

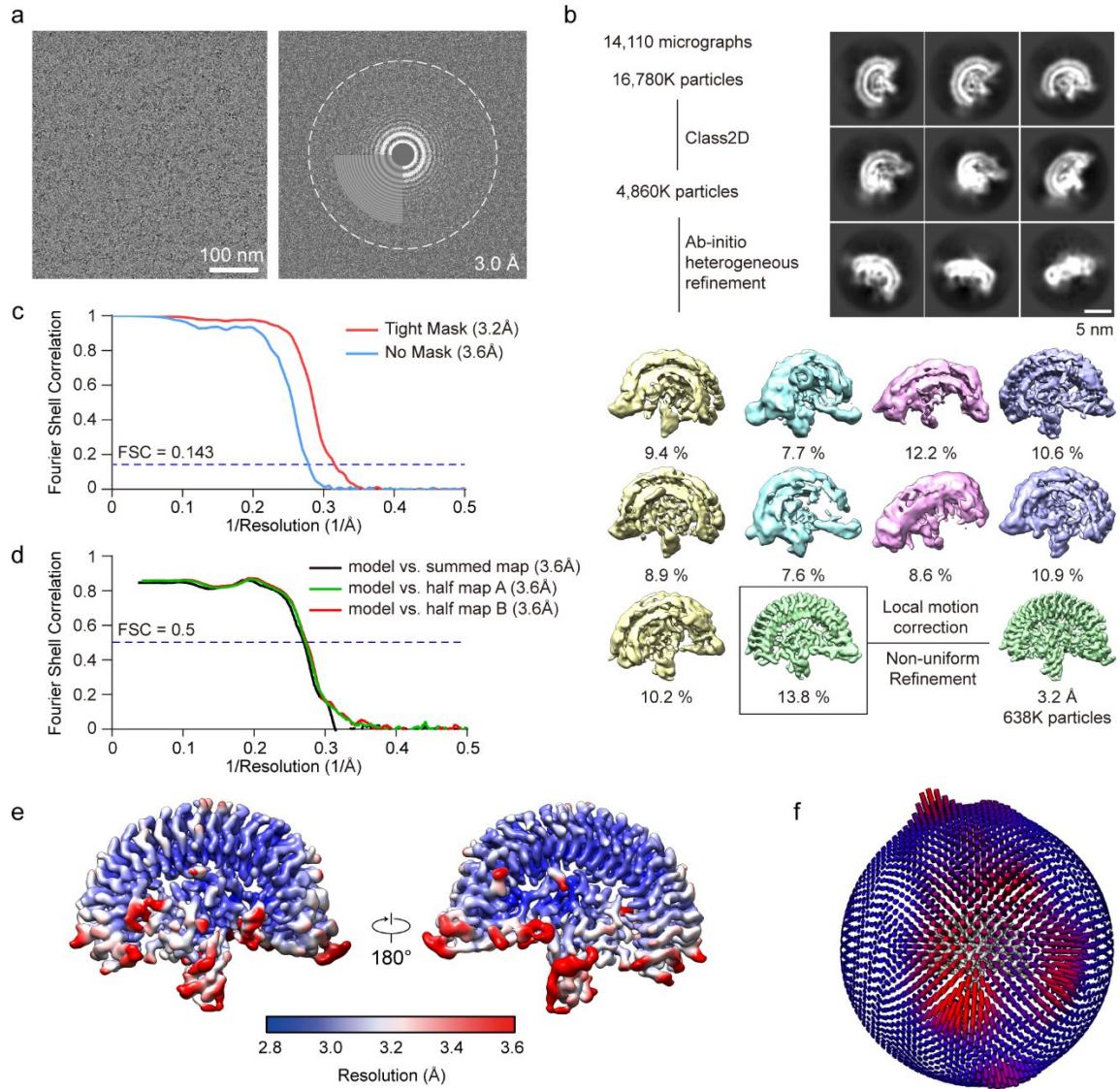
Supplementary Information

Structural basis of assembly and disassembly of the IGF/IGFBP/ALS ternary complex

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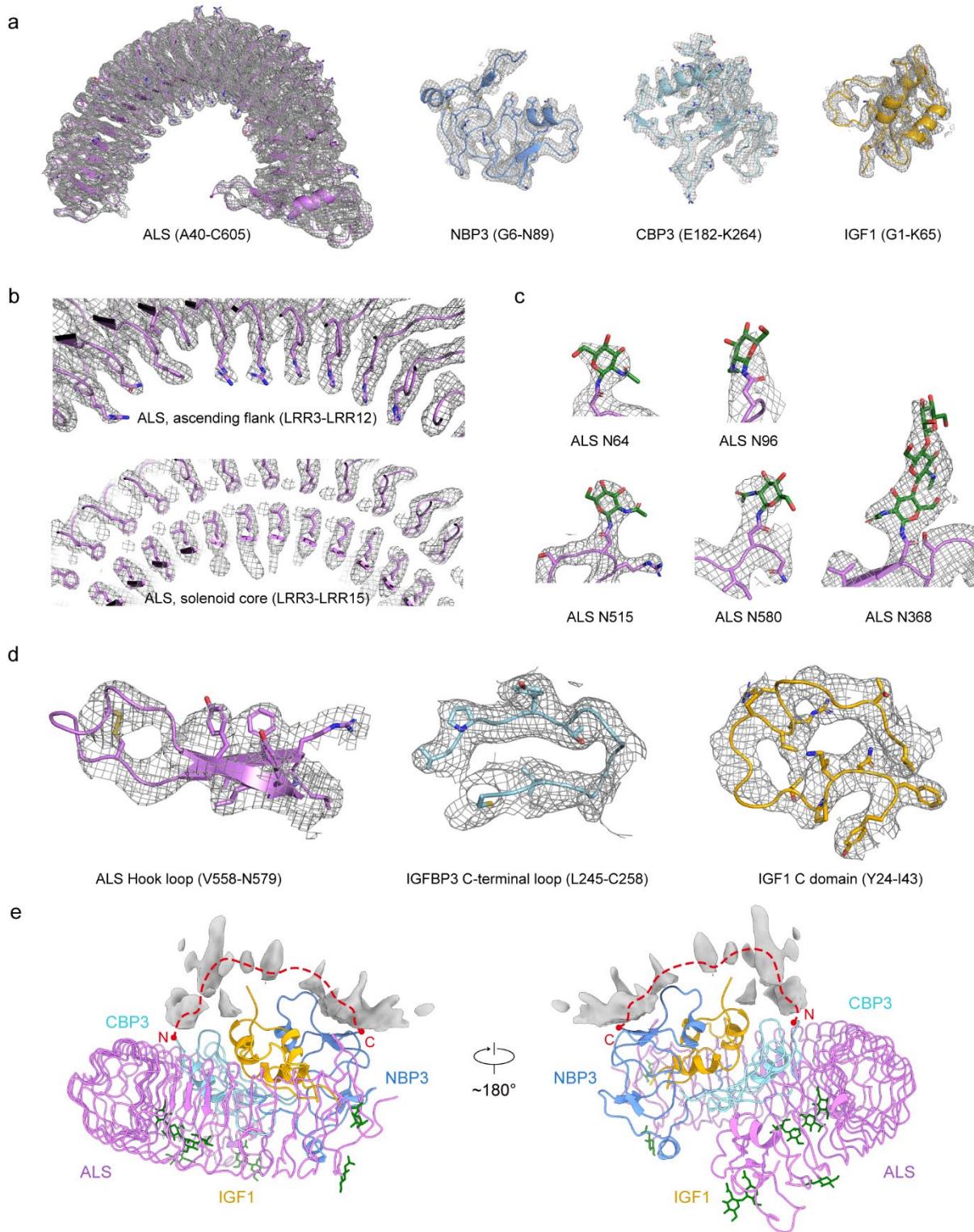
Supplementary Figures 1–7 and Legends

Supplementary Tables 1–6



Supplementary Fig. 1 | Cryo-EM analysis of IGF1/IGFBP3/ALS ternary complex.

- a**, Representative cryo-EM micrograph (left) and its Fourier transform (right) among 14,110 micrographs.
- b**, Data processing workflow of cryo-EM analysis and representative 2D class averages of the IGF1/IGFBP3/ALS ternary complex.
- c**, Gold-standard Fourier shell correlation (FSC) between two independently refined half maps in cryoSPARC (resolution cutoff at FSC = 0.143).
- d**, FSC curves for cross-validation: model versus summed map (black), model versus half map A (used in test refinement, green), model versus half map 2 (not used in test refinement, red).
- e**, Final cryo-EM map colored with local resolution.
- f**, Euler angle distribution of all particles used in the final 3D reconstructions. The height and color (from blue to red) of the cylinder bars are proportional to the number of particles in those views.



Supplementary Fig. 2 | Cryo-EM map for IGF1/IGFBP3/ALS complex.

a, Cryo-EM map and model of ALS (residues 40-605, contour level = 9), NBP3 (residues 6-89, contour level = 11), CBP3 (residues 182-264, contour level = 9), IGF1 (residues 1-65, contour level = 9).

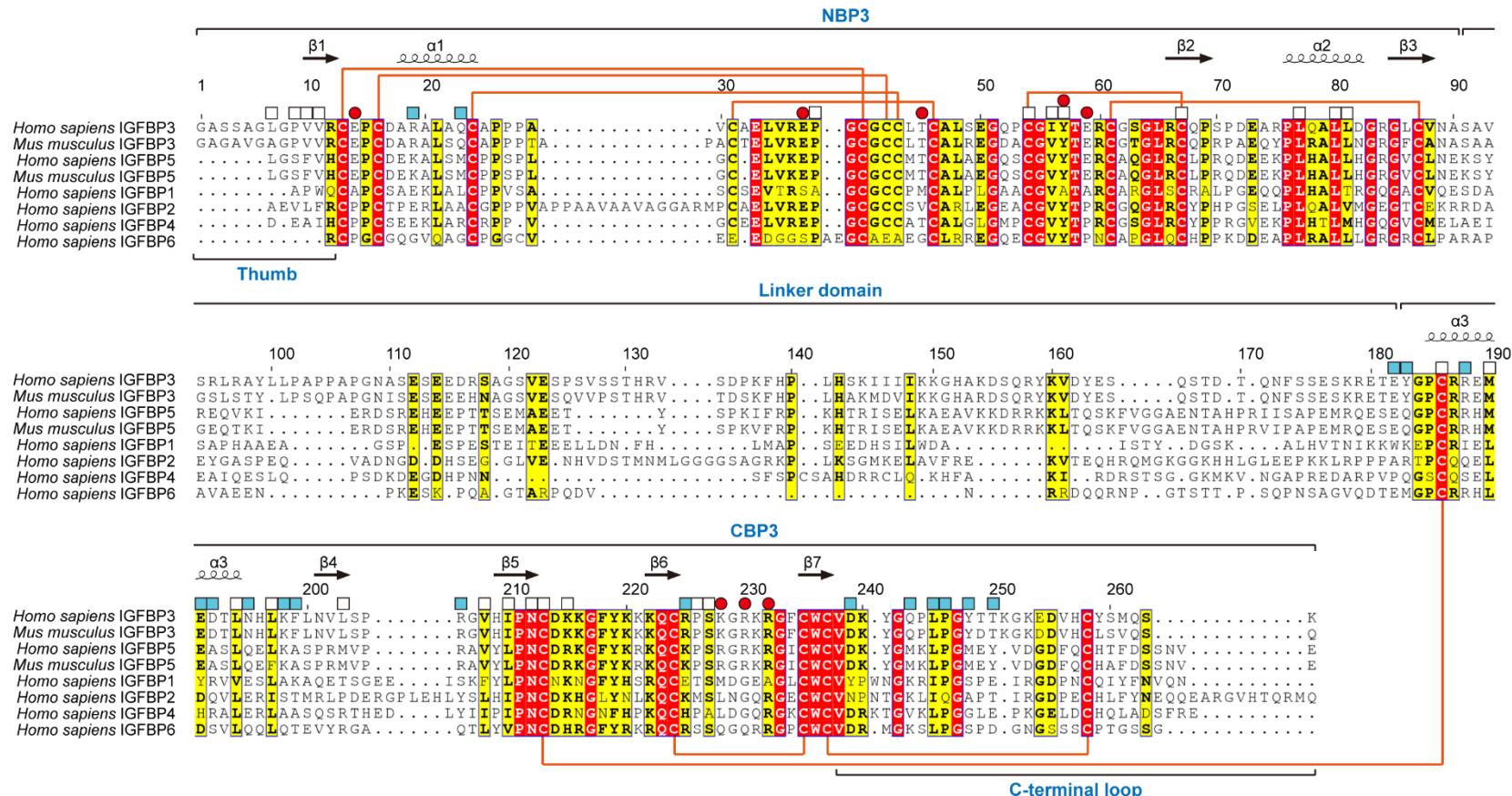
- b**, Cryo-EM map and model of ascending flank (top, LRR3-12) and solenoid core (bottom, LRR3-15) of ALS (contour level = 13).
- c**, Cryo-EM map and model of glycosylation (N64, N96, N368, N580, and N515) of ALS (contour level = 9).
- d**, Cryo-EM map and model of ALS hook loop (residues 558-579, contour level = 8.5), IGFBP3 C-terminal loop (residues 245-258, contour level = 8.5), and IGF1 C domain (residues 24-43, contour level = 8.5).
- e**, Cryo-EM density for central linker domain of IGFBP3 (surface level = 0.088). C-terminal end of NBP3 and N-terminal end of CBP3 are marked with red dots, and the potential path for central linker domain of IGFBP3 is indicated with dotted red line.

a

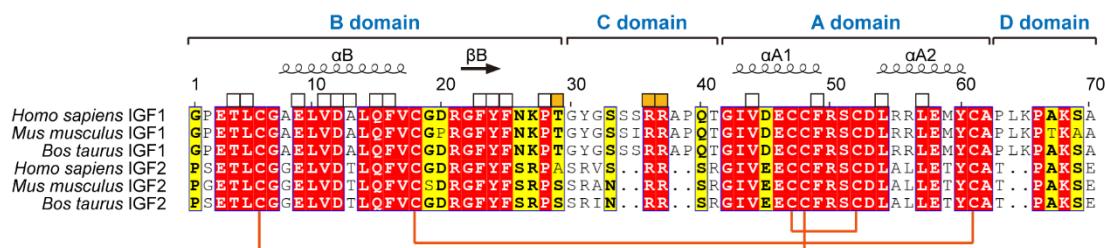


b

● ALS-ALS interaction □ IGFBP3-IGF1 interaction ● NBP3-CBP3 interaction ■ ALS-IGF1 interaction ■ ALS-IGFBP3 interaction ■ N-Glycosylation site — Disulfide Bond



c



Supplementary Fig. 3 | Multiple sequence alignment of ALS, IGFBPs, and IGFs.

- a**, Amino acid sequence alignment of human (*H. sapiens*, UniProt: P35858), rat (*R. norvegicus*, UniProt: P35859), mouse (*M. musculus*, UniProt: P70389), monkey (*P. hamadryas*, UniProt: O02833), and horse (*Equus caballus*, UniProt: F1P8U6) ALS. The LRR consensus sequence of ALS (xLxxLxLxxNxLxxLxxxxFxxLx) is labeled under the LRR motif alignment, and the conserved residues in the corresponding LRR motif are colored in red.
- b**, Amino acid sequence alignment of human IGFBP1-6 (*H. sapiens*, UniProt: P08833, P18065, P17936, P22692, P24593, and P24592) and mouse IGFBP3 and IGFBP 5 (*M. musculus*, UniProt: P47878 and Q07079).
- c**, Amino acid sequence alignment of human, mouse, and bovine IGF1 and IGF2 (*Homo sapiens*, UniProt: P05019 and P01344; *Mus musculus*, UniProt: P05017 and P09535; and *Bos taurus*, UniProt: P07455 and P07456). α helix (A8-V17) and β strand (G22-F25) of IGF1 B domain, α 1 helix (I43-F49) and α 2 helix (L54-Y60) of IGF1 A domain are labeled as α B, β B, α A1, α A2, respectively.
- (a-c)** Domains (LRRNT, LRR1~LRR19 motif, LRRCT, and for ALS; NBP3, linker domain, CBP3 for IGFBP3; B, C, A, and D domain for IGF1) and secondary structure (arrows for β stands and helices for α -helices) elements are noted above the alignment. Additional domain regions (hook loop for ALS; thumb for IGFBP3) are marked under the alignment. Purple circles indicate intramolecular interacting residues in ALS. White squares indicate interacting residues between IGF1 and IGFBP3. Red circles indicate residues for interaction between NBP3 and CBP3. Yellow squares indicate interacting residues at the ALS and IGF1 interface. Cyan squares indicate interacting residues between ALS and IGFBP3. N-glycosylation site (green hexagon) and disulfide bond (orange line) are also indicated. Red boxes indicate perfect sequence conservation, whereas yellow boxes show residues with >70% similarity based on physicochemical properties. The sequence alignment was created using T-Coffee (<http://tcoffee.crg.cat>) and ESPript servers (<http://escript.ibcp.fr>).

a

ALS residues	ALS residues	Interaction
R560	D593	Ionic
R595	D596	Ionic
	N579	Ionic
L531	N579	Hydrogen
Y576	F561	Hydrophobic
Y578		Hydrophobic
P559	V562	Hydrophobic
W532		Hydrophobic

b

BP3 residues (Cleft)	IGF1 residues	Interaction
L77	L57	Hydrophobic
L80	L54	Hydrophobic
L81	L5	Hydrophobic
I56	A13	Hydrophobic
C67	F16	Hydrophobic
C54		Hydrophobic
P38		Hydrophobic
V208		Hydrophobic
P226	V11	Hydrophobic
I210		Hydrophobic
L194	V44	Hydrophobic
M190		Hydrophobic
C186	F49	Hydrophobic
C213		Hydrophobic
N212	E9	Hydrogen
K215	T4	Hydrogen
S227	Q15	Hydrogen
Y57	D12	Hydrogen
S227		Hydrogen
BP3 residues (Thumb)	IGF1 residues	Interaction
P9		Hydrophobic
L197	F25	Hydrophobic
L203		Hydrophobic
V11	F23	Hydrophobic
V208		Hydrophobic
L7	Y24	Hydrophobic
V10	P28	Hydrophobic

c

NBP3 residues	CBP3 residues	Interaction
E37		Ionic
T45	R230	Hydrogen
E59		Ionic
Y57	R232	Hydrophobic
E14	K228	Ionic

d

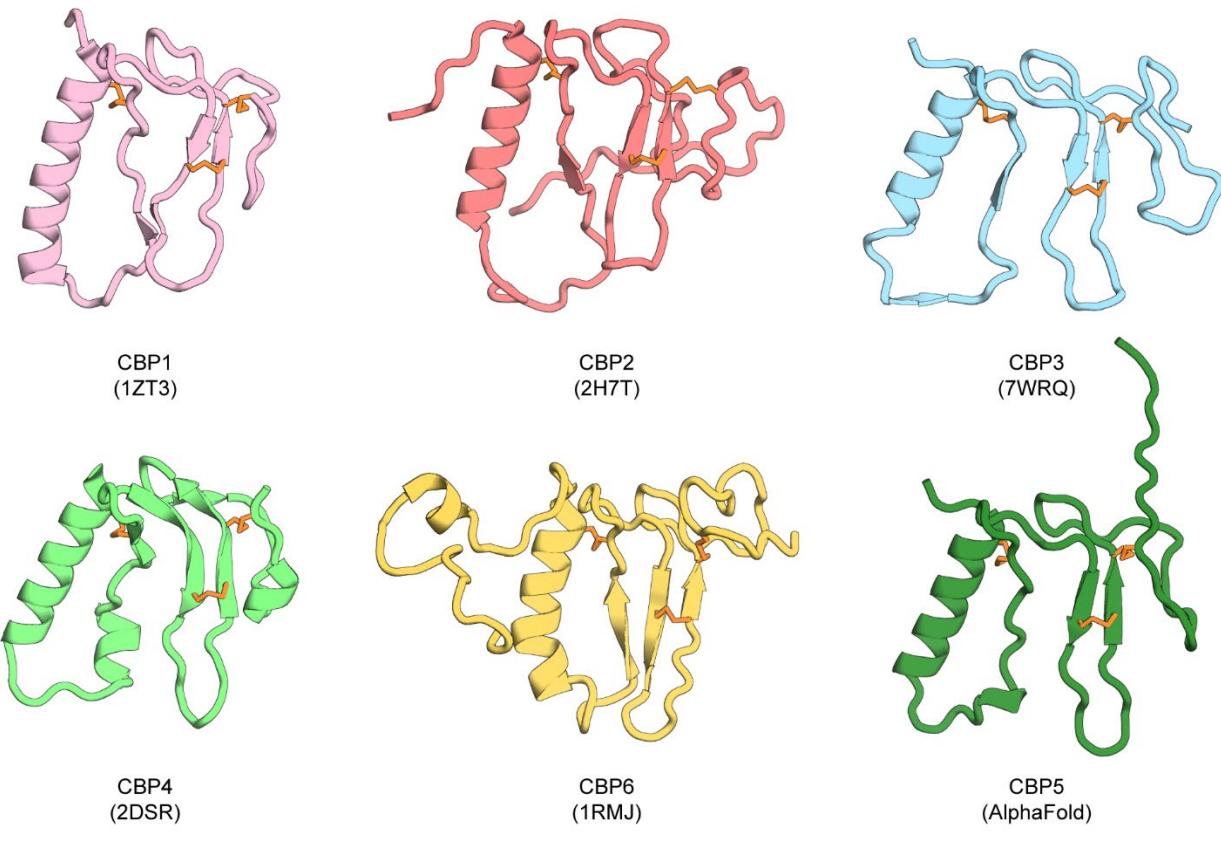
ALS residues	IGF1 residues	Interaction
H126	T29	Hydrogen
S171	R37	Hydrogen
W173		Hydrophobic
		Hydrophobic
D174	R36	Ionic
E198		Ionic

e

ALS residues (Concave)	BP3 residues (CBP3 helix)	Interaction
W179	F199	Hydrophobic
V200		Hydrophobic
E198	K198	Ionic
E222		Ionic
K246	E191	Ionic
Y248	N195	Hydrogen
R269	Y183	Hydrogen
R296	D192	Ionic
	E182	Hydrogen
E317		Ionic
E318	R188	Ionic
E341		Ionic
ALS residues (Concave)	BP3 residues (CBP3 loop)	Interaction
S390		Hydrogen
H392	Q243	Ionic
NAG ₃₆₈		Hydrogen
R414	D239	Ionic
Y462	L245	Hydrophobic
W486	P246	Hydrophobic
Y510	Y248	Hydrophobic
E529		Hydrogen
R530	T249	Hydrogen
ALS residues (Hook loop)	BP3 residues	Interaction
E567	R225	Ionic
D569	R206	Ionic
Q572	R19	Ionic
	Q23	Ionic

Supplementary Fig. 4 | Interactions among IGF1, IGFBP3, and ALS.

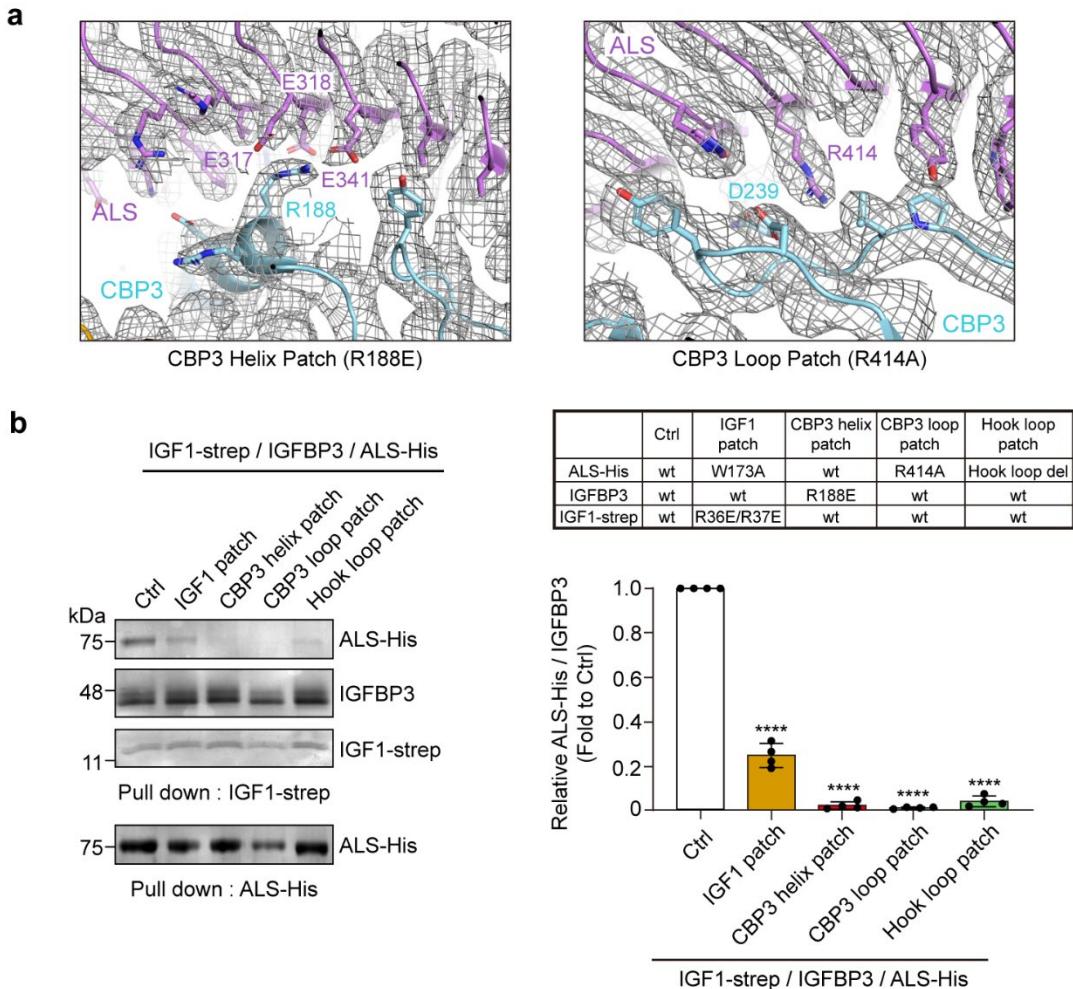
- a**, Key amino acid residues of ALS involved in ALS hook loop stabilization.
 - b**, Key amino acid residues involved in IGFBP3–IGF1 interactions.
 - c**, Key amino acid residues of IGFBP3 involved in NBP3–CBP3 interactions.
 - d**, Key amino acid residues involved in ALS–IGF1 interactions.
 - e**, Key amino acid residues involved in ALS–IGFBP3 interactions.
- (a-e)** Interaction properties are indicated in red (ionic interaction), black (hydrogen bond), and blue (hydrophobic interaction). The residues participating in the main chain interaction are underlined.



Ca rmsd	CBP1 (1ZT3)	CBP2 (2H7T)	CBP4 (2DSR)	CBP6 (1RMJ)
CBP3 (7WRQ)	2.040	3.118	1.132	3.789

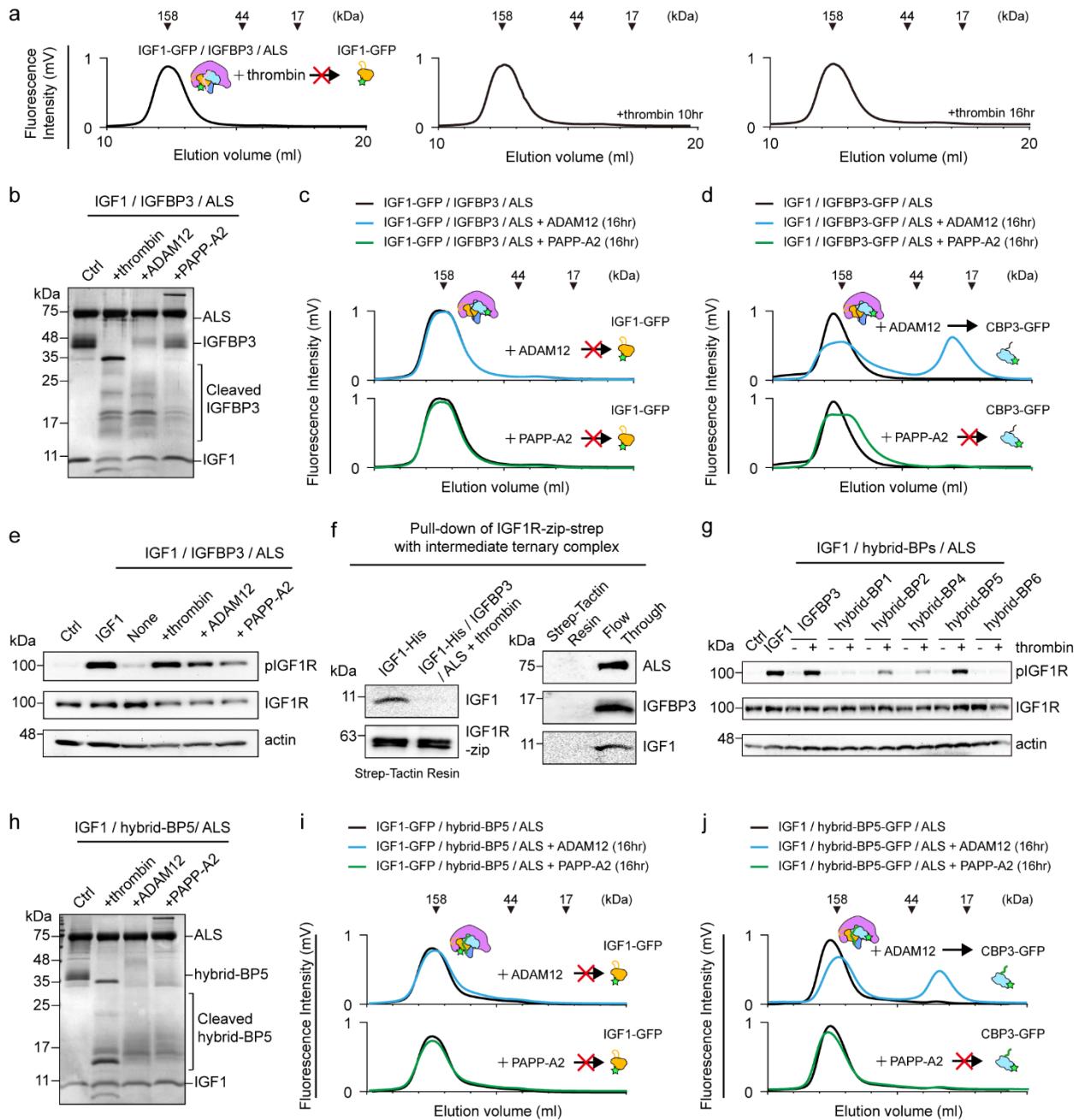
Supplementary Fig. 5 | Structural comparison of IGFBP C-terminal domains.

Structure of each C-terminal domain (CBP) in IGFBP1 (PDB ID: 1ZT3, blue), IGFBP2 (PDB ID: 2H7T, deep salmon), IGFBP3 (PDB ID: 7WRQ, cyan), IGFBP4 (PDB ID: 2DSR, lime), IGFBP6 (PDB ID: 1RMJ, yellow), and IGFBP5 (AlphaFold, forest). CBP1, CBP3, and CBP4 are from the structure of the IGF1/IGFBP complexes, whereas CBP2 and CBP6 are structures of IGFBP only. CBP5 is a structure predicted by AlphaFold (UniProt: P24593). Disulfide bridges (orange) are shown as sticks. Calculated Ca root mean square deviations of each domain are listed in the bottom table.



Supplementary Fig. 6 | Effects on ternary complex formation by mutations of key residues at the interface between ALS and the binary complex

- a**, Cryo-EM map and model of ALS and CBP3 in CBP3 helix patch (left, contour level = 8.5) and CBP3 loop patch (right, contour level = 9). Key residues for mutation and interacting residues are labeled.
- b**, SDS-PAGE for pull-down experiments with Expi 293F culture media co-expressing IGF1-strep, IGFBP3, and ALS-His WT or indicated mutants (left) and quantification of ALS bound to these binary complexes (right, bottom). The mutated key interacting residues at each patch and the construct combination for co-expression are indicated in the table (right, top). The components in the resulting complex were monitored by Coomassie staining after pull-down of IGF1-strep with Strep-Tactin resin. Expression of ALS mutants-His (W173A, R414A, and hook loop deletion) was confirmed by pull-down with Ni-NTA resin. Densitometric analyses of ALS-His/IGFBP3 ratios are shown (right, bottom). Data from four independent experiments ($n = 4$) were analyzed and expressed as mean \pm SD ($****P < 0.0001$ vs. control). P values by one-way ANOVA test followed by Sidak's multiple comparisons test.



Supplementary Fig. 7 | Release of CBP3 from the ternary complex by thrombin and ADAM12 proteolysis, not by PAPP-A2 proteolysis.

a, FSEC of the ternary complex (IGF1-GFP/IGFBP3/ALS-His) after thrombin digestion for the indicated time (0.2 unit thrombin/1 µg ternary complex for 0, 10 and 16h). Fluorescence signal (GFP) was monitored to examine the dissociation of IGF1 from the ternary complex.

b, SDS-PAGE analysis of the IGF1-His/IGFBP3/ALS ternary complex after proteolysis (0.2 unit thrombin/1 µg ternary complex for 10 h; 0.2 µg ADAM12/1 µg ternary complex for 16 h; 0.2 µg

PAPP-A2/1 µg ternary complex for 16 h). IGF1-His/IGFBP3/ALS ternary complex was used for control (Ctrl).

c and d, FSEC of the ternary complex IGF1-GFP/IGFBP3/ALS-His (**c**), or IGF1/IGFBP3-GFP/ALS-His (**d**) after digestion with ADAM12 (Cyan, 0.2 µg ADAM12/1 µg ternary complex for 16 h) or PAPP-A2 (Green, 0.2 µg PAPP-A2/1 µg ternary complex for 16 h). Fluorescence signal (GFP) was monitored to examine the dissociation of IGF1 (**c**) or CBP3 (**d**) from the indicated ternary complex.

e, Immunoblot analysis for IGF1R phosphorylation after treatment of the intermediate ternary complex. HEK293A cells were treated with IGF1-His (5 nM, positive control), IGF1-His /IGFBP3/ALS ternary complex (5 nM, none), or intermediate ternary complex (5 nM). The intermediate ternary complex was prepared by protease digestion (0.2 unit thrombin/1 µg ternary complex for 10 h; 0.2 µg ADAM12/1 µg ternary complex for 16 h; 0.2 µg PAPP-A2/1 µg ternary complex for 16 h) and subsequent SEC purification. No treatment was used for negative control (Ctrl).

f, Pull-down of the IGF1R ectodomain-leucine zipper-Strep (IGF1R-zip-strep) with Strep-Tactin resin after mixing IGF1R-zip-strep with IGF1-His or the intermediate ternary complex (IGF1-His/IGFBP3/ALS ternary complex after thrombin proteolysis). Immunoblot analysis was performed with anti-His and anti-Strep antibody to detect IGF1 bound to IGF1R-zip-strep and IGR1R-zip-strep bound to Strep-Tactin resin, respectively (left). Immunoblot analysis was also performed with anti-ALS, anti-IGFBP3, anti-His antibody to detect ALS, IGFBP3, and IGF-His which are bound to IGF1R-zip-strep or are contained in flow through (right).

g, Immunoblot analysis for IGF1R phosphorylation with intermediate ternary complex containing hybrid-BPs. IGF1-His (5 nM, positive control), IGF1-His/IGFBP3/ALS ternary complexes (5 nM), or IGF1-His/hybrid-BPs/ALS ternary complexes (5 nM) with and without thrombin digestion (0.2 unit thrombin/1 µg ternary complex for 10 h) were treated onto HEK293A cells. No treatment for negative control (Ctrl).

h, SDS-PAGE analysis of the IGF1-His/hybrid-BP5/ALS ternary complex after proteolysis (0.2 unit thrombin/1 µg ternary complex for 10 h; 0.2 µg ADAM12/1 µg ternary complex for 16 h; 0.2 µg PAPP-A2/1 µg ternary complex for 16 h). The IGF1/hybrid-BP5/ALS ternary complex was used for control (Ctrl).

i and j, FSEC of the ternary complex IGF1-GFP/hybrid-BP5/ALS-His (**i**), or IGF1/hybrid-BP5-GFP/ALS-His (**j**) after digestion with ADAM12 (Cyan, 0.2 µg ADAM12/1 µg ternary complex for 16 h) or PAPP-A2 (Green, 0.2 µg PAPP-A2/1 µg ternary complex for 16 h). Fluorescence signal (GFP) was monitored to examine the dissociation of IGF1 (**i**) or CBP3 (**j**) from the indicated ternary complex.

(b, e, f, g and h) Similar results were observed in three independent experiments.

Supplementary Table 1. Cryo-EM data collection, refinement, and validation statistics

Data collection and processing	
Magnification	105,000
Voltage (kV)	300
Electron exposure (e ⁻ /Å ²)	63.2
Defocus range (μm)	-0.9 ~ -2.7
Pixel size (Å)	0.883
Symmetry imposed	C1
Initial particle images (no.)	16,780,000
Final particle images (no.)	638,058
Map resolution (Å)	3.2
FSC threshold	0.143
Refinement	
Initial model used (PDB code)	2DSR
Map resolution (Å)	3.2
FSC threshold	0.143
Map sharpening B factor (Å ²)	0
Model composition	
Nonhydrogen atoms	6,247
Protein residues	798
Ligands	7
B factors (Å ²)	
Protein	130.19
Ligand	113.14
R.m.s. deviations	
Bond lengths (Å)	0.011
Bond angles (°)	1.274
Validation	
MolProbity score	1.55
Clashscore	0.96
Poor rotamers (%)	1.06
Ramachandran plot	
Favored (%)	79.62
Allowed (%)	19.87
Disallowed (%)	0.51

Supplementary Table 2. Acid labile subunit genes for Consurf analysis

Acid labile subunit genes	
Accession numbers	NP_004961.1 (<i>Homo sapiens</i>)
	NP_001351824.1 (<i>Mus musculus</i>)
	XP_038942850.1 (<i>Rattus norvegicus</i>)
	NP_001069431.1 (<i>Bos taurus</i>)
	XP_003198203.1 (<i>Danio rerio</i>)
	XP_001118634.1 (<i>Macaca mulatta</i>)
	XP_002940278.2 (<i>Xenopus tropicalis</i>)
	XP_425222.3 (<i>Gallus gallus</i>)
	XP_016784664.1 (<i>Pan troglodytes</i>)
	XP_038525519.1 (<i>Canis lupus familiaris</i>)
	NP_001041651.1 (<i>Sus scrofa</i>)
	XP_005081648.1 (<i>Mesocricetus auratus</i>)
	XP_004750818.1 (<i>Mustela putorius furo</i>)
	XP_029433221.1 (<i>Rhinatremma bivittatum</i>)
	XP_007906381.1 (<i>Callorhinchus milii</i>)
	XP_027678915.1 (<i>Chelonia mydas</i>)
	XP_034988039.1 (<i>Zootoca vivipara</i>)
	XP_026538289.1 (<i>Notechis scutatus</i>)
	XP_038043074.1 (<i>Anas platyrhynchos</i>)
	XP_035195408.1 (<i>Oxyura jamaicensis</i>)
	XP_021147646.1 (<i>Columba livia</i>)
XP_016081064.1 (<i>Ornithorhynchus anatinus</i>)	
XP_004705854.1 (<i>Echinops telfairi</i>)	
XP_037013030.1 (<i>Artibeus jamaicensis</i>)	
XP_003916392.1 (<i>Papio anubis</i>)	
XP_004057012.2 (<i>Gorilla gorilla gorilla</i>)	
XP_025772696.1 (<i>Puma concolor</i>)	
XP_030728025.1 (<i>Globicephala melas</i>)	
XP_033695959.1 (<i>Tursiops truncatus</i>)	
XP_023472407.1 (<i>Equus caballus</i>)	
XP_010948208.1 (<i>Camelus bactrianus</i>)	
XP_017896378.1 (<i>Capra hircus</i>)	
XP_027817538.1 (<i>Ovis aries</i>)	
XP_017496939.2 (<i>Manis javanica</i>)	
XP_008120350.1 (<i>Anolis carolinensis</i>)	
XP_009491144.1 (<i>Pelecanus crispus</i>)	
XP_032183150.1 (<i>Mustela erminea</i>)	
XP_020862174.1 (<i>Phascolarctos cinereus</i>)	
XP_010565355.1 (<i>Haliaeetus leucocephalus</i>)	
XP_032701926.1 (<i>Lontra canadensis</i>)	
XP_026714531.1 (<i>Athene cunicularia</i>)	
XP_032053301.1 (<i>Aythya fuligula</i>)	

Supplementary Table 3. Insulin-like growth factor-binding protein 3 genes for Consurf analysis

Insulin-like growth factor-binding protein 3 genes	
	NP_000589.2 (<i>Homo sapiens</i>)
	NP_036720.2 (<i>Rattus norvegicus</i>)
	NP_032369.2 (<i>Mus musculus</i>)
	NP_776981.1 (<i>Bos taurus</i>)
	NP_001005156.1 (<i>Sus scrofa</i>)
	XP_003639605.2 (<i>Canis lupus familiaris</i>)
	NP_001152748.1 (<i>Ovis aries</i>)
	NP_001247476.1 (<i>Macaca mulatta</i>)
	XP_519084.2 (<i>Pan troglodytes</i>)
Accession numbers	XP_023494800.1 (<i>Equus caballus</i>)
	XP_027307091.2 (<i>Anas platyrhynchos</i>)
	XP_006051369.2 (<i>Bubalus bubalis</i>)
	XP_008840408.1 (<i>Nannospalax galili</i>)
	XP_011760841.1 (<i>Macaca nemestrina</i>)
	XP_003896056.1 (<i>Papio anubis</i>)
	XP_025234541.1 (<i>Theropithecus gelada</i>)
	XP_009241083.1 (<i>Pongo abelii</i>)
	XP_027430511.1 (<i>Zalophus californianus</i>)
	XP_004283046.1 (<i>Orcinus orca</i>)
	XP_003407178.1 (<i>Loxodonta africana</i>)
	XP_020771079.1 (<i>Odocoileus virginianus texanus</i>)
	NP_001301148.1 (<i>Capra hircus</i>)
	XP_021503913.1 (<i>Meriones unguiculatus</i>)
	XP_004630408.1 (<i>Octodon degus</i>)
	XP_027692998.1 (<i>Vombatus ursinus</i>)
	XP_012618881.1 (<i>Microcebus murinus</i>)
	XP_023373655.1 (<i>Otolemur garnettii</i>)
	XP_029090218.1 (<i>Monodon monoceros</i>)
	XP_020032653.1 (<i>Castor canadensis</i>)
	XP_036859050.1 (<i>Manis javanica</i>)
	XP_026357817.1 (<i>Ursus arctos horribilis</i>)
	XP_025289672.1 (<i>Canis lupus dingo</i>)
	XP_023082252.1 (<i>Piliocolobus tephrosceles</i>)
	XP_017380055.1 (<i>Cebus imitator</i>)
	XP_028633334.1 (<i>Grammomys surdaster</i>)
	XP_032614032.1 (<i>Hylobates moloch</i>)
	XP_010372901.1 (<i>Rhinopithecus roxellana</i>)
	XP_037349128.1 (<i>Talpa occidentalis</i>)
	XP_005319317.1 (<i>Ictidomys tridecemlineatus</i>)
	XP_040500426.1 (<i>Ursus maritimus</i>)
	XP_023970916.1 (<i>Physeter catodon</i>)
	XP_026979864.1 (<i>Lagenorhynchus obliquidens</i>)

Supplementary Table 4. Insulin-like growth factor 1 genes for Consurf analysis

Insulin-like growth factor 1 genes	
	NP_000609.1 (<i>Homo sapiens</i>)
	NP_001104745.1 (<i>Mus musculus</i>)
	NP_001004384.1 (<i>Gallus gallus</i>)
	XP_012015032.1 (<i>Ovis aries</i>)
	XP_005206554.1 (<i>Bos taurus</i>)
	XP_005664255.1 (<i>Sus scrofa</i>)
	NP_001300784.1 (<i>Canis lupus familiaris</i>)
	NP_001075967.2 (<i>Equus caballus</i>)
	XP_519084.2 (<i>Pan troglodytes</i>)
Accession numbers	XP_023494800.1 (<i>Equus caballus</i>)
	NP_001247655.1 (<i>Macaca mulatta</i>)
	XP_005022610.1 (<i>Anas platyrhynchos</i>)
	XP_002936875.1 (<i>Xenopus tropicalis</i>)
	XP_006058875.1 (<i>Bubalus bubalis</i>)
	XP_005303753.1 (<i>Chrysemys picta bellii</i>)
	XP_003907099.1 (<i>Papio anubis</i>)
	XP_005482778.1 (<i>Zonotrichia albicollis</i>)
	XP_022455544.1 (<i>Delphinapterus leucas</i>)
	XP_003989226.1 (<i>Felis catus</i>)
	XP_005500337.1 (<i>Columba livia</i>)
	XP_006186104.1 (<i>Camelus ferus</i>)
	XP_005421104.1 (<i>Geospiza fortis</i>)
	XP_006141463.1 (<i>Tupaia chinensis</i>)
	XP_029455549.1 (<i>Rhinatremabivittatum</i>)
	XP_007072330.1 (<i>Chelonia mydas</i>)
	XP_005235639.1 (<i>Falco peregrinus</i>)
	XP_021243075.1 (<i>Numida meleagris</i>)
	XP_009092444.1 (<i>Serinus canaria</i>)
	XP_015483546.1 (<i>Parus major</i>)
	XP_009647544.1 (<i>Egretta garzetta</i>)
	XP_023792657.1 (<i>Cyanistes caeruleus</i>)
	XP_011760043.1 (<i>Macaca nemestrina</i>)
	XP_002915500.1 (<i>Ailuropoda melanoleuca</i>)
	XP_002199849.1 (<i>Taeniopygia guttata</i>)
	XP_007130866.1 (<i>Physeter catodon</i>)
	XP_010598061.1 (<i>Loxodonta africana</i>)
	XP_010988323.1 (<i>Camelus dromedarius</i>)
	XP_017536871.1 (<i>Manis javanica</i>)
	XP_026253508.1 (<i>Urocitellus parryii</i>)
	XP_008142970.1 (<i>Eptesicus fuscus</i>)
	XP_006205960.1 (<i>Vicugna pacos</i>)
	XP_021531233.1 (<i>Aotus nancymaae</i>)
	NP_001273775.1 (<i>Alligator sinensis</i>)
	XP_027536273.1 (<i>Neopelma chrysocephalum</i>)

Supplementary Table 5. Constructs for recombinant protein expression

Construct	Expression vector	Information
IGF1	pcDNA3.1	HindIII-IGF1(M1-A118)-stop-Xhol
IGF1-proA	pcDNA3.1	HindIII-IGF1(M1-A118)-thrombin site-Protein A tag-Xhol
IGF1-His	pcDNA3.1	HindIII-IGF1(M1-A118)-BamHI-6x His tag-XbaI
IGF1-Strep	pcDNA3.1	HindIII-IGF1(M1-A118)-Twin-Strep-tag-Xhol
IGF1-GFP-Strep	pcDNA3.1	HindIII-IGF1(M1-A118)-GGGGS linker-EGFP-Twin-Strep-tag-Xhol
IGF1(R36E/R37E)-Strep	pcDNA3.1	HindIII-IGF1(M1-A118, R36E/R37E)-Twin-Strep-tag-Xhol
IGFBP3	pcDNA3.1	Signal peptide-BamHI-IGFBP3(G28-K291)-stop-Xhol
IGFBP3-Strep	pcDNA3.1	Signal peptide-BamHI-IGFBP3(G28-K291)-Twin-Strep-tag-Xhol
Myc-IGFBP3-Strep	pcDNA3.1	Signal peptide-BamHI-Myc tag-IGFBP3(G28-K291)-NotI-Twin-Strep-tag-Xhol
IGFBP3-GFP-Strep	pcDNA3.1	Signal peptide-BamHI-IGFBP3(G28-K291)-(GGGGS) ₂ linker-EGFP-Twin-Strep-tag-Xhol
CLD3-CBP3-Strep	pcDNA3.1	Signal peptide-BamHI-IGFBP3(S118-K291)-Twin-Strep-tag-Xhol
IGFBP3(R188E)	pcDNA3.1	Signal peptide-BamHI-IGFBP3(G28-K291, R188E)-stop-Xhol
Hybrid-BP1-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD1(S108-W172)-CBP3(Y210-K291)-Twin-Strep-tag-Xhol
Hybrid-BP2-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD2(R135-A223)-CBP3(Y210-K291)-Twin-Strep-tag-Xhol
Hybrid-BP4-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD4(A104-P170)-CBP3(Y210-K291)-Twin-Strep-tag-Xhol
Hybrid-BP5-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD5(K104-E188)-CBP3(Y210-K291)-Twin-Strep-tag-Xhol
Hybrid-BP6-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD6(R108-E159)-CBP3(Y210-K291)-Twin-Strep-tag-Xhol
Hybrid-BP1-GFP-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD1(S108-W172)-CBP3(Y210-K291)-(GGGGS) ₂ linker-EGFP-Twin-Strep-tag-Xhol
Hybrid-BP2-GFP-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD2(R135-A223)-CBP3(Y210-K291)-(GGGGS) ₂ linker-EGFP-Twin-Strep-tag-Xhol
Hybrid-BP4-GFP-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD4(A104-P170)-CBP3(Y210-K291)-(GGGGS) ₂ linker-EGFP-Twin-Strep-tag-Xhol
Hybrid-BP5-GFP-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD5(K104-E188)-CBP3(Y210-K291)-(GGGGS) ₂ linker-EGFP-Twin-Strep-tag-Xhol

Hybrid-BP6- GFP-Strep	pcDNA3.4 TOPO	Signal peptide-BamHI-NBP3(G28-A118)-CLD6(R108-E159)-CBP3(Y210-K291)-(GGGGS) ₂ linker-EGFP-Twin-Strep-tag-Xhol
ALS	pcDNA3.1	Signal peptide-BamHI-ALS(A28-C605)-stop-Xhol
ALS-His	pcDNA3.1	Signal peptide-ALS(A28-C605)-BamHI-6x His tag-XbaI
ALS (W173A)-His	pcDNA3.1	Signal peptide- ALS(A28-C605, W173A)-BamHI-6x His tag-XbaI
ALS (R414A)-His	pcDNA3.1	Signal peptide- ALS(A28-C605, R414A)-BamHI-6x His tag-XbaI
ALS (hook loop del)-His	pcDNA3.1	Signal peptide- ALS(A28-560R, 579N-605C)-BamHI-6x His tag-XbaI
IGF1R-zip	pcDNA3.1	Signal peptide-BamHI-IGF1R ecto(E31-H935)-GCN4 zipper-NotI-Twin-Strep-tag-Xhol

Supplementary Table 6. Primer sequences used for cloning

Primer	Sequence (5'-3')
IGF1-F	TTC TCT CCA CAG AAG CTT ATG GGT AAG ATC AGC AGC CTT CCA A
IGF1-R	GCG GAA CCA GTC TAG ATC AGG CTG ATT TGG CAG GTT TGA
IGF1-proA-R	ACG CGG AAC CAG TCT AGA GGC TGA TTT GGC AGG TTT GA
IGF1-His-F	CCT TTC TCT CCA CAG AAG CTT ATG GGT AAG ATC AGC AGC C
IGF1-His-R	CAG TGA TGA TGA TGA TGA TGG GAT CCG GCT GAT TTG GCA GGT TTG
IGF1-Strep-R	GGA TGG CTC CAA GCG CTT GCG GCC GCG GCT GAT TTG GCA GGT TTG
IGF1-GFP-Strep-F	GCC AAA TCA GCC GGC GGT GGC GGC TCA ATG GTG AGC AAG GGC GAG GAG C
IGF1-GFP-Strep-R1	CTT GCT CAC CAT TGA GCC ACC GCC GGC TGA TTT GGC AGG TTT GAG G
IGF1-GFP-Strep-R2	CAG AGG TTG ATT CTC GAG TTA CTT CTC AAA CTG TGG GTG TGA C
IGF1 (R36E/R37E)-Strep-F1	ACC GGG TAT GGC TCC TCA TCC GAG AGG GCG CCT CAG ACA GGC ATT
IGF1 (R36E/R37E)-Strep-R1	AAT GCC TGT CTG AGG CGC CCT CTC GGA TGA GGA GCC ATA CCC GGT
IGF1 (R36E/R37E)-Strep-F2	GCT CCT CAT CCG AGG CGG CGC CTC AGA CAG
IGF1 (R36E/R37E)-Strep-R2	CTG TCT GAG GCG CCG CCT CGG ATG AGG AGC
IGFBP3-F	CCT TTG CGG CGG ATC CCG GCG CGA GCT CGG CGG GC
IGFBP3-R	GCG GAA CCA GTC TCT ACT TGC TCT GCA TGC TGT AGC AGT G
IGFBP3-Strep-F	CAC AGG ATC TAG TTC AGG TGG ATC CGG CGC GAG CTC GGC GGG CTT G
IGFBP3-Strep-R	GGA TGG CTC CAA GCG CTT GCG GCC TTG CTC TGC ATG CTG
Myc-IGFBP3-Strep-F1	CTC ATC TCA GAA GAG GAT CTG GGC GCG AGC TCG GCG GGC TTG GG
Myc-IGFBP3-Strep-F2	CTC ACA GGA TCT AGT TCA GGT GGA TCC GAA CAA AAA CTC ATC TCA GAA GAG G
Myc-IGFBP3-Strep-R	GGA TGG CTC CAA GCG CTT GCG GCC TTG CTC TGC ATG CTG
IGFBP3-GFP-Strep-F1	GGC GGT TCA GGC GGA GGT GGC TCT ATG GTG AGC AAG GGC GAG GAG C
IGFBP3-GFP-Strep-F2	GCT ACA GCA TGC AGA GCA AGG CGG CCG CAG GTG GAG GCG GTT CAG GCG G
IGFBP3-GFP-Strep-R	TTT GTA ATC CAG AGG TTG ATT CTC GAG TTA CTT CTC AAA CTG TGG GTG
CLD3-CBP3-Strep-F	GGA TCT AGT TCA GGT GGA TCC AGT GCC GTC AGC CGC CTG CG

CLD3-CBP3-Strep-R	GAG GAT GGC TCC AAG CGC TTG CCG CCG CCT TGC TCT GCA TGC TGT AGC
IGFBP3 (R188E)-F	GAC AGA ATA TGG TCC CTG CCG TGA AGA AAT GGA AGA CAC AC
IGFBP3 (R188E)-R	GTG TGT CTT CCA TTT CTT CAC GGC AGG GAC CAT ATT CTG TC
Hybrid-BP1-F	CGC GGG CTC TGC GTC AAC GCT TCT GAC GCC TCC GCT CCC
Hybrid-BP1-R	CCA TTT CTC TAC GGC AGG GAC CAT ACC ATT TTT TGA TGT TGG TGA CAT GG
Hybrid-BP2-F	CGC GGG CTC TGC GTC AAC GCT CCG GAC GCC GAG TAT GGC
Hybrid-BP2-R	CCA TTT CTC TAC GGC AGG GAC CAT AGG CAG GGG GTG GTC GCA GC
Hybrid-BP4-F	CGC GGG CTC TGC GTC AAC GCT GCG GAG ATC GAG GCC ATC C
Hybrid-BP4-R	CCA TTT CTC TAC GGC AGG GAC CAT AGG GCA CAG GCC GGG CAT CCT CCC
Hybrid-BP5-F	CGC GGG CTC TGC GTC AAC GCT AAG AGC TAC CGC GAG CAA GTC AAG ATC G
Hybrid-BP5-R	CCA TTT CTC TAC GGC AGG GAC CAT ACT CAG ACT CCT GTC TCA TCT CAG G
Hybrid-BP6-F	CGC GGG CTC TGC GTC AAC GCT CGC GCG CCT GCT GTT GCA GAG G
Hybrid-BP6-R	CCA TTT CTC TAC GGC AGG GAC CAT ACT CAG TGT CTT GGA CAC CCG C
Hybrid-BP-GFP-Strep-F	GGA GGA CGT GCA CTG CTA CAG CAT GCA GAG CAA GGC GGC CGC
Hybrid-BP-GFP-Strep-R	CCG GTA GGG ATC GAA CCC TTC TCG AGT TAC TTC TCA AAC TGT GGG TGT GAC C
ALS-F	GCT CCC TGC GCG GCC GCA TGA CTG GTT CCG CGT GGT TC
ALS-R	GAA CCA CGC GGA ACC AGT CAT GCG GCC GCG CAG GGA GC
ALS-His-F	TTC TCT CCA CAG AAG CTT ATG GCT CTG AGG AAA GGA GGG CT
ALS-His-R	ACG CGG AAC CAG TCT AGA GCA GGG AGC AAA ATG GGC TTC CGA CAA
ALS (W173A)-His-F	GGT CTC GGC AGC CTC GCG GAT CTC AAT CTC GG
ALS (W173A)-His-R	CCG AGA TTG AGA TCC GCG AGG CTG CCG AGA CC
ALS (R414A)-His-F	GGT CTC CGC GCT CTC TTC CTC AAG GAT AAC GGC CTC GTG GGC A
ALS (R414A)-His-R	GAA GAG AGC GCG GAG ACC AGA GAG ACC GGT GAA GGT GTG
ALS (hook loop del)-His-F1	CCA CTT TGC CTT TCT CTC CAC AGA AGC TTA TGG CTC TGA GGA AAG GAG GGC
ALS (hook loop del)-His-R1	CCC ACG ACC TCG GGA GGG CTG GCA CAG GTG ATG TTG TTG CGG GGC ACA GCA CTG GGG
ALS (hook loop del)-His-F2	CCC CAG TGC TGT GCC CCG CAA CAA CAT CAC CTG TGC C

ALS (hook loop del)-His-R2 GTG ATG ATG ATG ATG ATG GGA TCC GCA GGG AGC AAA ATG GGC

IGF1R-zip-F1 CTC ACA GGA TCT AGT TCA GGT GGA TCC GAA ATC TGC GGG CCA GGC ATC

IGF1R-zip-R1 GAG GAT GGC TCC AAG CGC TTG CGG CCG CAT GGA TGA AGT TTT CAT ATC C

IGF1R-zip-F2 GGC CAA AAC AGG ATA TGA AAA CTT CAT CCA TCG TTC GAT TCC AC

IGF1R-zip-R2 CGA ACT GAG GAT GGC TCC AAG CGC TTG CGG CCG CG
