SUPPLEMENTARY INFORMATION

Visible-Light Mediated Catalytic Asymmetric Radical

Deuteration at Non-Benzylic Positions

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1. Supplementary Methods

1.1. General Information

Unless otherwise stated all reactions were set up under nitrogen atmosphere utilizing ovendried glassware. Solvents were either purchased from Adamas or J&K (Sure/Seal bottles) or dried with activated 4 Å molecular sieves and stored in a glovebox under nitrogen atmosphere. Column chromatography was performed using silica gel (200-300 mesh). All other reagents were purchased from various commercial sources and used as received. A blue LED ($\lambda_{max} = 441$ nm, 30 W) was used as the light source for all the photo-reactions. Reactions were monitored by thinlayer chromatography on Leyan silica gel plates (60 GF254) which were rendered visible by ultraviolet light and/or spraying with basic KMnO₄ solution, followed by heating.

¹H NMR spectra were recorded on a Bruker AVIII 400 (400 MHz) spectrometer and are reported in ppm, relative to tetramethylsilane (TMS, $\delta 0$ ppm) or residual solvent signals (CDCl₃) referenced at δ 7.26 ppm, CD₃OD referenced at δ 3.31 ppm). Data are reported as follows: (brs = broad singlet, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet; coupling constant(s) in Hz; integration). ¹³C NMR spectra were recorded on a Bruker AVIII 400 (100 MHz) spectrometer and are reported in ppm, relative to residual solvent signals (CDCl₃ referenced at δ 77.0 ppm, CD₃OD referenced at δ 49.0 ppm). Note: due to quadrupole broadening and spin-spin coupling with boron, resonances of hydrogen atoms bonded to the boron atom are weak (sometimes absent due to partial deuteration) and are thus not integrated in ¹H NMR spectra. In addition, the two carbon atoms attached to the boron atom are not detected in the ¹³C NMR spectra for the same reason. High resolution mass spectra (HRMS) were performed at Instrumental Analysis Center of Shanghai Jiao Tong University with electrospray spectrometer Waters Micromass Q-TOF Premier Mass Spectrometer. Enantiomeric ratios (e.r.) were determined by chiral High Performance Liquid Chromatography (HPLC) analysis. UV detection was monitored at 214 nm at the same time. HPLC samples were dissolved in HPLC grade isopropanol (IPA) and hexane unless otherwise stated. HPLC analysis on chiral stationary phase was performed on a Shimadzu LC-2030 Plus-series instrument. Chiralpak AD-H, OD-H, OJ-H, IC-H, or AS-H columns were used with hexane and IPA being the eluents. Gas chromatographymass spectrometry (GC-MS) analysis were performed on a Shimadzu GCMS-QP2020NX system. Liquid chromatography-mass spectrometry (LC-MS) analysis were performed on a Shimadzu

LCMS 2020 system. Melting points were measured with microscope WRX-4 (Shanghai Yice). Optical rotations were measured on Anton Paar MCP100 automatic polarimeter using a 100 mm path-length cell at 589 nm and reported as follows; $[\alpha]_{\lambda}^{TC}$ (c = g/100 mL, solvent).

2. Supplementary Notes

2.1. Synthesis of Catalysts and Substrates

Note 1: All the thiols used in this study were synthesized from commercially available amino acids/peptides or sugars in 2-4 steps.

Note 2: The cost of D₂O is 2.72 \$/g from Aldrich and 0.63 \$/g from Bide, a domestic chemical supplier. For comparison, the costs of other commonly used deuterated sources from Aldrich are as follows: CD₃OD—13.31 \$/g; d_8 -^{*i*}PrOH—14.93 \$/g; D₂—149.1 \$/L (from Energy Chemical); NaBD₄—140.98 \$/g; LiAlD₄—1000.59 \$/g (prices accessed on June 13, 2022).

The photocatalyst 4CzIPN^1 and 4DPAIPN^2 **S2**, 3 **S3**, 4,5 **S4-S7**, 6,7 **S9-S14**, 6,7 **S1** and **S8**, ${}^{8-10}$ olefins, ${}^{11-15}$ and *N*-heterocyclic carbene (NHC)–BH₃ complexes 16,17 were synthesized according to the reported procedures.



The thiol catalyst **S4-S6** and **S9-S14** were synthesized as follows:^{6,7}



Typical procedure for the synthesis of S9: In a 250 mL round-bottomed flask equipped with a stir bar, I^6 (4.26 g, 8.7 mmol) was treated with HCl (4 N in 1,4-dioxane, 22.5 mL) and the reaction mixture was stirred at room temperature for 1 h. The excess HCl was removed under a stream of nitrogen atmosphere and the reaction mixture was concentrated in vacuo to give a foamy white solid. The residue was dissolved in CH₂Cl₂ (40 mL) and ^{*i*}Pr₂NEt (3.23 mL, 19 mmol) was added. (Boc-Cys-OH)₂ (1.84 g, 4.18 mmol) was added, followed by O-(7-Azabenzotriazol-1-yl)-*N*,*N*,*N'*,*N'*-tetramethyluronium hexafluorophosphate (HATU) (3.99 g, 10.4 mmol), and the reaction mixture was stirred at room temperature for 12 h. The reaction mixture was transferred to a separatory funnel, diluted with EtOAc (200 mL), and washed with aqueous citric acid (2 × 200 mL), saturated aqueous NaHCO₃ (1 × 200 mL) and brine (1 × 200 mL). The organic phases were then dried over Na₂SO₄, filtered, and concentrated in vacuo to afford the crude product **II**, which was used for the next step without further purification.

In a 100 mL round-bottomed flask equipped with a stir bar, **II** and tris(2carboxyethyl)phosphine hydrochloride (TCEP•HCl, 1.8 g, 6.2 mmol) were added and then dissolved in MeOH/H₂O (40 mL, v/v = 4/1). The reaction mixture was stirred at room temperature for 2 h, then the reaction was quenched with H₂O, extracted with CH₂Cl₂ (3 × 20 mL), and dried over Na₂SO₄. After evaporation, the residue was purified by flash column chromatography (PE/EA = 1/2) to give **S9** as a foamy white solid (1.53 g, 43% yield over 2 steps): $[\alpha]_{D}^{25} = -27.2$ (c = 0.36, CHCl₃); **m.p.** 174-176 °C; ¹**H** NMR (400 MHz, CDCl₃) $\delta = 7.83$ (d, J = 8.0 Hz, 1 H), 7.47-7.36 (m, 2 H), 7.35-7.22 (m, 3 H), 7.16 (d, J = 8.8 Hz, 1 H), 6.94 (brs, 1 H), 5.94 (d, J = 8.0 Hz, 1 H), 4.72-4.46 (m, 1 H), 4.44-4.25 (m, 1 H), 3.80-3.54 (m, 2 H), 3.06-2.85 (m, 7 H), 2.76-2.63 (m, 1 H), 2.53-2.40 (m, 1 H), 2.33-1.51 (m, 12 H), 1.42 (s, 9 H); ¹³C **NMR** (100 MHz, CDCl₃) $\delta = 172.7$, 170.8, 170.1, 169.7, 155.6, 137.5, 128.6, 127.8, 127.7, 79.6, 66.9, 66.7, 55.0, 53.4, 47.3, 39.6, 37.0, 36.1, 35.3, 28.2, 27.8, 25.9, 25.2, 24.7, 24.3; **HRMS** (ESI) calcd for C₂₉H₄₄N₅O₆S [M+H⁺]: 590.3007, found: 590.3014.

S4-S6 and S10-S14 were prepared similarly according to the above procedures.

S4, (*R*)-*tert*-butyl-2-(dimethylcarbamoyl)pyrrolidine-1-carboxylate was used instead of **I** following the above procedure. Colorless oil: $[α]_D^{25} = 16.3$ (c = 0.25, CHCl₃); ¹**H** NMR (400 MHz, CDCl₃) $\delta = 5.55$ (d, J = 8.0 Hz, 0.74 H), 5.38 (d, J = 9.6 Hz, 0.16 H), 5.11-5.00 (m, 0.14 H), 4.88-4.78 (m, 0.85 H), 4.70-4.55 (m, 0.74 H), 4.18-4.07 (m, 0.21 H), 3.88-3.57 (m, 2 H), 3.18-3.08 (m, 3 H), 3.04-2.95 (m, 3 H), 2.94-2.84 (m, 0.84 H), 2.84-2.65 (m, 1.20 H), 2.27-2.11 (m, 1.70 H), 2.04-1.87 (m, 2.30 H), 1.58 (t, J = 8.8 Hz, 1 H), 1.44 (s, 9 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 171.1$, 168.1, 155.0, 79.8, 57.3, 56.7, 54.1, 53.9, 47.3, 46.8, 36.9, 36.1, 35.8, 30.7, 28.5, 28.1, 26.7, 26.5, 24.5, 22.3; **HRMS** (ESI) calcd for C₁₅H₂₇N₃NaO₄S [M+Na⁺]: 368.1614, found: 368.1617.

S5, white solid: $[\alpha]_{D}^{25} = 34.6$ (c = 0.21, CHCl₃); **m.p.** 178-181 °C; ¹**H** NMR (400 MHz, CDCl₃) $\delta = 7.74$ (d, J = 8.0 Hz, 1 H), 7.44-7.23 (m, 5 H), 7.11-6.80 (m, 2 H), 5.90 (d, J = 7.6 Hz, 1 H), 4.63-4.46 (m, 1 H), 4.40-4.28 (m, 1 H), 3.75-3.59 (m, 2 H), 3.03-2.85 (m, 7 H), 2.76-2.65 (m, 1 H), 2.28-1.81 (m, 4 H), 1.64 (t, J = 8.8 Hz, 1 H), 1.55 (s, 3 H), 1.43 (s, 9 H), 1.33 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 173.0$, 170.6, 169.9, 169.6, 155.6, 137.3, 128.6, 127.9, 127.6, 79.6, 60.7, 57.0, 55.0, 53.3, 47.3, 36.9, 36.1, 28.2, 27.9, 27.2, 25.8, 25.0, 23.3; **HRMS** (ESI) calcd for C₂₇H₄₂N₅O₆S [M+H⁺]: 564.2850, found: 564.2847.

S6, white solid: $[\alpha]_{D}^{25} = -61.4$ (c = 0.36, CHCl₃); **m.p.** 187-188 °C; ¹H NMR (400 MHz, CDCl₃) $\delta = 7.31-7.17$ (m, 2 H), 6.65 (s, 1 H), 5.03-4.91 (m, 1 H), 4.58-4.47 (m, 1 H), 4.29-4.20 (m, 1 H), 3.76-3.58 (m, 2 H), 3.23-2.85 (m, 8 H), 2.74-2.63 (m, 1 H), 2.25-2.03 (m, 3 H), 1.99-1.86 (m, 1 H), 1.69-1.52 (m, 6 H), 1.51-1.36 (m, 12 H), 1.03-0.86 (m, 6 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 173.5$, 172.0, 170.9, 169.9, 155.7, 79.5, 61.1, 57.2, 55.0, 47.4, 46.7, 41.6, 37.1, 35.9, 28.3, 28.2, 27.8, 25.6, 25.2, 24.5, 23.6, 23.2, 21.9; HRMS (ESI) calcd for C₂₅H₄₅N₅NaO₆S [M+Na⁺]: 566.2983, found: 566.2988.

S10, white solid, $[\alpha]_D^{25} = -62.4$ (c = 0.34, CHCl₃); **m.p.** 158-161 °C; ¹H NMR (400 MHz, CDCl₃) $\delta = 7.55-7.32$ (m, 2 H), 7.31-7.13 (m, 5 H), 6.55 (brs, 1 H), 5.22-5.04 (m, 1 H), 4.62-4.48 (m, 1 H), 4.32-4.15 (m, 1 H), 3.79-3.55 (m, 2 H), 3.18-2.94 (m, 3 H), 2.92-2.81 (m, 6 H), 2.79-2.68 (m, 1 H), 2.27-2.01 (m, 3 H), 2.00-1.81 (m, 1 H), 1.66 (t, J = 8.8 Hz, 1 H), 1.52 (s, 3 H), 1.44 (s, 9 H), 1.24 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 173.2$, 171.0, 170.0, 155.7, 136.9, 129.3, 128.1, 126.5, 79.4, 61.1, 57.2, 55.1, 49.4, 47.4, 38.7, 37.0, 35.8, 28.3, 28.2, 27.6, 25.7, 25.3, 23.5; **HRMS** (ESI) calcd for C₂₈H₄₃N₅NaO₆S [M+Na⁺]: 600.2826, found: 600.2820.

S11, white solid, $[\alpha]_D^{25} = -34.7$ (c = 0.34, CHCl₃); **m.p.** 183-186 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.64$ (d, J = 8.4 Hz, 1 H), 7.51-7.43 (m, 2 H), 7.40-7.25 (m, 3 H), 6.86-6.61 (m, 2 H), 5.82 (d, J = 8.4 Hz, 1 H), 5.56 (d, J = 8.4 Hz, 1 H), 4.59-4.47 (m, 1 H), 4.25-4.14 (m, 1 H), 3.78-3.64 (m, 2 H), 2.83-2.69 (m, 4 H), 2.64-2.54 (m, 1 H), 2.24-2.03 (m, 3 H), 2.02-1.89 (m, 1 H), 1.58 (s, 3 H), 1.54-1.37 (m, 13 H); ¹³C **NMR** (100 MHz, CDCl₃) $\delta = 173.8$, 172.4, 170.9, 169.5, 155.1, 137.3, 128.5, 127.9, 127.8, 80.2, 61.0, 57.6, 57.0, 54.1, 47.8, 28.7, 28.2, 27.3, 26.4, 26.1, 25.1, 23.4; **HRMS** (ESI) calcd for C₂₆H₃₉N₅NaO₆S [M+Na⁺]: 572.2513, found: 572.2511.

S12, white solid, $[\alpha]_D^{25} = 37.9$ (c = 0.40, CHCl₃); **m.p.** 153-156 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.75$ (d, J = 8.0 Hz, 1 H), 7.52-7.42 (m, 2 H), 7.40-7.20 (m, 3 H), 6.92 (brs, 1 H), 6.12 (d, J = 8.4 Hz, 1 H), 5.66 (d, J = 7.6 Hz, 1 H), 4.63-4.41 (m, 1 H), 4.37-3.27 (m, 1 H), 3.81-3.63 (m, 5 H), 2.91-2.79 (m, 1 H), 2.72-2.61 (m, 1 H), 2.23-2.02 (m, 3 H), 2.00-1.85 (m, 1 H), 1.65-1.53 (m, 4 H), 1.48-1.33 (m, 12 H); ¹³C **NMR** (100 MHz, CDCl₃) $\delta = 173.6$, 171.6, 171.0, 169.7, 155.3, 136.5, 128.6, 128.1, 127.3, 79.9, 61.0, 57.1, 56.0, 54.7, 52.6, 47.5, 28.4, 28.2, 26.7, 25.8, 25.0, 24.0; **HRMS** (ESI) calcd for C₂₆H₃₈N₄NaO₇S [M+Na⁺]: 573.2353, found: 573.2348.

S13, white solid, $[\alpha]_{D}^{25} = 3.95$ (c = 0.38, CHCl₃); **m.p.** 204-205 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.75$ (d, J = 8.0 Hz, 1 H), 7.47-7.23 (m, 6 H), 7.03 (brs, 1 H), 5.71 (d, J = 7.6 Hz, 1 H), 4.62-4.48 (m, 1 H), 4.41-4.30 (m, 1 H), 3.76-3.51 (m, 4 H), 3.48-3.33 (m, 1 H), 3.21-2.92 (m, 2 H), 2.77-2.62 (m, 1 H), 2.28-2.13 (m, 2 H), 2.08-1.63 (m, 7 H), 1.56 (s, 3 H), 1.43 (s, 9 H), 1.30 (s, 3 H); ¹³C **NMR** (100 MHz, CDCl₃) $\delta = 172.9$, 170.5, 170.0, 167.9, 155.7, 137.3, 128.6, 127.9, 127.8, 79.5, 60.8, 57.1, 55.1, 54.8, 47.3, 46.3, 46.1, 28.3, 27.9, 27.5, 25.9, 25.8, 25.2, 23.7, 23.2; **HRMS** (ESI) calcd for C₂₉H₄₃N₅NaO₆S [M+Na⁺]: 612.2826, found: 612.2828.

S14, white solid, $[\alpha]_{D}^{25} = 29.7$ (c = 0.36, CHCl₃); **m.p.** 186-187 °C; ¹H NMR (400 MHz, CDCl₃) $\delta = 7.68$ (d, J = 8.0 Hz, 1 H), 7.42-7.34 (m, 3 H), 7.33-7.25 (m, 2 H), 7.22-7.15 (m, 1 H), 5.52-5.38 (m, 1 H), 5.12-4.96 (m, 1 H), 4.47-4.26 (m, 2 H), 4.00-3.86 (m, 1 H), 3.80-3.63 (m, 1

H), 2.91-2.80 (m, 1 H), 2.79-2.67 (m, 1 H), 2.29-2.12 (m, 1 H), 2.08-1.90 (m, 3 H), 1.69-1.60 (m, 2 H), 1.51-1.46 (m, 4 H), 1.43 (s, 9 H), 1.01-0.87 (m, 2 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 172.6, 170.70, 170.66, 155.8, 144.4, 128.2, 126.4, 80.7, 61.5, 54.4, 49.1, 47.8, 34.2, 29.2, 28.2, 25.4, 24.7, 22.5, 16.6; HRMS (ESI) calcd for C₂₅H₃₇N₄O₅S [M+H⁺]: 505.2479, found: 505.2474.$

The thiol catalyst **S8** was synthesized as follows:⁸⁻¹⁰



A stirred solution of compound III⁸ (7.7 g, 10.0 mmol) and thiourea (1.52 g, 20.0 mmol) in acetone (30 mL) was refluxed under N₂ atmosphere for 12 h. The reaction was cooled to room temperature, after which the solvent was removed under reduced pressure to give the isothiouronium salt as colorless foam. To a suspension of this salt in water (24 mL) and dichloromethane (36 mL) was added Na₂S₂O₅ (2.99 g, 15.73 mmol) and the mixture was stirred under reflux for 5 h until all the solid had dissolved. The reaction was cooled to room temperature, the organic layer was separated and the aqueous layer was extracted with dichloromethane. The combined organic phase was dried and concentrated under reduced pressure. The residue was purified by flash column chromatography to give the α,β -anomers IV (5.31 g, 73%) as colorless foam.

The α,β -anomers **IV** (1.0 g) was dissolved in CH₂Cl₂ (5 mL) under nitrogen and SnCl₄ (1.0 M in CH₂Cl₂, 2.5 equiv, 3.5 mL), followed by methane sulfonic acid (1.0 M in CH₂Cl₂, 2.0 equiv, 2.8 mL) was added slowly. The reaction mixture was stirred at 4 °C for 24 h. The reaction was diluted with EtOAc and quenched by addition of 1.0 M KHSO₄. The aqueous layer was extracted with EtOAc. The combined organic phase was washed with saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by flash column chromatography (PE/CH₂Cl₂/Ether = 14/10/1) to give the β -anomer **S8** as colorless foam (0.54 g, 54%); **m.p.** 91-93 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 8.19-8.10 (m, 2 H), 8.03-7.96 (m, 2 H), 7.95-7.88 (m, 2 H), 7.84-7.76 (m, 2 H), 7.47-7.40 (m, 2 H), 7.16-7.08 (m, 2 H), 7.06-6.99 (m, 2 H), 6.98-6.90 (m, 2 H), 6.02-5.92 (m, 1 H), 5.89 (d, *J* = 3.2 Hz, 1 H), 5.61 (dd, *J*₁ = 10.0 Hz, *J*₂ = 3.2 Hz, 1 H), 5.17 (d, *J* = 10.0 Hz, 1 H), 4.74 (dd, *J*₁ = 12.0 Hz, *J*₂ = 2.4 Hz, 1 H), 4.45 (dd, *J*₁ = 12.2 Hz, *J*₂ = 4.2 Hz, 1 H), 4.21-4.10 (m, 1 H), 2.64 (d, *J*= 10.0 Hz, 1 H), 1.38 (s, 9

H); ¹³C NMR (100 MHz, CDCl₃) δ = 166.0 (d, J_{C-F} = 254.0 Hz), 165.9 (d, J_{C-F} = 253.2 Hz), 165.8 (d, J_{C-F} = 252.8 Hz), 165.2, 165.0, 164.6, 164.3, 157.7, 132.39 (d, J_{C-F} = 9.2 Hz), 132.38 (d, J_{C-F} = 9.5 Hz), 132.33 (d, J_{C-F} = 9.4 Hz), 129.8, 126.04 (d, J = 3.0 Hz), 125.99, 125.7, 125.0-124.8 (m), 115.71 (d, J = 21.9 Hz), 115.63 (d, J = 21.9 Hz), 115.55 (d, J = 22.0 Hz), 76.9, 76.6, 73.1, 72.3, 66.1, 62.8, 35.2, 31.0; ¹⁹F NMR (376 MHz, CDCl₃) δ = -104.1, -104.5, -105.1; HRMS (ESI) calcd for C₃₈H₃₃F₃NaO₉S [M+Na⁺]: 745.1690, found: 745.1676.

2.2. Optimization Studies

$ \begin{array}{c} & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & &$										
	1a Fin (2.0 equiv) 2a 3a									
Entry	PC	R*SH	solvent	Yield/% ^[b]	D/% ^[c]	er ^[d]				
1	4DPAIPN	S1	toluene:D ₂ O (3:1)	56	96	48:52				
2	4DPAIPN	S2	toluene:D ₂ O (3:1)	79	90	51:49				
3	4DPAIPN	S 3	toluene:D ₂ O (3:1)	42	90	53:47				
4	4DPAIPN	S4	toluene:D ₂ O (3:1)	50	95	58:42				
5	4DPAIPN	S 5	toluene: $D_2O(3:1)$	67	94	93:7				
6	4DPAIPN	S6	toluene:D ₂ O (3:1)	49	96	93:7				
7	4DPAIPN	S7	toluene:D ₂ O (3:1)	68	94	92:8				
8	4DPAIPN	S10	toluene:D ₂ O (3:1)	78	92	86:14				
9	4DPAIPN	S 5	toluene:D ₂ O (1:1)	43	97	93:7				
10	4DPAIPN	S 5	toluene:D ₂ O (4:1)	71	90	93:7				
11	4DPAIPN	S 5	toluene	73	-	88:12				
12	4CzIPN	S 5	toluene:D ₂ O (3:1)	51	95	93:7				
13 ^[e]	4DPAIPN	S 5	toluene:D ₂ O (3:1)	73	94	93:7				
14 ^[f]	4DPAIPN	S 5	toluene:D ₂ O (3:1)	N.D.	-	-				
15 ^[g]	4DPAIPN	S 5	toluene:D ₂ O (3:1)	N.D.	-	-				
16 ^[h]	4DPAIPN	S 5	toluene: $D_2O(3:1)$	N.D.	-	-				

Supplementary Table 1. Optimization of deuteroboration^[a]

[a] Unless otherwise noted, all reactions were carried with **1a** (0.2 mmol), **2a** (0.1 mmol), PC (1 mol%), R*SH (15 mol%), toluene (0.75 mL), D₂O (0.25 mL) under 10 $^{\circ}$ C for 48 h with irradiation from a 30 W blue LED. The stirring rate for all the reactions is 400 r/min. [b] Isolated yield of **3a**. [c] Determined by ¹H NMR analysis of the isolated product. [d] Determined by chiral HPLC analysis. [e] Reaction time: 72 h. [f] No photocatalyst. [g] Without light irradiation. [h] No thiol catalyst. N.D. = Not detected.

P	'h₃SiH + ∫	∕_0 _	PC (1 R [*] SH	.0 mol%) (15 mol%)	Ph ₃ Si	\geq_0
(2.0	1i) equiv)	∽Ó 2k	toluene:D N ₂ , blue Ll	₂ O (X:1, 2 mL) ED, 10 °C, 72 h	∎ C D 4h)
Entry	PC	R*SH	X	Yield/% ^[b]	D/% ^[c]	er ^[d]
1	4CzIPN	S5	3	80	93	81:19
2	4CzIPN	S7	3	79	91	74:26
3 ^[e]	4CzIPN	S7	3	48	66	80:20
4	4DPAIPN	S 5	3	74	94	82:18
5	4DPAIPN	S 5	2	63	95	83:17
6	4DPAIPN	S5	1	52	98	82:18
7	4DPAIPN	S8	3	70	92	38:62
8	4DPAIPN	S9	3	14	90	78:22
9	4DPAIPN	S10	3	48	91	71:29
10	4DPAIPN	S11	3	67	91	69:31
11	4DPAIPN	S12	3	71	92	66:34
12	4DPAIPN	S13	3	24	90	77:23
13	4DPAIPN	S14	3	52	91	55:45

Supplementary Table 2. Optimization of deuterosilylation using olefin 2k^[a]

[a] Unless otherwise noted, all reactions were carried with **1i** (0.4 mmol), **2k** (0.2 mmol), PC (1 mol%), R*SH (15 mol%), toluene:D₂O (*X*:1, 2 mL) under 10 °C for 72 h with irradiation from a 30 W blue LED. The stirring rate for all the reactions is 400 r/min. [b] Isolated yield of **4h**. [c] Determined by ¹H NMR analysis of the isolated product. [d] Determined by chiral HPLC analysis. [e] Reaction conducted at 0 °C.



Supplementary Figure 1. Preliminary investigations on deuterosilylation reactions of acyclic 1,1-disubstituted olefins.



Supplementary Figure 2. Investigations on the use of recycled D₂O.



Supplementary Figure 3. Preliminary investigations on deuterophosphinoylation and deuterodifluoroalkylation reactions of **2t**.



Supplementary Figure 4. Investigations on the influence of stirring rate.

2.3. Synthesis and Characterization of Products



Typical procedure I: To an oven-dried 16×60 mm vial containing a dry Teflon stir bar were charged with 4DPAIPN (0.8 mg, 0.001 mmol), thiol catalyst S5 (9.0 mg, 0.015 mmol), and NHC-BH₃ 1a (22.2 mg, 0.2 mmol). After sequential addition of dry toluene (0.75 mL), D_2O (0.25 mL), and olefin **2a** (17.5 mg, 0.1 mmol), the reaction mixture was flushed with nitrogen gas for two minutes and then the vial was sealed with a cap and parafilm. The vial was placed in a cooling station and a 30 W blue LED ($\lambda_{max} = 441$ nm) was then placed on the top of the cooling station (Supplementary Figure 5), which is connected to a chiller to maintain the temperature of the cooling water at 10 °C. The reaction mixture was stirred at 10 °C under irradiation with a stirring rate of 400 r/min for 72 h. When the reaction is complete as monitored by TLC and GC, CH₂Cl₂ (10 mL) and H₂O (5 mL) were added, the organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (10 mL \times 3). The combined organic layer was washed with brine and dried over anhydrous Na₂SO₄. After filtration and evaporation, the residue was purified by preparative thin layer chromatography (eluent: petroleum ether/ethtyl acetate = 1/1) to afford **3a** (20.8 mg, 73%, 94% D) as a white solid: 93:7 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 60/40, 0.5 mL/min, λ = 214 nm, t_R (major) = 12.9 min, t_R (minor) = 14.8 min); $[\alpha]_{D}^{25} = -66.8 \ (c = 0.27, \text{ CHCl}_3); \text{ m.p. } 100-102 \ ^{\circ}\text{C}; ^{1}\text{H NMR} \ (400 \text{ MHz}, \text{CDCl}_3) \ \delta = 7.47-7.37 \ (\text{m}, 100-102 \ ^{\circ}\text{C}; ^{1}\text{C})$ 2 H), 7.37-7.28 (m, 2 H), 7.14-7.04 (m, 1 H), 6.76 (s, 2 H), 4.56 (d, J = 8.4 Hz, 1 H), 4.50-4.41 (m, 0.06 H), 4.27 (d, J = 8.4 Hz, 1 H), 3.67 (s, 6 H), 0.93 (d, J = 10.4 Hz, 1 H), 0.45 (d, J = 12.8 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 156.5, 137.6, 128.6, 124.0, 121.7, 121.6, 120.3, 69.2, 59.1 ($J_{C-D} = 21.9 \text{ Hz}$), 35.7; ¹¹**B NMR** (128 MHz, CDCl₃) $\delta = -30.3$ (t, J = 67.5 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



16 mm

A



HRMS (ESI)



Supplementary Figure 5. Reaction setup. (A) 8-mL vial used for the reaction. (B) 30 W blue LED ($\lambda_{max} = 441$ nm) used for the reaction. (C) A custom-made cooling station, connected to a chiller to maintain the reaction temperature at 10 °C. (D) Reaction setup (covered by a cardboard box when the light is on).

The following compounds 3-5 were prepared according to the above Typical Procedure I unless otherwise stated. All the racemic samples for HPLC measurement were also prepared according to this procedure but using Ph₃SiSH or CySH instead of a chiral thiol catalyst.

Synthesis of 3b:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1b** (27.9 mg, 0.2 mmol), **2a** (17.4 mg, 0.1 mmol), **S5** (8.6 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3b** (25.5 mg, 81%, 90% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as a white solid: 92:8 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 60/40, 0.5 mL/min, λ = 214 nm, t_R (major) = 10.4 min, t_R (minor) = 11.4 min); $[\alpha]_D^{25}$ = -72.8 (c = 0.17, CHCl₃); **m.p.** 64-65 °C; ¹H NMR (400 MHz, CDCl₃) δ = 7.49-7.39 (m, 2 H), 7.38-7.28 (m, 2 H), 7.14-7.04 (m, 1 H), 6.91-6.85 (m, 1 H), 6.83-6.76 (m, 1 H), 5.06-4.89 (m, 1 H), 4.57 (d, J = 8.4 Hz, 1 H), 4.47-4.34 (m, 0.1 H), 4.25 (d, J = 8.4 Hz, 1 H), 3.68 (s, 3 H), 1.43-1.29 (m, 6 H), 0.94 (d, J = 12.8 Hz, 1 H), 0.38 (d, J = 12.8 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 156.5, 137.7, 128.6, 124.0, 121.8, 120.8, 114.9, 69.4, 59.4 (t, J_{C-D} = 21.8 Hz), 49.5, 35.4, 23.1, 23.0; ¹¹B NMR (128 MHz, CDCl₃) δ = - 30.6 (m). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), 1c (33.4 mg, 0.2 mmol), 2a (17.4 mg, 0.1

3c, 83%, 94% D

94:6 er

2a

1c (2.0 equiv) mmol), **S5** (9.3 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3c** (28.4 mg, 83%, 94% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as a white solid: 94:6 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 75/25, 0.5 mL/min, λ = 214 nm, t_R (major) = 11.7 min, t_R (minor) = 13.1 min); $[\alpha]_D^{25}$ = -74.4 (c = 0.16, CHCl₃); **m.p.** 147-150 °C; ¹H NMR (400 MHz, CDCl₃) δ = 7.51-7.42 (m, 2 H), 7.39-7.28 (m, 2 H), 7.14-7.04 (m, 1 H), 6.91 (s, 2 H), 5.13-4.89 (m, 2 H), 4.58 (d, J = 8.4 Hz, 1 H), 4.49-4.39 (m, 0.06 H), 4.25 (d, J = 8.4 Hz, 1 H), 1.40-1.35 (m, 12 H), 0.94 (d, J = 12.8 Hz, 1 H), 0.36 (d, J = 12.8 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 156.5, 137.7, 128.6, 123.9, 121.7, 115.4, 69.4, 59.5 (t, J_{C-D} = 21.5 Hz), 49.1, 23.13, 23.11; ¹¹B NMR (128 MHz, CDCl₃) δ = -30.0 (t, J = 63.8 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 3d:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1d** (49.2 mg, 0.2 mmol), **2a** (17.6 mg, 0.1 mmol), **S5** (9.2 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3d** (35.6 mg, 84%, 94% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as an oil: 92:8 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 85/15, 0.5 mL/min, λ = 214 nm, t_R (major) = 16.2 min, t_R (minor) = 20.3 min); $[\alpha]_D^{25}$ = -61.2 (c = 0.10, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.52-7.40 (m, 2 H), 7.37-7.28 (m, 2 H), 7.14-7.02 (m, 1 H), 6.88 (s, 2 H), 4.70-4.51 (m, 3 H), 4.44-4.37 (m, 0.06 H), 4.23 (d, J = 8.4 Hz, 1 H), 2.01-1.68 (m, 10 H), 1.54-1.33 (m, 8 H), 1.32-1.10 (m, 2 H), 0.94 (d, J = 12.8 Hz, 1 H), 0.32 (d, J = 12.4 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ

= 156.5, 137.8, 128.6, 123.9, 121.6, 115.9, 69.6, 56.7, 33.9, 33.8, 25.5, 25.2; ¹¹**B** NMR (128 MHz, CDCl₃) δ = -29.9 (t, *J* = 56.6 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 3e:



The reaction of 4DPAIPN (0.9 mg, 0.001 mmol), **1e** (27.2 mg, 0.2 mmol), **2a** (17.4 mg, 0.1 mmol), **S5** (8.7 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3e** (15.3 mg, 49%, 95% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as an oil: 91:9 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 80/20, 0.5 mL/min, $\lambda = 214$ nm, t_R (major) = 26.1 min, t_R (minor) = 33.7 min); $[\alpha]_D^{25} = -78.5$ (c = 0.19, CHCl₃); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.47$ -7.37 (m, 2 H), 7.37-7.28 (m, 2 H), 7.15-7.03 (m, 1 H), 6.80 (s, 2 H), 5.95-5.73 (m, 1 H), 5.25 (d, J = 10.4 Hz, 1 H), 5.15 (d, J = 16.8 Hz, 1 H), 4.69 (d, J = 6.0 Hz, 2 H), 4.56 (d, J = 8.4 Hz, 1 H), 4.46-4.41 (m, 0.05 H), 4.25 (d, J = 8.4 Hz, 1 H), 3.69 (s, 3 H), 1.05-0.79 (m, 1 H), 0.48-0.33 (m, 1 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 156.5$, 137.7, 132.4, 128.6, 123.9, 121.7, 120.6, 119.1, 118.9, 69.3, 50.8, 35.7; ¹¹B NMR (128 MHz, CDCl₃) $\delta = -30.2$ (t, J = 70.2 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 3f:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1f** (37.1 mg, 0.2 mmol), **2a** (17.5 mg, 0.1 mmol), **S5** (8.9 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3f** (27.0 mg, 78%, 93% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as an oil: 89:11 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 60/40, 1.0 mL/min, λ = 214 nm, t_R (major) = 8.0 min, t_R (minor) = 10.0 min); $[\alpha]_D^{25}$ = -58.0 (c = 0.16, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.48-7.27 (m, 7 H), 7.21-7.03 (m, 3 H), 6.78 (d, J = 2.0 Hz, 1 H), 6.70 (d, J = 1.6 Hz, 1 H), 5.37-5.16 (m, 2 H), 4.54 (d, J = 8.4 Hz, 1 H), 4.46-4.36 (m, 0.07 H), 4.31-4.17 (m, 1 H), 3.70 (s, 3 H), 0.93 (d, J = 12.0 Hz, 1 H), 0.42 (d, J = 12.0 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 156.5, 137.7, 135.6, 128.9, 128.6, 128.3, 127.8, 123.9, 121.7, 120.8, 119.1, 69.3, 59.3 (t, J_{C-D} = 23.5 Hz), 51.9, 35.7; ¹¹B NMR (128 MHz, CDCl₃) δ = -30.1 (t, J = 68.4 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 3g:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1g** (32.5 mg, 0.2 mmol), **2a** (17.4 mg, 0.1 mmol), **S5** (8.5 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3g** (25.2 mg, 75%, 90% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as a white solid: 90:10 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 70/30, 0.5 mL/min, λ = 214 nm, t_R (major) = 18.6 min, t_R (minor) = 20.8 min); $[\alpha]_D^{25}$ = -79.6 (c = 0.11, CHCl₃); **m.p.** 145-147 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.45-7.28 (m, 6 H), 7.25-7.17 (m, 2 H), 7.09-6.94 (m, 1 H), 4.70-4.50 (m, 1.10 H), 4.37 (d, J = 8.4 Hz, 1 H), 3.87 (s, 6 H), 1.07-0.95 (m, 1 H), 0.77-0.59 (m, 1 H); ¹³C **NMR** (100 MHz, CDCl₃) δ = 156.5, 137.5, 133.0, 128.5, 124.2, 123.9, 121.4, 110.7, 68.9, 32.0; ¹¹B NMR (128 MHz, CDCl₃) δ = -30.3 (t, J = 84.8 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 3h:



The reaction of 4DPAIPN (1.1 mg, 0.001 mmol), **1h** (22.2 mg, 0.2 mmol), **2a** (17.4 mg, 0.1 mmol), **S5** (8.6 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3h** (20.9 mg, 73%, 94% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as a white solid: 93:7 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 60/40, 0.5 mL/min, λ = 214 nm, $t_{\rm R}$ (major)

= 16.5 min, $t_{\rm R}$ (minor) = 18.3 min); $[\alpha]_{\rm D}^{25}$ = -71.8 (c = 0.28, CHCl₃); **m.p.** 72-75 °C; ¹H NMR (400 MHz, CDCl₃) δ = 7.83 (s, 1 H), 7.43-7.29 (m, 4 H), 7.15-7.05 (m, 1 H), 4.60-4.45 (m, 1.06 H), 4.30 (d, J = 8.4 Hz, 1 H), 3.87 (s, 3 H), 3.66 (s, 3 H), 1.03-0.80 (m, 1 H), 0.70-0.50 (m, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 156.4, 141.4, 137.5, 128.7, 124.1, 121.4, 68.8, 58.6 (t, $J_{\rm C-D}$ = 21.6 Hz), 38.0, 33.5; ¹¹B NMR (128 MHz, CDCl₃) δ = -30.8 (t, J = 84.1 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of **3i**:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1a** (22.3 mg, 0.2 mmol), **2b** (23.3 mg, 0.1 mmol), **S5** (8.8 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3i** (20.0 mg, 58%, 95% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as a white solid: 89:11 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 85/15, 1.0 mL/min, λ = 214 nm, t_R (minor) = 14.1 min, t_R (major) = 15.2 min); $[\alpha]_D^{25} = -61.4$ (c = 0.36, CHCl₃); **m.p.** 114-117 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.38-7.28 (m, 4 H), 6.76 (s, 2 H), 4.55 (d, J = 8.4 Hz, 1 H), 4.47-4.37 (m, 0.05 H), 4.24 (d, J = 8.4 Hz, 1 H), 3.67 (s, 6 H), 1.29 (s, 9 H), 0.94 (d, J =12.4 Hz, 1 H), 0.45 (d, J = 12.8 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 156.7, 146.9, 134.9, 125.5, 121.4, 120.3, 69.3, 35.7, 34.3, 31.3; ¹¹B NMR (128 MHz, CDCl₃) δ = -30.3 (t, J = 65.9 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of **3j**:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1a** (22.3 mg, 0.2 mmol), **2c** (20.3 mg, 0.1 mmol), **S5** (8.5 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) afforded **3j** (20.4 mg, 65%, 95% D) (eluent: petroleum ether/ethtyl acetate = 1/1) as a white solid: 85:15 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 60/40, 0.5 mL/min, λ = 214 nm, t_R (major) = 11.1 min, t_R (minor) = 16.4 min); $[\alpha]_{D}^{25}$ = -54.5 (c = 0.17, CHCl₃); **m.p.** 104-106 °C; ¹H NMR (400 MHz, CDCl₃) δ = 7.00 (s, 2 H), 6.78-6.68 (m, 3 H), 4.54 (d, J = 8.4 Hz, 1 H), 4.44-4.37 (m, 0.05 H), 4.24 (d, J = 8.4 Hz, 1 H), 3.68 (s, 6 H), 2.29 (s, 6 H), 0.91 (d, J = 12.8 Hz, 1 H), 0.44 (d, J = 12.0 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = -30.3 (t, J = 67.4 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 3k:



The reaction of 4DPAIPN (0.9 mg, 0.001 mmol), **1a** (22.5 mg, 0.2 mmol), **2d** (18.8 mg, 0.1 mmol), **S5** (8.7 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **3k** (18.0 mg, 63%, 90% D) (eluent: CH₂Cl₂/MeOH = 20/1) as an oil: 97:3 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 0.5 mL/min, λ = 214 nm, t_R (major) = 102.0 min, t_R (minor) = 108.8 min); $[\alpha]_D^{25}$ = -74.0 (c = 0.05, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.37-7.24 (m, 2 H), 7.22-7.08 (m, 3 H), 6.71 (s, 2 H), 3.61-3.48 (m, 6.10 H), 2.61-2.38 (m, 2 H), 2.16-1.96 (m, 3 H), 1.84-1.69 (m, 1 H), 0.65-0.36 (m, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.3, 142.8, 128.4, 128.2, 126.0, 120.0, 63.8 (J_{C-D} = 20.7 Hz), 35.6, 33.0, 27.9, 18.1; ¹¹B NMR (128 MHz, CDCl₃) δ = -29.1 (t, J = 66.9 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 31:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1a** (22.7 mg, 0.2 mmol), **2e** (20.5 mg, 0.1 mmol), **S5** (8.9 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **3l** (23.3 mg, 74%, 90% D) (eluent: CH₂Cl₂/MeOH = 20/1) as an oil: 94:6 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 80/20, 1 mL/min, λ = 214 nm, *t*_R (major) = 12.9 min, *t*_R (minor) = 14.3 min); $[\alpha]_{D}^{25}$ = -46.9 (*c* = 0.27, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.16-7.05 (m, 2 H), 7.04-6.94 (m, 2 H), 6.74 (s, 2 H), 3.61-3.53 (m, 6.10 H), 2.58-2.40 (m, 2 H), 2.11-1.93 (m, 3 H), 1.82-1.69 (m, 1 H), 0.59-0.34 (m, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.6, 161.9, 159.5, 138.6, 129.7 (d, *J*_{C-F} = 8.3 Hz), 120.1, 115.2 (d, *J*_{C-F} = 22.2 Hz), 35.6, 33.0, 28.0, 18.1; ¹¹B NMR (128 MHz, CDCl₃) δ = -29.2 (t, *J* = 66.8 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ = -116.6. HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



The reaction of 4DPAIPN (0.9 mg, 0.001 mmol), 1a (22.3 mg, 0.2 mmol), 2f (21.5 mg, 0.1 mmol), S5 (8.9 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **3m** (19.1 mg, 58%, 90% D) (eluent: $CH_2Cl_2/MeOH = 20/1$) as an oil: 96:4 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 70/30, 1 mL/min, λ = 214 nm, $t_{\rm R}$ (major) = 10.7 min, $t_{\rm R}$ (minor) = 15.6 min); $[\alpha]_{\rm D}^{25}$ = -82.1 (c = 0.19, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.10-6.99 (m, 2 H), 6.91-6.80 (m , 2 H), 6.73 (s, 2 H), 3.79 (s, 3 H), 3.69-3.48 (m, 6.10 H), 2.60-2.40 (m, 2 H), 2.10-1.93 (m, 3 H), 1.81-1.67 (m, 1 H), 0.59 (d, J = 12.8 Hz, 1 H), 0.43 (d, J = 13.2 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.6, 157.5, 135.6, 129.1, 120.0, 113.8, 55.4, 35.6, 33.0, 27.9, 18.1; ¹¹B NMR (128 MHz, CDCl₃) δ = -29.1 (t, J = 67.6 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



ÒMe

Synthesis of 3n:



The reaction of 4DPAIPN (0.9 mg, 0.001 mmol), 1a (22.0 mg, 0.2 mmol), 2g (17.4 mg, 0.1 mmol), S5 (8.9 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **3n** (17.6 mg, 62%, 96% D) (eluent: PE/EA = 1/1) as a white solid: 94:6 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 75/25, 0.5 mL/min, λ = 214 nm, $t_{\rm R}$ (minor) = 19.1 min, $t_{\rm R}$ (major) = 21.9 min); $[\alpha]_{\rm D}^{25}$ = -49.6 (c = 0.14, CHCl₃); m.p. 79-81 °C; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3) \delta = 7.44-7.36 \text{ (m, 2 H)}, 7.35-7.28 \text{ (m, 2 H)}, 7.15-7.05 \text{ (m, 1 H)}, 6.75 \text{ (s, 2 H)},$ 4.26-4.17 (m, 0.04 H), 3.66 (s, 6 H), 2.68-2.57 (m, 1 H), 2.54-2.41 (m, 1 H), 2.36-2.25 (m, 1 H), 1.99-1.87 (m, 1 H), 0.81 (d, J = 12.4 Hz, 1 H), 0.34 (d, J = 12.8 Hz, 1 H); ¹³C NMR (100 MHz,

CDCl₃) $\delta = 174.8, 138.7, 128.4, 124.5, 123.7, 120.1, 35.7, 32.2, 25.5; {}^{11}B$ NMR (128 MHz, CDCl₃) $\delta = -29.7$ (t, J = 64.3 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).

HRMS (ESI)



Synthesis of **30**:



The reaction of 4DPAIPN (1.1 mg, 0.001 mmol), 1a (22.2 mg, 0.2 mmol), 2h (19.1 mg, 0.1 mmol), S5 (8.8 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **30** (17.2 mg, 57%, 92% D) (eluent: $CH_2Cl_2/MeOH = 20/1$) as a white solid: 93:7 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 75/25, 0.5 mL/min, λ = 214 nm, $t_{\rm R}$ (minor) = 18.4 min, $t_{\rm R}$ (major) = 21.3 min); $[\alpha]_{\rm P}^{25}$ = -52.2 (c = 0.07, CHCl₃); m.p. 96-100 °C; ¹H **NMR** (400 MHz, CDCl₃) δ = 7.40-7.29 (m, 2 H), 7.08-6.94 (m, 2 H), 6.76 (s, 2 H), 4.22-4.08 (m, 0.08 H), 3.67 (s, 6 H), 2.68-2.54 (m, 1 H), 2.54-2.40 (m, 1 H), 2.37-2.23 (m, 1 H), 1.97-1.84 (m, 1 H), 0.75 (d, J = 12.4 Hz, 1 H), 0.29 (d, J = 12.4 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) $\delta =$ 174.8, 160.9, 158.5, 134.7, 125.6 (d, $J_{C-F} = 7.9$ Hz), 120.1, 115.2 (d, $J_{C-F} = 22.2$ Hz), 35.7, 32.0, 25.6; ¹¹B NMR (128 MHz, CDCl₃) δ = -29.7 (t, J = 67.2 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ = -117.9. HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).





The reaction of 4DPAIPN (0.9 mg, 0.001 mmol), **1a** (22.1 mg, 0.2 mmol), **2i** (20.1 mg, 0.1 mmol), **S5** (8.9 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **3p** (18.6 mg, 60%, 95% D) (eluent: CH₂Cl₂/MeOH = 20/1) as an oil: 93:7 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 60/40, 0.5 mL/min, λ = 214 nm, t_R (minor) = 13.0 min, t_R (major) = 14.8 min); $[\alpha]_D^{25}$ = -35.6 (c = 0.16, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.31-7.22 (m, 2 H), 6.92-6.83 (m, 2 H), 6.75 (s, 2 H), 4.19-4.04 (m, 0.05 H), 3.78 (s, 3 H), 3.67 (s, 6 H), 2.68-2.39 (m, 2 H), 2.37-2.24 (m, 1 H), 1.97-1.82 (m, 1 H), 0.76 (d, J = 12.4 Hz, 1 H), 0.28 (d, J = 12.8 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = -29.7 (t, J = 64.2 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 4a:



The reaction of 4DPAIPN (1.8 mg, 0.002 mmol), **1i** (107.2 mg, 0.4 mmol), **2a** (35.2 mg, 0.2 mmol), **S7** (17.8 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) under room temperature afforded **4a** (72.3 mg, 83%, 90% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 92:8 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 1.0 mL/min, $\lambda = 214$ nm, t_R (major) = 7.6 min, t_R (minor) = 11.5 min); $[\alpha]_{D}^{25} = +2.61$ (c = 0.15, CHCl₃); **m.p.** 110-111 °C; ¹H NMR (400 MHz, CDCl₃) $\delta = 7.58-7.24$ (m, 19 H), 7.22-7.14 (m, 1 H), 4.71-4.51 (m, 0.1 H), 3.95 (d, J = 9.2 Hz, 1 H), 3.62 (d, J = 8.8 Hz, 1 H), 2.05 (d, J = 14.8 Hz, 1 H), 1.61 (d, J = 14.8 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 155.4$, 136.2, 135.4, 133.0, 130.2, 129.2, 128.3, 125.4, 122.7, 68.5, 17.9; HRMS (ESI) calcd for C₂₈H₂₄DNNaO₂Si [M+Na⁺]: 459.1610, found: 459.1602.

The reaction of 4DPAIPN (0.9 mg, 0.001 mmol), **1i** (52.3 mg, 0.2 mmol), **2a** (17.4 mg, 0.1 mmol), **S5** (8.8 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **4a** (37.1 mg, 85%, 89% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 88:12 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 1.0 mL/min, λ = 214 nm, t_R (major) = 7.6 min, t_R (minor) = 11.5 min); **¹H NMR** (400 MHz, CDCl₃) δ = 7.72-7.02 (m, 20 H), 4.66-4.50 (m, 0.11 H), 3.96 (d, *J* = 8.8 Hz, 1 H), 3.63 (d, *J* = 8.8 Hz, 1 H), 2.05 (d, *J* = 14.8 Hz, 1 H), 1.61 (d, *J* = 14.8 Hz, 1 H).

The reaction of 4DPAIPN (0.8 mg, 0.001 mmol), **1i** (52.5 mg, 0.2 mmol), **2a** (17.3 mg, 0.1 mmol), **S1** (6.2 mg, 0.015 mmol), toluene (0.75 mL) and D_2O (0.25 mL) under room temperature

afforded **4a** (30.5 mg, 70%, 96% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 46:54 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 1.0 mL/min, λ = 214 nm, t_R (minor) = 7.6 min, t_R (major) = 11.5 min); ¹H NMR (400 MHz, CDCl₃) δ = 7.75-7.09 (m, 20 H), 4.66-4.52 (m, 0.04 H), 3.96 (d, *J* = 9.2 Hz, 1 H), 3.63 (d, *J* = 9.2 Hz, 1 H), 2.05 (d, *J* = 14.8 Hz, 1 H), 1.61 (d, *J* = 14.8 Hz, 1 H).

Gram-scale synthesis of **4a** with **S7**:



The reaction of 4DPAIPN (12.0 mg, 0.015 mmol), **1i** (1.5261 g, 6.0 mmol), **2a** (525.3 mg, 3.0 mmol), **S7** (168.3 mg, 0.3 mmol), toluene (22.5 mL) and D₂O (7.5 mL) under room temperature afforded **4a** (1.08 g, 82%, 91% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 91:9 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 1.0 mL/min, λ = 214 nm, t_R (major) = 7.6 min, t_R (minor) = 11.4 min); ¹H NMR (400 MHz, CDCl₃) δ = 7.66-7.06 (m, 20 H), 4.73-4.45 (m, 0.09 H), 3.96 (d, *J* = 9.2 Hz, 1 H), 3.62 (d, *J* = 8.8 Hz, 1 H), 2.05 (d, *J* = 14.4 Hz, 1 H), 1.61 (d, *J* = 14.8 Hz, 1 H).

Synthesis of **4a** using recycled D₂O:



The reaction of 4DPAIPN (1.8 mg, 0.002 mmol), **1i** (104.2 mg, 0.4 mmol), **2a** (35.0 mg, 0.2 mmol), **S7** (16.9 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL, recycled from a 0.6 mmol scale reaction) under room temperature afforded **4a** (73.3 mg, 84%, 80% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 91:9 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 1.0 mL/min, λ = 214 nm, $t_{\rm R}$ (major) = 7.6 min, $t_{\rm R}$ (minor) = 11.3 min); ¹H NMR (400 MHz, CDCl₃) δ = 7.79-7.05 (m, 20

H), 4.68-4.50 (m, 0.20 H), 3.97 (d, *J* = 8.8 Hz, 1 H), 3.63 (d, *J* = 8.8 Hz, 1 H), 2.05 (d, *J* = 14.4 Hz, 1 H), 1.61 (d, *J* = 14.8 Hz, 1 H).

The reaction of 4DPAIPN (1.7 mg, 0.002 mmol), **1i** (104.1 mg, 0.4 mmol), **2a** (35.0 mg, 0.2 mmol), **S7** (17.0 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL, recycled from a 3.0 mmol scale reaction) under room temperature afforded **4a** (73.3 mg, 84%, 83% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 92:8 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 1.0 mL/min, λ = 214 nm, $t_{\rm R}$ (major) = 7.7 min, $t_{\rm R}$ (minor) = 11.5 min); ¹H NMR (400 MHz, CDCl₃) δ = 7.59-7.09 (m, 20 H), 4.64-4.54 (m, 0.17 H), 3.96 (d, *J* = 8.8 Hz, 1 H), 3.63 (d, *J* = 9.2 Hz, 1 H), 2.05 (d, *J* = 14.8 Hz, 1 H).

Synthesis of 4b:



The reaction of 4DPAIPN (1.7 mg, 0.002 mmol), **1i** (106.2 mg, 0.4 mmol), **2j** (29.9 mg, 0.2 mmol), **S5** (17.7 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **4b** (50.8 mg, 61%, 91% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 82:18 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 96/4, 1.0 mL/min, $\lambda = 214$ nm, t_R (minor) = 9.5 min, t_R (major) = 11.6 min); $[\alpha]_D^{25} = -17.3$ (c = 0.41, CHCl₃); **m.p.** 83-84 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.61$ -7.49 (m, 6 H), 7.49-7.32 (m, 9 H), 4.04-3.92 (m, 0.09 H), 3.76 (d, J = 8.8 Hz, 1 H), 3.54-3.40 (m, 2 H), 3.19-3.00 (m, 1 H), 2.06 (d, J = 14.4 Hz, 1 H), 1.58-1.22 (m, 5 H), 0.91 (t, J = 7.2 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃); 157.7, 135.3, 133.2, 130.1, 128.3, 68.7, 52.1 (t, $J_{C-D} = 21.8$ Hz), 40.7, 29.3, 19.9, 17.3, 13.7; **HRMS** (ESI) calcd for C₂₆H₂₈DNNaO₂Si [M+Na⁺]: 439.1923, found: 439.1921.

Synthesis of 4c:



The reaction of 4DPAIPN (0.8 mg, 0.001 mmol), **1i** (52.4 mg, 0.24 mmol), **2d** (18.9 mg, 0.1 mmol), **S7** (8.9 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **4c** (26.6 mg, 59%, 95% D) (eluent: petroleum ether/ethyl acetate = 5/1) as an oil: 95:5 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 80/20, 0.5 mL/min, λ = 214 nm, $t_{\rm R}$ (major) = 8.9 min, $t_{\rm R}$ (minor) = 9.8 min); $[\alpha]_{\rm D}^{25}$ = -2.4 (*c* = 0.13, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.52-7.20 (m, 18 H), 7.18-7.08 (m, 2 H), 4.15-4.00 (m, 0.05 H), 2.58-2.47 (m, 1 H), 2.46-2.33 (m, 1 H), 2.02-1.54 (m, 5 H), 1.53-1.41 (m, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.0, 141.4, 135.4, 133.9, 129.7, 129.3, 128.4, 128.0, 127.1, 32.5, 29.1, 18.1, 17.5; HRMS (ESI) calcd for C₃₀H₂₈DNNaOSi [M+Na⁺]: 471.1973, found: 471.1975.

Synthesis of 4d:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1i** (52.3 mg, 0.24 mmol), **2e** (20.5 mg, 0.1 mmol), **S5** (9.0 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **4d** (31.4 mg, 67%, 93% D) (eluent: petroleum ether/ethyl acetate = 1/1) as an oil: 92:8 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 80/20, 0.5 mL/min, λ = 214 nm, $t_{\rm R}$ (major) = 11.8 min, $t_{\rm R}$ (minor) = 12.7 min); $[\alpha]_{\rm D}^{25}$ = -10.6 (*c* = 0.11, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.47-7.26 (m, 15 H), 7.13-7.01 (m, 4 H), 4.07-3.96 (m, 0.07 H), 2.58-2.32 (m, 2 H), 1.98-1.73 (m, 3 H), 1.73-1.43 (m, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.2, 135.4, 133.8, 130.1, 130.0, 129.7, 128.0, 116.2, 116.0, 32.5, 29.2, 18.3, 17.6, ¹⁹F NMR (376 MHz, CDCl₃) δ = -114.9; HRMS (ESI) calcd for C₃₀H₂₇DFNNaOSi [M+Na⁺]: 489.1879, found: 489.1879.

Synthesis of 4e:



The reaction of 4DPAIPN (0.9 mg, 0.001 mmol), **1j** (40.6 mg, 0.2 mmol), **2d** (18.8 mg, 0.1 mmol), **S5** (8.9 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **4e** (20.1 mg, 52%, 95% D) (eluent: petroleum ether/ethyl acetate = 2/1) as an colorless oil: 95:5 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 90/10, 0.5 mL/min, λ = 214 nm, $t_{\rm R}$ (major) = 25.0 min, $t_{\rm R}$ (minor) = 30.0 min); $[\alpha]_{\rm D}^{25}$ = -16.8 (c = 0.17, CHCl₃); ¹H **NMR** (400 MHz, CDCl₃) δ = 7.45-7.19 (m, 13 H), 7.17-7.09 (m, 2 H), 4.04-3.88 (m, 0.05 H), 2.63-2.36 (m, 2 H), 2.00-1.84 (m, 2 H), 1.78-1.47 (m, 3 H), 1.41-1.30 (m, 1 H), 0.50 (s, 3 H); ¹³C **NMR** (100 MHz, CDCl₃) δ = 169.9, 141.4, 136.4, 135.5, 134.2, 134.1, 129.44, 129.41, 129.2, 128.3, 127.97, 127.95, 127.1, 32.6, 29.6, 19.7, 17.8, -3.8; **HRMS** (ESI) calcd for C₂₅H₂₆DNNaOSi [M+Na⁺]: 409.1817, found: 409.1716.

Synthesis of 4f:



The reaction of 4DPAIPN (1.0 mg, 0.001 mmol), **1k** (50.2 mg, 0.2 mmol), **2d** (18.7 mg, 0.1 mmol), **S5** (8.9 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **4f** (26.3 mg, 60%, 89% D) (eluent: petroleum ether/ethyl acetate = 2/1) as a white solid: 97:3 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 90/10, 0.5 mL/min, λ = 214 nm, t_R (major) = 9.8 min, t_R (minor) = 10.6 min); $[\alpha]_D^{25}$ = -58.5 (c = 0.19, CHCl₃); **m.p.** 108-112 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.45-7.34 (m, 2 H), 7.31-7.22 (m, 1 H), 7.19-7.09 (m, 2 H), 3.90-3.81 (m, 0.11 H), 2.64-2.44 (m, 2 H), 2.20-2.08 (m, 1 H), 2.07-1.74 (m, 3 H), 1.25-1.16 (m, 1 H), 1.04-0.93 (m, 1 H), 0.06 (s, 27 H); ¹³C **NMR** (100 MHz, CDCl₃) δ = 169.7, 141.7, 129.1, 128.3, 127.1, 32.6, 29.2, 17.9, 13.2, 1.1; **HRMS** (ESI) calcd for C₂₁H₄₀DNNaO₂Si₄ [M+Na⁺]: 459.2220, found: 459.2221.

Synthesis of 4g:



The reaction of 4DPAIPN (0.9 mg, 0.001 mmol), **11** (37 µL, 0.2 mmol), **2a** (17.4 mg, 0.1 mmol), **S5** (8.6 mg, 0.015 mmol), toluene (0.75 mL) and D₂O (0.25 mL) under room temperature afforded **4g** (19.0 mg, 53%, 91% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 88:12 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 80/20, 1.0 mL/min, λ = 214 nm, t_R (major) = 6.3 min, t_R (minor) = 7.6 min); $[\alpha]_D^{25} = -2.7$ (c = 0.09, CHCl₃); **m.p.** 103-105 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.75-6.99 (m, 15 H), 4.97-4.85 (m, 1 H), 4.60-4.44 (m, 0.09 H), 4.35 (d, J = 8.8 Hz, 1 H), 3.98 (d, J = 8.8 Hz, 1 H), 1.89-1.75 (m, 1 H), 1.47-1.37 (m, 1 H); ¹³**C NMR** (100 MHz, CDCl₃) δ = 155.5, 136.1, 135.0, 134.94, 133.91, 132.0, 131.8, 130.5, 130.4, 129.2, 128.5, 128.3, 125.3, 122.2, 122.0, 68.2, 17.1; **HRMS** (ESI) calcd for C₂₂H₂₀DNNaO₂Si [M+Na⁺]: 383.1297, found: 383.1298.

Synthesis of 4h:



The reaction of 4DPAIPN (1.6 mg, 0.002 mmol), **1i** (105.0 mg, 0.4 mmol), **2k** (13.2 mg, 0.2 mmol), **S5** (17.5 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **4h** (34.7 mg, 74%, 94% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 82:18 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 96/4, 1.0 mL/min, $\lambda = 214$ nm, t_R (major) = 10.2 min, t_R (minor) = 11.2 min); $[\alpha]_D^{25} = +5.53$ (c = 0.36, CHCl₃); **m.p.** 85-86 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.64-7.48$ (m, 6 H), 7.48-7.25 (m, 9 H), 4.78-4.67 (m, 0.06 H), 2.47-2.29 (m, 2 H), 2.20 (d, J = 14.4 Hz, 1 H), 2.00-1.90 (m, 1 H), 1.78 (d, J = 14.8 Hz, 1 H), 1.71-1.57 (m, 1 H); ¹³**C NMR** (100 MHz, CDCl₃) $\delta = 176.8$, 135.5, 133.6, 129.8, 128.1, 79.0 (t, $J_{C-D} = 22.7$ Hz), 30.6, 29.4, 21.3; **HRMS** (ESI) calcd for C₂₃H₂₁DNaO₂Si [M+Na⁺]: 382.1344, found: 382.1343.

Synthesis of 4i:



The reaction of 4DPAIPN (1.6 mg, 0.002 mmol), **1i** (104.3 mg, 0.4 mmol), **2l** (20.0 mg, 0.2 mmol), **S5** (16.8 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **4i** (61.8 mg, 85%, 92% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as an oil: 50:50 er (HPLC conditions: Chiralcel OJ-H column, hexane/*i*-PrOH = 99/1, 1.0 mL/min, $\lambda = 214$ nm, t_R (major) = 7.6 min, t_R (minor) = 11.2 min); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.65-7.47$ (m, 6 H), 7.45-7.29 (m, 9 H), 5.21-5.09 (m, 0.08 H), 1.95 (d, J = 14.8 Hz, 1 H), 1.75-1.62 (m, 4 H), 1.19 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 170.3$, 135.6, 134.5, 129.5, 127.9, 68.8 (t, $J_{C-D} = 23.1$ Hz), 23.4, 21.8, 21.0; HRMS (ESI) calcd for C₂₃H₂₃DNaO₂Si [M+Na⁺]: 384.1501, found: 384.1500.

Synthesis of 4j:



The reaction of 4DPAIPN (1.8 mg, 0.002 mmol), **1i** (104.2 mg, 0.4 mmol), **2m** (40.7 mg, 0.2 mmol), **S8** (22.1 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **4j** (61.6 mg, 66%, 94% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as an oil: 53:47 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 96/4, 1.0 mL/min, $\lambda = 214$ nm, t_R (major) = 12.9 min, t_R (minor) = 18.2 min); ¹H NMR (400 MHz, CDCl₃) $\delta = 7.62$ -7.16 (m, 20 H), 5.06 (s, 2 H), 4.67 (s, 1 H), 3.15-2.95 (m, 2 H), 1.97-1.83 (m, 0.06 H), 1.49 (d, J = 15.2 Hz, 1 H), 1.24 (d, J = 14.8 Hz, 1 H), 0.80 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 156.5$, 136.6, 135.6, 135.1, 129.5, 128.5, 128.1, 127.9, 66.6, 49.7, 20.5, 18.2; HRMS (ESI) calcd for C₃₀H₃₀DNNaO₂Si [M+Na⁺]: 489.2079, found: 489.2078.

Synthesis of 5a:



The reaction of 4DPAIPN (1.5 mg, 0.002 mmol), **1m** (40.6 mg, 0.2 mmol), **2n** (33.8 mg, 0.24 mmol), **S8** (21.7 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **5a** (42.9 mg, 72%, 93% D) (purified by chromatography on silica gel, eluent: dichloromethane/methanol = 30/1) as a white solid: 96:4 er, (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 65/35, 0.5 mL/min, λ = 214 nm, t_R (major) = 14.0 min, t_R (minor) =15.1 min); $[\alpha]_D^{25}$ = -80.6 (*c* = 0.21, CHCl₃); **m.p.** 172-173 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.87-7.65 (m, 4 H), 7.64-7.45 (m, 6 H), 7.34 (brs, 1 H), 3.37 (t, *J* = 11.2 Hz, 0.07 H), 2.44-2.09 (m, 4 H), 1.64-1.49 (m, 2 H), 1.02-0.84 (m, 6 H); ¹³**C NMR** (100 MHz, CDCl₃) δ = 170.6, 133.1 (d, J_{C-P} = 99.9 Hz), 132.24 (d, J_{C-P} = 2.6 Hz), 132.19 (d, J_{C-P} = 2.8 Hz), 131.1 (d, J_{C-P} = 7.6 Hz), 130.4 (d, J_{C-P} = 11.6 Hz), 30.1 (d, J_{C-P} = 70.1 Hz), 28.2, 26.7, 18.6; ³¹**P NMR** (162 MHz, CDCl₃) δ = 33.4; **HRMS** (ESI) calcd for C₂₀H₂₃DNNaO₂P [M+Na⁺]: 365.1500, found: 365.1497.

Synthesis of 5b:



The reaction of 4DPAIPN (1.7 mg, 0.002 mmol), **1n** (60.4 mg, 0.2 mmol), **2n** (34.6 mg, 0.24 mmol), **S8** (21.8 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **5b** (63.2 mg, 70%, 90% D) (purified by chromatography on silica gel, eluent: dichloromethane/methanol = 20/1) as a white solid: 95:5 er, (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 80/20, 1 mL/min, $\lambda = 214$ nm, t_R (minor) = 71.6 min, t_R (major) = 82.0 min); $[\alpha]_D^{25} = -104.4$ (c = 0.24, CHCl₃); **m.p.** 209-211 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 8.58$ (d, J = 13.2 Hz, 1 H), 8.32 (d, J = 13.6 Hz, 1 H), 8.01-7.83 (m, 6 H), 7.76-7.52 (m, 6 H), 7.46 (brs, 1 H), 3.43 (t, J = 13.4 mmodels and the solid states of the
11.2 Hz, 0.10 H), 2.62-2.51 (m, 1 H), 2.42-2.22 (m, 3 H), 1.58-1.45 (m, 2 H), 1.00 (s, 3 H), 0.92 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.6, 134.8 (d, J_{C-P} = 2.4 Hz), 134.7 (d, J_{C-P} = 2.5 Hz), 134.1 (d, J_{C-P} = 7.8 Hz), 132.6 (d, J_{C-P} = 12.6 Hz), 132.4 (d, J_{C-P} = 13.1 Hz), 132.3 (d, J_{C-P} = 9.1 Hz), 130.2 (d, J_{C-P} = 100.4 Hz), 129.1-128.7 (m), 128.5, 128.4, 127.9-127.8 (m), 127.4 (d, J_{C-P} = 98.4 Hz), 127.3, 127.2, 125.3-125.0 (m), 35.0, 32.1 (d, J_{C-P} = 11.7 Hz), 30.0 (d, J_{C-P} = 70.3 Hz), 28.3, 26.8, 18.7; ³¹P NMR (162 MHz, CDCl₃) δ = 33.7; HRMS (ESI) calcd for C₂₈H₂₈DNO₂P [M+H⁺]: 443.1993, found: 443.1995.

Synthesis of 5c:



The reaction of 4DPAIPN (1.9 mg, 0.002 mmol), **10** (46.1 mg, 0.2 mmol), **2n** (36.6 mg, 0.24 mmol), **S8** (22.0 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **5c** (55.5 mg, 75%, 94% D) (purified by chromatography on silica gel, eluent: dichloromethane/methanol = 30/1) as a white solid: 97:3 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 60/40, 0.5 mL/min, λ = 214 nm, t_R (major) = 15.5 min, t_R (minor) = 24.9 min); $[\alpha]_D^{25}$ = -76.0 (c = 0.24, CHCl₃); **m.p.** 115-118 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.72-7.50 (m, 4 H), 7.44-7.16 (m, 5 H), 3.44-3.25 (m, 0.06 H), 2.49-2.21 (m, 9 H), 2.19-2.03 (m, 1 H), 1.63-1.47 (m, 2 H), 0.95 (s, 3 H), 0.90 (s, 3 H); ¹³C **NMR** (100 MHz, CDCl₃) δ = 170.6, 142.74-142.61 (m), 131.1 (d, J_{C-P} = 9.3 Hz), 130.3 (d, J_{C-P} = 9.9 Hz), 130.1 (d, J_{C-P} = 102.4 Hz), 129.7-129.3 (m), 127.3 (d, J_{C-P} = 100.5 Hz), 56.7 (t, J_{C-D} = 24.7 Hz), 35.0, 32.0 (d, J_{C-P} = 11.6 Hz), 30.3 (d, J_{C-P} = 70.3 Hz), 28.2, 26.8, 21.6-21.4 (m), 18.7; ³¹P **NMR** (162 MHz, CDCl₃) δ = 33.7; **HRMS** (ESI) calcd for C₂₂H₂₈DNO₂P [M+H⁺]: 371.1993, found: 371.1992.

Synthesis of 5d:



The reaction of 4DPAIPN (2.0 mg, 0.002 mmol), **1p** (47.4 mg, 0.2 mmol), **2n** (34.2 mg, 0.24 mmol), **S8** (21.7 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **5d** (43.3 mg, 57%, 92%/58% D) (purified by chromatography on silica gel, eluent: dichloromethane/methanol = 30/1) as a white solid: 95:5 er (HPLC conditions: Chiralcel AS-H column, hexane/*i*-PrOH = 80/20, 0.5 mL/min, λ = 214 nm, $t_{\rm R}$ (minor) = 22.2 min, $t_{\rm R}$ (major) = 32.0 min); $[\alpha]_{\rm D}^{25}$ = -66.8 (*c* = 0.40, CHCl₃); **m.p.** 150-152 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.91-7.58 (m, 4 H), 7.37-7.09 (m, 5 H), 3.39-3.3.26 (m, 0.08 H), 2.46-2.24 (m, 2.42 H), 2.23-2.10 (m, 1 H), 1.67-1.49 (m, 2 H), 1.03-0.80 (m, 6 H); ¹³C **NMR** (100 MHz, CDCl₃) δ = 170.7, 166.5 (dd, J_1 = 10.8 Hz, J_2 = 3.3 Hz), 163.9 (dd, J_1 = 10.6 Hz, J_2 = 3.3 Hz), 133.7-133.4 (m), 132.8 (dd, J_1 = 11.1 Hz, J_2 = 8.9 Hz), 128.9 (dd, J_1 = 102.9 Hz, J_2 = 3.2 Hz), 126.1 (dd, J_1 = 101.4 Hz, J_2 = 3.1 Hz), 116.8-116.2 (m), 34.9, 32.0 (d, $J_{\rm C-P}$ = 11.7 Hz), 30.4 (d, $J_{\rm C-P}$ = 71.3 Hz), 28.2, 26.7, 18.7; ¹⁹**F NMR** (376 MHz, CDCl₃) δ = -105.47, -105.48; ³¹**P NMR** (162 MHz, CDCl₃) δ = 32.4; **HRMS** (ESI) calcd for C₂₀H₂₀D₂F₂NNaO₂P [M+Na⁺]: 402.1374, found: 402.1370.

Synthesis of **5e**:



The reaction of 4DPAIPN (1.9 mg, 0.002 mmol), **1q** (58.0 mg, 0.2 mmol), **2n** (34.7 mg, 0.2 mmol), **S8** (21.6 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **5e** (37.5 mg, 44%, 95% D) (purified by chromatography on silica gel, eluent: dichloromethane/methanol = 30/1) as a white solid: 95:5 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 70/30, 1 mL/min, $\lambda = 214$ nm, t_R (major) = 18.6 min, t_R (minor) = 22.4 min); $[\alpha]_D^{25} = -71.5$ (c = 70/30, 1 mL/min, $\lambda = 214$ nm, t_R (major) = 18.6 min, t_R (minor) = 22.4 min); $[\alpha]_D^{25}$

0.22, CHCl₃); **m.p.** 185-186 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.36-7.03 (m, 5 H), 7.01-6.83 (m, 2 H), 6.20-5.87 (m, 4 H), 3.37 (t, *J* = 11.2 Hz, 0.05 H), 2.42-2.19 (m, 3 H), 2.15-1.94 (m, 1 H), 1.66-1.50 (m, 2 H), 1.01-0.82 (m, 6 H); ¹³**C NMR** (100 MHz, CDCl₃) δ = 170.7, 151.1 (d, *J*_{C-P} = 2.9 Hz), 151.0 (d, *J*_{C-P} = 2.9 Hz), 148.5-148.1 (m), 126.3(d, *J*_{C-P} = 104.6 Hz), 126.4 (d, *J*_{C-P} = 9.9 Hz), 125.6 (d, *J*_{C-P} = 10.6 Hz), 123.4 (d, *J*_{C-P} = 102.9 Hz), 110.3 (d, *J*_{C-P} = 11.9 Hz), 109.8 (d, *J*_{C-P} = 12.5 Hz), 109.1 (d, *J*_{C-P} = 9.0 Hz), 108.9 (d, *J*_{C-P} = 9.4 Hz), 101.8 (d, *J*_{C-P} = 6.6 Hz), 35.0, 32.0 (d, *J*_{C-P} = 11.7 Hz), 30.4 (d, *J*_{C-P} = 71.6 Hz), 28.2, 26.8, 18.7; ³¹**P NMR** (162 MHz, CDCl₃) δ = 33.8; **HRMS** (ESI) calcd for C₂₂H₂₃DNNaO₆P [M+Na⁺]: 453.1296, found: 453.1298.

Synthesis of 5f:



The reaction of 4DPAIPN (2.0 mg, 0.002 mmol), **1m** (40.6 mg, 0.2 mmol), **2o** (34.9 mg, 0.24 mmol), **S8** (22.0 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **5f** (42.7 mg, 65%, 93%/50% D) (purified by chromatography on silica gel, eluent: dichloromethane/methanol = 30/1) as a white solid: 94:6 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 60/40, 0.5 mL/min, λ = 214 nm, t_R (major) = 12.1 min, t_R (minor) = 12.9 min); $[\alpha]_D^{25}$ = -78.1 (c = 0.28, CHCl₃); **m.p.** 149-153 °C; ¹H NMR (400 MHz, CDCl₃) δ = 7.85-7.66 (m, 4 H), 7.63-7.43 (m, 6 H), 6.70 (brs, 1 H), 3.54-3.44 (m, 0.07 H), 2.41-2.31 (m, 0.5 H), 2.30-1.94 (m, 3 H), 1.09-1.00 (m, 6 H); ¹³C NMR (100 MHz, CDCl₃) δ = 175.6, 132.8 (d, J_{C-P} = 99.8 Hz), 132.3-132.1 (m), 131.3, 130.8 (d, J_{C-P} = 9.1 Hz), 130.3 (d, J_{C-P} = 9.7 Hz), 129.0 (d, J_{C-P} = 6.2 Hz), 128.8 (d, J_{C-P} = 5.3 Hz), 45.6, 39.1 (d, J_{C-P} = 11.9 Hz), 29.6 (d, J_{C-P} = 70.7 Hz), 25.6, 22.6; ³¹P NMR (162 MHz, CDCl₃) δ = 32.4; HRMS (ESI) calcd for C₁₉H₂₀D₂NNaO₂P [M+Na⁺]: 352.1406, found: 352.1401.

Synthesis of 5g:



The reaction of 4DPAIPN (1.7 mg, 0.002 mmol), **1m** (40.3 mg, 0.2 mmol), **2p** (63.7 mg, 0.24 mmol), **S8** (22.6 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **5g** (57.9 mg, 62%, 96% D) (purified by chromatography on silica gel, eluent: dichloromethane/methanol = 30/1) as a white solid: >99:1 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 70/30, 1 mL/min, λ = 214 nm, t_R (minor) = 11.6 min, t_R (major) = 14.7 min); $[\alpha]_D^{25}$ = -147.1 (*c* = 0.28, CHCl₃); **m.p.** 234-236 °C; ¹**H NMR** (400 MHz, CDCl₃) δ = 7.81-7.61 (m, 2 H), 7.59-7.02 (m, 18 H), 5.82 (t, *J* = 10.6 Hz, 0.04 H), 2.84-2.66 (m, 1 H), 2.65-2.37 (m, 3 H), 2.32-2.09 (m, 2 H); ¹³**C NMR** (100 MHz, CDCl₃) δ = 168.3, 143.2 (d, J_{C-P} = 10.1 Hz), 133.1 (d, J_{C-P} = 100.6 Hz), 131.95 (d, J_{C-P} = 10.08 Hz), 131.86 (d, J_{C-P} = 2.9 Hz), 131.78 (d, J_{C-P} = 4.5 Hz), 128.5 (d, J_{C-P} = 4.7 Hz), 127.9 (d, J_{C-P} = 12.5 Hz), 127.1 (d, J_{C-P} = 31.3 Hz), 48.5 (d, J_{C-P} = 9.2 Hz), 33.5 (d, J_{C-P} = 70.2 Hz), 27.9, 27.4; ³¹**P NMR** (162 MHz, CDCl₃) δ = 29.4; **HRMS** (ESI) calcd for C₃₀H₂₆DNaO₃P [M+Na⁺]: 490.1653, found: 490.1649.

Synthesis of 5h:



The reaction of 4DPAIPN (1.9 mg, 0.002 mmol), **1r** (42.8 mg, 0.2 mmol), **2q** (33.0 mg, 0.24 mmol), **S8** (21.6 mg, 0.03 mmol), toluene (1.5 mL) and D₂O (0.5 mL) afforded **5h** (57.0 mg, 80%, 81% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 89:11 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 88/12, 1 mL/min, $\lambda = 214$ nm, t_R (minor) = 9.1 min, t_R (major) = 9.9 min); $[\alpha]_D^{25} = -83.1$ (c = 0.24, CHCl₃); **m.p.** 151-153 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 8.04-7.76$ (m, 4 H), 7.62-7.33 (m, 6

H), 4.81 (dd, $J_1 = 15.4$ Hz, $J_2 = 9.0$ Hz, 0.19 H), 3.06-2.76 (m, 1 H), 2.57-2.24 (m, 3 H), 1.86-1.73 (m, 1 H), 1.66-1.54 (m, 1 H), 1.09-0.87 (m, 6 H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 170.1$, 133.9 (d, $J_{C-P} = 82.8$ Hz), 131.8 (d, $J_{C-P} = 10.7$ Hz), 131.6 (d, $J_{C-P} = 3.1$ Hz), 131.5 (d, $J_{C-P} = 2.9$ Hz), 131.4, 130.7 (d, $J_{C-P} = 10.0$ Hz), 128.7 (d, $J_{C-P} = 12.1$ Hz), 128.2 (d, $J_{C-P} = 12.6$ Hz), 81.0-80.4 (m), 34.1 (d, $J_{C-P} = 55.9$ Hz), 34.0, 32.4 (d, $J_{C-P} = 10.1$ Hz), 27.2, 26.2, 19.8; ³¹P NMR (162 MHz, CDCl₃) $\delta = 42.6$; **HRMS** (ESI) calcd for C₂₀H₂₃DO₂PS [M+H⁺]: 360.1292, found: 360.1291.

Synthesis of 5i:



The reaction of 4DPAIPN (2.0 mg, 0.002 mmol), **1s** (30.5 mg, 0.2 mmol), **2p** (65.0 mg, 0.24 mmol), **S8** (22.2 mg, 0.03 mmol), toluene (1.5 ml) and D₂O (0.5 mL) afforded **5i** (51.9 mg, 62%, 97% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 5/1) as a white solid: 96:4 er (HPLC conditions: Chiralcel IC-H column, hexane/*i*-PrOH = 80/20, 0.5 mL/min, $\lambda = 214$ nm, t_R (minor) = 15.6 min, t_R (major) = 17.8 min); $[\alpha]_D^{25} = -195.4$ (c = 0.25, CHCl₃); **m.p.** 78-79 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.41$ -7.19 (m, 8 H), 7.17-7.09 (m, 2 H), 5.85-5.75 (m, 0.03 H), 4.27-3.90 (m, 4 H), 2.76-2.50 (m, 3 H), 2.34-2.15 (m, 2 H), 1.86-1.70 (m, 1 H), 1.29 (t, J = 7.0 Hz, 3 H), 1.24 (t, J = 7.2 Hz, 3 H); ¹³C **NMR** (100 MHz, CDCl₃) $\delta = 168.7$, 143.2 (d, $J_{C-P} = 15.4$ Hz), 128.9 (d, $J_{C-P} = 6.4$ Hz), 48.0 (d, $J_{C-P} = 28.7$ Hz), 127.1 (d, $J_{C-P} = 26.0$ Hz), 63.7 (d, $J_{C-P} = 6.3$ Hz), 61.9 (d, $J_{C-P} = 7.1$ Hz), 16.0 (d, $J_{C-P} = 7.5$ Hz); ³¹P **NMR** (162 MHz, CDCl₃) $\delta = 94.9$; **HRMS** (ESI) calcd for C₂₂H₂₆DNaO₄PS [M+Na⁺]: 442.1323, found: 442.1320.

Synthesis of 5j:



The reaction of 4DPAIPN (1.6 mg, 0.002 mmol), **1t** (37.0 mg, 0.19 mmol), **2q** (56.0 mg, 0.4 mmol), **S8** (21.6 mg, 0.03 mmol), toluene (1.5 ml) and D₂O (0.5 mL) afforded **5j** (42.3 mg, 67%, 88% D) (purified by chromatography on silica gel, eluent: petroleum ether/ethyl acetate = 2/1) as a white solid: 94:6 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 70/30, 0.8 mL/min, $\lambda = 214$ nm, t_R (minor) = 6.1 min, t_R (major) = 8.1 min); $[\alpha]_D^{25} = -32.4^{\circ}$ (c = 0.17, CHCl₃); **m.p.** 117-119 °C; ¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.88-7.75$ (m, 2 H), 7.75-7.62 (m, 2 H), 7.57-7.33 (m, 6 H), 4.55-4.43 (m, 0.12 H), 2.70-2.57 (m, 1 H), 2.54-2.36 (m, 2 H), 2.29-2.18 (m, 1 H), 1.79-1.68 (m, 1 H), 1.65-1.54 (m, 1 H), 1.50-0.55 (m, 9 H); ¹³**C NMR** (100 MHz, CDCl₃) $\delta = 170.1$, 132.9 (d, $J_{C-P} = 9.8$ Hz), 131.9 (d, $J_{C-P} = 9.1$ Hz), 131.5 (d, $J_{C-P} = 2.5$ Hz), 131.2 (d, $J_{C-P} = 2.5$ Hz), 130.0 (d, $J_{C-P} = 57.4$ Hz), 128.3 (d, $J_{C-P} = 9.9$ Hz), 128.5 (d, $J_{C-P} = 10.4$ Hz), 127.5 (d, $J_{C-P} = 55.0$ Hz), 81.4 (t, $J_{C-D} = 23.4$ Hz), 33.9, 32.6 (d, $J_{C-P} = 8.8$ Hz), 27.6 (d, $J_{C-P} = 36.5$ Hz), 27.2, 26.2, 19.4; ³¹**P NMR** (162 MHz, CDCl₃) $\delta = 16.2$ (d, J = 77.8 Hz); ¹¹**B NMR** (128 MHz, CDCl₃) $\delta = -39.4$; **HRMS** (ESI) calcd for C₂₀H₂₅BDNaO₂P [M+Na⁺]: 364.1718, found: 364.1726.

Synthesis of 6a:



Typical procedure II: To an oven-dried 16 x 60 mm vial containing a dry Teflon stir bar were charged with Hantzsch Ester (HE, 63.2 mg, 0.25 mmol), 1u (68.0 mg, 0.2 mmol), S8 (10.9 mg, 0.015 mmol), and (PhO)₂PO₂H (4.9 mg, 0.02 mmol), the vial was then taken into a N₂-filled glovebox where dry toluene (0.75 mL), 2q (14.2 mg, 0.1 mmol), and D₂O (0.25 mL) were added sequentially. The vial was sealed with a cap and parafilm and then taken out of the glovebox. The vial was placed in a cooling station and a 30 W blue LED ($\lambda = 441$ nm) was then placed on the top of the cooling station (Supplementary Figure 5), which is connected to a chiller to maintain the temperature of the cooling water at 10 °C. The reaction mixture was stirred at 10 °C under irradiation with a stirring rate of 400 r/min for 36 h. When the reaction is complete as monitored by TLC and GC, CH₂Cl₂ (10 mL) and H₂O (5 mL) were added, the organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (10 mL \times 3). The combined organic layer was washed with brine and dried over anhydrous Na₂SO₄. After filtration and evaporation, the residue was purified by chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 20/1 to 15/1 to 5/1) to afford 6a (28.1 mg, 71%, 94% D) as a faint yellow oil: 93:7 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 0.5 mL/min, λ = 214 nm, t_R (major) = 10.0 min, $t_{\rm R}$ (minor) = 10.9 min); $[\alpha]_{\rm D}^{25}$ = -44.8 (c = 0.16, CHCl₃); ¹H NMR (400 MHz, CDCl₃) $\delta = 4.43$ (dd, $J_1 = 9.2$ Hz, $J_2 = 1.2$ Hz, 0.06 H), 4.02 (tt, $J_1 = 11.6$ Hz, $J_2 = 3.5$ Hz, 1 H), 3.01 (tt, $J_1 = 11.8 \text{ Hz}, J_2 = 3.8 \text{ Hz}, 1 \text{ H}$, 2.66-2.26 (m, 6 H), 1.91-1.02 (m, 23 H), 0.99 (s, 3 H); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3) \delta = 170.8, 162.0 \text{ (t, } J_{\text{C-F}} = 27.1 \text{ Hz}), 118.9 \text{ (t, } J_{\text{C-F}} = 257.1 \text{ Hz}), 80.6 \text{ (t, } J_{\text{C-D}} = 257.1 \text{ Hz})$ 24.0 Hz), 57.1, 36.3-35.6 (m), 34.11, 34.08, 31.9, 31.8, 30.9, 29.0, 28.9, 27.1, 26.4, 26.3, 26.23, 26.20, 25.6, 25.5, 25.22, 25.17, 19.3; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -97.8$ (d, J = 279.4 Hz), -100.4 (d, J = 279.0 Hz); **HRMS** (ESI) calcd for C₂₂H₃₄DF₂NNaO₃ [M+Na⁺]: 423.2540, found: 423.2537.

The following compounds **6b-6f** were prepared according to the above **Typical Procedure II** unless otherwise stated. All the racemic samples for HPLC measurement were also prepared according to this procedure but using CySH instead of the thiol catalyst **S8**. Synthesis of 6b:



The reaction of HE (63.4 mg, 0.25 mmol), **1v** (51.8 mg, 0.2 mmol), **S8** (10.9 mg, 0.015 mmol), (PhO)₂PO₂H (5.1 mg, 0.02 mmol), toluene (0.75 mL), **2q** (14.4 mg, 0.1 mmol) and D₂O (0.25 mL) afforded **6b** (25.6 mg, 78%, 92% D) (eluent: petroleum ether/ethyl acetate = 20/1 to 10/1 to 5/1) as a faint yellow oil: 92:8 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 90/10, 0.5 mL/min, λ = 214 nm, t_R (major) = 10.6 min, t_R (minor) = 11.4 min); $[\alpha]_D^{25}$ = -40.1 (c = 0.18, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 4.56-4.40 (m, 1.08 H), 3.56-3.42 (m, 1 H), 2.66-2.44 (m, 3 H), 2.44-2.27 (m, 1 H), 1.86-1.74 (m, 1 H), 1.73-1.61 (m, 1 H), 1.45-1.37 (m, 6 H), 1.26-1.18 (m, 6 H), 1.05 (s, 3 H), 0.99 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.7, 161.7 (t, J_{C-F} = 27.3 Hz), 118.7 (t, J_{C-F} = 255.6 Hz), 48.4 (t, J = 7.5 Hz), 46.9, 35.9 (t, J = 22.9 Hz), 34.1, 31.8, 27.1, 26.2, 20.5, 20.4, 19.81, 19.78, 19.3; ¹⁹F NMR (376 MHz, CDCl₃) δ = -97.8 (d, J = 279.4 Hz), -100.6 (d, J = 279.4 Hz); HRMS (ESI) calcd for C₁₆H₂₆DF₂NNaO₃ [M+Na⁺]: 343.1914, found: 343.1914.

Synthesis of 6c:



The reaction of HE (63.4 mg, 0.25 mmol), **1u** (67.8 mg, 0.2 mmol), **S8** (10.8 mg, 0.015 mmol), (PhO)₂PO₂H (5.3 mg, 0.02 mmol), toluene (0.75 mL), **2p** (26.4 mg, 0.1 mmol) and D₂O (0.25 mL) afforded **6c** (25.8 mg, 49%, 92% D) (eluent: petroleum ether/ethyl acetate = 15/1 to 8/1) as a faint yellow oil: 97:3 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 95/5, 1 mL/min, λ = 214 nm, $t_{\rm R}$ (major) = 9.5 min, $t_{\rm R}$ (minor) = 10.6 min); $[\alpha]_{\rm D}^{25}$ = -113.3 (*c* = 0.16, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.40-7.25 (m, 6 H), 7.25-7.13 (m, 4 H), 5.70-5.64 (m, 0.08 H), 4.06-3.90 (m, 1 H), 3.05-2.90 (m, 1 H), 2.88-2.75 (m, 1 H), 2.72-2.49 (m, 3 H),

2.40-1.96 (m, 4 H), 1.86-0.99 (m, 18 H); ¹³C NMR (100 MHz, CDCl₃) δ = 169.1, 161.7, 143.3, 143.24, 143.21, 128.9, 128.8, 127.4, 127.11, 127.07, 127.06, 126.8, 57.1, 47.8, 47.7, 30.9, 29.1, 28.9, 27.3, 26.99, 26.97, 26.4, 26.3, 25.6, 25.5, 25.22, 25.20; ¹⁹F NMR (376 MHz, CDCl₃) δ = -100.1 (d, *J* = 285.0 Hz), -102.6 (d, *J* = 285.0 Hz); HRMS (ESI) calcd for C₃₂H₃₈DF₂NNaO₃ [M+Na⁺]: 547.2853, found: 547.2848.

Synthesis of 6d:



The reaction of HE (63.3 mg, 0.25 mmol), **1u** (67.5 mg, 0.2 mmol), **S8** (10.8 mg, 0.015 mmol), (PhO)₂PO₂H (5.0 mg, 0.02 mmol), toluene (0.75 mL), **2r** (13.0 mg, 0.1 mmol) and D₂O (0.25 mL) afforded **6d** (27.0 mg, 70%, 91% D) (eluent: petroleum ether/ethyl acetate = 20/1 to 15/1 to 10/1) as a faint yellow oil: 96:4 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 0.5 mL/min, λ = 214 nm, t_R (major) = 9.4 min, t_R (minor) = 10.4 min); $[\alpha]_D^{25}$ = -40.6 (c = 0.18, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 4.44 (dd, J_1 = 9.6 Hz, J_2 = 1.6 Hz, 0.09 H), 4.03 (tt, J_1 = 11.7 Hz, J_2 = 3.5 Hz, 1 H), 3.02 (tt, J_1 = 12.0 Hz, J_2 = 3.8 Hz, 1 H), 2.66-2.25 (m, 6 H), 1.99-0.99 (m, 24 H); ¹³C NMR (100 MHz, CDCl₃) δ = 175.5, 161.7 (t, J = 27.0 Hz), 118.70 (t, J = 257.4 Hz), 81.4 (t, J_{C-D} = 23.1 Hz), 57.2-57.0 (m), 44.1, 39.7, 39.6, 34.7-34.2 (m), 30.8, 28.9, 26.27, 26.25, 25.51, 25.47, 25.14, 25.09, 24.1, 21.0; ¹⁹F NMR (376 MHz, CDCl₃) δ = -97.0 (d, J = 281.6 Hz), -100.7 (d, J = 281.6 Hz); **HRMS** (ESI) calcd for C₂₁H₃₂DF₂NNaO₃ [M+Na⁺]: 409.2383, found: 409.2387.

Synthesis of 6e:



The reaction of HE (63.8 mg, 0.25 mmol), **1u** (67.8 mg, 0.2 mmol), **S8** (10.7 mg, 0.015 mmol), (PhO)₂PO₂H (5.2 mg, 0.02 mmol), toluene (0.75 mL), **2s** (17.0 mg, 0.1 mmol) and D₂O (0.25 mL) afforded **6e** (35.5 mg, 83%, 95% D) (eluent: petroleum ether/ethyl acetate = 20/1 to 15/1 to 7/1) as a faint yellow oil: 95:5 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 98/2, 1.0 mL/min, λ = 214 nm, t_R (major) = 9.6 min, t_R (minor) = 10.6 min); $[\alpha]_D^{25}$ = -35.0 (c = 0.16, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 4.60-4.53 (m, 0.05 H), 4.03 (tt, J_1 = 11.6 Hz, J_2 = 3.6 Hz, 1 H), 3.01 (tt, J_1 = 11.9 Hz, J_2 = 3.7 Hz, 1 H), 2.70-2.30 (m, 6 H), 1.94-1.01 (m, 28 H); ¹³C NMR (100 MHz, CDCl₃) δ = 170.7, 161.9 (t, J_{C-F} = 27.0 Hz), 119.0 (t, J_{C-F} = 257.7 Hz), 80.3 (J_{C-D} = 21.6 Hz), 57.2-57.0 (m), 43.6, 37.1 (t, J = 22.7 Hz), 35.7, 32.5, 30.9, 30.8, 29.0, 28.9, 27.3, 26.33, 26.32, 25.6, 25.5, 25.3, 25.20, 25.16, 24.8; ¹⁹F NMR (376 MHz, CDCl₃) δ = -97.8 (d, J = 280.9 Hz), -100.2 (d, J = 280.9 Hz); **HRMS** (ESI) calcd for C₂₄H₃₆DF₂NNaO₃ [M+Na⁺]: 449.2696, found: 449.2689.

Synthesis of 6f:



The reaction of HE (63.3 mg, 0.25 mmol), **1u** (67.6 mg, 0.2 mmol), **S8** (10.6 mg, 0.015 mmol), (PhO)₂PO₂H (5.0 mg, 0.02 mmol), toluene (0.75 mL), **2t** (11.6 mg, 0.1 mmol) and D₂O (0.25 mL) afforded **6f** (6.7 mg, 18%, 93% D) (eluent: petroleum ether/ethyl acetate = 20/1 to 15/1 to 7/1) as a faint yellow oil. Unreacted **2t** accounts for the rest of the mass balance. **6f**: 79:21 er (HPLC conditions: Chiralcel OJ-H column, hexane/*i*-PrOH = 99.7/0.3, 0.5 mL/min, λ = 214 nm, t_R (minor) = 40.6 min, t_R (major) = 46.5 min); $[\alpha]_p^{25}$ = -19.3 (c = 0.22, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 4.77-4.63 (m, 0.07 H), 4.07-3.93 (m, 1 H), 3.09-2.95 (m, 1 H), 2.68-2.30 (m, 6 H), 2.18-1.01 (m, 22 H); ¹³C NMR (100 MHz, CDCl₃) δ = 171.1, 161.8, 118.6 (t, J = 256.7 Hz), 57.3-57.0 (m), 41.1 (t, J = 23.2 Hz), 30.9, 29.1, 29.0, 28.2, 26.4, 25.60, 25.57, 25.2, 18.2; ¹⁹F NMR (376 MHz, CDCl₃) δ = -96.3 (d, J = 282.0 Hz), -99.1 (d, J = 282.0 Hz); HRMS (ESI) calcd for C₂₀H₃₁DF₂NO₃ [M+H⁺]: 373.2408, found: 373.2400.

2.4. Synthetic Applications

Synthesis of 7:



To a solution of **3a** (28.5 mg, 0.1 mmol) in CH₃CN (2 mL) was added HCl (2 M in water, 150 µL, 0.3 mmol) and pinacol (24.9 mg, 0.2 mmol). The reaction mixture was stirred at 30 °C for 1.5 h and then evaporated. The crude material was purified by flash column chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 5/1) to give pinacol boronate ester **7** (22.5 mg, 74%, 93% D) as a colorless oil: 93:7 er (HPLC conditions: Chiralcel OD-H column, hexane/i-PrOH = 80/20, 0.5 mL/min, λ = 214 nm, t_R (major) = 11.8 min, t_R (minor) = 13.2 min); $[\alpha]_D^{25}$ = -45.7 (c = 0.30, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.46-7.32 (m, 4 H), 7.24-7.10 (m, 1 H), 4.68-4.52 (m, 1.07 H), 4.06 (d, *J* = 8.8 Hz, 1 H), 1.41 (d, *J* = 16.4 Hz, 1 H), 1.34-1.15 (m, 12 H) 1.07 (d, *J* = 16.0 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 155.9, 136.4, 129.1, 125.4, 122.7, 83.8, 75.0, 68.6, 24.9, 24.8, 24.6; ¹¹B NMR (128 MHz, CDCl₃) δ = 32.9; HRMS (ESI) calcd for C₁₆H₂₁BDNNaO₄ [M+Na⁺]: 327.1597, found: 327.1601.

Synthesis of 8:



To a solution of **3a** (28.6 mg, 0.1 mmol) in CH₃CN (2 mL) was added selectfluor (107.5 mg, 0.3 mmol) at room temperature under nitrogen atmosphere. After addition, the reaction mixture was stirred at room temperature for 2 h and then evaporated. The crude material was purified by flash column chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 2/3) to give **8** (27.7 mg, 87%, 93% D) as a colorless oil: 93:7 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 65/35, 0.5 mL/min, λ = 214 nm, t_R (major) = 17.2 min, t_R (minor) = 19.4 min); $[\alpha]_D^{25}$ = -83.5 (*c* = 0.10, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.51-7.28 (m, 4 H), 7.19-7.03 (m, 1 H), 6.82 (s, 2 H), 4.74-4.49 (m, 1.07 H), 4.31 (d, *J* = 8.8 Hz, 1 H), 3.80 (s, 6 H), 1.03-0.71 (m, 1 H), 0.63-0.42 (m, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ = 156.3, 137.4, 128.8, 124.2, 121.9,

121.6, 69.6, 69.5, 54.7 (t, $J_{C-D} = 22.3 \text{ Hz}$), 36.3 (t, J = 4.8 Hz), 29.7; ¹¹**B NMR** (128 MHz, CDCl₃) $\delta = 5.26$; ¹⁹**F NMR** (376 MHz, CDCl₃) $\delta = -155.1$ (m), -157.2 (m); **HRMS** (ESI) calcd for $C_{15}H_{17}BDF_2N_3NaO_2$ [M+Na⁺]: 345.1415, found: 345.1419.

Synthesis of 9:



To a solution of **3a** (28.4 mg, 0.1 mmol) in EtOH (2 mL) was added NaOH (80.8 mg, 1 mmol), the reaction mixture was then heated to 80 °C for 10 h. After the reaction reached completion, EtOAc (5 mL) and H₂O (5 mL) were added. The organic layer was separated and the aqueous layer was extracted with EtOAc (5 mL x 3). The combined organic phase was dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 20/1) to afford **9** (15.9 mg, 61%, 92% D) as a colorless oil: 93:7 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 90/10, 0.5 mL/min, λ = 214 nm, t_R (major) = 83.4 min, t_R (minor) = 90.7 min); $[\alpha]_D^{25}$ = -6.8 (c = 0.53, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ = 7.18-7.04 (m, 2 H), 6.76 (s, 2 H), 6.68-6.57 (m, 3 H), 3.75 (d, J = 10.8 Hz, 1 H), 3.70-3.60 (m, 7 H), 3.31-2.33 (m, 2.08 H), 0.76-0.54 (m, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ = -29.5 (t, J = 67.8 Hz). HRMS analysis indicated that the BH₂ moiety of the product was also partially deuterated (see the first entry of the spectrum below).



Synthesis of 10:

To a solution of 4a (43.7 mg, 0.1 mmol) in dry CH₂Cl₂ (1 mL) was added BF₃•2AcOH (140 μ L, 1 mmol), the mixture was then heated to reflux for 5 h. The reaction mixture was cooled to ambient temperature, and poured into a saturated aqueous solution of sodium bicarbonate. The aqueous layer was extracted with EtOAc. The combined organic layer was dried over anhydrous Na_2SO_4 and concentrated. To a solution of the residue in THF-EtOH (1.2 mL, v/v = 1/1) was added KHCO₃ (10.5 mg, 0.1 mmol), KF (17.9 mg, 0.3 mmol), and a 30% aqueous solution of H_2O_2 (120 µL, 1 mmol). The mixture was heated to reflux for 12 h. When the reaction is complete as monitored by TLC, the mixture was poured into an aqueous solution of sodium bicarbonate. The aqueous layer was extracted with EtOAc (10 mL \times 3), and then the combined organic layer was washed with brine and dried over anhydrous Na₂SO₄. After filtration and evaporation, the residue was purified by chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 1/1) to afford **10** (11.7 mg, 60%, 90% D) as a white solid: 91:9 er (HPLC conditions: Chiralcel OD-H column, hexane/*i*-PrOH = 80/20, 0.5 mL/min, λ = 214 nm, t_R (major) = 18.1 min, $t_{\rm R}$ (minor) = 20.6 min); $[\alpha]_{\rm D}^{25}$ = -40.0 (c = 0.15, CHCl₃); m.p. 111-113 °C; ¹H NMR (400 MHz, CD₃OD) δ = 7.59-7.46 (m, 2 H), 7.45-7.34 (m, 2 H), 7.27-7.16 (m, 1 H), 4.66-4.49 (m, 1.10 H), 4.43 (d, J = 8.4 Hz, 1 H), 3.66 (d, J = 12.0 Hz, 1 H), 3.55 (d, J = 12.0 Hz, 1 H); ¹³C **NMR** (100 MHz, CD₃OD) δ = 158.7, 137.9, 130.1, 126.7, 124.0, 66.0, 60.3, 59.0 (t, J_{C-D} = 22.1 Hz); **HRMS** (ESI) calcd for $C_{10}H_{10}DNNaO_3$ [M+Na⁺]: 217.0694, found: 217.0696.

Synthesis of **11**:



Under nitrogen atmosphere, ^{*t*}BuOK (68.2 mg, 0.6 mmol), THF (2.0 mL), and ^{*t*}BuOOH (70% in water, 79.3 mg, 0.6 mmol) were added to a 10 mL flame-dried Schlenk flask at 0 °C. The mixture was stirred for 10 minutes, then a solution of **4a** (87.4 mg, 0.2 mmol) in 1 mL of THF

and 0.6 mL of TBAF (1.0 M in THF, 0.6 mmol) were added sequentially, the resulted mixture was stirred at 70 °C for 12 h. The reaction mixture was allowed to cool to room temperature before ether (5 mL) and H₂O (5 mL) were added. The organic layer was separated and the aqueous layer was extracted with ether (5 mL x 3). The combined organic phase was dried over anhydrous Na₂SO₄ and then concentrated under reduced pressure. The residue was purified by chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 5/1) to afford **11** (16.8 mg, 47%, 90% D) as a colorless oil : 91:9 er (HPLC conditions: Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, 1 mL/min, λ = 214 nm, $t_{\rm R}$ (major) = 10.9 min, $t_{\rm R}$ (minor) = 12.1 min); $[\alpha]_{\rm D}^{25}$ = -53.8 (*c* = 0.11, CHCl₃); ¹**H NMR** (400 MHz, CDCl₃) δ = 7.46-7.34 (m, 4 H), 7.23-7.15 (m, 1 H), 4.63-4.47 (m, 1.10 H), 4.02 (d, *J* = 8.4 Hz, 1 H), 1.33 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ = 155.7, 136.5, 129.1, 125.2, 122.0, 121.9, 68.5, 51.9 (t, $J_{\rm C-D}$ = 22.0 Hz), 18.3; **HRMS** (ESI) calcd for C₁₀H₁₀DNNaO₂ [M+Na⁺]: 201.0745, found: 201.0744.

Synthesis of 12:

To a solution of **4a** (43.7 mg, 0.1 mmol) in EtOH (1 mL) was added NaOH (41.2 mg, 1 mmol), the reaction mixture was then heated to reflux for 10 h. The reaction mixture was allowed to cool to room temperature, EtOAc (5 mL) and H₂O (5 mL) were added. The organic layer was separated and the aqueous layer was extracted with EtOAc (5 mL x 3). The combined organic phase was dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by chromatography on silica gel (eluent: petroleum ether/ethyl acetate = 5/1) to afford **12** (27.5 mg, 67%, 90% D) as a colorless oil: 92:8 er (HPLC conditions: Chiralcel OJ-H column, hexane/*i*-PrOH = 85/15, 1 mL/min, λ = 214 nm, t_R (minor) = 9.3 min, t_R (major) = 14.3 min); $[\alpha]_D^{25}$ = +9.3 (c = 0.23, CHCl₃); ¹**H NMR** (400 MHz, CDCl₃) δ = 7.55-7.44 (m, 6 H),7.43-7.24 (m, 9 H), 7.06 (t, J = 8.0 Hz, 2 H), 6.69 (t, J = 7.4 Hz, 1 H), 6.29 (d, J = 7.6 Hz, 2 H), 3.73-3.66 (m, 0.1 H), 3.61 (d, J = 11.2 Hz, 1 H)), 3.43 (d, J = 10.8 Hz, 1 H), 3.30-2.42 (m, 2 H), 1.87-1.16 (m, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ = 145.9, 135.6, 134.3, 129.6, 129.2, 128.0, 118.3, 114.1, 65.8, 16.7; **HRMS** (ESI) calcd for C₂₇H₂₇DNOSi [M+H⁺]: 411.1997, found: 411.2002.

2.5. Radical Trap Experiment



To an oven-dried 16 x 60 mm vial containing a dry Teflon stir bar were charged with photocatalyst 4DPAIPN (1.0 mg, 0.001 mmol), thiol catalyst **S5** (8.5 mg, 0.015 mmol) and NHC–BH₃ **1a** (21.8 mg, 0.2 mmol). After sequential addition of dry toluene (0.75 mL), D₂O (0.25 mL), and olefin **2a** (17.5 mg, 0.1 mmol), the reaction mixture was flushed with nitrogen gas for two minutes and then the vial was sealed with a cap and parafilm. The vial was placed in a cooling station and a 30 W blue LED was then placed on the top of the cooling station, which is connected to a chiller to maintain the temperature of the cooling water at 10 °C (**Supplementary Figure 5**). After stirring with irradiation for 24 hours, a small portion of the reaction mixture was diluted with MeCN and filtered. The filtrate was subjected to high resolution mass spectra (HRMS) analysis. Product **3a** was not detected, instead **13** was observed: HRMS (ESI) calculated for **13** C₁₄H₂₉BN₃O [M+H⁺]: 266.2398, found: 266.2402 (**Supplementary Figure 6**).



Supplementary Figure 6. The HRMS spectrum of the crude reaction mixture.

2.6. UV-Vis Absorption Spectra of 1a, 2a, and S5

UV-Vis absorption spectra of **1a**, **2a**, **S5** and their combinations were measured (in toluene, **Supplementary Figure 7-10**). None of the individual reaction components can absorb in the visible region and none of combinations showed obvious bathochromic shift, excluding the possibility of forming EDA complex under the reaction conditions.



Supplementary Figure 7. UV-Vis spectra of 1a, 2a, and their mixture.



Supplementary Figure 8. UV-Vis spectra of 1a, S5, and their mixture.



Supplementary Figure 9. UV-Vis spectra of 2a, S5, and their mixture.



Supplementary Figure 10. UV-Vis spectra of 1a, 2a, S5, and their mixture.

2.7. Cyclic Voltammetry Measurements

Cyclic voltammetry (CV) experiments of **2a**, **S5**, and 4DPAIPN were recorded on a CHI 600E electrochemical workstation. Electrolyte solution was prepared by dissolving the substrates (0.05 mmol, 5 mM) and tetrabutylammonium hexafluorophosphate (387.4 mg, 1 mmol, 100 mM) in MeCN (10 mL) and bubbling with nitrogen for two minutes. Measurements were performed in a 3-compartment electrochemical cell, in which glassy carbon electrode (GCE) was used as a working electrode, silver-silver chloride (Ag/AgCl) in saturated KCl as the reference electrode, and Pt wire as the counter electrode. The scan rate was set at 100 mV/s.



Supplementary Figure 11. CV curve of 2a.



Supplementary Figure 12. CV curve of S5.



Supplementary Figure 13. CV curve of S5 in the presence of H₂O.



Supplementary Figure 14. CV curve of 4DPAIPN in the presence of H₂O.

2.8. Stern-Volmer Experiments

Stern-Volmer luminescence quenching experiments were carried out with freshly prepared solutions of 4DPAIPN (10⁻⁵ M) at room temperature. For **1a** and **2a**, the solutions were prepared in toluene and was irradiated at 350 nm and the luminescence were measured at 523 nm. For **S5**, the solution was prepared in DMF due to its poor solubility in toluene and was irradiated at 350 nm and the luminescence were measured at 530 nm. For each sample, the luminescence was acquired three times and averaged (**Supplementary Tables 3-6**), the averages of the results were used for the graphical representation (**Supplementary Figures 15-18**).

	Species		Co	ncentration (mM	()
2	4DPAIPN		0.01		
	1 a			varied	
[Sub] (mM)	Scan 1	Scan 2	Scan 3	Average	I ₀ / I
0	1352	1351	1351	1351	1.000
5	1341	1342	1342	1342	1.007
10	1353	1353	1355	1354	0.998
15	1370	1371	1370	1370	0.986
20	1394	1393	1394	1394	0.969
25	1368	1368	1367	1368	0.988

Supplementary Table 3. Fluorescence quenching data with solutions of 4DPAIPN and 1a.



Supplementary Figure 15. Stern-Volmer plot of 4DPAIPN quenching with varying concentration of **1a**.

	Species		Co	ncentration (mN	(1)
2	4DPAIPN		0.01		
	2a			varied	
[Sub] (mM)	Scan 1	Scan 2	Scan 3	Average	I ₀ /I
0	1266	1266	1267	1266	1.000
5	1259	1258	1256	1258	1.006
10	1261	1251	1237	1250	1.013
15	1216	1212	1207	1212	1.045
20	1221	1220	1219	1220	1.038
25	1195	1196	1196	1196	1.059

Supplementary Table 4. Fluorescence quenching data with solutions of 4DPAIPN and 2a.



Supplementary Figure 16. Stern-Volmer plot of 4DPAIPN quenching with varying concentration of **2a**.

Species		Co	ncentration (mM	()	
	IDPAIPN		0.01		
	S 5			varied	
[Sub] (mM)	Scan 1	Scan 2	Scan 3	Average	I ₀ / I
0	1020	1021	1021	1021	1.000
5	946	943	948	946	1.079
10	874	874	874	874	1.168
15	818	819	818	818	1.248
20	764	766	767	766	1.333
25	723	723	723	723	1.412

Supplementary Table 5. Fluorescence quenching data with solutions of 4DPAIPN and S5.



Supplementary Figure 17. Stern-Volmer plot of 4DPAIPN quenching with varying concentration of **S5**.

	Species		Co	ncentration (mM	()
	4DPAIPN			0.01	
	D_2O			5000	
	S5			varied	
[Sub] (mM)	Scan 1	Scan 2	Scan 3	Average	I ₀ / I
0	1153	1156	1157	1155	1.000
1	1035	1033	1035	1034	1.117
2	969	969	969	969	1.192
3	913	913	914	913	1.265
4	868	868	868	868	1.331
5	843	843	843	843	1.370

Supplementary Table 6. Fluorescence quenching data with solutions of 4DPAIPN, and **S5** in the presence of D_2O (5.0 M).



Supplementary Figure 18. Stern-Volmer plot of 4DPAIPN quenching with varying concentration of S5 in the presence of D_2O (5.0 M).



Supplementary Figure 19. Summary of Stern-Volmer plots.

Supplementary Table 7. The influence of D₂O on reaction rate.



n.d. : Not Determined

We briefly investigated the initial product formation rates for reactions with or without D_2O and found that the reaction with D_2O was much faster than the reaction without D_2O as shown above.

2.9. Quantum Yield Measurements

2.9.1. Emission spectrum of light source

Emission spectrum of the blue LED (30 W) was recorded on a Steady-State & Time-Resolved Fluorescence Spectrofluorometer (**Supplementary Figure 20**). According to the spectrum, emission range of light source is 400–500 nm, and the maximum emission wavelength is 441 nm.



Supplementary Figure 20. Emission spectrum of the blue LED.

Quantum yield measurements were determined using standard ferrioxalate chemical actinometry as described by Yoon,¹⁸ Ritter,¹⁹ Alem án,²⁰ and Glorius.²¹ In this part, we used a 30 W blue LED ($\lambda = 441$ nm) as light source to determine the quantum yield.

2.9.2. Determination of the photon flux at $\lambda = 441$ nm:

The photon flux of the 30 W blue LED was determined by monitoring the photoreduction of Fe(III) in potassium ferrioxalate to Fe(II), upon complexation with 1,10-phenanthroline:



The following solutions were prepared:

a. Actinometer solution: 589.5 mg (1.2 mmol) of potassium ferrioxalate trihydrate and 278 μ L of H₂SO₄ 96% were added to a 100 mL volumetric flask and filled to the mark with Nanopure water.

b. Phenanthroline-buffer solution: 100 mg (0.55 mmol) of 1, 1,10-phenanthroline, 9.88 g (120.4 mmol) of sodium acetate and 2.0 mL of H_2SO_4 96% were added to a 100 mL volumetric flask and filled to the mark with Nanopure water.

1 mL of the actinometer solution was added to a vial (16×60 mm) and was irradiated by the 30 W blue LED for 4 seconds. After the irradiation, the mixture was quantitatively transferred to a 5 mL volumetric flask containing 1.0 mL of the phenanthroline-buffer solution. Then, the flask was filled to the mark with Nanopure water, wrapped up with aluminum foil, and was left in the dark for 1 h to ensure the quantitative formation of Fe^{II}(phen)₃²⁺ complex. This procedure was repeated one more time by changing the irradiation time to 6 seconds. Additionally, the experiment of a control sample was carried out under dark, following the same sample treatment.

The absorbance of each solution at $\lambda = 510$ nm was measured using a Perkin-Elmer Lambda 35 UV-Vis spectrophotometer, establishing the blank with Nanopure water. For each sample, the absorbance was acquired three times and averaged. According to Lambert-Beer law, the moles of Fe(II) in each sample are related to the absorbance:

$$n\left[Fe^{2+}\right] = \frac{V \cdot \Delta A}{l \cdot \varepsilon}$$

Where:

- ΔA is the absorbance difference between irradiated sample and non-irradiated sample.
- *V* is the volume (in L) of the measurement sample (5 mL).
- ε is the extinction coefficient of the complex Fe^{II}(phen)₃²⁺ at $\lambda = 510$ nm (11100 L mol⁻¹ cm⁻¹)
- l is the optical path of the sample in the spectrophotometer (1 cm).

The photon flux can be calculated using the following equation:

$$photon flux = \frac{n [Fe^{2+}]}{\phi \cdot t \cdot f}$$

Where:

- Φ is the quantum yield for the photoreduction of ferrioxalate at $\lambda = 441$ nm, which is $1.11.^{22,23}$
- t is the reaction time (4 s or 6 s).
- *f* is the fraction of light absorbed, and is calculated as $f = 1 10^{-A_{441}nm}$

where $A_{441 \text{ nm}}$ is the absorbance of the actinometer solution at $\lambda = 441$ nm, which is 0.3877 according to the UV-Vis spectra of actinometer solution (**Supplementary** Figure 21).



Supplementary Figure 21. UV-Vis spectra of actinometer solution (12 mM in aqueous solution).

The photon flux of the 30 W blue LED ($\lambda = 441$ nm) was thus determined to be 2.65*10⁻⁷ einsten s⁻¹ (**Supplementary Table 8**).

scan time	UV-Vis absorption at 510 nm			
	A_{4s}	A _{6s}	A _{dark}	
1	1.480	2.195	0.016	
2	1.644	2.452	0.019	
Average	1.562	2.324	0.017	
$\Delta A (=A-A_{dark})$	1.545	2.306	-	
n [Fe ²⁺] (mol)	6.96*10 ⁻⁷	1.04*10 ⁻⁶		
Photon flux	2.654*10 ⁻⁷	2.642*10 ⁻⁷		
(einstein/s)	(average) 2.648*10 ⁻⁷			

Supplementary Table 8. UV-Vis absorption data and calculated results of photo flux.

2.9.3. Determination of quantum yield at $\lambda = 441$ nm:

Once we have determined the photon flux of the 30 W blue LED ($\lambda = 441$ nm), the same equation was employed for the determination of the quantum yield of our photoreaction system. For that, the moles of product for a given time was determined.



Following the **Typical Procedure I**, the reaction of **1a** and **2a** was carried out under the standard conditions and was stopped after 2 h of irradiation. The solvent of reaction mixture was

evaporated in vacuo. The amount of product **3a** was determined to be $1.6*10^{-5}$ mol by ¹H NMR analysis of the crude reaction mixture using CH₂Br₂ as the internal standard. Another reaction with an irradiation time of 4 h was also carried out and the amount of **3a** was determined to be $2.6*10^{-5}$ mol.

$$\Phi' = \frac{n [prod.]}{photon flux \cdot t' \cdot f'}$$

Where:

- n [prod.] is the amount of **3a** (in mol) that has been formed during the irradiation time.
- t'(s) is the irradiation time (in seconds).
- f' is the fraction of light absorbed, and is calculated as $f'=1-10^{-A_{441\,nm}}$

where $A_{441 \text{ nm}}$ is the absorbance of PC solution at 441 nm, and was determined to be 2.5939 according to the UV-Vis spectra of 1 mM solution of 4DPAIPN in toluene (**Supplementary Figure 22**).



Supplementary Figure 22. UV-Vis spectra of 4DPAIPN solution (1 mM in toluene)

The quantum yield of the reaction was thus determined to be 0.0076 (average of two runs, **Supplementary Table 9**).

entry	reaction time/s	NMR yield/%	n[prod.]/mol	quantum yield (Φ')
1	7200	16	1.6*10 ⁻⁵	0.0084
2	14400	26	2.6*10 ⁻⁵	0.0068
$\Phi' = (0.0084 + 0.0068)/2 = 0.76\%$				

Supplementary Table 9. Quantum yield in the presence of D₂O.

We also measured the quantum yield of the reaction without D_2O , which is homogenous but also gave a very low value ($\Phi' = 0.28\%$, see the **Supplementary Table 10** below), indicating that the low quantum yield is largely related to the slow reaction rate.



Supplementary Table 10. Quantum yield in the absence of D₂O.

entry	reaction time/s	NMR yield/%	n[prod.]/mol	quantum yield (Φ')
1	14400	10	1.0*10 ⁻⁵	0.0026
2	28800	22	2.2*10 ⁻⁵	0.0029

 $\Phi' = (0.0026 + 0.0029)/2 = 0.28\%$

2.10. X-Ray Crystallographic Data of Compounds 3c, 4a, and 5a

2.10.1. X-Ray Crystallographic Data of Compound 3c



Supplementary Figure 23. X-Ray Structure of Compound **3c** (CCDC 2143290). Crystals suitable for X-ray structure analysis were obtained via recrystallization from tetrahydrofuran and pentane.

Supplementary Table 11. Crystal data and structure refinement for 3c.

3c		
$C_{19}H_{28}BN_3O_2$		
341.25		
173(2) K		
1.54178 A		
Monoclinic, P2(1)		
a = 5.8190(2) A alpha = 90 deg.		
b = 15.2860(4) A beta = 94.197(2) deg.		
gamma = 90 deg.		
1902.36(9) A^3		
4, 1.191 Mg/m^3		
0.609 mm^-1		
736		
0.180 x 0.160 x 0.140 mm		
3.554 to 68.137 deg.		
-6<=h<=6, -18<=k<=18, -25<=l<=25		
26076 / 6870 [R(int) = 0.0539]		
99.9 %		
Semi-empirical from equivalents		
Full-matrix least-squares on F^2		

Data / restraints / parameters	6870 / 1 / 471
Goodness-of-fit on F^2	1.049
Final R indices [I>2sigma(I)]	R1 = 0.0423, wR2 = 0.1063
R indices (all data)	R1 = 0.0484, wR2 = 0.1098
Absolute structure parameter	-0.12(11)
Extinction coefficient	n/a
Largest diff. peak and hole	0.193 and -0.188 e.A^-3

2.10.2 X-Ray Crystallographic Data of Compound 4a



Supplementary Figure 24. X-Ray Structure of Compound **4a** (CCDC 2106905). Crystals suitable for X-ray structure analysis were obtained via recrystallization from tetrahydrofuran and pentane.

Supplementary Table 12. Crystal data and structure refinement for 4a.

Identification code	4a
Empirical formula	$C_{28}H_{25}NO_2Si$
Formula weight	435.58
Temperature	299(2) K
Wavelength	1.54178 A
Crystal system, space group	Trigonal, P3(2)
Unit cell dimensions	a = 9.6154(4) A alpha = 90 deg.
	b = 9.6154(4) A beta = 90 deg.
	c = 44.006(3) A gamma = 120 deg.
Volume	3523.5(4) A^3

Z, Calculated density	6, 1.232 Mg/m^3
Absorption coefficient	1.070 mm^-1
F(000)	1380
Crystal size	0.200 x 0.200 x 0.200 mm
Theta range for data collection	3.012 to 68.567 deg.
Limiting indices	-11<=h<=11, -10<=k<=11, -51<=l<=49
Reflections collected / unique	29000 / 8470 [R(int) = 0.0795]
Completeness to theta $= 67.679$	99.8 %
Absorption correction	Semi-empirical from equivalents
Refinement method	Full-matrix least-squares on F^2
Data / restraints / parameters	8470 / 3 / 577
Goodness-of-fit on F^2	1.011
Final R indices [I>2sigma(I)]	R1 = 0.0470, wR2 = 0.1279
R indices (all data)	R1 = 0.0632, $wR2 = 0.1376$
Absolute structure parameter	0.031(16)
Extinction coefficient	n/a
Largest diff. peak and hole	0.328 and -0.218 e. A^-3

2.10.3 X-Ray Crystallographic Data of Compound 5a



Supplementary Figure 25. X-Ray Structure of Compound **5a** (CCDC 2106903). Crystals suitable for X-ray structure analysis were obtained via recrystallization from methanol and pentane.

Supplementary Table 13. Crystal data and structure refinement for 5a.

Identification code	5a
Empirical formula	$C_{20}H_{26}NO_3P$
Formula weight	359.39
Temperature	295(2) K
Wavelength	1.54178 A
Crystal system, space group	Hexagonal, P6(5)
Unit cell dimensions	a = 10.174(2) A alpha = 90.00(3) deg.
	b = 10.174(2) A beta = 90.00(3) deg.
	c = 34.443(7) A gamma = 120.00(3) deg.
Volume	3087.7(14) A^3
Z, Calculated density	6, 1.160 Mg/m^3
Absorption coefficient	1.318 mm^-1
F(000)	1152
Crystal size	0.200 x 0.200 x 0.200 mm
Theta range for data collection	5.019 to 68.322 deg.
Limiting indices	-12<=h<=12, -11<=k<=12, -41<=l<=41
Reflections collected / unique	32039 / 3772 [R(int) = 0.0447]
Completeness to theta $= 67.679$	99.9 %
Refinement method	Full-matrix least-squares on F^2
Data / restraints / parameters	3772 / 1 / 240
Goodness-of-fit on F^2	1.028
Final R indices [I>2sigma(I)]	R1 = 0.0329, wR2 = 0.0851
R indices (all data)	R1 = 0.0359, wR2 = 0.0884
Absolute structure parameter	0.038(8)
Extinction coefficient	n/a
Largest diff. peak and hole	0.128 and -0.119 e. A^-3

2.11. Computational Details

2.11.1. General Information

DFT calculations were performed in Gaussian 16 (C.01) program package.²⁴ All the structures were optimized using the ω B97xD functional²⁵ and the 6-31+G(d,p) basis set. Frequency calculations were carried out at that level of theory to analyze the nature of stationary points as transition states (one imaginary frequency) or minima (no imaginary frequencies). The Goodvibes program²⁶ was used to obtain the thermochemistry corrections at 283.15K, including quasi-harmonic corrections with the method proposed by Grimme,²⁷ with the cut-off value set as 100 wavenumbers. To further refine the potential energies, single point energy calculations were done with a larger basis set (6-311++G(3d,2p)) and PBE0²⁸ as the functional. Benchmarking calculations using M06-2X and ω B97xD were also done to further confirm the robustness of the method, showing similar performance for this reaction. The ω B97xD functional was selected for optimizations due to its general good performance in geometry optimization.²⁹ Implicit solvation was included in all the calculations, including geometry optimization-frequency calculations and single points, using the CPCM implicit solvent model^{30,31} and toluene as the solvent.

Conformation of the peptide catalyst was chosen based on the structural analysis of similar tetrapeptide catalysts by X-ray and DFT calculations reported by Miller and co-workers.³² Thiol **S6**, which showed the same enantioselectivity as thiol **S5**, was thus chosen as the catalyst for our calculation.

NCI analyses were performed using the NCIplot program^{33,34} and all the 3D representations were prepared in PyMOL.
2.11.2. Benchmarking of the computational method

Supplementary Table 14. Comparison of different DFT methods on the **TS**-*Re* vs **TS**-*Si* activation energy difference and calculated enantiomeric ratio. All energies in kcal/mol.

Method	$\Delta\Delta G_{\text{Re-Si}}^{\ddagger}$	er
PBE0	1.24	90:10
M062X	2.15	98:2
ωB97xD	2.14	98:2
Exp.	1.45	93:7

2.12 NMR Spectra and HPLC Spectra

¹H NMR (400 MHz, room temperature, CDCl₃)



100 90 fl (ppm) Supplementary Figure 27. ¹³C NMR spectrum of compound S4





Supplementary Figure 29. ¹³C NMR spectrum of compound S5



Supplementary Figure 31. ¹³C NMR spectrum of compound S6



Supplementary Figure 32. ¹H NMR spectrum of compound S8

¹³C NMR (100 MHz, room temperature, CDCl₃)

7.317 7.102 5.188 5.188 5.188 5.188 4.777 4.676 4.574 4.575 4.575 7.665	2,436 2,436 2,379 2,379 2,379 2,334 2,334 2,334 6,050 6,050 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 6,020 7,020 6,020 6,020 6,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,000 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,020 7,000 7,0000 7,0000000000	55.619 55.619 55.619 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.600 55.500 55.600 55.600 55.500 55.500 55.500 55.500 55.500 55.500 55.500 55.500 55.500 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.5000 55.50000 55.50000 55.50000 55.50000 55.50000 55.50000 55.500000000	081 265 076 823 178 029
0000000000000	0000000000000000	NU	-0000
22222222222			~~ @ @ @ @



Supplementary Figure 33. ¹³C NMR spectrum of compound S8



 $_{e7}$ - $_{e9}$ - $_{91}$ - $_{93}$ - $_{95}$ - $_{97}$ - $_{99}$ - $_{101}$ - $_{103}$ - $_{105}$ - $_{107}$ - $_{109}$ - $_{111}$ - $_{113}$ - $_{115}$ - $_{117}$ - $_{119}$ - $_{121}$ Supplementary Figure 34. ¹⁹F NMR spectrum of compound S8





Supplementary Figure 35. ¹H NMR spectrum of compound S9





Supplementary Figure 36. ¹³C NMR spectrum of compound S9







Supplementary Figure 40. ¹³C NMR spectrum of compound S11

.590



Supplementary Figure 42. ¹³C NMR spectrum of compound S12



Supplementary Figure 44. ¹³C NMR spectrum of compound S13

$^{1}H NMR (400 MHz, room temperature, CDCl_{3})$ 1.6441.6221.4891.4801.4711.4711.4720.9600.9600.9380.9270.9190.919939 666 958 HS . NHBoc ни Me S14 1.00-0.96 1.12 1.06-2.23-2.05 -.284 .10-3.01,2.95-1.03-.10 9.84 1.37 0.9 5.5 5.0 4.5 4.0 f1 (ppm) 7.5 8.5 8.0 7.0 3.0 2.0 1.5 9.0 6.5 6.0 3.5 2.5 1.0 0.5 0.0 -0. Supplementary Figure 45. ¹H NMR spectrum of compound S14 ¹³C NMR (100 MHz, room temperature, CDCl₃) 172.576 170.700 170.659 ~128.201 -155,836-144.371.34221 .29.189 .28.180 .25.375 .25.375 .22.345 .22.545 .16.638 80.661 77.318 77.000 77.000 76.681 -61.546 -54377 49.140 -41.829 HS O NHBoc HN Me S14

100 90 fl (ppm) 190 180 170 160 150 140 110 130 120 70 80 60 50 40 30 20 10 0 -10 Supplementary Figure 46. ¹³C NMR spectrum of compound S14



Supplementary Figure 48. ¹H NMR spectrum of compound 3a



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

Supplementary Figure 49. ¹³C NMR spectrum of compound 3a

¹¹B NMR (128 MHz, room temperature, CDCl₃)



Supplementary Figure 50. ¹¹B NMR spectrum of compound 3a

HPLC spectrum of racemic 3a

Vial#	: 11
Data File	: SQL-6-280-2-RACEMIC(1).lcd
Method File	: 40D-H-60-0.5-214.lcm
Date Acquired	: 12/9/2021 10:34:07 AM
Date Processed	: 1/25/2022 12:35:14 AM

<Chromatogram View>



Delector	A 214000			
Pesk #	Ret. Time	Height	Area	Area%
1	12.911	131195	2261994	49.950
2	14.695	115749	2266525	50.050
Total		246943	4528519	100.000

Supplementary Figure 51. HPLC spectrum of racemic 3a

HPLC spectrum of 3a

Sample Name Tray# Vial# Data File Method File Date Acquired Date Processed	: 14 : sql-6-354-CHIRAL(1).lcd : 40D-H-60-0.5-214.lcm : 12/31/2021 7:17:22 PM : 1/25/2022 1:34:49 AM
Date Processed	: 1/25/2022 12:34:49 AM

<Chromatogram View>



Supplementary Figure 52. HPLC spectrum of 3a



Supplementary Figure 54. ¹³C NMR spectrum of compound 3b



Vial#	: 5
Data File	: SQL-6-287-1-RAC-(3).lcd
Method File	: 40D-H-65-0.5-214.lcm
Date Acquired	: 12/11/2021 7:13:52 PM
Date Processed	: 1/25/2022 12:40:35 AM

<Chromatogram View>

SQL-6-287-1-RAC-(3).lcd



Supplementary Figure 56. HPLC spectrum of racemic 3b



Supplementary Figure 58. ¹H NMR spectrum of compound 3c



Supplementary Figure 60. ¹¹B NMR spectrum of compound 3c

HPLC spectrum of racemic 3c

:
:1
: 11
: sql-6-348-1-RAC-(4).lcd
: 40D-H-75-0.5-214.lcm
: 1/3/2022 3:28:10 PM
: 1/25/2022 12:42:12 AM

<Chromatogram View>



Supplementary Figure 61. HPLC spectrum of racemic 3c



<Chromatogram View>

sql-6-348-2-CHIRAL-(4).Icd m∨ 500 400-11.717 300-200-100-13.066 3c 0-2.5 5.0 7.5 10.0 0.0 12.5 min <Data Analysis> Height 3177 Area 4961 Area% 93.684 33448 529579 2019 33798 6.316

Supplementary Figure 62. HPLC spectrum of 3c



Supplementary Figure 64. ¹³C NMR spectrum of compound 3d





90 80 50 40 70 60 20 30 10 0 f1 (ppm) -10 -20 -30 -40 -50 -60 -70 -80 -90

Supplementary Figure 65. ¹¹B NMR spectrum of compound 3d

HPLC spectrum of racemic 3d

:1
: 10
: sql-7-74-1.lcd
: 4OD-H-85-0.5-214.lcm
: 2/21/2022 7:56:42 PM
: 2/21/2022 8:27:39 PM

<Chromatogram View>

sql-7-74-1.lcd m٧ 300 16.206 20.158 200-100-Ċ١ racemic 3d 0 0.0 2.5 5.0 7.5 10.0 12.5 15.0 17.5 20.0 min <Data Analysis> Detector A 214nm Pesk # | Ret. Tin 49.623 50.377 100.000 17804 Tota

Supplementary Figure 66. HPLC spectrum of racemic 3d











Supplementary Figure 70. ¹¹B NMR spectrum of compound 3e

HPLC spectrum of racemic 3e

1
:1
: 12
: sql-7-73-1-1.lcd
: 4OD-H-80-0.5-214.lcm
: 2/22/2022 2:17:55 PM
: 2/22/2022 4:32:32 PM

<Chromatogram View>



Delector	A 2140m			
Pesk #	Ret. Time	Height	Area	Area%
1	26.069	254307	8974856	50.360
2	33.465	201219	8846546	49.640
Total		455526	17821402	100.000

Supplementary Figure 71. HPLC spectrum of racemic 3e

HPLC spectrum of 3e

1	
Sample Name	
Tray#	:1
Vial#	: 20
Data File	: sql-7-73-2-2.lcd
Method File	: 4OD-H-80-0.5-214.lcm
Date Acquired	: 2/21/2022 11:47:26 PM
Date Processed	: 2/22/2022 9:11:31 AM

<Chromatogram View>



Supplementary Figure 72. HPLC spectrum of 3e



Supplementary Figure 74. ¹³C NMR spectrum of compound 3f





Supplementary Figure 76. HPLC spectrum of racemic 3f



Supplementary Figure 78. ¹H NMR spectrum of compound 3g









10 0 -10 f1 (ppm) 80 90 70 60 50 40 30 20 -20 -30 -40 -50 -60 -70 -80 -90 Supplementary Figure 80. ¹¹B NMR spectrum of compound 3g

HPLC spectrum of racemic **3g**

:1
: 11
: sql-6-360-1-4-CHIRAL-(5).lcd
: 4OD-H-70-0.5-214.lcm
: 1/7/2022 4:50:51 PM
: 1/25/2022 12:43:05 AM

<Chromatogram View>



Supplementary Figure 81. HPLC spectrum of racemic 3g

HPLC spectrum of 3g

:1
: 10
: sql-6-360-2-1-CHIRAL-(5).lcd
: 4OD-H-70-0.5-214.lcm
: 1/7/2022 5:16:57 PM
: 1/25/2022 12:43:42 AM

<Chromatogram View>

sql-6-360-2-1-CHIRAL-(5).lcd m٧ 1000 18.622 750-500-Ph 250-3g 20.814 0 5.0 10.0 12.5 15.0 17.5 2.5 7.5 0.0 20.0 min <Data Analysis> Detector Pesk # <u>14nm</u> et. Time leigh <u>rea%</u> 90.293 211405 2177773 9.707 800

Supplementary Figure 82. HPLC spectrum of 3g



Supplementary Figure 84. ¹³C NMR spectrum of compound 3h



-30.174 -30.825 -31.488



Supplementary Figure 86. HPLC spectrum of racemic 3h



Vial#	10
Data File	: sql-6-353-2-(3N)-CHIRAL-(2).lcd
Method File	: 40D-H-60-0.5-214.lcm
Date Acquired	: 1/4/2022 11:34:10 PM
Date Processed	: 1/25/2022 12:37:49 AM

<Chromatogram View>



Supplementary Figure 88. ¹H NMR spectrum of compound 3i





Supplementary Figure 90. ¹¹B NMR spectrum of compound 3i



<Chromatogram View>



Supplementary Figure 91. HPLC spectrum of racemic 3i

HPLC spectrum of 3i

-	
Sample Name	:
Tray#	:1
Vial#	: 47
Data File	: XMC-4-327b-1.lcd
Method File	: 3AD-H-85-1-214.lcm
Date Acquired	: 2/11/2022 2:54:07 PM
Date Processed	2/11/2022 3:53:58 PM

<Chromatogram View>

XMC-4-327b-1.lcd m∨ 300 15.229 200 100-14.095 3i ^tBu 0 7.5 2.5 10.0 0.0 5.0 12.5 15.0 min <Data Analysis> <u>214nm</u> Ret. Time 14.09 Detector A Pesk # Height 35648 Area% 11.051 88.949 100.000 Area 886744 71376 80243

Supplementary Figure 92. HPLC spectrum of 3i



Supplementary Figure 94. ¹³C NMR spectrum of compound 3j
¹¹B NMR (128 MHz, room temperature, CDCl₃)



Supplementary Figure 96. HPLC spectrum of racemic 3j



Supplementary Figure 98. ¹H NMR spectrum of compound 3k



Supplementary Figure 100. ¹¹B NMR spectrum of compound 3k

HPLC spectrum of racemic 3k

1	
Sample Name	
Sample Name	-
Trav#	- 1
ii ay n	
Vial#	· 28
Vicin	. 20
Data File	: sal-6-382-1-4-(10).lcd
Mathad File	0 A D 11 00 0 5 04 4 400 MINU am
Method File	. 3AD-H-90-0.5-214-100MIN.ICM
Data Acquired	1/20/2022 2:51:06 DM
Date Acquired	. 1/20/2022 2.01.00 FW
Date Processed	· 1/25/2022 1:07:36 AM
Date FIOLESSEU	. 1/20/2022 1.04.00 AM

<Chromatogram View>



1 030 1	Ret. Hille	ricigitt	Alca	/ ucu/u
1	101.893	126566	17323282	50.030
2	108.284	115079	17302226	49.970
Total		241645	34625508	100.000

Supplementary Figure 101. HPLC spectrum of racemic 3k

HPLC spectrum of **3k**

Tray#	:1
Vial#	: 29
Data File	: sql-7-4-(10).lcd
Method File	: 3AD-H-90-0.5-214-100MIN.Icm
Date Acquired	: 1/20/2022 4:52:17 PM
Date Processed	: 1/25/2022 1:04:53 AM

<Chromatogram View>



Supplementary Figure 102. HPLC spectrum of 3k



272 114 114 1097 097 997 737 737	5573 573 5608 5608 0073 0073 970 970 970 970 970 970 977 777 777 777
NNNNNNNN 0000	<u>«««««««««««««««««««««««««»»»»»»»»»»»»</u>



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

Supplementary Figure 104. ¹³C NMR spectrum of compound 31

¹¹B NMR (128 MHz, room temperature, CDCl₃)



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 **Supplementary Figure 106**. ¹⁹F NMR spectrum of compound **3**I



Supplementary Figure 107. HPLC spectrum of racemic 31

HPLC spectrum of 31

Tray#	:1
Vial#	:1
Data File	: sql-7-29-2-1.lcd
Method File	: 40D-H-80-1.0-214.lcm
Date Acquired	: 1/25/2022 5:53:25 AM
Date Processed	: 1/25/2022 8:22:21 AM

<Chromatogram View>



Supplementary Figure 108. HPLC spectrum of 31



Supplementary Figure 110. ¹³C NMR spectrum of compound 3m

¹¹B NMR (128 MHz, room temperature, CDCl₃)





Area 104143

10347

20761868

5.0

racemic 3m

2.5

Heid

<u>214nm</u> Ret. Time 10.717

15.410

ÒМе

7.5

250-

0-

Detector A Pesk #

Tot

0.0

10.0

<Data Analysis>

Area% 50.161 49.839 100.000 12.5

15.0

17.5

min



<Chromatogram View>



Supplementary Figure 113. HPLC spectrum of 3m





Supplementary Figure 114. ¹H NMR spectrum of compound 3n



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

Supplementary Figure 115. ¹³C NMR spectrum of compound **3n** ¹¹B NMR (128 MHz, room temperature, CDCl₃)





Supplementary Figure 116. ¹¹B NMR spectrum of compound 3n



Supplementary Figure 117. HPLC spectrum of racemic 3n

HPLC spectrum of 3n

Tray#	:1
Viaĺ#	: 12
Data File	: sql-6-357-1.lcd
Method File	: 3AD-H-75-0.5-214-50MIN.lcm
Date Acquired	: 2/2/2022 11:57:23 AM
Date Processed	: 2/2/2022 12:22:19 PM

<Chromatogram View>

sql-6-357-1.lcd m∨ 896 5 500-250-3n 19.137 0 22.5 2.5 20.0 5.0 7.5 12.5 10.0 15.0 17.5 0.0 min <Data Analysis> <u>.214nm</u> Ret. Time 19.137 Detector A Pesk # Height 3426 Area 96216 Area% 1570100 511 545 21.89 94 100 2 Tota

Supplementary Figure 118. HPLC spectrum of 3n



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 Supplementary Figure 120. ¹³C NMR spectrum of compound **30** ¹¹B NMR (128 MHz, room temperature, CDCl₃)



^{10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210} **Supplementary Figure 122**. ¹⁹F NMR spectrum of compound **30**

HPLC spectrum of racemic **30** Tray# :1 Vial# :10 Data File : sql-6-398-1-(12).lcd Method File : 30-H-75-0.5-214-50MIN.lcm Date Acquired :1/25/2022 2:31:00 AM Date Processed :1/25/2022 8:49:47 AM

<Chromatogram View>



Pesk #	Ret. Time	Height	Area	Area%
1	18.344	176393	4524873	50.149
2	20.928	157007	4498063	49.851
Total		333400	9022936	100.000

Supplementary Figure 123. HPLC spectrum of racemic 30

HPLC spectrum of 30

Data File : sql-7-19-4-(12).lcd Method File : 3AD-H-75-0.5-214-50MIN.lcm Date Acquired : 1/25/2022 2:58:06 AM Date Processed : 1/25/2022 8:55:00 AM	Tray# Vial#	: 1 : 11	
Method File : 3ÅD-H-75-Ò.5 ² 214-50MIN.Icm Date Acquired : 1/25/2022 2:58:06 AM Date Processed : 1/25/2022 8:55:00 AM	Data File	: sql-7-19-4-(12).lcd	
Date Acquired : 1/25/2022 2:58:06 AM Date Processed : 1/25/2022 8:55:00 AM	Method File	: 3AD-H-75-0.5-214-50MIN.Icm	
Date Processed : 1/25/2022 8:55:00 AM	Date Acquired	1/25/2022 2:58:06 AM	
	Date Processed	: 1/25/2022 8:55:00 AM	

<Chromatogram View>



Supplementary Figure 124. HPLC spectrum of 30



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1(ppm)

Supplementary Figure 126. ¹³C NMR spectrum of compound 3p

¹¹B NMR (128 MHz, room temperature, CDCl₃)





Supplementary Figure 128. HPLC spectrum of racemic 3p



Supplementary Figure 129. HPLC spectrum of 3p



Supplementary Figure 131. ¹H NMR spectrum of compound 4a (S5 as the thiol catalyst)



Supplementary Figure 133. ¹³C NMR spectrum of compound 4a



Supplementary Figure 134. HPLC spectrum of racemic 4a

HPLC spectrum of 4a (S7 as the thiol catalyst)

Sample Name	:
Tray#	1
Vial#	47
Data File	: sql-6-349-3.lcd
Method File	: 3AD-H-90-1-214.lcm
Date Acquired	2/25/2022 9:34:52 PM
Date Acquired	: 2/25/2022 9:34:52 PM
Date Processed	: 2/25/2022 11:04:17 PM

<Chromatogram View>

sql-6-349-3.lcd m٧ 750-7.584 500- Ph_3S Đ Ρ'n 250-4a S7 as the thiol catalyst 11.476 0 2.5 10.0 5.0 7.5 0.0 12.5 min <Data Analysis> Detector Pesk # <u>214nm</u> Area% 91.624 Heigh Area 6514(8.376 100.000 1 476 318 595547 7110179 591530 Tota

Supplementary Figure 135. HPLC spectrum of 4a (S7 as the thiol catalyst)



<Chromatogram View>



Supplementary Figure 136. HPLC spectrum of 4a (S5 as the thiol catalyst)

HPLC spectrum of 4a (S1 as the thiol catalyst)

Sample Name	:	
Tray#	:1	
Vial#	: 13	
Data File	: SQL-7-15.lcd	
Method File	: 3AD-H-90-1-214.lcm	
Date Acquired	: 1/23/2022 3:29:41 PM	
Date Processed	: 2/28/2022 10:25:34 AM	

<Chromatogram View>



Supplementary Figure 137. HPLC spectrum of 4a (S1 as the thiol catalyst)

¹H NMR (400 MHz, room temperature, CDCl₃)



Supplementary Figure 138. ¹H NMR spectrum of compound 4a (gram-scale reaction)



Tray# Vial#	: 1 : 46	
Data File	sql-5-320-1.lcd	
Method File Date Acquired	: 3AD-H-90-1-214.lcm : 2/25/2022 9:19:32 PM	
Date Processed	: 2/25/2022 11:04:00 PM	
	P	

<Chromatogram View>



Supplementary Figure 139. HPLC spectrum of racemic 4a



Supplementary Figure 140. HPLC spectrum of 4a (gram-scale reaction)

¹H NMR (400 MHz, room temperature, CDCl₃)



Supplementary Figure 141. ¹H NMR spectrum of compound **4a** (D₂O recycled from a 0.6 mmol scale reaction)



Supplementary Figure 142. HPLC spectrum of racemic 4a



HPLC spectrum of **4a** (D₂O recycled from a 0.6 mmol scale reaction)

Supplementary Figure 143. HPLC spectrum of **4a** (D₂O recycled from a 0.6 mmol scale reaction)

¹H NMR (400 MHz, room temperature, CDCl₃)



Supplementary Figure 144. ¹H NMR spectrum of compound **4a** (D₂O recycled from a 3.0 mmol scale reaction)



Supplementary Figure 145. HPLC spectrum of racemic 4a

HPLC spectrum of **4a** (D₂O recycled from a 3.0 mmol scale reaction)

: 30
: sql-7-151-2.lcd
: 3AD-H-90-1-214.lcm
: 6/16/2022 6:28:30 PM
: 6/16/2022 8:47:33 PM

<Chromatogram View>



Supplementary Figure 146. HPLC spectrum of 4a (D₂O recycled from a 3.0 mmol scale reaction)

¹H NMR (400 MHz, room temperature, CDCl₃)

N0N0N00F004000		00-0400000-0-00
44000N0040-0N0	000000000000000000000000000000000000000	-00400-000000-00C
00000444444000	000000000000000000000000000000000000000	1044440000000C
	444666666666666666666666666666666666666	



Supplementary Figure 148. ¹³C NMR spectrum of compound 4b

HPLC spectrum of racemic 4b

Data File
Method File
Date Processed

: 21-RACEMIC-SQL-5-379-1.lcd : 3AD-H-96-1-214-20min.lcm : 8/8/2021 12:13:54 PM



Supplementary Figure 149. HPLC spectrum of racemic 4b

HPLC spectrum of 4b

Data File	
Method File	
Date Processed	

21-CHIRAL-SQL-5-380.lcd	
3AD-H-96-1-214-20min.lcn	n
8/8/2021 12:14:28 PM	



Supplementary Figure 150. HPLC spectrum of 4b



Supplementary Figure 152. ¹³C NMR spectrum of compound 4c

HPLC spectrum of racemic 4c Sample Name : Tray# :1 Vial# :46 Data File : SQL-6-391-1-4.Icd Method File :3AD-H-80-0.5-214.Icm Date Acquired :1/24/2022 7:21:08 PM Date Processed :1/24/2022 7:46:22 PM

<Chromatogram View>





Supplementary Figure 153. HPLC spectrum of racemic 4c

HPLC spectrum of 4c

Sample Name	
Tray#	:1
Vial#	: 47
Data File	: SQL-7-35-1-1.lcd
Method File	: 3AD-H-80-0.5-214.lcm
Date Acquired	: 1/24/2022 7:47:15 PM
Date Processed	: 1/24/2022 8:12:39 PM

<Chromatogram View>

SQL-7-35-1-1.lcd m٧ 88 750-D Ph₃Si Ó N 500-Ρh 4c 250-9.802 0-10.0 2.5 7.5 5.0 0.0 min <Data Analysis> Detector A 214nm Pesk # Ret. Tim Height 8453 400 4.754

Supplementary Figure 154. HPLC spectrum of 4c



Supplementary Figure 156. ¹³C NMR spectrum of compound 4d

¹⁹F NMR (376 MHz, room temperature, CDCl₃)



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

Supplementary Figure 157. ¹⁹F NMR spectrum of compound 4d

HPLC spectrum of racemic 4d

Sample Name	:
Tray#	:1
Viaĺ#	: 20
Data File	: sql-7-72-1.lcd
Method File	: 4OD-H-80-0.5-214.lcm
Date Acquired	: 2/20/2022 11:00:16 PM
Date Processed	: 2/20/2022 11:32:56 PM

<Chromatogram View>



Supplementary Figure 158. HPLC spectrum of racemic 4d



Supplementary Figure 160. ¹H NMR spectrum of compound 4e



Supplementary Figure 161. ¹³C NMR spectrum of compound 4e



Sample Name 1 Tray# 1 Vial# 28 Data File sql-7-59-7 Method File 4OD-H-90 Date Acquired 2/20/2022 Date Processed 2/20/2022	1-8.lcd 0-0.5-214.lcm 2 12:03:21 AM
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<Chromatogram View>

E



Supplementary Figure 162. HPLC spectrum of racemic 4e



Supplementary Figure 164. ¹H NMR spectrum of compound 4f
¹³C NMR (100 MHz, room temperature, CDCl₃)



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

Supplementary Figure 165. ¹³C NMR spectrum of compound 4f

HPLC spectrum of racemic 4f

Sample Name	
Tray#	:1
Vial#	: 19
Data File	: sql-7-58-1.lcd
Method File	: 4OD-H-90-0.5-214.lcm
Date Acquired	: 2/16/2022 12:59:30 AM
Date Processed	: 2/16/2022 1:18:36 AM

<Chromatogram View>



Supplementary Figure 166. HPLC spectrum of racemic 4f

 HPLC spectrum of 4f
 I

 Sample Name
 1

 Tray#
 1

 Vial#
 20

 Data File
 sql-7-58-2.lcd

 Method File
 40D-H-90-0.5-214.lcm

 Date Acquired
 2/16/2022 1:40:28 AM

 Date Processed
 2/16/2022 1:53:57 AM

<Chromatogram View>



Supplementary Figure 167. HPLC spectrum of 4f





Supplementary Figure 168. ¹H NMR spectrum of compound 4g



<Chromatogram View>



Supplementary Figure 170. HPLC spectrum of racemic 4g



Supplementary Figure 172. ¹H NMR spectrum of compound 4h



HPLC spectrum of 4h

Data File	
Method File	
Date Processed	

SQL-5-375-6.lcd 3AD-H-96-1-214-20min.lcm 9/2/2021 9:07:29 AM



Supplementary Figure 176. ¹H NMR spectrum of compound 4i





210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

Supplementary Figure 177. ¹³C NMR spectrum of compound 4i

HPLC spectrum of racemic 4i

Tray#	:1
Vial#	: 19
Data File	: sql-5-284-1-1.lcd
Method File	: 10J-H-99-1-214.lcm
Date Acquired	: 2/24/2022 9:03:24 AM
Date Processed	: 2/24/2022 9:41:20 AM

<Chromatogram View>



Supplementary Figure 178. HPLC spectrum of racemic 4i



Supplementary Figure 180. ¹H NMR spectrum of compound 4j

¹³C NMR (100 MHz, room temperature, CDCl₃)

	DNATENO				
0	0010000	NON	00	-	4 1~
4	00044000		60	9	4 4
ó	6666666 N	000	Ω	9	50
õ	0000000	N N 60	(C)	0	00
	x x x x x x x	アアア	9	-	CV
		\checkmark	1	1	11





Supplementary Figure 181. ¹³C NMR spectrum of compound 4j

HPLC spectrum of racemic 4j

Termult	
ITdy#	.1
Viaĺ#	: 19
Data File	: sql-6-217-1-1-3.lcd
Method File	: 3AD-H-96-1-214.lcm
Date Acquired	: 11/19/2021 5:07:52 PM
Date Processed	: 2/26/2022 12:31:48 AM

<Chromatogram View>



Supplementary Figure 182. HPLC spectrum of racemic 4j



Supplementary Figure 183. HPLC spectrum of 4j

¹H NMR (400 MHz, room temperature, CDCl₃) 1.5721.5681.5621.5531.5531.5511.5511.55461.5461.5461.5420.9030.9040.000



210 200 190 180 170 160 150 140 130 120 110 100 90 f1 (ppm) 80 70 60 50 40 30 20 10 0 -10 Supplementary Figure 185. ¹³C NMR spectrum of compound 5a

³¹P NMR (162 MHz, room temperature, CDCl₃)



120 -40 -60 f1 (ppm) 140 100 80 60 40 -100 20 0 -20 -80 -120 -140 -160 -180 -200 -220 -240

Supplementary Figure 186. ³¹P NMR spectrum of compound 5a

HPLC spectrum of racemic 5a

1	
Data File	: (39-racemic)SQL-4-235-1.lcd
Method File	: AD-H-65-0.5-214.lcm
Date Processed	: 7/17/2021 12:34:24 PM



(39-racemic)SQL-4-235-1.lcd m٧ 750 14.087 5.00 500 Ĥ 250 racemic 5a 0-12.5 2.5 5.0 7.5 10.0 0.0 15.0 min <Data Analysis> DetA 214nm Pesk # Ret. Time 1 14.08 Area% 49.830 50.170 100.000 Heigh Area 2878 ∠ Tota

Supplementary Figure 187. HPLC spectrum of racemic 5a

HPLC spectrum of 5a

Date Processed : 7/17/2021 12:34:	-258-2lcd 4.lcm :07 PM
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LC2V #	Ret. Hille	Height	Alea	Alea /o	
1	13.990	536413	10961497	95.919	
2	15.059	19860	466318	4.081	
Total		556273	11427816	100.000	

Supplementary Figure 188. HPLC spectrum of 5a





Supplementary Figure 189. ¹H NMR spectrum of compound 5b





Supplementary Figure 191. ³¹P NMR spectrum of compound 5b

HPLC spectrum of racemic 5b

Data File	
Method File	
Date Processed	

: (40-racemic)SQL-4-323-1.lcd : 4OD-H-80-1-214.lcm : 7/17/2021 12:35:12 PM



Supplementary Figure 192. HPLC spectrum of racemic 5b

HPLC spectrum of 5b

Data File
Method File
Date Processed

(40-chiral)SQL-4-327.lcd
4OD-H-80-1-214.lcm
7/17/2021 12:34:55 PM



Supplementary Figure 193. HPLC spectrum of 5b



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 Supplementary Figure 195. ¹³C NMR spectrum of compound 5c



62 60 58 56 54 52 50 48 46 44 42 40 38 36 34 32 30 28 26 24 22 20 18 16 14 12 10 8 f1 (ppm) 6 4 2 0 -2 -4

Supplementary Figure 196. ³¹P NMR spectrum of compound 5c

HPLC spectrum of racemic 5c Data File : (41-rac Method File : AD-H-4 Date Processed : 7/17/20

: (41-racemic)SQL-4-243-1.lcd : AD-H-60-0.5-214.lcm : 7/17/2021 12:35:49 PM

```
<Chromatogram View>
```



Supplementary Figure 197. HPLC spectrum of racemic 5c

HPLC spectrum of 5c

Data File (41-c) Method File 3AD- Date Processed 7/17/2	H-60-0.5-214.lcm 2021 12:35:35 PM
---------------------------------------------------------------	--------------------------------------



Supplementary Figure 198. HPLC spectrum of 5c

¹H NMR (400 MHz, room temperature, CDCl₃)





Supplementary Figure 199. ¹H NMR spectrum of compound 5d



140 120 100 80 60 40 -20 -40 -60 f1 (ppm) 20 0 -80 -100 -120 -140 -160 -180 -200 -220 -240 Supplementary Figure 201. ³¹P NMR spectrum of compound 5d

¹⁹F NMR (376 MHz, room temperature, CDCl₃)

-105.468



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

Supplementary Figure 202. ¹⁹F NMR spectrum of compound 5d

HPLC spectrum of racemic 5d

Data File
Method File
Date Processed

: ((42-racemic)SQL	-4-256-1.lcd
	2AS-H-80-0.5-21	4.lcm
1	7/17/2021 12:36:	05 PM



Supplementary Figure 203. HPLC spectrum of racemic 5d

HPLC spectrum of **5d** Data File Method File Date Processed

: (42-chiral)SQL-4-301.lcd : 2AS-H-80-0.5-214.lcm : 7/17/2021 12:36:24 PM





.916

Supplementary Figure 205. ¹H NMR spectrum of compound 5e



-40 -60 f1 (ppm) 140 120 100 80 60 40 -200 -220 20 -20 -80 -100 -120 -140 0 -160 -180 -240 Supplementary Figure 207. ³¹P NMR spectrum of compound 5e

HPLC spectrum of racemic **5e**

Data File	: (43-racemic)SQL-4-336-1.lcd
Method File	: 3AD-H-70-1-214.lcm
Date Processed	: 7/17/2021 12:36:52 PM



Supplementary Figure 208. HPLC spectrum of racemic 5e

HPLC spectrum of **5e** Data File Method File Date Processed

(43-chiral)SQL-4-348-.lcd 3AD-H-70-1-214.lcm 7/17/2021 12:36:39 PM



(43-chiral)SQL-4-348-.lcd



Supplementary Figure 209. HPLC spectrum of 5e



Supplementary Figure 211. ¹³C NMR spectrum of compound 5f

³¹P NMR (162 MHz, room temperature, CDCl₃)



-40 -60 f1 (ppm) 140 120 100 80 60 40 20 Ó -20 -80 -100 -120 -140 -160 -180 -200 -220 -240

Supplementary Figure 212. ³¹P NMR spectrum of compound 5f

HPLC spectrum of racmic 5f

Data File Method File Date Processed : (45-racemic)SQL-4-257-1.lcd : 3AD-H-60-0.5-214.lcm : 7/17/2021 12:37:54 PM

<Chromatogram View>



Supplementary Figure 213. HPLC spectrum of racemic 5f

HPLC spectrum of 5f

Data File	
Method File	
Date Processed	

: (45-chiral) SQL-4-288.lcd : 3AD-H-60-0.5-214.lcm : 7/17/2021 12:37:41 PM



Supplementary Figure 214. HPLC spectrum of 5f





Supplementary Figure 215. ¹H NMR spectrum of compound 5g



140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 **Supplementary Figure 217**. ³¹P NMR spectrum of compound **5**g

HPLC spectrum of racemic **5g**

-	-
Data File	: 46-RACEMIC-SQL-4-289-1.lcd
Method File	: 3AD-H-70-1-214.lcm
Date Processed	: 7/17/2021 12:40:21 PM



Supplementary Figure 218. HPLC spectrum of racemic 5g

HPLC spectrum of **5g** Data File Method File Date Processed

: 46-CHIRAL-SQL-4-287-4.lcd : 3AD-H-70-1-214.lcm : 7/17/2021 12:38:56 PM



¹H NMR (400 MHz, room temperature, CDCl₃)

Construction of the second sec



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 Supplementary Figure 221. ¹³C NMR spectrum of compound 5h

173

³¹P NMR (162 MHz, room temperature, CDCl₃)



140 120 -40 -60 f1 (ppm) 100 80 60 40 20 0 -20 -80 -100 -120 -140 -160 -180 -200 -220 -240

Supplementary Figure 222. ³¹P NMR spectrum of compound 5h

HPLC spectrum of racemic 5h

: SQL-4-293-1.lcd
: 40D-H-88-12-1-214.lcm
: 12/29/2020 10:09:02 AM



Supplementary Figure 223. HPLC spectrum of racemic 5h

HPLC spectrum of 5h

: SQL-4-293-2-1.lcd : 4OD-H-88-12-1-214.lcm : 12/29/2020 10:48:54 AM
. 12/29/2020 10.40.04 AM



Supplementary Figure 224. HPLC spectrum of 5h





Supplementary Figure 225. ¹H NMR spectrum of compound 5i



Supplementary Figure 227. ³¹P NMR spectrum of compound 5i

HPLC spectrum of racemic **5i**

Data File	: (47-racemic)SQL-4-311-1.lcd
Method File	: 6IC-H-80-0.5-214.lcm
Date Processed	: 7/17/2021 12:41:00 PM



Supplementary Figure 228. HPLC spectrum of racemic 5i

HPLC spectrum of 5i

Data File Method File Date Processed

: (47-chiral)SQL-4-311-2.lcd : 6IC-H-80-0.5-214.lcm : 7/17/2021 12:40:39 PM



(47-chiral)SQL-4-311-2.lcd m٧ 1000-17.820 750-P٢ P١ 500-O Ó C Đ 5i 250-15.632 0 5.0 15.0 2.5 12.5 17.5 7.5 10.0 0.0 min <Data Analysis> 214nm Time 632 Ret 348 14170277 14737410 152 000

Supplementary Figure 229. HPLC spectrum of 5i



Supplementary Figure 231. ¹³C NMR spectrum of compound 5j





10 0 -10 f1 (ppm) 90 80 70 60 50 40 20 -20 -30 30 -40 -50 -60 -70 -80 -90 Supplementary Figure 233. ¹¹B NMR spectrum of compound 5j

HPLC spectrum of racemic **5**j

1	
Data File	: 48-sql-6-21-1racemic.lcd
Method File	: 3AD-H-70-0.8-214.lcm
Date Processed	: 8/8/2021 1:50:08 PM



Supplementary Figure 234. HPLC spectrum of racemic 5j

HPLC spectrum of 5j

Data File
Method File
Date Processed

ł	48-SQL-6-21-2chiral.lcd
i	3AD-H-70-0.8-214.lcm
2	8/8/2021 1:50:54 PM



Supplementary Figure 235. HPLC spectrum of 5j
¹H NMR (400 MHz, room temperature, CDCl₃) 1^{1} H NMR (400 MHz, room temperature, CDCl₃)



Supplementary Figure 236. ¹H NMR spectrum of compound 6a

¹³C NMR (100 MHz, room temperature, CDCl₃)



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 Supplementary Figure 237. ¹³C NMR spectrum of compound **6a**





HPLC spectrum of racemic 6a

: XM
: 3AE
: 8/5/



Supplementary Figure 239. HPLC spectrum of racemic 6a

HPLC spectrum of 6a

.529

Data File	: xmc-4-146-1-1.lcd
Method File	: 3AD-H-90-0.5-214.lcm
Date Processed	: 10/19/2021 2:44:29 PM



0.000



Supplementary Figure 241. ¹H NMR spectrum of compound 6b



190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

Supplementary Figure 242. ¹³C NMR spectrum of compound 6b

¹⁹F NMR (376 MHz, room temperature, CDCl₃)





184

HPLC spectrum of racemic 6b

Data File	: xmc-2-61a-1-100501 lcd
Method File	40D-H-90-0 5-214 lcm
Date Processed	: 10/5/2021 9:19:44 PM



Delector	A Z 141111			
Pesk #	Ret. Time	Height	Area	Area%
1	10.659	196362	2303539	50.103
2	11.469	180255	2294061	49.897
Total		376617	4597600	100.000

Supplementary Figure 244. HPLC spectrum of racemic 6b

HPLC spectrum of **6b** Data File Method File Date Processed

: xmc-4-155-1-1-02.lcd
: 40D-H-90-0.5-214.lcm
: 10/19/2021 8:02:22 PM



Supplementary Figure 245. HPLC spectrum of 6b

¹H NMR (400 MHz, room temperature, CDCl₃)



Supplementary Figure 246. ¹H NMR spectrum of compound 6c

¹³C NMR (100 MHz, room temperature, CDCl₃)





Supplementary Figure 247. ¹³C NMR spectrum of compound 6c





Supplementary Figure 249. HPLC spectrum of racemic 6c

HPLC spectrum of **6c** Data File Method File Date Processed

: xmc-4-165-1.lcd : 3AD-H-95-1-214.lcm : 10/28/2021 10:49:37 PM



Supplementary Figure 250. HPLC spectrum of 6c





Supplementary Figure 251. ¹H NMR spectrum of compound 6d



-10 -60

Supplementary Figure 253. ¹⁹F NMR spectrum of compound 6d

HPLC spectrum of racemic **6d**



Delector A 214mm			
Pesk # Ret. Time	Height	Area	Area%
1 9.462	765269	8570586	50.105
2 10.475	683338	8534804	49.895
Total	1448607	17105391	100.000

Supplementary Figure 254. HPLC spectrum of racemic 6d

HPLC spectrum of 6d

Data File Method File Date Processed

: xmc-2-156-1.lcd
: 3AD-H-90-0.5-214.lcm
: 10/19/2021 2:44:16 PM



Supplementary Figure 255. HPLC spectrum of 6d

¹H NMR (400 MHz, room temperature, CDCl₃)



Supplementary Figure 256. ¹H NMR spectrum of compound 6e

¹³C NMR (100 MHz, room temperature, CDCl₃)



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 Supplementary Figure 257. ¹³C NMR spectrum of compound **6e**





10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 Supplementary Figure 258. ¹⁹F NMR spectrum of compound 6e

HPLC spectrum of racemic 6e



: xmc-4-21-2-01.lcd : 40D-H-98-1-214.lcm : 10/13/2021 6:37:53 PM



xmc-4-21-2-01.lcd m∨ 400 9.697 300-554 6 200 Ô , racemic **6e** 100-0-2.5 5.0 7.5 12.5 10.0 0.0 min <Data Analysis> Detector A 214nm Pesk # Ret. Time 1 9.697 Height 327019 Area 5003046 Area 50 49.704 10 554 4944142 9947188 Tota

Supplementary Figure 259. HPLC spectrum of racemic 6e

HPLC spectrum of 6e

Data File	
Method File	
Date Processed	

: xmc-2-157-1-1.lcd : 40D-H-98-1-214.lcm : 10/18/2021 10:33:06 PM



Supplementary Figure 260. HPLC spectrum of 6e





Supplementary Figure 261. ¹H NMR spectrum of compound 6f



Supplementary Figure 263. ¹⁹F NMR spectrum of compound 6f





Supplementary Figure 264. HPLC spectrum of racemic 6f

HPLC spectrum of 6f

Method File Date Acquired Date Processed	: 10J-H-99.7-0.5-214.lcm : 6/14/2022 11:53:08 PM : 6/15/2022 9:26:27 AM	
<chromatogram '<="" th=""><th>View></th><th></th></chromatogram>	View>	



Supplementary Figure 265. HPLC spectrum of 6f







HPLC spectrum of racemic **7**

Vial#	:1	
Data File	: sql-7-31-1-(Y3).lcd	
Method File	: 40D-H-80-0.5-214.lcm	
Date Acquired	: 1/22/2022 7:06:47 PM	
Date Processed	: 1/25/2022 1:39:01 AM	

<Chromatogram View>



Supplementary Figure 269. HPLC spectrum of racemic 7



Supplementary Figure 271. ¹H NMR spectrum of compound 8



Supplementary Figure 273. ¹¹B NMR spectrum of compound 8

¹⁹F NMR (376 MHz, room temperature, CDCl₃)

~155.133

10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

Supplementary Figure 274. ¹⁹F NMR spectrum of compound 8

HPLC spectrum of racemic 8

	Matrix 241-72 Data File : 240-1- Date Acquired : 2/1/20 Date Processed : 2/1/20	25-2.1cd 1-65-0.5-214.1cm 22 12:33:38 PM 22 1:23:11 PM
--	------------------------------------------------------------------------------------------	-----------------------------------------------------------------

<Chromatogram View>



Supplementary Figure 275. HPLC spectrum of racemic 8



Supplementary Figure 277. ¹H NMR spectrum of compound 9



Supplementary Figure 279. ¹¹B NMR spectrum of compound 9

HPLC spectrum of racemic **9**

Sample Marile	-
Tray#	:1
Vial#	: 32
Data File	: SQL-7-27-8-(Y2).lcd
Method File	: 40D-H-90-0.5-214-40min.lcm
Date Acquired	: 1/22/2022 12:23:53 AM
Date Processed	: 1/25/2022 1:36:17 AM

<Chromatogram View>



Pesk #	Ret. Time	Height	Area	Area%
1	83.332	85130	9105288	49.534
2	88.586	70990	9276560	50.466
Total		156120	18381848	100.000

Supplementary Figure 280. HPLC spectrum of racemic 9

HPLC spectrum of 9

Tray#	:1
Viaĺ#	: 33
Data File	: SQL-7-36-(Y3).lcd
Method File	: 40D-H-90-0.5-214-40min.lcm
Date Acquired	: 1/22/2022 2:04:59 AM
Date Processed	: 1/25/2022 1:37:25 AM

<Chromatogram View>

SQL-7-36-(Y3).lcd m٧ 150 100-83.391 ЮH 50-H₂ D NHPh 9 90.730 0 75 50 25 100 min <Data Analysis> 214nm Ret. Tim Detector A leigh 67 Area% 93,248 6.752 100.000 90 730 4130 71812 7859227 Tota

Supplementary Figure 281. HPLC spectrum of 9



20 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 Supplementary Figure 283. ¹³C NMR spectrum of compound 10

HPLC spectrum of racemic 10



Supplementary Figure 284. HPLC spectrum of racemic 10

HPLC spectrum of 10

Date Acquired Date Processed	: 9/1/2021 8:54:08 PM : 9/1/2021 11:04:48 PM	
Data File Method File	SQL-6-96.lcd : 40D-H-80-0.5-214.lcm	



Supplementary Figure 285. HPLC spectrum of 10



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 **Supplementary Figure 287.** ¹³C NMR spectrum of compound **11**

HPLC spectrum of racemic **11**

Data File	: sql-6-88-4.lcd
Method File	: 3AD-H-90-1-214.lcm
Date Processed	: 9/1/2021 4:27:09 PM



HPLC spectrum of 11

: sql-6-106.lcd : 3AD-H-90-1-214.lcm : 9/21/2021 4:03:02 PM



Supplementary Figure 289. HPLC spectrum of 11



Supplementary Figure 291. ¹³C NMR spectrum of compound 12



HPLC spectrum of 12

Data File	sql-6-97-2-chiral003.lcd
Method File	: 10J-H-85-1-214.lcm
Date Acquired	: 9/8/2021 10:00:59 PM
Date Processed	: 9/8/2021 11:30:18 PM

<Chromatogram View>

sql-6-97-2-chiral003.lcd m∨ 1000 750-OH Ph₃Si 500-14.261 NΗ Đ Ρh 12 250-9.310 0-15.0 2.5 7.5 17.5 12.5 0.0 5.0 10.0 20.0 min <Data Analysis> Detector A 214nm Pesk # Ret. Time 9.310 9.310 Heigh Area 3466 3/16 91.809 100.000 14 261 385 461 42319679 Tot:

Supplementary Figure 293. HPLC spectrum of 12

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