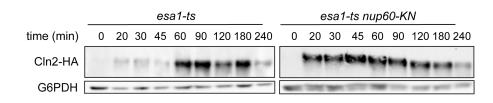
Appendix

Nuclear Pore Complex Acetylation Regulates mRNA Export and Cell Cycle Commitment in Budding Yeast

Mercè Gomar-Alba, Vasilisa Pozharskaia et al.

Table of contents

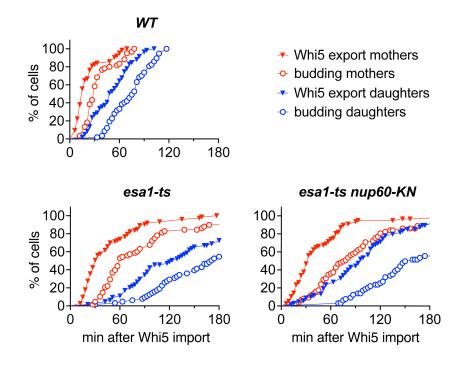
| Appendix Figure S1. | 2 |
|----------------------|----|
| Appendix Figure S2. | 3 |
| Appendix Figure S3. | 4 |
| Appendix Figure S4. | 6 |
| Appendix Figure S5. | 7 |
| Appendix Figure S6. | 8 |
| Appendix Figure S7. | 9 |
| Appendix Figure S8. | 10 |
| Appendix Figure S9. | 11 |
| Appendix Figure S10. | 12 |
| Appendix Figure S11. | 13 |
| Appendix Figure S12. | 15 |
| Appendix Figure S13. | 16 |
| Appendix Table S1. | 17 |



Appendix Figure S1.

nup60-KN mutation partially rescues the delay in synthesis of the G1/S cyclin Cln2 in *esa1-ts* cells.

Independent biological replicates of the experiment shown in Figure 2D. Cells of the indicated strains were arrested in G1 by treatment with a-factor for 2.5 h at 25 °C, shifted to 37 °C for 1 h and released from the G1 arrest at 37 °C. Samples for total protein extracts were collected at the indicated times after a-factor washout and the amount of Cln2-HA protein was assessed by western blot. G6PDH was used as loading control.



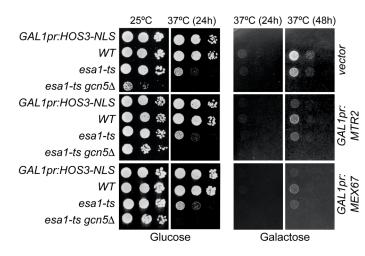
Appendix Figure S2.

Whi5-mCherry nuclear export and budding for cells in Figure 2E.

Whi5-mCherry nuclear export was scored in the fluorescence channel, and budding was scored in bright-field images (maximum projections of 3 z-confocal slices spaced 0.5μ m).

В Α vector SAC3 3860-3905 SAC3*, SSY1, YDR161W, NBP2, CWC15, SEC1, TRM82* W vector SAC3 464-3905 SAC3*, SSY1, YDR161W, NBP2, CWC15, SEC1* HOS3-NLS SAC3 1-855 NUM1*, CTH1, GIR2, ENT5, YDR154C, CPR1, RPA14, YDR157w, SAC3 GAL1pr:HOS3-NLS SUS1 YBR109W-A*, ALG1, YSA1, SUS1, CYC8, YBR113W, RAD16 GAL1pr: SAC3 SEM1 BCP1*, TFC6, ESC2, tKD1, SEM1, CDC40, ESF1, YDR365v HOS3-NLS THP1 MDM20*, YOL075C, YOL073C, THP1, EMI5, YOL070C, NUF2 GAL1pr:HOS3-NLS SEC63*,OSW1,YOR256C,CDC31,HNT3,RPT4,GCD1,RPN8,YOR262W, CDC31 Galactose Glucose YOR263C.DSE3 С GAL1pr:HOS3-NLS WΤ vector vector SAC3 3860-3905 SAC3 464-3905 S SAC3 464-3905 SAC3 1-855 SAC3 1-855 SUS1 vector GAL1pr: HOS3-NLS SEM1 SAC3 3860-3905 THP1 SAC3 464-3905 SAC3 1-855 CDC31 0 Galactose Glucose Glucose Galactose Glucose Galactose

D



Appendix Figure S3.

Overexpression of the TREX-2 complex component Sac3 rescues the toxicity of *HOS3-NLS* overexpression.

(A) Toxicity of full-length *SAC3* overexpression. 10-fold serial dilutions of wild-type (*WT*), *HOS3-NLS-GFP* and *GAL1pr:HOS3-NLS-GFP* cultures transformed with an empty vector or the *GAL1pr:SAC3-HA* plasmid were spotted onto SC-Glu and SC-Gal medium and incubated at 25 °C for 3 days.

(B) List of high-copy (2μ) plasmids from a tiling genome library (Jones et al., 2008) containing TREX-2 complex genes (*SAC3*, *SUS1*, *CDC31*, *SEM1* and *THP1*) together with

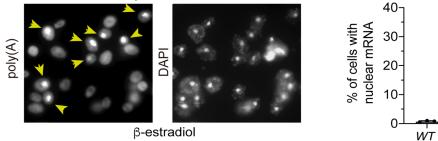
4

neighboring genes. Asterisks (*) indicate that the corresponding ORFs are incomplete. Nucleotides of *SAC3* (full length 3905 nt) included in each plasmid are indicated.

(C) A high-copy plasmid containing SAC3(464-3905) is not toxic and relieves the toxicity of HOS3-NLS over-expression. 10-fold serial dilutions of wild-type (*WT*) and GAL1pr:HOS3-NLS-GFP cultures, transformed with an empty vector or the indicated multicopy plasmids, were spotted onto SC-Glu and SC-Gal medium and incubated at 25 °C for 3 days. Note that SAC3(464-3905) rescues growth of GAL1pr:HOS3-NLS but that the overlapping plasmid SAC3(3860-3905), lacking all of SAC3 ORF but 45 nucleotides at its 3', does not. The "vector" and SAC3(464-3905) sections of the left image are also shown in Figure 3C for simplicity.

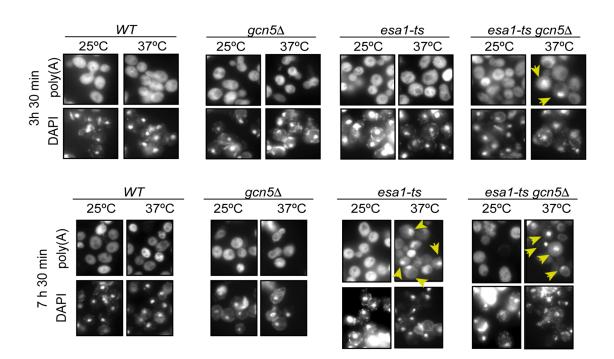
(D) Overexpression of *MEX67* and *MTR2* does not rescue the growth defect of *esa1-ts* and *esa1-ts* $gcn5\Delta$ cells. 10-fold serial dilutions of the indicated strains transformed with the indicated plasmids spotted onto SC-Glu and SC-Gal medium. Due to poor growth at 37 °C on SC-gal, plates were pre-incubated for 24 or 48h at 25 °C and later incubated at indicated temperatures for up to 5 days from plating.

GAL1pr:HOS3-NLS



GAL1pr: HOS3-NLS

В



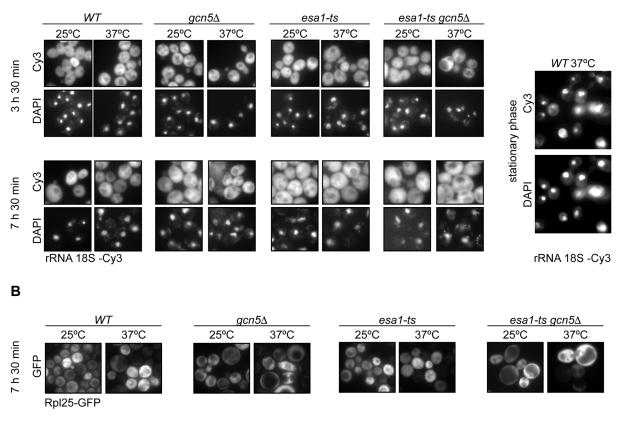
Appendix Figure S4.

Hos3-NLS overexpression or depletion of Esa1 impairs export of poly(A) RNA.

(A) Overexpression of Hos3-NLS promotes nuclear accumulation of mRNA. Cultures of the indicated strains were treated with β-estradiol (90 nM) to induce Hos3-NLS. After induction overnight, cells were fixed and FISH was performed using a Cy3-Oligo(dT) probe. Arrows point to polyadenylated RNA in the nucleus, which was visualized by DAPI staining (left). The fraction of cells with nuclear mRNA accumulation was determined for the indicated strains and conditions (right).

(B) Representative images of cells processed for poly(A) FISH as in (A) after incubation in the indicated conditions. Associated with Figure 3E.



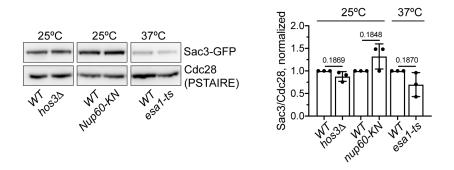


Appendix Figure S5.

Depletion of Esa1 or Gcn5 does not affect export of rRNA.

(A) Wild-type, $gcn5\Delta$, esa1-ts and $gcn5\Delta$ esa1-ts cultures were incubated at 25°C or 37°C at the indicated times. In all cases, cells were fixed and in situ hybridization was performed using Cy3-TXGTTCCTCGTTAAGGXATTTACATTGTACTXCC-Cy3 to target 18S rRNA and monitor ribosomal 40S subunit nucleo/cytoplasmic distribution. DNA was visualized by DAPI staining. Cells from stationary cultures exposed to heat shock during 4h were used as positive control for nuclear accumulation of 18S rRNA.

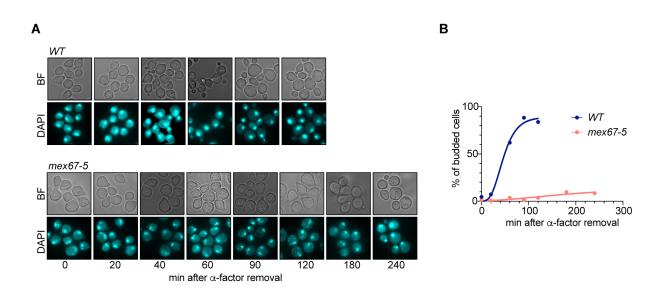
(B) Wild-type, $gcn5\Delta$, esal-ts and $gcn5\Delta$ esal-ts cultures transformed with an Rpl25-GFP plasmid as a reporter for ribosomal 60S subunit nucleo/cytoplasmic distribution were incubated at 25°C or 37°C for 7h and 30 min and imaged at the indicated conditions using fluorescence microscopy.



Appendix Figure S6.

Sac3 protein levels are not significantly affected by *HOS3* deletion, *nup60-KN* mutation and Esa1 inactivation.

Cells of the indicated strains were grown at 25 °C (and shifted to 37 °C for 2 h when indicated), and then collected for total protein extraction. Experiment was repeated three times, the result of one representative western blot is shown (*left*). The amount of Sac3-GFP protein was assessed by western blot, normalized to Cdc28 (PSTAIR) and further normalized to the WT at the corresponding temperature (*right*). Exact *p*-values from one sample t-test are given.

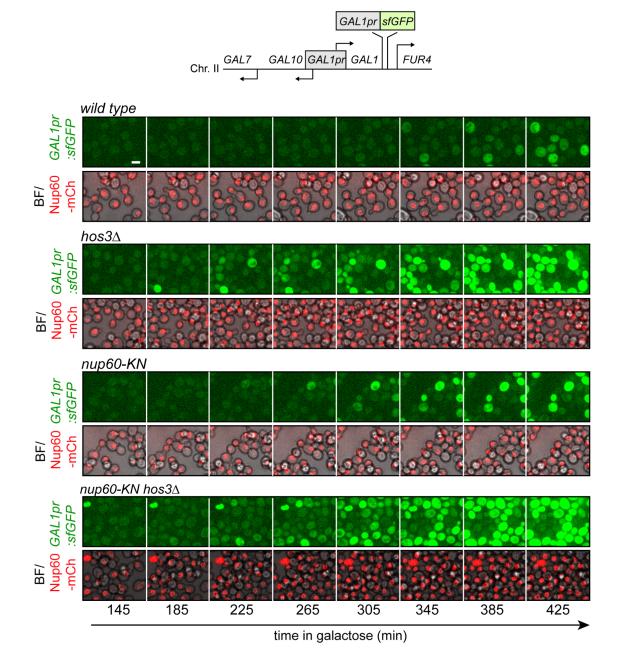


Appendix Figure S7.

Bud emergence defects in the mex67-5 thermosensitive mutant.

(A) Bright field (BF) images of wild type (*WT*) and *mex67-5* cells at the indicated times after the a-factor washout. Cells were arrested in G1 by treatment with a-factor for 2.5 h at 25 °C, shifted to 37 °C for 1 h and released from the G1 arrest at 37 °C. The DNA was visualized by DAPI staining.

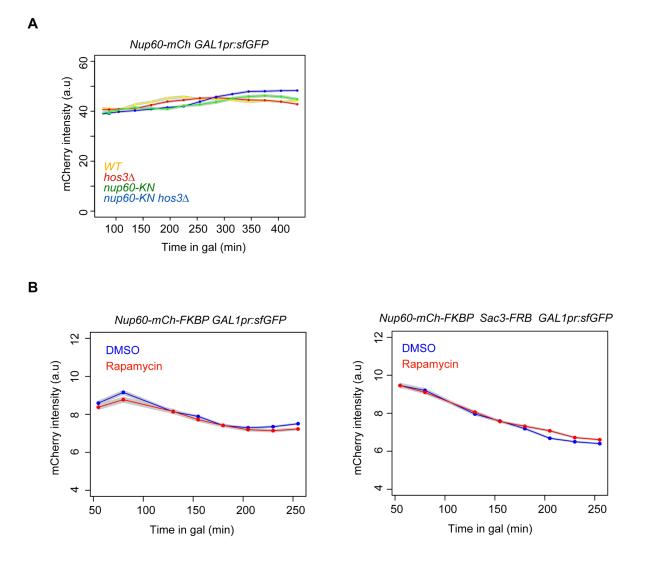
(B) Cells were fixed at the indicated times and the presence of buds was assessed by microscopy. At least 200 cells were scored for each strain and time point.



Appendix Figure S8.

Daughter-cell specific Nup60 deacetylation inhibits GAL1 expression.

(*Top*) The *GAL1pr:sfGFP* reporter was integrated on Chr. II between the *GAL1* and *FUR4* loci. (*Bottom*) Time Lapse microscopy of *WT*, $hos3\Delta$, nup60-KN $hos3\Delta$ and nup60-KN cells expressing *GAL1pr:sfGFP* and Nup60-mCherry at the indicated times of galactose induction. Scale bar, 4 µm.

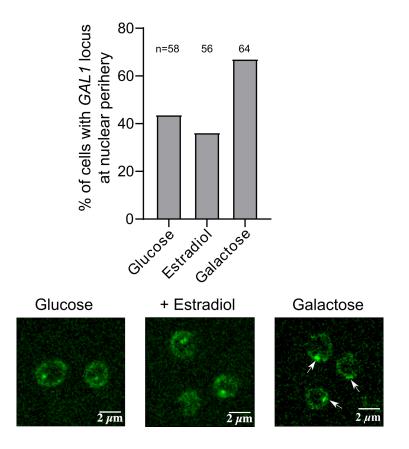


Appendix Figure S9.

Nup60 protein levels upon galactose induction of *GAL1pr:sfGFP* are not changed by the acetyl-mimic allele of Nup60 (*nup60-KN*) or Hos3 and Sac3 anchoring to NPCs.

(A) Cultures of WT, $hos3\Delta$, nup60-KN and nup60-KN $hos3\Delta$ were shifted to galactose and imaged by Time Lapse microscopy to monitor Nup60-mCherry fluorescence during 7 hours of galactose induction of GAL1pr:sfGFP expression. Nuclear fluorescence was scored by segmentation of the nuclear area in the mCherry channel and total fluorescence of Nup60-mCherry was quantified as in Figure 6B. At least 200 cells were scored for each strain and time point. Shaded areas indicate the SEM.

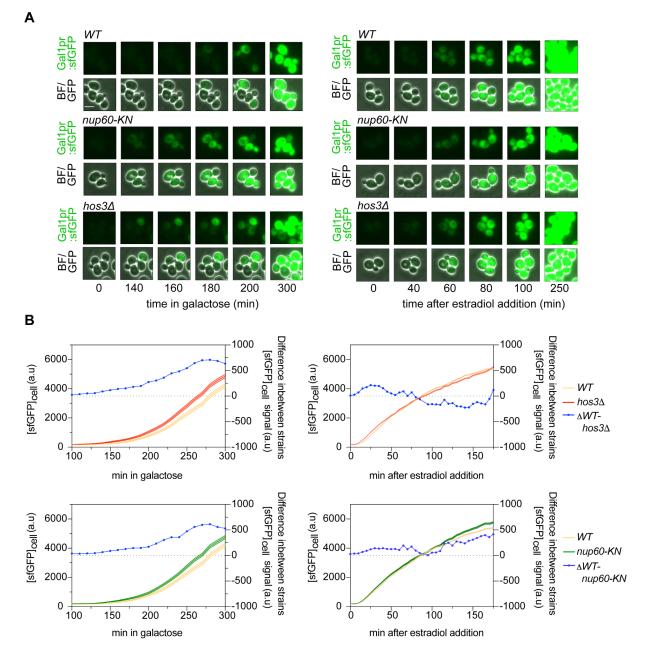
(B) Cells expressing either *Nup60-mCherry-FKBP GAL1pr:sfGFP* or *Nup60-mCherry-FKBP Sac3-FRB GAL1pr:sfGFP* were incubated with rapamycin for FRB-FKBP heterodimerization or DMSO as control as in Figure 7A. Cells were imaged upon rapamycin and galactose addition and the *Nup60-mCherry* fluorescence over time was monitored as in A.



Appendix Figure S10.

The ß-estradiol-dependent GAL4-VP16 transactivator does not increase the perinuclear localization of *GAL1,10* locus.

GAL1 locus localization in cells incubated with 2% glucose (repression), or 30 minutes after addition of 2% galactose or 90 μ M β-estradiol (induction). Localisation was scored by time-lapse microscopy of GAL10::LacO cells expressing LacI-GFP and the β-estradiol-dependent transactivator (GEV). Gene localization was scored as "perinuclear" (arrows) when the nuclear focus was in contact with the nuclear periphery signal (Nup49-GFP). *n* indicates the number of cells scored, which were pooled from two independent experiments.



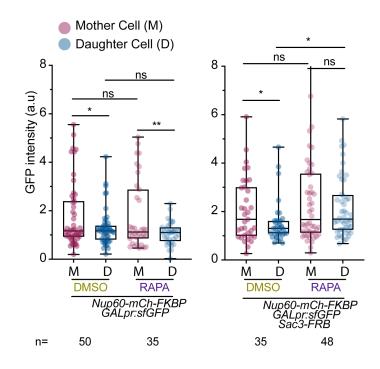
Appendix Figure S11.

Independent biological replicate of the microfluidics time lapse microscopy experiment shown in Figure 5E.

(A) GFP images, and composite of bright field and GFP, from time-lapse microscopy of WT, *nup60-KN* and *hos3* Δ cells expressing *GAL1pr:sfGFP* at the indicated times after addition of galactose (left panels) or β -estradiol (right panels). Scale bar, 4 μ m.

(B) sfGFP expression of WT, hos3A and nup60-KN strain after switching to 2% galactose

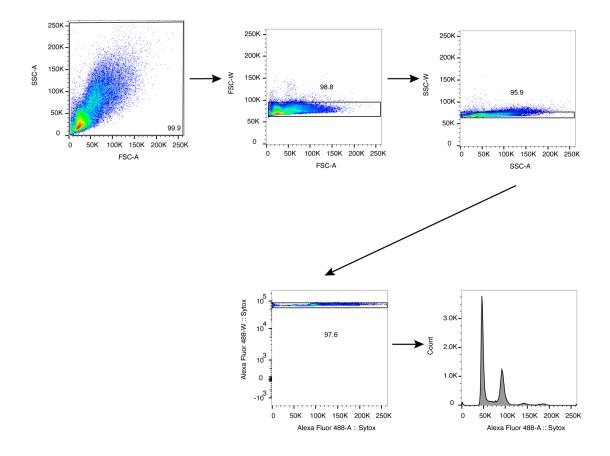
(top) or estradiol (bottom) containing media was monitored. Mean intensity of the sfGFP signal quantified from sum projections for each strain (*WT*, *hos3* Δ and *WT*, *nup60-KN* – left and right panel correspondingly) and the difference of mean intensity in between the strains (Δ WT-*hos3* Δ and Δ WT-*nup60-KN*)) is displayed. At least 450 cells have been quantified for each strain and time point. Shaded areas indicate the SEM.



Appendix Figure S12.

Mother/daughter pairs were quantified as in Figure 7B at 200 min after galactose addition.

Boxes include 50% of data points, the line represents the median and whiskers extend to maximum and minimum values. ***, $p \le 0.001$; **, $p \le 0.01$; *, $p \le 0.05$; ns, p > 0.05, two-tailed paired t-test for M-D comparisons, unpaired for comparisons between strains.



Appendix Figure S13.

Gating FACS strategy used in Figure 1D.

Shown are wild-type cells 240 minutes after the release from alpha factor block.

Appendix Table S1.

Saccharomyces cerevisiae strains used in this work.

| Name | Strain | Genotype | Genetic background | Source |
|---------|--|---|-----------------------|-----------------------|
| YMM1 | wild type (WT) | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 | S288c | |
| YMM5088 | wild type (WT) | MATa his3 leu2 met15 ura3 | BY4741 | |
| YMM5737 | gcn5∆ | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 gcn5∆∷kanMX6 | S288c | This study |
| YMM5671 | esal-ts | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 esa1-L254P::KANMX | S288c | This study |
| YMM5686 | gcn5∆ esa1-ts | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 esa1-L254P::KANMX gcn5∆::kanMX6 | S288c | This study |
| YMM2936 | HOS3-NLS-GFP | <i>MATa ura3-52 his3∆200 leu2 ls2-801 ade2-101 trp1∆63 HOS3-GFP::KAN</i> | S288c | Kumar et al., 2018 |
| YMM3073 | GAL1pr:HOS3-NLS- GFP MYO1-mCherry | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 natNT2::GAL1pr-HOS3-NLS-GFP::KAN MYO1-mCherry::hphNT1 | S288c | Kumar et al., 2018 |
| YMM5121 | GAL1pr:HOS3-NLS- GFP | ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 natNT2::GAL1pr-HOS3-NLS-GFP::KAN | S288c | This study |
| YMM5123 | GAL1pr:HOS3(EN)- NLS-GFP | ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 natNT2::GALpr-hos3-EN(H196E D231A)-NLS-GFP::KAN | S288c | This study |
| YMM3861 | GAL1pr:HOS3-NLS- GFP MYO1-mCherry ADGEV | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 natNT2::GAL1pr-HOS3-NLS-GFP::KAN MYO1-mCherry::hphNT1 ADHpr:GAL4-ER-VP16::URA3 (ADGEV) | S288c | Kumar et al., 2018 |
| YMM5761 | NUP60-GFP | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 NUP60-GFP∷HIS3MX6 | S288c | This study |
| YMM5763 | nup60-KN-GFP | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 nup60(K467N)-GFP::HIS3MX6 | S288c | This study |
| YMM5769 | esa1-ts NUP60-GFP | ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 NUP60-GFP::HIS3MX6 esa1-L254P::kanMX | S288c | This study |
| YMM5771 | esa1-ts nup60-KN-GFP | ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 nup60(K467N)-GFP::HIS3MX6 esa1-L254P::kanMX | S288c | This study |
| YMM5027 | CLN2-HA | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 CLN2-6xHA∷HIS3 | S288c | This study |
| JCY2452 | esa1-ts NUP60-GFP CLN2-HA | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 NUP60-GFP::HIS3MX6 esa1-L254P::kanMX CLN2-6xHA::hphNT1 | S288c | This study |

| JCY2450 | esa1-ts nup60-KN-GFP CLN2-HA | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 nup60(K467N)-GFP::HIS3MX6 esa1-L254P::kanMX CLN2-6xHA::hphNT1 | S288c | This study |
|---------|---|---|--------|------------------------|
| YMM5036 | mex67-5 | MATa leu2Δ1 ura3-52 trp1Δ63 mex67-5::natNT2 | FY86 | Scarcelli et al., 2007 |
| YMM5117 | SAC3-GFP NUP49-3xmCherry | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 SAC3-GFP::KAN NUP49-3xmCherry::hphNT1 | S288c | This study |
| YMM5119 | hos3∆ SAC3-GFP NUP49-3xmCherry | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 SAC3-GFP::KAN NUP49-3xmCherry::hphNT1 hos3A::natNT2 | S288c | This study |
| YMM5351 | SAC3-GFP NUP49-3xmCherry | MATα ura3-52 his3Δ200 leu2 lys2-801 ade2-101 trp1Δ63 SAC3-GFP::TRP NUP49-3xmCherry::hphNT1 | S288c | This study |
| YMM5353 | SAC3-GFP nup60-KN NUP49-3xmCherry | ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 SAC3-GFP::TRP NUP49-3xmCherry::hphNT1 nup60(K467N) | S288c | This study |
| YMM5675 | SAC3-GFP NUP49-3xmCherry esa1-ts | MATα ura3-52 his3Δ200 leu2 lys2-801 ade2-101 trp1Δ63 SAC3-GFP::TRP1 NUP49-3xmCherry::hphNT1 esa1-L254P::kanMX | S288c | This study |
| YMM5549 | GAL1pr:sfGFP-CLN 2PEST nup60-mCherry | MATa his3 leu2 met15 ura3 GAL1pr:sfGFP-CLN2PEST::KAN NUP60-mCherry::hphNT1 | BY4741 | This study |
| YMM5622 | hos3A GAL1pr:sfGFP-CLN 2PEST nup60-mCherry | MATa his3 leu2 met15 ura3 GAL1pr:sfGFP-CLN2PEST::KAN hos3A::natNT2 NUP60-mCherry::hphNT1 | BY4741 | This study |
| YMM5557 | GAL1pr:sfGFP-CLN 2PEST nup60-KN-mCherry | MATa his3 leu2 met15 ura3 GAL1pr:sfGFP-CLN2PEST::KAN nup60(K467N)-mCherry::hphNT1 | BY4741 | This study |
| YMM5721 | hos3A GAL1pr:sfGFP-CLN 2PEST nupP60-KN-mCherr y | MATa his3 leu2 met15 ura3 GAL1pr:sfGFP-CLN2PEST::KAN hos3A::natNT2 nup60(K467N)-mCherry::hphNT1 | BY4741 | This study |
| YMM5653 | nup60-mCherry-FK BP SAC3-FRB-GFP | MATα his3Δ1 leu2Δ0 ura3Δ0 LYS+, Can1::Ste2pr-Leu2, Lyp1::, tor1-1, Fpr1::Ura NUP60-mCherry-FKBP::natNT2 SAC3-FRB-GFP::KAN | BY4742 | This study |
| YMM5637 | 2PEST | MATα his3Δ1 leu2Δ0 ura3Δ0 LYS+, Can1::Ste2pr-Leu2, Lyp1::, tor1-1, Fpr1::Ura NUP60-mCherry-FKBP::natNT2 GAL1pr:sfGFP-CLN2PEST::KAN | BY4742 | This study |
| YMM5657 | 2PEST | MATα his3Δ1 leu2Δ0 ura3Δ0 LYS+, Can1::Ste2pr-Leu2, Lyp1::, tor1-1, Fpr1::Ura NUP60-mCherry-FKBP::natNT2 GAL1pr:sfGFP-CLN2PEST::KAN SAC3-FRB::hphNT1 | BY4742 | This study |

| YMM5844 | SAC3-mCherry-FKB | MATa his $3\Delta 1$ leu $2\Delta 0$ ura $3\Delta 0$ LYS+, | BY4742 | This study |
|-------------|---|---|--------|-------------------------|
| 1 111113644 | P NUP60-FRB WHI5-GFP esal-ts | Can1::Ste2pr-Leu2, Lyp1::, tor1-1, Fpr1::Ura SAC3-mCherry-FKBP::natNT2 NUP60-FRB::hphNT1 esa1-L254P::kanMX WHI5-GFP::HIS3MX6 | D1+/+2 | This study |
| YMM5848 | SAC3-mCherry-FKB P NUP60-FRB WHI5-GFP | MATα his3Δ1 leu2Δ0 ura3Δ0 LYS+, Can1::Ste2pr-Leu2, Lyp1::, tor1-1, Fpr1::Ura SAC3-mCherry-FKBP::natNT2NT2 NUP60-FRB::hphNT1 WHI5-GFP::HIS3MX6 | BY4742 | This study |
| YMM5850 | NUP60-GFP WHI5-mCherry | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 NUP60-GFP::HIS3MX6 WHI5-mCherry::hphNT1 | S288c | This study |
| YMM5854 | NUP60-GFP WH15-mCherry esa1-ts | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 esa1-L254P::kanMX NUP60-GFP::HIS3MX6 WHI5-mCherry::hphNT1 | S288c | This study |
| YMM5860 | nup60-KN-GFP WH15-mCherry esa1-ts | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 nup60(K467N)-GFP::HIS3MX6 WHI5-mCherry::hphNT1 esa1-L254P::kanMX | S288c | This study |
| YMM5773 | gcn5∆ esa1-ts NUP60-GFP | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 NUP60-GFP::HIS3MX6 esa1-L254P::kanMX gcn5A::kanMX6 | S288c | This study |
| YMM5775 | gcn5∆ esa1-ts nup60-KN-GFP | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 nup60-KN-GFP::HIS3MX6 esa1-L254P::kanMX gcn5A::kanMX6 | S288c | This study |
| YMM5019 | CLN2-PP7 NLS-PCP-EGFP WHI5-tdTomato | MATa, ADE2, leu2-3, ura3, trp1-1, his3-11,15, can1-100, psi+, WHI5- tdTOMATO::KAN, URA::NAT::pCYC1-NLS-PCP-GFP-ADH1ter m::ura3 CLN2- 24xPP7SL::loxP | W303 | Neurohr et al., 2018 |
| YMM6020 | CLN2-PP7 NLS-PCP-EGFP WHI5-tdTomato NUP60-6xHA | MATa, ADE2, leu2-3, ura3, trp1-1, his3-11,15, can1-100, psi+, WHI5- tdTOMATO::KAN, URA::NAT::pCYC1-NLS-PCP-GFP-ADH1ter m::ura3 CLN2- 24xPP7SL::loxP NUP60-6xHA::HIS3MX6 | W303 | This study |
| YMM6022 | CLN2-PP7 NLS-PCP-EGFP WH15-tdTomato NUP60-6xHA esa1-ts | MATa, ADE2, leu2-3, ura3, trp1-1, his3-11,15, can1-100, psi+, WHI5- tdTOMATO::KAN, URA::NAT::pCYC1-NLS-PCP-GFP-ADH1ter m::ura3 CLN2- 24xPP7SL::loxP NUP60-6xHA::HIS3MX6 esa1-ts::hphNT1 | W303 | This study |
| YMM6026 | CLN2-PP7 NLS-PCP-EGFP WHI5-tdTomato nup60-KN-6xHA esa1-ts | <i>MATa</i> , <i>ADE2</i> , <i>leu2-3</i> , <i>ura3</i> , <i>trp1-1</i> , <i>his3-11,15</i> , <i>can1-100</i> , <i>psi+</i> , <i>WHI5- tdTOMATO::KAN</i> , <i>URA::NAT::pCYC1-NLS-PCP-GFP-ADH1ter</i> <i>m::ura3 CLN2- 24xPP7SL::loxP</i> <i>nup60(K467N)-6xHA::HIS3MX6</i> <i>esa1-ts::hphNT1</i> | W303 | This study |
| YMM6077 | NUP60-GFP | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 NUP60-GFP::HIS3MX6 | S288c | This study |

| YMM6081 | nup60-KR-GFP | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 nup60(K467R)-GFP::HIS3MX6 | S288c | This study |
|---------|---|---|-------|---------------------|
| YMM6085 | NUP60-GFP esal-ts | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 NUP60-GFP::HIS3MX6 esa1-ts::kanMX | S288c | This study |
| YMM6087 | nup60-KR-GFP esa1-ts | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 nup60(K467R)-GFP::HIS3MX6 esa1-ts::kanMX | S288c | This study |
| YMM6070 | $hatl\Delta$ | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 hat1∆∷natNT2 | S288c | This study |
| YMM6072 | gcn5 Δ hat1 Δ | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 gcn5∆::kanMX6 hat1∆::natNT2 | S288c | This study |
| YMM6070 | esal-ts hat $l\Delta$ | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 hat1∆∷natNT2 esa1-ts∷kanMX | S288c | This study |
| YMM3836 | Gal10-LacO LacI-GFP NUP49-GFP | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 LacI-GFP::HIS Gal10-LacO::TRP NUP49-GFP | W303 | Susan Gasser lab |
| YMM6104 | Gal10-LacO LacI-GFP NUP49-GFP Gal4-ER-VP16 (ADEGV) | MATa ura3-52 his3A200 leu2 lys2-801 ade2-101 trp1A63 LacI-GFP::HIS Gal10-LacO::TRP NUP49-GFP Adh1pr-Gal4-ER-VP16::URA | W303 | This study |
| YMM6101 | Gal4-ER-VP16(ADE GV) | MATα leu2-3,112 trp1-1 can1-100 ura3-1 ade2-1 his3-11,15 Gal4-ER-VP16::ADE | W303 | This study |
| YMM6140 | sfGFP-CLN2PEST Nup60-mCherry Gal4-ER-VP16(ADE GV) | MATα leu2-3,112 trp1-1 can1-100 ura3-1 ade2-1 his3-11,15 GAL1pr:sfGFP-CLN2PEST::KAN Nup60mCherry::hphNT1 Gal4-ER-VP16::ADE | W303 | This study |
| YMM6142 | sfGFP-CLN2PEST Nup60KN-mCherry Gal4-ER-VP16(ADE GV) | MATα leu2-3,112 trp1-1 can1-100 ura3-1 ade2-1 his3-11,15 GAL1pr:sfGFP-CLN2PEST::KAN Nup60(K467N)mCherry::hphNT1 Gal4-ER-VP16::ADE | W303 | This study |
| YMM6144 | sfGFP-CLN2PEST hos3A Nup60-mCherry Gal4-ER-VP16(ADE GV) | MATα leu2-3,112 trp1-1 can1-100 ura3-1 ade2-1 his3-11,15 hos3Δ::natNT2 GAL1pr:sfGFP-CLN2PEST::KAN Nup60mCherry::hphNT1 Gal4-ER-VP16::ADE | W303 | This study |
| YMM6138 | NUP60-GFP gcn5∆ | MATa ura3-52 his3∆200 leu2 lys2-801 ade2-101 trp1∆63 NUP60-GFP::HIS3 gcn5∆::kanMX6 | S288c | This study |