

Figure S1 Morphology of PDOs before and after the passage.

PDOs (HCT64-1T) were mechanically disrupted by pipetting, split in the indicated ratio, and cultured in ENR medium. Images were collected at four days after plating using stereomicroscope. (Bar = 400 μ M)

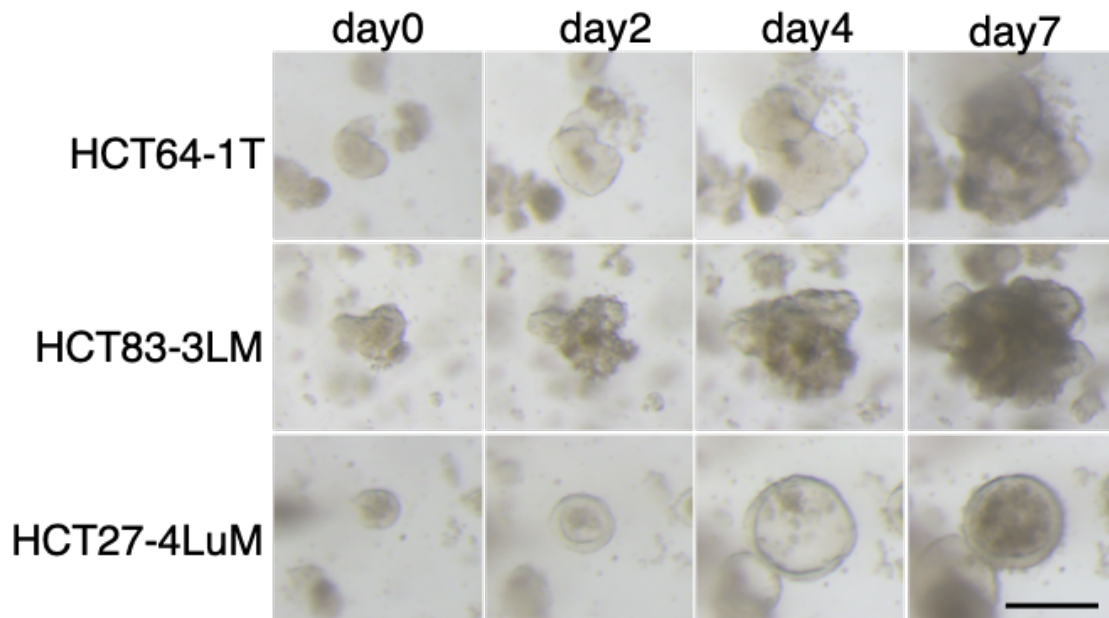


Figure S2. Morphological change during organoid growth.

Organoids were broken into small pieces by pipetting, expanded 1:4 in Matrigel and cultured in ENR medium (day0). Medium was changed every 3-4 days. To examine the morphology of organoids during the culture, each organoid was followed by identifying their location manually. Three PDOs representing type 0 (HCT64-1T), type 1 (HCT83-3LM) and type 5 (HCT27-4LuM) were shown. Image was taken at indicated days. Bar = 200 μ m

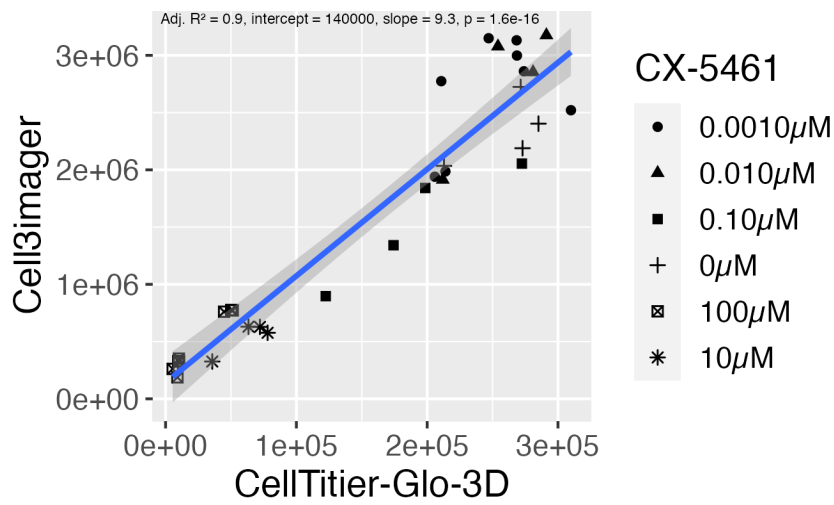


Figure S3 The correlation of cell viabilities as measured by Cell3imager and CellTiter Glo-3D.

PDO (HCT64-1T) was cultured in 96 well dishes with the indicated concentrations of CX-5461 for 10 days. Cell Viability was assessed by imaging using Cell3imager and by quantifying ATP using CellTiterGlo-3D. Linear regression line and confidence interval were shown in blue and gray, respectively.

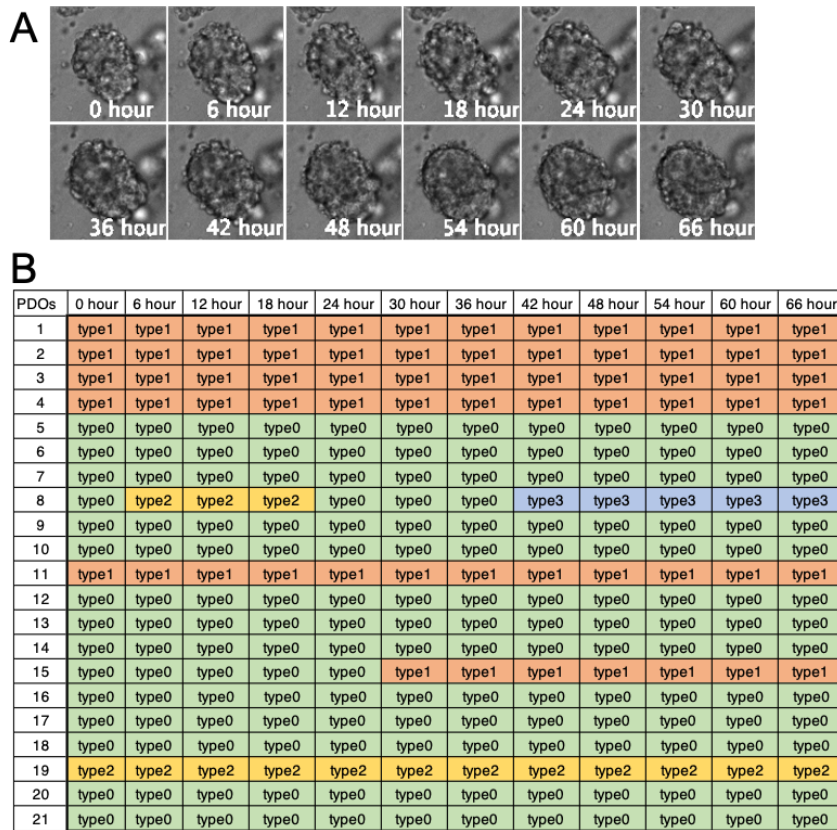


Figure S4 Analysis of time-lapse imaging of PDOs.

(A) PDOs were cultured in Matrigel, and images were taken from 4 days to 7 days after expansion. Single organoid images were manually cropped and used for classification.

(B) Morphological type of 21 organoids in each time point was determined by three referees and the summary was shown.

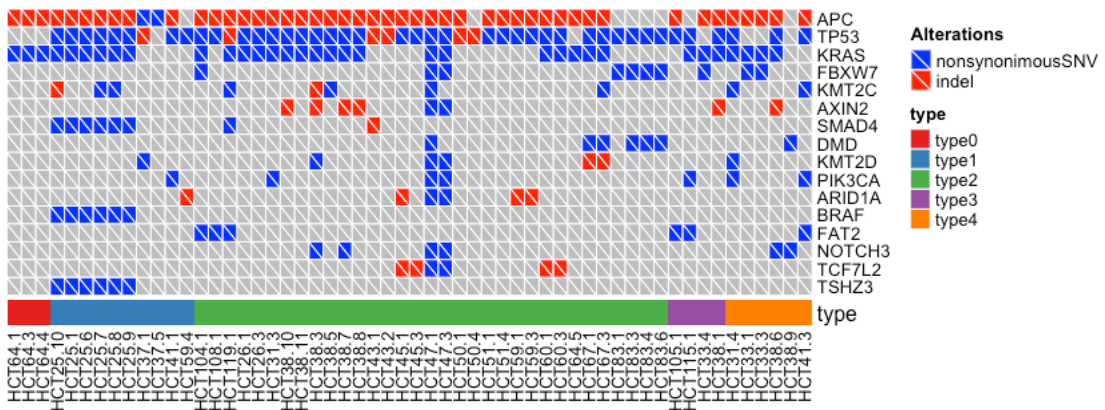


Figure S5. Mutation profile of PDOs.

Somatic mutations detected in multiple organoids were shown.