nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

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Stat	ISTICS		
For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.			
n/a 0	Confirmed		
	The exact	sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement	
	🔀 A stateme	ent on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly	
	The statis	tical test(s) used AND whether they are one- or two-sided non tests should be described solely by name; describe more complex techniques in the Methods section.	
	A description of all covariates tested		
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons		
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)		
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.		
	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings		
For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes			
\square Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated			
Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.			
Software and code			
Policy	information	about <u>availability of computer code</u>	
Dat	a collection	Sciex SIMCA, MassLynx 4.1, QuanLynx,	
Dat	a analysis	Prism, Excel,	
For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.			
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Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Our proteomic data sets are available on line at massive.ucsd.edu under the accession numbers MSV000087642 and MSV000087641. All raw data is provided in source data.

Field-specific reporting		
☐ Life sciences	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection. Behavioural & social sciences Ecological, evolutionary & environmental sciences be document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf	
ror a reference copy or t	ne document with an sections, see <u>nature.com/documents/nr-reporting-summary-nat.pdr</u>	
Life scier	nces study design	
	close on these points even when the disclosure is negative.	
Sample size	Sample size was dictated by the standards in the filed and our ability to isolate quiescent cells. Wojcik, C., DeMartino, G.N. (2002)J. Biol. Chem. 277(8): 61886197. and Sieber et al (2016) Cell 164(3): 420432.	
Data exclusions	No data was excluded from this study	
Replication	No replicate experiments were excluded from this study. All replicate experiments displayed effects similar to those presented in the manscript.	
Randomization	Our proteomics samples were randomized and blinded to ensure no bias in subsequent analysis	
Blinding	Our proteomics samples were randomized and blinded to ensure no bias in subsequent analysis	
Materials & exp n/a Involved in th Antibodies Eukaryotic Palaeontol Animals an Human res Clinical dat Dual use re	Cell lines ChIP-seq Flow cytometry Degy and archaeology MRI-based neuroimaging d other organisms earch participants	
Antibodies		
Antibodies used	Anti-ATP5a (Abcam #15H4C4 lot#GR3306993-25), Anti-K48 linkage (ABcam #EP8589 lot:6), anti-20S (George Demartino), anti-Rpt2(George Demartino), anti-Rpt5 (George Demartino), anti-Rpn12 (George Demartino), anti-tubulin (DSHB# E7), Anti-actin(DSHB# JLA20)	
Validation	These antibodies were validated in multiple cells lines and drosophila by Western blot. Cells were also confirm free of mycoplasma by commercial kit.	
Eukaryotic c	ell lines	
Policy information	about <u>cell lines</u>	
Cell line source(s	3t3 cells (ATCC CRL-1658), HEK293T cells (ATCC CRL-3216)	

Cell line source(s)

Authentication

Mycoplasma contamination

Commonly misidentified lines (See ICLAC register)

State CRL-1658), HEK293T cells (ATCC CRL-3216)

These cells were validated by their ability to differentiate into myofibers in vitro.

No contamination was detected

no commoly misidentified lines (See ICLAC register)

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research		
Laboratory animals	drosophila (oregon R) adult females, neurospora crassa	
Wild animals	N/A	
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Field-collected samples	N/A	
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Ethics oversight	No ethical guidance was required for this study.	

Note that full information on the approval of the study protocol must also be provided in the manuscript.