

Supplementary Materials for
**Single-cell multiomics sequencing reveals the reprogramming defects in
embryos generated by round spermatid injection**

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The PDF file includes:

Figs. S1 to S14
Legends for tables S1 to S7

Other Supplementary Material for this manuscript includes the following:

Tables S1 to S7

Supplementary Tables:

Table S1. The developmental potential of pre-implantation and post-implantation mouse embryos.

Table S2. Basic summary of single-cell multi-omics sequencing data in each individual cell that passed quality control.

Table S3. DEGs between ICSI and ROSI embryos at different stages.

Table S4. DEGs associated with NDRs and DEGs associated with DMRs.

Table S5. DEGs, NDRs, and DMRs amended by A366 treatment.

Table S6. List of some small compounds related to epigenetic modifications.

Table S7. Target sequences of siRNA, the primers for qPCR, liDNaseI-qPCR, bisulfite genomic PCR, and cDNA-cloning.

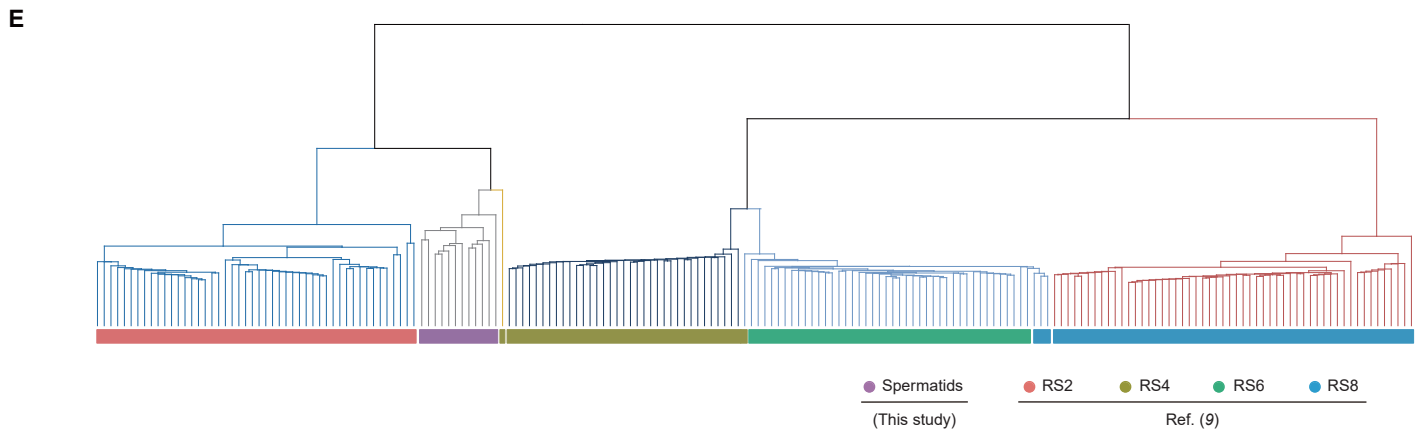
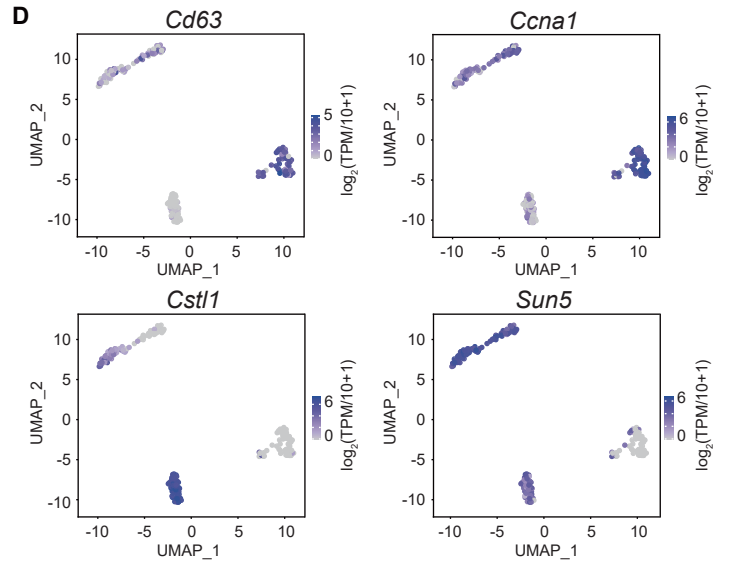
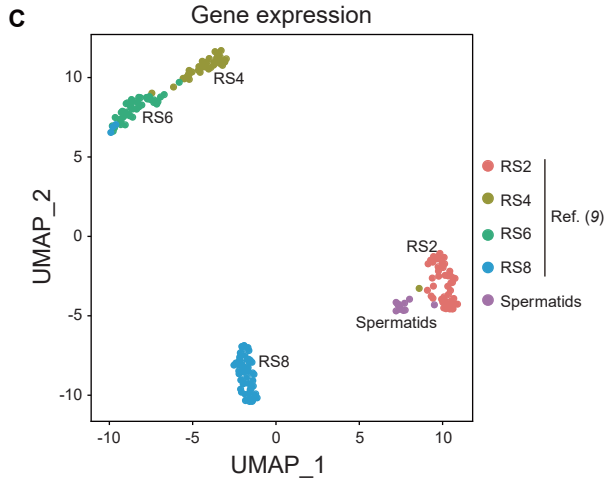
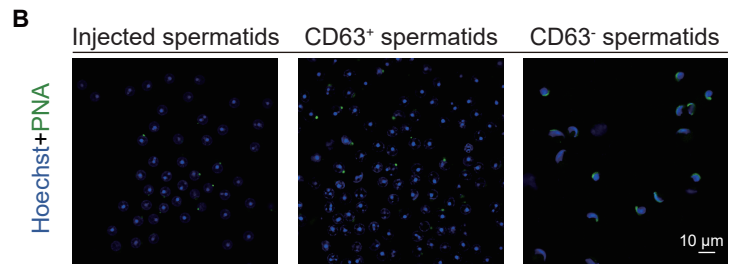
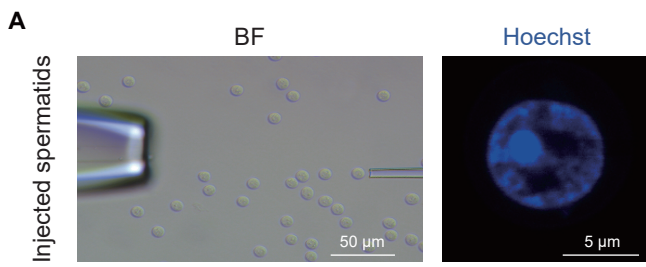


Figure S1. Characterization of the early-stage round spermatids used in this study.

(A) Representative images showing the morphology of round spermatids used in this study (left). Hoechst 33342 staining showing a representative round spermatid with a centrally located chromatin granule within the nucleus (right).

(B) Immunofluorescence of PNA in round spermatids used in this study (left), and the early-stage round spermatids (CD63⁺, middle) as well as late-stage round spermatids (CD63⁻, right) sorted by FACS as previously described (9).

(C) UMAP plot showing transcriptome data of round spermatids used in this study (n = 12) by integrating with the datasets of different stages of round spermatids in a previous study (9) (n = 184). Cells are colored by stage and experimental group.

(D) UMAP plots showing the expression levels of marker genes in the round spermatids at different developmental stages. The color from gray to blue indicates low to high expression levels.

(E) K-means clustering of round spermatids used in this study and different stages of round spermatids in a previous study (9) based on the gene expression data.

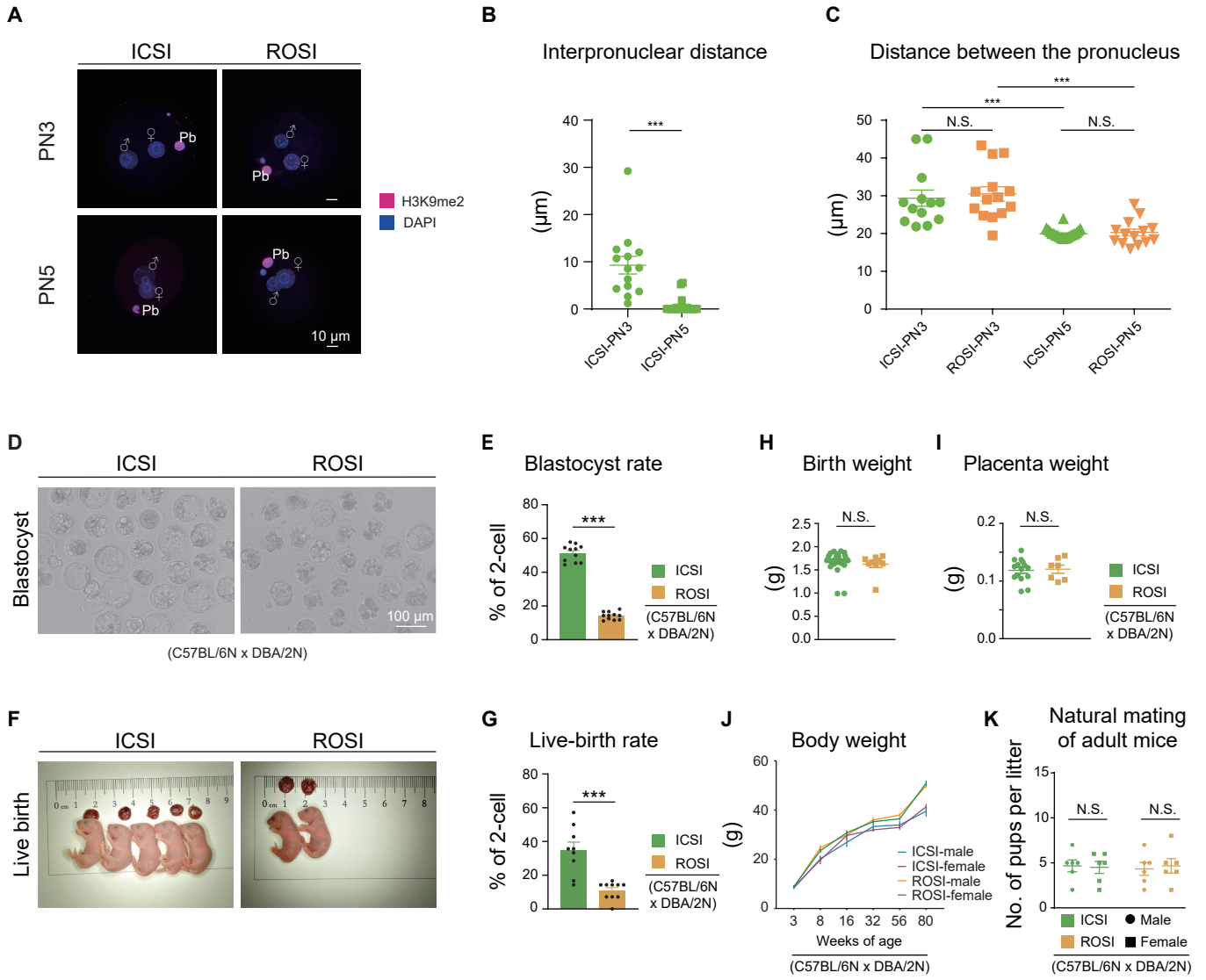


Figure S2. The developmental progress of pronuclear stage embryos derived by ICSI and ROSI at 7.5 h and 12 h post-injection, and the developmental potential of ROSI embryos.

(A) Representative immunofluorescence images of embryos at the pronuclear stages derived by ICSI and ROSI at 7.5 h (PN3) and 12 h (PN5) post-injection. Embryos are fixed and permeabilized using 4% paraformaldehyde containing 0.2% Triton X-100 for 20 min at room temperature. Blue: DAPI, red: H3K9me2. Pb: polar body, ♀: female pronucleus, ♂: male pronucleus. Scale bars, 10 μ m.

(B) Dot plot showing the interpronuclear distance of embryos at the pronuclear stages derived by ICSI at 7.5 h (PN3, n = 14) and 12 h (PN5, n = 20) post-injection. *** $P < 0.001$, unpaired two-tailed Student's t test.

(C) Dot plot showing the geometric center distance between two pronuclei of embryos at the pronuclear stages derived by ICSI (PN3, n = 13; PN5, n = 20) and ROSI (PN3, n = 14; PN5, n = 14) in three biological replicates. *** $P < 0.001$, N.S., not significant, unpaired two-tailed Student's t test.

(D) Representative ROSI and ICSI blastocysts at 94 h post-injection. Scale bar, 100 μ m.

(E) Blastocyst rate of ROSI and ICSI embryos in eleven biological replicates. *** $P < 0.001$, unpaired two-tailed Student's t test.

(F) Representative alive fetuses produced from ROSI and ICSI groups at E19.5.

(G) Live-birth rate of ROSI and ICSI embryos. *** $P < 0.001$, unpaired two-tailed Student's t test. Nine (ICSI) and ten (ROSI) replicates are performed.

(H) Dot plot showing the birth weight (ICSI, n = 26; ROSI, n = 7) of alive fetuses from ROSI and ICSI groups in five biological replicates. N.S., not significant, unpaired two-tailed Student's t test.

(I) Dot plot showing the placenta weight (ICSI, n = 15; ROSI, n = 7) of alive fetuses from ROSI and ICSI groups in five biological replicates. N.S., not significant, unpaired two-tailed Student's t test.

(J) Line chart showing the body weight of mice (from 3 to 80 week) from ROSI and ICSI groups. n = 3 per group.

(K) Dot plot showing the litter size of natural mating of adult mice from ROSI and ICSI groups in six biological replicates. N.S., not significant, unpaired two-tailed Student's t test.

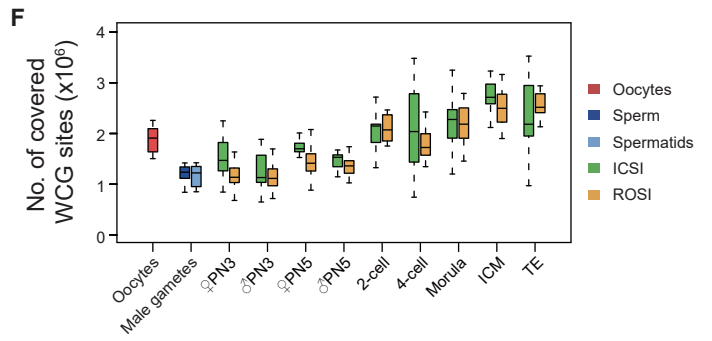
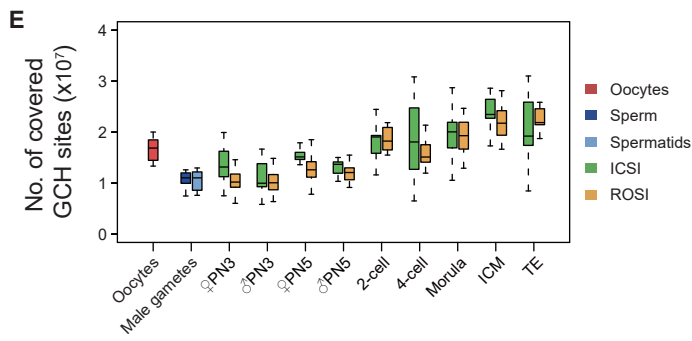
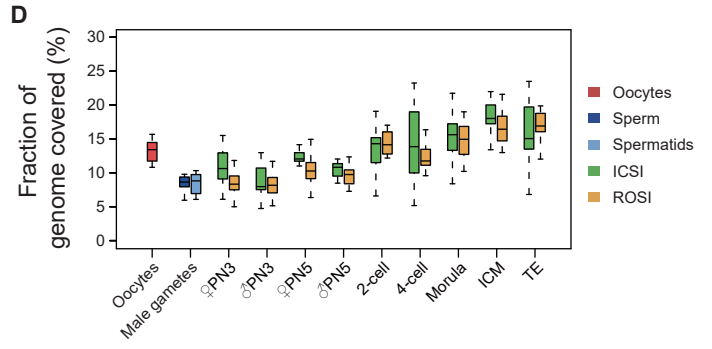
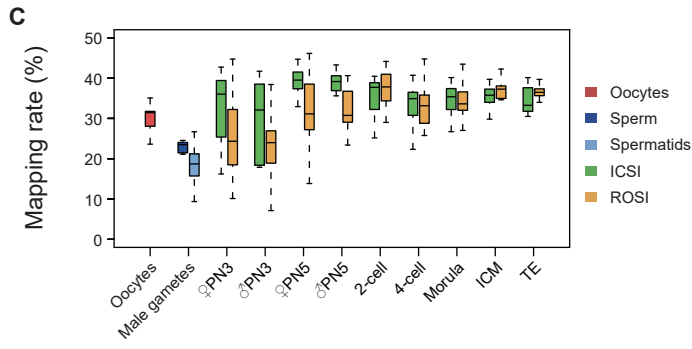
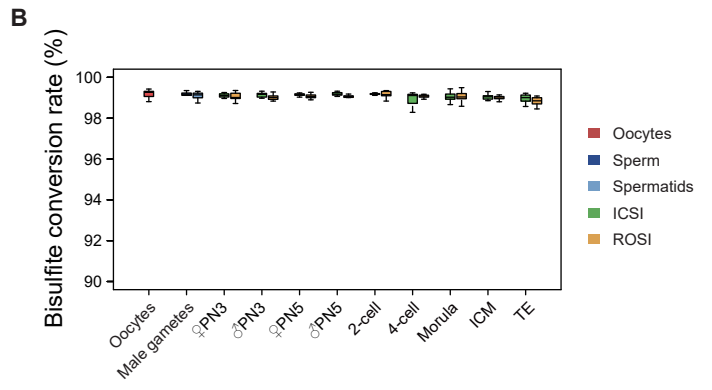
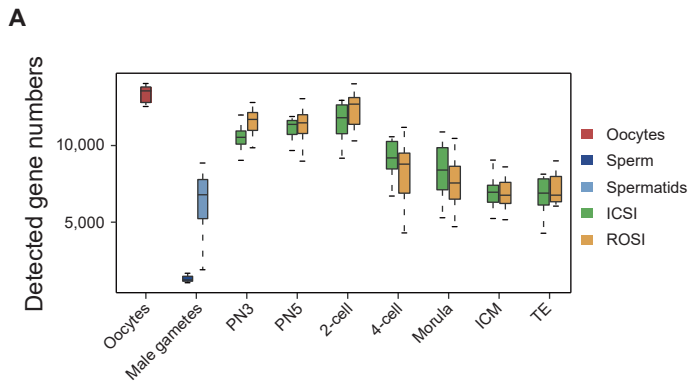


Figure S3. Quality control of single-cell multi-omics sequencing datasets.

(A) Boxplot showing the gene number detected in each individual cell.

(B-D) Boxplots showing the bisulfite conversion rate (B), mapping rate (C), and genome coverage (D) of DNA part data in each individual cell.

(E-F) Boxplots showing the numbers of GCH (GCA/GCT/GCC) sites (E) and WCG (ACG/TCG) sites (F) covered at $1\times$ depth of DNA part data in each individual cell.

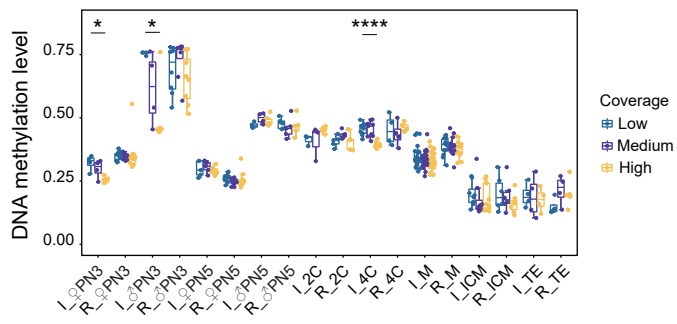
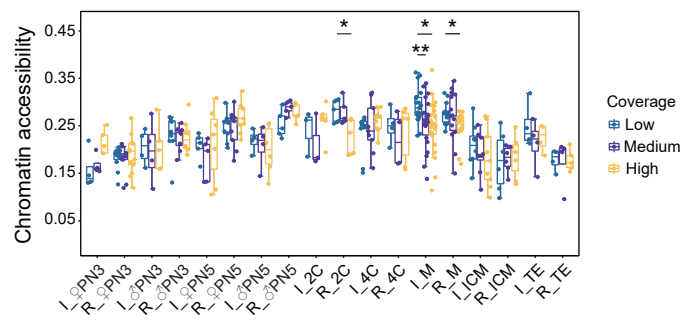
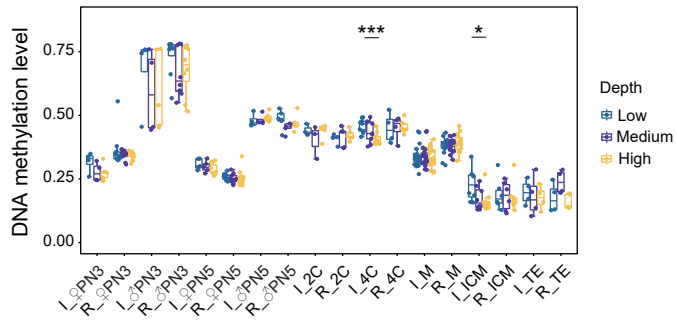
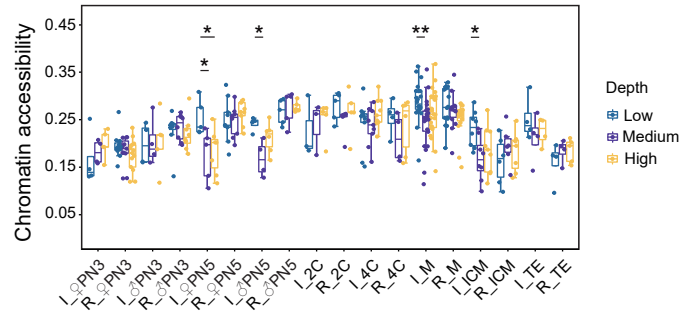
A**B****C****D**

Figure S4. The dynamics of chromatin accessibility and DNA methylation are mainly consistent in groups with different coverage or depth.

(A-B) Boxplots showing the DNA methylation level (A) and chromatin accessibility (B) of each individual cell with low (blue), medium (purple), and high (yellow) genomic coverage.

(C-D) Boxplots showing the DNA methylation level (C) and chromatin accessibility (D) of each individual cell with low (blue), medium (purple), and high (yellow) sequencing depth.

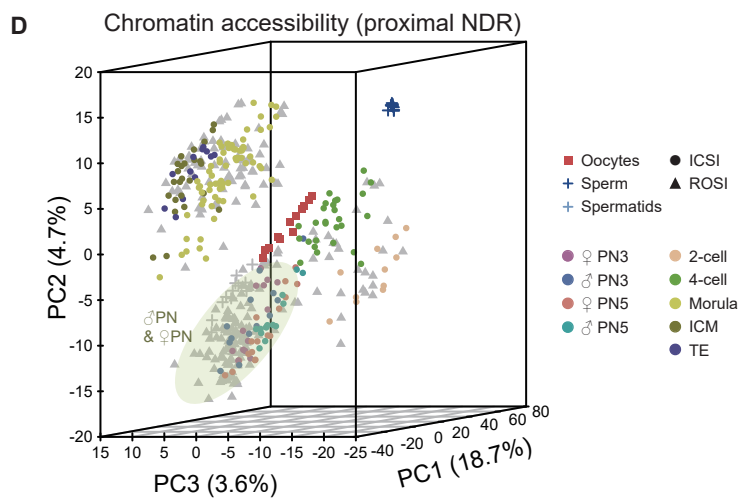
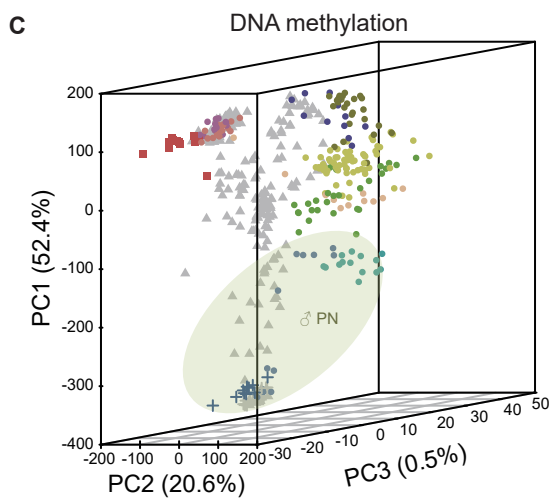
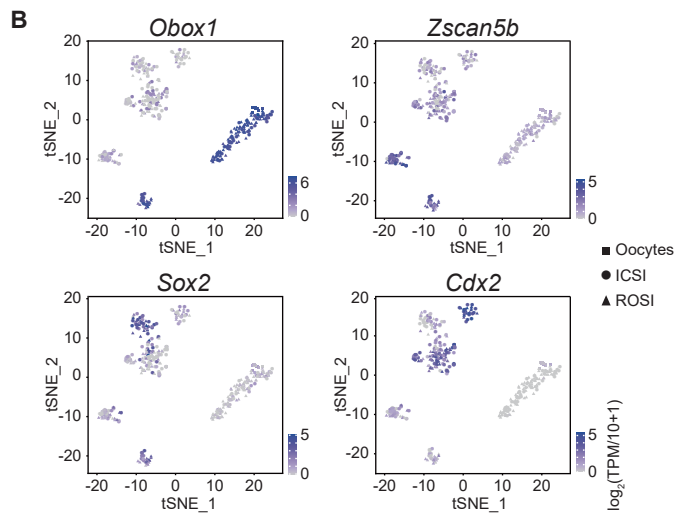
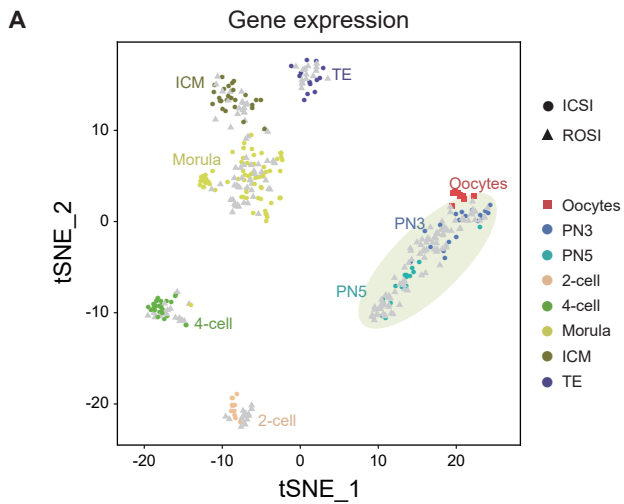


Figure S5. The dimensionality reduction analysis of transcriptome, chromatin accessibility, and DNA methylation data in the pre-implantation mouse embryos.

(A) Dimensionality reduction analysis of gene expression data using t-SNE. Each dot represents a single cell. ICSI samples are dots and are colored by the developmental stages, while ROSI samples are triangles and are in grey. Yellow-green circle indicates the pronuclei.

(B) t-SNE plots showing the expression levels of marker genes, *Obox1* (oocyte and 1-cell), *Zscan5b* (2-cell), *Sox2* (ICM), *Cdx2* (TE). The color from gray to blue indicates low to high expression levels.

(C-D) Dimensionality reduction analysis of DNA methylation data in 500-bp windows (C) and chromatin accessibility in proximal NDRs (D) using PCA. Each dot represents a single cell. ICSI samples are dots and are colored by the developmental stages, while ROSI samples are triangles and are in grey. Yellow-green circle indicates male pronucleus (C), male and female pronuclei (D), respectively.

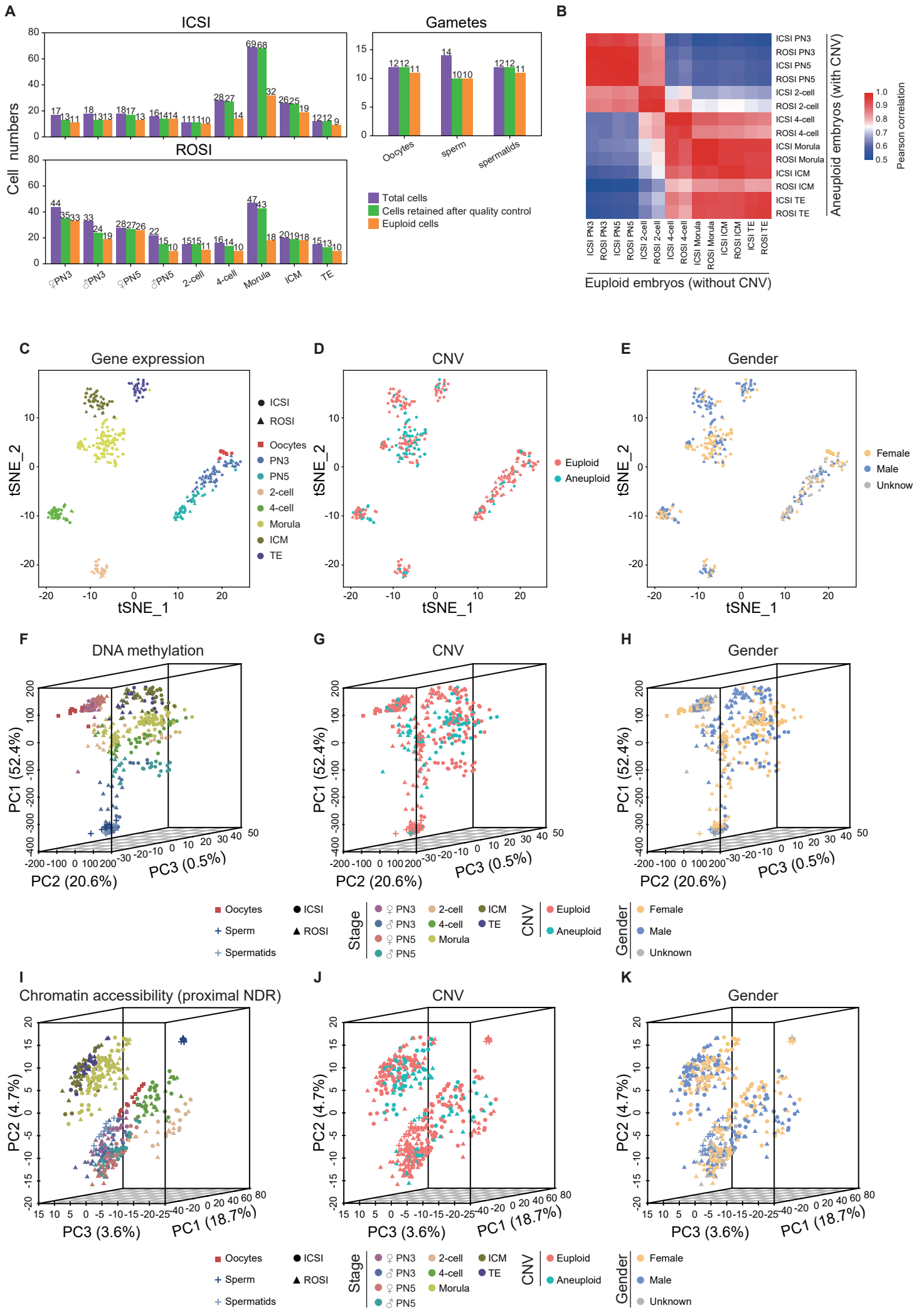


Figure S6. The overview of transcriptome, chromatin accessibility, and DNA methylation in euploid and aneuploid cells, and embryos with different gender derived by ICSI and ROSI.

(A) Bar plots showing the distributions of the number of total cells, cells retained after quality control, and euploid cells in ICSI (top), ROSI (bottom) as well as gametes (right) group, respectively.

(B) Heatmap showing the pearson correlation of transcriptome data between euploid and aneuploid cells from the pronuclear stage to blastocyst stage.

(C-E) Dimensionality reduction analysis of gene expression data colored by the developmental stages (C), CNVs (D), and gender information (E) using t-SNE. The dots represent the ICSI embryos and the triangles represent the ROSI embryos in fig. S6 C-E. The gray color in fig. S6E represents the embryos with unknown gender.

(F-H) Dimensionality reduction analysis of DNA methylation data in 500-bp windows colored by the developmental stages (F), CNVs (G), and gender information (H) using PCA. ROSI samples are triangles and ICSI samples are dots in fig. S6 F-H. The gray color in fig. S6H represents the embryos with unknown gender.

(I-K) Dimensionality reduction analysis of chromatin accessibility data colored by stage (I), CNVs (J), and gender information (K) using PCA. ROSI samples are triangles and ICSI samples are dots in fig. S6 I-K. The gray color in fig. S6K represents the embryos with unknown gender.

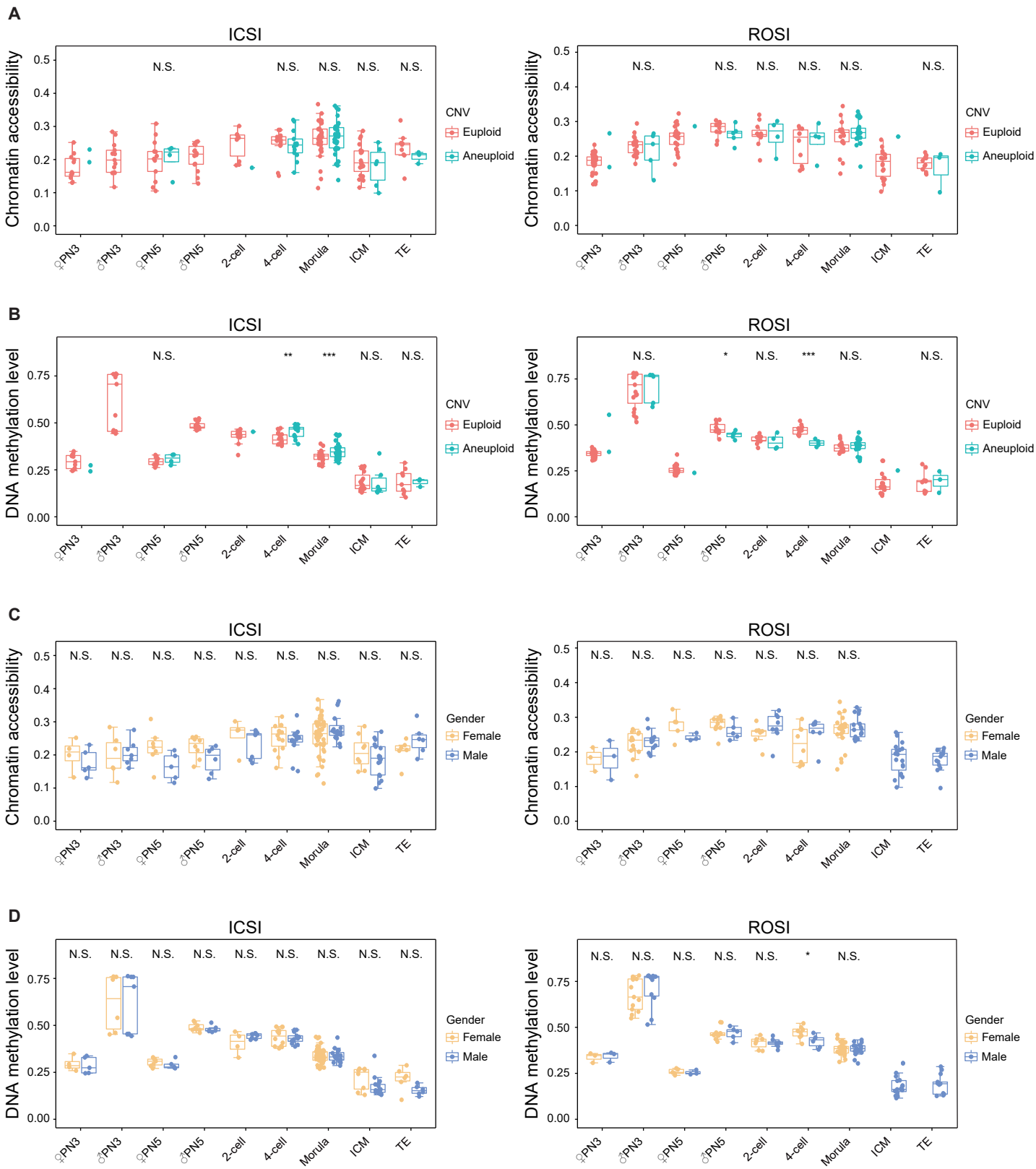


Figure S7. The dynamics of chromatin accessibility and DNA methylation in euploid and aneuploid cells, and embryos with different gender derived by ICSI and ROSI.

(A) Boxplots showing the chromatin accessibility in euploid and aneuploid cells in ICSI (left) and ROSI (right) embryos. N.S., not significant, unpaired two-tailed Student's *t* test.

(B) Boxplots showing the DNA methylation level in euploid and aneuploid cells in ICSI (left) and ROSI (right) embryos. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, N.S., not significant, unpaired two-tailed Student's *t* test.

(C) Boxplots showing the chromatin accessibility in ICSI (left) and ROSI (right) embryos with different gender. N.S., not significant, unpaired two-tailed Student's *t* test.

(D) Boxplots showing the DNA methylation level in ICSI (left) and ROSI (right) embryos with different gender. * $P < 0.05$, N.S., not significant, unpaired two-tailed Student's *t* test.

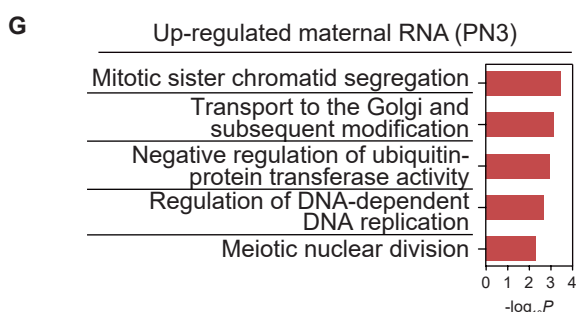
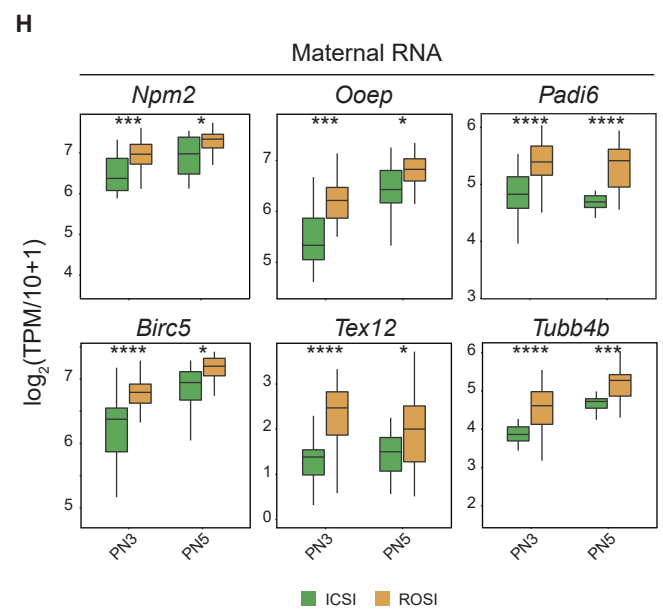
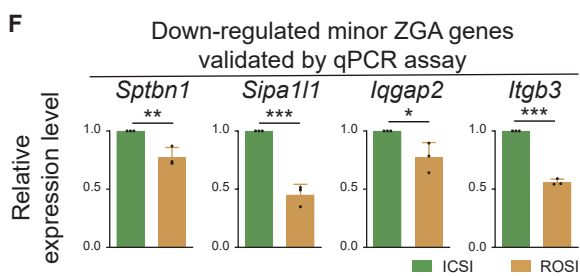
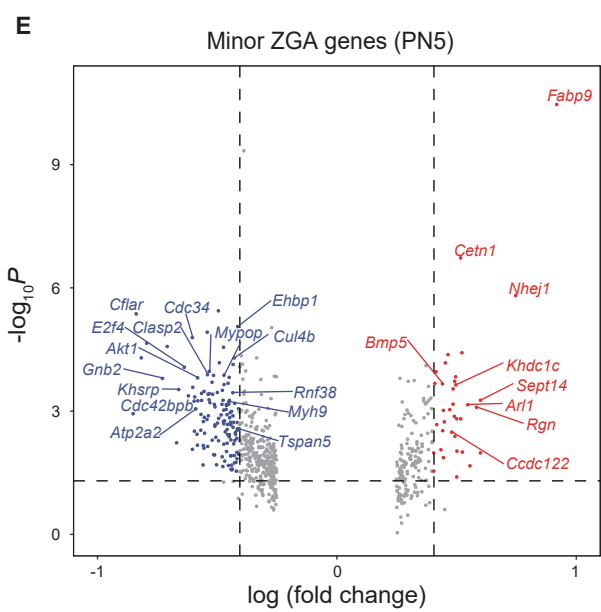
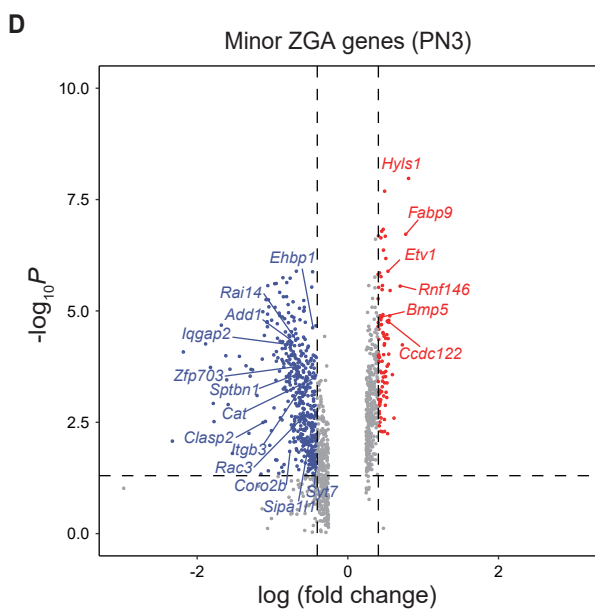
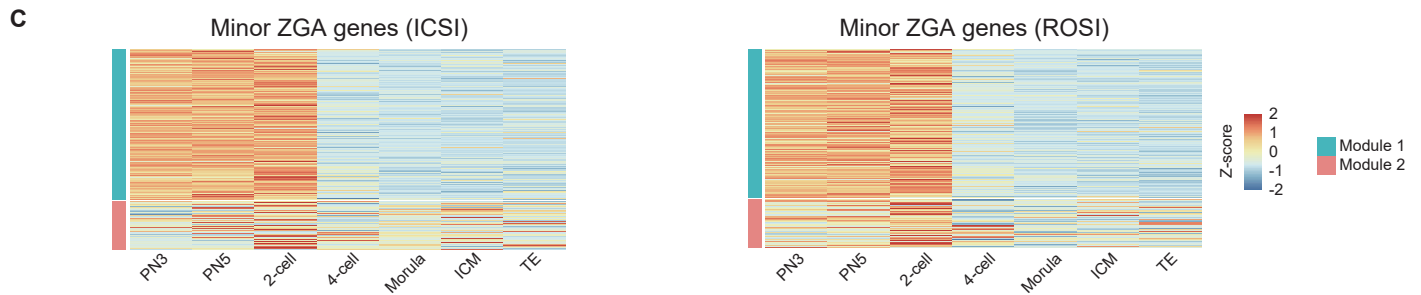
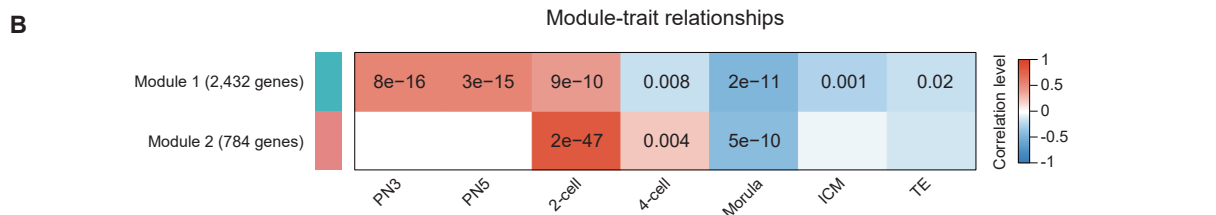
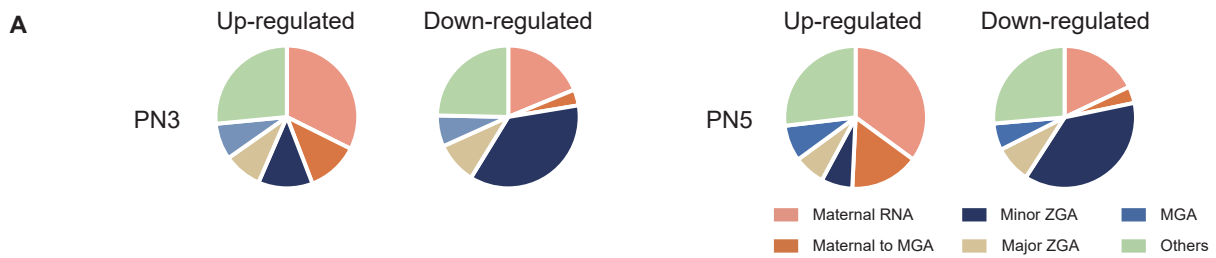


Figure S8. The mis-expression of maternal to zygotic transition associated genes in ROSI embryos.

- (A) The proportion of each gene cluster in DEGs at the PN3 (left) and PN5 (right) stages.
- (B) Heatmap showing the correlation between modules and different stages of minor ZGA genes. The number of genes in each module is marked on the left.
- (C) Heatmaps showing the expression level of each module in ICSI (left) and ROSI (right) embryos.
- (D-E) Volcano plots showing the differential gene expression of minor ZGA genes at the PN3 (D) and PN5 (E) stages. Up-regulated and down-regulated minor ZGA genes in ROSI embryos are shown as red dots and blue dots, respectively.
- (F) Bar graphs showing the relative expression levels of 4 representative down-regulated minor ZGA genes at the PN3 stage validated by qPCR assay in three biological replicates. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, unpaired two-tailed Student's t test.
- (G) Enriched GO terms (powered by Metascape) of up-regulated maternal RNA genes in ROSI embryos (PN3). The x-axis represents the significance of relative GO terms ($-\log_{10}P$, hypergeometric testing).
- (H) Expression levels ($\log_2(\text{TPM}/10+1)$) of up-regulated maternal RNA genes in ROSI and ICSI embryos at the PN3 and PN5 stages. * $P < 0.05$, *** $P < 0.001$, **** $P < 0.0001$, likelihood ratio test.

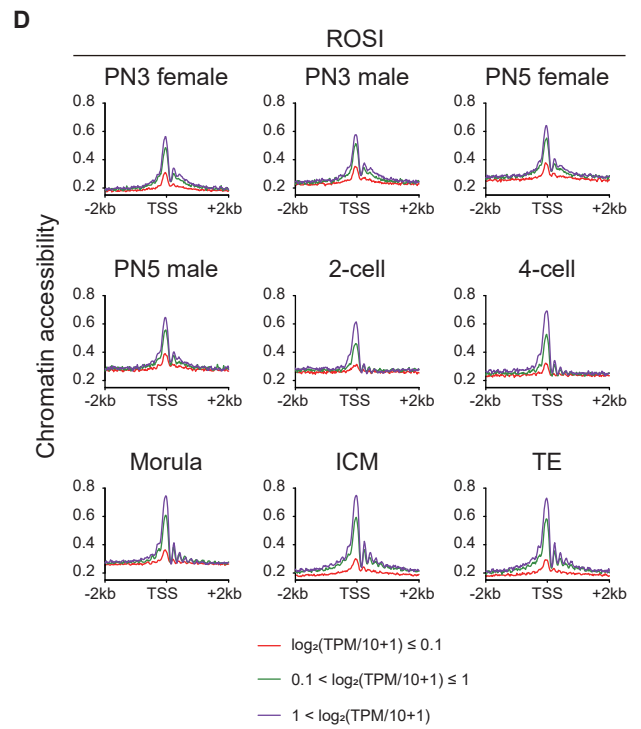
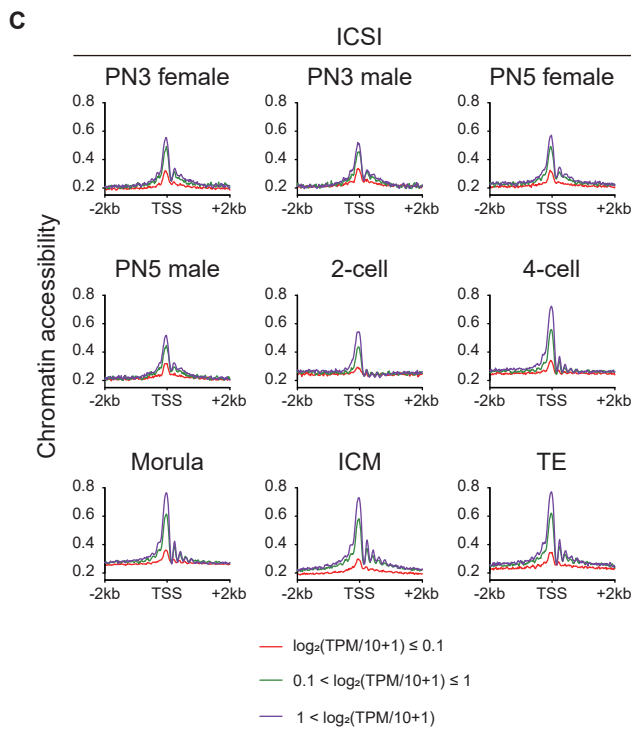
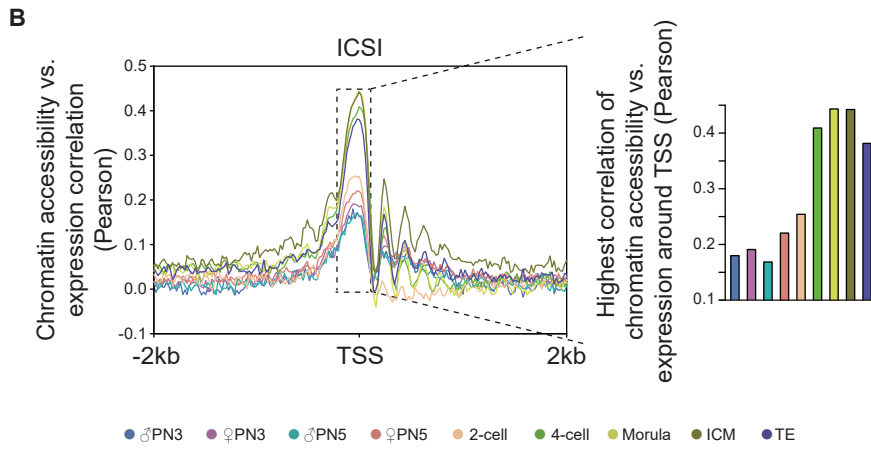
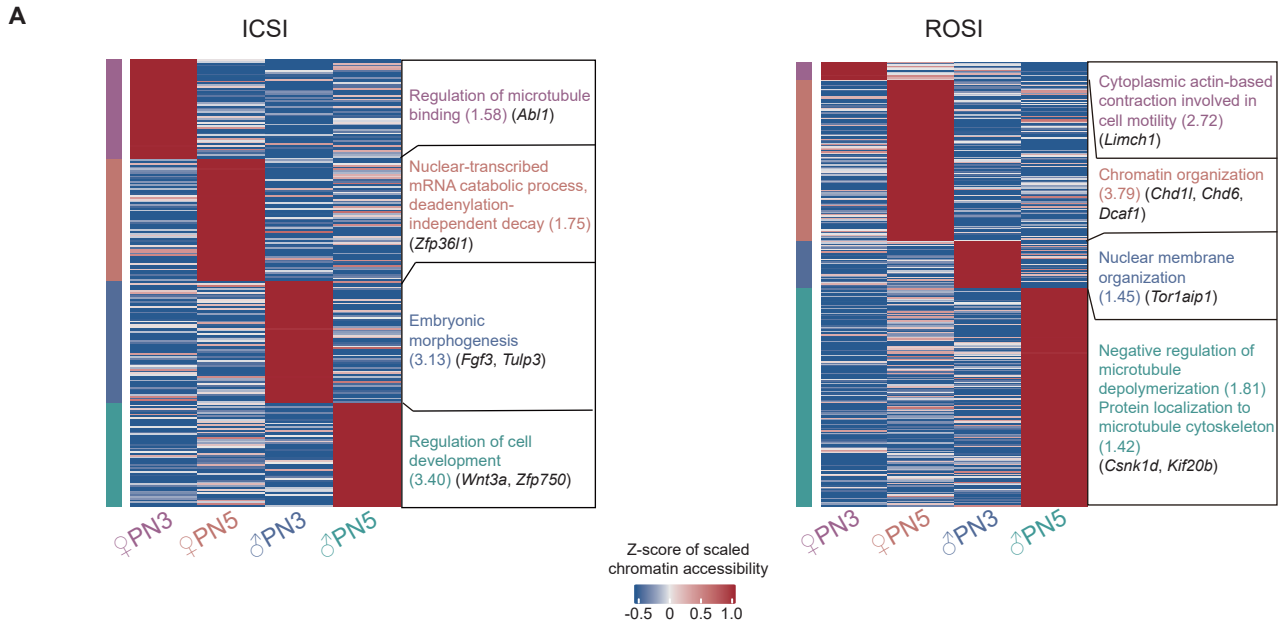


Figure S9. Chromatin accessibility reprogramming in ICSI and ROSI embryos.

(A) Heatmaps showing the scaled chromatin accessibility of stage-specific proximal NDRs in ICSI (left) and ROSI embryos (right) at the PN3 and PN5 stages. Representative GO terms and stage-specific proximal NDRs associated genes at each stage are listed on the right.

(B) Pearson correlation of chromatin accessibility and gene expression level around promoter regions (TSSs \pm 2 kb) of all the RefSeq genes in ICSI embryos at each stage. Bar plot on the right showing the highest correlation between chromatin accessibility and gene expression level around TSS in ICSI embryos at each stage.

(C-D) Chromatin accessibility on the promoters with different expression levels in ICSI (C) and ROSI (D) embryos at each stage. The genes are divided into three groups: lowly expressed ($\log_2(\text{TPM}/10+1) \leq 0.1$), intermediately expressed ($0.1 < \log_2(\text{TPM}/10+1) \leq 1$), and highly expressed ($\log_2(\text{TPM}/10+1) > 1$).

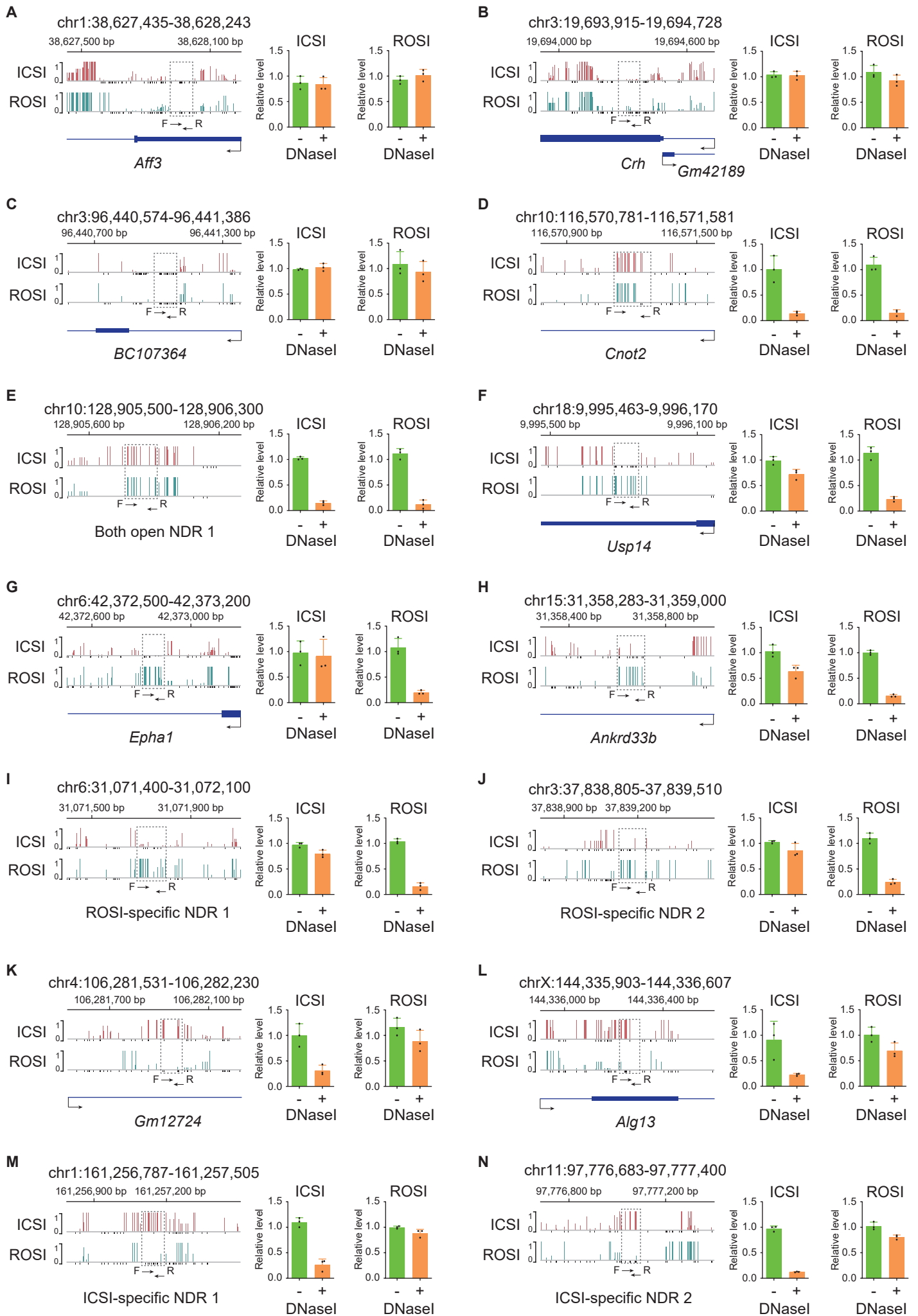


Figure S10. The validation of representative loci detected as either open or closed chromatin regions in ICSI and ROSI 4-cell embryos by liDNaseI-qPCR assay.

(A-C) Validation of 3 representative loci (*Aff3*, *Crh*, *BC107364*) detected as both closed chromatin regions by liDNaseI-qPCR assay.

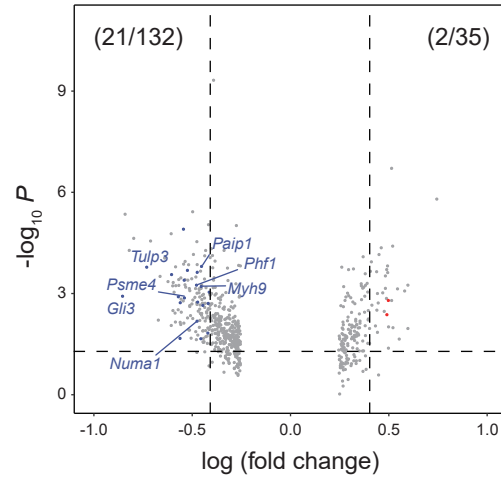
(D-E) Validation of 2 representative loci (*Cnot2*, both open NDR 1) detected as both open chromatin regions by liDNaseI-qPCR assay.

(F-J) Validation of 5 representative loci (*Usp14*, *Epha1*, *Ankrd33b*, ROSI-specific open loci1, ROSI-specific open loci2) detected as ROSI-specific open chromatin regions by liDNaseI-qPCR assay.

(K-N) Validation of 4 representative loci (*Gm12724*, *Alg13*, ICSI-specific open loci1, ICSI-specific open loci 2) detected as ICSI-specific open chromatin regions by liDNaseI-qPCR assay. Three biological replicates are performed for each locus above and the error bar indicates standard deviation.

A

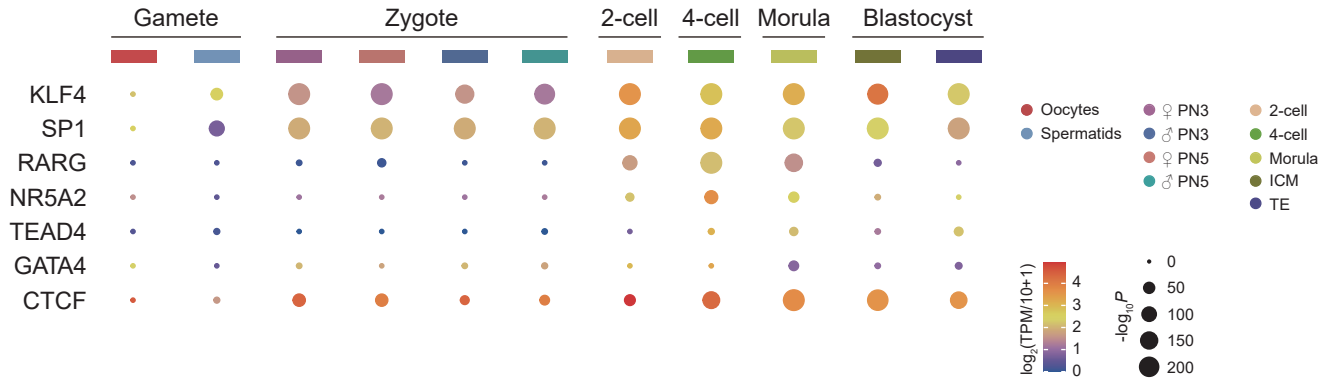
Abnormal minor ZGA genes associated with differential NDRs (PN5)



Down-regulated
 GO terms
 Establishment of spindle localization
 $-\log_{10}P$
 2.95

B

Motif enrichment in distal NDRs in ICSI embryos



C

Motif enrichment in differential distal NDRs between ICSI and ROSI embryos

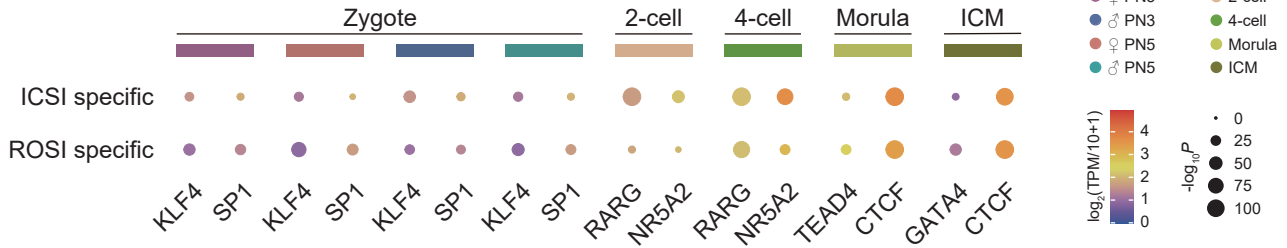


Figure S11. The relationship between chromatin accessibility and gene expression in ROSI embryos.

(A) Volcano plot showing the relationship between differential gene expression of minor ZGA genes and chromatin accessibility of proximal NDRs (PN5). The blue dots represent genes whose expression levels are significantly lower in ROSI embryos and occupied with the ICSI specific-proximal NDRs (absolute GCH methylation difference > 0.1 ; $P < 0.05$), and the red dots are vice versa.

(B) Transcription factor motifs enrichment analysis of distal NDRs in ICSI embryos. Only Transcription factors expressed ($\text{TPM} \geq 10$) and with motif enrichment $P < 10^{-10}$ in at least one stage are included.

(C) Transcription factor motifs enrichment analysis in differential distal NDRs between ROSI and ICSI embryos. Only Transcription factors expressed ($\text{TPM} \geq 10$) and with motif enrichment $P < 10^{-10}$ in at least one stage are included.

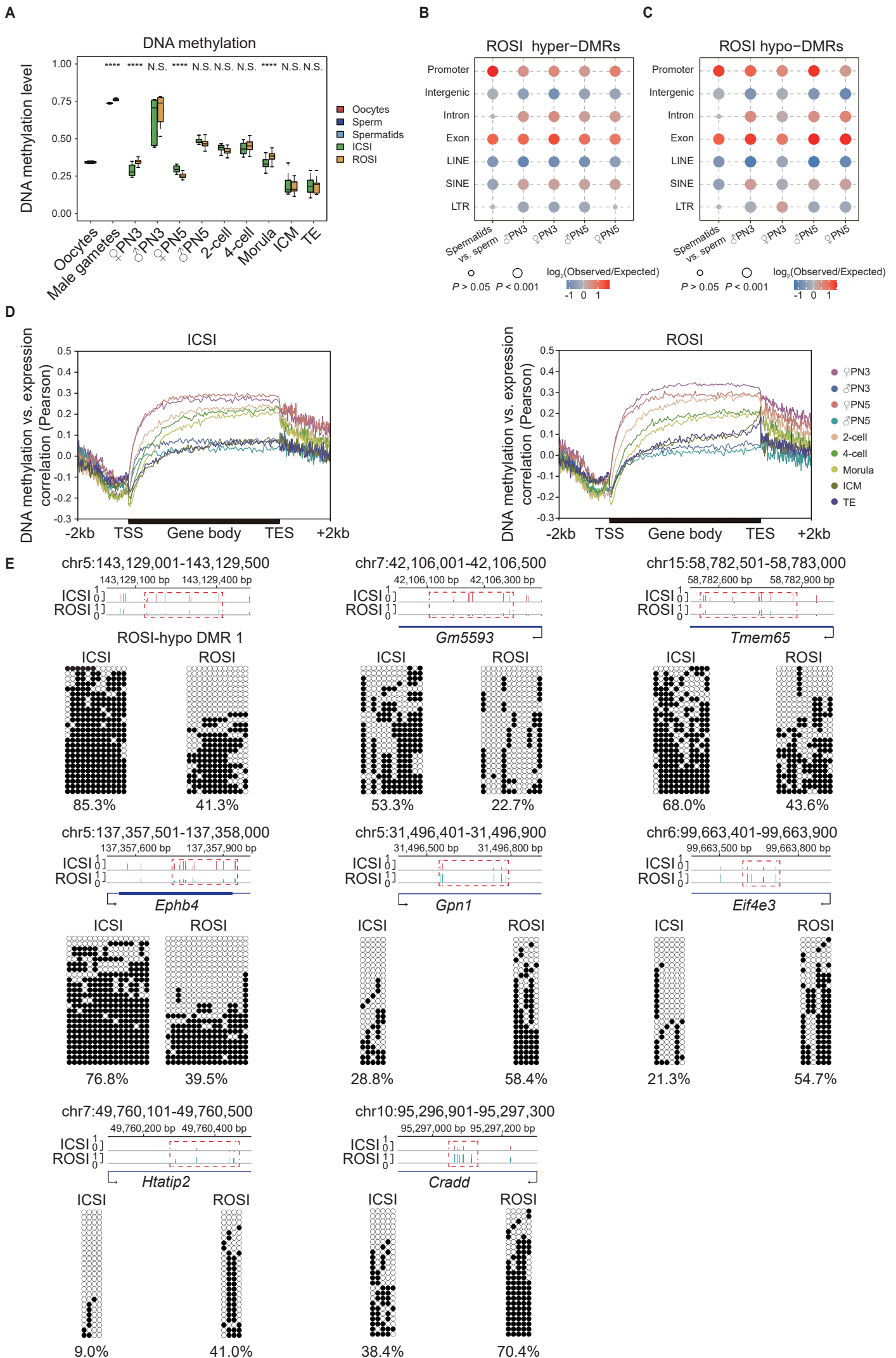


Figure S12. DNA methylation reprogramming alterations from gamete to pre-implantation embryos.

(A) Boxplot showing the average DNA methylation levels in gametes, ROSI, and ICSI embryos. The top and bottom of the boxes indicate the first and third quartiles, while the lines inside the boxes represent the median level. **** $P < 0.0001$, N.S., not significant, unpaired two-tailed Student's t test.

(B-C) Relative enrichment analysis of hyper-DMRs (B) and hypo-DMRs (C) at different genomic regions between ROSI and ICSI embryos from gamete to zygote (PN3 and PN5 stages). The color indicates relative enrichment score. The size of circles represents the P value of enrichment (chi-squared test).

(D) Pearson correlation of DNA methylation and gene expression level along gene bodies, 2 kb upstream of TSS, and 2 kb downstream of TES of all the RefSeq genes in ICSI (left) and ROSI (right) embryos at each stage.

(E) 4 representative loci (ROSI-hypo DMR 1, *Gm5593*, *Tmem65*, *Ephb4*) detected as ROSI hypo-DMRs and 4 representative loci (*Gpn1*, *Eif4e3*, *Htatip2*, *Cradd*) detected as ROSI hyper-DMRs in ICSI and ROSI embryos at morula stage by bisulfite genomic PCR-based Sanger sequencing.

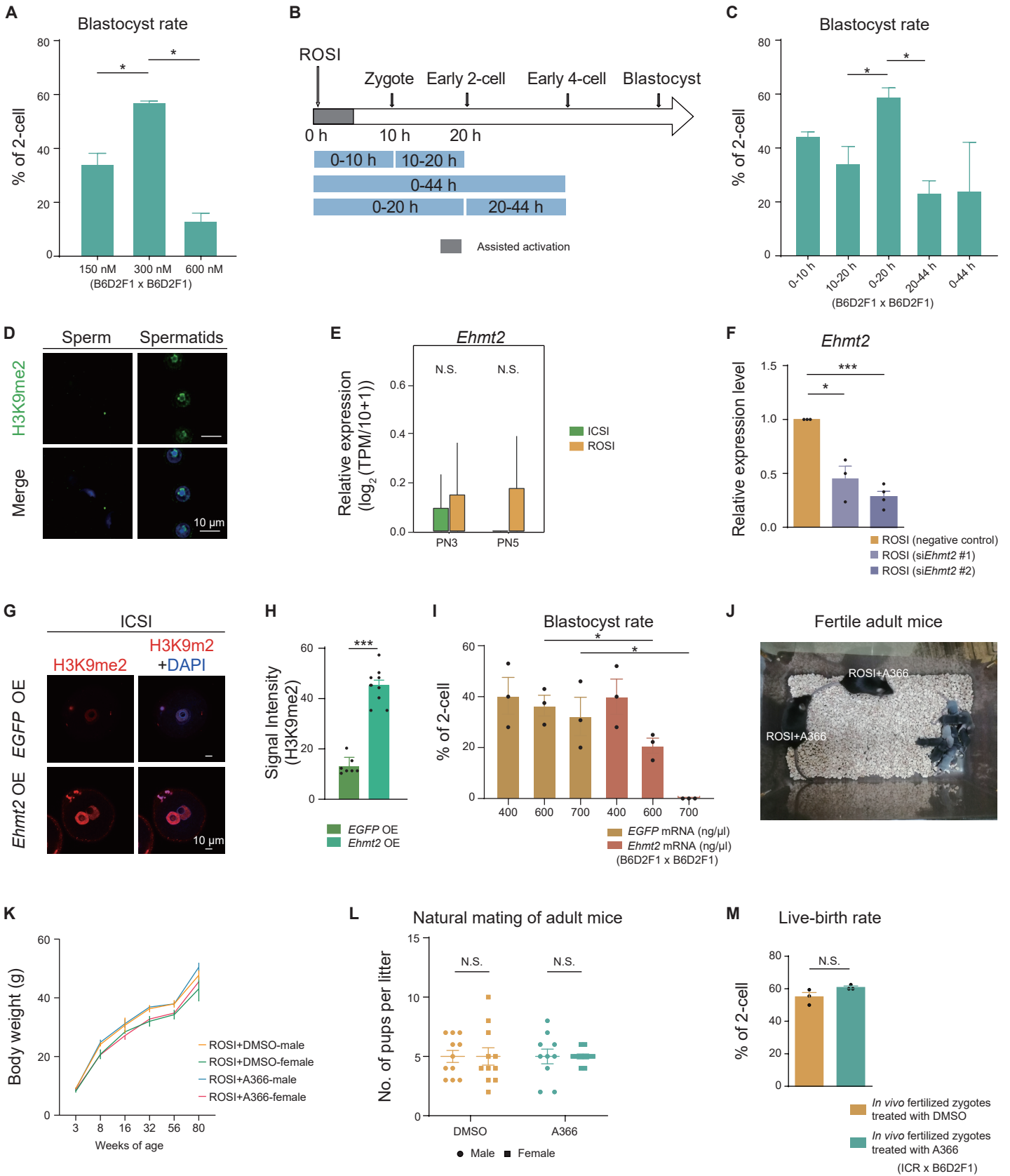


Figure S13. The optimization of A366 treatment and long-term outcomes of A366-treated ROSI mice.

(A) Blastocyst rate of the ROSI embryos treated by A366 under different concentrations in three biological replicates. $*P < 0.05$, unpaired two-tailed Student's t test.

(B) Schematic diagram showing the time frame for A366 treatment optimization.

(C) Blastocyst rate of the ROSI embryos treated by A366 under different time frames in three biological replicates. $*P < 0.05$, unpaired two-tailed Student's t test.

(D) Immunofluorescence of H3K9me2 in sperm and round spermatids used in this study. Scale bars, 10 μm .

(E) Relative expression ($\log_2(\text{TPM}/10+1)$) of *Ehmt2* in ROSI and ICSI embryos at the PN3 and PN5 stages, N.S., not significant, likelihood ratio test.

(F) Relative expression level of *Ehmt2* in ROSI embryos injected with si*Ehmt2* #1 and si*Ehmt2* #2 compared to the negative control at 22 h post-injection in three biological replicates. $*P < 0.05$, $***P < 0.001$, unpaired two-tailed Student's t test.

(G) Immunofluorescence of H3K9me2 in ICSI embryos (PN5) that overexpressed *Ehmt2*. Scale bars, 10 μm .

(H) Bar graph showing the fluorescence intensity of H3K9me2 in ICSI embryos (PN5) that overexpressed *Ehmt2* ($n = 9$) and *EGFP* ($n = 7$) in three biological replicates. $***P < 0.001$, unpaired two-tailed Student's t test.

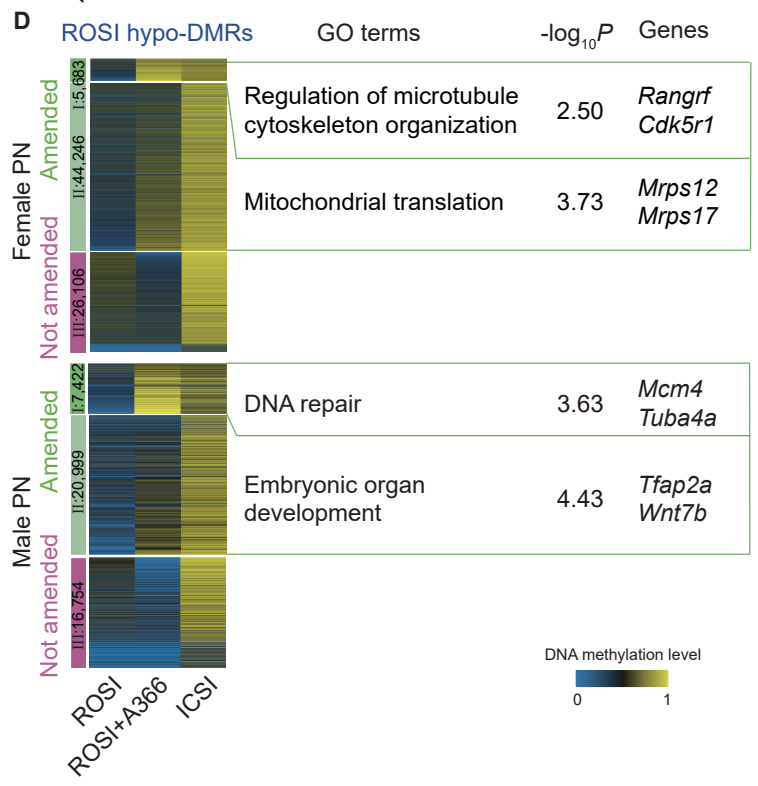
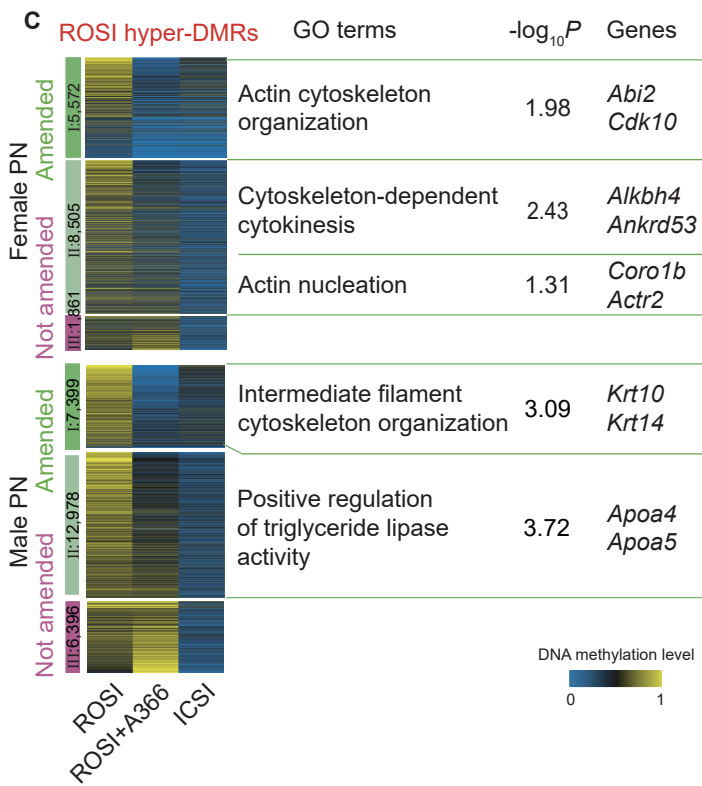
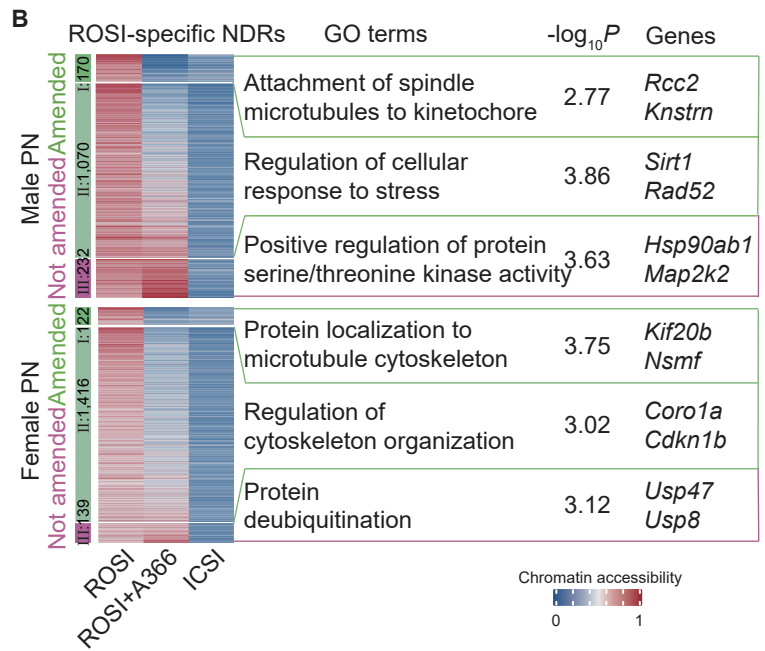
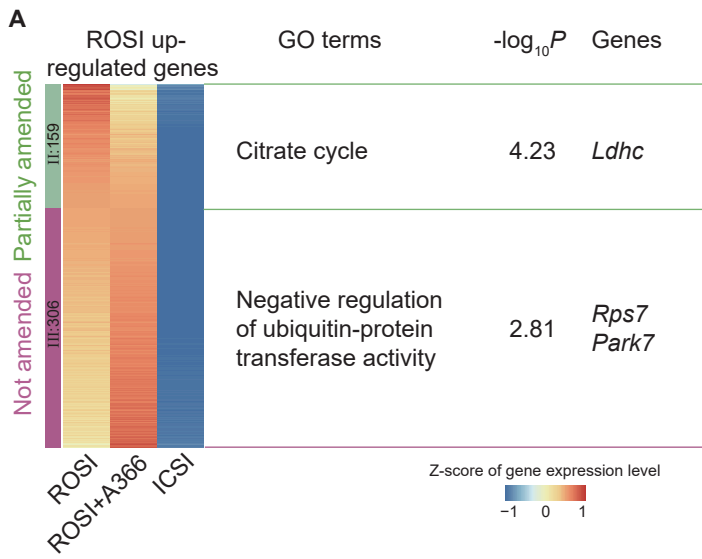
(I) Blastocyst rate of ROSI embryos injected with *Ehmt2* mRNA under different concentrations in three biological replicates. $*P < 0.05$, unpaired two-tailed Student's t test.

(J) Representative adult mice from A366-treated ROSI embryos and their pups.

(K) Line chart showing the body weight of mice (from 3 to 80 week) from ROSI group treated by A366 and the untreated control. $n = 3$ per group.

(L) Dot plot showing the litter size of natural mating adult mice from ROSI group treated by A366 ($n = 10$) and the untreated control ($n = 11$) in three biological replicates. N.S., not significant, unpaired two-tailed Student's t test.

(M) Live-birth rate of *in vivo* fertilized zygotes treated by A366 and the untreated control (DMSO) in three biological replicates. N.S., not significant, unpaired two-tailed Student's t test.



E DMRs in female PN overlapped with H3K9me2 ChIP peaks in oocytes from ref. (40)

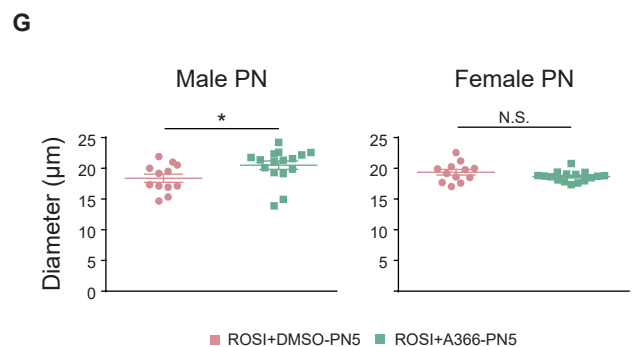
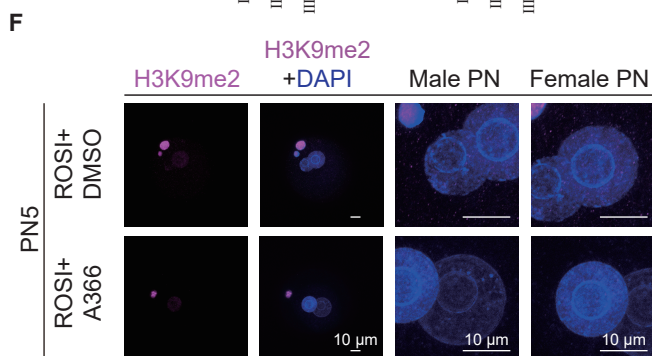
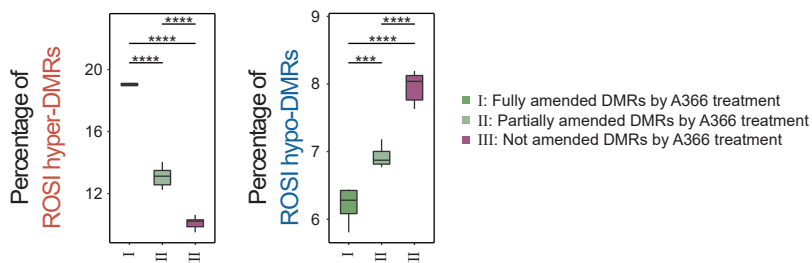


Figure S14. The amending of gene expression, chromatin accessibility, and DNA methylation in ROSI embryos by A366 treatment.

(A) Heatmap showing the expression alterations of ROSI up-regulated genes at the PN5 stage after A366 treatment. I: gene expression fully amended by A366; II: gene expression partially amended by A366; III: gene expression not amended by A366. The number of genes in each category is shown on the left. Representative GO terms and genes in each category are shown on the right.

(B) Heatmaps showing the chromatin accessibility alterations of ROSI-specific NDRs at the PN5 stage after A366 treatment. I: chromatin accessibility fully amended by A366; II: chromatin accessibility partially amended by A366; III: chromatin accessibility not amended by A366. The number of NDRs in each category is shown on the left. Representative GO terms and genes in each category are shown on the right.

(C-D) Heatmaps showing the DNA methylation alterations of ROSI hyper-DMRs (C) and hypo-DMRs (D) at the PN5 stage after A366 treatment. I: DNA methylation fully amended by A366; II: DNA methylation partially amended by A366; III: DNA methylation not amended by A366. The number of DMRs in each category is shown on the left. Representative GO terms and genes in each category are shown on the right.

(E) Boxplots showing the percentage of overlapping between randomly selected DMRs within each category (fig. S14, C and D) and H3K9me2 ChIP peaks from ref. (40). One third of DMRs in each category are randomly selected independently to calculate the overlapping percentage and are repeated for 10 times to estimate the significance of difference. *** $P < 0.001$, **** $P < 0.0001$, unpaired two-tailed Student's t test.

(F) Immunofluorescence of H3K9me2 in A366-treated ROSI embryos and the untreated control (PN5). Embryos are fixed and permeabilized using 4% paraformaldehyde containing 0.2% Triton X-100 for 20 min at room temperature. The magnified image of male and female pronuclei are shown on the right. Scale bars, 10 μm .

(G) Dot plots showing the diameter of male (left) and female (right) pronuclei in A366-treated ROSI embryos ($n = 14$) and the untreated control ($n = 14$) (PN5) in three biological replicates. * $P < 0.05$, N.S., not significant, unpaired two-tailed Student's t test.