Supplementary Information

Development of ⁶⁸Ga-labeled Hepatitis E virus nanoparticles for targeted drug delivery and diagnostics with PET

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Experimental Procedures

All chemicals and solvents were obtained from commercial providers and they were used without further purification.

Cell culture:

Cell culturing flasks and well plates were obtained from Corning Inc. (Corning, NY, U.S.A.). McCoy's 5A (for HCT 116 cells) and Dulbecco's Modified Eagle Medium (RAW 264.7 cells) cell culture media were purchased from ATCC (Manassas, VA, U.S.A.) and Corning, respectively. DMEM (for Hep G2), CO₂-independent cell medium, GlutaMAX (1×), sodium pyruvate (100 mM), fetal bovine serum (FBS), Penicillin-Streptomycin (10,000 U/ml), non-essential amino acids (100×NEAA), Dulbecco's phosphate buffer saline (10×DPBS), TrypLETM Express (1×), and sterile vacuum filter/storage bottle unit (250–500 ml, 0.22 µm PES membrane) were purchased from Life Technologies Gibco (Carlsbad, CA, U.S.A.). The Heracell VIOS 160i incubator (Thermo Fisher Scientific, Waltham, MA, U.S.A.) was maintained at 37 °C, 5% CO₂, and 95% relative humidity.

Cell culture media preparation:

The cell culture media were supplemented with 10% FBS, 1% GlutaMAX, 1% sodium pyruvate and 1% Penicillin-Streptomycin, and aseptically filtered through sterile 0.22- μ m filter before use. In addition to this general procedure, DMEM medium for Hep G2 cell line was further supplemented with 1% NEAA.

<u>Cell studies of [68Ga]Ga-DOTA-HEVNPs and controls in hepatocytes, macrophages and colorectal cancer cells:</u>

The cell medium was removed and collected into a scintillation tube at the designed time. *Free fraction collection*: the cells were washed with 1 mL of ice-cold 0.01 M PBS (pH 7.4) and the PBS was collected in the same tube. *Membrane-bound fraction collection*: the addition of 1 mL of ice-cold glycine buffer (0.05 M, pH 2.8). The cells were incubated for 5 min on ice. The buffer was then collected in a new scintillation tube. This procedure was repeated once and the buffer was collected in the same tube. The treated cells were washed with 1 mL of ice-cold 0.01 M PBS (pH 7.4) and this was then collected in the same tube. *Internalized fraction*

collection: 1 mL of 1 M NaOH was added and the cells were incubated for 10 min at RT. NaOH was collected in a new scintillation tube. The cells were washed twice with 1 mL of ice-cold 0.01 M PBS and the buffer was collected both times in the same tube. The radioactivity was detected in each tube using the Wizard automated gamma-counter. The percent internalization was normalized with total added activity and calculated as: %Internalization = 100 × CPM of internalized fraction/SUM of the CPM in all fractions. The measurements were done in triplicate for every time point and sample under study.



Results

Fig.S1: Radio-TLC chromatograms of pure [⁶⁸Ga]Ga-DOTA-HEVNPs (top) and free ⁶⁸Ga-radionuclide (bottom); conditions: Whatman 1 paper chromatography with 0.5 mM DTPA.

Tissue	15 min		30 min		60 min		120 min	
	%ID/g	S.D.	%ID/g	S.D.	%ID/g	S.D.	%ID/g	S.D.
Urine	97.34	83.66	222.49	149.24	142.61	98.55	46.45	55.48
Blood	3.9	1.47	3.42	0.7	2.87	2.32	2.11	0.67
Pancreas	0.52	0.22	0.6	0.08	0.49	0.43	0.48	0.18
Spleen	12.11	2.28	12.14	3.53	17.21	7.7	15.05	3.44
Kidney	2.12	0.26	1.63	0.25	1.48	1.03	1.09	0.17
Gallbladder	11.51	11.23	21.53	12.8	17.4	10.58	15.16	8.8
Liver	55.04	13.83	69.28	10.86	96.34	58.65	62.82	11.04
Lung	3.99	1.64	4.09	1.09	3.42	2.94	2.49	0.77
Heart	1.77	0.75	1.59	0.32	1.24	1.01	1.06	0.37
Muscle	0.46	0.21	0.6	0.19	0.42	0.42	0.44	0.19
Occipital bone	1.29	0.85	1.32	0.84	1.44	1.09	1.62	0.8
Bone with marrow	2.18	0.89	2.3	0.87	3.42	3.1	3.2	0.99
Brain	0.16	0.1	0.16	0.08	0.13	0.12	0.13	0.06
Stomach	0.25	0.12	0.23	0.09	0.3	0.33	0.23	0.09
Small intestine	0.4	0.06	0.46	0.13	0.6	0.49	0.6	0.27
Large intestine	0.23	0.07	0.28	0.09	0.28	0.24	0.48	0.16
Skin	1.06	0.53	1.11	0.54	0.86	0.76	1.08	0.3

Table S1: The average %ID/g values obtained from the *ex vivo* biodistribution of the [68 Ga]Ga-DOTA-HEVNPs tracer in healthy mice (n = 3-5). S.D.: standard deviation.





Fig.S2: Membrane-bound fractions of [68 Ga]Ga-DOTA-HEVNPs and [68 Ga]GaCl₃ in (A) hepatocytes (Hep G2) and (B) macrophages (RAW 264.7). The comparison between the two cell lines is shown for [68 Ga]Ga-DOTA-HEVNPs (C). The cell uptake assay was done at 37 °C up to 2 h after the initiation of the incubation. The columns represent the average ± standard deviation (n = 3).