Toluenesulfonic acid (PTSA, 860 mg, 4.52 mmol, 1.00 equiv.) was added, and the resulting reaction mixture was stirred for 4 h at the same temperature. Then the reaction was added a sat. aq. solution of NaHCO₃ (40 mL) and diluted with EtOAc (30 mL). The layers were separated, and the aqueous phase was extracted with EtOAc (3 × 30 mL). The combined organic phase was washed with brine (30 mL), dried (MgSO₄), and the solvent removed in vacuo. Alcohol **22** (1.04 g, 2.67 mmol, 59%, 90% brsm) was obtained after purification by flash chromatography (SiO₂, 5% EtOAc in hexane), and unreacted starting material was reisolated. All spectroscopic and physical data were in agreement with those reported in the literature.² *R*₁ (5% EtOA in hexane, visualized by KMnO₄ stain) = 0.28; $[\alpha]_D^{25} = +2.8$ (c = 0.1, MeOH) [Lit.² $[\alpha]_D^{20} = +2.9$ (c 0.1, MeOH)]; ¹H NMR (400 MHz, CDCl₃): δ 5.55 – 5.34 (m, 2H), 3.80 – 3.71 (m, 2H), 3.65 – 3.58 (m, 2H), 2.36 – 2.22 (m, 2H), 2.08 – 2.00 (m, 2H), 0.96 (t, *J* = 7.6 Hz, 3H), 0.91 (s, 9H), 0.89 (s, 9H), 0.10 (s, 3H), 0.09 – 0.08 (m, 6H), 0.07 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 133.8, 124.7, 75.4, 74.8, 63.9, 32.1, 26.1 (3C), 26.0 (3C), 21.0, 18.3, 18.2, 14.2, -4.3, -4.3, -4.4, -4.5.

(2S,3S,Z)-2,3-Bis((tert-butyldimethylsilyl)oxy)oct-5-enal (9)



NaHCO₃ (1.02 g, 12.2 mmol, 5.70 equiv.) and Dess-Martin periodinane (DMP, 1.63 mg, 3.85 mmol, 1.80 equiv.) was added to a stirred solution of alcohol **22** (830 mg, 2.14 mmol, 1.00 equiv.) in CH₂Cl₂ (65.0 mL) at ambient temperature. The resulting suspension was stirred overnight and quenched by the addition of sat. aq. Na₂S₂O₃ (30 mL). The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 30 mL). The combined organic phase was washed with brine, dried (Na₂SO₄), filtrated, and the solvent removed in vacuo. The crude product was purified by flash chromatography (SiO₂, 2.5% EtOAc in hexane) to afford the desired aldehyde **9** (735 mg, 1.90 mmol, 89%) as a clear oil. All spectroscopic and physical data were in agreement with those reported in the literature.² *R*_f (5% EtOAc in hexane, visualized by KMnO₄ stain) = 0.48; $[\alpha]_D^{25} = +3.0$ (*c* 0.5, CHCl₃) [Lit.² $[\alpha]_D^{20} = +3.0$ (0.5, CHCl₃)]; ¹H NMR (400 MHz, CDCl₃): δ 9.56 – 9.55 (m, 1H), 5.53 – 5.43 (m, 1H), 5.29 – 5.21 (m, 1H), 3.95 – 3.89 (m, 2H), 2.47 – 2.39 (m, 1H), 2.24 – 2.18 (m, 1H), 2.11 – 1.98 (m, 2H), 0.96 (t, *J* = 7.5 Hz, 3H), 0.92 (s, 9H), 0.88 (s, 9H), 0.08 (s, 6H), 0.07 (s, 6H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 204.0, 135.1, 123.9, 81.1, 76.3, 31.7, 26.0 (3C), 26.0 (3C), 20.9, 18.4, 18.3, 14.2, -4.5, -4.5, -4.6, -4.7.

(2E,4R,5S,7Z)-4,5-Bis((tert-butyldimethylsilyl)oxy)deca-2,7-dienal (10)



To a solution of aldehyde **9** (650 mg, 1.68 mmol, 1.00 equiv.) in toluene (33 mL) was added (triphenylphosphoranylidene)acetaldehyde (510 mg, 1.68 mmol, 1.00 equiv.). The suspension was warmed to 95 °C in an oil bath and stirred for 6 h at that temperature. Then another equivalent of (triphenylphosphoranylidene)acetaldehyde (510 mg, 1.68 mmol, 1.00 equiv.). The suspension was warmed back to 95 °C and stirred for an additional 14 h. After cooling to room temperature, the solvent was evaporated, and the crude product was purified by flash chromatography (SiO₂, 2% EtOAc in hexane) to afford the α , β -unsaturated aldehyde **10** (490 mg, 1.19 mmol, 71%, 90% brsm, *E/Z* >98:2) as a yellow oil. Unreacted starting material was reisolated. *R*_f (5% EtOAc in hexane, visualized by UV and KMnO₄ stain) = 0.40; $[\alpha]_D^{25}$ = +13.2 (*c* 0.6, MeOH); ¹H NMR (400 MHz, CDCl₃): δ 9.58 (d, *J* = 8.0 Hz, 1H), 6.88 (dd, *J* = 15.7, 5.3 Hz, 1H), 6.24 (ddd, *J* = 15.7, 8.1, 1.5 Hz, 1H), 5.50 – 5.43 (m, 1H), 5.39 – 5.32 (m, 1H), 4.30 (ddd, *J* = 5.2, 3.6, 1.5 Hz, 1H), 3.74 (td, *J* = 6.2, 3.6 Hz, 1H), 2.32 – 2.20 (m, 2H), 2.07 – 1.99 (m, 2H), 0.96 (t, *J* = 7.5 Hz, 3H), 0.91 (s, 9H), 0.86 (s, 9H), 0.06 (s, 3H), 0.05 (s, 3H), 0.04 (s, 3H), 0.02 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 193.7, 157.7, 134.2, 132.5, 124.4, 75.4, 32.1, 26.0 (6C), 21.0, 18.4, 18.3, 14.2, -4.1, -4.3, -4.4; HRMS (ESI): Exact mass calculated for C₂₂H₄₄NaO₃Si₂: 435.2721, found 435.2721.

(5*R*,6*S*)-5-((1*E*,3*E*)-4-lodobuta-1,3-dien-1-yl)-2,2,3,3,8,8,9,9-octamethyl-6-((*Z*)-pent-2-en-1-yl)-4,7-dioxa-3,8disiladecane (4)



Vinyl iodide 4 was prepared according to the published procedure of Takai and coworkers.³ CrCl₂ (119 mg, 0.969 mmol, 8.00 equiv.) was placed in a flask and dried under high vacuum using a heat gun. After cooling to room temperature, the flask was protected from light and dry THF (0.17 mL) and dioxane (2.95 mL) was added. The suspension was cooled to 0 °C before CHI₃ (119 mg, 0.303 mmol, 2.50 equiv.) was added in one portion. The resulting mixture was stirred at ambient temperature for 2 h, wherein the suspension turned from dark-green to red-brown. Then, the reaction mixture was cooled back to 0 °C, followed by dropwise addition of the α,β -unsaturated aldehyde **10** (50.0 mg, 0.121 mmol, 1.00 equiv.) dissolved in dioxane (0.21 mL). Stirring was continued for another 2 h at room temperature. When deemed complete, the reaction mixture was added sat. aq. NH₄Cl (2.5 mL), and the aqueous phase was extracted with Et₂O (3 × 3 mL). The combined organic layers were washed successively with a saturated aqueous solution of Na₂S₂O₃ (5 × 2 mL) and brine (5 × 2 mL), dried (Na₂SO₄), filtrated, and concentrated in vacuo. The crude product was purified by flash chromatography (SiO₂, hexane) to yield the desired product 4 (53 mg, 0.100 mmol, 82%) as a clear oil. R_f (hexane, visualized by UV and KMnO₄ stain) = 0.32; $[\alpha]_D^{20}$ = +1.1 (*c* 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.04 (ddd, *J* = 14.4, 10.7, 0.8 Hz, 1H), 6.27 (d, J = 14.4 Hz, 1H), 6.04 (dd, J = 15.4, 10.6 Hz, 1H), 5.72 (dd, J = 15.5, 6.9 Hz, 1H), 5.46 - 5.31 (m, 2H), 3.98 (ddd, J = 15.4, 10.6 Hz, 1H), 5.72 (dd, J = 15.5, 6.9 Hz, 1H), 5.46 - 5.31 (m, 2H), 3.98 (ddd, J = 15.4, 10.6 Hz, 1H), 5.72 (dd, J = 15.5, 6.9 Hz, 1H), 5.46 - 5.31 (m, 2H), 3.98 (ddd, J = 15.4, 10.6 Hz, 1H), 5.72 (dd, J = 15.5, 6.9 Hz, 1H), 5.46 - 5.31 (m, 2H), 3.98 (ddd, J = 15.4, 10.6 Hz, 1H), 5.72 (dd, J = 15.4, 10.6 Hz, 1H), 5.72 (dd, J = 15.5, 6.9 Hz, 1H), 5.46 - 5.31 (m, 2H), 3.98 (ddd, J = 15.4, 10.6 Hz, 1H), 5.72 (dd, J = 15.4, 10.6 Hz, 1H), 5.46 - 5.31 (m, 2H), 3.98 (ddd, J = 15.4, 10.6 Hz, 1H), 5.72 (dd, J = 15.5, 6.9 Hz, 1H), 5.46 - 5.31 (m, 2H), 3.98 (ddd, J = 15.4, 10.6 Hz, 1H), 5.46 - 5.31 (m, 2H), 3.98 (ddd, J = 15.4, 10.6 Hz, 1H), 5.46 - 5.31 (m, 2H), 5.46 (m, 2H), 5. 7.1, 4.1, 1.1 Hz, 1H), 3.61 (td, J = 6.1, 4.0 Hz, 1H), 2.26 - 2.13 (m, 2H), 2.05 - 1.98 (m, 2H), 0.95 (t, J = 7.5 Hz, 1H), 0.88 (s, 9H), 0.86 (s, 9H), 0.03 - 0.02 (m, 9H), 0.00 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 145.1, 135.4, 133.5, 131.1, 125.3, 78.6, 76.7, 76.4, 31.8, 26.1 (3C), 26.1 (3C), 20.9, 18.4, 18.3, 14.3, -4.0, -4.0, -4.3, -4.5; HRMS (ESI): Exact mass calculated for C23H45INaO2Si2: 559.1895, found 559.1895. This compound is sensitive to heat, light and oxygen. Prudent care should be made during handling and storage. The above procedure was repeated on a 0.5 mmol scale of compound 10.

7-Methoxy-7-oxoheptanoic acid (11)



The selective monohydrolysis of diester **7** was performed according to the procedure of Niwayam.⁴ Compound **7** (6.00 g, 31.8 mmol, 1.00 equiv.) was dissolved in THF (50 mL), and H₂O (500 mL) was added. The solution was cooled to 0 °C in an ice bath. To this mixture was added 0.25 M NaOH (200 mL) in small portions until detection of the dicarboxylic acid was observed by thin-layer chromatography. Then, the reaction was immediately acidified by addition of 1.0 M HCl (75 mL) at 0 °C, saturated with brine (100 mL), and extracted with EtOAc (3 × 200 mL). The combined organic layer was dried (Na₂SO₄), filtrated, and the solvent removed in vacuo. The crude product was purified by flash chromatography (SiO₂, hexane/EtOAc, 1:1) to afford the desired product **11** (3.90 g, 22.3 mmol, 70%) as a clear oil. *R*_f (60% EtOAc in hexane, visualized by bromocrestol green stain) = 0.30; ¹H NMR (400 MHz, CD₃OD): δ 4.87 (s, 1H), 3.65 (s, 3H), 2.31 (dt, *J* = 17.0, 7.4 Hz, 4H), 1.62 (pd, *J* = 7.5, 5.8 Hz, 4H), 1.40 – 1.32 (m, 2H); ¹³C(¹H) NMR (100 MHz, CD₃OD): δ 177.5, 175.8, 52.0, 34.7, 34.6, 29.6, 25.7, 25.7; HRMS (ESI): Exact mass calculated for C₈H₁₄NaO₄: 197.0784, found 197.0784.

Methyl 7-oxo-9-(trimethylsilyl)non-8-ynoate (12)



Ketone 12 was prepared according to the published procedure of Rodriguez and Spur.⁵ Anhydrous N,Ndimethylformamide (30 µL, 1.95 x 10⁻³ mmol, 0.01 mol%) was added to a stirred solution of carboxylic acid 11 (3.30 g, 18.9 mmol, 1.00 equiv.) in anhydrous CH₂Cl₂ (51 mL). The reaction mixture was cooled to 0 °C, and oxalyl chloride (3.30 mL, 37.8 mmol, 2.00 equiv.) was added in a dropwise manner. The resulting solution was allowed to slowly warm to room temperature and stirred overnight. After removal of the volatile compounds by reduced pressure, the crude product was dissolved in anhydrous CH₂Cl₂ (24 mL) and transferred to a flame-dried flask with bis(trimethylsilyl)acetylene (3.21 g, 18.9 mmol, 1.00 equiv.) under argon. The resulting solution was added in a dropwise manner to a stirred solution of AICI₃ (3.27 g, 24.6 mmol, 1.30 equiv.) in anhydrous CH₂Cl₂ (24 mL) at 0 °C. The resulting reaction mixture was stirred at 0 °C for 30 min, warmed up to room temperature over a period of 45 min and then cooled back down to 0 °C. Then, the reaction mixture was added 1 M HCl (50 mL) and stirred for an additional 10 min. The resulting thick suspension was vacuumfiltrated through a short plug of silica gel directly into a separatory funnel and the plug was washed with additional fresh CH₂Cl₂ (50 mL). The phases were separated and the aqueous phase was extracted with CH₂Cl₂ (3 × 50 mL). The combined organic phase was dried (Na₂SO₄) and concentrated in vacuo. The crude product was purified by flash chromatography (SiO₂, 10% EtOAc in hexane) to yield the desired product 12 (2.85 g, 11.2 mmol, 59%) as a colorless oil. The spectroscopic data were in agreement with those reported in literature.⁵ $R_{\rm f}$ (10% EtOAc in hexane, visualized by UV and KMnO₄ stain) = 0.25; ¹H NMR (400 MHz, CDCl₃): δ 3.66 (s, 3H), 2.56 (td, J = 7.3, 2.6 Hz, 2H), 2.31 (t, J = 7.5 Hz, 2H), 1.69 - 1.62 (m, 4H), 1.38 – 1.30 (m, 2H), 0.23 (s, 9H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 187.8, 174.2, 102.1, 97.9, 51.6, 45.1, 33.9, 28.5, 24.7, 23.6, -0.6 (3C); HRMS (ESI): Exact mass calculated for C₁₃H₂₂NaO₃Si: 277.1230, found 277.1230.

Methyl (S)-7-hydroxy-9-(trimethylsilyl)non-8-ynoate ((S)-13)



Ketone 12 (1.80 g, 7.08 mmol, 1.00 equiv.) was azeotropically dried with 2-methyltetrahydrofuran (2 x 2 mL) and then placed under high vacuum for 30 min. The flask was vented with argon and cooled to -10 °C. (S)-Alpine-borane solution (28.0 mL, 0.5 M in THF, 14.2 mmol, 2.00 equiv.) was added in one portion, and most of the THF solvent was immediately removed in vacuo with efficient stirring while warming up to 0° C. The resulting, highly viscous reaction mixture was then allowed to warm to ambient temperature and stirred overnight. Next, the reaction mixture was cooled to 0 °C and acetaldehyde (0.80 mL, 620 mg, 0.785 g/mL at 25 °C, 2.00 equiv.) was added dropwise. After 15 min, Et₂O (28.0 mL) was added, followed by dropwise addition of ethanolamine (1.72 mL, 1.72 g, 1.012 g/mL, 4.00 equiv.). The reaction was stirred for 30 min at 0 °C, then warmed to room temperature and stirred for an additional 1 h. The white, solid 9-BBN/ethanolamine complex was removed by filtration and the filtrate was washed with water (2 × 20 mL). The organic phase was dried (Na₂SO₄), filtrated, concentrated in vacuo and then purified by flash chromatography (SiO₂, gradient elution, 10 - 20% EtOAc in hexane) to give the desired product (S)-13 (1.72 mg, 6.72 mmol, 95%, 93% ee (see below)) as a clear oil. All spectroscopic data were in agreement with those reported in literature.⁵ Rf (20% EtOAc in hexane, visualized by KMnO₄ stain) = 0.29; $[\alpha]_D^{20}$ = -0.56 (c 1.3, CHCl₃) [Lit.⁵ $[\alpha]_D^{20}$ = -0.50 (c 0.63, CHCl₃)]; ¹H NMR (400 MHz, CD₃OD): δ 4.27 (t, J = 6.7 Hz, 1H), 3.65 (s, 3H), 2.33 (t, J = 7.4 Hz, 2H), 1.67 - 1.59 (m, 4H), 1.49 - 1.42 (m, 2H), 1.40 - 1.34 (m, 2H), 0.15 (s, 9H); ¹³C{¹H} NMR (100 MHz, CD₃OD): δ 175.9, 108.8, 88.8, 63.0, 52.0, 38.6, 34.7, 29.7, 26.0, 25.9, -0.1 (3C); HRMS (ESI): Exact mass calculated for $C_{13}H_{24}NaO_3Si$: 279.1387, found 279.1386.

Methyl 7-hydroxy-9-(trimethylsilyl)non-8-ynoate (rac 13)



Ketone **12** (50 mg, 0,197 mmol, 1.00 equiv.) was azeotropically dried with 2-methyltetrahydrofuran (2 × 1 mL) and then placed under high vacuum for 30 min. The flask was cooled to 0 °C and 9-borabicyclo[3.3.1]nonane solution (9-BBN, 0.80 mL, 0.5 M in THF, 0.393 mmol, 2.00 equiv.) was added. Approximately half the solvent volume was removed under vacuum at room temperature, and the reaction was stirred for 72 h. Acetaldehyde (11 μ L, 0.197 mmol, 1.00 equiv.) was added in a dropwise manner and the reaction was allowed to stir for an additional 1 h. The reaction mixture was diluted with Et₂O (1.1 mL), then ethanolamine (12 μ L, 0.197 mmol, 1.00 equiv.) was added. After 30 min, the suspension was concentrated in vacuo. H₂O (2 mL) was added, and the aqueous phase was extracted with Et₂O (3 × 2 mL). The combined organic phases were dried (Na₂SO₄), filtrated and concentrated in vacuo. The crude product was purified by flash chromatography (SiO₂, gradient elution, 10 - 20% EtOAc in hexane) to give racemic **13** (41 mg, 0.160 mmol, 81%) as a clear oil. The obtained spectral data matched those given for compound (*S*)-**13**.

(S)-9-Methoxy-9-oxo-1-(trimethylsilyl)non-1-yn-3-yl 2-naphthoate ((S)-23)



The propargylic alcohol (S)-13 (30 mg, 0.117 mmol, 1.00 equiv.) was dissolved in CH₂Cl₂ (0.50 mL) and cooled to 0 °C. Triethylamine (0.049 mL, 0.327 mmol, 3.00 equiv.) was added dropwise followed by 4-(dimethylamino)pyridine (DMAP, 1.40 mg, 0.0117 mmol, 10 mol%). Next, 2-naphthoyl chloride (27 mg, 0.140 mmol, 1.20 equiv.) was added, and the resulting suspension was allowed to slowly warm up to room temperature and stirred overnight. The solvent was removed under a gentle stream of argon, and then hexane (2.5 mL) and phosphate buffer (2.5 mL) was added. The phases were separated after 5 min of stirring, and the aqueous phase was extracted in the same manner two more times. The combined organic phase was dried (Na₂SO₄), filtrated and the solvent removed in vacuo. The material thus obtained was purified by flash chromatography (SiO₂, gradient elution, 0 - 10% EtOAc in hexane) to afford the desired compound (S)-23 (44 mg, 0.107 mmol, 91%) as a clear oil. The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/*i*PrOH 99/1, 1 mL/min): t_r (e₁, minor) = 10.89 min and t_r (e₂, major) = 11.76 min. R_f (10% EtOAc in hexane, visualized by UV and KMnO₄ stain) = 0.32; $[\alpha]_{D}^{20}$ = +15.0 (*c* 0.7, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.64 – 8.63 (m, 1H), 8.08 (dd, J = 8.6, 1.7 Hz, 1H), 7.97 (dd, J = 8.1, 1.5 Hz, 1H), 7.89 (dd, J = 8.5, 2.2 Hz, 2H), 7.58 (dddd, J = 19.7, 8.2, 6.9, 1.4 Hz, 2H), 5.70 (t, J = 6.6 Hz, 3H), 3.66 (s, 3H), 2.33 (t, J = 7.5, 2H), 1.98 - 1.91 (m, 2H), 1.78 (p, J = 7.5 Hz, 2H), 1.61 - 1.54 (m, 2H), 1.46 - 1.40 (m, 2H), 0.19 (s, 9H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.3, 165.8, 135.8, 132.6, 131.5, 129.6, 128.5, 128.3, 127.9, 127.4, 126.8, 125.5, 102.8, 90.8, 65.1, 51.6, 34.9, 34.1, 28.8, 24.9, -0.02 (3C); HRMS (ESI): Exact mass calculated for $C_{24}H_{30}NaO_4Si$: 433.1806, found 433.1806.

The exact same procedure was followed for racemic **13**. Yield: 56 mg (93%). The obtained spectral data matched those given for compound (S)-**23**.

Methyl (S)-7-((tert-butyldimethylsilyl)oxy)-9-(trimethylsilyl)non-8-ynoate (24)



Imidazole (720 mg, 10.3 mmol, 2.00 equiv.) and *tert*-butyldimethylsilyl chloride (930 mg, 6.18 mmol, 1.50 equiv.) were added in a successive manner to a stirred solution of propargylic alcohol (*S*)-**13** (1.32 g, 5.16 mmol, 1.00 equiv.) in CH₂Cl₂ (15 mL) at room temperature. The reaction was stirred overnight and quenched by the addition of a half-sat. aq. solution of NaH₂PO₄ (40 mL). The phases were separated, and the aqueous phase was extracted with CH₂Cl₂ (3 × 40 mL). The combined organic phases were dried (Na₂SO₄), filtrated, and the solvent removed in vacuo. The desired product **24** (1.79 mg, 4.85 mmol, 94%) was obtain as a clear oil after purification by flash chromatography (SiO₂, 2.5% EtOAc in hexane). All spectroscopic data were in agreement with those reported in literature.⁵ *R*_f (5% EtOAc in hexane, visualized by UV and KMnO₄ stain) = 0.32; $[\alpha]_D^{20}$ = -36.3 (*c* 0.4, CHCl₃) [Lit.⁵ $[\alpha]_D^{25}$ = -37.0 (*c* 0.6, CHCl₃)]; ¹H NMR (400 MHz, CDCl₃): δ 4.30 (t, *J* = 6.6 Hz, 1H), 3.65 (s, 3H), 2.30 (t, *J* = 7.5 Hz, 2H), 1.67 – 1.58 (m, 4H), 1.47 – 1.30 (m, 4H), 0.89 (s, 9H), 0.14 (s, 9H), 0.12 (s, 3H), 0.09 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.3, 107.9, 88.6, 63.4, 51.6, 38.4, 34.1, 28.9, 26.0 (3C), 25.0, 25.0, 18.4, -0.03 (3C), -4.3, -4.8; HRMS (ESI): Exact mass calculated for C₁₉H₃₈NaO₃Si₂: 393.2252, found 393.2251.

Methyl (S)-7-((tert-butyldimethylsilyl)oxy)non-8-ynoate (5)



TMS-protected alkyne **24** (1.50 g, 4.05 mmol, 1.00 equiv) was dissolved in methanol (39 mL) and cooled to 0 °C. K₂CO₃ (570 mg, 4.05 mmol, 1.00 equiv.) was added in one portion, and the reaction was warmed to room temperature. The reaction was followed with TLC analysis (5% EtOAc in hexane) until completion (the product can be observed just below the starting material). After 2 h of stirring, the reaction mixture was cooled back to 0 °C, quenched by the addition of phosphate buffer (45 mL, pH = 7) and stirred for an additional 5 min. NaCl (~3 g) was added, and the aqueous phase was extracted with hexane (5 × 45 mL). The combined organic phase was dried (MgSO₄), filtrated and concentrated in vacuo. The crude product was purified by flash chromatography (SiO₂, 5% EtOAc in hexane) to give the desired compound **5** (1.15 g, 3.85 mmol, 95%) as a clear oil. All spectroscopic data were in agreement with those reported in literature.⁵ *R*_f (5% EtOAc in hexane, visualized by KMnO₄ stain) = 0.25; $[\alpha]_D^{20} = -34.9$ (*c* 1.5, CHCl₃) [Lit.⁵ $[\alpha]_D^{20} = -36.0$ (*c* 0.5, CHCl₃)]; ¹H NMR (400 MHz, CDCl₃): δ 4.33 (td, *J* = 6.4, 2.1 Hz, 1H), 3.66 (s, 3H), 2.36 (d, *J* = 2.1 Hz, 1H), 2.30 (t, *J* = 7.5 Hz, 3H), 1.70 - 1.60 (m, 4H), 1.49 - 1.40 (m, 2H), 1.37 - 1.32 (m, 2H), 0.89 (s, 9H), 0.13 (s, 3H), 0.10 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.3, 85.7, 72.1, 62.8, 51.6, 38.5, 34.1, 28.9, 25.9 (3C), 25.0, 24.9, 18.4, -4.4, -4.9; HRMS (ESI): Exact mass calculated for C₁₆H₃₀NaO₃Si: 321.1856, found 321.1856.

Methyl (S,E)-7-((tert-butyldimethylsilyl)oxy)-9-(tributylstannyl)non-8-enoate (25)



Compound **25** was prepared by following the protocol of Sulikowski and coworkers.⁶ Alkyne **5** (980 mg, 3.29 mmol, 1.00 equiv.) was dissolved in benzene (68 mL). Bu₃SnH (2.70 mL, 10.1 mmol, 3.00 equiv.) and AIBN (90.0 mg, 0.550 mmol, 16 mol%) was added. The reaction was heated to 80 °C with stirring. After 2 h, the reaction mixture was cooled back to room temperature and the solvent was removed in vacuo. The crude product was purified by flash chromatography (SiO₂, 2.5% EtOAc in hexane) to obtain the stannane product **25** (1.80 g, 3.06 mmol, 93%) as a clear oil. *R*_I (5% EtOAc in hexane, visualized by KMnO₄ stain) = 0.48; $[\alpha]_D^{20} = -12.5$ (*c* 1.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.00 (d, *J* = 19.0 Hz, 1H), 5.89 (dd, *J* = 19.0, 5.6 Hz, 1H), 4.00 (q, *J* = 5.9 Hz, 1H), 3.66 (s, 3H), 2.29 (t, *J* = 7.6 Hz, 2H), 1.64 – 1.25 (m, 26H), 0.90 – 0.86 (m, 18H), 0.03 (s, 3H), 0.02 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.4, 152.0, 126.6, 51.6, 38.0, 34.2, 29.3, 29.3 (3C), 27.4 (3C), 26.1 (3C), 25.2, 25.1, 18.5, 13.9 (3C), 9.6 (3C), -4.1, -4.6; HRMS (ESI): Exact mass calculated for C₂₈H₅₈NaO₃Si¹¹⁶Sn: 609.3065, found 609.3064.

Methyl (S,E)-7-((tert-butyldimethylsilyl)oxy)-9-iodonon-8-enoate (14)



Stannane **25** (1.66 g, 2.82 mmol, 1.00 equiv.) was dissolved in dry CH₂Cl₂ (8.0 mL). A solution of I₂ (1.07 g, 4.21 mmol, 1.50 equiv.) in dry CH₂Cl₂ (11 mL) was added dropwise at room temperature until the resulting solution maintained a pink color. The reaction was stirred for 20 min, followed by addition of saturated aqueous Na₂S₂O₃ (5 mL), H₂O (5 mL), and saturated aqueous NaHCO₃ (5 mL). After stirring for 5 min, the phases were separated and the aqueous phase was extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄), filtrated, and the solvent removed in vacuo. The crude product was purified by flash column chromatography (SiO₂, 5% Et₂O in hexane) to obtain the vinyl iodide **14** (1.13 g, 2.65 mmol, 94%) as a clear oil. All spectroscopic and physical data were in agreement with those previously reported.⁵ *R*₁ (5% Et₂O in hexane, visualized by UV and KMnO₄ stain) = 0.26; $[\alpha]_D^{20} = -17.7$ (*c* 0.9, CHCl₃) [Lit.⁵ $[\alpha]_D^{20} = -18.5$ (*c* 0.4, CHCl₃)]; ¹H NMR (400 MHz, CDCl₃): δ 6.49 (dd, *J* = 14.4, 6.0 Hz, 1H), 6.20 - 6.16 (m, 1H), 4.05 (qd, *J* = 6.0, 1.3 Hz, 1H), 3.66 (s, 3H), 2.29 (t, *J* = 7.5 Hz, 2H), 1.65 - 1.59 (m, 2H), 1.50 - 1.42 (m, 2H), 1.36 - 1.28 (m, 4H), 0.88 (m, 9H), 0.03 (s, 3H), 0.02 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.3, 149.4, 75.8, 75.2, 51.6, 37.4, 34.2, 29.2, 26.0 (3C), 25.0, 24.6, 18.4, -4.4, -4.7; HRMS (ESI): Exact mass calculated for C₁₆H₃₁INaO₃Si: 449.0979, found 449.0979. This compound is sensitive to heat, light and oxygen and prudent care should be made when handling and storing it.

Methyl (S,E)-7-((tert-butyldimethylsilyl)oxy)undec-8-en-10-ynoate (3)



Vinyl iodide **14** (1.06 g, 2.49 mmol, 1.00 equiv.) was dissolved in dry THF (14.0 mL) and cooled to 0 °C. Pd(PPh₃)₂Cl₂ (90 mg, 0.12 mmol, 5 mol%) and Cul (57 mg, 0.30 mmol, 12 mol%) was added in one portion followed by dropwise addition of Et₃N (0.50 g, 0.70 mL, 4.98 mmol, 2.00 equiv.). Ethynyltrimethylsilane (62 mg, 0.90 mL, 6.23 mmol, 2.50 equiv.) was dissolved in a minimum amount of dry THF and added in a dropwise manner. The reaction mixture was allowed to slowly warm up to room temperature and stirred in the dark for 18 h. After completion, the suspension was filtrated through a plug of silica gel (10% EtOAc in hexane). The solvent was removed in vacuo, and the crude product thus obtained was used directly in the next reaction without any further purification. $R_{\rm f}$ (5% EtOAc in hexane, visualized by UV and KMnO₄ stain) = 0.26.

The crude TMS-acetylene **15** was dissolved in methanol (22 mL) and cooled to 0 °C. K₂CO₃ (690 mg, 4.99 mmol, 2.00 equiv.) was added in one portion, and the reaction was warmed to room temperature. The reaction was followed with TLC analysis (10% EtOAc in hexane) until completion (the product can be observed just below the starting material). After 2 h of stirring, the reaction mixture was cooled back to 0 °C, quenched by the addition of phosphate buffer (30 mL, pH = 7) and stirred for an additional 5 min. NaCl (1.80 g) was added, and the aqueous phase was extracted with hexane (5 × 30 mL). The combined organic phases were dried (Na₂SO₄), filtrated and concentrated in vacuo. The crude product was purified by flash chromatography (SiO₂, 5% EtOAc in hexane) to give the desired compound **3** (655 mg, 2.02 mmol, 81% over two steps) as a clear oil. *R*_f (5% EtOAc in hexane, visualized by KMnO₄ stain) = 0.15; $[\alpha]_D^{20}$ = -7.3 (*c* 0.3, MeOH); ¹H NMR (400 MHz, CDCl₃): δ 6.24 – 6.18 (ddd, *J* = 15.9, 5.3, 0.6 Hz, 1H), 5.65 – 5.60 (ddd, *J* = 15.9, 2.3, 1.6 Hz, 1H), 4.18 4.14 (qd, *J* = 5.8, 1.6 Hz, 1H), 3.67 (s, 3H), 2.87 – 2.86 (d, *J* = 2.3 Hz, 1H), 2.32 – 2.28 (t, *J* = 7.5 Hz, 2H), 1.65 – 1.58 (m, 2H), 1.51 – 1.44 (m, 2H), 1.36 – 1.27 (m, 4H), 0.89 (s, 9H), 0.04 (s, 3H), 0.03 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ

174.3, 148.6, 107.7, 82.2, 77.4, 72.3, 51.6, 37.6, 34.2, 29.3, 26.0 (3C), 25.0, 24.6, 18.3, -4.4, -4.7; HRMS (ESI): Exact mass calculated for $C_{18}H_{32}NaO_3Si$: 347.2013, found 347.2013.

Methyl (7*S*,8*E*,12*E*,14*E*,16*R*,17*S*,19*Z*)-7,16,17-tris((*tert*-butyldimethylsilyl)oxy)docosa-8,12,14,19-tetraen-10-ynoate (16)



To a stirred solution of vinyliodine 4 (120 mg, 0.224 mmol, 1.00 equiv.) in benzene (0.4 mL) and Et₂NH (0.96 mL) was added Pd(PPh₃)₄ (7.8 mg, 6.8 µmol, 3 mol%) at ambient temperature and the reaction mixture was stirred for 30 min in the dark. Cul (4.0 mg, 21 µmol, 9 mol%) and a solution of alkyne 3 (72 mg, 0.224 mmol, 1.00 equiv.) in a minimum amount of Et₂NH were added in a sequential manner and the resulting suspension was stirred overnight. After completion, the reaction mixture was guenched by the addition of sat. aq. NH₄Cl (6 mL) and diluted with Et₂O (6 mL). The layers were separated, and the aqueous layer was extracted (Et₂O, 3 × 6 mL). The combined organic phase was washed with brine (6 mL), dried over MgSO₄, filtrated, and the solvent removed in vacuo. Alkyne 16 (156 mg, 0.212 mmol, 94%) was obtained as a pale yellow oil after purification by flash column chromatography (SiO₂, 1% EtOAc in hexane). R_f (10% EtOAc in hexane, visualized by UV and KMnO₄ stain) = 0.50; $[\alpha]_D^{20}$ = -15.5 (c 1.1, MeOH); ¹H NMR (400 MHz, C₆D₆): δ 6.65 (dd, J = 15.4, 10.8 Hz, 1H), 6.22 - 6.15 (m, 2H), 5.98 (dt, J = 15.8, 1.9 Hz, 1H), 5.82 - 5.73 (m, 2H), 5.59 - 5.46 (m, 2H), 4.10 (dd, J = 6.9, 4.5 Hz, 1H), 3.72 (td, J = 5.8, 4.4, Hz, 1H), 3.36 (s, 3H), 2.40 (t, J = 6.1 Hz, 2H), 2.11 - 2.02 (m, 4H), 1.55 -1.47 (m, 2H), 1.42 - 1.32 (m, 2H), 1.27 - 1.18 (m, 2H), 1.17 - 1.09 (m, 2H), 1.00 (s, 9H), 0.99 (s, 9H), 0.96 (s, 9H), 0.94 (t, J = 7.5 Hz, 3H), 0.10 (s, 3H), 0.09 (s, 6H), 0.07 (s, 3H), 0.05 (s, 3H), 0.03 (s, 3H); ¹³C{¹H} NMR (100 MHz, C₆D₆): δ 173.3, 146.6, 141.2, 137.2, 133.6, 131.4, 125.6, 112.0, 109.7, 91.5, 90.2, 77.1, 76.9, 72.9, 50.9, 38.0, 34.0, 32.0, 29.4, 26.2 (6C), 26.1 (3C), 25.2, 25.0, 21.3, 18.5, 18.4 (2C), 14.4, -3.8, -3.9, -4.1, -4.3, -4.5, -4.7; HRMS (ESI): Exact mass calculated for C₄₁H₇₆NaO₅Si₃: 755.4893, found 755.4892.

Methyl (7S,8E,12E,14E,16R,17S,19Z)-7,16,17-trihydroxydocosa-8,12,14,19-tetraen-10-ynoate (17)



TBS-protected triol **16** (81.0 mg, 0.110 mmol, 1.00 equiv.) was dissolved in THF (1.8 mL) and cooled to 0 °C. TBAF (1 M in THF, 0.828 mL, 0.828 mmol, 7.50 equiv.) was added dropwise, and the resulting mixture was stirred over night. After completion, the reaction mixture was quenched by the addition of phosphate buffer (pH = 7, 1.5 mL). Brine (7.5 mL) and EtOAc (9 mL) was added, and the phases were separated. The aqueous phase was extracted with EtOAc (3 × 9 mL), and the combined organic layer was dried (Na₂SO₄), filtrated and the solvent removed in vacuo. The crude product was purified by flash chromatography (SiO₂, EtOAc/heptane 1:1) to obtain the desired triol **17** (40.2 mg, 0.103 mmol, 93%) as a clear oil. *R*₁ (EtOAc/Heptane 1:1, visualized by UV and KMnO₄ stain) = 0.19; $[\alpha]_D^{20} = +40.0$ (*c* 0.9, MeOH); ¹H NMR (400 MHz, CD₃OD): δ 6.58 (dd, *J* = 15.4, 10.8 Hz, 1H), 6.35 (dd, *J* = 15.6, 10.7 Hz, 1H), 6.07 (dd, *J* = 15.8, 6.1 Hz, 1H), 5.92 (dd, *J* = 15.3, 6.6 Hz, 1H), 5.82 (ddd, *J* = 15.8, 2.4, 1.4 Hz, 1H), 5.75 (dd, *J* = 15.4, 2.4 Hz, 1H), 5.53 – 5.39 (m, 2H), 4.08 (qd, *J* = 6.2, 1.4 Hz, 1H), 4.00 (ddd, *J* = 6.5, 5.0, 1.2 Hz, 1H), 3.52 (dt, *J* = 8.2, 4.9 Hz, 1H), 2.33 (t, *J* = 7.4 Hz, 2H), 2.19 – 2.11 (m, 1H), 2.10 – 2.01 (m, 2H), 1.62 (p, *J* = 7.4 HZ, 1H), 1.54 – 1.47 (m, 2H), 1.41 – 1.29 (m, 4H), 0.97 (t, *J* = 7.5 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CD₃OD): δ 176.0, 147.1, 142.2, 136.7, 134.4, 132.3, 126.2, 112.3, 110.5, 91.3, 90.0, 76.0, 75.9, 72.7, 52.0, 37.8, 34.7, 31.8, 30.1, 26.1, 25.9, 21.7, 14.6; HRMS (ESI): Exact mass calculated for C₂₃H₃₄NaO₅: 413.2298, found 413.2297.

RvD2_{n-3 DPA} methyl ester (2)



Compound 17 was prepared by following the protocol reported by Näf et al.⁷ Compound 17 (20.0 mg, 51.2 µmol, 1.00 equiv.) was dissolved in a 1:1 mixture of 1-propanol/ H_2O (2.4 mL, pH = 7.0). Zinc powder (730 mg, 11.2 mmol, 218 equiv.) and KCN (47.0 mg, 0.722 mmol, 14.1 equiv.) was added in a successive manner, and the resulting suspension was stirred at ambient temperature under protection of argon in the dark. After stirring for 20 hours, the reaction mixture was filtered through a short pad of Celite® and the pad was washed with EtOAc (10 mL). Sat aq. NH₄Cl (8 mL) was added to the filtrate and the phases were separated. The aqueous phase was extracted with EtOAc (3 × 6 mL), and the combined organic layer was concentrated in vacuo, but not to complete dryness [R_f (10% MeOH in CH₂Cl₂, visualized by UV and KMnO₄ stain) = 0.26]. The crude product was immediately dissolved in a mixture of toluene/MeOH (3:2, 0.92 mL), followed by dropwise addition of TMS-diazomethane (2.0 M in Et₂O, 33.3 µL, 66.6 µmol, 1.30 equiv.) until the yellow color persisted. After completion (~1 h), the reaction was quenched with sat. aq. NH₄Cl (2 mL). The layers were separated, and the aq. phase was extracted with Et₂O (3 × 2 mL). The combined organic layer was evaporated, and the crude product was purified by flash column chromatography (SiO₂, gradient elution, 2 – 5% MeOH in CH₂Cl₂) to afford RvD2_{n-3 DPA} methyl ester 2 (12.5 mg, 31.8 µmol, 59%) as a clear oil. The chemical purity (>96%) was determined by HPLC analysis (Eclipse XDBC₁₈, MeOH/H₂O 70:30, 1.0 mL/min): t_r (major) = 11.16 min, t_r (minor) = 12.10 min. R_f (5% MeOH in CH₂Cl₂, visualized by UV and KMnO₄ stain) = 0.26; $[\alpha]_{D}^{20}$ = +23.8 (c 0.3, MeOH); ¹H NMR (400 MHz, CD₃OD): δ 6.76 (ddd, J = 14.2, 9.9, 3.4 Hz, 2H), 6.39 (dd, J = 14.8, 11.5 Hz, 1H), 6.30 (dd, J = 14.3, 10.8 Hz, 1H), 6.06 - 5.98 (m, 2H), 5.85 (ddd, J = 14.9, 12.0, 5.3 Hz, 1H), 5.72 (dd, J = 15.1, 6.6 Hz, 1H), 5.60 – 5.45 (m, 2H), 4.15 (q, J = 6.4 Hz, 1H), 4.04 – 3.97 (m, 1H), 3.66 (s, 3H), 3.56 - 3.49 (m, 1H), 2.34 (t, J = 7.4 Hz, 2H), 2.27 - 2.10 (m, 2H), 2.04 (p, J = 7.3 Hz, 2H), 1.65 - 1.61 (m, 2H), 1.58 - 1.47 (m, 2H), 1.42 – 1.30 (m, 4H), 1.00 (t, J = 7.6 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CD₃OD): δ 176.0, 138.8, 135.6, 134.6, 134.5, 133.4, 130.4, 130.3, 129.1, 126.6, 126.5, 76.2, 75.9, 73.1, 52.0, 38.2, 37.3, 34.7, 30.1, 26.7, 26.2, 26.0, 14.2; HRMS

(ESI): Exact mass calculated for $C_{23}H_{36}NaO_5$: 415.2455, found 415.2456; UV (MeOH): λ_{max} 288, 301, 315 nm (log ϵ = 7.02).

RvD2n-3 DPA (1)



Following a literature procedure,⁹ a solution of $RvD2_{n-3 DPA}$ methyl ester (2, 5.0 µg, 13 nmol) in MeOH was concentrated under a gentle stream of nitrogen gas, dissolved in THF (500 µL), and cooled to -78 °C. To the resulting solution was added 1 M LiOH (50 µL, 50 µmol) and distilled water (one drop, ~20 µL) and the reaction mixture was stirred in a 4 °C cold room for 24 h. The reaction mixture was then concentrated under a gentle stream of nitrogen gas and reconstituted with MeOH (500 µL). The chemical yield of the $RvD2_{n-3 DPA}$ free acid **1** was 68% (3.3 µg, 8.9 nmol) post saponification (based on UV–Vis).





Figure S-1. UV spectrum of endogenous $RvD2_{n-3 DPA}(1)$.



Figure S-2. MS/MS fragmentation spectra from biological $RvD2_{n-3}DPA$ (1) and synthetic material of 1. Lipid mediators were extracted using C18 SPE from (A) inflammatory exudates were collected from mice 2 h after inoculation of *E. coli* (10⁵ CFU) via peritoneal lavage (B) human serum was obtained from commercial sources and MS/MS spectra for endogenous $RvD2_{n-3}DPA$ (1), together with those of (C) synthetic 1, were obtained using lipid mediator profiling. Results are from n = 3 mice for A and n = 3 determination for B and C. lons in bold denoted deduced ions.

Lipid Mediator Profiling

Human serum (0.5 mL; Sigma Aldrich) and mouse exudates were extracted and lipid mediators were identified and quantified as described.¹⁰ In brief, after the addition of ice-cold methanol containing d_5 -RvD2, samples were placed at -20 °C (45 min) to allow for protein precipitation. The lipid mediators were extracted using an ExtraHera liquid handling system (Biotage) and solid phase extraction techniques with Isolute C18 500 mg columns (Biotage). Methyl formates were collected, brought to dryness and resuspended in phase (methanol/water, 1:1, vol/vol) for injection on a Shimadzu LC-20AD HPLC and a Shimadzu SIL-20AC autoinjector, paired with a QTrap 6500+ (Sciex). The QTRAP was operated in negative ion mode using a multiple reaction monitoring method coupled with an information dependent acquisition to an Enhanced Product Ion experiment. An Agilent Poroshell 120 EC-C18 column (100 mm × 4.6 mm × 2.7 µm) was kept at 50 °C and mediators eluted using a mobile phase consisting of methanol/water/acetic acid of 20:80:0.01 (vol/vol/vol) that was ramped to 50:50:0.01 (vol/vol/vol) over 0.5 min and then to 80:20:0.01 (vol/vol/vol) form 2 min to 11 min, maintained till 14.5 min and then rapidly ramped to 98:2:0.01 (vol/vol/vol) for the next 0.1 min. This was subsequently maintained at 98:2:0.01 (vol/vol/vol) for 5.4 min, and the flow rate was maintained at 0.5 ml/min and the presence of RvD2_{n-3 DPA}(**1**) was monitored using multiple reaction monitoring with QM set to an *m/z* 377 and Q3 to *m/z* of 143.

MS/MS spectral matching was conducted in accordance with published criteria with the matching of retention time and at least 6 diagnostic ions to that of reference standard, with a minimum of one backbone fragment being identified.¹⁰ Furthermore, we also required a minimum of 70% match between the biological and reference standard spectra as determined by the SCIEX OS library match algorithm.

Evaluations of bone marrow-derived macrophage differentiation

The experiments strictly adhered to UK Home Office regulations (Guidance on the Operation of Animals, Scientific Procedures Act, 1986) and Laboratory Animal Science Association (LASA) Guidelines (Guiding Principles on Good Practice for Animal Welfare and Ethical Review Bodies, 3rd Edition, 2015) and according to protocols detailed in a UK Home Office approved protocol (P998AB295).

E. coli peritonitis

Male C57/Bl6 mice were inoculated with 10^5 CFU of *E. coli* via i.p. injection. After 2 h the peritoneum was lavages with 4 mL of sterile PBS, 2 volumes of ice-cold methanol containing 500 pg of deuterium labelled d₅-RvD2 were added to the lavages and lipid mediators were extracted as detailed below.

Bone marrow-derived macrophage differentiation

Generation of bone marrow-derived macrophages (BMDM). Male C57BL6/J mice were culled using CO₂ and bone marrow (BM) from femurs and tibias flushed using Ca²⁺- and Mg²⁺ free DPBS (Sigma, D8537). BM cells were left to adhere to 10 cm² petri dishes for 45 min at 37 °C and 5% CO₂ and non-adherent cells were washed away. Cells were subsequently cultured using RPMI (Sigma, R8758) complemented with 10% fetal bovine serum (Gibco, 10270106), 1% Penicillin/Streptomycin (Gibco, 15140122) and 20 ng/mL of GM-CSF (Peprotech, 315-03-100UG), for 7 days, with media refreshed every 3 days. BMDM were detached by incubating for 15 min at 37 °C and 5% CO₂ with 5 mM EDTA in Ca²⁺- and Mg²⁺ free DPBS and plated in black 96-well plate (Greiner, 655090) at a number of 50,000 per well and nuclei were stained using 1 µg/mL Hoechst H3342 (Thermofisher, P37165) for 15 min at 37 °C. pHrodo Greenlabelled *S. aureus* (5 µg/well)

or pHrodo Red- labelled zymosan A bioparticles (2.5 µg/well) (Invitrogen, P35367 and P35364) were opsonized by incubating in 20% (v/v) human serum (Sigma, H4522) for 30 min at 37 °C. BMDM were incubated with RvD2_{n-3 DPA} (1) (10 nM to 0.01 nM) for 15 min or vehicle (0.01% EtOH in RPMI) at 37 °C and 5% CO₂, before adding 5 µg/well of opsonized pHrodo Green-labelled *S. aureus* bioparticles. The accumulation of pHrodo Green or pHrodo Red fluorescence, indicative of bioparticle engulfment in phago-lysosomes, was then recorded using a Celldiscoverer 7 high content imaging system (Zeiss) during the subsequent 120 min.

¹H- and ¹³C{¹H} NMR Spectra



Figure S-3. ¹H NMR spectrum of compound 18.



Figure S-4. ¹³C{¹H} NMR spectrum of compound 18.



Figure S-5. ¹H NMR spectrum of compound 19.



Figure S-6. ¹³C{¹H} NMR spectrum of compound 19.



Figure S-7. ¹H NMR spectrum of compound 6.



Figure S-8. ¹³C{¹H} NMR spectrum of compound 6.



Figure S-9. ¹H NMR spectrum of compound 21.



Figure S-10. ¹³C{¹H} NMR spectrum of compound 21.







Figure S-12. ¹³C{¹H} NMR spectrum of compound 22.











Figure S-15. ¹H NMR spectrum of compound 10.







Figure S-17. ¹H NMR spectrum of compound 4.



Figure S-18. ¹³C{¹H} NMR spectrum of compound 4.



Figure S-19. ¹H NMR spectrum of compound 11.



Figure S-20. ¹³C{¹H} NMR spectrum of compound 11.







Figure S-22. ¹³C{¹H} NMR spectrum of compound 12.



Figure S-24. ¹³C{¹H} NMR spectrum of compound (S)-13.







Figure S-27. ¹H NMR spectrum of compound 24.



Figure S-28. ¹³C{¹H} NMR spectrum of compound 24.







Figure S-30. ¹³C{¹H} NMR spectrum of compound 5.





Figure S-33. ¹H NMR spectrum of compound 14.



Figure S-34. ¹³C{¹H} NMR spectrum of compound 14.







Figure S-36. ¹³C{¹H} NMR spectrum of compound 3.







Figure S-38. ¹³C{¹H} NMR spectrum of compound 16.



Figure S-39. ¹H NMR spectrum of compound 17.



Figure S-40. ¹³C{¹H} NMR spectrum of compound 17.



Figure S-42. Expansion of the olefinic region in the ¹H NMR spectrum of RvD2_{n-3 DPA} methyl ester 2.





HPLC Chromatograms

Data File C:\Chem32\2\Data\Naft midland Rac AFR2 2020-09-02 07-54-48.D Sample Name: Naft midland Rac AFR2

Aca Operator · SYSTEM							
Acq. operator . SISIEM							
Sample Operator : SYSTEM							
Acq. Instrument : LC1260-1 Location : P1-A-01							
Injection Date : 9/2/2020 7:55:33 AM							
Inj Volume : 15.000 µl							
Acq. Method : C:\CHEM32\2\METHODS\Krial 10% iPrOH i heksan.M							
Last changed : 9/2/2020 8:14:09 AM by SYSTEM							
(modified after loading)							
Analysis Method : C:\CHEM32\2\METHODS\Krial 10% iPrOH i heksan.M							
Last changed : 9/2/2020 8:17:45 AM by SYSTEM							
(modified after loading)							
Sample Info : AD-H kolonne, 254 nm, 1 mL/min, 1% iPrOH i heksan							
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Additional Info : Peak(s) manually integrated							
VWD1 A, Wavelength=254 nm (Naft midland Rac AFR2 2020-09-02 07-54-48.D)							
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Figure S-45. HPLC chromatogram of racemic 23.

Data File C:\Chem32\2\Data\Naft midland optisk AFR 2020-09-02 08-19-06.D Sample Name: Naft midland optisk AFR

Acq. Operator : SYSTEM								
Sample Operator : SYSTEM								
Acq. Instrument : LCI260-1 Location : PI-A-01								
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Acg. Method : C:\CHEM32\2\METHODS\Krial 10% iPrOH i heksan.M								
Last changed : 9/2/2020 8:17:45 AM by SYSTEM								
(modified after loading)								
Analysis Method : C:\CHEM32\2\METHODS\Krial 10% iPrOH i heksan.M								
Last changed : 9/2/2020 8:43:38 AM by SYSTEM								
(modified after loading) Sample Info : AD-H kolonne, 254 nm, 1 mL/min, 1% iPrOH i beksan								
optisk (S) naft Amalie								
Additional Info : Peak(s) manually integrated								
WUT A, wavelength=254 nm (Natt midland optisk AFR 2020-09-02 08-19-06.D)								
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Figure S-46. HPLC chromatogram of (S)-23.

Data File C:\Chem32\2\Data\RvD2n-3 DPA Me KCN_Zn 2021-01-29 13-32-24.D Sample Name: RvD2n-3 DPA Me KCN/Zn

Acq. Operator	SYSTEM							
Sample Operator	SYSTEM							
Acq. Instrument : LC1260-1 Location : P1-A-01								
Injection Date	Injection Date : 1/29/2021 1:33:06 PM							
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Analysis Method	C:\CHEM	32\2\METHODS	S\uio-test.M					
Last changed	: 1/29/20	21 1:53:50 F	M by SYSTEM					
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Additional Info	Poak(c)	manually ir	tograted					
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Figure S-47. HPLC chromatogram of RvD2_{n-3 DPA} methyl ester (2).

UV-Vis Chromatograms



Figure S-48. UV-Vis chromatogram of $RvD2_{n-3 DPA}$ methyl ester (2).

HRMS Chromatograms

Analysis Info Sample Name Method	AFR-95 ESI_pos_50	_1500_os.m			Acquisition Date Analysis Name	9/2/2020 9:38:35 D:\Data\maxis20	6 AM 20\16939.d
Acquisition Pa	arameter						
Source Type Focus Scan Begin Scan End	ESI Not active 50 m/z 1500 m/z	lon Pol Set Ca Set En Set Ch Set Co	larity pillary d Plate Offset arging Voltage rona	Positiv 3500 V -500 V e 2000 V 0 nA	e /	Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve Set APCI Heater	0.4 Bar 200 °C 4.0 I/min Waste 0 °C
Intens x10 ⁵ _		435.2	721			+MS, 0	0.0-1.0min #2-57
-		433.2	/21		,	OTBS	3
2.5-						ŌТВS 10	
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1.0-			43	6.2745			
0.5-					437.2720		
0.0 433	434	435	436	,I., ., .,	437 43	438.2731 	
Meas. m/z 435.2721	Ion Formula C28H39O2Si C22H44NaO3Si2 C25H39O6 C24H36N4NaO2 C22H31N10 C23H40NaO6	m/z err [435.2714 435.2721 435.2741 435.2730 435.2728 435.2717	ppm] -1.7 0.0 4.6 2.1 1.5 -0.9				

Figure S-49. HRSM spectrum of compound 10.



Figure S-50. HRSM spectrum of compound 4.

Figure S-51. HRMS spectrum of compound 11.

Figure S-52. HRMS spectrum of compound 12.

Figure S-53. HRMS spectrum of compound (S)-13.

Figure S-54. HRMS spectrum of compound (S)-23.

Figure S-55. HRMS spectrum of compound 24.

Figure S-56. HRMS spectrum of compound 5.

Figure S-57. HRMS spectrum of compound 25.

Figure S-58. HRMS spectrum of compound 14.

Figure S-59. HRMS spectrum of compound 3.

Figure S-61. HRMS spectrum of compound 17.

Figure S-62. HRMS spectrum of RvD2_{n-3 DPA} methyl ester 2.

References

- Tungen, J.; Gerstmann, L.; Vik, A.; De Matteis, R.; Colas, R.; Dalli, J.; Chiang, N.; Serhan, C.; Kalesse, M.; Hansen, T. Resolving Inflammation: Synthesis, Configurational Assignment, and Biological Evaluations of RvD1_{n-3 DPA}. *Chemistry*, **2019**, *25*, 1476.
- Tungen, J. E.; Primdahl, K. G.; Hansen, T. V. The First Total Synthesis of the Lipid Mediator PD2_{n-3 DPA}. J. Nat. Prod., 2020, 83, 2255.
- 3. Takai, K.; Nitta, K.; Utimoto, K. Simple and selective method for RCHO → (E)-RCH=CHX conversion by means of a CHX₃− CrCl₂ system. *J. Am. Chem. Soc.*, **1986**, *108*, 7408.
- 4. Niwayama, S. Highly Efficient Selective Monohydrolysis of Symmetric Diesters. J. Org. Chem., 2000, 65, 5834.
- Rodriguez, A. R.; Spur, B. W. First total syntheses of the pro-resolving lipid mediators 7(S),13(R),20(S)-Resolvin T1 and 7(S),13(R)-Resolvin T4. *Tetrahedron Lett.*, **2020**, *61*, 151473.
- 6. Boer, R. E.; Gimnez-Bastida, J. A.; Boutaud, O.; Jana, S.; Schneider, C.; Sulikowski, G. A. Total Synthesis and Biological Activity of the Arachidonic Acid Metabolite Hemiketal E 2. *Org. Lett.*, **2018**, *20*, 4020.
- 7. Näf, F.; Decorzant, R.; Thommen, W.; Willhalm, B.; Ohloff, G. The Four Isomeric 1,3,5-Undecatrienes. Synthesis and configurational assignment. *Helv. Chim. Acta.*, **1975**, *58*, 1016.
- 8. Li, J.; Leong, M. M.; Stewart A.; Rizzacasa, M. A. Total synthesis of the endogenous inflammation resolving lipid resolvin D2 using a common lynchpin. *Beilstein J. Org. Chem.*, **2013**, *9*, 2762.
- Chang, M.; Rao, M. K.; Reddanna, P.; Li, C. H.; Tu, C.-P. D.; Corey, E. J.; Reddy, C. C. I. Specificity of the glutathione S-transferases in the conversion of leukotriene A4 to leukotriene C4. *Arch. Biochem. Biophys.*, **1987**, 259, 536.
- Palmas, F.; Clarke, J.; Colas, R. A.; Gomez, E. A.; Keogh, A.; Boylan, M.; McEvoy, N.; McElvaney, O. J.; McElvaney, O.; Alalqam, R.; McElvaney, N. G.; Curley, G. F.; Dalli, J. Dysregulated plasma lipid mediator profiles in critically ill COVID-19 patients. *PLoS ONE*, **2021**, *16(8)*, e0256226.