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# Supplemental Information

# How insulin-like growth factor I binds

## to a hybrid insulin receptor type 1

## insulin-like growth factor receptor

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#### A **Protein Sequence of IR-Bzip**

B



- - 801 ADDIPGPVTWEPRPENSIFLKWPEPENPNGLILMYEIKYGSQVEDQRECVSRQEYRKYGGAKLNRLNPGNYTARIQATSL 880
	- 881 SGNGSWTDPVFFYVQAKTGYENFIHRMKQLEDKVEELLSKNYHLENEVARLKKLVGERSSSEQKLISEEDLN 952

### **Figure S1. Protein sequences of the two constructs use to form HybZip, Related to STAR Methods**.

(**A**) IR-Bzip, comprising residues 1-928 of the human IR-B isoform followed by the 33-residue GCN4 zipper sequence (*orange*).

(**B**) IGF-1Rzip, comprising residues 1-905 of human IGF-1R followed by the 33-residue GCN4 zipper sequence (*orange*) followed by a three-serine spacer and the c-myc tag sequence EQKLISEEDLN (*blue*).



**Figure S2. Purification and characterization of HybZip, Related to STAR Methods**.

(**A**) Western blots of protein product obtained after serial elution from both 9E10 mAb and 18-44 mAb beads, showing presence of both species in the material (lanes with *bar* on right-hand side of each blot). Left-hand lanes in each blot correspond to pH 3 eluted material obtained from final column wash. Superfluous boundary has been removed from both blot images.

(**B**) Size-exclusion chromatogram obtained from protein product post affinity purification. The dominant peak corresponds to  $(\alpha\beta)_4$  species, with a shoulder corresponding to  $(\alpha\beta)_2$  species.

(C) Enrichment of  $(\alpha\beta)_2$  species following re-run of pooled shoulder fractions from (B).

(**D**) Schematic illustrating leucine-zipper cross-linking of receptor ectodomain dimers to form hybrid tetrameric species. All four tetrameric species will be affinity co-purified with the desired hybrid dimer, but can be separated from it by size-exclusion chromatography (see panels B and C).

(E) Non-reducing SDS-PAGE gel of pooled  $(\alpha\beta)_2$  fractions depicted in (C). Superfluous boundary and blank lanes (to the right) have been removed from the gel image.

(**F**) Competitive displacement assay of labelled IGF-I bound to holo-IGF-1R (*n* = 9) and to HybZip (*n* = 12) by unlabelled IGF-I. Error bars reflect standard error of the mean and are omitted when smaller than marker size.  $B/B<sub>0</sub>$  = percentage of binding in the absence of competing ligand.

(**G**) Competitive displacement assay of labelled insulin bound to holo-IR-B (*n* = 9) and to HybZip (*n* = 9) by unlabelled insulin. Error bars reflect standard error of the mean and are omitted when smaller than marker size.  $B/B_0$  = percentage of binding in the absence of competing ligand.



### **Figure S3. CryoEM reconstruction of IGF-I-complexed HybZip, Related to STAR Methods**.

(**A**) Representative image obtained after motion correction (scale bar = 200 Å)

(**B**) Contrast transfer function associated with (A).

(**C**) 2D classes of particles selected from the four data sets (one data set per row; individual class images have been cropped to facilitate enlarged display).

(**D**) Flowchart showing the process of 3D reconstruction of IGF-I-complexed HybZip.

Full details are provided in the **STAR Methods Detail** section.



## **Figure S4. Quality of the 3D reconstructions of the two receptor volumes, Related to STAR Methods.**

(**A**),(**C**) Gold-standard Fourier shell correlation (GSFSC) plots for the half-maps associated with the reconstruction of the "head" and "leg" region, respectively, of the IGF-I-complexed HybZip.

(**B**), (**D**) Angular distribution of particles contributing to the reconstruction of the "head" and "leg" region, respectively, of the IGF-I-complexed HybZip.

(**E**) Left panel: overlay of the "head" and "leg" regions maps, colored according to the local resolution. Right panel: ribbon diagram of IGF-I-complexed HybZip, oriented and scaled to match the local-resolution map in the left panel.

(**F**) CryoEM potential density for IR CT segment, residues Lys687-Pro718. Contour level 0.201, display restricted to within 3.0 Å of depicted residues.

(**G**) CryoEM potential density for IGF-1R domain L1, residues Tyr28-Ser35. Contour level 0.230, display restricted to within 3.0 Å of depicted residues.

(**H**) CryoEM potential density for B domain helix of IGF-I, residues Cys6-Cys18. Contour level 0.242, display restricted to within 3.0 Å of depicted residues.

(**I**) CryoEM potential density for IGF-1R CT segment, residues Glu685-Pro706. Contour level 0.201, display restricted to within 3.4 Å of depicted residues.

(**J**) CryoEM potential density for C domain of IGF-I, residues Phe23-Tyr31. Contour level 0.160, display restricted to within 3.0 Å of depicted residues.

**Table S1. CryoEM data collection parameters, related to STAR METHODS**

Data Set:				
Pixel size (Å)	1.06	1.06	1.06	1.06
Dose rate (e $.pypi^{-1}$ .sec $^{-1}$ )	8	6	b	6
Total dose (e <sup>-</sup> .Å <sup>-2</sup> )	50	52	52	52
No. frames/movie	36	50	50	50
$\mathcal{C}_s$	2.7	2.7	2.7	2.7
k٧	300	300	300	300